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# Fluid biomarkers of the neurovascular unit in cerebrovascular disease and vascular cognitive disorders: A systematic review and meta-analysis

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# ABSTRACT

Background: The disruption of the neurovascular unit (NVU), which maintains the integrity of the blood brain barrier (BBB), has been identified as a critical mechanism in the development of cerebrovascular and neurodegenerative disorders. However, the understanding of the pathophysiological mechanisms linking NVU dysfunction to the disorders is incomplete, and reliable blood biomarkers to measure NVU dysfunction are yet to be established. This systematic review and meta-analysis aimed to identify biomarkers associated with BBB dysfunction in large vessel disease, small vessel disease (SVD) and vascular cognitive disorders (VCD). Methods: A literature search was conducted in PubMed, EMBASE, Scopus and PsychINFO to identify blood biomarkers related to dysfunction of the NVU in disorders with vascular pathologies published until 20 November 2023. Studies that assayed one or more specific markers in human serum or plasma were included. Quality of studies was assessed using the Newcastle-Ottawa Quality Assessment Scale. Effects were pooled and methodological heterogeneity examined using the random effects model. Results: A total of 112 studies were included in this review. Where study numbers allowed, biomarkers were analysed using random effect meta-analysis for VCD (1 biomarker; 5 studies) and cerebrovascular disorders, including stroke and SVD (9 biomarkers; 29 studies) while all remaining biomarkers (n = 17 biomarkers; 78 studies) were examined through qualitative analysis. Results of the meta-analysis revealed that cerebrospinal fluid/serum albumin quotient (Q-Alb) reliably differentiates VCD patients from healthy controls (MD = 2.77; 95 % CI = 1.97–3.57; p < 0.0001) while commonly measured biomarkers of endothelial dysfunction (VEGF, VCAM-1, ICAM-1, vWF and E-selectin) and neuronal injury (NfL) were significantly elevated in vascular pathologies. A qualitative assessment of non-meta-analysed biomarkers revealed NSE, NfL, vWF, ICAM-1, VCAM-1, lipocalin-2,

1, ICAM-1, vWF and E-selectin) and neuronal injury (NfL) were significantly elevated in vascular pathologies. A qualitative assessment of non-meta-analysed biomarkers revealed NSE, NfL, vWF, ICAM-1, VCAM-1, lipocalin-2, MMP-2 and MMP-9 levels to be upregulated in VCD, although these findings were not consistently replicated. *Conclusions:* This review identifies several promising biomarkers of NVU dysfunction which require further validation. A panel of biomarkers representing multiple pathophysiological pathways may offer greater discriminative power in distinguishing possible disease mechanisms of VCD.

#### Introduction

Vascular cognitive disorders (VCD), also referred to as vascular cognitive impairment (VCI) or sometimes vascular cognitive impairment and dementia (VCID), are the second most common cause of dementia after Alzheimer's disease (AD), affecting at least 7.2 million people worldwide [1]. VCD represent a heterogeneous group of disorders with multiple pathogenetic mechanisms, with the fundamental characteristic attributable to brain injury arising from cerebrovascular disorders that affect the small and large vessels of the brain [2]. One important

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mechanism is the breakdown of the blood brain barrier (BBB) owing to the disruption of the neurovascular unit (NVU), a mechanism shared between VCD and neurodegenerative disorders [3].

This paper focuses on the NVU as the basic structural and functional subunit of the BBB. At the cellular level, the NVU is the building block of the brain parenchyma and comprises a basement membrane, endothelial cells, pericytes, vascular smooth muscle cells, glial cells (astrocytes, microglia, oligodendrocytes) and neurons [3]. The NVU is responsible for the regulation of cerebral blood flow, protection of the brain from blood borne toxins, and delivery of nutrients to, and clearance of waste products from, the brain [4]. Dysfunction of the NVU has been attributed to the degradation of endothelial cells, pericytes and loss of tight junction proteins [4]. Even though some BBB breakdown occurs with normal ageing [5], it has been found to be more pronounced in individuals with mild cognitive impairment, suggesting its association with early cognitive impairment [6]. Disruption of the NVU occurs in small and large cerebral vessel disease, following stroke [7], and in neurodegenerative disease [2], making it a key area of interest in the pathogenesis of VCD and neurodegenerative dementias.

Activation or disruption of the NVU induces release of molecules, such as soluble platelet derived growth factor receptor  $\beta$  (sPDGFR $\beta$ ) [8], occludin and claudin-5 [9] into the circulation, characteristic for specific cellular components [10]. The detection of these molecules as potential biomarkers in both CSF and blood could therefore serve as an indicator of the integrity of the NVU, and thereby the maintenance of the blood-brain barrier (BBB), cerebral homeostasis and cerebral blood flow [11]. While there are a number of reviews on biomarkers associated with VCD in general [2,12-14], none has solely categorised them as activated components of the NVU. The focal points of this review, aim to address gaps in the literature, and include; (1) a focus on blood and other fluid biomarkers, since the literature has mainly been on CSF, (2) a focus on blood testing would provide for an economical, minimally invasive, and accessible alternative that would allow clinicians to screen for VCD readily, (3) to date there has been no meta-analysis of NVU biomarkers in blood. This review provides an updated summary of existing blood biomarkers related to NVU pathology in large vessel disease, SVD and VCD. Though the primary focus of the biomarkers was to assist with diagnosis and mechanistic understanding of VCD, data were incorporated from the stroke literature as stroke is closely related to VCD and has been the subject of significant investigation in this regard. We examined all published literature on blood biomarkers of the NVU and performed meta-analyses on those with three or more published studies with relevant data format.

#### Methods

This systematic review was conducted in accordance with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidelines (Supplementary Table S1) [15]. A protocol was registered with PROSPERO (ID: CRD42023389103).

#### Database search strategy

A systematic search was conducted to identify blood-based biomarkers along the spectrum of VCD and cerebrovascular pathologies. The search was conducted in PubMed, Scopus, Embase (via Ovid) and PsychINFO (via ProQuest) for studies from inception until 20 November 2023. Search terms included combinations of the Medical Subject Headings and keywords for three themes: VCD, NVU and blood biomarkers (Supplementary Table S2). No restriction for publication period was set. Filters were applied for language and to exclude non-journal publications. Two authors (G.K.H. and T.J.) independently performed a search of the terms and evaluated all titles and abstracts using the prespecified eligibility criteria (see section below). Removal of duplicates and screening was performed via Rayyan software [16]. During the screening phase, bibliography lists of nine review articles were checked for additional primary studies using the snowball sampling design. In the event of any disagreement, a third author (S.H.) was consulted to reach a consensus.

#### Inclusion and exclusion criteria

Eligible studies met the following inclusion criteria: (1) conducted in individuals with vascular neurocognitive disorders (VCD) or cerebrovascular disorders (large vessel stroke or small vessel disease); (2) conducted in humans; (3) assayed one or more specific markers in serum or plasma; (4) assessed a biomarker relevant to a component of the NVU (extracellular matrix, tight junctions, pericytes, neuroglia and endothelial cells); (5) English language only and (6) original, primary manuscripts. In addition to the mentioned inclusion criteria, studies shortlisted for the meta-analysis had to have a case-control study design and include a healthy control group.

Studies were excluded from both the meta-analysis and systematic review if they belonged to any of the following categories: (1) animal studies; (2) non-cerebrovascular based studies, (3) studies involving vascular surgery or aneurysms, (4) studies conducted on other neurological diseases that may cause cognitive impairment such as Alzheimer's disease, Parkinson's disease, encephalopathy, traumatic brain injury and depression, (5) studies on biomarkers not related to a NVU component, (6) studies which only analysed biomarkers in tissues other than blood or CSF (e.g. Urine and saliva); and (7) other publication types (reviews, editorials, comments etc.). Full text of shortlisted studies was obtained for further review and data extraction.

#### Data extraction

Data extraction was performed by two authors (G.K.H. and T.J.). Data indices included the primary author's name, year of publication, mean age of subjects, sample size for cases and/or controls, number (or %) of female subjects, specimen type (serum or plasma), method of measurement, name of study or country or place of recruitment (where applicable), assessed blood biomarker(s), association measure and outcomes pertaining to the biomarker of interest. In case of missing or unclear data, authors were contacted by email. Meta-analyses were performed when data (expressed as mean  $\pm$  SD) were available for a minimum of three studies per biomarker to obtain reliable estimates. Studies without a healthy control group were excluded from the meta-analysis. Extracted values for stroke case groups were from baseline scores and if not provided, values close to the onset of stroke were extracted.

#### Qualitative analysis

To perform a comprehensive review of the topic, shortlisted studies that met the general inclusion criteria for the review but not the metaanalysis were summarised qualitatively. This was the case for most studies on VCD as there were insufficient articles on biomarker measurements related to the NVU. Studies on cerebrovascular disorders lacking the statistical reporting values of interest, healthy controls or if they were present in a count of less than 3 for a biomarker, were also summarised qualitatively.

#### Quantitative analysis

The test for skewness was performed using the method described in Wan et al. [17] as implemented in the R package 'metamedian' for studies reporting median, first and third quartiles. Means and SDs were estimated based on quartiles, minimum, maximum and sample sizes using two methods [17] (1) under the normality assumption for the biomarkers by Wan et al. [17,18] (2) under non-normal distributions for the biomarkers by McGrath et al. [19], as implemented in the R package 'estmeansd'.

Meta-analysis was performed using raw mean difference (MD) between cases and controls. Due to differences in biomarker measures and large variability across studies, studies were removed from the metaanalysis if their 95 % confidence intervals (CI) do not overlap with the CI of the pooled effect size (outliers). Random effects model was selected to address known methodological and analytical heterogeneity. Statistical heterogeneity was assessed using the I<sup>2</sup> statistic (values of 25 %, 50 % and 75 % indicate small, moderate and large heterogeneity, respectively).

Considering the large variations in estimated and reported means, SDs and MDs within each biomarker, the meta-analysis was repeated using the unit-free standardised mean difference (SMD), effect size measure and sensitivity of statistical inference on the pooled results were assessed.

Publication bias within the meta-analysed studies was examined using the Egger's regression test for funnel plot asymmetry as implemented in R package 'metafor'. The effect size calculations, metaanalyses and forest plots were done using the appropriate functions in the R packages 'metafor' [20] and 'dmetar' [21].

#### Quality evaluation

The quality of studies was assessed using a modified version of the Newcastle-Ottawa Quality Assessment Scale (NOS) (Supplementary Table S6–7). The adaptation of questions was validated by consultation with senior authors (P.S. and A.P.). Two authors (G.K.H. and T.J.) independently appraised three aspects of the scale: selection of study groups, comparability of the groups and the outcome of interest. The maximum number of points awarded to each section was three, one and two for case-control studies respectively and two for each section for all other study types. Final NOS score for each study ranged from 0 (lowest quality) to 6 (highest quality). Studies with a score of 0–2 were identified as low quality, 3–4 as medium quality and those between 5 and 6 considered to be of high quality. Discrepancies in the quality assessment were resolved by consensus.



Fig. 1. PRISMA flowchart of Identification and Selection of studies based on the eligibility criteria.

#### Table 1

Classification of identified blood fluid biomarkers based on NVU component.

