



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

REFERENCES

- Arthur, J.C., Gharaibeh, R.Z., Uronis, J.M., Perez-Chanona, E., Sha, W., Tomkovich, S., Mühlbauer, M., Fodor, A.A., and Jobin, C. (2013). VSL#3 probiotic modifies mucosal microbial composition but does not reduce colitis-associated colorectal cancer. *Sci. Rep.* 3, 2868.
- Cait, A., Mooney, A., Poyntz, H., Shortt, N., Jones, A., Gestin, A., Gell, K., Grooby, A., O'Sullivan, D., Tang, J.S., et al. (2021). Potential Association Between Dietary Fibre and Humoral Response to the Seasonal Influenza Vaccine. *Front. Immunol.* 12, 765528.
- McQuade, J.L., Daniel, C.R., Hess, K.R., Mak, C., Wang, D.Y., Rai, R.R., Park, J.J., Haydu, L.E., Spencer, C., Wongchenko, M., et al. (2018). Association of body-mass index and outcomes in patients with metastatic melanoma treated with targeted therapy, immunotherapy, or chemotherapy: a retrospective, multicohort analysis. *Lancet Oncol.* 19, 310–322.
- Meza, L.A., Dizman, N., Bergerot, P.G., Dorff, T.B., Lyou, Y., Frankel, P.H., Mira, V., Llamas, M., Hsu, J., Busra Zengin, Z., et al. (2021). First results of a randomized phase IB study comparing nivolumab/ipilimumab with or without CBM-588 in patients with metastatic renal cell carcinoma. *J Clin Oncol.* 39, 4513.
- Nomura, M., Nagatomo, R., Doi, K., Shimizu, J., Baba, K., Saito, T., Matsumoto, S., Inoue, K., and Muto, M. (2020). Association of Short-Chain Fatty Acids in the Gut Microbiome With Clinical Response to Treatment With Nivolumab or Pembrolizumab in Patients With Solid Cancer Tumors. *JAMA Netw. Open* 3, e202895.
- Spencer, C.N., McQuade, J.L., Gopalakrishnan, V., McCulloch, J.A., Vetzou, M., Cogdill, A.P., Khan, M.A.W., Zhang, X., White, M.G., Peterson, C.B., et al. (2021). Dietary fiber and probiotics influence the gut microbiome and melanoma immunotherapy response. *Science* 374, 1632–1640.
- Tangestani, H., Emamat, H., Ghalandari, H., and Shab-Bidar, S. (2020). Whole Grains, Dietary Fibers and the Human Gut Microbiota: A Systematic Review of Existing Literature. *Recent Pat. Food Nutr. Agric.* 11, 235–248.
- Tomita, Y., Ikeda, T., Sakata, S., Saruwatari, K., Sato, R., Iyama, S., Jodai, T., Akaike, K., Ishizuka, S., Saeki, S., and Sakagami, T. (2020). Association of Probiotic *Clostridium butyricum* Therapy with Survival and Response to Immune Checkpoint Blockade in Patients with Lung Cancer. *Cancer Immunol. Res.* 8, 1236–1242.
- Wagenaar, C.A., van de Put, M., Bisschops, M., Walrabenstein, W., de Jonge, C.S., Herrema, H., and van Schaardenburg, D. (2021). The Effect of Dietary Interventions on Chronic Inflammatory Diseases in Relation to the Microbiome: A Systematic Review. *Nutrients* 13, 3208.
- Yonekura, S., Terrisse, S., Costa Silva, C.A., Lafarge, A., Iebba, V., Ferrere, G., Goubet, A.-G., Fahrner, J.-E., Lahmar, I., Ueda, K., et al. (2021). Cancer induces a stress ileopathy depending on B-adrenergic receptors and promoting dysbiosis that contribute to carcinogenesis. *Cancer Discov.* Published online December 20, 2021. <https://doi.org/10.1158/2159-8290.CD-21-0999>.

