



Review Article

Endocrinol Metab 2022;37:559-574

A Key Metabolic Regulator of Bone and Cartilage Health

Elizabeth Pérez-Hernández¹, Jesús Javier Pastrana-Carballo², Fernando Gómez-Chávez², Ramesh C. Gupta³, Nury Pérez-Hernández²

¹Medical Unit of High Specialty of Traumatology, Orthopedics and Rehabilitation "Dr. Victorio de la Fuente Narváez", Mexican Social Security Institute; ²National School of Medicine and Homeopathy, National Polytechnic Institute, Mexico City, Mexico; ³School of Agricultural Sciences and Rural Development (SASRD) Nagaland University, Medziphema, India

Taurine, a cysteine-derived zwitterionic sulfonic acid, is a common ingredient in energy drinks and is naturally found in fish and other seafood. In humans, taurine is produced mainly in the liver, and it can also be obtained from food. In target tissues, such as the retina, heart, and skeletal muscle, it functions as an essential antioxidant, osmolyte, and antiapoptotic agent. Taurine is also involved in energy metabolism and calcium homeostasis. Taurine plays a considerable role in bone growth and development, and high-profile reports have demonstrated the importance of its metabolism for bone health. However, these reports have not been collated for more than 10 years. Therefore, this review focuses on taurine-bone interactions and covers recently discovered aspects of taurine's effects on osteoblastogenesis, osteoclastogenesis, bone structure, and bone pathologies (e.g., osteoporosis and fracture healing), with due attention to the taurine-cartilage relationship.

Keywords: Taurine; Osteoblasts; Osteoclasts; Osteoporosis; Cartilage; Osteoarthritis

INTRODUCTION

Taurine, a nonessential amino acid, is a frequently used ingredient in energy drinks, as it improves athletic and mental performance. This amino acid is also an essential regulator of physiological processes and pathologies, such as metabolism and cancer development; this aspect was either unknown or ignored until the last decade, when it attracted the attention of scientists [1-3].

Taurine was discovered in Germany in 1827 by Tiedemann and Gmelin [4], who isolated it from ox (Bos taurus) bile. Ten years later, Demarcay [5] named this amino acid taurine, and 20 years later, Jacobsen and Smith [6] found that its structure contains sulfur. Since then, the natural occurrence of taurine has been recognized in a wide variety of invertebrate and vertebrate tissues [6] in some plants, algae, and fungi [7]. In mammals, except felines and foxes, which cannot synthesize taurine, the total taurine pool is balanced between synthesis and absorption from food. In humans, taurine plays a functional role in vital organs, such as the brain, eyes, kidneys, and heart. It performs several primary physiological functions, including osmotic regulation, and has antioxidant, antiapoptotic, and anti-inflammatory effects [1]. The mechanisms through which taurine exerts these effects

Received: 24 February 2022, Revised: 28 May 2022, Accepted: 7 June 2022

Corresponding authors: Nury Pérez-Hernández

National School of Medicine and Homeopathy, National Polytechnic Institute, 07320, Mexico City, Mexico

Tel: +52-5729-6000 ext. 55537, Fax: +52-5729-6000, E-mail: nperezh@ipn.mx

Ramesh C. Gupta

School of Agricultural Sciences and Rural Development (SASRD) Nagaland University, Medziphema-797106, India

Tel: +91-3862-247102, Fax: +91-3862-247113,

E-mail: rameshgupta1954@gmail.com

Copyright © 2022 Korean Endocrine Society

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (https://creativecommons.org/ licenses/by-nc/4.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

EnM

are not completely known, but are currently being unveiled. This review aims to provide an overview of taurine—bone interactions based on discoveries in the last decade. We summarize how this amino acid affects bone development and functions, as well as orthopedic disorders, including bone cancer.

PROPERTIES OF TAURINE

Taurine (2-aminoethane-1-sulfonic acid) is a sulfur-containing amino acid with a β -amino group and an acidic sulfonic group (R-SO₃H) separated by two methylene (CH₂) moieties. Unlike several other amino acids, taurine does not contain a carboxyl group; therefore, it is considered a nonproteinogenic amino acid. Taurine's molecular formula is C₂H₇NO₃S; its exact molecular mass is 125.014664 g/mol; its melting point is 300°C; and its solubility in water is 80.7 mg/mL. Although taurine has an isoelectric point of 5.15, it is considered a zwitterion at physiological pH, with a pK_a value of 1.05 and pK_b value of 9. Taurine is commercially manufactured from ethylene oxide or

monoethanolamine [8].

BIOSYNTHESIS OF TAURINE

The primary source of taurine for humans is the diet, especially seafood and meat [9]. Human milk or formula compensates for insufficient synthesis in infants and children, which develops until adulthood. Endogenous synthesis in adults occurs in discrete amounts, ranging from 0.4 to 1.0 mmol per day, depending on protein intake and hepatic function [10]. *De novo* production of taurine has been mainly found in the liver and brain; however, other tissues, including the kidney, skeletal muscle, white and brown adipose tissues, pineal gland, and retina, can also synthesize it [6]. There are three putative pathways for taurine production from cysteine. The first is the cysteine sulfinate pathway (I), which includes two reactions: decarboxylation of cysteine sulfinate to hypotaurine by cysteine sulfinate decarboxylase, and oxidation of hypotaurine to taurine by hypotaurine dehydrogenase [11]. In pathway (II), cysteine sulfinate is con-

