

HHS Public Access

J Invest Dermatol. Author manuscript; available in PMC 2012 September 01.

Published in final edited form as:

Author manuscript

J Invest Dermatol. 2012 March ; 132(3 0 1): 593-600. doi:10.1038/jid.2011.376.

TNFAIP3 Gene Polymorphisms Are Associated with Response to TNF Blockade in Psoriasis

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Abstract

The *TNFAIP3* gene has been associated with psoriasis, rheumatoid arthritis, type 1 diabetes mellitus, systemic lupus erythematosus and celiac disease. *TNFAIP3* encodes A20, a TNF- α -inducible zinc finger protein thought to limit NF- κ B mediated immune responses. In this study we report association of response of psoriasis to TNF blockers with two *TNFAIP3* SNPs (rs2230926 in exon 3 and rs610604 in intron 6) and their haplotypes. Treatment response was self-evaluated using a 0–5 visual analog scale in 433 psoriasis patients who received TNF blockers. Confirmation was sought in 199 psoriasis and psoriatic arthritis patients from Toronto who were followed prospectively. Response variables were dichotomized separately in the two cohorts, yielding similar proportions of good responses. While significant association between dosage of the G allele of rs610604 and good combined response to all TNF blockers (OR = 1.50, p_{corr} = 0.050) and etanercept (OR = 1.64, p_{corr} = 0.016). The rs2230926 T–rs610604 G haplotype was similarly associated. By demonstrating an association with therapeutic response, these results provide a clinically relevant functional correlate to the recently described genetic association between psoriasis and *TNFAIP3*.

Keywords

psoriasis; TNFAIP3; tumor necrosis factor; pharmacogenetics

URLs

Plink software: http://pngu.mgh.harvard.edu/purcell/plink/ G*Power software: http://www.psycho.uni-duesseldorf.de/aap/projects/gpower/

CONFLICT OF INTEREST

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The authors state no conflict of interest.

INTRODUCTION

Psoriasis is a common immune mediated disorder of the skin, nails, scalp and joints, with a multifactorial genetic basis. Recent association studies have identified 25 susceptibility loci reaching genome-wide significance for psoriasis (Ellinghaus et al., 2010; Huffmeier et al., 2010; Nair et al., 2009; Strange et al., 2010; Stuart et al., 2010; Sun et al., 2010). The HLA-C region on chromosome 6p21.3 emerged as the strongest associated locus in all populations studied, accompanied by several genes involved in the immune and inflammatory pathways (Elder et al., 2010). Biological drugs targeting tumor necrosis factor-a (TNF-a, encoded by the TNF gene which happens to be located 303 kb centromeric of HLA-C), have been highly successful in treating severe psoriasis vulgaris (PsV), as well as the subset of approximately 25% of PsV patients (Gottlieb et al., 2003; Leonardi et al., 2003; Papp et al., 2005) diagnosed with psoriatic arthritis (PsA). Nevertheless, there is marked heterogeneity in treatment response to these medications. Multiple studies evaluating the antipsoriatic efficacy of anti-TNF agents have demonstrated efficacy of 75% reduction in Psoriasis Area Severity Index (PASI, a composite score that measures erythema, induration and scaling weighted for the body surface involved) in 45-70% patients depending on the dosage and duration of treatment (Sobell et al., 2009).

Compared with traditional systemic therapies for psoriasis such as phototherapy and methotrexate, treatment with the anti-TNF drugs poses a considerable financial burden. A year of biologic therapy can cost between US \$10,000–\$25,000 based on the dosage and treatment regimen prescribed (Cordoro and Feldman, 2007) while methotrexate and narrow-band ultraviolet B (NB-UVB) phototherapy would cost \$623 and \$3,692 respectively (Pearce *et al.*, 2006). Given these costs, accurate predictors of therapeutic response to anti-TNF agents would be of great value in making decisions on treatment options.

Previous studies addressing genetic predictors of therapeutic response have investigated promoter region polymorphisms in the *TNF* gene in rheumatoid arthritis (RA), ankylosing spondylitis (AS), Crohn's disease and ulcerative colitis (Kooloos *et al.*, 2007; Maxwell *et al.*, 2008; Musone *et al.*, 2008; O'Rielly *et al.*, 2009; Pierik *et al.*, 2006; Seitz *et al.*, 2007). We carried out a survey-based evaluation of *TNF* promoter region polymorphisms in 343 psoriasis patients treated with TNF antagonists, and found an association with the TNF-238 G/A polymorphism (OR=2.03, p=0.044) (Tejasvi *et al.*, 2008). However, this association was only nominally significant, prompting evaluation of additional patients as well as additional potential genetic loci.

