

Citation: Seimetz M, Parajuli N, Pichl A, Bednorz M, Ghofrani HA, Schermuly RT, et al. (2015) Cigarette Smoke-Induced Emphysema and Pulmonary Hypertension Can Be Prevented by Phosphodiesterase 4 and 5 Inhibition in Mice. PLoS ONE 10(6): e0129327. doi:10.1371/journal. pone.0129327

Academic Editor: Tim Lahm, Indiana University, UNITED STATES

Received: October 18, 2014

Accepted: May 8, 2015

Published: June 9, 2015

Copyright: © 2015 Seimetz et al. This is an open access article distributed under the terms of the <u>Creative Commons Attribution License</u>, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by German Research Foundation (DFG), Excellence Cluster Cardio-Pulmonary System (ECCPS).

Competing Interests: The authors have declared that no competing interests exist.

RESEARCH ARTICLE

Cigarette Smoke-Induced Emphysema and Pulmonary Hypertension Can Be Prevented by Phosphodiesterase 4 and 5 Inhibition in Mice

Michael Seimetz[®], Nirmal Parajuli[®], Alexandra Pichl, Mariola Bednorz, Hossein Ardeschir Ghofrani, Ralph Theo Schermuly, Werner Seeger, Friedrich Grimminger, Norbert Weissmann*

Universities of Giessen and Marburg Lung Center (UGMLC), Excellence Cluster Cardio-Pulmonary System (ECCPS), Member of the German Center for Lung Research (DZL), Giessen, Germany

• These authors contributed equally to this work.

* Norbert.Weissmann@innere.med.uni-giessen.de

Abstract

Rationale

Chronic obstructive pulmonary disease (COPD) is a widespread disease, with no curative therapies available. Recent findings suggest a key role of NO and sGC-cGMP signaling for the pathogenesis of the disease. Previous data suggest a downregulation/inactivation of the cGMP producing soluble guanylate cyclase, and sGC stimulation prevented cigarette smoke-induced emphysema and pulmonary hypertension (PH) in mice. We thus aimed to investigate if the inhibition of the cGMP degrading phosphodiesterase (PDE)5 has similar effects. Results were compared to the effects of a PDE 4 inhibitor (cAMP elevating) and a combination of both.

Methods

C57BL6/J mice were chronically exposed to cigarette smoke and in parallel either treated with Tadalafil (PDE5 inhibitor), Piclamilast (PDE4 inhibitor) or both. Functional measurements (lung compliance, hemodynamics) and structural investigations (alveolar and vascular morphometry) as well as the heart ratio were determined after 6 months of tobacco smoke exposure. In addition, the number of alveolar macrophages in the respective lungs was counted.

Results

Preventive treatment with Tadalafil, Piclamilast or a combination of both almost completely prevented the development of emphysema, the increase in lung compliance, tidal volume, structural remodeling of the lung vasculature, right ventricular systolic pressure, and right ventricular hypertrophy induced by cigarette smoke exposure. Single, but not combination treatment prevented or reduced smoke-induced increase in alveolar macrophages.

Conclusion

Cigarette smoke-induced emphysema and PH could be prevented by inhibition of the phosphodiesterases 4 and 5 in mice.

Introduction

Chronic obstructive pulmonary disease (COPD) is a collective term for chronic bronchitis and emphysema and is one of the major causes of death worldwide [1]. On the one hand, airway inflammation and remodeling represent characteristic features. On the other hand, there is destruction of the elastic architecture of the lung which leads to enlargement of distal airspaces, causing emphysema [2]. In addition, COPD/emphysema is increasingly viewed as a systemic disease, involving skeletal muscle wasting, diaphragmatic dysfunction, and systemic inflammation [3]. Influx of inflammatory cells, imbalance of proteolytic and anti-proteolytic activity, increased oxidative stress with the rise in number of apoptotic cells and decreased proliferation might be important events underlying COPD [4-6].

Recent observations suggest an essential role of endothelial dysfunction and pulmonary hypertension (PH) for the development of COPD [7]. In animal models, cigarette smoke-induced emphysema was accompanied by pulmonary vascular remodeling and PH [8–11], and we and others could recently show that such vascular alterations even preceded alveolar destruction [8, 9]. The observations in animal models are supported by human studies showing that vascular remodeling can already occur in smokers without COPD [12, 13]. Moreover, the involvement of the pulmonary vasculature in COPD is reflected by the fact that up to 70% of COPD patients also suffer from PH [14].

Our own previous studies investigating nitric oxide (NO) as well as sGC-cGMP signaling revealed that 1) an inhibition of the NO-producing enzyme inducible NO synthase (iNOS) [9] and 2) a stimulation of the soluble guarylate cyclase (sGC) [15] prevented cigarette smoke-induced emphysema and PH. Furthermore, a curative approach using an iNOS inhibitor in diseased animals resulted in lung regeneration within three months [9]. Functionally essential sGC subunits have been shown to be downregulated upon smoke exposure in mice and in human COPD, suggesting a key role of cyclic guanosine monophosphate (cGMP) reduction as a driver of smoke-induced lung emphysema and PH [15]. Against this background the aim of the present study was to test the hypothesis that, similar to cGMP enrichment by sGC stimulation, inhibition (using Tadalafil) of the downstream cGMP degrading enzyme, phosphodiesterase (PDE)5, would prevent cigarette smoke-induced emphysema and PH in mice as well. For comparison we used a specific PDE4 inhibitor, Piclamilast [16], to assess the effects of a cAMP elevating agent. A combination of both inhibitors should reveal potential synergistic effects, possibly needed if single PDE inhibitor treatment would have no or only partially preventive effects. Despite the fact that PDE5 inhibitors have already been used in clinical trials in COPD patients with and without PH [17-25], this study provides the thus far missing data of long term treatment with PDE inhibitors and assesses detailed effects on alveolar and vascular structure which was not possible in the respective human studies.

Methods

All experiments were performed according to institutional guidelines complying with national and international regulations. The study was approved by the Regierungspräsidium Giessen (Hessen, Germany). All analyses/exposures were done in a blinded, randomized fashion. For the study design sample size estimation was performed prior to the experiments.