SARS-CoV-2 pirates the kidneys: A scar(y) story

Jochen Reiser,^{1,*} Ryan Spear,¹ and Shengyuan Luo¹

¹Department of Internal Medicine, Rush University, Chicago, IL 60612, USA

*Correspondence: jochen_reiser@rush.edu
<https://doi.org/10.1016/j.cmet.2022.02.005>

SARS-CoV-2 can cause diverse severe and lasting damage to the kidneys. In the latest issue of *Cell Stem Cell*, Jansen et al. utilized data gleaned from human kidney autopsies and human induced pluripotent stem cell-derived kidney organoids to investigate the direct effects of SARS-CoV-2 infection on kidney cells. They found that such infections resulted in renal scarring (notably, tubulointerstitial fibrosis).

As the coronavirus disease 2019 (COVID-19) global pandemic continues for a third year, more has been unveiled about its culprit, the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2). Aside from triggering a possibly dire innate immune response, it also causes severe respiratory illness and a myriad of acute and chronic multi-organ damage, with the kidneys being one of the most frequently targeted organs outside the lungs. Accumulating evidence suggests the presence of SARS-CoV-2 in some but not all kidneys from COVID-19 cases, yet at higher concentrations compared to other organs (Puelles et al., 2020). Renal manifestations of COVID-19 can range from acute kidney injury (AKI), hematuria, proteinuria, podocytopathy, and tubulointerstitial fibrosis (Diao et al., 2021). Importantly, a substantial number of individuals

with COVID-19 experience post-acute sequelae, a syndrome now referred to as “long COVID,” many of whom suffered worse outcomes of AKI such as rapid kidney function decline and chronic kidney disease (CKD) (Bowe et al., 2021). One key to understanding these phenomena is to determine if the virus infects kidney cells directly and, if so, if this leads to short- and/or long-term renal damage.

In the latest issue of *Cell Stem Cell*, Jansen et al. comprehensively investigated whether direct infection of the kidneys by SARS-CoV-2 introduces a pro-fibrotic phenotype (Jansen et al., 2022). The authors compared kidneys of individuals with COVID-19 compared to matched controls. With data derived from 61 human autopsies and single-RNA nucleus sequencing, the study associated increased interstitial fibrosis with the

presence of higher activity in proinflammatory and fibrosis-driving pathways (notably, tumor necrosis factor alpha [TNF α], transforming growth factor beta, nuclear factor κ B, and JAK-STAT). Taking this to the next level, the study utilized human induced pluripotent stem cell (hiPSC)-derived kidney organoids to fine map cellular locations of SARS-CoV-2 infection and accompanying signaling changes. In addition to verifying upregulated activity of pro-fibrotic pathways found in SARS-CoV-2-infected autopsied kidneys, the team identified evidence of a molecular crosstalk between proximal tubular cells and PDGFR α /b⁺ mesenchymal cells (a key source of myofibroblasts) that further contributed to fibrosis.

The findings by Jansen and colleagues are exciting and supported by both human tissues and hiPSC-derived