TNFAIP3 was originally identified as a TNF inducible gene which, at least in some settings, functions as a negative feedback inhibitor of TNF signaling (Opipari *et al.*, 1990; Werner *et al.*, 2008). Recent genome-wide association studies (GWAS) have revealed associations between *TNFAIP3* and psoriasis (Nair *et al.*, 2009; Strange *et al.*, 2010), rheumatoid arthritis (RA) (Plenge *et al.*, 2007; Thomson *et al.*, 2007), type 1 diabetes mellitus (Fung *et al.*, 2009), systemic lupus erythematosus (SLE) (Graham *et al.*, 2008), celiac disease (Trynka *et al.*, 2009), and chronic sinusitis (Cormier *et al.*, 2009). Its protein product, A20, functions as a dual enzyme: initially it removes Lys⁶³-linked ubiquitin chains from RIP1, an essential mediator of the proximal TNFR signaling complex. Later, it functions as E3 ligase, leading

to polyubiquitinylation of RIP1 via Lys⁴⁸ that targets RIP1 for proteasomal degradation, resulting in termination of TNF-induced NF-κB signaling (Liu *et al.*, 2005; Wertz *et al.*, 2004). A20 can also remove K63-linked polyubiquitin chains from TRAF6 and RIP2, inhibiting NF-kB activation by TLR4 and NOD2, respectively (Vereecke *et al.*, 2009). A20 also blocks NF-kB activation induced by TAK1 overexpression, which signals downstream of RIP1, RIP2 and TRAF6 (Zetoune *et al.*, 2001). It can also target TRAF2 for lysosomal degradation, independent of its ubiquitin modifying properties (Li *et al.*, 2009). Thus A20 acts at multiple steps in the NF-kB signaling pathway (Vereecke *et al.*, 2009).

A20 binds to ABIN-1, a protein encoded by the *TNIP1* gene, which, interestingly, is also associated with psoriasis (Nair *et al.*, 2009) and SLE (Gateva *et al.*, 2009; Han *et al.*, 2009). This interaction also temporally limits NF- κ B activation (Heyninck *et al.*, 1999), at least in part by binding of ABIN-1 to NEMO/IKK- γ (Mauro *et al.*, 2006).

In a genome-wide association study (GWAS) of response to TNF blockers in rheumatoid arthritis (RA), 89 patients were analyzed using 283,348 single nucleotide polymorphisms (SNPs), yielding nine signals reaching nominal significance (p < 0.05) (Liu *et al.*, 2008). In a very recent GWAS, a total of 1286 anti-TNF-treated patients were analyzed in a 3-stage design (Plant *et al.*, 2011). No loci yielded statistical evidence reaching genome-wide significance, emphasizing the need for much larger cohorts for unbiased estimation of pharmacogenetic responses to TNF antagonists and indeed to other biologics and immunomodulators in general.

A recent pharmacogenomic study looking into the profile of early gene expression profile among responders and non-responders to anti-TNF therapy in RA revealed early downregulation of expression of *TNFAIP3* levels secondary to TNF neutralization in responders (Koczan *et al.*, 2008). However, to date there are no published pharmacogenetic studies of the relationship between the *TNFAIP3* locus and responsiveness to anti-TNF agents in psoriasis. In this study, we evaluate the role of two polymorphisms in the *TNFAIP3* gene as predictors of clinical response to TNF blockade in patients with psoriasis and/or PsA from two study sites.

RESULTS

We studied 433 psoriasis patients from Michigan and 199 patients from Toronto who received treatment with any of three different TNF blockers – etanercept, infliximab and adalimumab. We compared the two samples for 19 demographic and phenotypic variables (Table 1). Highly significant differences were observed between the means of three variables—the Toronto sample had more PsA patients (93.0% vs. 58.3%, p = 2.9×10^{-20}), while the Michigan sample had a higher mean age (51.4 yrs vs. 46.0 yrs, p = 8.6×10^{-7}) and a higher percentage of patients with a family history of psoriasis (60.6% vs. 42.9 %, p = 4.5×10^{-5}). The Michigan sample also had a later age at PsA onset (37.6 yrs vs. 33.6 yrs, p = 0.0028). A nominally significant difference (p = 0.019) in the percentage of patients with inflammatory bowel disease did not remain so after correction for multiple testing. No significant difference between the two samples was seen in the percentage of good response to treatment with any of the anti-TNF agents, either individually or combined. Furthermore,

the frequencies of the allele imparting risk for psoriasis did not differ significantly between the two groups for either of the two analyzed *TNFAIP3* SNPs.