NVU Component	Blood Biomarker	Number of VCD Studies		Number of Cerebrovascular Studies				
		Plasma	Serum	Plasma	Serum			
BBB Permeability	Q-Alb*	CSF/Plasma: 2	CSF/Serum: 4 [23,25,		CSF/Serum:1 [55]			
		[22,24]	26,87]					
Endothelial	vWF	4 [67,72-74]	-	14 [27-30,36,110-118]	-			
Dysfunction	PECAM-1	-	-	1 [116]	1 [119]			
	Endothelin-1	-	_	1 [76]	-			
	ICAM-1	2 [67,68]	1 [120]	12 [36,38,41,43,113,116,117,121-125]	5 [39,42,64,126,127]			
	VCAM-1	2 [37,67]	1 [66]	11 [36,38,40,41,46,113,122-125,128]	6 [37,39,42,64,126,129]			
	E-selectin	2 [37,67]	_	15 [27,28,36,38,40,41,45-47,65,86,	6 [37,39,42,44,64,129]			
				116,122,124,125]				
	P-selectin	1 [68]	-	9 [28,40,45,47,86,111,114,116,128]	2 [42,64]			
	VEGF	-	1 [70]	6 [31,86,113,130-132]	12 [32-35,51,64,133-138]			
Tight junction	VE-Cadherin	-	_	-	1 [139]			
proteins	Claudin-5	-	-	-	4 [9,138,140,141]			
	Occludin	-	-	-	7 [9,79,138,140-143]			
	ZO-1	-	-	-	3 [9,79,140]			
Neuronal Injury	NSE	-	2 [91,144]	3 [54,112,114]	8 [32,52,53,138,145-148]			
	NfL	-	3 [105–107]	4 [46,149-151]	7 [11,48,49,152-155]			
	NFH	-	_	-	2 [156,157]			
	Total Tau	-	_	1 [158]	1 [147]			
	Neurogranin	-	-	1 [158]	-			
	Neuronal	-	1 [159]	-	-			
	Pentraxin 2							
Neuroglial Injury	Lipocalin 2	2 [94,95]	-	-	-			
	GFAP	-	1 [106]	-	2 [93,134]			
	S100β	-	2 [91,108]	2 [46,112]	12 [32,33,50,51,129,134,138,145,			
					147,156,160,161]			
	Galectin-3	-	1 [162]	-	-			
	MgAb	-	1 [163]	-	-			
Extracellular Matrix	MMP-2	-	1 [109]	2 [86,123]	-			
	MMP-3	-	-	1 [123]	1 [135]			
	MMP-9	1 [85]	-	8 [55,86,112,122-124,164,165]	5 [64,79,134,135,138]			

\* Both CSF and plasma samples are required for the measurement of the CSF/Plasma albumin quotient. The numbers represent the quantity of studies, with the respective references in superscript.

# Results

# Search results and study characteristics

The search strategy from four databases yielded 2796 results (Fig. 1). An additional 10 studies were included from reviews and manual searching. We screened 2806 titles and abstracts and reviewed 359 full-text articles, finally including 112 studies. In total, 27 biomarkers were identified as indicators of NVU dysfunction (Table 1). Five out of the 26 studies met the inclusion criteria for VCD and were included in the metaanalysis (Fig. 1& 2), while the remaining 21 were included in the qualitative synthesis (Fig. 1& Supplementary Table S3). Eighty-six studies on cerebrovascular disorders were shortlisted out of which, 29 were meta-analysed and 57 summarised qualitatively (Fig. 1). A list of all included studies is provided in the Supplementary section (Supplementary Tables S3–5, S8).

#### Quality assessment and publication bias

Assessment of technical aspects of the available data, such as quality of individual manuscripts, publication bias and qualitative analyses are provided in Supplementary Tables S6–7.

#### Quantitative analysis

For those studies reporting quartiles, means and SDs were estimated assuming both normally and non-normally distributed outcome variables. Some biomarkers were found to have skewed distributions

Q-Alb Author(s) (Year)	N1:N2	Mean1(SD1)	Mean2(SD2)	Weight (%)								MD [95%CI]
Hagnelius et al., 2008	44:73	9.71(4.39)	6.83(2.35)	17.9%			ŀ					2.88 [1.48, 4.28]
Kettunen et al., 2022	30:45	7.90(2.60)	6.30(2.30)	22.0%			-					1.60 [0.45, 2.75]
Skillback et al., 2017	255:292	8.90(8.20)	5.90(2.20)	24.0%								3.00 [1.96, 4.04]
Janelidze et al., 2017	28:65	8.70(3.70)	6.30(2.90)	16.1%								2.40 [0.86, 3.94]
Musaeus et al., 2020	48:34	9.45(3.89)	5.47(1.87)	20.0%				,				3.98 [2.71, 5.25]
RE Model (Q = 7.93, df = 4,	, p = 0.09; l <sup>2</sup> = 5	$i0.0\%, \tau^2 = 0.41)$							-			2.77 [1.97, 3.57]
					0	1	2	3	4	5	6	

Fig. 2. Meta-analysis plot for CSF-Serum albumin quotient (Q-Alb) in vascular neurocognitive disorders (VCD).

Abbreviations - N1: Number of case subjects; N2: Number of control subjects; Mean 1: Mean of case subjects; SD 1: Standard deviation of case subjects; Mean 2: Mean difference of controls; SD 2: Standard deviation of controls.

The summary results within parentheses followed by the RE model correspond to the test of heterogeneity among the studies. P-value for significance of the MD is shown in supplementary table S9.

(Supplementary Table S8). Both MD and SMD measures were metaanalysed, with comprehensive results including p values reported within the text shown in supplementary tables S9-S10. The metaanalysis of MD with means and SDs estimated assuming non-normal distribution are shown in Forest plots (Figs. 2 and 3).

#### Meta-analysis of VCD biomarkers of BBB integrity

The Q-Alb quotient was the only biomarker of VCD with sufficient replicate studies (n = 5) that it could be meta-analysed (Supplementary Table S9 and Fig. 2). Q-Alb quotient was upregulated in VCD patients (n = 405) relative to healthy controls (n = 509) [22–26] (MD = 2.77; 95 % CI = 1.97–3.57; p < 0.0001, moderate heterogeneity across studies;  $I^2 = 50.0$  %).

#### Meta- analysis of NVU component biomarkers in cerebrovascular disorders

#### Endothelial dysfunction

Four of 14 studies on von Willebrand factor (vWF), a glycoprotein crucial to hemostasis, involving stroke and healthy controls were metaanalysed (Fig. 3.1A). The vWF levels were significantly upregulated in people with cerebrovascular disease compared to healthy controls (370 stroke and 3513 controls; MD = 21.77; 95% CI = 12.07 to 31.48; p = <0.001, with moderate heterogeneity across studies;  $I^2 = 58.0\%$ ) [27–30]. Five of 18 studies on vascular endothelial growth factor (VEGF), a master regulator of angiogenesis, were meta-analysed (Fig. 3.1B), with VEGF elevated in stroke patients (n = 365) relative to healthy controls (n = 315) (MD = 132.08; 95% CI = 47.75–216.41; p = 0.002,  $I^2 = 99.4$ %) [31–35].

A) vWF											
Author(s) (Year)	N1:N2	Mean1(SD1)	Mean2(SD2)	Weight (%)						MD [95	%CI]
Lavallee et al., 2013	59:59	147.88(52.35)	123.88(37.88)	19.5%						24.00 [ 7.51	1, 40.49]
Wannamethee et al., 2013	192:3219	150.20(50.10)	136.80(45.10)	35.7%		•				13.40 [ 6.14	4, 20.66]
Cherian et al., 2003	90:205	183.80(69.23)	164.10(69.71)	18.6%			· · ·			19.70 [ 2.51	1, 36.89]
Nadar et al., 2005	29:30	150.00(29.00)	117.00(17.00)	26.2%						33.00 [20.82	2, 45.18]
RE Model (Q = 7.71, df = 3	. p = 0.05; l <sup>2</sup>	= 58.0%, τ <sup>2</sup> = 54.95	i)							21.77 [12.0]	07, 31.48]
						1	1			i	
				0		10	20		30	50	
B) VEGF											
Author(s) (Year)	N1:N2	Mean1(SD1)	Mean2(SD2)	Weight (%)		1				MD [959	%CI]
Zheng et al., 2017	82:78	149.03(44.97)	83.66(15.45)	23.0%		<b>⊢∎</b> -i				65.37 [ 55.05	5, 75.69]
Golubev et al., 2019	43:20	350.36(193.81)	323.28(305.20)	) 13.7%						27.09 [-118.68,	8, 172.85]
Matsuo et al., 2013	171:171	569.00(13.00)	471.00(13.00)	23.1%		•				98.00 [ 95.24,	, 100.76]
Slevin et al., 2000	29:26	410.00(71.00)	245.00(28.00)	22.5%			<b></b>			165.00 [ 137.01,	, 192.99]
Klimenko et al., 2016	40:20	487.70(297.84)	185.31(66.56)	17.7%						302.39 [ 205.59,	), 399.19]
RE Model (Q = 77.34, df =	4, p < .01; l <sup>2</sup>	= 99.4%, τ <sup>2</sup> = 8015.	40)							132.08 [ 47.75,	i, 216.41]
				-200		0 10	n 200	300	400		
				200			200		-100		
C) VCAW-1	N11-N12	Moon1(SD1)	Maan2(SD2)	Woight (%)						MD IOF	0/ CII
Author(s) (Tear)	INT.INZ	Wearr(SDT)	meanz(SDZ)	weight (%)						MD [95	70CIJ
Rouhl et al., 2012	48:43	756.55(213.96)	644.41(105.90)	15.6%				-		112.14 [ 43.84,	, 180.45]
Zuliani et al., 2008	40:30	719.84(59.36)	564.74(50.24)	18.2%					<b>⊢</b> ∎	155.10 [ 129.38,	, 180.82]
Janes et al., 2019	35:35	561.70(175.10)	572.20(208.70)	13.9%				•		-10.50 [-100.75	5, 79.75]
Zhang et al., 2020	35:33	343.50(181.70)	151.75(58.21)	16.0%					<b>⊢−−−−</b>	191.75 [ 128.36,	, 255.14]
Prugger et al., 2013	95:190	454.06(127.46)	451.27(117.12)	18.0%				-		2.79 [ -27.78	8, 33.36]
Abdulle et al., 2010	40:42	296.03(331.85)	413.62(956.26)	3.7%			•			-117.59 [-424.53,	, 189.35]
Selakovic et al., 2003	15:15	687.24(156.96)	550.00(50.00)	14.5%					<b>_</b>	137.24 [ 53.88,	, 220.60]
RE Model (Q = 73.07, df =	6, p < .01; l*	= 89.8%, τ° = 6112.	25)	<b></b>	1					90.98 [ 24.64,	, 157.31]
				-500	-400	-200	)	0	200	300	
D) ICAM-1											
Author(s) (Year)	N1:N2	Mean1(SD1)	Mean2(SD2)	Weight (%	)		1			MD [95	5%CI]
Fassbender et al., 1999	64:67	280.00(40.00)	255.00(122.78	3) 13.5%						25.00 [ -5.99	9, 55.99]
Rouhl et al., 2012	48:43	516.58(128.69)	528.36(70.15	) 11.1%	,					-11.78 [-53.79	9, 30.23]
Abdulle et al., 2010	40:42	119.88(25.74)	76.14(51.10)	16.4%						43.74 [ 26.35	5, 61.13]
Prugger et al., 2013	95:190	313.23(101.02)	284.27(71.57	) 15.3%						28.96 [ 6.24	4, 51.68]
Zhang et al., 2020	35:33	93.99(55.94)	30.51(15.83)	16.0%					L	63.48 [ 44.18	8, 82.78]
Hassan et al., 2003	110:50	425.30(163.62)	341.90(77.18	) 12.1%						83.40 [ 46.08,	8, 120.72]
Janes et al., 2019	35:35	217.70(41.20)	213.80(49.50	) 15.6%						3.90 [-17.44	4, 25.24]
RE Model (Q = 29.22, df =	6, p < .01; l <sup>2</sup>	= 82.1%, τ <sup>2</sup> = 705.7	8)							34.54 [ 12.29	9, 56.78]
				-100	-50		0	50	100	150	

#### Fig. 3.1. Meta-analyses of biomarkers in cerebrovascular disorders.

Abbreviations – N1: Number of case subjects; N2: Number of control subjects; Mean 1: Mean of case subjects; SD 1: Standard deviation of case subjects; Mean 2: Mean difference of controls; SD 2: Standard deviation of controls.