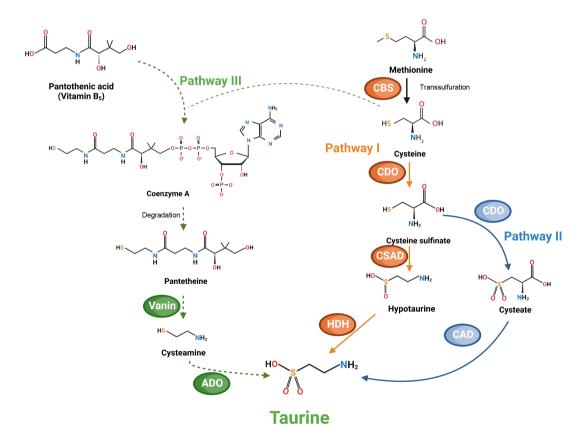


Fig. 1. Biosynthetic pathways of taurine in humans. Orange and blue arrows indicate synthesis pathways (I) and (II) from cysteine, respectively. Synthesis pathway (III) is indicated in green arrows [13] (created with BioRender.com). CBS, cystathionine β-synthase; CDO, cysteine dioxygenase; CSAD, cysteine sulfinate decarboxylase; HDH, hypotaurine dehydrogenase; CAD, candysteic acid decarboxylase; ADO, 2-aminoethanethiol dioxygenase.

verted to cysteate and taurine by cysteine dioxygenase and cysteate decarboxylase, respectively. The third pathway (III) uses cysteamine produced by the degradation of coenzyme A, and finally, cysteamine is transformed into hypotaurine by 2-aminoethanethiol dioxygenase (Fig. 1) [12,13].

ABSORPTION, DISTRIBUTION, AND EXCRETION OF TAURINE

The taurine supply in the human body is covered by intestinal uptake and biosynthesis, mainly in the liver. After oral ingestion, intestinal transport moves taurine by two carriers: proton-coupled amino acid transporter 1 (PAT1) and sodium- and chloride-dependent taurine transporter (TAUT). PAT1 has pH-dependent activity on the millimolar order and functions as the primary transporter at higher taurine concentrations; this protein also transports glycine, alanine, and proline [14]. TAUT is a beta-alanine transporter; unlike PAT1, its taurine affinity is in the micromolar range, but with low transport capacity [15]. In addi-

tion to endogenous synthesis, taurine content in the liver (approximately 10 mM) is maintained by portal vein transport and taurine conjugated with bile acids, which are products of cholesterol catabolism. Taurine metabolism involves the formation of 5-glutamyl-taurine and taurocholate, and its excretion occurs in the kidney through urine or by the enterohepatic cycle through feces conjugated to bile acids (Fig. 2) [13,14,16].

CELLULAR FUNCTIONS OF TAURINE

Antioxidant activity

Taurine is frequently classified as an antioxidant; it scavenges reactive oxygen species (ROS), reduces ROS generation, and alleviates the harmful effects of oxidative stress. Free radical scavenging by taurine depends on its concentration; it is effective against peroxyl radicals (ROO•), superoxide anion (O2•¯), nitric oxide (NO•), and peroxynitrite (ONOO¯), but it does not scavenge H₂O₂ [17]. However, the main antioxidant property of taurine is attributed to its mitochondrial protective effect, which

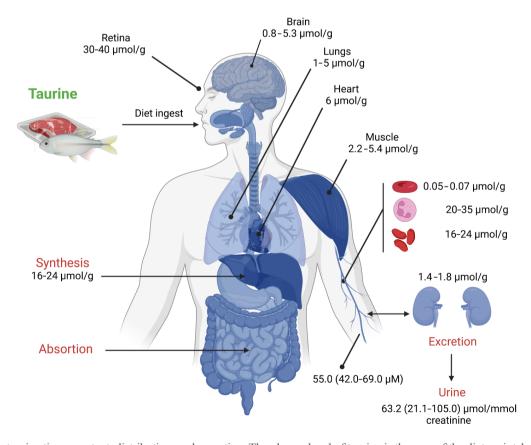


Fig. 2. Human taurine tissue content, distribution, and excretion. The plasma level of taurine is the sum of the dietary intake and the endogenous synthesis by the liver, while the kidney is the primary organ for its excretion. Tissues with remarkably high taurine accumulation include the retina, heart, lung, and skeletal muscle, as well as some hematopoietic cells such as erythrocytes, platelets, and white blood cells. Data represents the amount per day. Data taken from [13] (created with BioRender.com).

EnM

blocks ROS generation [18]. Taurine reacts with uridines of mitochondrial tRNAs to form 5-taurinomethyluridine (τ m5U) and 5-taurinomethyl-2-thiouridine (\tau m5s2U), thereby improving the codon-anticodon interaction between the uridine-uridine-guanosine (UUG) codon and taurine-modified adenosine-adenosineuridine, optimizing the translation of encoded proteins that are rich in UUG regions of subunits 5 (mt-ND5) and 6 (mt-ND6) of the respiratory chain complex I [19]. Another antioxidant mechanism of taurine involves normalizing the enzymatic machinery against oxidative stress in the cell. Many studies have described the restoration of levels of catalase, glutathione peroxidase, glutathione, superoxide dismutase (SOD), and thioredoxin reductase by taurine treatment after exposure to oxidative inducers [1,20], with the most recent studies focusing on diabetes-induced oxidative stress [21]. In contrast, the generation of taurochloramine (TauCl) and taurobromide (TauBr) by the reaction of taurine hypochlorous and hypobromous acids derived from the myeloperoxidase system in activated neutrophils is an indirect antioxidant mechanism of this amino acid [22]. Taurine haloamines significantly induce antioxidant enzyme expression, promoting the nuclear translocation of nuclear factor E2-related factor 1 (Nrf2) (Fig. 3) [23]. The following sections describe additional properties of these taurine analogs.

