



Metabolic Syndrome and Skin Diseases

Yu Hu^{1,2}, Yun Zhu², Ni Lian¹, Min Chen^{1*}, Andrzej Bartke² and Rong Yuan^{2*}

¹ Jiangsu Key Laboratory of Molecular Biology for Skin Diseases and Sexually Transmitted Infections, Institute of Dermatology, Chinese Academy of Medical Sciences and Peking Union Medical College, Nanjing, China, ² Department of Internal Medicine, Southern Illinois University School of Medicine, Springfield, IL, United States

The increasing prevalence of Metabolic syndrome (MetS) is a worldwide health problem, and the association between MetS and skin diseases has recently attracted growing attention. In this review, we summarize the associations between MetS and skin diseases, such as psoriasis, acne vulgaris, hidradenitis suppurativa, androgenetic alopecia, acanthosis nigricans, and atopic dermatitis. To discuss the potential common mechanisms underlying MetS and skin diseases, we focus on insulin signaling and insulin resistance, as well as chronic inflammation including adipokines and proinflammatory cytokines related to molecular mechanisms. A better understanding of the relationship between MetS and skin diseases contributes to early diagnosis and prevention, as well as providing clues for developing novel therapeutic strategies.

OPEN ACCESS

Edited by:

Damian G. Romero, University of Mississippi Medical Center, United States

Reviewed by:

Julio Sartori-Valinotti, Mayo Clinic, United States Anne-Francoise Burnol, Institut National de la Santé et de la Recherche Médicale (INSERM), France

*Correspondence:

Min Chen drchenmin@126.com Rong Yuan ryuan@siumed.edu

Specialty section:

This article was submitted to Cellular Endocrinology, a section of the journal Frontiers in Endocrinology

Received: 29 August 2019 Accepted: 29 October 2019 Published: 20 November 2019

Citation:

Hu Y, Zhu Y, Lian N, Chen M, Bartke A and Yuan R (2019) Metabolic Syndrome and Skin Diseases. Front. Endocrinol. 10:788. doi: 10.3389/fendo.2019.00788 Keywords: metabolic syndrome, skin diseases, insulin resistance, adipokine, proinflammatory cytokine

INTRODUCTION

Metabolic syndrome (MetS), first described in 1988 by GM Reaven as "syndrome X," is a group of abnormalities which includes abdominal obesity, hypertension, insulin resistance, and dyslipidemia (1, 2). It is estimated that about one-quarter to one-third of the world's population is affected by MetS (3). Growing evidence from clinical studies suggests that there exists a strong connection between MetS and skin diseases (4, 5). However, mechanistic insights regarding the relationship of metabolic syndrome and skin diseases are notably lacking. In this review, we will summarize the epidemiological evidence for the association between MetS and skin diseases, as well as discuss the potential common mechanisms underlying MetS and skin diseases.

EPIDEMIOLOGICAL EVIDENCE OF THE CONNECTION BETWEEN METS AND SKIN DISEASES

Psoriasis

Psoriasis is one of the most common chronic inflammatory skin diseases, with the prevalence ranging from 0.51 to 11.43% in adults and from 0 to 1.37% in children (6). The association between psoriasis and MetS has been investigated in numerous studies. Two recent systematic reviews demonstrated that psoriasis patients have an increased risk for MetS (7, 8). Singh and his colleagues (7) reviewed 17 articles involving 3,791 psoriasis patients in a group of 28,939 participants from January 1946 to June 2016, and reported a higher prevalence of both MetS and the individual components of MetS including abdominal obesity, elevated fasting plasma glucose, and high blood pressure in the psoriasis patients. The same group then updated the data from June 2016 to January 2017 and conducted a meta-analysis. The meta-analysis included 35 studies with 1,450,188 participants, of whom 46,714 were psoriasis patients.

They found that, compared to the general population, psoriasis patients had a higher prevalence of MetS with the pooled odds ratio (OR) of 2.14 (95% CI 1.84–2.48) (9). Another systematic review and meta-analysis investigated 14 papers including 25,042 psoriasis patients and also found that 31.4% psoriasis patients had MetS with the pooled OR of 1.42 (95% CI, 1.28–1.55) (8). When examining the association between disease severity and MetS, a dose-dependent relationship between Psoriasis Area and Severity Index (PASI) and the prevalence of MetS was observed (9). The adjusted ORs for MetS in severe, moderate, and mild psoriasis were 1.98, 1.56, and 1.22, respectively. Additionally, a recent population-based study in United Kingdom also supported a positive correlation between PASI and the prevalence of MetS, although it was not significant (OR = 1.18, 95% CI, 0.97–1.44, p = 0.099) (10).

Acne Vulgaris

Acne vulgaris is an inflammatory disorder of the pilosebaceous unit which affects many adolescents and young adults. A crosssectional study was performed by comparing the metabolic conditions in 100 male acne patients and 100 male controls. It showed that the prevalence of metabolic syndrome tends to be higher in acne patients (17%) compared with controls (9%) (p = 0.09), and also that prevalence of insulin resistance is significantly higher in acne patients (22%) than controls (11%) (p = 0.03) (11). Another study of 243 acne patients and 156 controls also found that fasting insulin levels were significantly higher in acne patients (12). However, a recent nationwide study in Israel showed that acne and metabolism have a complicated relationship. In this study, including 600,404 adolescents, the researchers found that overweight or obesity is inversely associated with acne in a dose-dependent manner, suggesting that excessive body mass index (BMI) has a protective effect against acne (13). The possible mechanism of the protective effect is increased aromatase activity, which leads to a greater conversion of testosterone to estradiol (14). Estrogen is known to act by anti-androgenic effect, inhibiting sebum secretion and attenuating proinflammatory cytokine activity (15). Moreover, obesity is reported to reduce the conversion of testosterone to the more physiologically active dihydrotestosterone by suppressing the activity of $5-\alpha$ reductase-II (16).

Hidradenitis Suppurativa

Hidradenitis suppurativa (HS) is a chronic inflammatory skin disease involving the follicular portion of folliculopilosebaceous units with an estimated prevalence from 0.05 to 4.10% (17). The accumulating data have shown a positive association between HS and MetS (18–20). A cross-sectional study in Israel, which included 3,207 HS patients and 6,412 controls, revealed that HS was significantly associated with MetS (OR 1.61, 95% CI 1.36–1.89) (18). When comparing the prevalence of MetS in both hospital HS and population HS groups vs. non-HS groups, the ORs were 3.89 (95%CI, 1.90–7.98) for hospital HS and 2.08 (95%CI, 1.61–2.69) for population HS (19). Another retrospective review, which enrolled 366 HS patients and 366 controls of all races, also indicated that patients with HS might have a greater risk for MetS. They found the prevalence of MetS

in HS patients (50.6%) was significantly higher than controls (30.2%, p = 0.001) (20).

Androgenetic Alopecia

Androgenetic alopecia (AGA) is a common type of progressive non-scarring hair loss in both men and women. The incidence of AGA varies by race and age. Around 30% of Caucasian men will have AGA at the age of 30, 50% at the age of 50, and 80% at the age of 70. The prevalence of AGA in Caucasian women is about 19%. Asians are less affected by AGA than both male and female Caucasians (21). Numerous studies have reported a strong association between AGA and MetS (22-26), although one case-control study (27) in Turkey of 74 male AGA patients and 42 controls, found that there was no difference in the rate of MetS. However, another case-control study by Dharam and his colleagues found a higher prevalence of MetS in AGA patients (53%) than controls (17%) (P < 0.001) (26). Similar findings were reported in a hospital-based cross-sectional study in India, in which 19 of 85 (22.4%) AGA patients were affected with MetS compared with 8 of 85 (9.4%) controls (p = 0.021) (22). However, when analyzing the relationship between each component of MetS and AGA, different results were reported. A survey conducted in Taiwan noted that high-density lipoprotein cholesterol (HDL-C) (OR 2.36, 95% CI 1.41–3.95; *P* = 0.001) was the most significant factor associated with AGA (25). Another case-control study reported that waist circumference (>102 cm) was the most significant risk factor for AGA patients to develop MetS with the value of 1.25 (95% CI = 1.10-1.42, P < 0.001) (28). Similarly, the research in female AGA patients revealed waist circumference (OR 5.6, 95% CI 2.2 -13.9, P = 0.0002) and hypertension (OR 3.5, 95% CI 1.3–8.9, P = 0.008) were the most important factors for AGA (29).

Acanthosis Nigricans

Acanthosis nigricans (AN) is a common skin disorder characterized by hyperpigmented, velvety patches and plaques involving the intertriginous areas. The prevalence of AN varies among different ages and ethnicities, reaching 25% in the general population and even over 60% in overweight and obese children (30). There is solid evidence for the existence of a link between AN and MetS (31-33). For overweight and obese women, the incidence of MetS is significantly higher in AN patients (60%) than controls (37.6%) (p = 0.0001)(31). A case-control study, which included 100 children who were overweight or obese, showed that 73% of them had a diagnosis of MetS with a strong association to AN (OR 1,872 [95% CI: 112.9-31,028]) (32). Another cross-sectional study also found that AH in overweight and obese children might be a clinical indicator of MetS, with the elevation of body fat, blood pressure, insulin, and homeostasis model assessment index (33). Furthermore, a 2-year multicenter case-control study including 123 patients at the age of 38.83 \pm 8.62 years in India, found that facial AN was strongly related to impaired glucose tolerance, increased waist-hip ratio (WHR), and increased BMI. The authors suggest that the facial AN might be a potential clinical marker of MetS (34).

Atopic Dermatitis

Atopic dermatitis (AD) is a chronic pruritic inflammatory dermatosis which is commonly associated with other atopic disorders including food allergies, asthma, hay fever, and allergic rhinitis. The worldwide prevalence of AD is approximately 15-20% in children and 1-3% in adults (35). The association between AD and MetS has not been fully clarified. A crosssectional study in 5,007 Korean adults reported that MetS (OR 2.92, 95% CI 1.49-5.73), central obesity (OR 1.73, 95% CI 1.09-2.75), and hypertriglyceridemia (OR 2.20, 95% CI 1.19-4.05) correlated positively with AD in women (36). However, unlike the psoriasis cohort, the prevalence ratios (PR) for MetSassociated components including hypertension (PR 0.83, 95% CI 0.81-0.85), hyperlipidemia (PR 0.94, 95% CI 0.91-0.95), and diabetes (PR 0.82, 95% CI 0.80-0.86) showed no difference in AD patients compared to non-AD patients (37). Moreover, a recent systematic review, which included 14 studies evaluating the relationship between AD and MetS, reported that the association between AD and MetS was not causal. The associations between hypertension, hyperglycemia, cholesterol levels, and AD all remain unclear. According to the systematic review, central obesity is the only component that correlates positively with AD (38). This phenomenon was also reported in a case-control study and a systematic review. These two studies both reported that obesity is associated with increased prevalence and severity of AD (39, 40).