Exposure to smoke

Male C57BL6/J mice, body weight (19 to 20 g) were purchased from Charles River Deutschland, Sulzfeld, Germany. Mice were divided randomly into five groups (10 animals, each). Group 1: healthy control (no smoke exposure); group 2: control (Placebo [solvent application], smoke exposure), group 3: Piclamilast treatment (10 mg/kg body weight, smoke exposure); group 4: Tadalafil (10 mg/kg body weight, smoke exposure) and group 5: combination treatment with Piclamilast and Tadalafil (10 mg/kg body weight each, smoke exposure). The condition of the animals was monitored daily prior to treatment. Smoke challenged animals were whole body exposed [9] to tobacco smoke of 3R4F cigarettes (Kentucky Tobacco Research and Development Center, USA) at a particle concentration of 140 mg/m³ for 6 h/day, 5 days/week for a period of 6 months. 200 µl of solutions of Piclamilast and Tadalafil were freshly prepared and suspended in 4.0% methocel and 1.3% polyethyleneglycol via an Ultraturrax (IKA Staufen, Germany). To avoid foam formation, two drops of Antifoam C (Sigma-Aldrich) were added. The suspension was applied by gavage daily. After six months, functional parameters were assessed and lungs were fixed for morphometric analyses. Missing data are due to technical problems (e.g., catheter placement).

Lung function tests

Animals were anaesthetized with ketamine (60 mg/kg body weight) and xylazine (10 mg/kg body weight) intraperitoneally and anticoagulated with heparin (1000 U/kg). The trachea was cannulated, and the lungs were ventilated with room air at a tidal volume of 200 μ l and a frequency of 150 breaths per minute. Animals were maintained at physiological temperature throughout the experiment. A tracheal cannula was connected to a pneumotachometer (Hugo Sachs GmbH, Germany) and the tidal volume, resistance and dynamic compliance were evaluated using the HSE PULMODYN software (Hugo Sachs Electronics, March-Hugstetten, Germany) as described previously [9].

In vivo hemodynamics

Animals were anaesthetized with ketamine (60 mg/kg body weight) and xylazine (10 mg/kg body weight) intraperitoneally and anticoagulated with heparin (1000 U/kg). The trachea was cannulated, and the lungs were ventilated with room air at a tidal volume of 200 μ l and a frequency of 150 breaths per minute. Animals were maintained at physiological temperature throughout the experiment. Right ventricular systolic pressure (RVSP) was measured by inserting a PE-80 tube into the right ventricle via the right jugular vein and the systemic arterial pressure was determined by carotid artery catheterization as described previously [15, 26].

Fixation of the lung

Lungs were flushed blood-free with saline via the pulmonary artery. For morphometry, left lungs were fixed with 4.5% paraformaldehyde in phosphate-buffered saline (pH 7.0) via the trachea at a pressure of 12 cm H_2O and via the pulmonary artery at a vascular pressure of 22 cm H_2O . Investigations were performed from 3 μ m sections of paraffin embedded lungs.

Alveolar morphometry

The development of emphysema was determined from lung sections stained with hematoxylin and eosin as described previously [27] and measured by quantitative morphometry (parameters: mean linear intercept, air space, and septal wall thickness [9]).

Vascular morphometry

The degree of muscularization was determined from stained lung sections using antibodies against α -smooth muscle actin and von Willebrand factor as described before [9, 28]. Small (20–70 µm outer diameter), medium (70–150 µm outer diameter) and large (>150 µm outer diameter) vessels were classified as non-muscular (no smooth muscle cells detectable with actin staining), partially muscularized (at least one smooth muscle cell up to 75% of circumference with actin staining), and fully muscularized (>75% of circumference with actin staining). Vascular lumen area was determined from the same vessels analyzed for the degree of muscularization and is given as mean lumen area.

Right ventricular hypertrophy

Right ventricular hypertrophy was quantified by the ratio of the right ventricular (RV) and the left ventricular + septum (S) mass. Hearts were removed directly after lung fixation and the RV was dissected from the LV+S. Values were determined from dried heart tissue [15].

Quantification of the number of alveolar macrophages in lung sections

Macrophage counting was performed in 3- μ m sections from paraffin-embedded lungs. After heating at 58°C, lung sections were deparaffinized in xylene and rehydrated. The endogenous peroxidase activity was quenched with 15% (v/v) H₂O₂ in methanol. For staining, a 1:10 dilution of macrophage marker F4/80 antigen-specific antibody (rat monoclonal to F4/80 [ab6640], Abcam, Cambridge, UK) was used. Immune complexes were visualized with a peroxidase-conjugated secondary antibody and NovaRED Peroxidase (HRP) Substrate Kit (Vector labs, LINARIS, Wertheim-Bettingen, Germany) and Hematoxylin solution was used for counterstaining of the sections. The quantification of macrophages and measurement of the lung areas were carried out microscopically using a Qwin macro program from Leica (Wetzlar, Germany). For each lung, the total number of macrophages per lung section was counted. Data were calculated as amount of macrophages/mm².

Data analysis

All data are given as mean \pm SEM. Differences between more than two groups were assessed by ANOVA followed by Student-Newman-Keuls or Dunnett's post hoc test. P values <0.05 were considered as significant. All statistical calculations were performed using GraphPad Prism software.

Results

Effect of Piclamilast and/or Tadalafil on cigarette smoke-induced lung emphysema development

Tobacco smoke-exposure of mice for 6 months resulted in development of lung emphysema, quantified by an increase in airspace, mean linear intercept, and a decrease in septal wall thickness (Fig 1). These structural alterations were reflected by respective changes in lung compliance, tidal volume as well as airway resistance *in vivo* (Fig 2). Treatment with either Piclamilast,



Fig 1. Effect of Piclamilast and/or Tadalafil on cigarette smoke-induced lung emphysema development assessed by structural parameters. Mice were exposed to cigarette smoke for 6 months and treated in parallel with Piclamilast (10 mg/kg body weight/day) and/or Tadalafil (10 mg/kg body weight/ day). Time matched controls received solvent only. (A) Air space, (B) septal wall thickness and (C) mean linear intercept. (D) Representative histology from mice lung sections stained with hematoxylin and eosin. Data are given as mean \pm SEM from n = 5–6, each. Bars indicate significant differences using ANOVA followed by Student-Newman-Keuls post hoc test (*P<0.05;** P<0.01; ***P<0.001).

doi:10.1371/journal.pone.0129327.g001



Fig 2. Effect of Piclamilast and/or Tadalafil on cigarette smoke-induced lung emphysema development assessed by *in vivo* lung functional parameters. Mice were exposed to cigarette smoke for 6 months and treated in parallel with Piclamilast (10 mg/kg body weight/day) and/or Tadalafil (10 mg/kg body weight/day). Time matched controls received solvent only. (A) dynamic lung compliance, (B) tidal volume and (C) airway resistance. Data are given as mean \pm SEM from n = 6–9, each. Bars indicate significant differences using ANOVA followed by Student-Newman-Keuls post hoc test (* P<0.05**; P<0.01; ***P<0.001).

doi:10.1371/journal.pone.0129327.g002

Tadalafil or a combination of both in parallel with smoke exposure prevented the development of lung emphysema as evident from the structural as well as the functional parameters which were not different from untreated non-exposed control mice (Figs $\underline{1}$ and $\underline{2}$). No differences between Piclamilast, Tadalafil or the combination therapy could be detected.