The summary results within parentheses followed by the RE model correspond to the test of heterogeneity among the studies. P-value for significance of the MD is shown in supplementary table S9.

A) E-Selectin													
Author(s) (Year)	N1:N2	Mean1(SD1)	Mean2(SD2)	Weight (%)									MD [95%CI]
Nielsen et al., 2020	31:20	10.36(4.52)	9.03(6.15)	11.7%		•							1.34 [-1.79, 4.47]
Kozuka et al., 2002	52:86	54.90(30.10)	39.05(13.83)	8.6%		-							15.85 [ 7.16, 24.54]
Zuliani et al., 2008	40:30	49.78(11.64)	46.73(11.20)	10.6%		-	_						3.06 [-2.34, 8.45]
Fassbender et al., 1999	64:67	49.05(17.28)	43.00(24.56)	9.5%									6.05 [-1.20, 13.30]
Rouhl et al., 2012	48:43	33.44(27.23)	30.13(11.70)	8.7%		-							3.31 [-5.15, 11.77]
Zhang et al., 2020	35:33	6.28(4.25)	1.32(0.91)	12.3%									4.96 [ 3.52, 6.40]
Prugger et al., 2013	95:190	43.58(32.59)	22.17(14.29)	9.7%				-	•				21.41 [14.55, 28.27]
Lanes et al., 2003	90:205	37 70(16 30)	22.40(21.06)	11.0%		-							6 00 [-1 03 13 03]
Selakovic et al., 2003	15:15	51.13(17.57)	30.00(5.00)	8.3%									21.13 [11.89, 30.37]
	10.10	,	,	0.070									
RE Model (Q = 56.93, df = 9, p < .01; l <sup>2</sup> = 89.3%, r <sup>2</sup> = 44.14)												9.20 [ 4.62, 13.79]	
			-1	0	0		10		20	30	40		
B) P-Selectin													
Author(s) (Year)	N1:N2	Mean1(SD1)	Mean2(SD2)	Weight (%)									MD [95%CI]
Janes et al., 2019	35:35	19.90(0.90)	20.10(1.00)	28.5%		Ŵ							-0.20 [ -0.65, 0.25]
Rouhl et al., 2012	48:43	73.93(134.58)	66.60(43.11)	9.1%	,								7.33 [-32.87, 47.52]
Kozuka et al., 2002	52:86	52.64(23.80)	26.01(7.40)	26.9%									26.63 [ 19.97, 33.28]
Pletsch-Borba et al., 2020	335:2418	45,89(64,48)	27,84(16,40)	26.8%			— <b>—</b> —						18.05 [ 11.11, 24.98]
Cherian et al., 2003	90:205	322.20(178.09)	267.30(140.88)	8.7%							-		54.90 [ 13.36, 96.44]
RE Model (Q = 95.13, df =	4, p < .01; l <sup>2</sup> =	94.4%, τ <sup>2</sup> = 198.17)	,			-							17.40 [ 2.67, 32.12]
						1			1				
				-40		0			50		100		
C) NfL													
Author(s) (Year)	N1:N2 Mea	an1(SD1)	Mean2(SD2)	Weight (%)									MD [95%CI]
Nielsen et al., 2020	31:29 53	.10(71.38)	15.95(11.94)	46.8%									37.16 [ 11.65, 62.66]
Gattringer et al., 2017	79:53 109	.33(161.91)	37.22(18.76)	30.1%			,						72.11 [ 36.05, 108.17]
Ahn et al., 2022	15:8 45	60(84.92)	14.22(1.95)	23.1%									31.38 [-11.62, 74.37]
RE Model (O = 2.92 df = 3	$2 n = 0.23 I^2 =$	$30.5\% t^2 = 133.94$	()			_			_				46 33 [ 22 97 69 69]
	L, p = 0.20, 1 =	00.070, 1 - 100.047	1		i			1		1			40.00 [ 22.07, 00.00]
			-50		0			50		100		150	
D) S100B													
Author(s) (Year)	N1:N2	Mean1(SD <sup>2</sup>	1) Mean	2(SD2)	Weight (%	6)							MD [95%CI]
Nielsen et al., 2020	31:29	0.05(0.00)	0.04	(0.00)	25.1%	-							0.01 [0.01, 0.01]
Skalny et al., 2017	30:30	0.11(0.01)	0.08	(0.01)	25.0%								0.03 [0.02, 0.03]
Petzold et al., 2008	33:20	0.10(0.01)	0.02	(0.00)	25.0%						<b>-</b>		0.07 [0.07, 0.08]
Klimenko et al., 2016	40:20	0.10(0.01)	0.08	0.01)	25.0%			-					0.02 [0.02, 0.02]
RE Model (Q = 1049.31, d	lf = 3, p < .01; l <sup>2</sup>	$= 99.7\%, \tau^2 = 0.00)$											0.03 [0.01, 0.06]
									1	1	1		
						U	0.02		0.04	0.06	0.08		
E) NSE													
Author(s) (Year)	N1:N2	Mean1(SD1) Me	ean2(SD2) We	eight (%)									MD [95%CI]
Gojska-Grymajlo et al., 20	18 67:15	25.01(19.64) 1	2.39(7.97) 2	3.1%				•					12.62 [ 6.42, 18.82]
Bharosay et al., 2012	150:101	22.68(7.69)	7.48(1.52) 3	8.1%				-	<b>—</b>				15.20 [13.93, 16.47]
Selakovic et al., 2005	55:16	12.67(2.58)	4.48(0.89) 3	8.8%	-								8.19 [7.38, 9.00]
RE Model (Q = 84,05. df =	2. p < .01; l <sup>2</sup> =	96.2%, $\tau^2 = 14.38$ )											11.89 [ 7.23, 16.54]
									1				
1				5			0		15		20		

#### Fig. 3.2. Meta-analyses of biomarkers in cerebrovascular disorders.

Abbreviations – N1: Number of case subjects; N2: Number of control subjects; Mean 1: Mean of case subjects; SD 1: Standard deviation of case subjects; Mean 2: Mean difference of controls; SD 2: Standard deviation of controls.

The summary results within parentheses followed by the RE model correspond to the test of heterogeneity among the studies. P-value for significance of the MD is shown in supplementary table S9.

# Neuroglial and neuronal injury

Patients with cerebrovascular diseases showed increased levels of vascular cellular adhesion molecule-1 (VCAM-1) (7 studies; 308 stroke patients and 388 neurologically normal controls; MD = 90.98; 95 % CI = 24.64 to 157.31; p = 0.007; I<sup>2</sup> = 89.8 %) (Fig. 3.1C) [36–42] as well as upregulated intercellular adhesion molecule-1 (ICAM-1) levels (427 subjects and 460 controls; MD = 34.54; 95 % CI = 12.29–56.78; p = 0.002; I<sup>2</sup> = 82.1 %) (Fig. 3.1D) [36,38-40,42-44]. Blood levels of E-selectin (10 studies; 505 subjects with stroke and 724 controls; MD = 9.20; 95 % CI = 4.62–13.79; p < 0.001, I<sup>2</sup> = 89.3 %) (Fig. 3.2A) [28,36, 37,39-42,44-46] and P-selectin (5 studies; 560 subjects with stroke or SVD and 2787 controls; MD = 17.4; 95 % CI = 2.67–32.12; p = 0.02, I<sup>2</sup> = 94.4 %) were increased in cerebrovascular diseases compared to healthy controls [28,40,42,45,47] (Fig. 3.2B).

NfL, a major component of the neuronal cytoskeleton, was significantly higher in subjects with stroke or SVD (3 studies; 125 subjects) compared to 90 healthy controls (MD = 46.33; 95 % CI = 22.97–69.69; p = 0.0001;  $I^2 = 30.5$  %) [46,48,49](Fig. 3.2C).

Four studies analysed differences in S100 calcium-binding protein B (S100B), a commonly used astrocyte marker, levels between 134 patients with stroke and 99 healthy controls. S100B levels were elevated in stroke patients relative to controls (MD = 0.03; 95 % CI = 0.01-0.06; p =

0.02, I<sup>2</sup> = 99.7 %) (Fig. 3.2D) [33,46,50,51]. Three studies comparing circulating neuron specific enolase (NSE) concentrations, an enzyme released from neurons during injury, between 272 stroke patients and 132 controls, and reported significantly upregulated levels in stroke subjects (MD =11.89; 95 % CI = 7.23–16.54; p < 0.001, I<sup>2</sup> = 96.2 %) (Fig. 3.2E) [52–54].

#### Publication bias

Statistical examination using the Egger's regression test revealed no publication bias for the list of studies included in the final meta-analysis (Supplementary Table S9, column 0).

### Discussion

This review summarises the evidence on biomarkers associated with NVU pathology and identifies altered markers in patients with VCD or cerebrovascular disorders such as stroke. The data indicates a diversity of blood biomarkers associated with cerebrovascular disease with or without cognitive impairment. The biomarkers can be theoretically linked to multiple disease mechanisms, including increased BBB permeability caused by endothelial dysfunction or disruption of tight



#### Fig. 4. Overview of the NVU and its associated blood biomarkers.

The NVU is comprised of endothelial cells, a basement membrane, pericytes, glial cells and neurons. It is postulated that hypoperfusion and reduced cerebral blood flow arising from vascular pathologies trigger a sequence of events, such as the release of ROS which can activate pericytes and astrocytes to secrete MMPs as well as other cellular components. MMPs have been linked to the degradation of the basement membrane and junction proteins, which can be detected in the brain tissue or peripheral circulation. Additionally, ischemic conditions can disrupt pericyte-endothelium interaction causing the release of soluble PDGFR $\beta$  and disruption of junction proteins. Activated pericytes release MMP-9 further contributing to the breakdown of the BBB. Increase in activated MMPs, ROS and cytokines can potentially induce neuronal dysfunction and damage. With the disruption of the BBB, blood proteins such as albumin enter the brain tissue and CSF. Additionally, proteins indicative of neuronal injury (NfL, NfH, NSE), synaptic dysfunction (neurogranin, neuronal pentraxin-2), astrocyte injury (S100B, GFAP and lipocalin-2) and pericyte dysfunction (sPDGFR $\beta$ ) may enter the bloodstream. In response to inflammation, endothelial cells release various molecules such as VCAM-1, ICAM-1, Pselectin, E-selectin, endothelin, VEGF and vWF into the blood. Most of the derived biomarkers apart from Tau, microglial antibodies and galectin-3 are included here. Image created by first author GKH using the program BioRender.com.

and adherens junctions, as well as injury to neurons, neuroglia and the interstitial matrix (Table 1, Fig. 4).

# BBB integrity

The meta-analysis of plasma Q-Alb levels, as a marker of increased BBB permeability, showed an increase in VaD, subcortical small vessel disease and CADASIL patients when compared to healthy individuals [22–26] and were found to correlate with poor neurological outcomes in stroke [55]. Q-Alb levels were also elevated in other neurodegenerative disorders such as AD, Parkinson's disease dementia and frontotemporal dementia [24], but one study found that Q-Alb levels were highest in participants with VaD and mixed dementia compared to those with AD [25]. Furthermore, this association was not observed in AD cohorts without vascular risk factors [56,57]. Q-Alb may therefore be a useful biomarker of a compromised BBB and, by inference, NVU integrity, but more research is needed to determine its efficacy as a marker for differential diagnosis. Even though, it is the most widely used biomarker for BBB permeability, it is important to note that the calculation of Q-Alb levels requires the analysis of both serum and CSF samples.

Interestingly, Q-Alb has been found to correlate with sPDGFR $\beta$  levels [58,59]. Shed by pericytes, sPDGFR $\beta$  is another valuable biomarker of

NVU disruption given its association with cognitive dysfunction in the CSF and serum of AD subjects [60,61].

The review also identified blood biomarkers of endothelial cells, junction proteins and neuronal injury, all key components of the NVU. While the evidence for most of these markers is limited, they may serve as potential avenues for future work to establish blood biomarkers for VCD in relation to other brain diseases.