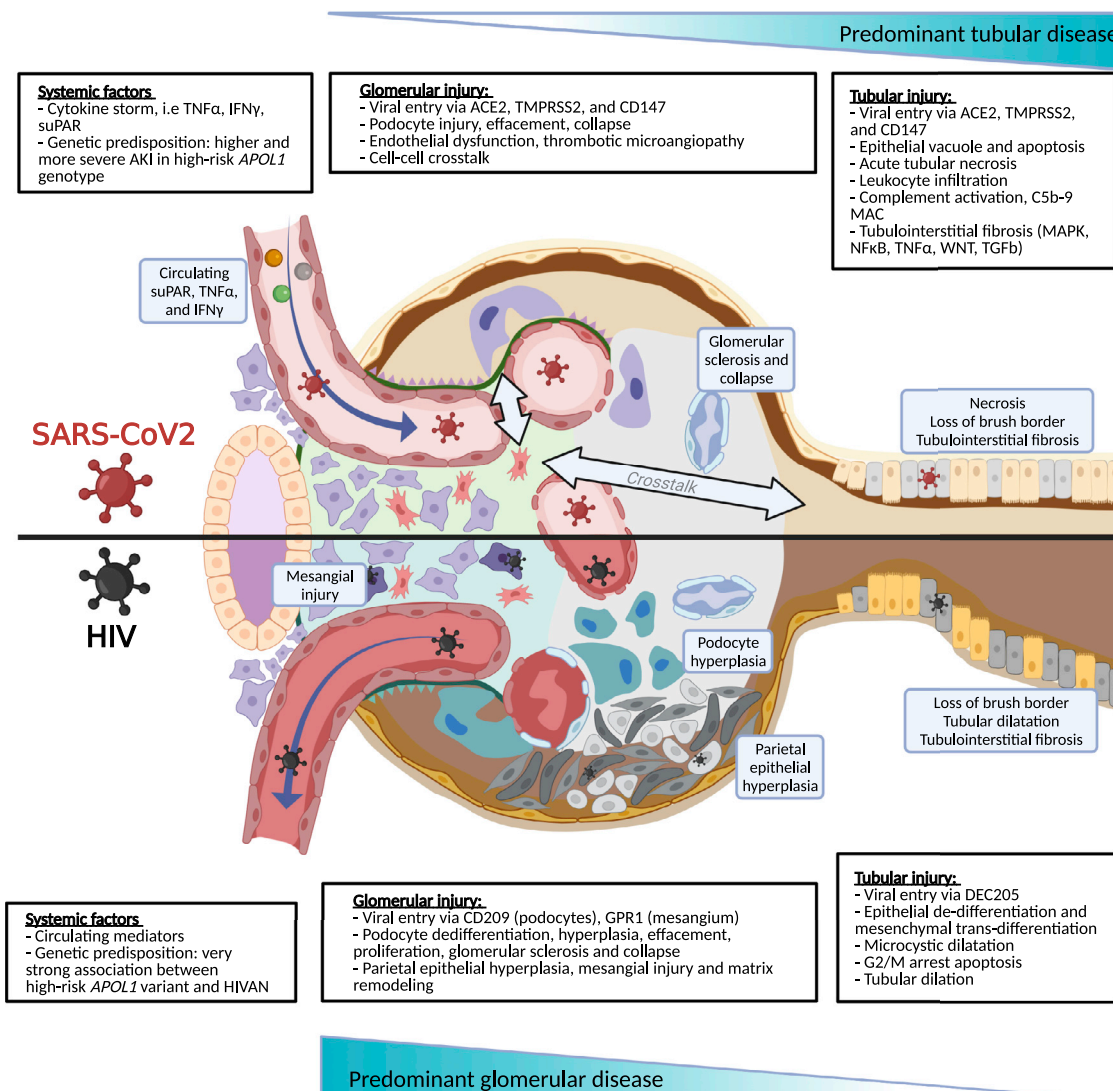


Figure 1. Schematic overview of the kidney infected by SARS-CoV-2 or HIV

Mechanisms and details of SARS-CoV-2 and human immunodeficiency virus (HIV) and their impact on the kidney. SARS-CoV-2, more than HIV, triggers an innate immune response that imposes systemic effects on the kidney, impacting glomerular injury and tubular injury. Both viruses can also infect kidney cells directly, which in the case of SARS-CoV-2 may contribute to fibrosis. Notably, systemic and cell-specific effects are compounded, which can increase risk further upon additional factors (e.g., suPAR, *APOL1* risk variants).

organoids. They can serve as strong evidence to support the notion that SARS-CoV-2 can directly drive renal fibrosis, a CKD hallmark, suggesting a likely impact on the worse AKI outcomes during COVID-19 that are often observed. These findings also bridged a knowledge gap, linking acute viral illness to kidney disease during long-COVID. hiPSC-derived organoids have the advantage of being able to assess intercellular interactions, which possibly explains the greater extent of cytopathic changes seen in organoids compared to other studies using SARS-CoV-2-infected *ex vivo* models comprised

solely of tubular epithelial cells and that lacked cytopathic changes (Omer et al., 2021). Jansen et al. provided the first hints that SARS-CoV-2 infection in kidney organoids is amenable to a treatment with a protease inhibitor, providing a glimpse into potential future treatments for COVID-19-related kidney injury. The findings also stimulate further clinical and translational investigations for testing SARS-CoV-2 tropism in the kidneys of infected individuals to identify potential virus-free kidneys for transplantation procured from individuals who died of fulminant COVID-19 (Lee et al., 2022).