Effect of Piclamilast and/or Tadalafil treatment on smoke-induced pulmonary hypertension, pulmonary vascular remodeling, and right heart hypertrophy

Tobacco smoke-exposed mice developed pulmonary hypertension as determined by increased right ventricular systolic pressure (Fig 3A). Moreover, smoke exposure resulted in vascular remodeling reflected by an increased degree of muscularization in all categories of vessel diameters assessed (Fig 4A-4C) and decreased the vascular lumen area in all vessels (Fig 4D-4F). Furthermore, the right ventricle was augmented upon chronic smoke exposure shown by the ratio of RV/LV+S (Fig 3B) compared to control mice. Treatment of mice with Piclamilast and/ or Tadalafil resulted in a complete protection against the development of pulmonary hypertension as displayed in Figs 3–5. Of note, the lumen of large vessels was significantly decreased after the combination therapy compared to both single treatments (Fig 4F).

Systemic effects of smoke exposure and inhibition of phosphodiesterases

To determine possible systemic consequences of right heart hypertrophy and PH seen in smoke-exposed mice, systemic arterial pressure (SAP) was measured via *A. carotis*. These data revealed a significant decrease of SAP in untreated smoke-exposed mice compared to control mice which was prevented by Tadalafil and the combination therapy, but not by the sole application of Piclamilast (Fig 6).



Fig 3. Effect of Piclamilast and/or Tadalafil on cigarette smoke-induced pulmonary hypertension in mice assessed by functional parameters. Mice were exposed to cigarette smoke for 6 months and treated in parallel with Piclamilast (10 mg/kg body weight/day) and/or Tadalafil (10 mg/kg body weight/day). Time matched controls received solvent only. (A) Right ventricular systolic pressure quantified by right heart catheterization in anesthetized animals. (B) Right heart hypertrophy, given as the ratio of right ventricular (RV) mass to left ventricular plus septum (LV+S) mass, from dried heart tissue; left ventricular weights did not differ between the different groups. Data are given as mean \pm SEM from n = 5–8, each. Bars indicate significant differences using ANOVA followed by Student-Newman-Keuls post hoc test (**P<0.01; ***P<0.001).

doi:10.1371/journal.pone.0129327.g003



Fig 4. Effect of Piclamilast and/or Tadalafil on cigarette smoke-induced pulmonary vascular remodeling in mice assessed by structural parameters. Mice were exposed to cigarette smoke for 6 months and treated in parallel with Piclamilast (10 mg/kg body weight/day) and/or Tadalafil (10 mg/kg body weight/day). Time matched controls received solvent only. (A-C) Degree of muscularization of (A) small pulmonary arteries (outer diameter 20–70 μ m), (B) medium vessels (outer diameter >70 to 150 μ m), and (C) large vessel (outer diameter >150 μ m), as a percentage of total vessel count for fully muscularized (full), partially muscularized (partial), and non-muscularized (non) vessels. (D-F) Mean vascular lumen area of (D) small pulmonary vessels (outer diameter 20–70 μ m), (E) medium vessels (outer diameter >70 to 150 μ m), and (F) large vessel (outer diameter >150 μ m). Data are given as mean ± SEM from n = 6, each. Bars indicate significant differences using ANOVA followed by Student-Newman-Keuls post hoc test (*P<0.05; **P<0.01; ***P<0.001).

doi:10.1371/journal.pone.0129327.g004

Anti-inflammatory effect of Piclamilast and/or Tadalafil

Based on already described anti-inflammatory action of PDE4 and -5 inhibitors, we counted the number of alveolar macrophages as an indicator of inflammatory response in lung sections derived from our smoke-exposed and control mice. Tobacco smoke-exposed mice showed a significant increase of alveolar macrophages per mm², which was significantly attenuated by Piclamilast. Tadalafil treatment showed a trend toward reduction, and interestingly, no such effect was evident for the combination therapy (Fig 7).

Discussion

The present study answers the question of whether an enrichment of the second messengers cGMP and cAMP by PDE inhibition could potentially be used for long-term treatment of cigarette smoke-induced emphysema and PH. Indeed, we demonstrated in a long-term mouse study that the PDE5 inhibitor Tadalafil and the PDE4 inhibitor Piclamilast as well as a combination of both prevented smoke-induced emphysema and pulmonary hypertension. Synergistic effects of the combination therapy could not be observed since single therapies have already had strong effects, almost to the level of unexposed control animals. The decline of systemic



Fig 5. Effect of Piclamilast and/or Tadalafil on cigarette smoke-induced pulmonary vascular remodeling in mice—histology. Representative histology from lung sections stained with antibodies against α-smooth muscle actin and von Willebrand factor.

doi:10.1371/journal.pone.0129327.g005

arterial pressure (SAP) caused by cigarette smoke exposure could be prevented by Tadalafil as well as by the combination of Tadalafil and Piclamilast but not by Piclamilast treatment alone. Although much is known about PDE inhibitors in lung diseases and such inhibitors are used in the clinic, human studies do not allow one to assess effects of PDE inhibitors on the alveolar and vascular structure. These drawbacks can be overcome by experimental studies in mice as in our investigation.

Although with these experiments we only investigated a preventive and not a curative treatment, any means to stop the progression of structural and functional deterioration in COPD would be of value if transferable to the human situation. Furthermore, it is conceivable that even longer-term treatment with these agents could cure the disease, the holy grail of lung emphysema treatment, a possibility which warrants further investigation.

Previous data suggest an important role of NO as well as sGC-cGMP signaling in the physiology and pathophysiology of pulmonary vasculature [29] as well as in airway disease [15].