Calcium homeostasis

Taurine can preserve intracellular calcium (Ca²⁺) concentrations by inhibiting influx from extracellular routes or releasing it from intracellular pools. One of the known mechanisms underlying

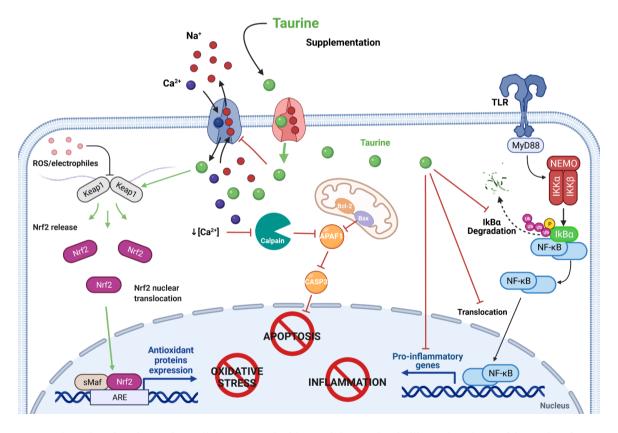


Fig. 3. Cytoprotective roles of taurine against cell damage. Antioxidant activity: taurine facilitates the release of the nuclear factor E2-related factor (Nrf2), a redox-sensitive transcription factor, by the Kelch-like ECH-associated protein 1 (Keap1), and the translocation of Nrf2 to the nucleus and its binding to the antioxidant response element (ARE) activates the gene transcription of antioxidant enzymes. Anti-apoptotic activity: taurine's contribution to calcium homeostasis blocks the activation of the calpain-dependent apoptotic cascade. Anti-inflammatory activity: taurine represses the degradation of the inhibitor of nuclear factor-κΒ (NF-κΒ) alpha (IκΒα) and keeps NF-κΒ inactivated; it also prevents its translocation to the nucleus and reduces the transcription of genes encoding proinflammatory cytokines (created with BioRender.com). APAF1, apoptosis protease-activating factor-1; Bax, Bcl-2-associated X protein; BCl-2, B-cell lymphoma 2; CASP3, caspase-3; Ikβα, nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor, alpha; Keap1, Kelch-like ECH-associated protein 1; MyD88, innate immune signal transduction adaptor; Nrf2, NEMO, NF-kappa-B essential modulator; sMaF, musculoaponeurotic fibrosarcoma; TLR, Toll-like receptor.

this effect is the extrusion of taurine and sodium (Na⁺) from the cell via taurine/Na⁺ symport, which limits Ca²⁺ overload through Na⁺/Ca²⁺ exchange in ischemia-reperfusion injuries [24]. This effect suggests the therapeutic use of taurine in surgery for cardiac damage, heart transplantation, and myocardial infarction [25]. Taurine also regulates the activity of Ca²⁺-ATPase in the sarcoplasmic reticulum, ultimately maintaining Ca²⁺ concentrations, which has implications for diastolic dysfunction of the heart or impaired fibers type I and II of the skeletal muscle [26,27]. The role of taurine in calcium-sensitive excitation-contraction in muscular pathologies has been used to test the hypothesis that increasing taurine intake can improve skeletal muscle function in a mouse model of Duchenne muscular dystrophy [28]. Taurine also modulates the calcium influx produced by coupling glutamate to N-methyl-d-aspartate receptors, preventing membrane depolarization by opening chloride channels or inhibiting L-, P/Q-, and N-type voltage-gated Ca²⁺ channels [29].

Cell death

Closely related to its antioxidant activity, taurine maintains normal cellular function and regulates apoptosis (Fig. 3). The interruption of the respiratory chain by a deficiency in complex I has been demonstrated in taurine-deficient models. In fact, mitochondrial damage leads to increased membrane permeability. inducing cytochrome c release, apoptosome formation, and activation of caspase-9 and caspase-3 [30]. Taurine also inhibits apoptosis induced by toxic agents, such as tert-butyl hydroperoxide [31], acrylamide [32], and arsenic [33], in cell cultures that activate the phosphatidylinositol 3-kinase pathway/Akt and inhibit nuclear translocation of nuclear factor-kappa B (NF-κB). Interestingly, taurine supplementation protects osteocyte-like cells from cell death triggered by ROS in vitro [34]. This nonclassical amino acid is also involved in neuronal cell death protection against elevated circulating corticosteroids due to chronic stress, by positively regulating extracellular signal-regulated kinase and Nrf2 [35]. Therefore, taurine has been suggested as a potential neuroprotective agent, particularly for glaucoma treatment [36]. More recently, taurine has been demonstrated to limit arsenic-induced cell death by downregulating pyroptosis and autophagy in vitro and in vivo, thereby reducing hepatotoxicity [37]. In addition, taurine has been proposed as a potential therapeutic agent for treating alcohol-induced cell death and nonalcoholic fatty liver disease resulting from obesity [38,39].

Energy metabolism

Although taurine biosynthesis is limited in humans, it also plays

a vital role in energy metabolism. Under taurine deficiency, skeletal and heart muscles, liver, and adipose tissue show metabolic and physical dysfunction, highlighting the downregulation of genes involved in fatty acid metabolism [40]. Taurine supplementation in the diet can also strengthen energy metabolism for muscle performance, cardiac function, and liver activity [25,41,42]. For example, in obesity models, taurine treatment lowers fatty acid oxidation, decreases weight, and improves hyperglycemia and hyperinsulinemia. Details of these effects and their molecular mechanisms have recently been compiled elsewhere [40,43].