Covariate analyses with age at onset, BMI and presence of PsA were performed in both cohorts, along with worst-ever total body surface area (TBSA) in the Michigan cohort. Age at onset was significantly and negatively associated with response to TNF blockers in the Michigan cohort (OR=0.973, p=0.00016). Although not reaching nominal significance, age at onset also trended towards negative association with all all anti-TNF responses in the Toronto sample (OR=0.982 for all TNF blockers combined).

Data were analyzed for linear trend of association between drug response and genotypes and haplotypes of one disease-associated SNP in intron 6 (rs610604, MAF = 0.338) (Nair *et al.*, 2009) and another in exon 3 of TNFAIP3 (rs2230926, MAF = 0.029). The latter was the only tested exonic SNP that was polymorphic in our sample. In the Michigan sample, association was observed between dosage of the G allele of rs610604 and good response to all TNF blockers (OR = 1.74, $p_{nom} = 0.0027$, $p_{corr} = 0.0084$)(Table 2). Combined analysis of the two samples using a fixed-effects model showed association between dosage of the G allele of rs610604 and good response to all elle of rs610604 and good response to all TNF blockers combined (OR= 1.50, p_{nom} = 0.010, $p_{corr} = 0.050$). Despite a similar OR, this association was no longer significant when the combined analysis used a random-effects model (OR= 1.39, p_{nom} = 0.22, $p_{corr} = 0.77$) (Table 2). Age at onset of disease was included as a covariate in all association models and the nested logistic regression models testing association for each combination of SNP and drug revealed that the associations of age at onset and SNP rs610604 with drug response were independent of each other (no significant interaction).

Haplotype analysis (Table 3) revealed that dosage of the rs2230926 T–rs610604 G haplotype was associated with good response to all TNF blockers (OR = 1.82, $p_{nom} = 0.0012$, $p_{corr} = 0.0060$) in the Michigan cohort. Combined analysis of both groups under a fixed effects model showed association between the rs2230926 T–rs610604 G haplotype and response to all TNF blockers (OR = 1.55, $p_{nom} = 0.0051$, $p_{corr} = 0.031$). The association of haplotype rs2230926 T–rs610604 G with response to TNF blockers in the Michigan cohort is driven mainly by rs610604; conditional haplotype testing showed that the independent contribution of the rs610604 G allele to the association is significant ($p_{nom} = 0.0016$) whereas the independent contribution of the rs230926 T allele is not ($p_{nom} = 0.90$).

DISCUSSION

The *TNFAIP3* gene has been shown to be associated with psoriasis in a recent genome-wide association study (Nair *et al.*, 2009), a finding recently confirmed strongly in Caucasian (Strange *et al.*, 2010) and suggestively in Chinese (Sun *et al.*, 2010) populations. Here we demonstrate that polymorphisms in *TNFAIP3* are associated with response to anti-TNF agents in psoriasis patients. In the Michigan sample, good response to etanercept alone and all TNF blockers combined was positively associated with dosage of the G allele of rs610604, a psoriasis susceptibility allele (Nair *et al.*, 2009). The proportion of good responders to anti-TNF treatment was substantially larger for people carrying two copies of this allele compared to carriers of no copies, both for etanercept (90.7% vs. 70.3%, p =

0.0027) and for all TNF blockers combined (88.1% vs. 70.7%, p = 0.0075). To a lesser degree, the same holds true for heterozygous carriers of the G risk allele compared to non-carriers (79.1% vs. 70.3%, p = 0.067 for etanercept and 80.1% vs. 70.7%, p = 0.034 for all TNF blockers). We also found an association of haplotype rs2230926 T–rs610604 G with good response to etanercept or all TNF blockers (Table 3).

Here we also report significant association between age at onset and treatment response that is independent of *TNFAIP3* genotype; a later age at onset is associated with poorer response to treatment. The profile of genetic risk factors, especially the role of *HLA-Cw6*, differs between early and late onset psoriasis (Chen *et al.*, 2011; Gudjonsson *et al.*, 2002), which may influence the effectiveness of TNF blockers. The use of phenotypes as predictors of response to anti-TNF agents in RA showed that age of the patient was not associated with treatment response (Eder *et al.*, 2010) and in AS age was negatively associated with response to treatment (Arends *et al.*, 2011). Since we did not collect age at treatment data, a direct comparison was not possible. Other covariates analyzed did not show any association with treatment response.