PLOS ONE





Fig 6. Systemic effects of smoke exposure and treatment with Piclamilast and/or Tadalafil in mice. Mice were exposed to cigarette smoke for 6 months and treated in parallel with Piclamilast (10 mg/kg body weight/day) and/or Tadalafil (10 mg/kg body weight/day). Time matched controls were received solvent only. Systemic arterial pressure determined by *A. carotis* catheterization and quantified in anesthetized animals. Data are given as mean \pm SEM from n = 5–8, each. Bars indicate significant differences using ANOVA followed by Student-Newman-Keuls post hoc test (**P<0.01; ***P<0.001).

doi:10.1371/journal.pone.0129327.g006



Fig 7. Effect of smoke exposure, Piclamilast and/or Tadalafil treatment on the number of alveolar macrophages in mice. Lung paraffin sections were stained with a macrophage-specific antibody (F4/80) and respective alveolar macrophages from whole sections were counted and calculated as number of macrophages/mm². Data are given as mean \pm SEM from n = 5, each. Bars indicate significant differences using ANOVA with Dunnett's post hoc test (**P*<0.05).

doi:10.1371/journal.pone.0129327.g007

This system controls many vascular functions such as vascular tone, homeostasis, structure and remodeling [30, 31]. An imbalance of this system may result in PH and other pulmonary diseases [26, 32]. cGMP is not only an important regulator of short-term changes in smooth muscle tone but also in long-term responses to chronic pro-proliferative signals [33]. Nitric oxide (NO), produced by NO synthases (NOS), can activate sGC, which subsequently increases the levels of cGMP [34]. However, iNOS upregulation has been shown to cause emphysema in mice, which may be explained by iNOS-dependent sGC oxidation and inactivation [9].

In line with the previous finding that sGC stimulation by Riociguat can prevent from smoke-induced PH and emphysema in mice and guinea pigs [15], the PDE5 inhibitor Tadalafil also prevented such smoke-induced changes as demonstrated in the present study. PDE5 is highly specific for cGMP hydrolysis and is thought to be the most active cGMP hydrolyzing PDE in smooth muscle cells under basal conditions [33]. Although the mechanisms were not investigated in our study, it is well known that nitric oxide, atrial natriuretic peptide (ANP) and several other vasodilators use cGMP to regulate the vascular tone [33]. cGMP activates the cGMP-dependent protein kinase (PKG). An increase in cGMP results in a decrease of the intracellular Ca^{2+} concentration which leads to vasodilation [33]. The cGMP effects on contraction in smooth muscle seem to be specifically mediated by PKG but not by cAMP-dependent protein kinase (PKA). This finding was validated using PKG-1-deficient mice where cGMP-induced relaxation of aortic smooth muscle was completely abolished, whereas cAMP-dependent relaxation was not affected [35]. Phosphorylation of specific PKA substrates such as regulatory myosin-binding subunit of myosin phosphatase [36], IP3 receptor associated cGMP kinase substrate (IRAG) [37] and calcium-activated maxi K⁺ (BK_{ca}) channels [38], contribute to the reduction of intracellular calcium which finally causes decreased muscular tone [39]. Both cGMP and cAMP modulate acute smooth muscle relaxation, mainly through the reduction of intracellular calcium or activation of myosin phosphatase [33]. In addition to vascular tone regulation, which can also affect vascular remodeling processes e.g., by altered shear stress, cGMP elevating therapies have previously been shown to protect the organism from increased proliferation underlying pulmonary vascular remodeling [26, 30, 32, 33]. Such effects may explain the protection from vascular remodeling and PH in our study. The protection from emphysema is in line with the concept that molecular pulmonary vascular alterations drive smoke-induced emphysema development [9]. Alternatively, the protection from emphysema by PDE5 inhibition could also be independent of its vascular effects. Since only the single therapies, but not the combination therapy, prevented vascular narrowing in large vessels, it can be speculated that a parallel inhibition of both PDEs has negative effects by inhibiting anti-proliferative pathways differentially regulated in large vessels compared to small vessels.

Interestingly, application of the PDE4 inhibitor which was shown to increase cAMP [40], had quite similar effects to those observed after PDE5 inhibition. This finding is again in line with the suggestion that pulmonary vascular alterations can drive emphysema development— cAMP-elevating therapies have been shown in different animal models to protect against non-smoke-induced forms of PH [41, 42] as well and prostanoid therapy is used for treatment of human PH [43, 44]. Although the pathways of generation and regulation of cAMP and cGMP are different, the final effects in terms of vasodilation, anti-inflammation, and anti-proliferation are quite similar and not always unrelated. For instance, it could be shown that PDE5 inhibitors reduce proliferation of bovine coronary artery SMCs via cGMP elevation and subsequent inhibition of PDE3, a cGMP-inhibitable cAMP-hydrolyzing PDE [45]. These data demonstrate the possibility of increasing cAMP in a cGMP-dependent manner. Potential additional therapeutic effects of the combination of Tadalafil and Piclamilast could not be evaluated in our study as single application of each inhibitor has already had a maximum effect. Thus, investigations determining possible additive effects must be performed with lower dosages of the inhibitors.

It could be demonstrated *in vitro* that the oxidative burst in sputum from asthma and COPD patients could be reduced by Piclamilast [46]. Such anti-oxidant, but also anti-apoptotic and anti-proliferative properties of Piclamilast were recently confirmed *in vitro* by Mata and colleagues [16]. Although not shown in the present study, these Piclamilast-dependent effects could also, at least in part, explain the prevention of emphysema and vascular remodeling in smoke-exposed mice upon PDE4 inhibition.

A limitation of our study is that we could not measure cAMP/cGMP levels in our mice directly when using lung homogenate. This issue can be related either 1) to methodological limitations of our cAMP and cGMP measurement approach or 2) cell type-specific elevation of cAMP and cGMP concentration, not being reflected by lung homogenate measurements. Also variations in drug preparation, dosing, and administration strategies may result in significant differences in drug levels and, therefore, target effects. However, it was previously shown that PDE 4 and 5 inhibitors including Tadalafil and Piclamilast can elevate cGMP and cAMP levels and have been used *in vivo* in rat, rabbit, guinea pig and mouse models [27, 42, 47–56].

The decrease in SAP upon smoke exposure can principally be explained by either a direct effect of tobacco smoke on the systemic circulation or an impaired left-ventricular function due to PH-associated reductions in pre-load and cardiac output [57]. Since treatment with the PDE5 inhibitor or a combination of PDE4 and 5 inhibitors prevented PH and its corresponding pre-load effects, the latter explanation does not seem valid. Instead the data suggest a direct effect of tobacco smoke on the systemic circulation. Interestingly, only the presence of the PDE5 inhibitor could prevent the systemic effects, suggesting that cAMP elevation alone does not counteract systemic hemodynamic effects of tobacco smoke.