Inflammation

Although the antioxidant properties of the taurine derivatives, TauCl and TauBr, have already been noted, their anti-inflammatory properties are remarkable. TauCl inhibits the overproduction of inflammatory mediators, such as tumor necrosis factor-α, interleukin-1, -2, -6, -8, 10, and -12, monocyte chemo-attractant protein-1 and -2, cyclooxygenase (COX)-2, and prostaglandin E2 (PGE2) [44]. The regulation of all these molecules converges on the inactivation of IkappaB kinase, which phosphorylates the inhibitor kappa B-alpha, thereby preventing its degradation and promoting its union with NF-κB involved in the synthesis of proinflammatory mediators (Fig. 3) [45]. The beneficial effects of taurine have recently been explained by the brain injury and inflammation mechanisms in patients with type 2 diabetes [21,46]. Furthermore, new insights have been gained into how the microbiome regulates taurine and healthy aging. Ahmadi et al. [47] reported that a human-origin probiotic cocktail from a healthy infant gut prevented high-fat diet-induced inflammation in elderly mice in a manner dependent on increased taurine abundance in the gut, favoring healthy aging. Additionally, topically applied taurine can regulate skin inflammation caused by ultraviolet light exposure, enabling this amino acid to be suitable for the amelioration of dermatitis and other skin pathologies [48]. The anti-inflammatory role of taurine has been addressed in other conditions, such as adult mouse lungs exposed to cigarette smoke [49], carbon tetrachloride-induced testicular and kidney damage [50], and ulcerative colitis-colorectal cancer [51]. As the inflammatory effect of taurine has become markedly relevant, it is now being used as a biopharmaceutical coupled to collagen sponges to improve wound healing and tissue proliferation [52]. Furthermore, evidence from clinical trials has shown that taurine can significantly decrease the levels of protein C reactive and malondialdehyde 56 days after its supplementation, thereby exhibiting beneficial effects on inflammato-

EnM

ry and oxidative stress [53].

Osmotic regulation

Osmoregulation in cells involves the maintenance of salt and water balance, with adjustment of their osmolyte content to minimize injury to the membrane under osmotic disequilibrium. Unlike charged osmolytes, taurine is a neutral zwitterion released or introduced by cells to compensate for deviations from normotonicity, which can occur by different mechanisms [54]. Swelling-induced taurine efflux is energy-independent, temperature-sensitive, and pH-sensitive, and is regulated by non-hydrolytic adenosine triphosphate (ATP) binding via a volumeregulated anion channel [55]. Although taurine efflux is believed to be Ca²⁺-independent, it was recently demonstrated that it depends on Ca²⁺ in an insensitive fraction of human retinal Müller cells [56]. Under conditions of hypertonicity, taurine accumulation is mediated by TAUT based on in vivo studies and cultured astrocytes, which has been suggested to be a concerted uptake/biosynthesis interaction; however, this notion has not been well established [57]. Fig. 3 summarizes the functions of taurine in cell protection.

TAURINE AND BONE

Bone is a mineralized connective tissue that constitutes the skeletal system and provides mechanical support and protection for organs and bone marrow, as well as conferring strength, toughness, and capacity for continuous growth and remodeling throughout post-fetal life [58]. Osteoblasts, osteoclasts, and osteocytes, the bone cells, are immersed in an organic and inorganic matrix composed of collagen type I, a smaller proportion of non-collagen proteins, and nanosized hydroxyapatite crystals [59].

Two processes are involved in bone growth: intramembranous ossification and endochondral ossification. Intramembranous ossification involves the transition from mesenchymal cells to osteoblasts, mainly by the action of the Runt-related transcription factor 2 (RUNX2) [60], while endochondral ossification involves the formation of a hyaline cartilage template, which is replaced by mineralized bone tissue by the action of SRY-box transcription factor (SOX9) [61]. Once the bone has formed, bone remodeling is described as either the formation of bone by osteoblasts or resorption by osteoclasts.

Different amino acids act as precursors of bioactive macromolecules, such as second messengers, cytokines, or neurotransmitters, that control various cellular processes that affect the skeletal system [62]. Although taurine is not a structural component of proteins, it is metabolically involved in many processes that influence bone development [63]. Interestingly, the influence of taurine on bone growth was elucidated less than 10 years ago by Roman-Garcia, who found that vitamin B12 deficiency induced growth retardation and osteoporosis in a mice model parallel with low taurine production by the liver. This effect results in the abrogation of the growth hormone-insulin-like growth factor I axis (GH/IGF-I) and GH resistance. As GH controls taurine synthesis in a vitamin-B12 dependent manner, this amino acid is consequently an upstream regulator of IGF-I production in the liver and a regulator in osteoblasts [64]. Although this study helps to understand the role of taurine in bone development at the molecular level, the impact of taurine at the macro level and on the microstructure has scarcely been explored. Furthermore, few studies have been conducted in the last two decades. At the time of this review, the only study that evaluated the impact of taurine ingestion on human bones revealed significantly higher serum osteocalcin levels in patients with type II diabetes than in controls, suggesting that taurine may modulate bone mineralization [65]. This finding is supported by studies using animal models, which revealed the positive effects of taurine on bone development. For instance, in a mouse model of protein malnutrition, increases in growth plate thickness and improvement in bone volume, trabecular number, connection density, and total porosity were observed after taurine consumption for 12 weeks. Taurine is proposed to exert these effects by elevating IGF-1 levels and promoting the phosphorylation of Janus kinase 2-signal transducer and activator of transcription 5, as these markers are enhanced in the serum, liver, and tibia-growth plate [66]. Consistent with this study, male rats fed a diet containing 2% taurine for 6 weeks had significantly higher femur bone mineral content (BMC)/ weight than control rats, suggesting that taurine may have a beneficial effect on bone even though the markers, alkaline phosphatase (ALP) and osteocalcin were not affected [67]. The third study involved the effect of taurine supplementation in the diet of broiler birds, which, after 42 days, presented notably higher levels of serum ALP and bone-specific ALP than control birds. In an analysis of BMC, birds fed a taurine-supplemented diet had the highest percentage of phosphorous, calcium, and phosphate composition, indicating that taurine supplementation boosted the bone mineral composition of meat-type poultry birds [68].