MECHANISMS OF METS AND SKIN DISEASES

Although the exact mechanisms of the association of MetS and skin diseases remain vague, it is of great importance to identify the potential common pathways behind MetS and associated skin diseases in order to develop novel therapeutic strategies. Of all the potential pathological mechanisms of MetS, emerging evidence suggests that impaired insulin signaling and increased insulin resistance, as well as elevated status of chronic inflammation, are major risk factors that may induce skin diseases (41–43).

Insulin Signaling and Insulin Resistance

Insulin, secreted by the beta cells of the pancreas, is involved in glucose homeostasis, lipid metabolism, and anabolic processes by activating the insulin signaling pathways. Insulin initiates its function by binding to insulin receptor (IR). After stimulation, IR is activated via autophosphorylation and leads to the phosphorylation and activation of the downstream substrates, such as insulin receptor substrates (IRS-1, IRS-2) and Shc. Then the phosphorylated tyrosine residues of IRS and Shc can activate the two main insulin signaling pathways: the phosphatidylinositol-3-kinase (PI3K)/Akt pathway and the mitogen-activated protein kinases (MAPK)/Ras pathway (44). In the PI3K/Akt pathway, the phosphorylated tyrosine residues of IRS activate PI3K and phosphorylates phosphatidylinositol (4, 5)-bisphosphate (PIP2), leading to the formation of phosphatidylinositol 3-5 three phosphoric acid (PIP3). Then, PIP3 facilitates the phosphorylation of Akt by 3-Phosphoinositide-dependent protein kinase-1 (PDK1). Once activated, Akt phosphorylates tuberous sclerosis complex 2 (TSC2) and suppresses its inhibitory effect on mechanistic target of rapamycin complex 1 (mTORC1), thus upregulating the activity of mTORC1, which regulates protein synthesis and cell growth (45). The activated Akt may also phosphorylate the downstream target FOXO1, leading to nuclear export and inhibiting FOXO1 transcriptional activity (46). FOXO1 is involved in adipogenesis and gluconeogenesis by controlling the expression of lipogenic and gluconeogenic genes, as well as cell proliferation and apoptosis (47, 48). The other branch of the insulin signaling pathway is the MAPK/Ras pathway, which is mediated by the growth factor binding protein 2 (Grb2). Active Ras then interacts with, and leads to, the activation of downstream Raf and MAP kinases ERK1/ERK2. After activation, ERK1/2 regulates gene expression, cell proliferation, differentiation, and cell growth by translocating to the nucleus and catalyzing the phosphorylation of transcription factors, such as Elk-1 and Sep-1a (Figure 1) (49, 50).

Insulin resistance is defined as the reduced responsiveness of target tissues to normal insulin levels and is widely accepted as the primary mechanism of the pathophysiology of MetS. Though many cell types express insulin receptors, liver, skeletal muscle, white adipose tissue, and brain are the main tissues responsible for glucose homeostasis (51). In insulin resistance, the cells are resistant to insulin and fail to maintain the blood glucose level. To achieve a normal blood glucose level, pancreatic β -cells secret excessive insulin, which leads to hyperinsulinemia (52). With further resistance to insulin, the function of β-cells is impaired and becomes insufficient to maintain the normal blood glucose range, resulting in hyperglycemia. A high plasma glucose level also results from an impaired inhibitory action of insulin on hepatic glucose production, a reduction of glycogen synthesis in hepatocytes and the inability of skeletal muscle and adipocytes to take up glucose (53). The possible pathophysiologic mechanisms that provoke insulin resistance in MetS include defective insulin signaling, impaired glucose disposal, lipotoxicity, and inflammatory cytokines.

The two main target cell types in skin diseases, keratinocytes and fibroblasts, both have insulin receptors and IGF receptors. Insulin has been reported to cross the dermal-epidermal junction to affect keratinocytes (54). The growth-promoting effects of insulin in keratinocytes and fibroblasts rely on the following mechanisms. First, insulin resistance and compensatory hyperinsulinemia decrease the expression of insulin-like growth factor binding proteins (IGFBPs) (55). IGFBPs bind to IGFs and extend their half-life, thereby controlling the transport of IGFs to target tissues and regulating the level of circulation IGFs (56). The high levels of free IGF-1 lead to the proliferation and differentiation of fibroblasts and keratinocytes (57). IGFBPs also exert antiproliferative effects by blocking the interaction of IGF and IGF-receptors because the affinity of IGFs for IGFBPs is similar to that of the IGFs and IGF-receptors (58). Second, under low insulin concentrations, insulin is more likely to bind to its classic receptor and then regulate glucose metabolism. Under insulin resistance and



compensatory hyperinsulinemia condition, insulin might have a potential to bind to IGF-receptors and stimulate the proliferation of keratinocytes and fibroblasts (59). However, it is reported that the affinity of insulin binding to IGF-receptors is approximately 2000-fold lower than binding to IR (60). Thus, we propose that the compensatory hyperinsulinemia exceeds the binding capacity of IR, allowing the insulin to bind to IGF-receptors. These two mechanisms have been proposed as the possible pathogenesis of acanthosis nigricans. In brief, hyperinsulinemia in acanthosis nigricans patients not only can increase the binding of insulin and IGF-receptors, but also can reduce IGFBPs, thus increasing biologically active IGF-1, and resulting in the development of hyperkeratosis and papillomatosis (54, 59).

As for acne vulgaris and hidradenitis suppurativa, mechanistic target of rapamycin complex 1 (mTORC1) is the key element that associates with insulin signaling and insulin resistance (61-64). Western diet and puberty, which are the two main risk factors of acne vulgaris, increase insulin/IGF-1 signaling and activate Akt. Active AKT stimulates mTORC1 activity by inhibiting TSC2. Ribosomal S6 kinase 1(S6K1), which is a downstream target of mTORC1, contributes to the serine/threonine phosphorylation of IRS, thus inducing insulin resistance by antagonizing tyrosine phosphorylation (65). mTORC1 also promotes lipid synthesis via activating the transcription factor sterol regulatory element binding protein 1 (SREBP-1) and inducing the expression of acetyl CoA carboxylase (ACC), which is the rate-limiting enzyme of fatty acid synthesis (66, 67). In acne vulgaris, overexpression of free fatty acids such as sapienic acid, oleic acid, and palmitic acid promotes propionibacterium acnes biofilm formation, stimulates T helper cell 17 (Th17)-driven inflammation, and disturbs follicular keratinization and barrier function (68). The role of mTORC1 in hidradenitis suppurativa is related to its function in cell proliferation, especially in sebaceous glands and keratinocytes. Taken together, Western diet and puberty, in the conditions of acne and hidradenitis suppurativa, increase insulin signaling and activate mTORC1. Increased mTORC1 activity then contributes to insulin resistance, which means acne vulgaris and hidradenitis suppurativa are not only skin diseases but also are more like systemic diseases.

A recent study has reported that the PI3K/Akt pathway is overexpressed in peripheral blood mononuclear cells (PBMCs) and keratinocytes of psoriasis patients (69, 70). Compared with normal skin and uninvolved psoriatic skin, the psoriatic skin has a repressed expression of FOXO1 (71). As described above, Akt phosphorylates FOXO and leads to its nuclear export, thus promoting cell proliferation by suppressing its function of activating cell cycle inhibitors (p27KIP1 and p21) and repressing cell cycle activators (cyclin D1/D2) (72). Thereby, overactivation of PI3K and Akt and the downregulation of FOXO contribute to proliferation of keratinocytes, which might be the potential role of PI3K/Akt signaling in psoriasis. Another downstream molecule of PI3K/Akt pathway, mTOR and its phosphorylation, have also been reported to be hyperactivated in lesional psoriatic skin. Buerger and his colleagues found that mTOR is activated throughout the whole epidermis of psoriatic skin, particularly in the basal layer, and the downstream molecules S6K-1 and ribosomal protein S6 are only activated in suprabasal layers in lesional skin (73). Their group also proposed a model where mTORC1 functions as a central switch between keratinocyte proliferation and differentiation (74). In the basal layer of healthy skin, the mTORC1 signaling pathway controls proliferation and blocks differentiation. When keratinocytes leave the basal layer, the mTORC1 pathway is switched off and this leads to the differentiation. Nevertheless, in the psoriatic condition, the mTORC1 signaling pathway is permanently switched on and results in epidermal hyperplasia by promoting hyperproliferation and inhibiting differentiation of keratinocytes (74, 75). Recent studies also revealed that inflammatory cytokines are associated with the activation of the mTORC1 signaling cascade. Patel and his colleagues found that tumor necrosis factor α (TNF- α) can activate mTOR and then stimulate IL-6, CXCL8, and VEGF secretion from both HaCat cells and primary human keratinocytes (76). Another study reported IL-17A-mediated inflammatory stimulation in keratinocytes activates PI3K/AKT/mTOR signaling and inhibits autophagy (70). Consistent with the previous studies, the Buerger laboratory found that inflammatory cytokines, such as IL-1β, IL-17A, and TNF-α, promote the mTOR pathway via PI3K signaling and lead to enhanced proliferation in psoriatic keratinocytes (77). Then, growth factors and relevant cytokines (IL-17A, TNF-a, and IL-1B) in psoriasis activate mTOR and promote keratinocyte hyperproliferation and inhibit differentiation, which might be the mechanism underlying the PI3K/Akt pathway and psoriasis (78). Therefore, we hypothesize that mTOR may function as a key target by connecting psoriasis and MetS through insulin resistance. mTOR causes the serine/threonine phosphorylation of IRS by activating S6K, and reduces its ability to be phosphorylated on tyrosine residues, which results in impaired insulin signaling and insulin resistance (52). Moreover, insulin resistance can exacerbate inactivation of the PI3K/AKT signaling pathway, which is characteristic of metabolic syndrome (79, 80).