Although clinical studies using Sildenafil reported no change in exercise capacity, quality of life and V/Q mismatch, long term data such as in our mouse study (when comparing the life-span of mice and humans) are not available for PDE5 inhibitors in lung emphysema. Concerning PDE4 inhibitors, Roflumilast has recently been approved as anti-inflammatory therapy for treatment of COPD. However, these clinical studies do not allow the assessment of long-term effects on alveolar and lung vascular structure.

A recent clinical study using Tadalafil in COPD patients demonstrated pulmonary vasodilation, but the exercise capacity and quality of life remained unchanged [23]. Although disappointing at first, limitations of this study were:"...no right heart catheterisation for patient selection and assessment of treatment response,...use of fairly modest indicators of pulmonary vascular disease for assessment of inclusion in the study..." (false positive diagnosis at inclusion). Of interest, the authors suggest higher effects of Tadalafil in COPD patients with severe PH. This suggestion is supported by a small randomized controlled clinical trial using Sildenafil treatment in patients with severe PH. It was reported that the PAP was decreased, associated with an increase in the 6-min walk distance [24]. Since Sildenafil improves exercise capacity and quality of life in PAH patients [58] and pulmonary hemodynamics at rest and during exercise in patients with COPD-associated PH, it seems that inhibition of PDE5 could be a reliable target for patients suffering from vascular alterations. This suggestion is supported by our observation that protection against vascular remodeling and PH in cigarette smoke-exposed mice was paralleled with prevention from emphysema development. Taken together, these data support the hypothesis of an essential role of the vasculature for the development of emphysema/ COPD.

Based on the facts that PDE4 inhibitors suppress inflammatory processes in several cell types involved in COPD [59], inhibit cellular trafficking and microvascular leakage, produce reactive oxygen species, express cell adhesion molecules [60] and mediate lung smooth muscle relaxation, selective inhibitors have been developed for the treatment of COPD [61]. For instance, the PDE4 inhibitor Roflumilast has been used as anti-inflammatory therapy for the

treatment of COPD. It reduces the numbers of neutrophils (35%) and eosinophils (50%) in sputum of COPD patients [62]. Along this line Piclamilast treatment significantly prevented the smoke-induced increase in the number of alveolar macrophages in our study. Interestingly, Tadalafil tended to have the same effect. However, no such tendency was observed for the combined treatment with Piclamilast and Tadalafil, questioning that the prevention from PH and emphysema was related to macrophage/inflammatory effects. Independently, several clinical studies have demonstrated that Roflumilast [63, 64] and Cilomilast [61] can improve lung function, quality of life and reduce the frequency of exacerbations. A beneficial effect of Piclamilast in sputum of COPD patients has been reported *in vitro* [46], and now in our animal model of cigarette smoke-induced emphysema and PH. However, a clinical trial using Piclamilast in COPD or COPD-PH patients has yet to be conducted.

In conclusion, our present data substantiate the recent suggestion, which was derived from investigations of guanylate cyclase stimulators, that sGC stimulation and possible downstream signaling via cGMP can, in long-term studies, prevent lung structural and functional deterioration induced by tobacco smoke. Comparison to the effects of a PDE4 inhibitor showed in addition that another, but mechanistically quite similar pathway, the AC-cAMP signaling, may mediate comparable protective mechanisms since the elevation not only of cGMP, but also cAMP resulted in protection from cigarette smoke-induced emphysema and PH.

Supporting Information

S1 ARRIVE Checklist. (PDF)

Acknowledgments

The authors thank Guelsen Acikgoez, Lisa Fröhlich and Karin Quanz for technical assistance. This work was supported by German Research Foundation (DFG), Excellence Cluster Cardio-Pulmonary System (ECCPS). Tadalafil and Piclamilast were a kind gift from ALTANA Pharma, Konstanz, Germany.

Author Contributions

Conceived and designed the experiments: MS NP NW. Performed the experiments: MS NP AP MB. Analyzed the data: MS NP AP NW. Contributed reagents/materials/analysis tools: HAG RTS WS FG NW. Wrote the paper: MS NP AP NW.

References

- Murray CJ, Lopez AD. Alternative projections of mortality and disability by cause 1990–2020: Global Burden of Disease Study. Lancet. 1997; 349(9064):1498–504. Epub 1997/05/24. doi: <u>10.1016/S0140-6736(96)07492-2</u> PMID: <u>9167458</u>.
- Black PN, Ching PS, Beaumont B, Ranasinghe S, Taylor G, Merrilees MJ. Changes in elastic fibres in the small airways and alveoli in COPD. Eur Respir J. 2008; 31(5):998–1004. Epub 2008/01/25. doi: <u>10.</u> <u>1183/09031936.00017207</u> PMID: <u>18216063</u>.
- Agusti AG, Noguera A, Sauleda J, Sala E, Pons J, Busquets X. Systemic effects of chronic obstructive pulmonary disease. Eur Respir J. 2003; 21(2):347–60. Epub 2003/03/01. PMID: <u>12608452</u>.
- Barnes PJ. Chronic obstructive pulmonary disease. N Engl J Med. 2000; 343(4):269–80. Epub 2000/ 07/27. doi: <u>10.1056/NEJM200007273430407</u> PMID: <u>10911010</u>.
- Repine JE, Bast A, Lankhorst I. Oxidative stress in chronic obstructive pulmonary disease. Oxidative Stress Study Group. American journal of respiratory and critical care medicine. 1997; 156(2 Pt 1):341–57. Epub 1997/08/01. doi: <u>10.1164/ajrccm.156.2.9611013</u> PMID: <u>9279209</u>.