In contrast to previous studies, the oral administration of taurine to mice for 8 weeks decreased cortical and trabecular bone thickness and reduced the density of secondary osteons [69]. Although this previous study suggests the negative effect of prolonged taurine exposure on the bone tissue microstructure, the lack of sufficient studies prevents the drawing of conclusions. Undoubtedly, the effect of taurine on the typical bone structure must be studied thoroughly, as most analyses have focused on its positive effect on damage caused by osteoporosis; such studies are described in the next section.

Taurine and osteoblasts

Osteoblasts are bone-building cells that synthesize collagen matrix and mineralize bone. The mesenchymal progenitors of osteoblasts are activated by specific transcription factors that operate at a distinct time point during differentiation, defining various stages of the osteoblast lineage. The two master transcription regulators in osteoblast differentiation are SOX9 and

RUNX2, the first of which mediates the differentiation of mesenchymal stem cells into chondrogenic progenitors but is not expressed in mature osteoblasts. In contrast, RUNX2 is necessary to differentiate osteoblasts from precursors [70]. The regulation of RUNX2 involves several pathways, such as Hedgehog, Notch, Wnt, bone morphogenetic protein (BMP), and fibroblast growth factor (FGF) signaling [71]. Besides SOX9 and RUNX2, other transcription factors, such as osterix, play essential roles in osteoblastic differentiation.

The relationship between taurine and osteoblasts is being elucidated. To date, two studies have shown that taurine stimulates osteogenesis. In MG-63 human osteoblast-like cells, taurine was recently shown to increase the expression levels of RUNX2 and intermediates of the BMP pathway, such as SMAD-1,5,8. These effects enhance the activity of ALP, which is a marker of osteogenic differentiation (Fig. 4) [72]. In mesenchymal stem

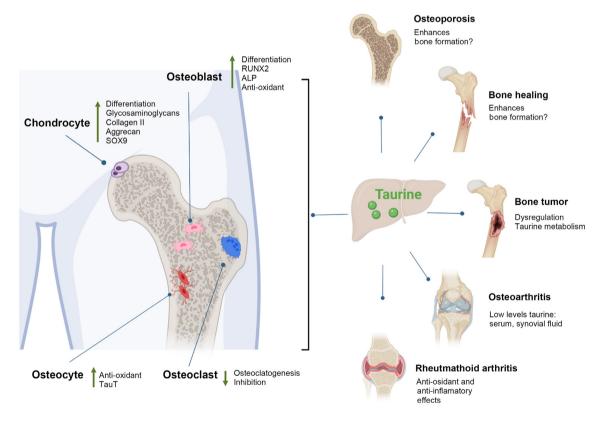


Fig. 4. Potential actions of taurine in bone and cartilage. Taurine promotes osteogenesis by upregulating Runt-related transcription factor 2 (RUNX2), as well as increasing alkaline phosphatase (ALP) activity and calcium deposition by osteoblasts; it also plays an antioxidant role in osteoblasts and osteocytes. In osteoclastogenesis, taurine exerts an inhibitory function. In chondrogenic differentiation, taurine promotes the synthesis of extracellular matrix components such as glycosaminoglycans and collagen, in addition to increasing the expression of SRY-box transcription factor 9 (SOX9). In bone and cartilage health, taurine promotes the formation of bone calluses in the consolidation process and promotes osseointegration. In pathological conditions such as osteoporosis, taurine supplementation increases bone mass, while in osteoarthritis, rheumatoid arthritis, and some bone tumors there are alterations in the metabolism of taurine (created with BioRender.com). TauT, taurine transporter.

cells, taurine promotes osteoblastogenesis, increasing ALP activity and calcium deposition owing to the upregulation of osteopontin, RUNX2, and osterix mRNA [73]. Taurine also exerted cytoprotective effects on osteoblasts under stress, as demonstrated in osteoblast-like UMR-106 cells. Pre-treatment with taurine can evade apoptosis induced by H_2O_2 , an effect attributed to the overexpression of Nrf2 and β -catenin, which reduce oxidative damage [74]. In contrast, taurine was found to reduce apoptosis in MC3T3-E1 cells (osteoblast precursor cells) by inhibiting cytochrome C release and caspase-3 and caspase-9 activation [75].

The consumption of taurine by preosteoblasts or osteoblasts must play an important role in the differentiation or protective effect; thus, the transport of taurine has attracted attention. In 2006, TAUT mRNA and protein expression was detected in a murine osteoblast cell line (MC3T3-E1) and primary human osteoblasts [76]. Two years later, taurine uptake via TauT was found to be dependent on the presence of extracellular calcium, which was decreased by L-type calcium channel blockers and oxidative agents, such as H₂O₂, in MC3T3-E1 cells [77]. Although the participation of taurine in osteoblast differentiation and maintenance is evident, more studies are necessary to understand the detailed mechanism by which these events occur.

Taurine and osteoclasts

As cells complementary to osteoblasts, osteoclasts are multinuclear giant cells that can degrade bone by secreting H⁺, Cl⁻, cathepsin K, and matrix metalloproteinases (MMPs). Osteoclasts are derived from myeloid lineage cells, mainly in response to receptor activator of NF-kB ligand (RANKL) and macrophage colony-stimulating factor [78]. RANKL is expressed by osteoblasts and osteocytes and regulates osteoclast differentiation, bone resorption, and osteoprotegerin (OPG), which is also secreted by osteoblasts. OPG is a negative regulator of osteoclastogenesis by binding with RANKL and impeding receptor interaction, ultimately playing a vital role in bone homeostasis (Fig. 4) [79]. RANKL-RANK generates a downstream signaling cascade of adaptors/kinases, activating transcription factors that induce the expression of osteoclastogenic markers, such as tartrate-resistant acid phosphatase (TRAP) and cathepsin K [78].