#### Endothelial dysfunction

Significant elevation of all cell adhesion molecules (CAMs) in cerebrovascular disorders, (Fig. 3.1, Supplementary Table S8) were observed in the meta-analyses when MD was analysed. CAMs are essential for mediating leukocyte recruitment to sites of tissue damage across the vascular endothelium and can be released as soluble forms into the bloodstream as part of a feedback mechanism [62]. Studies that measured multiple CAMs found only one protein family, either VCAM-1, ICAM-1 or E-selectin to be associated with severity of SVD or lower cognitive scores (Supplementary Table S5). The association between ICAM-1 and poor cognition was particularly noteworthy being observed in a longitudinal study that followed up with participants for 16 years [63]. Furthermore, increased expression of CAMs in cerebrovascular pathologies correlate with imaging markers such as lacunes (small areas of brain tissue damage/small subcortical infarcts), white matter hyperintensities (WMH) [64] and cerebral microbleeds suggesting potential use of these markers as indicators of cognitive impairment [65]. However, levels of these proteins in the blood of VCD and cerebrovascular disorders have been inconsistent. For example, VCAM-1 and E-selectin have been linked to poorer cognition and VCD in some studies [66,67] but the association with ICAM-1 and P-selectin is conflicting [68] (Supplementary Table S3).

Our meta-analysis also found upregulated blood vWF and VEGF levels in cerebrovascular disorders compared to healthy controls, with a large effect size for VEGF (Fig. 3.1). VEGF is a protein produced by vascular cells and neurons and is known to be a master regulator of angiogenesis involved in growth, maintenance and vascular repair; furthermore, VEGF mediates neuronal survival [31,69]'. One study found blood VEGF to be associated with VaD incidence [70] however, increased levels of circulating VEGF could also be a reflection of ageing [71]. Similarly, vWF levels were able to distinguish VCD subjects from healthy controls [72,73] and positively correlated with poor cognition and WMH [74]. Plasma vWF is derived mainly from endothelial cells rather than platelets; therefore, vWF is a gold standard measurement of endothelial damage [45]. Other less commonly studied proteins, such as PECAM-1 (CD31), which has been linked to worsening neurological outcome and endothelin-1, a predictor of VaD in cognitively intact adults [75], were also upregulated in the serum of stroke patients [76].

# Tight and adherens junction proteins

Interestingly, qualitative analysis of results from our search found less commonly studied blood biomarkers such as the tight junctions (TJs) and adherens junctions to be present in the peripheral circulation of humans (Supplementary Table S5). Functional characteristics of TJs are attributed to various proteins, namely the claudins, occludin and cytoplasmic zonula occludens (ZO), which are responsible for regulating the permeability of endothelial cells in the cerebrovasculature. Disruption of these junctions may contribute to increased BBB leakage [77]. Claudin-5, occludin and ZO-1 were each upregulated in stroke patients compared to healthy controls. Specifically, increased levels of claudin-5 and ZO-1 were associated with worsening outcomes as a result of haemorrhagic transformation, while increased occludin levels were associated with severe cognitive impairment (Supplementary Table S5). The strength of correlation analyses between some of these proteins and stroke outcomes have been inconsistent in other studies. VE-cadherin. an adherens junction protein involved in regulating the adhesion of brain endothelial cells [78], was significantly upregulated in the blood of stroke patients [79], congruent with a recent study, where VE-cadherin levels were seen to be reduced in the brain and increased in the CSF of preclinical AD patients [80]. Though there are currently no studies measuring the levels of junction proteins in the blood of humans with VCD, BBB impairment resulting from degradation of TJs has been reported in endothelial cell culture models [81] and animal models of hypoxia-ischemia [82]. Analysing these structural proteins may provide possible insights into the pathophysiology of cognitive impairment associated with vascular pathologies as their degradation is an early indication of NVU dysfunction. The anatomical location of TJs and adherens junctions between adjacent endothelial cells of the BBB, may pose a challenge for their measurement in the systemic circulation. However, with the utilization of sensitive and specific assays, they may provide a novel avenue for monitoring the prognosis of vascular contributions to cognitive impairment.

# Extracellular matrix proteins

Increased permeability of the BBB can be attributed to MMP related proteolytic degradation of the basal lamina and TJs [83]. MMP-9 is activated in hypoxic conditions [84], found to be elevated in VCD [85] and stroke patients [86] as well as associated with cognitive decline and worsening neurological outcomes. Though elevated blood MMP-2 levels have also been reported in VCD subjects with multiple infarcts and stroke [87], results for MMP-3 levels in cerebrovascular disorders were conflicting (Supplementary Table S5).

#### Neuroglial and neuronal injury

Findings from the meta-analysis revealed that subjects with cerebrovascular disorders have higher levels of NfL and NSE in their blood than do healthy controls (Fig. 3.2). NfL levels also correlated with neuroimaging lesions such as lacunes, microbleeds as well as, cognitive function in VCD and SVD. Notably, serum NfL successfully predicted 7year changes in cognition in patients with a pure form of VaD. Similarly, a recent meta-analysis of 19 studies in 4237 patients with cerebrovascular events by Sanchez, Martirosian [88] found blood NfL levels to be a suitable diagnostic marker for distinguishing between cerebrovascular subtypes.

Qualitative evidence suggests similar patterns with neurofilament heavy chain (NfH) levels as it differentiated patients with neurological disorders from controls and correlated with infarct volume (Supplementary Table S5). There are fewer studies on NfH levels in VCD, but elevated levels have been seen in the CSF of VaD and AD patients in comparison to controls [89]. Neurofilaments with light, intermediate, and heavy chains are important for the maintenance of the cytoskeleton and together with NSE, have been widely recognised as markers of neuronal injury. In the event of hypoxia-ischemia, these molecules leak out of neurons, pass through the BBB and enter the peripheral circulation [90]. A positive correlation between serum NSE and WMH with NSE blood levels is a predictive marker in the progression of cognitive impairment [91].

S100B, glial fibrillary acidic protein (GFAP) and lipocalin-2 are proteins expressed in astrocytes, the most abundant glial cell type, and is critical for a healthy NVU. Therefore, elevation of these proteins serve as possible indicators of astrocyte injury [92]. A meta-analysis of studies revealed that S100B is upregulated in cerebrovascular disorders compared to controls (Fig. 3.2). S100B is not normally present in the peripheral circulation but is released following stroke and has been commonly implicated in brain injury. Elevated levels of serum GFAP have been found in ischemic stroke [93] and intracranial haemorrhage <sup>e</sup> [65] with lipocalin-2 increased in the blood of VaD [94] and AD patients [95]. Though activated astrocytes have been reported to contribute to the breakdown of the BBB [96], the exact pathogenic mechanisms involving lipocalin-2 and its diagnostic capability in cognitive impairment is unclear.

Many of these biomarkers were analysed in relatively few studies and with small sample sizes. As biomarkers such as GFAP and NfL are elevated in various brain disorders, their specificity for neurodegeneration needs to be evaluated in greater detail. The issue of whether NfL can be considered a primary marker of NVU integrity remains unresolved. As a major constituent of the neuro-axonal cytoskeleton, elevated NfL is typically reported in the CSF as a reflection of axonal damage, either primary or secondary to processes such as Wallerian degeneration [97-99]. Presence of elevated levels of NfL in the blood may therefore be a downstream effect of compromised BBB integrity, as has been reported in several brain diseases [100-103]. By contrast, others have failed to identify an association between NfL and BBB permeability [104]. Therefore, altered fluid levels of NfL in VCD, as identified in this review, cannot be regarded as a specific marker of NVU integrity. Further studies are warranted to validate the efficacy of these proteins as useful markers of BBB-related pathology and cognitive decline in the periphery.

#### Small vessel disease and cognitive impairment

There are only a limited number of studies investigating biomarkers of SVD and cognitive impairment. Out of these studies, vWF [67,72-74],

NSE [105], NfL [88,105-107], S100 $\beta$  [91,108] and Lipocalin-2 [94,95] appeared to be the most robust biomarkers across studies .

#### Strengths and limitations

To our knowledge, this is the first systematic review and metaanalysis on blood biomarkers pertaining to NVU pathology in both VCD and cerebrovascular disease. This study accounted for selection bias by having two independent reviewers perform screening of initial results. In addition, the extraction of eligible studies from reference lists maximised the inclusion of relevant studies. Various statistical tests were conducted using both SMD and MD measures (Supplementary Tables S8–10) before results were described based on non-normal estimation with MD measurements (Supplementary Table S9), therefore, allowing for the recognition of heterogeneity in biomarker levels. Also, there was limited disagreement (6%) among the three analysed domains when the quality of each study was assessed independently by two reviewers.

Our review has several limitations. Due to the limited number of studies related to NVU components in VCD subjects, we were unable to perform quantitative analysis to conclude statistical significance. Most of our meta-analyses have low study numbers which could have contributed to low statistical power. Some studies with wide deviations in measurements were removed from the meta-analyses. Additionally, the use of mean and SD values could have limited the inherent variance and not taken into account the skewness of the data. Some of the biomarkers were analysed in stroke cohorts but not in VCD patients. Hence, their actual potential as a diagnostic marker in VCD is not known. Despite exhaustive searches across four databases, we might have missed some eligible studies and data from a small number of studies had to be extrapolated from the bar graphs as there were no responses to requests for unprocessed data from authors. Apart from NfL and vWF, high heterogeneity was present in the analyses for all other biomarkers. This could have been attributed to a variety of factors including variability of fluid biomarkers, study subjects and sample size. It has also been argued that Egger's test on MD can lead to an overestimation of false positive outcomes [21]. Finally, as the biomarkers were analysed separately, it is unclear to what extent they are correlated.

#### Conclusion

Multiple blood biomarkers were quantitatively altered in VCD and cerebrovascular disorders. However, only nine biomarkers for cerebrovascular disorders and one for VCD could be meta-analysed, highlighting the limited research in VCD. Tight and adherens junctions (Claudin-5, Occludin and VE-cadherin), which have only been studied in animals and in a small number of human participants with stroke, should be analysed for their potential diagnostic role in VCD alongside other markers such as VEGF, NfL, vWF, Q-Alb and MMP-2/9. The diagnosis of VCD is likely to require a diverse panel of markers considering the heterogeneous nature of this disorder with multiple pathogenic mechanisms and more importantly, for the identification of various subtypes. In fact, one study found a panel of biomarkers associated with increased BBB permeability such as albumin quotient and MMP-2 amidst other inflammatory biomarkers to be the most predictive model for the diagnosis of subcortical ischemic vascular dementia [109]. As NVU is a hallmark of many neurological diseases, measuring its function could aid not only differential diagnosis but also prognosis and disease monitoring. Blood biomarkers of BBB permeability, pericytes and the integrity of endothelial cells, junction proteins, neurons and glia therefore deserve much greater enquiry.

# CRediT authorship contribution statement

**Gurpreet Kaur Hansra:** Writing – original draft, Project administration, Methodology, Formal analysis, Data curation, Conceptualization. Tharusha Jayasena: Writing – review & editing, Data curation. Satoshi Hosoki: Writing – review & editing, Data curation. Anne Poljak: Writing – review & editing. Ben Chun Pan Lam: Writing – review & editing, Formal analysis. Ruslan Rust: Writing – review & editing. Abhay Sagare: Writing – review & editing. Berislav Zlokovic: Writing – review & editing. Anbupalam Thalamuthu: Writing – review & editing, Formal analysis. Perminder S. Sachdev: Writing – review & editing, Supervision.

#### Declaration of competing interest

None.

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#### Supplementary materials

Supplementary material associated with this article can be found, in the online version, at doi:10.1016/j.cccb.2024.100216.