- Churg A, Cosio M, Wright JL. Mechanisms of cigarette smoke-induced COPD: insights from animal models. Am J Physiol Lung Cell Mol Physiol. 2008; 294(4):L612–31. PMID: <u>18223159</u>. doi: <u>10.1152/</u> <u>ajplung.00390.2007</u>
- Wright JL, Levy RD, Churg A. Pulmonary hypertension in chronic obstructive pulmonary disease: current theories of pathogenesis and their implications for treatment. Thorax. 2005; 60(7):605–9. Epub 2005/07/05. doi: 60/7/605 [pii] doi: <u>10.1136/thx.2005.042994</u> PMID: <u>15994270</u>; PubMed Central PMCID: PMC1747459.
- Ferrer E, Peinado VI, Diez M, Carrasco JL, Musri MM, Martinez A, et al. Effects of cigarette smoke on endothelial function of pulmonary arteries in the guinea pig. Respir Res. 2009; 10:76. PMID: <u>19682386</u>. doi: <u>10.1186/1465-9921-10-76</u>
- Seimetz M, Parajuli N, Pichl A, Veit F, Kwapiszewska G, Weisel FC, et al. Inducible NOS inhibition reverses tobacco-smoke-induced emphysema and pulmonary hypertension in mice. Cell. 147(2):293–305. Epub 2011/10/18. doi: S0092-8674(11)01005-1 [pii] doi: 10.1016/j.cell.2011.08.035 PMID: 22000010.
- Wright JL, Churg A. Cigarette smoke causes physiologic and morphologic changes of emphysema in the guinea pig. Am Rev Respir Dis. 1990; 142(6 Pt 1):1422–8. PMID: <u>2252262</u>.
- Wright JL, Churg A. Effect of long-term cigarette smoke exposure on pulmonary vascular structure and function in the guinea pig. Experimental lung research. 1991; 17(6):997–1009. Epub 1991/11/01. PMID: 1769357.
- 12. Peinado VI, Pizarro S, Barbera JA. Pulmonary vascular involvement in COPD. Chest. 2008; 134 (4):808–14. PMID: <u>18842913</u>. doi: <u>10.1378/chest.08-0820</u>
- Santos S, Peinado VI, Ramirez J, Melgosa T, Roca J, Rodriguez-Roisin R, et al. Characterization of pulmonary vascular remodelling in smokers and patients with mild COPD. Eur Respir J. 2002; 19 (4):632–8. PMID: <u>11998991</u>.
- Minai OA, Chaouat A, Adnot S. Pulmonary hypertension in COPD: epidemiology, significance, and management: pulmonary vascular disease: the global perspective. Chest. 137(6 Suppl):39S–51S. Epub 2010/06/11. doi: 137/6_suppl/39S [pii] doi: 10.1378/chest.10-0087 PMID: 20522579.
- Weissmann N, Lobo B, Pichl A, Parajuli N, Seimetz M, Puig-Pey R, et al. Stimulation of soluble guanylate cyclase prevents cigarette smoke-induced pulmonary hypertension and emphysema. American journal of respiratory and critical care medicine. 2014; 189(11):1359–73. doi: <u>10.1164/rccm.201311-</u> <u>2037OC PMID: 24738736</u>.
- Mata M, Pallardo F, Morcillo EJ, Cortijo J. Piclamilast inhibits the pro-apoptotic and anti-proliferative responses of A549 cells exposed to H(2)O(2) via mechanisms involving AP-1 activation. Free radical research. 2012; 46(5):690–9. doi: 10.3109/10715762.2012.669040 PMID: 22360706.
- Blanco I, Gimeno E, Munoz PA, Pizarro S, Gistau C, Rodriguez-Roisin R, et al. Hemodynamic and gas exchange effects of sildenafil in patients with chronic obstructive pulmonary disease and pulmonary hypertension. American journal of respiratory and critical care medicine. 2010; 181(3):270–8. Epub 2009/ 10/31. doi: <u>10.1164/rccm.200907-0988OC</u> PMID: <u>19875684</u>.
- Blanco I, Santos S, Gea J, Guell R, Torres F, Gimeno-Santos E, et al. Sildenafil to improve respiratory rehabilitation outcomes in COPD: a controlled trial. Eur Respir J. 2013; 42(4):982–92. Epub 2013/02/ 23. doi: 10.1183/09031936.00176312 PMID: 23429918.
- Holverda S, Rietema H, Bogaard HJ, Westerhof N, Postmus PE, Boonstra A, et al. Acute effects of sildenafil on exercise pulmonary hemodynamics and capacity in patients with COPD. Pulmonary pharmacology & therapeutics. 2008; 21(3):558–64. Epub 2008/03/18. doi: <u>10.1016/j.pupt.2008.01.012</u> PMID: <u>18342559</u>.
- Rietema H, Holverda S, Bogaard HJ, Marcus JT, Smit HJ, Westerhof N, et al. Sildenafil treatment in COPD does not affect stroke volume or exercise capacity. Eur Respir J. 2008; 31(4):759–64. Epub 2007/12/21. doi: <u>10.1183/09031936.00114207</u> PMID: <u>18094009</u>.
- Bull T, Badesch DB. Sildenafil for COPD: a randomized crossover trial. Copd. 2012; 9(3):211–2. Epub 2012/05/17. doi: <u>10.3109/15412555.2012.683966</u> PMID: <u>22587530</u>.
- Lederer DJ, Bartels MN, Schluger NW, Brogan F, Jellen P, Thomashow BM, et al. Sildenafil for chronic obstructive pulmonary disease: a randomized crossover trial. Copd. 2012; 9(3):268–75. Epub 2012/03/ 01. doi: 10.3109/15412555.2011.651180 PMID: 22360383.
- Goudie AR, Lipworth BJ, Hopkinson PJ, Wei L, Struthers AD. Tadalafil in patients with chronic obstructive pulmonary disease: a randomised, double-blind, parallel-group, placebo-controlled trial. The lancet Respiratory medicine. 2014; 2(4):293–300. doi: <u>10.1016/S2213-2600(14)70013-X</u> PMID: <u>24717626</u>.
- Rao RS, Singh S, Sharma BB, Agarwal VV, Singh V. Sildenafil improves six-minute walk distance in chronic obstructive pulmonary disease: a randomised, double-blind, placebo-controlled trial. The Indian journal of chest diseases & allied sciences. 2011; 53(2):81–5. Epub 2011/05/07. PMID: <u>21545068</u>.