We did not find any association of drug response and *TNFAIP3* genotype in the Toronto replication sample. This outcome could be due to limited power. For the strongest effect observed in the Michigan sample (OR of 1.83 for etanercept response and G allele dosage at rs6010604), the Toronto sample has only 43% power to detect association at a nominal significance level of 0.05 and 29% power when significance is corrected for multiple testing. Because of the winner's curse (Goring *et al.*, 2001), the estimated effect size in the Michigan sample is likely biased upwards, and power for a more realistic OR of 1.50 (midway between 1.83 and the lower bound of its corrected 95% confidence interval), is only 22% and 13% at a nominal and corrected significance level of 0.05, respectively. To achieve 80% power to detect an OR of 1.83 at a nominal or corrected significance of 0.05, sample size for the Toronto cohort would need to be increased 2.5- or 3.2-fold; for an OR of 1.50 80% power requires 5.5- or 6.9-fold increases in sample size.

The lack of association in the Toronto sample could also be due to heterogeneity; i.e., response to anti-TNF drugs is genuinely more poorly associated with *TNFAIP3* genotype in the Toronto population than in the Michigan population. The estimated I² coefficient of heterogeneity for association, of both etanercept response and combined TNF blocker response with r610604 genotype, is greater than zero (I² = 27.5% and 54.3%, respectively), leading to a reduced significance of the combined analysis for these two associations when using a random-effects model versus fixed-effects model (Table 2). However, neither of these I² coefficients is significant (95% confidence interval = 0.0–72.0% and 0.0–88.8%, respectively), nor is the related Cochran's Q-test of heterogeneity (p = 0.24 and 0.14, respectively). It is important to note that these results are inconclusive, since the power of heterogeneity tests is known to be low when the number of strata is small.

Other potential limitations of this study could also be responsible for the different results for the two samples. We combined skin and joint responses that were measured individually in Toronto to approximate the undifferentiated self-evaluated response for the Michigan sample. Also, the response to medication in the Michigan cohort was assessed

retrospectively, which may introduce recall error (though not recall bias since it is unlikely that recall accuracy is differentially affected by TNFAIP3 genotype). The similar proportion of good responders in the two samples to each of the anti-TNF therapies (Table 1) provides some confidence that the two response metrics, though measured in different ways, are reasonably similar. Nevertheless, the much higher proportion of PsA in the Toronto sample could contribute to the observed lack of association if the response of the joint symptoms of people with PsA to anti-TNF therapy is more poorly correlated with TNFAIP3 genotype than is response of the skin symptoms of people with PsA or purely cutaneous psoriasis (PsC). Table S1 compares skin-only and joint-only responses in the Toronto sample. Despite limited power, this analysis yields no indication that joint symptoms are more poorly associated with TNFAIP3 genotype than are skin symptoms—for rs610604, the OR of response of joint vs. skin phenotypes is 1.17 vs. 0.77 for etancercept and 1.08 vs. 0.65 for all TNF blockers combined. Unfortunately, a similar comparison in the Michigan sample is not possible because this information was not collected. In lieu of this, Table S2 compares response to TNF blockers in the Michigan sample for people with PsA versus those with PsC. The strength of association with SNP rs610604 is approximately equal for both phenotypes, being slightly lower in PsA (OR = 1.67 vs. 1.77 for all TNF blockers combined). Hence neither sample supports the notion that response of joint symptoms to anti-TNF therapy is less well associated with TNFAIP3 genotype than is response of skin symptoms, although the small sample sizes prevent any definitive conclusions.

In conclusion, we demonstrate in this study that the G allele of SNP rs610604 located in the *TNFAIP3* gene and its haplotype with the T allele of rs2230926 are markers of good response to anti-TNF agents. The results of this study using a limited sample set calls for confirmation by additional studies with substantially larger number of samples, prospective patient and physician evaluated responses, and gene and protein expression levels measured before and after therapy. Such efforts would also provide phenotype data and material needed for a GWAS of anti-TNF agents. Further association studies on *TNFAIP3* and other genetically associated loci could result in a panel of polymorphisms that can be routinely typed in a clinical setting to help the physician make an informed decision on administration of anti-TNF therapy.

MATERIALS AND METHODS

Subjects

The Michigan sample included 433 patients with psoriasis who received anti-TNF agents from 2005–2009 and completed a questionnaire concerning their response to the drug therapy. The Toronto sample consisted of 199 patients with psoriasis vulgaris who received anti-TNF therapy at Toronto Western Hospital, Center for Prognosis Studies in the Rheumatic Diseases. A physician (dermatologist and/or rheumatologist) diagnosed the presence of psoriasis and PsA in all patients. Only 3 of the 199 Toronto patients did not have cutaneous lesions.