Although the main function of the MAPK/Ras pathway was thought to be the regulation of mitogenesis, it has been reported that ERK1/2 can activate upstream MEK, reduce hepatic Akt phosphorylation, and contribute to insulin resistance (80). Furthermore, p-ERK1/2 has been found to be increased in psoriatic skin, which results in the abnormal proliferation and differentiation of keratinocytes (81, 82). ERK phosphorylation can also activate Th17 cells and then promote the secretion of inflammatory cytokines TNF- α , IL-6, and IL-17A, which positively correlate with psoriasis. A recent study reported that ERK inhibitor JSI287 alleviates imiquimod-induced psoriasis-like dermatitis by suppressing the ERK/IL-17 signaling pathway (83).

Chronic Inflammation

It is evident that MetS is a chronic inflammatory status with increased levels of proinflammatory cytokines, inflammatory biomarkers, and altered adipokines. Liver, intestine, and adipose tissue are the major three sites of the initiators of inflammation in MetS, of which, adipose tissue is vital (84). Because of nutritional excess in MetS, adipose tissue mass expands and adipocytes become hypertrophic, which can lead to decreased insulin sensitivity, hypoxia, altered autophagy, and apoptosis of adipocytes (85). The dysfunction of adipose tissue and impaired condition of adipocytes can also lead to overproduction of proinflammatory cytokines, such as interleukin 6 (IL-6), TNF- α , and C-reactive protein (CRP), thus resulting in macrophage infiltration (41, 42). Adipokines, which are mainly secreted by adipocytes, also play a significant role in MetS. In regard to the relationship between adipocytes and skin diseases, previous studies mostly focused on their role in regulating hair regeneration, wound healing, and skin aging (86, 87). Recently, the secretion of cytokines and adipokines by adipocytes has received increased attention, which might provide a novel angle in understanding the pathological and molecular connections between MetS and skin diseases (Figure 2).



mediators could migrate into the systemic circulation and the release of these proinflammatory cytokines results in the chronic systemic inflammation and MetS. The activation of NFkB signaling pathway is a commom pathway underlying skin diseases and MetS, in the canonical NFkB pathway, the dimer of p65 and p50 is in an inactive state by the inhibition of $I_{\kappa}B$ in cytoplasm. Upon stimulation by various cytokine receptors, IKKs are activated, leading to phosphorylate $I_{\kappa}B$, which results in their degradation and enables NF κ B translocate to the nucleus to induce target gene expression. The activation of NF κ B then produces a wide range of chemokines and cytokines, which leads to the formation of a feed forward loop, and establishs a chronic inflammatory environment.

Adipokines

Adipokines represent a group of hormones and cytokines secreted by adipose tissue, which are widely involved in a variety of biological processes, including energy homeostasis, inflammation, insulin resistance, and cell proliferation (88). Impaired adipose tissue function, such as adipose tissue inflammation, adipocyte hypertrophy, and ectopic fat accumulation, causes an adverse adipose secretion and contributes to metabolic and inflammatory diseases (87). Adipokines can be classified as pro-inflammatory or anti-inflammatory, depending on their effect on inflammation (89). Chemerin, leptin, resistin, and classical cytokines like TNF- α and IL-6, are pro-inflammatory adipokines. Anti-inflammatory adipokines include adiponectin, secreted frizzled-related protein 5 (SFRP5), and omentin-1.

Chemerin has been found to be involved in energy metabolism, adipogenesis, and inflammation. The role of chemerin in body metabolism depends on its role in regulating food intake, glucose homeostasis, and body weight. In adipose tissue, chemerin is involved in the processes of hyperplasia, angiogenesis, and inflammation by targeting adipocytes, endothelial cells, and immune cells, separately. In both mice and humans, chemerin levels are positively correlated with body mass index (BMI), inflammatory cytokines, and percentage of body fat (90, 91). Notably, the expression of chemerin is increased in psoriasis patients and is correlated to PASI (92). A recent study found that chemerin induces the inflammatory response in keratinocytes through reactive oxygen species (ROS)-sirtuin 1 (sirt1)-NF κ B signaling (93).

Leptin is mainly produced by adipocytes and is associated with waist circumference, insulin resistance, and MetS (94, 95). Moreover, leptin levels have been found to be higher in psoriasis, Behçet's syndrome, skin tags, and acanthosis nigricans (96–100). The role of leptin in psoriasis is clearly related to its proinflammatory actions. Leptin is reported to induce the secretion of IL-6, TNF- α , IL-1, IL-8, CXCL-1, CXCL-8, and IL-12 (101, 102). These proinflammatory mediators not only lead to insulin resistance and modulate lipid metabolism in MetS, but also contribute to the development of psoriasis. In Behçet's syndrome, apart from the role of leptin in inflammation, leptin also enhances the release of nitric oxide from endothelial cells and is involved in the pathophysiology of vascular lesions (97). In patients with skin tags, serum leptin is significantly upregulated and positive correlations are also found between serum leptin and insulin resistance calculated by homeostasis model assessment (HOMA-IR). Importantly, serum leptin and number of skin tags are significantly higher in patients with MetS compared to patients without MetS (98, 103), indicating that leptin may play a common pathologic role, although the underlying mechanism remains to be elucidated. An immunohistochemical study reported that the angiogenic effect and induction of cellular proliferation of leptin might contribute to the development of skin tags (104).

Resistin is another pro-inflammatory adipokine which is mainly produced by white adipose tissue in mice. In humans, resistin is mainly produced by cell populations such as macrophages, PBMCs, and bone marrow cells (105, 106).The previous data regarding correlation between resistin and MetS or MetS-components was inconsistent (107). However, it has been found that resistin plays an important role in inflammation by increasing the secretion of several inflammatory cytokines including TNF-α, CXCL8, IL-12, IL-1β, and IL-6 (108, 109). Interestingly, resistin has been shown to be associated with psoriasis, AD, AN, and Behçet disease (100, 110-112). Concerning the role of resistin in psoriasis, in addition to its effect on proinflammatory cytokine production, its connection with proprotein convertase subtilisin/kexin 9 (PCSK9) may represent another key mechanism. PCSK9 is a chaperone protein of lowdensity lipoprotein (LDL), which promotes the degradation of LDL receptors and, therefore, increases the concentration of circulating LDL. PCSK9 inhibitor is already approved to treat hyperlipoproteinemia (113, 114). Our previous study reported that suppressing PCSK9 inhibited the hyper-proliferation of keratinocytes and reduced the psoriasis-like inflammation via the NF κ B pathway (115). Interestingly, according to the structure analysis, PCSK9 C-terminal cysteine-rich domain (CRD) reveals structural homology with resistin (116). Moreover, resistin has also been shown to exhibit a positive correlation with PCSK9 levels by enhancing mRNA expression and protein stability (117, 118). As a result, resistin may act as an activator of PCSK9 in the pathogenesis of psoriasis. Regarding its role in AD, resistin is reported to increase the expression of monocyte chemoattractant protein 1 (MCP-1), vascular cell adhesion molecule 1 (VCAM-1), and intercellular adhesion molecule 1 (ICAM-1) in endothelial cells, which contributes to inflammation in AD (116). The expression of resistin was found to be upregulated in AN and Behçet disease, but further studies are still needed to reveal the underlying mechanisms (119, 120).

Adiponectin, as an anti-inflammatory adipokine, has three oligomeric isoforms: a low-molecular-weight trimer, a middle-molecular-weight hexamer, and a high-molecular-weight complex (121). High-molecular-weight adiponectin is reported to be the most biologically active of the three. The circulating levels of adiponectin are decreased in MetS, obesity, and type-2 diabetes mellitus (122). Furthermore, due to its association with severity and incidence of MetS, adiponectin is reported to function as a predictable marker for MetS (123, 124). The major

mechanism underlying the relationship between adiponectin and MetS may be the potent insulin-sensitizing effect of adiponectin. Adiponectin can also activate AMPK and PPAR-α pathways via binding to two receptors, including AdipoR1 and AdipoR2 (125). Growing data show that adiponectin is decreased in psoriasis, acne vulgaris, AN, chronic urticaria, and AD (99, 100, 126, 127). It has been suggested that adiponectin suppresses IL-17 synthesis through AdipoR1 and regulates skin inflammation; therefore, it might be a therapeutic target for psoriasis (128). The role of adiponectin in acne vulgaris may be not only related with its antiinflammatory effects but also associated through its function in inhibiting mTORC1 activity by activating AMP-activated protein kinase (126). In other words, decreased adiponectin levels in acne vulgaris could upregulate activity of the mTORC1 pathway and increase the synthesis of pro-inflammatory cytokines. Although it has been reported that adiponectin is reduced in AN, chronic urticaria, and AD patients, the exact pathogenic role is still not fully understood, which might due to the anti-inflammatory role of adiponectin.

Proinflammatory Cytokines Related Molecular Mechanisms

The "inflammatory skin march," first identified in psoriasis patients with systemic inflammatory condition, might be one of the main mechanisms underlying the connection between MetS and some skin diseases. As for psoriasis, activated myeloid dendritic cells produce IL-12 and IL-23 to promote the development of Th1, Th17, and Th22 cells in psoriasis patients, thus leading to the overproduction of proinflammatory cytokines in psoriatic lesions, including interferon-y (IFN- y), IL-1, IL-17, IL-6, IL-12, IL-22, IL-23, and TNF-α (129). These cutaneous inflammatory mediators can migrate into the systemic circulation, which is identified as "inflammatory skin march." The release of these proinflammatory cytokines can result in chronic systemic inflammation, which induces insulin resistance, obesity, hypertension, and MetS (130, 131). Moreover, atopic dermatitis is a T-cell-mediated disease, with the lesions also characterized by increased expression of cytokines produced by Th2, Th17, and Th22 cells (132). The "inflammatory skin march" is also the reason for the systemic inflammation in AD patients (133). Since the "inflammatory skin march" is reported in psoriasis and AD, it is reasonable to hypothesize that this mechanism could be the common reason for skin inflammation in global inflammatory-related diseases, such as MesS, obesity and, cardiovascular diseases.