- Alp S, Skrygan M, Schmidt WE, Bastian A. Sildenafil improves hemodynamic parameters in COPD-an investigation of six patients. Pulmonary pharmacology & therapeutics. 2006; 19(6):386–90. doi: <u>10.</u> <u>1016/j.pupt.2005.09.006</u> PMID: <u>16291503</u>.
- Dumitrascu R, Weissmann N, Ghofrani HA, Dony E, Beuerlein K, Schmidt H, et al. Activation of soluble guanylate cyclase reverses experimental pulmonary hypertension and vascular remodeling. Circulation. 2006; 113(2):286–95. Epub 2006/01/05. doi: CIRCULATIONAHA.105.581405 [pii] doi: <u>10.1161/</u> <u>CIRCULATIONAHA.105.581405</u> PMID: <u>16391154</u>.
- Woyda K, Koebrich S, Reiss I, Rudloff S, Pullamsetti SS, Ruhlmann A, et al. Inhibition of phosphodiesterase 4 enhances lung alveolarisation in neonatal mice exposed to hyperoxia. Eur Respir J. 2009; 33 (4):861–70. doi: <u>10.1183/09031936.00109008</u> PMID: <u>19010982</u>.
- Weissmann N, Dietrich A, Fuchs B, Kalwa H, Ay M, Dumitrascu R, et al. Classical transient receptor potential channel 6 (TRPC6) is essential for hypoxic pulmonary vasoconstriction and alveolar gas exchange. Proc Natl Acad Sci U S A. 2006; 103(50):19093–8. Epub 2006/12/05. doi: 0606728103 [pii] doi: 10.1073/pnas.0606728103 PMID: 17142322; PubMed Central PMCID: PMC1748182.
- Hoeper MM, Halank M, Wilkens H, Gunther A, Weimann G, Gebert I, et al. Riociguat for interstitial lung disease and pulmonary hypertension: a pilot trial. Eur Respir J. 2013; 41(4):853–60. doi: <u>10.1183/</u>09031936.00213911 PMID: 22936711.
- Schermuly RT, Janssen W, Weissmann N, Stasch JP, Grimminger F, Ghofrani HA. Riociguat for the treatment of pulmonary hypertension. Expert Opin Investig Drugs. 20(4):567–76. Epub 2011/03/12. doi: 10.1517/13543784.2011.565048 PMID: 21391889.
- Schermuly RT, Stasch JP, Pullamsetti SS, Middendorff R, Muller D, Schluter KD, et al. Expression and function of soluble guanylate cyclase in pulmonary arterial hypertension. Eur Respir J. 2008; 32 (4):881–91. Epub 2008/06/14. doi: 09031936.00114407 [pii] doi: <u>10.1183/09031936.00114407</u> PMID: <u>18550612</u>.
- Evgenov OV, Pacher P, Schmidt PM, Hasko G, Schmidt HH, Stasch JP. NO-independent stimulators and activators of soluble guanylate cyclase: discovery and therapeutic potential. Nat Rev Drug Discov. 2006; 5(9):755–68. Epub 2006/09/07. doi: nrd2038 [pii] doi: <u>10.1038/nrd2038</u> PMID: <u>16955067</u>; PubMed Central PMCID: PMC2225477.
- Rybalkin SD, Yan C, Bornfeldt KE, Beavo JA. Cyclic GMP phosphodiesterases and regulation of smooth muscle function. Circulation research. 2003; 93(4):280–91. doi: <u>10.1161/01.RES.0000087541.</u> <u>15600.2B</u> PMID: <u>12933699</u>.
- Murad F. Cyclic guanosine monophosphate as a mediator of vasodilation. The Journal of clinical investigation. 1986; 78(1):1–5. doi: <u>10.1172/JCI112536</u> PMID: <u>2873150</u>; PubMed Central PMCID: PMC329522.
- Hofmann F, Ammendola A, Schlossmann J. Rising behind NO: cGMP-dependent protein kinases. Journal of cell science. 2000; 113 (Pt 10):1671–6. PMID: <u>10769198</u>.
- Surks HK, Mochizuki N, Kasai Y, Georgescu SP, Tang KM, Ito M, et al. Regulation of myosin phosphatase by a specific interaction with cGMP- dependent protein kinase lalpha. Science. 1999; 286 (5444):1583–7. PMID: 10567269.
- Schlossmann J, Ammendola A, Ashman K, Zong X, Huber A, Neubauer G, et al. Regulation of intracellular calcium by a signalling complex of IRAG, IP3 receptor and cGMP kinase lbeta. Nature. 2000; 404 (6774):197–201. doi: <u>10.1038/35004606</u> PMID: <u>10724174</u>.
- Fukao M, Mason HS, Britton FC, Kenyon JL, Horowitz B, Keef KD. Cyclic GMP-dependent protein kinase activates cloned BKCa channels expressed in mammalian cells by direct phosphorylation at serine 1072. The Journal of biological chemistry. 1999; 274(16):10927–35. PMID: <u>10196172</u>.
- Schlossmann J, Feil R, Hofmann F. Signaling through NO and cGMP-dependent protein kinases. Annals of medicine. 2003; 35(1):21–7. PMID: <u>12693609</u>.
- 40. Hertz AL, Bender AT, Smith KC, Gilchrist M, Amieux PS, Aderem A, et al. Elevated cyclic AMP and PDE4 inhibition induce chemokine expression in human monocyte-derived macrophages. Proc Natl Acad Sci U S A. 2009; 106(51):21978–83. doi: <u>10.1073/pnas.0911684106</u> PMID: <u>19959669</u>; PubMed Central PMCID: PMC2799834.
- Izikki M, Raffestin B, Klar J, Hatzelmann A, Marx D, Tenor H, et al. Effects of roflumilast, a phosphodiesterase-4 inhibitor, on hypoxia- and monocrotaline-induced pulmonary hypertension in rats. The Journal of pharmacology and experimental therapeutics. 2009; 330(1):54–62. doi: <u>10.1124/jpet.108.148742</u> PMID: <u>19386793</u>.
- Schermuly RT, Kreisselmeier KP, Ghofrani HA, Samidurai A, Pullamsetti S, Weissmann N, et al. Antiremodeling effects of iloprost and the dual-selective phosphodiesterase 3/4 inhibitor tolafentrine in chronic experimental pulmonary hypertension. Circulation research. 2004; 94(8):1101–8. doi: <u>10.1161/01.</u> <u>RES.0000126050.41296.8E</u> PMID: <u>15031263</u>.