#### References

- M. Dichgans, D. Leys, Vascular cognitive impairment, Circ. Res. 120 (2017) 573–591, https://doi.org/10.1161/CIRCRESAHA.116.308426.
- [2] S. Hosoki, G.K. Hansra, T. Jayasena, et al., Molecular biomarkers for vascular cognitive impairment and dementia, Nature Rev. Neuro. 19 (2023) 737–753, https://doi.org/10.1038/s41582-023-00884-1.
- [3] C. Iadecola, The neurovascular unit coming of age: a journey through neurovascular coupling in health and disease, Neuron 96 (2017) 17–42, https:// doi.org/10.1016/j.neuron.2017.07.030.
- [4] B.V. Zlokovic, Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders, Nature Rev. Neurosci. 12 (2011) 723–738, https:// doi.org/10.1038/nrn3114.
- [5] E.G. Knox, M.R. Aburto, G. Clarke, et al., The blood-brain barrier in aging and neurodegeneration, Mol. Psychiatr. 27 (2022) 2659–2673, https://doi.org/ 10.1038/s41380-022-01511-z.
- [6] A. Montagne, S.R. Barnes, M.D. Sweeney, et al., Blood-brain barrier breakdown in the aging human hippocampus, Neuron 85 (2015) 296–302, https://doi.org/ 10.1016/j.neuron.2014.12.032.
- [7] Z. Nadareishvili, A.N. Simpkins, E. Hitomi, et al., Post-stroke blood-brain barrier disruption and poor functional outcome in patients receiving thrombolytic therapy, Cerebrovasc. Dis. 47 (2019) 135–142, https://doi.org/10.1159/ 000499666.
- [8] A.P. Sagare, M.D. Sweeney, J. Makshanoff, et al., Shedding of soluble plateletderived growth factor receptor-β from human brain pericytes, Neurosci. Lett. 607 (2015) 97–101, https://doi.org/10.1016/j.neulet.2015.09.025, 20150925.
- [9] A. Lasek-Bal, A. Kokot, D. Gendosz de Carrillo, et al., Plasma levels of occludin and claudin-5 in acute stroke are correlated with the type and location of stroke but not with the neurological state of patients-preliminary data, Brain Sci. 10 (2020) 20201109, https://doi.org/10.3390/brainsci10110831.
- [10] K. Makris, A. Haliassos, M. Chondrogianni, et al., Blood biomarkers in ischemic stroke: potential role and challenges in clinical practice and research, Crit. Rev. Clin. Lab. Sci. 55 (2018) 294–328, https://doi.org/10.1080/ 10408363.2018.1461190, 20180418.
- [11] S. Tiedt, A.M. Buchan, M. Dichgans, et al., The neurovascular unit and systemic biology in stroke — Implications for translation and treatment, Nature Rev. Neurol. 18 (2022) 597–612, https://doi.org/10.1038/s41582-022-00703-z.
- [12] S. Hosoki, T. Tanaka, M. Ihara, Diagnostic and prognostic blood biomarkers in vascular dementia: from the viewpoint of ischemic stroke, Neurochem. Int. 146 (2021) 105015, https://doi.org/10.1016/j.neuint.2021.105015, 20210326.
- [13] V. Cipollini, F. Troili, F. Giubilei, Emerging biomarkers in vascular cognitive impairment and dementia: from pathophysiological pathways to clinical application, Int. J. Mol. Sci. 20 (2019) 20190608, https://doi.org/10.3390/ ijms20112812.
- [14] A. Jagtap, S. Gawande, S. Sharma, Biomarkers in vascular dementia: a recent update, Biomark. Genom. Med. 7 (2015) 43–56, https://doi.org/10.1016/j. bgm.2014.11.001.
- [15] D. Moher, A. Liberati, J. Tetzlaff, et al., Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement, PLoS. Med. 6 (2009) e1000097, https://doi.org/10.1371/journal.pmed.1000097, 20090721.

- [16] M. Ouzzani, H. Hammady, Z. Fedorowicz, et al., Rayyan a web and mobile app for systematic reviews. systematic reviews, Syst. Rev. 5 (2016), https://doi.org/ 10.1186/s13643-016-0384-4.
- [17] X. Wan, W. Wang, J. Liu, et al., Estimating the sample mean and standard deviation from the sample size, median, range and/or interquartile range, BMC. Med. Res. Methodol. 14 (2014) 135, https://doi.org/10.1186/1471-2288-14-135.
- [18] J. Shi, D. Luo, X. Wan, et al., Detecting the skewness of data from the five-number summary and its application in meta-analysis, Stat. Methods Med. Res. 32 (2023) 1338–1360, https://doi.org/10.1177/09622802231172043.
- [19] S. McGrath, X. Zhao, R. Steele, et al., Estimating the sample mean and standard deviation from commonly reported quantiles in meta-analysis, Stat. Methods Med. Res. 29 (2020) 2520–2537, https://doi.org/10.1177/0962280219889080.
- [20] W. Viechtbauer, Conducting meta-analyses in r with the metafor package, J. Stat. Softw. 36 (2010) 1–48, https://doi.org/10.18637/jss.v036.i03.
- [21] M. Harrer, P. Cuijpers, T.A. Furukawa, et al., Doing Meta-Analysis With R: A Hands-On Guide, 1st ed, Chapman & Hall/CRC Press, Boca Raton, FL and London, 2021.
- [22] C.S. Musaeus, H.S. Gleerup, P. Høgh, et al., Cerebrospinal fluid/plasma albumin ratio as a biomarker for blood-brain barrier impairment across neurodegenerative dementias, J. Alzheimers. Dis. 75 (2020) 429–436, https://doi.org/10.3233/jad-200168.
- [23] T. Skillbäck, L. Delsing, J. Synnergren, et al., CSF/serum albumin ratio in dementias: a cross-sectional study on 1861 patients, Neurobiol. Aging 59 (2017) 1–9, https://doi.org/10.1016/j.neurobiolaging.2017.06.028, 20170711.
- [24] S. Janelidze, J. Hertze, K. Nägga, et al., Increased blood-brain barrier permeability is associated with dementia and diabetes but not amyloid pathology or APOE genotype, Neurobiol. Aging 51 (2017) 104–112, https://doi.org/ 10.1016/j.neurobiolaging.2016.11.017, 20161205.
- [25] N.O. Hagnelius, L.O. Wahlund, T.K Nilsson, CSF/serum folate gradient: physiology and determinants with special reference to dementia, Dement. Geriatr. Cogn. Disord. 25 (2008) 516–523, https://doi.org/10.1159/000129696, 20080507.
- [26] P. Kettunen, M. Bjerke, C. Eckerström, et al., Blood-brain barrier dysfunction and reduced cerebrospinal fluid levels of soluble amyloid precursor protein-β in patients with subcortical small-vessel disease, Alzheimer's. Dement. Diagnosis Assess. Dis. Monitoring 14 (2022) e12296, https://doi.org/10.1002/dad2.12296.
- [27] S.K. Nadar, G.Y. Lip, K.W. Lee, et al., Circulating endothelial cells in acute ischaemic stroke, Thromb. Haemost. 94 (2005) 707–712, https://doi.org/ 10.1160/th04-12-0795.
- [28] P. Cherian, G.J. Hankey, J.W. Eikelboom, et al., Endothelial and platelet activation in acute ischemic stroke and its etiological subtypes, Stroke 34 (2003) 2132–2137, https://doi.org/10.1161/01.Str.000086466.32421.F4, 20030807.
- [29] S.G. Wannamethee, A.G. Shaper, P.H. Whincup, et al., Adiposity, adipokines, and risk of incident stroke in older men, Stroke 44 (2013) 3–8, https://doi.org/ 10.1161/strokeaha.112.670331, 20121127.
- [30] P.C. Lavallée, J. Labreuche, D. Faille, et al., Circulating markers of endothelial dysfunction and platelet activation in patients with severe symptomatic cerebral small vessel disease, Cerebrovasc. Dis. 36 (2013) 131–138, https://doi.org/ 10.1159/000353671, 20130911.
- [31] R. Matsuo, T. Ago, M. Kamouchi, et al., Clinical significance of plasma VEGF value in ischemic stroke - research for biomarkers in ischemic stroke (REBIOS) study, BMC. Neurol. 13 (2013) 32, https://doi.org/10.1186/1471-2377-13-32.
- [32] A. Golubev, M. Petrova, A. Grechko, et al., Molecular markers of ischemic stroke, Gen. Reanimatol. 15 (2019) 11–22, https://doi.org/10.15360/1813-9779-2019-5-11-22.
- [33] L.L. Klimenko, A.V. Skalny, A.A. Turna, et al., Serum trace element profiles, prolactin, and cortisol in transient ischemic attack patients, Biol. Trace Elem. Res. 172 (2016) 93–100, https://doi.org/10.1007/s12011-015-0586-y, 20151214.
- [34] M. Slevin, J. Krupinski, A. Slowik, et al., Serial measurement of vascular endothelial growth factor and transforming growth factor-beta1 in serum of patients with acute ischemic stroke, Stroke 31 (2000) 1863–1870, https://doi. org/10.1161/01.str.31.8.1863.
- [35] J. Zheng, J. Sun, L. Yang, et al., The potential role of vascular endothelial growth factor as a new biomarker in severe intracerebral hemorrhage, J. Clin. Lab. Anal. 31 (2017) 20161220, https://doi.org/10.1002/jcla.22076.
- [36] C. Prugger, G. Luc, B. Haas, et al., Multiple biomarkers for the prediction of ischemic stroke: the PRIME study, Arterioscler. Thromb. Vasc. Biol. 33 (2013) 659–666, https://doi.org/10.1161/atvbaha.112.300109, 20130117.
- [37] G. Zuliani, M. Cavalieri, M. Galvani, et al., Markers of endothelial dysfunction in older subjects with late onset Alzheimer's disease or vascular dementia, J. Neurol. Sci. 272 (2008) 164–170, https://doi.org/10.1016/j.jns.2008.05.020, 20080702.
- [38] A.M. Abdulle, J.Y. Pathan, N. Moussa, et al., Association between homocysteine and endothelial dysfunction markers in stroke disease, Nutr. Neurosci. 13 (2010) 2–6, https://doi.org/10.1179/147683010X12611460763562.
- [39] X. Zhang, L. Wang, Z. Han, et al., KLF4 alleviates cerebral vascular injury by ameliorating vascular endothelial inflammation and regulating tight junction protein expression following ischemic stroke, J. Neuroinflammation 17 (2020) 107, https://doi.org/10.1186/s12974-020-01780-x.
- [40] R.P. Rouhl, J.G. Damoiseaux, J. Lodder, et al., Vascular inflammation in cerebral small vessel disease, Neurobiol. Aging 33 (2012) 1800–1806, https://doi.org/ 10.1016/j.neurobiolaging.2011.04.008, 20110523.
- [41] V. Selaković, M. Colić, M. Jovanović, et al., Cerebrospinal fluid and plasma concentration of soluble intercellular adhesion molecule 1, vascular cell adhesion molecule 1 and endothelial leukocyte adhesion molecule in patients with acute

ischemic brain disease, Vojnosanit. Pregl. 60 (2003) 139–146, https://doi.org/ 10.2298/vsp0302139s.