The use of hiPSC-kidney organoids is not without caveats, however. As the authors pointed out, the protocols to differentiate hiPSCs to model the adult kidney in its entirety are still suboptimal. While excluding extra-renal factors, such as intensive care treatment, the model also isolates kidney cells from the involvement of immune cells. For example, the recruitment of CD68+ macrophages, CD56+ natural killer cells, and CD8+ T cells, alongside strong C5b-9 deposition, was found with tubular necrosis in the kidneys of patients with COVID-19 (Diao et al., 2021). Jansen et al. also observed an

interaction between podocytes and the mesenchymal endothelial progenitor population. The cytologic consequences were beyond the study's scope and therefore not described, but one must recognize podocytopathy, glomerular collapsing, and sclerosis as indispensable parts of the COVID-19-related kidney disease spectrum, in which innate immunity likely plays an essential role. The massive release of immune mediators such as soluble urokinase plasminogen activator receptor (suPAR), TNF α , and interferon gamma, i.e., a cytokine storm, signifies severe COVID-19 and confers substantial risk for AKI (Azam et al., 2020; Karki et al., 2021). Biomarkers predictive of outcomes that are elevated early during disease progression, such as suPAR, have been successfully applied in clinical practice to guide immunomodulative treatment and improve prognosis (Kyriazopoulou et al., 2021). Future studies aiming to examine viral triggers, innate immune mediators, and kidney cell reactivity in concert will need to be performed. In the meantime, Jansen et al. allow us to draw the conclusion that the renal presence of SARS-CoV-2 can bring about pro-inflammatory and pro-fibrotic molecular changes that may prime the kidneys for further damage (including hits from circulating mediators such as suPAR and TNF α), ultimately leading to lasting kidney disease in long COVID.

The study by Jansen et al. provides additional insights to allow for a comparison of kidneys infected by SARS-CoV-2 and other viral infections, especially human immunodeficiency virus (HIV) infection (Figure 1). Both SARS-CoV-2 and HIV directly infect podocytes and tubular epithelial cells. Collapsing glomerulopathy, tubulointerstitial fibrosis, and mesangial injury can be seen in both COVID-19 and HIV infection and are perhaps dependent on the amount of the innate immune activation that is mounted as an anti-infection response. Lessons

learned from our experience with HIV infection may offer insight into the study of COVID-19-related kidney disease. Is there a set of COVID-19 gene products that converges pro-fibrotic and other pathways in diverse kidney cells and structures? African Americans develop COVID-19 associated with stronger AKI more often than other racial and ethnic groups (Hung et al., 2022). Are socioeconomic factors the sole drivers of these disparities, or is there a role for other genetic factors and environmental factors? It is noteworthy that untreated HIV infection caused glomerulopathy in 50% of African Americans with two *APOL1* risk alleles. Data from patients with COVID-19 also suggested AKI, proteinuria, and podocytopathy were more prevalent among those with the high-risk *APOL1* genotype. Will there be a need for screening and early detection of CKD in long COVID? Which antivirals and what duration of treatment will ensure clearance of SARS-CoV-2 in the kidneys? A stage is now set for these topics to be addressed.

ACKNOWLEDGMENTS

The authors thank Vineet Gupta, Ph.D. for critical reading of the manuscript. This work was supported by the National Institute of Diabetes and Digestive and Kidney Diseases (R01DK125858, R01DK109720, and R01DK113761) to J.R.