Proinflammatory cytokines mediate their effects via binding to their own cytokine receptors and then activating several downstream pathways, including the JAK-STAT pathway, MAPK pathway, and NF κ B pathway, of which, the NF κ B pathway has been studied thoroughly. The NF κ B transcription factor family consists five members, NF κ B1 (p105/p50), NF κ B2 (p100/p52), RelA (p65), RelB, and c-Rel. The activation of NF κ B relies on two major signaling pathways, the canonical and non-canonical pathways, of which, proinflammatory cytokines are associated with the former one, and the non-canonical pathway is associated with differentiation and maturation of immune cells (134). In the canonical NF κ B signaling pathway, the dimer of p65 and

p50 is in an inactive state by the inhibition of IkB in cytoplasm. Upon stimulation by various cytokine receptors, such as TNF receptor superfamily members, IkB kinase (IKKs) are activated, leading to phosphorylate IkB, which results in their degradation, and enables $NF\kappa B$ translocate to the nucleus to induce the expression of downstream genes, including TNF-α, as well as IL-1, IL-6, and IL-12. The proinflammatory cytokines activate NFkB and then induce the expression of these cytokines, leading to the formation of a feed-forward loop. Recent studies have also shown that NFkB exhibits an anti-inflammatory role, directly inhibiting the expression of inflammatory cytokines by inducing the apoptosis of leukocytes and by upregulating the expression of anti-inflammatory cytokines such as IL-10 (135). Apart from the positive loop, activated NFkB in the nucleus can promote the expression of I κ B- α , which binds to and inactivates NF κ B in both cytoplasma and nucleus, resulting in a negative feedback loop of the NF κ B pathway (136).

The association between NF κ B and metabolic syndrome has been studied extensively in obesity and insulin resistance. In obesity, proinflammatory cytokines, free fatty acids, and microbiota-derived LPS can activate IKK β by directly binding to cytokine receptors and Toll-like receptor 4 in adipocytes, macrophage and muscle (137, 138). The activated IKK β not only phosphorylates the serine site of IRS, but also activates mTOR and S6K1 by suppressing the TSC1/2 in adipocytes and hepatocytes, both of which contribute to the induction of insulin resistance (139). Besides the inhibition of insulin signaling, activated IKK β can induce the translocation of NF κ B to nuclear by phosphorylating I κ B. The activation of NF κ B then produces a wide range of chemokines and cytokines in most metabolic tissues, leading to chronic inflammation (138).

In psoriasis, numerous effector cells including keratinocytes, Th17 cells, and dendritic cells activated by TNF-α, plasmin and TLRs could produce cytokines and chemokines via the NFkB pathway, which leads to psoriatic phenotypes (140). Moreover, our recent studies reported that metabolicassociated genes PCSK9 and nuclear receptor interacting protein 1 (NRIP1) both play important roles in psoriasis via the NFkB pathway (115, 141). It has been reported that PCSK9 silencing can suppress atherosclerosis by inhibiting the TLR4/NFkB signaling pathway (142). Also, suppressing PCSK9 can inhibit the NFkB mediated inflammatory response in macrophages (143). In our study, we found that suppressing PCSK9 inhibited the hyper-proliferation of keratinocytes and reduced the RelA/p65-mediated inflammatory reaction induced by imiquimod (115). Likewise, NRIP1, acting as co-activator or co-repressor for several nuclear receptors and transcriptional factors, was also reported to interact with NFkB, thus participating in the inflammatory and metabolic processes. NRIP1 functions as a co-activator in macrophages for NFkB subunit RelA, and cAMP-responsive element binding protein (CREB)-binding protein (CBP), and promotes the TLR-induced production of proinflammatory cytokines such as TNF- α , IL-1 β , and IL-6 (144). NF κ B (RelA) can inversely resolute the inflammatory response by targeting and degrading NRIP1 via recruiting the SCF (Skp, culin, F-box-containing) E3 ligase complex (145).Our laboratory revealed that suppression of NRIP1 in CD4+ T cells that were isolated from psoriasis patients could downregulate the expression of RelA/p65 and decrease the secretion of IL-17, thus inhibiting the inflammation in psoriasis (141). Importantly, together, PCSK9 and NRIP1 may be two potential therapeutic targets for both psoriasis and metabolic syndrome by regulating the NF κ B-associated chronic inflammatory status.

Adipose-Derived Mesenchymal Stem Cells (ADMSCs) Therapy in MetS and Skin Diseases

To date, applications of ADMSCs have been studied in a variety of skin diseases, such as alopecia, scars, wound healing, and psoriasis. Although the therapeutic mechanisms of ADMSCs in these diseases remain vague, it has been reported that the reduction of inflammation, the increase of immune cell apoptosis, the promotion of collagen synthesis, and the improvement of angiogenesis might be possibilities (146, 147). It is well known that ADMSCs have the properties of immunomodulatory, which can inhibit inflammation by suppressing the production of proinflammatory cytokines, such as IL-6, IL-17, and TNF- α (148). Moreover, because of the multipotent function, anabolic activity, and immunomodulatory effect, ADMSCs are reported to be a promising therapy for MetS (149). According to the study of MSC transplantation in obese mice, the MSC-based therapies not only reduce obesityassociated metabolic syndromes including non-alcoholic fatty liver disease, non-alcoholic steatohepatitis, glucose intolerance, and inflammation, but also ameliorate high-fat diet-induced obesity and hyperlipidemia (150). A systematic review showed that ADMSCs therapy provides positive effects on body weight, lipid profiles, glucose metabolism homeostasis, non-alcoholic fatty liver disease, and systemic inflammation, which reflects that ADMSCs therapy could be a potential and promising strategy in obesity (151). Furthermore, ADMSCs therapy has been reported in both an imiquimod (IMQ)-induced psoriasis mouse model and in psoriasis patients. In the imiquimodinduced psoriasis mouse model, the intradermal administration of ADMSCs inhibits IL-17A and TNF-a, and suppresses the IMQ-induced inflammation (152). In psoriasis patients (five patients from three case reports), those who received the intravenous injection of ADMSCs, all showed great improvement of psoriatic erythema, scaling, and induration by significant decrease of PASI scores with no severe adverse events (153-155). Considering the chronic inflammatory status in both MetS and psoriasis, along with the results of all these clinical trials, ADMSCs therapy shows great potential to have clinical benefits for patients with combined psoriasis and MetS by reducing inflammation through an immunomodulatory cascade of events (156). Nevertheless, a recent study indicated that mesenchymal stem cells isolated from MetS patients have abnormal apoptosis, autophagy, and mitochondria function, which may limit their therapeutic potential (157). Moreover, mesenchymal stem cells isolated from obese patients have an altered secretome profile such as increased IL-6 and decreased adiponectin (158). Thus, improving the function of the isolated mesenchymal stem cells by genetic treatment may provide a new approach for better clinical outcomes. Several studies have reported that the function of ADMSC can be improved by genetic methods, such as Toll-like receptors depletion, and chemical treatment such as melatonin and transforming growth factor- β (TGF- β) pretreatment (159–161). However, more clinical studies are needed to further evaluate the safety and efficacy of ADMSCs therapy.

CONCLUSION

Evidence shows a strong connection between MetS and skin diseases, indicating that some skin diseases may be the cutaneous manifestations of systemic disorders. In this review, we have shown the epidemiological evidence for the connection between MetS and skin diseases, including psoriasis, acne vulgaris, hidradenitis suppurativa, androgenetic alopecia, acanthosis nigricans, and atopic dermatitis. Although the exact relationship between MetS and skin diseases is still unclear, insulin signaling, insulin resistance, and chronic inflammation are believed to contribute. Understanding the mechanisms underlying MetS and skin diseases will help to improve clinical outcomes and guide the development of new therapeutic treatments.

REFERENCES

- Favaro L, Masini F, Maffei ML, Botti G. [Syndrome X]. Recenti Prog Med. (1989) 80:281–5.
- Rochlani Y, Pothineni NV, Kovelamudi S, Mehta JL. Metabolic syndrome: pathophysiology, management, and modulation by natural compounds. *Ther Adv Cardiovasc Dis.* (2017) 11:215–25. doi: 10.1177/17539447177 11379
- Samson SL, Garber AJ. Metabolic syndrome. Endocrinol Metab Clin North Am. (2014) 43:1–23. doi: 10.1016/j.ecl.2013.09.009
- Karadag AS, Lavery MJ. Skin and the metabolic syndrome. *Clin Dermatol.* (2018) 36:1–2. doi: 10.1016/j.clindermatol.2017.09.001
- Gui XY, Yu XL, Jin HZ, Zuo YG, Wu C. Prevalence of metabolic syndrome in Chinese psoriasis patients: a hospital-based cross-sectional study. J Diabetes Investig. (2018) 9:39–43. doi: 10.1111/jdi.12663
- Michalek IM, Loring B, John SM. A systematic review of worldwide epidemiology of psoriasis. J Eur Acad Dermatol Venereol. (2017) 31:205– 12. doi: 10.1111/jdv.13854
- Singh S, Young P, Armstrong AW. Relationship between psoriasis and metabolic syndrome: a systematic review. *G Ital Dermatol Venereol.* (2016) 151:663–77.
- Rodriguez-Zuniga MJM, Garcia-Perdomo HA. Systematic review and metaanalysis of the association between psoriasis and metabolic syndrome. *J Am Acad Dermatol.* (2017) 77:657–66 e658. doi: 10.1016/j.jaad.2017.0 4.1133
- Singh S, Young P, Armstrong AW. An update on psoriasis and metabolic syndrome: a meta-analysis of observational studies. *PLoS ONE*. (2017) 12:e0181039. doi: 10.1371/journal.pone.0181039
- Fernandez-Armenteros JM, Gomez-Arbones X, Buti-Soler M, Betriu-Bars A, Sanmartin-Novell V, Ortega-Bravo M, et al. Psoriasis, metabolic syndrome and cardiovascular risk factors. A population-based study. J Eur Acad Dermatol Venereol. (2019) 33:128–35. doi: 10.1111/jdv.15159
- Nagpal M, De D, Handa S, Pal A, Sachdeva N. Insulin resistance and metabolic syndrome in young men with acne. *JAMA Dermatol.* (2016) 152:399–404. doi: 10.1001/jamadermatol.2015.4499
- 12. Emiroglu N, Cengiz FP, Kemeriz F. Insulin resistance in severe acne vulgaris. Postepy Dermatol Alergol. (2015) 32:281–5. doi: 10.5114/pdia.2015.53047

AUTHOR CONTRIBUTIONS

YH and NL collected data. YH, YZ, and RY wrote the manuscript. RY, AB, and MC revised the manuscript.