Like osteoblasts, the effect of taurine on osteoclasts has not been explored in detail, and the most remarkable effect is its inhibition of osteoclastogenesis. Several studies in the first decade of the millennium supported this notion; however, crucial details were not obtained until 2010. Taurine was found to inhibit osteoclastogenesis of bone marrow cells co-cultured with osteo-

blasts without the influence of OPG and RANKL, whereas TAUT small interfering RNA reversed this effect. This finding indicates that the inhibition of osteoclastogenesis by taurine is dependent on its transporter [76]. Similarly, as ROS are crucial in osteoclast differentiation [80], taurine inhibits LPS-induced osteoclastogenesis in RAW264.7 cells, preventing ROS generation by inhibiting COX-2 expression and JNK phosphorylation [81]. A recent study reported evidence supporting the inhibition of osteoclastogenesis by taurine, indicating that TauCl also impeded RANKL-induced osteoclastic differentiation of RAW 264.7, modifying the mRNA of markers, such as TRAP, cathepsin K, and calcitonin receptor [82,83].

Taurine and osteocytes

Osteocytes are the most abundant cells in the bone and are embedded in a lacuna-canalicular network inside the mineralized matrix, which can sense mechanical forces and induce signals to osteoblasts and osteoclasts to initiate remodeling [84]. As mechanosensitive cells, osteocytes are equipped with integrin complexes, such as \(\beta \) integrin, and ion channels, such as transient receptor potential vanilloid-type 4 (Trpv4), which serve as their main mechanosensory system for physical stimuli. In addition to assisting with the mechanical adaptation of the bone, osteocytes are a significant source of RANKL, controlling osteoclast differentiation in a positive direction and decreasing the expression level of OPG, or vice versa. Stimulatory factors from osteocytes, such as lipids (PGE2), growth factors (e.g., IGF-1), glycoproteins (e.g., Wnt), free radicals (e.g., NO), and nucleotides (e.g., ATP), have substantial effects on osteoblastogenesis and matrix formation [85].

In contrast to osteoblasts and osteoclasts, the relationship between taurine and osteocytes had not been described until 2 years ago [34], and more findings are yet to be obtained. Interestingly, primary osteocytes and IDGSW3 cells can synthesize taurine as a self-protective mechanism, encoding enzymes that convert cysteine into cysteine sulfinate and hypotaurine [34]. In that study, the researchers also found that taurine supplementation protected IDG-SW3 and MLO-Y4 cells from death triggered by H₂O₂ treatment (Fig. 4). Additionally, the gene of the taurine uptake transporter, *SLC6A6*, was highly expressed in IDG-SW3 cells under exogenous taurine supplementation, and sclerostin, a well-known negative regulator of the Wnt signaling pathway, was inhibited by taurine.

Taurine and bone healing

Bone healing is the process of tissue reconstitution in the pres-

ence of fractures or bone defects. The major metabolic phases of fracture healing include inflammation, endochondral ossification, and coupled remodeling [86]. The role of taurine and its derivatives in bone healing has been highlighted. The effects of oral administration of taurine on rabbits with osteotomy in the early stages of bone healing included the formation of a callus rich in osseous components compared with that of control rats, which produced a callus rich in fibrous components [87]. In another model of glucocorticoid-induced rabbit osteonecrosis, taurine administration normalized the expression of mitochondrial transcription factor A and the complex responsible for the synthesis of ATP; thus, mitochondrial function prevented the development of osteonecrosis [88]. Additionally, tauroursodeoxycholic acid, a common bile acid, induces new blood vessel formation and enhances bone formation after implantation, as revealed by a three-dimensional micro-computed tomographic analysis of rabbit calvarial defects [89].

Finally, taurine incorporation into biomaterials to improve the osseointegration of bioactive scaffolds for bone defect regeneration has been explored. The three-dimensional combination of polylactic acid, polycaprolactone, gelatin nanofibers, and taurine was found to be suitable for cell growth and proliferation. Furthermore, bone defects treated with this scaffold resulted in higher new bone formation than that observed in the control group, illustrating the potential use of taurine in biomaterials [90].

TAURINE AND BONE DISORDERS

Taurine and osteoporosis

Osteoporosis is a chronic condition in which bone mass declines. According to the World Health Organization, adults whose bone mineral density (BMD) or BMC values are at least 2.5 standard deviations less than the mean BMD for normal adults have osteoporosis [91]. Clinically, osteoporosis is characterized by back pain caused by vertebral deterioration, fractures, and weight loss. Osteoporosis-related fractures in the USA are associated with an estimated direct cost of \$17 billion per year, with an expected increase to \$25.3 billion by 2025 [92].

The relationship between taurine and osteoporosis has been studied using different approaches; most notably, metabolomic studies have shown low taurine levels in the blood of patients with osteoporosis. Previous studies have shown that postmenopausal Brazilian women with osteopenia or osteoporosis had reduced taurine levels in plasma compared to healthy volunteers [93]. The same results were obtained for Caucasian pre-menopausal women in the USA [94], Japanese women with low estradiol and BMD levels [95] and older Chinese adults with low BMD and fracture [96]. In addition to its serum level, taurine levels in urine are lower in menopausal women than in normal women [97]; thus, low taurine levels in the serum and urine highlight the potential of this amino acid as a biomarker of osteoporosis and bone loss. This result may be related to the high plasma homocysteine (Hcy) levels measured in patients with osteoporosis and low BMD, as recently highlighted elsewhere [98]. Increased Hcy levels result in the dysregulation of the transsulfuration pathway, as L-cysteine is synthesized from Hcy and taurine is synthesized from L-cysteine. In osteoporosis, a decrease in taurine synthesis causes the dysregulation of calcium and vitamin D, which are essential elements for bone growth and resorption [99].