All patients were of European Caucasian descent. Informed consent was obtained from all subjects under protocols adherent to the Declaration of Helsinki Principles, which were approved by the Institutional Review Boards of the participating institutions.

Phenotypes

For the Michigan sample, response to treatment with anti-TNF drugs (etanercept, adalimumab, and infliximab) was self-evaluated using a 0–5 visual analogue scale, 0 being not effective and 5 being very effective. Responses were dichotomized as good if they scored from 3 to 5 and poor if they scored 0 to 2. For those patients treated with more than one anti-TNF drug (17.7% of the sample received two and 5.5% all three drugs), a combined response to all TNF blockers was computed as the mean of the raw 0–5 response for all drugs used by the patient; the mean response was then dichotomized to responders and non-responders using a 2.5 threshold.

In Toronto, rheumatologists evaluated the response to treatment for the same three anti-TNF drugs. Only response to the first anti-TNF agent prescribed was considered. For skin symptoms, a 50% or greater reduction from the patient's baseline PASI was considered a good response. For joint symptoms, a good response was defined as a 50% or better decrease in the actively inflamed (swollen and/or tender) joint count or 50% or better improvement in BASDAI (Bath Ankylosing Spondylitis Disease Activity Index) at 6 months from the initiation of treatment. The skin and joint responses of the Toronto patients were combined into a single metric for most of the analyses—if either or both responses were good, the combined skin-joint response to the first anti-TNF drug, the combined response for all TNF blockers was simply the combination of all single drug responses across the sample.

Demographic data including age, sex, race, and history of psoriasis and other autoimmune diseases in patients and family members were recorded for both samples. Body mass index, fingernail involvement, age at onset and duration of psoriasis and PsA were also recorded.

Genotyping

In Ann Arbor, genomic DNA was prepared from peripheral blood mononuclear cells or Epstein-Barr virus-immortalized lymphoblastoid cell lines, as previously described (Nair *et al.*, 1995). In Toronto, DNA was extracted from peripheral blood using a modified salting out procedure (Gentra Puregene Blood Kit, Qiagen, Valencia, CA). The genotypes were determined by Taqman assay (Applied Biosystems, Foster City, CA).

Statistical analysis

The Michigan and Toronto samples were compared for a variety of epidemiological and phenotypic variables using two-sample tests for the equality of the means. For variables expressed as a percentage, Fisher's exact test was used. All other variables except age were first transformed using the optimal Box-Cox power transform (power of 0.5 for age at PsV onset and duration of PsV, 0.7 for age at PsA onset, 0.2 for duration of PsA, and -0.1 for BMI) in order to achieve near-normality of the residuals of the appropriate regression model. Means for age, age at onset of PsV, and age at onset of PsA were then tested with the two-sample t-test for unequal variances. Means for duration of PsV, duration of PsA, and BMI were tested with a multiple regression model that included linear (duration PsA), linear and quadratic (BMI), or linear, quadratic, and cubic (duration PsV) terms for age and that used a Huber-White heteroscedasticity consistent covariance matrix.

Data were analyzed for linear trend of association between drug response and genotypes and haplotypes of two SNPs in TNFAIP3 using logistic regression, where drug response was coded as a binary variable (0 = poor, 1 = good), and the genotype predictor variable was expressed as the dosage of the SNP allele or two SNP haplotype. Association between drug response and several covariates (worst-ever TBSA, BMI, age at onset, and PsA) was also tested with logistic regression. The Box-Tidwell procedure was applied to association testing of TBSA, BMI, and age at onset to determine whether a power transformation was necessary to achieve a linear relationship between the logit of drug response and the covariate, which is an important assumption of logistic regression. In all cases, the deviation between a model using the optimal linearizing power transformation of the covariate and a model using the untransformed covariate was nonsignificant, so untransformed covariates were used in all tests. Age at onset, the only covariate to show significant association with drug response, was included in all tests assessing association between drug response and genotypes and haplotypes of the two SNPs in TNFAIP3. Interaction between age at onset and SNP genotype was assessed with a likelihood ratio test applied to nested logistic regression models with and without a multiplicative interaction term. Conditional haplotypebased association testing was performed by comparing the odds of disease for pairs of haplotypes that have different alleles for the SNP under scrutiny but the same allele for the other SNP.