FUNDING

This work was funded by U.S. NIH grant AG046432, internal research seed grants of Southern Illinois University School of Medicine, CAMS Innovation Fund for Medical Sciences (CIFMS-2017-I2M-1-017) and China Scholarship Council (No. 201806210434). U.S. NIH grant AG046432 and internal research seed grants of Southern Illinois University School of Medicine to Dr. RY supported collecting data, reviewing papers and writing the manuscript. CAMS Innovation Fund for Medical Sciences (CIFMS-2017-I2M-1-017) and China Scholarship Council (No. 201806210434) to YH supported collecting data, reviewing papers and writing the manuscript.

ACKNOWLEDGMENTS

Lisa Hensley kindly helped with editing the manuscript.

- Snast I, Dalal A, Twig G, Astman N, Kedem R, Levin D, et al. Acne and obesity: a nationwide study of 600,404 adolescents. J Am Acad Dermatol. (2019) 81:723–9. doi: 10.1016/j.jaad.2019.04.009
- Cohen PG. The hypogonadal-obesity cycle: role of aromatase in modulating the testosterone-estradiol shunt–a major factor in the genesis of morbid obesity. *Med Hypotheses*. (1999) 52:49–51. doi: 10.1054/mehy.1997.0624
- Ju Q, Tao T, Hu T, Karadag AS, Al-Khuzaei S, Chen W. Sex hormones and acne. *Clin Dermatol.* (2017) 35:130– 7. doi: 10.1016/j.clindermatol.2016.10.004
- Bechis SK, Otsetov AG, Ge R, Wang Z, Vangel MG, Wu CL, et al. Age and obesity promote methylation and suppression of 5alpha-reductase 2: implications for personalized therapy of benign prostatic hyperplasia. *J Urol.* (2015) 194:1031–7. doi: 10.1016/j.juro.2015.04.079
- Saunte DML, Jemec GBE. Hidradenitis suppurativa: advances in diagnosis and treatment. JAMA. (2017) 318:2019–32. doi: 10.1001/jama.2017.16691
- Shalom G, Freud T, Harman-Boehm I, Polishchuk I, Cohen AD. Hidradenitis suppurativa and metabolic syndrome: a comparative cross-sectional study of 3207 patients. Br J Dermatol. (2015) 173:464–70. doi: 10.1111/bjd.13777
- Miller IM, Ellervik C, Vinding GR, Zarchi K, Ibler KS, Knudsen KM, et al. Association of metabolic syndrome and hidradenitis suppurativa. JAMA Dermatol. (2014) 150:1273–80. doi: 10.1001/jamadermatol.2014.1165
- Gold DA, Reeder VJ, Mahan MG, Hamzavi IH. The prevalence of metabolic syndrome in patients with hidradenitis suppurativa. J Am Acad Dermatol. (2014) 70:699–703. doi: 10.1016/j.jaad.2013.11.014
- Lolli F, Pallotti F, Rossi A, Fortuna MC, Caro G, Lenzi A, et al. Androgenetic alopecia: a review. *Endocrine*. (2017) 57:9–17. doi: 10.1007/s12020-017-1280-y
- Gopinath H, Upadya GM. Metabolic syndrome in androgenic alopecia. *Indian J Dermatol Venereol Leprol.* (2016) 82:404– 8. doi: 10.4103/0378-6323.174421
- 23. Agamia NF, Abou Youssif T, El-Hadidy A, El-Abd A. Benign prostatic hyperplasia, metabolic syndrome and androgenic alopecia: is there a possible relationship? *Arab J Urol.* (2016) 14:157–62. doi: 10.1016/j.aju.2016.01.003
- 24. Ertas R, Orscelik O, Kartal D, Dogan A, Ertas SK, Aydogdu EG, et al. Androgenetic alopecia as an indicator of metabolic syndrome and cardiovascular risk. *Blood Press.* (2016) 25:141–8. doi: 10.3109/08037051.2015.1111021

- Su LH, Chen TH. Association of androgenetic alopecia with metabolic syndrome in men: a community-based survey. Br J Dermatol. (2010) 163:371–7. doi: 10.1111/j.1365-2133.2010.09816.x
- Dharam Kumar KC, Kishan Kumar YH, Neladimmanahally V. Association of androgenetic alopecia with metabolic syndrome: a case-control study on 100 patients in a tertiary care hospital in South India. *Indian J Endocrinol Metab.* (2018) 22:196–9. doi: 10.4103/ijem.IJEM_650_17
- Ozbas Gok S, Akin Belli A, Dervis E. Is there really relationship between androgenetic alopecia and metabolic syndrome? *Dermatol Res Pract.* (2015) 2015;980310. doi: 10.1155/2015/980310
- Bakry OA, Shoeib MA, El Shafiee MK, Hassan A. Androgenetic alopecia, metabolic syndrome, and insulin resistance: Is there any association? A case-control study. *Indian Dermatol Online J.* (2014) 5:276– 81. doi: 10.4103/2229-5178.137776
- El Sayed MH, Abdallah MA, Aly DG, Khater NH. Association of metabolic syndrome with female pattern hair loss in women: a case-control study. *Int J Dermatol.* (2016) 55:1131–7. doi: 10.1111/ijd.13303
- Karadag AS, You Y, Danarti R, Al-Khuzaei S, Chen W. Acanthosis nigricans and the metabolic syndrome. *Clin Dermatol.* (2018) 36:48– 53. doi: 10.1016/j.clindermatol.2017.09.008
- Ayaz T, Baydur Sahin S, Sahin OZ. Relation of Acanthosis nigricans to metabolic syndrome in overweight and obese women. *Metab Syndr Relat Disord*. (2014) 12:320–3. doi: 10.1089/met.2013.0145
- Velazquez-Bautista M, Lopez-Sandoval JJ, Gonzalez-Hita M, Vazquez-Valls E, Cabrera-Valencia IZ, Torres-Mendoza BM. Association of metabolic syndrome with low birth weight, intake of high-calorie diets and acanthosis nigricans in children and adolescents with overweight and obesity. *Endocrinol Diabetes Nutr.* (2017) 64:11–7. doi: 10.1016/j.endien.2016. 09.002
- Palhares H, Zaidan PC, Dib FCM, Silva APD, Resende DCS, Borges MF. Association between acanthosis nigricans and other cardiometabolic risk factors in children and adolescents with overweight and obesity. *Rev Paul Pediatr.* (2018) 36:301–8. doi: 10.1590/1984-0462/;2018;36;3;00017
- Panda S, Das A, Lahiri K, Chatterjee M, Padhi T, Rathi S, et al. Facial acanthosis nigricans: a morphological marker of metabolic syndrome. *Indian J Dermatol.* (2017) 62:591–7. doi: 10.4103/ijd.JJD_545_17
- Nutten S. Atopic dermatitis: global epidemiology and risk factors. Ann Nutr Metab. (2015) 66 (Suppl. 1):8–16. doi: 10.1159/000370220
- Lee JH, Jung HM, Han KD, Lee SH, Lee JY, Park YG, et al. Association between metabolic syndrome and atopic dermatitis in korean adults. *Acta Derm Venereol.* (2017) 97:77–80. doi: 10.2340/00015555-2441
- Radtke MA, Schafer I, Glaeske G, Jacobi A, Augustin M. Prevalence and comorbidities in adults with psoriasis compared to atopic eczema. *J Eur Acad Dermatol Venereol.* (2017) 31:151–7. doi: 10.1111/jdv.13813
- Ali Z, Ulrik CS, Agner T, Thomsen SF. Association between atopic dermatitis and the metabolic syndrome: a systematic review. *Dermatology*. (2018) 234:79–85. doi: 10.1159/000491593
- Silverberg JI, Kleiman E, Lev-Tov H, Silverberg NB, Durkin HG, Joks R, et al. Association between obesity and atopic dermatitis in childhood: a case-control study. J Allergy Clin Immunol. (2011) 127:1180–6 e1181. doi: 10.1016/j.jaci.2011.01.063
- Zhang A, Silverberg JI. Association of atopic dermatitis with being overweight and obese: a systematic review and metaanalysis. J Am Acad Dermatol. (2015) 72:606–16 e604. doi: 10.1016/j.jaad.2014.12.013
- McCracken E, Monaghan M, Sreenivasan S. Pathophysiology of the metabolic syndrome. *Clin Dermatol.* (2018) 36:14– 20. doi: 10.1016/j.clindermatol.2017.09.004
- 42. Eckel RH, Grundy SM, Zimmet PZ. The metabolic syndrome. *Lancet*. (2005) 365:1415–28. doi: 10.1016/S0140-6736(05)66378-7
- Roth MM, Leader N, Kroumpouzos G. Gynecologic and andrologic dermatology and the metabolic syndrome. *Clin Dermatol.* (2018) 36:72– 80. doi: 10.1016/j.clindermatol.2017.09.013
- Beale EG. Insulin signaling and insulin resistance. J Investig Med. (2013) 61:11-4. doi: 10.2310/JIM.0b013e3182746f95
- Engelman JA, Luo J, Cantley LC. The evolution of phosphatidylinositol 3kinases as regulators of growth and metabolism. *Nat Rev Genet.* (2006) 7:606–19. doi: 10.1038/nrg1879