- 43. Olschewski H. Inhaled iloprost for the treatment of pulmonary hypertension. European respiratory review: an official journal of the European Respiratory Society. 2009; 18(111):29–34. doi: <u>10.1183/09059180.00011111</u> PMID: <u>20956120</u>.
- Olschewski H, Simonneau G, Galie N, Higenbottam T, Naeije R, Rubin LJ, et al. Inhaled iloprost for severe pulmonary hypertension. N Engl J Med. 2002; 347(5):322–9. doi: <u>10.1056/NEJMoa020204</u> PMID: <u>12151469</u>.
- Osinski MT, Rauch BH, Schror K. Antimitogenic actions of organic nitrates are potentiated by sildenafil and mediated via activation of protein kinase A. Molecular pharmacology. 2001; 59(5):1044–50. PMID: <u>11306686</u>.
- Beeh KM, Beier J, Lerch C, Schulz AK, Buhl R. Effects of piclamilast, a selective phosphodiesterase-4 inhibitor, on oxidative burst of sputum cells from mild asthmatics and stable COPD patients. Lung. 2004; 182(6):369–77. PMID: <u>15765929</u>.
- Teixeira CE, Priviero FB, Webb RC. Differential effects of the phosphodiesterase type 5 inhibitors sildenafil, vardenafil, and tadalafil in rat aorta. The Journal of pharmacology and experimental therapeutics. 2006; 316(2):654–61. doi: <u>10.1124/jpet.105.092544</u> PMID: <u>16204472</u>.
- Bundschuh DS, Eltze M, Barsig J, Wollin L, Hatzelmann A, Beume R. In vivo efficacy in airway disease models of roflumilast, a novel orally active PDE4 inhibitor. The Journal of pharmacology and experimental therapeutics. 2001; 297(1):280–90. PMID: 11259555.
- de Visser YP, Walther FJ, Laghmani el H, Steendijk P, Middeldorp M, van der Laarse A, et al. Phosphodiesterase 4 inhibition attenuates persistent heart and lung injury by neonatal hyperoxia in rats. Am J Physiol Lung Cell Mol Physiol. 2012; 302(1):L56–67. doi: <u>10.1152/ajplung.00041.2011</u> PMID: <u>21949154</u>; PubMed Central PMCID: PMC3349366.
- Dony E, Lai YJ, Dumitrascu R, Pullamsetti SS, Savai R, Ghofrani HA, et al. Partial reversal of experimental pulmonary hypertension by phosphodiesterase-3/4 inhibition. Eur Respir J. 2008; 31(3):599–610. doi: 10.1183/09031936.00002007 PMID: 18032446.
- Schermuly RT, Kreisselmeier KP, Ghofrani HA, Yilmaz H, Butrous G, Ermert L, et al. Chronic sildenafil treatment inhibits monocrotaline-induced pulmonary hypertension in rats. American journal of respiratory and critical care medicine. 2004; 169(1):39–45. doi: 10.1164/rccm.200302-282OC PMID: 12958054.
- Selige J, Tenor H, Hatzelmann A, Dunkern T. Cytokine-dependent balance of mitogenic effects in primary human lung fibroblasts related to cyclic AMP signaling and phosphodiesterase 4 inhibition. Journal of cellular physiology. 2010; 223(2):317–26. doi: <u>10.1002/jcp.22037</u> PMID: <u>20082309</u>.
- Udalov S, Dumitrascu R, Pullamsetti SS, Al-tamari HM, Weissmann N, Ghofrani HA, et al. Effects of phosphodiesterase 4 inhibition on bleomycin-induced pulmonary fibrosis in mice. BMC Pulm Med. 2010; 10:26. doi: <u>10.1186/1471-2466-10-26</u> PMID: <u>20444277</u>; PubMed Central PMCID: PMC2881047.
- Weissmann N, Gerigk B, Kocer O, Nollen M, Hackemack S, Ghofrani HA, et al. Hypoxia-induced pulmonary hypertension: different impact of iloprost, sildenafil, and nitric oxide. Respir Med. 2007; 101 (10):2125–32. doi: <u>10.1016/j.rmed.2007.05.025</u> PMID: <u>17643279</u>.
- 55. Weissmann N, Peters DM, Klopping C, Kruger K, Pilat C, Katta S, et al. Structural and functional prevention of hypoxia-induced pulmonary hypertension by individualized exercise training in mice. Am J Physiol Lung Cell Mol Physiol. 2014; 306(11):L986–95. doi: <u>10.1152/ajplung.00275.2013</u> PMID: 24705723.
- Yang J, Li X, Al-Lamki RS, Wu C, Weiss A, Berk J, et al. Sildenafil potentiates bone morphogenetic protein signaling in pulmonary arterial smooth muscle cells and in experimental pulmonary hypertension. Arterioscler Thromb Vasc Biol. 2013; 33(1):34–42. doi: <u>10.1161/ATVBAHA.112.300121</u> PMID: <u>23139294</u>.
- Chin KM, Kim NH, Rubin LJ. The right ventricle in pulmonary hypertension. Coronary artery disease. 2005; 16(1):13–8. Epub 2005/01/18. PMID: <u>15654194</u>.
- Galie N, Brundage BH, Ghofrani HA, Oudiz RJ, Simonneau G, Safdar Z, et al. Tadalafil therapy for pulmonary arterial hypertension. Circulation. 2009; 119(22):2894–903. Epub 2009/05/28. doi: CIRCULA-TIONAHA.108.839274 [pii] doi: 10.1161/CIRCULATIONAHA.108.839274 PMID: 19470885.
- Huang Z, Mancini JA. Phosphodiesterase 4 inhibitors for the treatment of asthma and COPD. Current medicinal chemistry. 2006; 13(27):3253–62. Epub 2006/12/16. PMID: 17168849.
- 60. Sanz MJ, Cortijo J, Morcillo EJ. PDE4 inhibitors as new anti-inflammatory drugs: effects on cell trafficking and cell adhesion molecules expression. Pharmacology & therapeutics. 2005; 106(3):269–97. Epub 2005/06/01. doi: <u>10.1016/j.pharmthera.2004.12.001</u> PMID: <u>15922015</u>.
- Brown WM. Treating COPD with PDE 4 inhibitors. International journal of chronic obstructive pulmonary disease. 2007; 2(4):517–33. Epub 2008/02/14. PMID: <u>18268925</u>; PubMed Central PMCID: PMC2699952.

- Grootendorst DC, Gauw SA, Verhoosel RM, Sterk PJ, Hospers JJ, Bredenbroker D, et al. Reduction in sputum neutrophil and eosinophil numbers by the PDE4 inhibitor roflumilast in patients with COPD. Thorax. 2007; 62(12):1081–7. Epub 2007/06/19. doi: <u>10.1136/thx.2006.075937</u> PMID: <u>17573446</u>; PubMed Central PMCID: PMC2094292.
- **63.** Cowan C. Roflumilast for asthma and chronic obstructive pulmonary disease. Issues in emerging health technologies. 2005;(74:):1–4. Epub 2005/12/02. PMID: <u>16317827</u>.
- Rabe KF, Bateman ED, O'Donnell D, Witte S, Bredenbroker D, Bethke TD. Roflumilast–an oral anti-inflammatory treatment for chronic obstructive pulmonary disease: a randomised controlled trial. Lancet. 2005; 366(9485):563–71. Epub 2005/08/16. doi: <u>10.1016/S0140-6736(05)67100-0</u> PMID: <u>16099292</u>.