The advantage of restoring plasma taurine levels in osteoporosis models has been analyzed; various treatments, including the administration of bone collagen peptides from Bos grunniens [98] alendronate sodium [99] and natural extracts [100], increase the concentrations of serum taurine and normalizes bone turnover markers and biomechanical parameters. Taurine supplementation alleviates osteoporosis by increasing 1,25(OH)2D3, IGF-1, transforming growth factor (TGF)-β, BMC, and BMD levels in ovariectomized rats [101]. It also increases bone mass and improves resistance to force by promoting, serum osteocalcin, and ALP activity in postmenopausal rats [102].

Taurine and bone tumors

Bone tumors are rare neoplasms that encompass a broad group of benign and malignant entities that affect the bone, cartilage, and connective tissues [103]. Sarcomas of bone present an annual incidence of 0.2% in adults and account for 5% of childhood malignancies; as they are benign tumors, they are asymptomatic and are only detected as incidental findings. The three most frequent bone malignancies are osteosarcoma and Ewing's sarcoma in adolescents, and chondrosarcoma in adults. Although standard treatment includes surgery, chemotherapy, and radiation, surgical resection does not ensure complete removal of the tumor, and recurrence and metastasis can result in a high mortality rate. Therefore, a better understanding of the biology of bone sarcomas is necessary to improve their outcomes, identify and exploit unique characteristics, and propose new treatment targets. Many metabolomic studies have been conducted to search for metabolic biomarkers associated with tumor prediction and diagnosis; however, there are limited data to provide a comprehensive metabolic picture of bone tumors. Nonetheless, taurine has appeared in some of these studies as a possible metabolic marker of bone sarcomas. For example, a serum untargeted metabolomics analysis of 65 patients diagnosed with osteosarcoma revealed higher levels of ascorbic acid, niacinamide, and taurine than in healthy controls [104]. A similar study revealed differences in the synthesis and degradation of ketone bodies and taurine metabolism compared to normal serum [105]. Furthermore, the response of osteosarcoma cells to some treatments highlights taurine as a participant, with the intracellular metabolic profile of MG-63 human OS cells induced by the chemotherapy agent cisplatin and decreased taurine levels, which reflect the activation of defense mechanisms against oxidative stress [106]. Similarly, treatment of Saos-2 osteosarcoma cells with nanographene oxide sheets reduced the levels of several amino acids, such as taurine, which is also related to oxidative stress.

Finally, the action of taurine or its derivatives is important in bone tumor treatment. For example, taurolidine is cytotoxic to osteosarcoma cells and supports the effect of doxorubicin and carboplatin [107]. In addition, taurine chloramine exerts cytotoxic effects against osteosarcoma cell lines (HOS, SAOS-2, and MG-63) by inducing the loss of mitochondrial membrane potential, activating the intrinsic mitochondrial apoptosis pathway [108].

TAURINE AND CARTILAGE

The cartilage is a flexible connective tissue in the skeletal system, which maintains fluid joint motion by coating the surfaces between the bones. Cartilage is a semi-rigid, avascular, and aneural tissue, composed of chondrocytes embedded in an extracellular matrix of collagen fibers, proteoglycans, and other noncollagenous proteins. The proliferation and differentiation of marrow-derived mesenchymal stem cells into chondrocytes also known as chondrogenesis—are complex processes that involve positive regulators, such as FGF, TGF-β/BMP, Wnt/Bcatenin, Hedgehog, Notch, and vascular endothelial growth factor pathways. Furthermore, pro-chondrogenic transcription factors, such as SOX9, RUNX, and trico-rhino phalangeal syndrome type 1 (Trps1), also control chondrogenesis [109].

Taurine and chondrocytes

Taurine participates not only in the maintenance of the bone, but also in cartilage maintenance, and could effectively promote chondrogenic differentiation of human umbilical cord-derived mesenchymal stem cells, enhancing the synthesis of glycosaminoglycans and upregulating the expression of collagen II, aggrecan, and SOX9 chondrocyte mRNA [110]. Similar results were found for human articular chondrocytes treated with taurine, such as the promotion of proliferation, maintenance of the phenotype-enhanced accumulation of glycosaminoglycans and collagens, and increased expression level of SOX9 (Fig. 4) [111]. Similarly, osteoblasts express the TAUT for its uptake under certain conditions; chondrocytes also express this carrier, which is upregulated in hypertonic conditions to protect the cell in hyperosmolarity environments [112].

Taurine and osteoarthritis

Cartilage problems include tears and injuries caused by sports accidents, genetic factors, and various types of arthritis. Within the last group, osteoarthritis (OA) is the most common form, defined as a degenerative cartilage disorder that affects diarthrodial joints. The estimated prevalence of OA is 303.1 69 million worldwide among adults aged ≥65 years, representing the primary cause of disability in the elderly population [113].

Metabolism is considerably altered in OA. Normal chondrocytes utilize glucose via glycolysis, and oxidative phosphorylation maintains mitochondrial function under optimal conditions. Normal energy homeostasis is mainly supported by the AMP activated protein kinase-NAD dependent protein deacetylase sirtuin 1– peroxisome proliferator activated receptor γ co-activator 1α (AMPK–SIRT1–PGC1α) pathway; however, chondrocytes in OA switch from oxidative phosphorylation to glycolysis, impairing mitochondrial metabolism and accumulating ROS, thereby inhibiting AMPK signaling [114]. Furthermore, accumulating evidence shows that other metabolic pathways, such as taurine metabolism, are deregulated in addition to glycolysis. Particularly, metabolomic analysis revealed that in serum samples of OA patients, taurine is significantly lower than that in healthy controls, with a sensitivity and specificity $\geq 80\%$ [115]; the same tendency was observed in the subchondral bone [116] and synovial fluid [117]. Overall, both studies suggested that taurine is relevant to sclerosis. Complementarily, targeted metabolomics also revealed higher levels of taurine, such as phenylalanine and proline, in osteophyte cartilage samples than in control samples, which were related to collagen dissolution or the destruction of boundary layers involved in osteophyte formation [118].