- Birkenkamp KU, Coffer PJ. Regulation of cell survival and proliferation by the FOXO (Forkhead box, class O) subfamily of Forkhead transcription factors. *Biochem Soc Trans.* (2003) 31(Pt 1):292–7. doi: 10.1042/bst0310292
- Tia N, Singh AK, Pandey P, Azad CS, Chaudhary P, Gambhir IS. Role of Forkhead Box O (FOXO) transcription factor in aging and diseases. *Gene.* (2018) 648:97–105. doi: 10.1016/j.gene.2018.01.051
- Tzivion G, Dobson M, Ramakrishnan G. FoxO transcription factors; Regulation by AKT and 14-3-3 proteins. *Biochim Biophys Acta.* (2011) 1813:1938–45. doi: 10.1016/j.bbamcr.2011.06.002
- Taniguchi CM, Emanuelli B, Kahn CR. Critical nodes in signalling pathways: insights into insulin action. Nat Rev Mol Cell Biol. (2006) 7:85– 96. doi: 10.1038/nrm1837
- Mebratu Y, Tesfaigzi Y. How ERK1/2 activation controls cell proliferation and cell death: Is subcellular localization the answer? *Cell Cycle.* (2009) 8:1168–75. doi: 10.4161/cc.8.8.8147
- Petersen MC, Shulman GI. Mechanisms of insulin action and insulin resistance. *Physiol Rev.* (2018) 98:2133– 223. doi: 10.1152/physrev.00063.2017
- Lann D, LeRoith D. Insulin resistance as the underlying cause for the metabolic syndrome. *Med Clin North Am.* (2007) 91:1063–77, viii. doi: 10.1016/j.mcna.2007.06.012
- Yaribeygi H, Farrokhi FR, Butler AE, Sahebkar A. Insulin resistance: review of the underlying molecular mechanisms. *J Cell Physiol.* (2019) 234:8152– 61. doi: 10.1002/jcp.27603
- Cruz PD Jr, Hud JA Jr. Excess insulin binding to insulin-like growth factor receptors: proposed mechanism for acanthosis nigricans. J Invest Dermatol. (1992) 98(Suppl. 6):82S-5S. doi: 10.1111/1523-1747.ep12462293
- 55. Nam SY, Lee EJ, Kim KR, Cha BS, Song YD, Lim SK, et al. Effect of obesity on total and free insulin-like growth factor (IGF)-1, and their relationship to IGF-binding protein (BP)-1, IGFBP-2, IGFBP-3, insulin, and growth hormone. *Int J Obes Relat Metab Disord.* (1997) 21:355– 9. doi: 10.1038/sj.ijo.0800412
- Baxter RC. IGF binding proteins in cancer: mechanistic and clinical insights. Nat Rev Cancer. (2014) 14:329–41. doi: 10.1038/nrc3720
- Scheinfeld NS. Obesity and dermatology. Clin Dermatol. (2004) 22:303– 9. doi: 10.1016/j.clindermatol.2004.01.001
- Marzec KA, Baxter RC, Martin JL. Targeting insulin-like growth factor binding protein-3 signaling in triple-negative breast cancer. *Biomed Res Int.* (2015) 2015:638526. doi: 10.1155/2015/638526
- Phiske MM. An approach to acanthosis nigricans. *Indian Dermatol Online J.* (2014) 5:239–49. doi: 10.4103/2229-5178.137765
- Sciacca L, Cassarino MF, Genua M, Pandini G, Le Moli R, Squatrito S, et al. Insulin analogues differently activate insulin receptor isoforms and post-receptor signalling. *Diabetologia*. (2010) 53:1743–53. doi: 10.1007/s00125-010-1760-6
- Monfrecola G, Lembo S, Caiazzo G, De Vita V, Di Caprio R, Balato A, et al. Mechanistic target of rapamycin (mTOR) expression is increased in acne patients' skin. *Exp Dermatol.* (2016) 25:153–5. doi: 10.1111/exd.12885
- 62. Agamia NF, Abdallah DM, Sorour O, Mourad B, Younan DN. Skin expression of mammalian target of rapamycin and forkhead box transcription factor O1, and serum insulin-like growth factor-1 in patients with acne vulgaris and their relationship with diet. *Br J Dermatol.* (2016) 174:1299–307. doi: 10.1111/bjd.14409
- Monfrecola G, Balato A, Caiazzo G, De Vita V, Di Caprio R, Donnarumma M, et al. Mammalian target of rapamycin, insulin resistance and hidradenitis suppurativa: a possible metabolic loop. *J Eur Acad Dermatol Venereol.* (2016) 30:1631–3. doi: 10.1111/jdv.13233
- Marasca C, Balato A, Annunziata MC, Cacciapuoti S, Fabbrocini G. Insulin resistance, mTOR and hidradenitis suppurativa. J Eur Acad Dermatol Venereol. (2018) 33:e106–7. doi: 10.1111/jdv.15271
- Blagosklonny MV. TOR-centric view on insulin resistance and diabetic complications: perspective for endocrinologists and gerontologists. *Cell Death Dis.* (2013) 4:e964. doi: 10.1038/cddis.2013.506
- Bakan I, Laplante M. Connecting mTORC1 signaling to SREBP-1 activation. Curr Opin Lipidol. (2012) 23:226–34. doi: 10.1097/MOL.0b013e328352dd03
- 67. Sun Y, He W, Luo M, Zhou Y, Chang G, Ren W, et al. SREBP1 regulates tumorigenesis and prognosis of pancreatic

cancer through targeting lipid metabolism. *Tumour Biol.* (2015) 36:4133-41. doi: 10.1007/s13277-015-3047-5

- Melnik BC. Acne vulgaris: the metabolic syndrome of the pilosebaceous follicle. *Clin Dermatol.* (2018) 36:29– 40. doi: 10.1016/j.clindermatol.2017.09.006
- Ochaion A, Bar-Yehuda S, Cohen S, Barer F, Patoka R, Amital H, et al. The anti-inflammatory target A(3) adenosine receptor is over-expressed in rheumatoid arthritis, psoriasis and Crohn's disease. *Cell Immunol.* (2009) 258:115–22. doi: 10.1016/j.cellimm.2009.03.020
- Varshney P, Saini N. PI3K/AKT/mTOR activation and autophagy inhibition plays a key role in increased cholesterol during IL-17A mediated inflammatory response in psoriasis. *Biochim Biophys Acta Mol Basis Dis.* (2018) 1864(5 Pt A):1795–803. doi: 10.1016/j.bbadis.2018.02.003
- Liu Y, Luo W, Chen S. Comparison of gene expression profiles reveals aberrant expression of FOXO1, Aurora A/B and EZH2 in lesional psoriatic skins. *Mol Biol Rep.* (2011) 38:4219–24. doi: 10.1007/s11033-010-0544-x
- Greer EL, Brunet A. FOXO transcription factors at the interface between longevity and tumor suppression. Oncogene. (2005) 24:7410– 25. doi: 10.1038/sj.onc.1209086
- 73. Buerger C, Malisiewicz B, Eiser A, Hardt K, Boehncke WH. Mammalian target of rapamycin and its downstream signalling components are activated in psoriatic skin. *Br J Dermatol.* (2013) 169:156–9. doi: 10.1111/bjd.12271
- Buerger C. Epidermal mTORC1 signaling contributes to the pathogenesis of psoriasis and could serve as a therapeutic target. *Front Immunol.* (2018) 9:2786. doi: 10.3389/fimmu.2018.02786
- 75. Burger C, Shirsath N, Lang V, Diehl S, Kaufmann R, Weigert A, et al. Blocking mTOR signalling with rapamycin ameliorates imiquimod-induced psoriasis in mice. Acta Derm Venereol. (2017) 97:1087–94. doi: 10.2340/00015555-2724
- Patel AB, Tsilioni I, Weng Z, Theoharides TC. TNF stimulates IL-6, CXCL8 and VEGF secretion from human keratinocytes via activation of mTOR, inhibited by tetramethoxyluteolin. *Exp Dermatol.* (2018) 27:135– 43. doi: 10.1111/exd.13461
- Buerger C, Shirsath N, Lang V, Berard A, Diehl S, Kaufmann R, et al. Inflammation dependent mTORC1 signaling interferes with the switch from keratinocyte proliferation to differentiation. *PLoS ONE.* (2017) 12:e0180853. doi: 10.1371/journal.pone.0180853
- Zhang M, Zhang X. The role of PI3K/AKT/FOXO signaling in psoriasis. Arch Dermatol Res. (2019) 311:83–91. doi: 10.1007/s00403-018-1879-8
- Huang X, Liu G, Guo J, Su Z. The PI3K/AKT pathway in obesity and type 2 diabetes. *Int J Biol Sci.* (2018) 14:1483–96. doi: 10.7150/ijbs.27173
- Guo S. Insulin signaling, resistance, and the metabolic syndrome: insights from mouse models into disease mechanisms. *J Endocrinol.* (2014) 220:T1– 23. doi: 10.1530/JOE-13-0327
- Yu XJ, Li CY, Dai HY, Cai DX, Wang KY, Xu YH, et al. Expression and localization of the activated mitogen-activated protein kinase in lesional psoriatic skin. *Exp Mol Pathol.* (2007) 83:413–8. doi: 10.1016/j.yexmp.2007.05.002
- Johansen C, Kragballe K, Westergaard M, Henningsen J, Kristiansen K, Iversen L. The mitogen-activated protein kinases p38 and ERK1/2 are increased in lesional psoriatic skin. *Br J Dermatol.* (2005) 152:37–42. doi: 10.1111/j.1365-2133.2004.06304.x
- Huang X, Yu P, Liu M, Deng Y, Dong Y, Liu Q, et al. ERK inhibitor JSI287 alleviates imiquimod-induced mice skin lesions by ERK/IL-17 signaling pathway. *Int Immunopharmacol.* (2019) 66:236– 41. doi: 10.1016/j.intimp.2018.11.031
- Mendrick DL, Diehl AM, Topor LS, Dietert RR, Will Y, La Merrill MA, et al. Metabolic syndrome and associated diseases: from the bench to the clinic. *Toxicol Sci.* (2018) 162:36–42. doi: 10.1093/toxsci/kfx233
- Kloting N, Bluher M. Adipocyte dysfunction, inflammation and metabolic syndrome. *Rev Endocr Metab Disord.* (2014) 15:277–87. doi: 10.1007/s11154-014-9301-0
- Horsley V, Watt F. Repeal and replace: adipocyte regeneration in wound repair. Cell Stem Cell. (2017) 20:424–6. doi: 10.1016/j.stem.2017.03.015
- Rivera-Gonzalez G, Shook B, Horsley V. Adipocytes in skin health and disease. *Cold Spring Harb Perspect Med.* (2014) 4:a015271. doi: 10.1101/cshperspect.a015271