For individual cohorts, nominal significance of association was assessed using 100,000 random permutations of the drug response variable. A special set of 100,000 restricted permutations of drug response labels was generated to determine significance that was corrected for multiple testing of eight single marker tests (4 drug treatments \times 2 SNPs), twelve individual haplotype tests (4 drugs \times 3 haplotypes), or four omnibus haplotype tests (4 drugs \times 1 omnibus test). Responses for all SNP–drug combinations were permuted as intact vectors to maintain all dependencies among responses, and response vectors were permuted randomly within strata formed by groups of patients receiving the same combination of drugs to ensure constant sample size among all permutations in the presence of missing genotype data. The best association p-value among all tests for each iteration of the permutation procedure was saved, the saved p-values were sorted into ascending order, and the corrected p-value determined as the fractional rank within this vector of the best observed p-value across all tests.

Meta-analysis was used to test for association across both cohorts combined. For single markers, standard fixed and random-effects models were used. Asymptotic p-values were reported for nominal significance, and the restricted permutations generated for the Michigan and Toronto samples were used to determine significance corrected for multiple testing. For haplotypes, fixed-effects meta-analysis of both cohorts was formulated as a logistic regression model with sample cohort as an additional covariate, and permutations were used to assess both nominal and corrected significance. Heterogeneity of ORs between the two samples was tested with Cochran's Q statistic and the I² heterogeneity index. All association analyses were carried out with version 1.0.7 of PLINK (Purcell *et al.*, 2007).

Power calculations were carried out with version 3.1.2 of G*Power (Faul *et al.*, 2007) using an exact unconditional test of allelic association, which under Hardy-Weinberg equilibrium

is essentially equivalent to the permutational version of the logistic regression test used in this study. Risk allele frequencies and the ratio of good to poor responders in the underlying population were estimated from the Toronto sample; the expected ORs for association were estimated from the Michigan sample. A type I error rate of 0.021 was used to compute power of association tests that are corrected for multiple testing, which is equivalent to a corrected significance of 0.05 when testing single SNPs in the Toronto sample.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

This research was supported by NIH grants R01 AR042742 and R01 AR050511, the Ann Arbor Veterans Affairs Hospital (JTE), the Dudley and Dawn Holmes Fund, the Babcock Memorial Trust, a Canadian Institutes of Health Research New Emerging Team (NET) Grant, the Krembil Foundation, and Arthritis Society of Canada National Research Initiative Grant. We thank Sarah Laponsa, Anna Pero and Lynda Hodges for their tireless efforts in recruiting patients and data entry.

Abbreviations

GWAS	genome-wide association study
MAF	minor allele frequency
NF-ĸB	nuclear factor kappa light chain enhancer of activated B cells
SNP	single nucleotide polymorphism
OR	odds ratio
PsV	psoriasis vulgaris
PsA	psoriatic arthritis
PsC	cutaneous psoriasis without arthritis
TNF	tumor necrosis factor-a
TNFAIP3	tumor necrosis factor alpha induced protein 3

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Table 1

Phenotypic comparison of Michigan and Toronto samples

	Micl	nigan sar	nple	Tor	onto san	ple		
Variable (units)	n ³	mean	$^{\rm sd^4}$	n ³	mean	sd^4	p-value ⁵	
Age at study entry (yrs)	433	51.4	13.9	199	46.0	12.0	$8.6 imes 10^{-7}$	
Age at PsV onset (yrs)	433	26.9	15.5	196	26.2	13.7	0.75	
Duration of psoriasis (yrs)	433	24.4	15.6	196	19.7	12.4	0.055	
Age at PsA onset (yrs)	214	37.6	13.6	185	33.6	12.6	0.0028	
Duration of PsA (yrs)	214	14.8	12.3	185	12.3	9.3	0.61	
BMI $(kg/m^2)^I$	431	30.5	7.2	192	29.1	5.9	0.10	
Males (%)	433	43.2	2.4	199	38.7	3.5	0.30	
PsA (%)	376	58.3	2.6	199	93.0	1.8	$2.9 imes 10^{-20}$	
Nail involvement (%)	417	79.4	2.0	199	84.4	2.6	0.15	
IBD (%) ²	405	3.2	0.9	199	2.0	1.0	0.60	
Autoimmune disease (%)	402	5.7	1.2	197	4.6	1.5	0.70	
Family history PsV (%)	419	60.6	2.4	198	42.9	3.5	$4.5 imes 10^{-5}$	
Family history IBD (%)	415	11.8	1.6	194	5.7	1.7	0.019	
rs610604 G allele frequency (%)	432	37.5	1.6	188	38.6	2.5	0.75	
rs2230926 T allele frequency (%)	430	97.1	0.6	188	96.8	0.9	0.86	
Good etanercept response (%)	380	77.4	2.1	139	83.5	3.2	0.15	
Good adalimumab response (%)	89	77.5	4.4	31	74.2	7.0	0.81	
Good infliximab response (%)	89	80.9	4.2	29	82.8	4.2	1.00	
Good TNF blocker response (%)	433	77.6	2.0	199	81.9	2.7	0.25	
l Body mass index					- -			
² Irritable bowel disease								
3 Number of non-missing observation	s							
⁴ Standard deviation								
\mathcal{S} Nominal p-value for the test of the e	quality	of the va	riable r	neans fo	or the Mi	chigan a	nd Toronto samples	<i>i</i>

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Tests for duration of PsV, duration of PsA, and BMI were adjusted for the effects of age. See Materials and Methods for details of testing procedure.