- [42] F. Janes, A. Cifù, M.E. Pessa, et al., ADMA as a possible marker of endothelial damage. A study in young asymptomatic patients with cerebral small vessel disease, Sci. Rep. 9 (2019) 14207, https://doi.org/10.1038/s41598-019-50778-
- [43] A. Hassan, B.J. Hunt, M. O'Sullivan, et al., Markers of endothelial dysfunction in lacunar infarction and ischaemic leukoaraiosis, Brain 126 (2003) 424–432, https://doi.org/10.1093/brain/awg040.
- [44] K. Fassbender, T. Bertsch, O. Mielke, et al., Adhesion molecules in cerebrovascular diseases. Evidence for an inflammatory endothelial activation in cerebral large- and small-vessel disease, Stroke 30 (1999) 1647–1650, https:// doi.org/10.1161/01.str.30.8.1647.
- [45] K. Kozuka, T. Kohriyama, E. Nomura, et al., Endothelial markers and adhesion molecules in acute ischemic stroke-sequential change and differences in stroke subtype, Atherosclerosis 161 (2002) 161–168, https://doi.org/10.1016/s0021-9150(01)00635-9.
- [46] H.H. Nielsen, C.B. Soares, S.S. Høgedal, et al., Acute neurofilament light chain plasma levels correlate with stroke severity and clinical outcome in ischemic stroke patients, Front. Neurol. 11 (2020) 448, https://doi.org/10.3389/ fneur.2020.00448, 20200611.
- [47] L. Pletsch-Borba, M. Grafetstätter, A. Hüsing, et al., Vascular injury biomarkers and stroke risk: a population-based study, Neurology. 94 (2020) e2337–e2345, https://doi.org/10.1212/wnl.00000000009391, 20200505.
- [48] J.W. Ahn, J. Hwang, M. Lee, et al., Serum neurofilament light chain levels are correlated with the infarct volume in patients with acute ischemic stroke, Medicine (Baltimore) 101 (2022).
- [49] T. Gattringer, D. Pinter, C. Enzinger, et al., Serum neurofilament light is sensitive to active cerebral small vessel disease, Neurology. 89 (2017) 2108–2114, https:// doi.org/10.1212/wnl.00000000004645, 20171018.
- [50] A. Petzold, P. Michel, M. Stock, et al., Glial and axonal body fluid biomarkers are related to infarct volume, severity, and outcome, J. Stroke Cerebrovasc. Dis. 17 (2008) 196–203, https://doi.org/10.1016/j.jstrokecerebrovasdis.2008.02.002.
- [51] A.V. Skalny, L.L. Klimenko, A.A. Turna, et al., Serum trace elements are associated with hemostasis, lipid spectrum and inflammatory markers in men suffering from acute ischemic stroke, Metab. Brain Dis. 32 (2017) 779–788, https://doi.org/10.1007/s11011-017-9967-6, 20170220.
- [52] A. Gójska-Grymajło, M. Zieliński, A. Wardowska, et al., CXCR7+ and CXCR4+ stem cells and neuron specific enolase in acute ischemic stroke patients, Neurochem. Int. 120 (2018) 134–139, https://doi.org/10.1016/j. neuint.2018.08.009, 20180817.
- [53] A. Bharosay, V.V. Bharosay, M. Varma, et al., Correlation of brain biomarker neuron specific enolase (NSE) with degree of disability and neurological worsening in cerebrovascular stroke, Indian J. Clin. Biochem. 27 (2012) 186–190, https://doi.org/10.1007/s12291-011-0172-9, 20111108.
- [54] V. Selakovic, R. Raicevic, L. Radenovic, The increase of neuron-specific enolase in cerebrospinal fluid and plasma as a marker of neuronal damage in patients with acute brain infarction, J. Clin. Neurosci. 12 (2005) 542–547, https://doi.org/ 10.1016/j.jocn.2004.07.019.
- [55] R. Brouns, A. Wauters, D. De Surgeloose, et al., Biochemical markers for bloodbrain barrier dysfunction in acute ischemic stroke correlate with evolution and outcome, Eur. Neurol. 65 (2011) 23–31, https://doi.org/10.1159/000321965.
- [56] K. Blennow, A. Wallin, C. Uhlemann, et al., White-matter lesions on CT in Alzheimer patients: relation to clinical symptomatology and vascular factors, Acta Neurol. Scand. 83 (1991) 187–193, https://doi.org/10.1111/j.1600-0404.1991.tb04675.x.
- [57] K. Blennow, A. Wallin, P. Fredman, et al., Blood-brain barrier disturbance in patients with Alzheimer's disease is related to vascular factors, Acta Neurol. Scand. 81 (1990) 323–326, https://doi.org/10.1111/j.1600-0404.1990.tb01563.
- [58] A. Montagne, D.A. Nation, A.P. Sagare, et al., APOE4 leads to blood-brain barrier dysfunction predicting cognitive decline, Nature 581 (2020) 71–76, https://doi. org/10.1038/s41586-020-2247-3.
- $[59] X. Lv, M. Zhang, Z. Cheng, et al., Changes in CSF sPDGFR\beta level and their association with blood-brain barrier breakdown in Alzheimer's disease with or without small cerebrovascular lesions, Alzheimers. Res. Ther. 15 (2023) 51,$ https://doi.org/10.1186/s13195-023-01199-5, 20230314.
- [60] J.S. Miners, P.G. Kehoe, S. Love, et al., CSF evidence of pericyte damage in Alzheimer's disease is associated with markers of blood-brain barrier dysfunction and disease pathology, Alzheimers. Res. Ther. 11 (2019) 81, https://doi.org/ 10.1186/s13195-019-0534-8, 20190914.
- [61] M.D. Sweeney, A.P. Sagare, M. Pachicano, et al., A novel sensitive assay for detection of a biomarker of pericyte injury in cerebrospinal fluid, Alzheimers. Dement. 16 (2020) 821–830, https://doi.org/10.1002/alz.12061, 20200416.
- [62] B.A. Kallmann, V. Hummel, T. Lindenlaub, et al., Cytokine-induced modulation of cellular adhesion to human cerebral endothelial cells is mediated by soluble vascular cell adhesion molecule-1, Brain 123 (Pt 4) (2000) 687–697, https://doi. org/10.1093/brain/123.4.687.
- [63] S.B. Rafnsson, I.J. Deary, F.B. Smith, et al., Cognitive decline and markers of inflammation and hemostasis: the Edinburgh artery study, J. Am. Geriatr. Soc. 55 (2007) 700–707, https://doi.org/10.1111/j.1532-5415.2007.01158.x.
- [64] D.D. Zhang, Y. Cao, J.Y. Mu, et al., Inflammatory biomarkers and cerebral small vessel disease: a community-based cohort study, Stroke Vasc. Neurol. 7 (2022) 302–309, https://doi.org/10.1136/svn-2021-001102, 20220308.

- [65] Z. Huang, Q. Yin, W. Sun, et al., Microbleeds in ischemic stroke are associated with lower serum adiponectin and higher soluble E-selectin levels, J. Neurol. Sci. 334 (2013) 83–87, https://doi.org/10.1016/j.jns.2013.07.2513, 20130802.
- [66] N. El Husseini, C. Bushnell, C.M. Brown, et al., Vascular cellular adhesion molecule-1 (VCAM-1) and memory impairment in African-Americans after small vessel-type stroke, J. Stroke Cerebrovasc. Dis. 29 (2020) 104646, https://doi.org/ 10.1016/j.jstrokecerebrovasdis.2020.104646, 20200214.
- [67] S.P. Rensma, T.T. van Sloten, A.J.H.M. Houben, et al., Microvascular dysfunction is associated with worse cognitive performance, Hypertension 75 (2020) 237–245, https://doi.org/10.1161/HYPERTENSIONAHA.119.13023.
- [68] J. Staszewski, E. Skrobowska, R. Piusińska-Macoch, et al., IL-1α and IL-6 predict vascular events or death in patients with cerebral small vessel disease—Data from the SHEF-CSVD study, Adv. Med. Sci. 64 (2019) 258–266, https://doi.org/ 10.1016/j.advms.2019.02.003.
- [69] R. Rust, Insights into the dual role of angiogenesis following stroke, J. Cereb. Blood Flow Metab. 40 (2020) 1167–1171, https://doi.org/10.1177/ 0271678X20906815, 20200216.
- [70] K. Trares, M. Bhardwaj, L. Perna, et al., Association of the inflammation-related proteome with dementia development at older age: results from a large, prospective, population-based cohort study, Alzheimers. Res. Ther. 14 (2022) 128, https://doi.org/10.1186/s13195-022-01063-y, 20220909.
- [71] A. Chakraborty, M. Chatterjee, H. Twaalfhoven, et al., Vascular endothelial growth factor remains unchanged in cerebrospinal fluid of patients with Alzheimer's disease and vascular dementia, Alzheimers. Res. Ther. 10 (2018) 58, https://doi.org/10.1186/s13195-018-0385-8.
- [72] N.O. Hagnelius, K. Boman, T.K. Nilsson, Fibrinolysis and von Willebrand factor in Alzheimer's disease and vascular dementia-a case-referent study, Thromb. Res. 126 (2010) 35–38, https://doi.org/10.1016/j.thromres.2009.10.001, 20091029.
- [73] F. Pescini, I. Donnini, F. Cesari, et al., Circulating biomarkers in cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy patients, J. Stroke Cerebrovasc. Dis. 26 (2017) 823–833, https://doi.org/10.1016/j.jstrokecerebrovasdis.2016.10.027, 20161118.
- [74] S. Kuipers, L.M. Overmars, B. van Es, et al., A cluster of blood-based protein biomarkers reflecting coagulation relates to the burden of cerebral small vessel disease, J. Cereb. Blood Flow Metab. 42 (2022) 1282–1293, https://doi.org/ 10.1177/0271678X221077339, 20220127.
- [75] H. Holm, K. Nägga, E.D. Nilsson, et al., Biomarkers of microvascular endothelial dysfunction predict incident dementia: a population-based prospective study, J. Intern. Med. 282 (2017) 94–101, https://doi.org/10.1111/joim.12621, 20170504.
- [76] V. Sapira, I.M. Cojocaru, G. Lilios, et al., Study of endothelin-1 in acute ischemic stroke, Rom. J. Intern. Med. 48 (2010) 329–332.
- [77] J.J. Lochhead, J. Yang, P.T. Ronaldson, et al., Structure, function, and regulation of the blood-brain barrier tight junction in central nervous system disorders, Front. Physiol. 11 (2020) 914, https://doi.org/10.3389/fphys.2020.00914, 20200806.
- [78] W. Li, Z. Chen, I. Chin, et al., The role of VE-cadherin in blood-brain barrier integrity under central nervous system pathological conditions, Curr. Neuropharmacol. 16 (2018) 1375–1384, https://doi.org/10.2174/ 1570159X16666180222164809
- [79] Y. He, Q. Yang, H. Liu, et al., Effect of blood pressure on early neurological deterioration of acute ischemic stroke patients with intravenous rt-PA thrombolysis may be mediated through oxidative stress induced blood-brain barrier disruption and AQP4 upregulation, J. Stroke Cerebrovasc. Dis. 29 (2020) 104997, https://doi.org/10.1016/j.jstrokecerebrovasdis.2020.104997, 20200613
- [80] R. Tarawneh, R.S. Kasper, J. Sanford, et al., Vascular endothelial-cadherin as a marker of endothelial injury in preclinical Alzheimer disease, Ann. Clin. Transl. Neurol. 9 (2022) 1926–1940, https://doi.org/10.1002/acn3.51685, 20221107.
- [81] S. Page, A. Munsell, A.J Al-Ahmad, Cerebral hypoxia/ischemia selectively disrupts tight junctions complexes in stem cell-derived human brain microvascular endothelial cells, Fluids. Barriers. CNS. 13 (2016) 16, https://doi. org/10.1186/s12987-016-0042-1.
- [82] K. Toyama, J.M. Spin, A.C. Deng, et al., MicroRNA-mediated therapy modulating blood-brain barrier disruption improves vascular cognitive impairment, Arterioscler. Thromb. Vasc. Biol. 38 (2018) 1392–1406, https://doi.org/ 10.1161/ATVBAHA.118.310822.
- [83] F. Wang, Y. Cao, L. Ma, et al., Dysfunction of cerebrovascular endothelial cells: prelude to vascular dementia, Front. Aging Neurosci. 10 (2018), https://doi.org/ 10.3389/fnagi.2018.00376. Review.
- [84] S.M. Stamatovic, A.M. Johnson, R.F. Keep, et al., Junctional proteins of the bloodbrain barrier: new insights into function and dysfunction, Tissue Barriers 4 (2016) e1154641, https://doi.org/10.1080/21688370.2016.1154641, 20160226.
- [85] M. Gong, J. Jia, Contribution of blood-brain barrier-related blood-borne factors for Alzheimer's disease vs. vascular dementia diagnosis: a pilot study, Front. Neurosci. 16 (2022). Original Research.
- [86] A. Khan, A. Parray, N. Akhtar, et al., Corneal nerve loss in patients with TIA and acute ischemic stroke in relation to circulating markers of inflammation and vascular integrity, Sci. Rep. 12 (2022) 3332, https://doi.org/10.1038/s41598-022-07353-7, 20220228.
- [87] G.A. Rosenberg, Metalloproteinases and neurodegenerative diseases: pathophysiological and therapeutic perspectives, Metalloproteinases. Med. 5 (2015) 39–50.
- [88] J.D. Sanchez, R.A. Martirosian, K.T. Mun, et al., Temporal patterning of neurofilament light as a blood-based biomarker for stroke: a systematic review

and meta-analysis, Front. Neurol. 13 (2022) 841898, https://doi.org/10.3389/fneur.2022.841898, 20220516.