DECLARATION OF INTERESTS

J.R. is cofounder, scientific advisory board co-chair, and shareholder of Walden Biosciences, a kidney therapeutic company. Other authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

REFERENCES

Azam, T.U., Shadid, H.R., Blakely, P., O'Hayer, P., Berlin, H., Pan, M., Zhao, P., Zhao, L., Pennathur, S., Pop-Busui, R., et al. (2020). Soluble urokinase receptor (SuPAR) in COVID-19-related AKI. *J. Am. Soc. Nephrol.* 31, 2725–2735.

Bowe, B., Xie, Y., Xu, E., and Al-Aly, Z. (2021). Kidney outcomes in long COVID. *J. Am. Soc. Nephrol.* 32, 2851–2862.

Diao, B., Wang, C., Wang, R., Feng, Z., Zhang, J., Yang, H., Tan, Y., Wang, H., Wang, C., Liu, L., et al. (2021). Human kidney is a target for novel severe acute respiratory syndrome coronavirus 2 infection. *Nat. Commun.* 12, 2506.

Hung, A.M., Shah, S.C., Bick, A.G., Yu, Z., Chen, H.C., Hunt, C.M., Wendt, F., Wilson, O., Greevy, R.A., Chung, C.P., et al.; VA Million Veteran Program COVID-19 Science Initiative (2022). *APOL1* risk variants, acute kidney injury, and death in participants with African ancestry hospitalized with COVID-19 from the Million Veteran Program. *JAMA Intern. Med.* Published online January 28, 2022. <https://doi.org/10.1001/jamainternmed.2021.8538>.

Jansen, J., Reimer, K.C., Nagai, J.S., Varghese, F.S., Overheul, G.J., de Beer, M., Rovers, R., Daviran, D., Fermin, L.A.S., Willemsen, B., et al.; COVID Moonshot consortium (2022). SARS-CoV-2 infects the human kidney and drives fibrosis in kidney organoids. *Cell Stem Cell* 29, 217–231.e8.

Karki, R., Sharma, B.R., Tuladhar, S., Williams, E.P., Zalduendo, L., Samir, P., Zheng, M., Sundaram, B., Banoth, B., Malireddi, R.K.S., et al. (2021). Synergism of TNF- α and IFN- γ triggers inflammatory cell death, tissue damage, and mortality in SARS-CoV-2 infection and cytokine shock syndromes. *Cell* 184, 149–168.e17.

Kyriazopoulou, E., Poulakou, G., Milionis, H., Metallidis, S., Adamis, G., Tsiakos, K., Fragkou, A., Rapti, A., Damoulari, C., Fantoni, M., et al. (2021). Early treatment of COVID-19 with anakinra guided by soluble urokinase plasminogen receptor plasma levels: a double-blind, randomized controlled phase 3 trial. *Nat. Med.* 27, 1752–1760.

Lee, K., Desai, N.M., Resnick, J., Li, M., Johanson, A., Pekosz, A., Rabb, H., and Mankowski, J.L. (2022). Successful kidney transplantation from a deceased donor with severe COVID-19 respiratory illness with undetectable SARS-CoV-2 in donor kidney and aorta. *Am. J. Transplant.* Published online January 13, 2022. <https://doi.org/10.1111/ajt.16956>.

Omer, D., Pleniceanu, O., Gnatek, Y., Namestnikov, M., Cohen-Zontag, O., Goldberg, S., Friedman, Y.E., Friedman, N., Mandelboim, M., Vitner, E.B., et al. (2021). Human kidney spheroids and monolayers provide insights into SARS-CoV-2 renal interactions. *J. Am. Soc. Nephrol.* 32, 2242–2254.

Puelles, V.G., Lütgehetmann, M., Lindenmeyer, M.T., Sperhake, J.P., Wong, M.N., Allweiss, L., Chilla, S., Heinemann, A., Wanner, N., Liu, S., et al. (2020). Multiorgan and renal tropism of SARS-CoV-2. *N. Engl. J. Med.* 383, 590–592.