				Michigan sam	ple			Toronto Sample	8		Combine	1 analysis (Fixed/random effects)
SNP	Drug	Genotype	Good response ¹	Poor response ¹	OR^2	\mathbf{P}^3	Good response ¹	Poor response ¹	OR^2	ЪЗ	OR^2	P3
		TT	275 (.945)	79 (.919)			101 (.927)	19 (.950)				
	etanercept	TG	16 (.055)	6 (.070)	1.77	0.19(0.74)	8 (.073)	1 (.050)	0.63	0.67 (0.99)	1.53/1.53	0.30/0.30 (0.92/0.89)
		GG	0 (.000)	1 (.011)			0 (000) 0	0 (.000)				
		TT	54 (.915)	28 (.933)			20 (.952)	5 (.833)				
	adalimumab	TG	5 (.085)	2 (.067)	0.66	0.64~(1.00)	1 (.048)	1 (.167)	2.39	0.59 (0.98)	1.12/1.12	0.89/0.89 (1.00/1.00)
		GG	0 (.000)	0 (.000)			0 (.000)	0 (.000)				
152250920		TT	69 (.958)	15 (.882)			23 (.958)	5 (1.00)				
	infliximab	TG	3 (.042)	2 (.118)	3.00	0.26 (0.92)	1 (.042)	0 (.000) 0	0.00	1.00(1.00)	3.00/3.00	0.26/0.26 (0.89/0.86)
		GG	0 (.000)	0 (.000)			0 (000) 0	0 (.000)				
		TT	315 (.956)	91 (.944)			144 (.935)	29 (.935)				
	combined	TG	18 (.044)	5 (.037)	1.40	0.45(0.98)	10 (.065)	2 (.065)	06.0	0.89~(1.00)	1.26/1.26	0.55/0.55 (0.99/0.99)
		GG	0 (.020)	1 (.019)			0 (.000)	0 (.000)				
		GG	49 (.167)	5 (.058)			14 (.128)	2 (.100)				
	etanercept	GT	140 (.478)	37 (.430)	1.83	$0.0020\ (0.0068)$	55 (.505)	10 (.500)	1.10	0.79~(1.00)	1.64/1.57	0.0043/0.051 (0.016/0.31)
		ΤΤ	104 (.355)	44 (.512)			40 (.367)	8 (.400)				
		GG	3 (.052)	1 (.033)			4 (.190)	0 (.000)				
	adalimumab	GT	30 (.517)	12 (.400)	1.67	0.22 (0.79)	8 (.381)	3 (.500)	1.42	0.65(0.99)	1.61/1.61	0.19/0.19 (0.75/0.70)
rs610604		ΤΤ	25 (.431)	17 (.567)			9 (.429)	3 (.500)				
		GG	11 (.153)	2 (.118)			3 (.125)	2 (.400)				
	infliximab	GT	35 (.486)	7 (.412)	1.34	0.48(0.99)	14 (.583)	2 (.400)	0.40	0.26 (0.72)	1.05/0.90	0.89/0.85 (1.00/1.00)
		ΤΤ	26 (.361)	8 (.471)			7 (.292)	1 (.200)				
	combined	GG	52 (.156)	7 (.033)	1.74	0.0027 (0.0084)	21 (.136)	4 (.129)	1.01	0.98 (1.00)	1.50/1.39	0.010/0.22 (0.050/0.76)

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Association of genotypes for two TNFAIP3 SNPs with a combined skin-joint response to TNF blockers for psoriasis patients in the Michigan, Toronto,