- [89] A. Petzold, G. Keir, J. Warren, et al., A systematic review and meta-analysis of CSF neurofilament protein levels as biomarkers in dementia, Neurodegener. Dis. 4 (2007) 185–194, https://doi.org/10.1159/000101843.
- [90] A. Petzold, Neurofilament phosphoforms: surrogate markers for axonal injury, degeneration and loss, J. Neurol. Sci. 233 (2005) 183–198, https://doi.org/ 10.1016/j.jns.2005.03.015.
- [91] M. Polyakova, K. Mueller, K. Arelin, et al., Increased serum NSE and S100B indicate neuronal and glial alterations in subjects under 71 years with mild neurocognitive disorder/mild cognitive impairment, Front. Cell Neurosci. 16 (2022) 788150, https://doi.org/10.3389/fncel.2022.788150, 20220714.
- [92] E.C. Kugler, J. Greenwood, R.B. MacDonald, The "Neuro-Glial-Vascular" unit: the role of glia in neurovascular unit formation and dysfunction, Front. Cell Dev. Biol. 9 (2021), https://doi.org/10.3389/fcell.2021.732820. Review.
- [93] G.C. O'Connell, C.G. Smothers, S.A. Gandhi, Newly-identified blood biomarkers of neurological damage are correlated with infarct volume in patients with acute ischemic stroke, J. Clin. Neurosci. 94 (2021) 107–113, https://doi.org/10.1016/j. jocn.2021.10.015, 20211019.
- [94] J.H. Kim, P.W. Ko, H.W. Lee, et al., Astrocyte-derived lipocalin-2 mediates hippocampal damage and cognitive deficits in experimental models of vascular dementia, Glia 65 (2017) 1471–1490, https://doi.org/10.1002/glia.23174, 20170605.
- [95] P. Hermann, A. Villar-Piqué, M. Schmitz, et al., Plasma lipocalin 2 in Alzheimer's disease: potential utility in the differential diagnosis and relationship with other biomarkers, Alzheimers. Res. Ther. 14 (2022) 9, https://doi.org/10.1186/ s13195-021-00955-9, 20220113.
- [96] Y.N. Li, R. Pan, X.J. Qin, et al., Ischemic neurons activate astrocytes to disrupt endothelial barrier via increasing VEGF expression, J. Neurochem. 129 (2014) 120–129, https://doi.org/10.1111/jnc.12611, 20131206.
- [97] Z. Alirezaei, M.H. Pourhanifeh, S. Borran, et al., Neurofilament light chain as a biomarker, and correlation with magnetic resonance imaging in diagnosis of CNSrelated disorders, Mol. Neurobiol. 57 (2020) 469–491, https://doi.org/10.1007/ s12035-019-01698-3, 20190805.
- [98] S. Narayanan, A. Shanker, T. Khera, et al., Neurofilament light: a narrative review on biomarker utility, Fac. Rev. 10 (2021) 46, https://doi.org/10.12703/r/10-46, 20210507.
- [99] S.-H. Kim, M.K. Choi, N.Y. Park, et al., Serum neurofilament light chain levels as a biomarker of neuroaxonal injury and severity of oxaliplatin-induced peripheral neuropathy, Sci. Rep. 10 (2020) 7995, https://doi.org/10.1038/s41598-020-64511-5.
- [100] J. Jessen Krut, T. Mellberg, R.W. Price, et al., Biomarker evidence of axonal injury in neuroasymptomatic HIV-1 patients, PLoS. One 9 (2014) e88591, https://doi. org/10.1371/journal.pone.0088591, 20140211.
- [101] Y.Y. Wong, C.Y. Wu, D. Yu, et al., Biofluid markers of blood-brain barrier disruption and neurodegeneration in Lewy body spectrum diseases: a systematic review and meta-analysis, Parkinsonism. Relat. Disord. 101 (2022) 119–128, https://doi.org/10.1016/j.parkreldis.2022.06.004, 20220620.
- [102] C.H. Wai, J. Jin, M. Cyrklaff, et al., Neurofilament light chain plasma levels are associated with area of brain damage in experimental cerebral malaria, Sci. Rep. 12 (2022) 10726, https://doi.org/10.1038/s41598-022-14291-x.
- [103] T. Uher, M. McComb, S. Galkin, et al., Neurofilament levels are associated with blood-brain barrier integrity, lymphocyte extravasation, and risk factors following the first demyelinating event in multiple sclerosis, Mult. Scler. 27 (2021) 220–231, https://doi.org/10.1177/1352458520912379, 20200407.
- [104] M. Kalm, M. Boström, Å. Sandelius, et al., Serum concentrations of the axonal injury marker neurofilament light protein are not influenced by blood-brain barrier permeability, Brain Res. 1668 (2017) 12–19, https://doi.org/10.1016/j. brainres.2017.05.011.
- [105] G. Gravesteijn, J.W. Rutten, I.M.W. Verberk, et al., Serum neurofilament light correlates with CADASIL disease severity and survival, Ann. Clin. Transl. Neurol. 6 (2019) 46–56, https://doi.org/10.1002/acn3.678, 20181120.
- [106] A. Huss, A. Abdelhak, B. Mayer, et al., Association of serum GFAP with functional and neurocognitive outcome in sporadic small vessel disease, Biomedicines. 10 (2022) 20220802, https://doi.org/10.3390/biomedicines10081869.
- [107] M. Duering, M.J. Konieczny, S. Tiedt, et al., Serum neurofilament light chain levels are related to small vessel disease burden, J. Stroke 20 (2018) 228–238, https://doi.org/10.5853/jos.2017.02565, 20180531.
- [108] Q. Gao, Y. Fan, L-Y Mu, et al., S100B and ADMA in cerebral small vessel disease and cognitive dysfunction, J. Neurol. Sci. 354 (2015) 27–32, https://doi.org/ 10.1016/j.jns.2015.04.031.
- [109] G.A. Rosenberg, J. Prestopnik, J.C. Adair, et al., Validation of biomarkers in subcortical ischaemic vascular disease of the Binswanger type: approach to targeted treatment trials, J. Neurol. Neurosurg. Psychiatr. 86 (2015) 1324–1330, https://doi.org/10.1136/jnnp-2014-309421, 20150124.
- [110] W.O. Tobin, J.A. Kinsella, G.F. Kavanagh, et al., Profile of von Willebrand factor antigen and von Willebrand factor propeptide in an overall TIA and ischaemic stroke population and amongst subtypes, J. Neurol. Sci. 375 (2017) 404–410, https://doi.org/10.1016/j.jns.2017.02.045, 20170224.
- [111] G.Y. Lip, A.D. Blann, I.S. Farooqi, et al., Sequential alterations in haemorheology, endothelial dysfunction, platelet activation and thrombogenesis in relation to prognosis following acute stroke: the West Birmingham Stroke Project, Blood Coagul, Fibrinolysis 13 (2002) 339–347, https://doi.org/10.1097/00001721-200206000-00010.

- [112] A. Bustamante, E. López-Cancio, S. Pich, et al., Blood biomarkers for the early diagnosis of stroke: the stroke-chip study, Stroke 48 (2017) 2419–2425, https:// doi.org/10.1161/strokeaha.117.017076, 20170717.
- [113] F. Arba, A. Giannini, B. Piccardi, et al., Small vessel disease and biomarkers of endothelial dysfunction after ischaemic stroke, Eur. Stroke J. 4 (2019) 119–126, https://doi.org/10.1177/2396987318805905, 20181022.
- [114] H.C. Segal, A.I. Burgess, D.L. Poole, et al., Population-based study of blood biomarkers in prediction of subacute recurrent stroke, Stroke 45 (2014) 2912–2917, https://doi.org/10.1161/STROKEAHA.114.005592.
- [115] D. Faille, J. Labreuche, E. Meseguer, et al., Endothelial markers are associated with thrombolysis resistance in acute stroke patients, Eur. J. Neurol. 21 (2014) 643–647, https://doi.org/10.1111/ene.12369, 20140204.
- [116] A.L.C.F. Lehmann, D.F. Alfieri, M.C.M. de Araújo, et al., Immune-inflammatory, coagulation, adhesion, and imaging biomarkers combined in machine learning models improve the prediction of death 1 year after ischemic stroke, Clin. Exp. Med. 22 (2022) 111–123, https://doi.org/10.1007/s10238-021-00732-w.
- [117] X. Wang, F.M. Chappell, M. Valdes Hernandez, et al., Endothelial function, inflammation, thrombosis, and basal ganglia perivascular spaces in patients with stroke, J. Stroke Cerebrovasc. Dis. 25 (2016) 2925–2931, https://doi.org/ 10.1016/j.jstrokecerebrovasdis.2016.08.007, 20160827.
- [118] E. Cuadrado-Godia, A. Ois, E. Garcia-Ramallo, et al., Biomarkers to predict clinical progression in small vessel disease strokes: prognostic role of albuminuria and oxidized LDL cholesterol, Atherosclerosis 219 (2011) 368–372, https://doi. org/10.1016/j.atherosclerosis.2011.07.114, 20110804.
- [119] J. Zaremba, J. Losy, sPECAM-1 in serum and CSF of acute ischaemic stroke patients, Acta Neurol. Scand. 106 (2002) 292–298, https://doi.org/10.1034/ j.1600-0404.2002.01339.x.
- [120] M.A. Gregory, L. Manuel-Apolinar, S. Sánchez-Garcia, et al., Soluble Intercellular adhesion molecule-1 (sICAM-1) as a biomarker of vascular cognitive impairment in older adults, Dement. Geriatr. Cogn. Disord. 47 (2019) 243–253, https://doi. org/10.1159/000500068, 20190813.
- [121] H.S. Markus, B. Hunt, K. Palmer, et al., Markers of endothelial and hemostatic activation and progression of cerebral white matter hyperintensities: longitudinal results of the Austrian Stroke Prevention Study, Stroke 36 (2005) 1410–1414, https://doi.org/10.1161/01.STR.0000169924.60783.d4, 20050519.
- [122] L.M. Varela, E. Meseguer, B. Lapergue, et al., Changes in high-density lipoproteins related to outcomes in patients with acute stroke, J. Clin. Med. 9 (2020) 20200717, https://doi.org/10.3390/jcm9072269.
- [123] E. Meseguer, D. Diallo, J. Labreuche, et al., Osteopontin predicts three-month outcome in stroke patients treated by reperfusion therapies, J. Clin. Med. 9 (2020) 20201213, https://doi.org/10.3390/jcm9124028.
- [124] T. Bogoslovsky, M. Spatz, A. Chaudhry, et al., Circulating CD133+CD34+ progenitor cells inversely correlate with soluble ICAM-1 in early ischemic stroke patients, J. Transl. Med. 9 (2011) 145, https://doi.org/10.1186/1479-5876-9-145, 20110826.
- [125] M. van Dinther, M.T. Schram, J.F.A. Jansen, et al., Extracerebral microvascular dysfunction is related to brain MRI markers of cerebral small vessel disease: the Maastricht Study, Geroscience 44 (2022) 147–157, https://doi.org/10.1007/ s11357-021-00493-0, 20211123.
- [126] J.R. Batuca, M.C. Amaral, C. Favas, et al., Antibodies against HDL Components in Ischaemic stroke and coronary artery disease, Thromb. Haemost. 118 (2018) 1088–1100, https://doi.org/10.1055/s-0038-1645857, 20180503.
- [127] Y. Cui, X.H. Wang, Y. Zhao, et al., Change of serum biomarkers to postthrombolytic symptomatic intracranial hemorrhage in stroke, Front. Neurol. 13 (2022) 889746, https://doi.org/10.3389/fneur.2022.889746, 20220602.
- [128] F.E. de Leeuw, M. de Kleine, C.J. Frijns, et al., Endothelial cell activation is associated with cerebral white matter lesions in patients with cerebrovascular disease, Ann. N. Y. Acad. Sci. 977 (2002) 306–314, https://doi.org/10.1111/ j.1749-6632.2002.tb04831.x.
- [129] S. Richard, L. Lagerstedt, P.R. Burkhard, et al., E-selectin and vascular cell adhesion molecule-1 as biomarkers of 3-month outcome in cerebrovascular diseases, J. Inflamm. (Lond) 12 (2015) 61, https://doi.org/10.1186/s12950-015-0106-z, 20151104.
- [130] A. Lasek-Bal, H. Jedrzejowska-Szypulka, S. Student, et al., The importance of selected markers of inflammation and blood-brain barrier damage for short-term ischemic stroke prognosis, J. Physiol. Pharmacol. 70 (2019) 20190722, https:// doi.org/10.26402/jpp.2019.2.04.
- [131] F. Yu, J. Lu, Z. Li, et al., Correlation of plasma vascular endothelial growth factor and endostatin levels with symptomatic intra- and extracranial atherosclerotic stenosis in a Chinese Han population, J. Stroke Cerebrovasc. Dis. 26 (2017) 1061–1070, https://doi.org/10.1016/j.jstrokecerebrovasdis.2016.12.021, 20170208.
- [132] H.S. Kwon, Y.S. Kim, H.H. Park, et al., Increased VEGF and decreased SDF-1α in patients with silent brain infarction are associated with better prognosis after first-ever acute lacunar stroke, J. Stroke Cerebrovasc. Dis. 24 (2015) 704–710, https://doi.org/10.1016/j.jstrokecerebrovasdis.2014.11.021, 20150116.
- [133] S.C. Lee, K.Y. Lee, Y.J. Kim, et al., Serum VEGF levels in acute ischaemic strokes are correlated with long-term prognosis, Eur. J. Neurol. 17 (2010) 45–51, https:// doi.org/10.1111/j.1468-1331.2009.02731.x, 20090629.
- [134] L. Simani, M. Ramezani, E. Mohammadi, et al., Association of changed serum brain biomarkers with perihematomal edema and early clinical outcome in primary ICH patients, Neurologist. 27 (2022) 168–172, https://doi.org/10.1097/ nrl.00000000000000000, 20220701.
- [135] N. Li, Y.F. Liu, L. Ma, et al., Association of molecular markers with perihematomal edema and clinical outcome in intracerebral hemorrhage, Stroke 44 (2013) 658–663, https://doi.org/10.1161/strokeaha.112.673590, 20130206.