- Trayhurn P, Wood IS. Adipokines: inflammation and the pleiotropic role of white adipose tissue. Br J Nutr. (2004) 92:347–55. doi: 10.1079/BJN200 41213
- Kwon H, Pessin JE. Adipokines mediate inflammation and insulin resistance. Front Endocrinol. (2013) 4:71. doi: 10.3389/fendo.2013.00071
- Ernst MC, Issa M, Goralski KB, Sinal CJ. Chemerin exacerbates glucose intolerance in mouse models of obesity and diabetes. *Endocrinology*. (2010) 151:1998–2007. doi: 10.1210/en.2009-1098
- Chakaroun R, Raschpichler M, Kloting N, Oberbach A, Flehmig G, Kern M, et al. Effects of weight loss and exercise on chemerin serum concentrations and adipose tissue expression in human obesity. *Metabolism*. (2012) 61:706– 14. doi: 10.1016/j.metabol.2011.10.008
- Coban M, Tasli L, Turgut S, Ozkan S, Tunc Ata M, Akin F. Association of adipokines, insulin resistance, hypertension and dyslipidemia in patients with psoriasis vulgaris. *Ann Dermatol.* (2016) 28:74–9. doi: 10.5021/ad.2016.28.1.74
- Wang Y, Huo J, Zhang D, Hu G, Zhang Y. Chemerin/ChemR23 axis triggers an inflammatory response in keratinocytes through ROS-sirt1-NFkappaB signaling. J Cell Biochem. (2019) 120:6459–70. doi: 10.1002/jcb. 27936
- 94. Gannage-Yared MH, Khalife S, Semaan M, Fares F, Jambart S, Halaby G. Serum adiponectin and leptin levels in relation to the metabolic syndrome, androgenic profile and somatotropic axis in healthy non-diabetic elderly men. *Eur J Endocrinol.* (2006) 155:167–76. doi: 10.1530/eje.1.02175
- 95. Franks PW, Brage S, Luan J, Ekelund U, Rahman M, Farooqi IS, et al. Leptin predicts a worsening of the features of the metabolic syndrome independently of obesity. *Obes Res.* (2005) 13:1476–84. doi: 10.1038/oby.2005.178
- Aly DG, Abdallah IY, Hanafy NS, Elsaie ML, Hafiz NA. Elevated serum leptin levels in nonobese patients with psoriasis. J Drugs Dermatol. (2013) 12:e25–9.
- Evereklioglu C, Inaloz HS, Kirtak N, Doganay S, Bulbul M, Ozerol E, et al. Serum leptin concentration is increased in patients with Behcet's syndrome and is correlated with disease activity. *Br J Dermatol.* (2002) 147:331–6. doi: 10.1046/j.1365-2133.2002.04703.x
- 98. Shaheen MA, Abdel Fattah NS, Sayed YA, Saad AA. Assessment of serum leptin, insulin resistance and metabolic syndrome in patients with skin tags. J Eur Acad Dermatol Venereol. (2012) 26:1552–7. doi: 10.1111/j.1468-3083.2011.04401.x
- 99. Han B, Wu WH, Bae JM, Son SJ, Lee JH, Han TY. Serum leptin and adiponectin levels in atopic dermatitis (AD) and their relation to disease severity. J Am Acad Dermatol. (2016) 75:629–31. doi: 10.1016/j.jaad.2016.04.036
- 100. Atwa M, Emara A, Balata M, Youssef N, Bayoumy N, Sherif A, et al. Serum leptin, adiponectin, and resistin among adult patients with acanthosis nigricans: correlations with insulin resistance and risk factors for cardiovascular disease. *Int J Dermatol.* (2014) 53:e410–20. doi: 10.1111/ijd.12340
- 101. Ommen P, Stjernholm T, Kragstrup T, Raaby L, Johansen C, Stenderup K, et al. The role of leptin in psoriasis comprises a proinflammatory response by the dermal fibroblast. Br J Dermatol. (2016) 174:187–90. doi: 10.1111/bjd.13969
- 102. Johnston A, Arnadottir S, Gudjonsson JE, Aphale A, Sigmarsdottir AA, Gunnarsson SI, et al. Obesity in psoriasis: leptin and resistin as mediators of cutaneous inflammation. Br J Dermatol. (2008) 159:342– 50. doi: 10.1111/j.1365-2133.2008.08655.x
- Putra IB, Siregar R, Jusuf NK, Ginting O, Nurhayati R. Correlation between serum leptin level with type and number of lesion skin tag. *Open Access Maced J Med Sci.* (2019) 7:53–55. doi: 10.3889/oamjms.2019.020
- 104. Seleit I, Bakry OA, Samaka RM, Samy M. Immunohistochemical evaluation of leptin role in skin tags. Ultrastruct Pathol. (2015) 39:235– 44. doi: 10.3109/01913123.2015.1006744
- 105. Patel L, Buckels AC, Kinghorn JJ, Murdock PR, Holbrook JD, Plumpton C, et al. Resistin is expressed in human macrophages and directly regulated by PPAR gamma activators. *Biochem Biophys Res Commun.* (2003) 300:472– 6. doi: 10.1016/S0006-291X(02)02841-3
- 106. Jamaluddin MS, Weakley SM, Yao Q, Chen C. Resistin: functional roles and therapeutic considerations for cardiovascular disease. *Br J Pharmacol.* (2012) 165:622–32. doi: 10.1111/j.1476-5381.2011.01369.x

- Mostafazadeh M, Haiaty S, Rastqar A, Keshvari M. Correlation between resistin level and metabolic syndrome component: a review. *Horm Metab Res.* (2018) 50:521–36. doi: 10.1055/a-0637-1975
- 108. Gao F, Si F, Feng S, Yi Q, Liu R. Resistin enhances inflammatory cytokine production in coronary artery tissues by activating the NF-κB signaling %J BioMed Research International. (2016) 2016:8. doi: 10.1155/2016/3296437
- Zuniga MC, Raghuraman G, Hitchner E, Weyand C, Robinson W, Zhou W. PKC-epsilon and TLR4 synergistically regulate resistin-mediated inflammation in human macrophages. *Atherosclerosis.* (2017) 259:51– 9. doi: 10.1016/j.atherosclerosis.2017.02.021
- Kyriakou A, Patsatsi A, Sotiriadis D, Goulis DG. Serum leptin, resistin, and adiponectin concentrations in psoriasis: a meta-analysis of observational studies. *Dermatology*. (2017) 233:378–89. doi: 10.1159/000481882
- Banihani SA, Abu-Alia KF, Khabour OF, Alzoubi KH. Association between resistin gene polymorphisms and atopic dermatitis. *Biomolecules*. (2018) 8:17. doi: 10.3390/biom8020017
- Lee YH, Song GG. Association of circulating resistin, leptin, adiponectin and visfatin levels with Behcet disease: a meta-analysis. *Clin Exp Dermatol.* (2018) 43:536–45. doi: 10.1111/ced.13383
- 113. Lavecchia A, Cerchia C. Recent advances in developing PCSK9 inhibitors for lipid-lowering therapy. *Future Med Chem.* (2019) 11:423–41. doi: 10.4155/fmc-2018-0294
- 114. Spitthover R, Roseler T, Julius U, Heigl F, Schettler VJJ, Kuhn R, et al. Real-world study: Escalating targeted lipid-lowering treatment with PCSK9-inhibitors and lipoprotein apheresis. J Clin Apher. (2019) 34:423– 33. doi: 10.1002/jca.21695
- Luan C, Chen X, Zhu Y, Osland JM, Gerber SD, Dodds M, et al. Potentiation of psoriasis-like inflammation by PCSK9. *J Invest Dermatol.* (2019) 139:859– 67. doi: 10.1016/j.jid.2018.07.046
- 116. Machura E, Szczepanska M, Ziora K, Ziora D, Swietochowska E, Barc-Czarnecka M, et al. Evaluation of adipokines: apelin, visfatin, and resistin in children with atopic dermatitis. *Mediators Inflamm.* (2013) 2013:760691. doi: 10.1155/2013/760691
- 117. Li S, Xu RX, Zhang Y, Guo YL, Zhu CG, Liu G, et al. Relation of resistin to proprotein convertase subtilisin-kexin type 9 levels in coronary artery disease patients with different nutritional status. *J Endocrinol Invest.* (2015) 38:1291–9. doi: 10.1007/s40618-015-0310-y
- 118. Melone M, Wilsie L, Palyha O, Strack A, Rashid S. Discovery of a new role of human resistin in hepatocyte low-density lipoprotein receptor suppression mediated in part by proprotein convertase subtilisin/kexin type 9. J Am Coll Cardiol. (2012) 59:1697–705. doi: 10.1016/j.jacc.2011.11.064
- 119. Cantarini L, Pucino V, Vitale A, Talarico R, Lucherini OM, Magnotti F, et al. Immunometabolic biomarkers of inflammation in Behcet's disease: relationship with epidemiological profile, disease activity and therapeutic regimens. *Clin Exp Immunol.* (2016) 184:197–207. doi: 10.1111/cei. 12768
- 120. Turkcu FM, Sahin A, Cingu AK, Kaya S, Yuksel H, Cinar Y, et al. Serum omentin, resistin and tumour necrosis factor-alpha levels in Behcet patients with and without ocular involvement. *Graefes Arch Clin Exp Ophthalmol.* (2015) 253:1565–8. doi: 10.1007/s00417-015-3016-0
- 121. Pajvani UB, Du X, Combs TP, Berg AH, Rajala MW, Schulthess T, et al. Structure-function studies of the adipocyte-secreted hormone Acrp30/adiponectin. Implications fpr metabolic regulation and bioactivity. *J Biol Chem.* (2003) 278:9073–85. doi: 10.1074/jbc.M207198200
- 122. Frankenberg ADV, Reis AF, Gerchman F. Relationships between adiponectin levels, the metabolic syndrome, and type 2 diabetes: a literature review. *Arch Endocrinol Metab.* (2017) 61:614–22. doi: 10.1590/2359-399700000 0316
- 123. Bae YJ, Kim SH, Chung JH, Song SW, Kim KS, Kim MK, et al. Evaluation of adiposity-related biomarkers as metabolic syndrome indicators. *Clin Nutr Res.* (2013) 2:91–9. doi: 10.7762/cnr.2013.2.2.91
- 124. Kim JY, Ahn SV, Yoon JH, Koh SB, Yoon J, Yoo BS, et al. Prospective study of serum adiponectin and incident metabolic syndrome: the ARIRANG study. *Diabetes Care*. (2013) 36:1547–53. doi: 10.2337/dc12-0223
- 125. Kadowaki T, Yamauchi T, Kubota N, Hara K, Ueki K, Tobe K. Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. J Clin Invest. (2006) 116:1784–92. doi: 10.1172/JCI29126