Table 2

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SNP Drug Genotype Good response ¹ Poor response ¹ OR ² P ³ OR P ³ P ³ OR P ³ P ³ OR P ³					Michigan sam	ple			Toronto Sample			Compined	analysis (Fixed effects)
GT 165 (493) 41 (483) 77 (.500) 15 (.484) TT 118 (.351) 49 (.483) 56 (.364) 12 (.387) ¹ Frequency and proportion of genotype among good and poor responders to drug therapy 56 (.364) 12 (.387) ² Odds ratio for association of drug response and <i>TNFAIP3</i> SNP in a logistic regression model using SNP allele dosages and age at onset 12 (.387)	SNP	Drug	Genotype	Good response ¹	Poor response ¹	OR^2	\mathbf{P}^{3}	Good response ¹	Poor response ¹	OR^2	ЪЗ	OR ²	\mathbf{P}^3
TT 118 (.351) 49 (.483) 56 (.364) 12 (.387) I I I I I I I I I I I I I I I I I I I I I I I I I I I			GT	165 (.493)	41 (.483)			77 (.500)	15 (.484)				
¹ Frequency and proportion of genotype among good and poor responders to drug therapy ² Odds ratio for association of drug response and <i>TNFAIP3</i> SNP in a logistic regression model using SNP allele dosages and age at onset			ΤΤ	118 (.351)	49 (.483)			56 (.364)	12 (.387)				
² Odds ratio for association of drug response and <i>TNFAIP3</i> SNP in a logistic regression model using SNP allele dosages and age at onset	I Frequency an	d proportion	of genotype an	nong good and poor r	responders to drug t	herapy							
	² Odds ratio for	r association	of drug respons	se and TNFAIP3 SNF	o in a logistic regres	sion model using	g SNP allel	e dosages and age at	onset				

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Association of TNFAIP3 haplotypes and a combined skin-joint response to TNF blockers for psoriasis patients in the Michigan, Toronto, and combined samples. Age at onset of disease is included as a covariate in the association model.

			Michiga	n Sample			oronto	Sample		Com	bined ans	lysis6
Drug	Haplotype ^I	freq ²	OR^3	${ m P_{nom}}^4$	${ m P_{corr}}^5$	freq ²	OR^3	${ m P_{nom}}^4$	${\rm P_{corr}}^5$	OR^3	$P_{nom}^{}4$	P_{corr}^{2}
	TG	.409/.269	1.92	0.00077	0.0038	.381/.350	1.10	0.83	1.00	1.71	0.0017	0.011
	GT	.025/.040	0.55	0.18	0.82	.037/.025	1.60	0.44	1.00	0.67	0.36	0.96
etanercept	TT	.566/.691	0.58	0.0039	0.023	.583/.625	0.85	0.71	1.00	0.63	0.0063	0.039
	omnibus			0.0030	0.0067	I		0.81	0.98		0.0077	0.023
	TG	.303/.232	1.63	0.26	0.87	.381/.250	1.42	0.67	1.00	1.57	0.23	0.83
- 1	GT	.032/.032	1.21	0.60	1.00	.024/.083	0.42	0.29	0.99	0.87	0.92	1.00
adalimumaD	TT	.665/.736	0.63	0.24	0.84	.595/.667	0.86	0.85	1.00	0.67	0.25	0.86
	omnibus			0.36	0.86	I		0.68	0.97		0.38	0.82
	TG	.391/.322	1.32	0.53	0.99	.417/.600	0.40	0.24	0.79	1.02	0.96	1.00
	GT	.013/.056	0.21	0.14	0.76	.021/.000	8	0.73	1.00	0.29	0.17	0.87
	TT	.595/.622	06.0	0.81	1.00	.563/.400	2.10	0.40	0.88	1.08	0.81	1.00
	omnibus			0.25	0.74	I		0.27	0.79		0.41	0.91
	TG	.404/.279	1.82	0.0012	0.0060	.386/.371	1.01	96.0	1.00	1.55	0.0051	0.031
	GT	.025/.031	0.73	0.49	0.99	.032/.032	1.11	0.61	1.00	0.81	0.63	1.00
combined	TT	.571/.690	0.59	0.0034	0.016	.581/.597	0.98	0.93	1.00	0.67	0.0095	0.066
	omnibus		I	0.0055	0.014		Ι	0.96	1.00		0.021	0.066
<i>l</i> Haplotype for	r SNPs rs223092	26 and rs6106	04; omn	ibus refers t	o all haple	otypes combi	ined					
² Frequency of	haplotypes in gc	od responde:	rs and po	oor responde	SIC							
$^{\mathcal{J}}$ Odds ratio foi	r association of c	lrug response	and TN	<i>FAIP3</i> hapl	otype in a	logistic regre	ession m	odel using	g haplotyj	be dosag	es	
⁴ Nominal p-va	lue for association	on of drug re.	sponse a	nd TNFAIP	3 haplotyp	e e						
⁵ Corrected p-v	alue for associat	tion of drug r	esponse	and TNFAL	P3 haploty	pe						
6 Fixed effects	meta-analysis											