- [136] E. López-Cancio, A.C. Ricciardi, T. Sobrino, et al., Reported prestroke physical activity is associated with vascular endothelial growth factor expression and good outcomes after stroke, J. Stroke Cerebrovasc. Dis. 26 (2017) 425–430, https:// doi.org/10.1016/j.jstrokecerebrovasdis.2016.10.004, 20161028.
- [137] P. Dassan, G. Keir, H.R. Jäger, et al., Value of measuring serum vascular endothelial growth factor levels in diagnosing acute ischemic stroke, Int. J. Stroke 7 (2012) 454–459, https://doi.org/10.1111/j.1747-4949.2011.00677.x, 20111122.
- [138] R. Kazmierski, S. Michalak, A. Wencel-Warot, et al., Serum tight-junction proteins predict hemorrhagic transformation in ischemic stroke patients, Neurology. 79 (2012) 1677–1685, https://doi.org/10.1212/WNL.0b013e31826e9a83, 20120919.
- [139] X. He, D.R. Li, C. Cui, et al., Clinical significance of serum MCP-1 and VE-cadherin levels in patients with acute cerebral infarction, Eur. Rev. Med. Pharmacol. Sci. 21 (2017) 804–808.
- [140] X. Jiao, P. He, Y. Li, et al., The role of circulating tight junction proteins in evaluating blood brain barrier disruption following intracranial hemorrhage, Dis. Markers 2015 (2015) 860120, https://doi.org/10.1155/2015/860120.
- [141] S. Shi, Z. Qi, Q. Ma, et al., Normobaric hyperoxia reduces blood occludin fragments in rats and patients with acute ischemic stroke, Stroke 48 (2017) 2848–2854, https://doi.org/10.1161/strokeaha.117.017713, 20170920.
- [142] J.Y. Li, Z.Y. Cai, X.H. Sun, et al., Blood-brain barrier dysfunction and myelin basic protein in survival of amyotrophic lateral sclerosis with or without frontotemporal dementia, Neurol. Sci. 43 (2022) 3201–3210, https://doi.org/ 10.1007/s10072-021-05731-z, 20211126.
- [143] W. Li, Z. Qi, H. Kang, et al., Serum occludin as a biomarker to predict the severity of acute ischemic stroke, hemorrhagic transformation, and patient prognosis, Aging Dis. 11 (2020) 1395–1406, https://doi.org/10.14336/ad.2020.0119, 20201201.
- [144] Y. Shen, H.M. Gao, Serum somatostatin and neuron-specific enolase might be biochemical markers of vascular dementia in the early stage, Int. J. Clin. Exp. Med. 8 (2015) 19471–19475, 20151015.
- [145] S. González-García, A. González-Quevedo, M. Peña-Sánchez, et al., Serum neuronspecific enolase and S100 calcium binding protein B biomarker levels do not improve diagnosis of acute stroke, J. R. Coll. Physicians Edinb. 42 (2012) 199–204, https://doi.org/10.4997/jrcpe.2012.302.
- [146] S. Zaheer, M. Beg, I. Rizvi, et al., Correlation between serum neuron specific enolase and functional neurological outcome in patients of acute ischemic stroke, Ann. Indian Acad. Neurol. 16 (2013) 504–508, https://doi.org/10.4103/0972-2327.120442.
- [147] J. Onatsu, R. Vanninen, P. JÄkÄlÄ, et al., Tau, S100B and NSE as blood biomarkers in acute cerebrovascular events, In. Vivo (Brooklyn) 34 (2020) 2577–2586, https://doi.org/10.21873/invivo.12075.
- [148] B.J. Kim, Y.J. Kim, S.H. Ahn, et al., The second elevation of neuron-specific enolase peak after ischemic stroke is associated with hemorrhagic transformation, J. Stroke Cerebrovasc. Dis. 23 (2014) 2437–2443, https://doi.org/10.1016/j. jstrokecerebrovasdis.2014.05.020, 20140831.
- [149] J. Wu, D. Wu, Y. Liang, et al., Plasma neurofilament light chain: a biomarker predicting severity in patients with acute ischemic stroke, Medicine (Baltimore) 101 (2022) e29692, https://doi.org/10.1097/md.00000000029692, 20220701.
- [150] T.F. Gendron, M.K. Badi, M.G. Heckman, et al., Plasma neurofilament light predicts mortality in patients with stroke, Sci. Transl. Med. 12 (2020), https:// doi.org/10.1126/scitranslmed.aay1913.
- [151] Y. Qu, C.-C. Tan, X.-N. Shen, et al., Association of plasma neurofilament light with small vessel disease burden in nondemented elderly, Stroke 52 (2021) 896–904, https://doi.org/10.1161/STROKEAHA.120.030302.
- [152] Y. Peng, Q. Li, L. Qin, et al., Combination of serum neurofilament light chain levels and MRI markers to predict cognitive function in ischemic stroke, Neurorehabil. Neural Repair. 35 (2021) 247–255, https://doi.org/10.1177/ 1545968321989354, 20210201.
- [153] F. Pujol-Calderón, E. Portelius, H. Zetterberg, et al., Neurofilament changes in serum and cerebrospinal fluid after acute ischemic stroke, Neurosci. Lett. 698 (2019) 58–63, https://doi.org/10.1016/j.neulet.2018.12.042, 20181229.
- [154] T. Uphaus, S. Bittner, S. Gröschel, et al., NFL (neurofilament light chain) levels as a predictive marker for long-term outcome after ischemic stroke, Stroke 50 (2019) 3077–3084, https://doi.org/10.1161/STROKEAHA.119.026410.
- [155] N. Peters, E. van Leijsen, A.M. Tuladhar, et al., Serum neurofilament light chain is associated with incident lacunes in progressive cerebral small vessel disease, J. Stroke 22 (2020) 369–376, https://doi.org/10.5853/jos.2019.02845, 20200929.
- [156] J. Sellner, A. Patel, P. Dassan, et al., Hyperacute detection of neurofilament heavy chain in serum following stroke: a transient sign, Neurochem. Res. 36 (2011) 2287–2291, https://doi.org/10.1007/s11064-011-0553-8, 20110727.
- [157] P. Singh, J. Yan, R. Hull, et al., Levels of phosphorylated axonal neurofilament subunit H (PNFH) are increased in acute ischemic stroke, J. Neurol. Sci. 304 (2011) 117–121, https://doi.org/10.1016/j.jns.2011.01.025, 20110223.
- [158] A. De Vos, D. Jacobs, H. Struyfs, et al., C-terminal neurogranin is increased in cerebrospinal fluid but unchanged in plasma in Alzheimer's disease, Alzheimers Dement. 11 (2015) 1461–1469, https://doi.org/10.1016/j.jalz.2015.05.012, 20150616.
- [159] K. Shao, S. Shan, W. Ru, et al., Association between serum NPTX2 and cognitive function in patients with vascular dementia, Brain Behav. 10 (2020) e01779, https://doi.org/10.1002/brb3.1779, 20200803.

#### G.K. Hansra et al.

- [160] D.W. Choi, T.S. Kim, Y.S. Kim, et al., Elevated plasma biomarkers of inflammation in acute ischemic stroke patients with underlying dementia, BMC. Neurol. 20 (2020) 293, https://doi.org/10.1186/s12883-020-01859-1, 20200805.
  [161] C. Foerch, B. Otto, O.C. Singer, et al., Serum S100B predicts a malignant course of
- [161] C. Foerch, B. Otto, O.C. Singer, et al., Serum S100B predicts a malignant course of infarction in patients with acute middle cerebral artery occlusion, Stroke 35 (2004) 2160–2164, https://doi.org/10.1161/01.STR.0000138730.03264.ac.
- [162] Q. Wang, K. Wang, Y. Ma, et al., Serum galectin-3 as a potential predictive biomarker is associated with poststroke cognitive impairment, Oxid. Med. Cell Longev. 2021 (2021) 5827812, https://doi.org/10.1155/2021/5827812.
- [163] G.K. Davis, N.S. Baboolal, D. Seales, et al., Potential biomarkers for dementia in Trinidad and Tobago, Neurosci. Lett. 424 (2007) 27–30, https://doi.org/ 10.1016/j.neulet.2007.07.011.
- [164] T.L. Barr, L.L. Latour, K.Y. Lee, et al., Blood-brain barrier disruption in humans is independently associated with increased matrix metalloproteinase-9, Stroke 41 (2010) e123–e128, https://doi.org/10.1161/strokeaha.109.570515, 20091224.
- [165] J. Serena, M. Blanco, M. Castellanos, et al., The prediction of malignant cerebral infarction by molecular brain barrier disruption markers, Stroke 36 (2005) 1921–1926, https://doi.org/10.1161/01.Str.0000177870.14967.94, 20050811.