- 126. Cerman AA, Aktas E, Altunay IK, Arici JE, Tulunay A, Ozturk FY. Dietary glycemic factors, insulin resistance, and adiponectin levels in acne vulgaris. J Am Acad Dermatol. (2016) 75:155–62. doi: 10.1016/j.jaad.2016.0 2.1220
- 127. Trinh HK, Pham DL, Ban GY, Lee HY, Park HS, Ye YM. Altered systemic adipokines in patients with chronic urticaria. Int Arch Allergy Immunol. (2016) 171:102–10. doi: 10.1159/000452626
- 128. Shibata S, Tada Y, Hau CS, Mitsui A, Kamata M, Asano Y, et al. Adiponectin regulates psoriasiform skin inflammation by suppressing IL-17 production from gammadelta-T cells. *Nat Commun.* (2015) 6:7687. doi: 10.1038/ncomms8687
- Baliwag J, Barnes DH, Johnston A. Cytokines in psoriasis. *Cytokine*. (2015) 73:342–50. doi: 10.1016/j.cyto.2014.12.014
- Furue M, Kadono T. "Inflammatory skin march" in atopic dermatitis and psoriasis. *Inflamm Res.* (2017) 66:833–42. doi: 10.1007/s00011-017-1065-z
- Davidovici BB, Sattar N, Prinz J, Puig L, Emery P, Barker JN, et al. Psoriasis and systemic inflammatory diseases: potential mechanistic links between skin disease and co-morbid conditions. *J Invest Dermatol.* (2010) 130:1785– 96. doi: 10.1038/jid.2010.103
- Leung DY, Guttman-Yassky E. Deciphering the complexities of atopic dermatitis: shifting paradigms in treatment approaches. J Allergy Clin Immunol. (2014) 134:769–79. doi: 10.1016/j.jaci.2014.08.008
- 133. Brunner PM, Silverberg JI, Guttman-Yassky E, Paller AS, Kabashima K, Amagai M, et al. Increasing comorbidities suggest that atopic dermatitis is a systemic disorder. J Invest Dermatol. (2017) 137:18–25. doi: 10.1016/j.jid.2016.08.022
- Shih VF, Tsui R, Caldwell A, Hoffmann A. A single NFkappaB system for both canonical and non-canonical signaling. *Cell Res.* (2011) 21:86– 102. doi: 10.1038/cr.2010.161
- 135. Lawrence T. The nuclear factor NF-kappaB pathway in inflammation. *Cold Spring Harb Perspect Biol.* (2009) 1:a001651. doi: 10.1101/cshperspect.a001651
- Ferreiro DU, Komives EA. Molecular mechanisms of system control of NF-kappaB signaling by IkappaBalpha. *Biochemistry*. (2010) 49:1560– 7. doi: 10.1021/bi901948j
- 137. Shi H, Kokoeva MV, Inouye K, Tzameli I, Yin H, Flier JS. TLR4 links innate immunity and fatty acid-induced insulin resistance. J Clin Invest. (2006) 116:3015–25. doi: 10.1172/JCI28898
- Catrysse L, van Loo G. Inflammation and the metabolic syndrome: the tissue-specific functions of NF-kappaB. *Trends Cell Biol.* (2017) 27:417– 29. doi: 10.1016/j.tcb.2017.01.006
- 139. Zhang J, Gao Z, Yin J, Quon MJ, Ye J. S6K directly phosphorylates IRS-1 on Ser-270 to promote insulin resistance in response to TNF-(alpha) signaling through IKK2. J Biol Chem. (2008) 283:35375–82. doi: 10.1074/jbc.M806480200
- 140. Sun L, Zhang XJAI. The immunological and genetic aspects in psoriasis. Appl Inform. (2014) 1:3. doi: 10.1186/s40535-014-0003-1
- 141. Luan C, Chen X, Hu Y, Hao Z, Osland JM, Chen X, et al. Overexpression and potential roles of NRIP1 in psoriasis. *Oncotarget.* (2016) 7:74236– 46. doi: 10.18632/oncotarget.12371
- 142. Tang ZH, Peng J, Ren Z, Yang J, Li TT, Li TH, et al. New role of PCSK9 in atherosclerotic inflammation promotion involving the TLR4/NF-kappaB pathway. *Atherosclerosis.* (2017) 262:113–22. doi: 10.1016/j.atherosclerosis.2017.04.023
- 143. Ricci C, Ruscica M, Camera M, Rossetti L, Macchi C, Colciago A, et al. PCSK9 induces a pro-inflammatory response in macrophages. *Sci Rep.* (2018) 8:2267. doi: 10.1038/s41598-018-20425-x
- 144. Zschiedrich I, Hardeland U, Krones-Herzig A, Berriel Diaz M, Vegiopoulos A, Muggenburg J, et al. Coactivator function of RIP140 for NFkappaB/RelA-dependent cytokine gene expression. *Blood.* (2008) 112:264–76. doi: 10.1182/blood-2007-11-121699
- 145. Ho PC, Tsui YC, Feng X, Greaves DR, Wei LN. NF-kappaB-mediated degradation of the coactivator RIP140 regulates inflammatory responses and contributes to endotoxin tolerance. *Nat Immunol.* (2012) 13:379– 86. doi: 10.1038/ni.2238
- Caplan AI, Correa D. The MSC: an injury drugstore. *Cell Stem Cell*. (2011) 9:11–5. doi: 10.1016/j.stem.2011.06.008

- Gaur M, Dobke M, Lunyak VV. Mesenchymal stem cells from adipose tissue in clinical applications for dermatological indications and skin aging. *Int J Mol Sci.* (2017) 18:208. doi: 10.3390/ijms18010208
- 148. Ma S, Xie N, Li W, Yuan B, Shi Y, Wang Y. Immunobiology of mesenchymal stem cells. *Cell Death Differ*. (2014) 21:216–25. doi: 10.1038/cdd.2013.158
- Matsushita K. Mesenchymal stem cells and metabolic syndrome: current understanding and potential clinical implications. *Stem Cells Int.* (2016) 2016:2892840. doi: 10.1155/2016/2892840
- Lee CW, Hsiao WT, Lee OK. Mesenchymal stromal cell-based therapies reduce obesity and metabolic syndromes induced by a high-fat diet. *Transl Res.* (2017) 182:61–74 e68. doi: 10.1016/j.trsl.2016.11.003
- 151. Saleh F, Itani L, Calugi S, Grave RD, El Ghoch M. Adipose-derived mesenchymal stem cells in the treatment of obesity: a systematic review of longitudinal studies on preclinical evidence. *Curr Stem Cell Res Ther.* (2018) 13:466–75. doi: 10.2174/1574888X13666180515160008
- 152. Rokunohe A, Matsuzaki Y, Rokunohe D, Sakuraba Y, Fukui T, Nakano H, et al. Immunosuppressive effect of adipose-derived stromal cells on imiquimod-induced psoriasis in mice. *J Dermatol Sci.* (2016) 82:50–3. doi: 10.1016/j.jdermsci.2015.12.007
- Comella K, Parlo M, Daly R, Dominessy K. First-in-man intravenous implantation of stromal vascular fraction in psoriasis: a case study. *Int Med Case Rep J.* (2018) 11:59–64. doi: 10.2147/IMCRJ.S163612
- 154. Chen H, Niu JW, Ning HM, Pan X, Li XB, Li Y, et al. Treatment of psoriasis with mesenchymal stem cells. Am J Med. (2016) 129:e13– 14. doi: 10.1016/j.amjmed.2015.11.001
- 155. De Jesus MM, Santiago JS, Trinidad CV, See ME, Semon KR, Fernandez MO Jr, et al. Autologous adipose-derived mesenchymal stromal cells for the treatment of psoriasis vulgaris and psoriatic arthritis: a case report. *Cell Transplant.* (2016) 25:2063–9. doi: 10.3727/096368916X691998
- 156. Kim KH, Blasco-Morente G, Cuende N, Arias-Santiago S. Mesenchymal stromal cells: properties and role in management of cutaneous diseases. J Eur Acad Dermatol Venereol. (2017) 31:414–23. doi: 10.1111/jdv.13934

- 157. Kornicka K, Houston J, Marycz K. Dysfunction of mesenchymal stem cells isolated from metabolic syndrome and type 2 diabetic patients as result of oxidative stress and autophagy may limit their potential therapeutic use. *Stem Cell Rev.* (2018) 14:337–45. doi: 10.1007/s12015-018-9809-x
- 158. Silva KR, Liechocki S, Carneiro JR, Claudio-da-Silva C, Maya-Monteiro CM, Borojevic R, et al. Stromal-vascular fraction content and adipose stem cell behavior are altered in morbid obese and post bariatric surgery exobese women. *Stem Cell Res Ther.* (2015) 6:72. doi: 10.1186/s13287-015-0029-x
- Shirjang S, Mansoori B, Solali S, Hagh MF, Shamsasenjan K. Toll-like receptors as a key regulator of mesenchymal stem cell function: an up-todate review. *Cell Immunol.* (2017) 315:1–10. doi: 10.1016/j.cellimm.2016. 12.005
- 160. Tang Y, Cai B, Yuan F, He X, Lin X, Wang J, et al. Melatonin pretreatment improves the survival and function of transplanted mesenchymal stem cells after focal cerebral ischemia. *Cell Transplant.* (2014) 23:1279– 91. doi: 10.3727/096368913X667510
- Ghosh D, McGrail DJ, Dawson MR. TGF-beta1 pretreatment improves the function of mesenchymal stem cells in the wound bed. *Front Cell Dev Biol.* (2017) 5:28. doi: 10.3389/fcell.2017.00028

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Hu, Zhu, Lian, Chen, Bartke and Yuan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.