In addition to taurine's participation as a crucial element in OA, various studies have highlighted its beneficial effects as a possible treatment. For example, daily taurine intake induces high SOD activity in the blood of patients with knee OA [119],

whereas taurine tail vein injection relieved OA symptoms in a rat surgery model in a dose- and time-dependent manner, alleviating allodynia and knee swelling inhibition. A histopathological analysis revealed the suppression of matrix loss and cartilage degeneration and the downregulation of matrix metalloproteinase-3 (MMP-3) and CCAAT-enhancer-binding protein homologous protein (CHOP) expression [120]. Finally, *in vitro* studies have revealed that taurine exerted an antioxidant effect by alleviating H₂O₂-induced endoplasmic reticulum stress and subsequently inhibiting patient-derived chondrocyte apoptosis in different grades of OA [121].

Taurine and rheumatoid arthritis

Besides OA, rheumatoid arthritis (RA), another common chronic joint disorder, is an autoimmune inflammatory condition that involves degradation of the extracellular matrix of cartilage. The global estimated prevalence of RA is 0.46% [122]. Although there is no cure for RA, early treatment and supportive treatments can reduce the risk of joint damage and offer better living conditions. In the search for new treatments, several studies have evaluated natural products or existing drugs, and metabolomic approaches have analyzed their effectiveness. Among these, taurine has been identified as a biomarker with the potential to monitor the efficacy of pharmacotherapy. Chinese medicine formulas can regulate taurine and hypotaurine metabolism in the urine samples of the RA rat model [123-125]. Furthermore, the serum metabolic profiles of patients diagnosed with RA showed alterations in taurine levels in response to rituximab [126] and methotrexate treatment [127].

The use of taurine as a therapeutic candidate in RA has been suggested because of its antioxidant and anti-inflammatory properties; this subject was reviewed in detail recently using animal and *in vitro* research [128]. In brief, these studies suggest the benefits of taurine and its derivatives in managing RA in humans. Fig. 4 summarizes the potential roles of taurine in bone and cartilage under normal and pathological conditions.

CONCLUSION

Overall, the present data indicate that taurine plays an important role in bone development and function. We emphasized the ability of taurine to control osteoblastogenesis, osteoclastogenesis, and maintenance of chondrocyte phenotype. Indeed, taurine is emerging as a key player in bone disorders and a promising therapeutic approach for diseases such as osteoporosis and OA. However, more research and clinical studies are needed to elu-

cidate the details of the taurine-bone relationship and the therapeutic efficacy of taurine.

CONFLICTS OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGMENTS

We acknowledge financial support for this work from SIP 20220544.

ORCID

Elizabeth Pérez-Hernández https://orcid.org/0000-0001-5834-3960

Nury Pérez-Hernández https://orcid.org/0000-0001-9806-6267 Ramesh C. Gupta https://orcid.org/0000-0001-8597-619X

REFERENCES

- Lambert IH, Kristensen DM, Holm JB, Mortensen OH. Physiological role of taurine: from organism to organelle. Acta Physiol (Oxf) 2015;213:191-212.
- Ribeiro RA, Bonfleur ML, Batista TM, Borck PC, Carneiro EM. Regulation of glucose and lipid metabolism by the pancreatic and extra-pancreatic actions of taurine. Amino Acids 2018;50:1511-24.
- Yu X, Hu L, Li S, Shen J, Wang D, Xu R, et al. Long noncoding RNA taurine upregulated gene 1 promotes osteosarcoma cell metastasis by mediating HIF-1α via miR-143-5p. Cell Death Dis 2019;10:280.
- 4. Tiedemann F, Gmelin L. Einige neue bestandtheile der galle des ochsen. Ann Phys 1827;85:326-37.
- 5. Demarcay H. Ueber die Natur der Galle. Ann Pharm 1838; 27:270-91.
- Jacobsen JG, Smith LH. Biochemistry and physiology of taurine and taurine derivatives. Physiol Rev 1968;48:424-511
- 7. Kataoka H, Ohnishi N. Occurrence of taurine in plants. Agric Biol Chem 2014;50:1887-8.
- 8. Wishart DS, Feunang YD, Marcu A, Guo AC, Liang K, Vazquez-Fresno R, et al. HMDB 4.0: the human metabolome database for 2018. Nucleic Acids Res 2018;46(D1): D608-17.

- Ma CC, Butler D, Milligan V, Hammann BA, Luo H, Brazdil JF, et al. Continuous process for the production of taurine from monoethanolamine. Ind Eng Chem Res 2020;59: 13007-15.
- Laidlaw SA, Grosvenor M, Kopple JD. The taurine content of common foodstuffs. JPEN J Parenter Enteral Nutr 1990; 14:183-8.
- 11. Chawla D. Taurine and neonatal nutrition. Indian J Pediatr 2018;85:829.
- Tappaz ML. Taurine biosynthetic enzymes and taurine transporter: molecular identification and regulations. Neurochem Res 2004;29:83-96.
- Sarenac TM, Mikov M. Bile acid synthesis: from nature to the chemical modification and synthesis and their applications as drugs and nutrients. Front Pharmacol 2018;9:939.
- Huxtable RJ. Biochemistry of Sulfur. Boston: Springer;
 1986. Chapter 4, Taurine and the oxidative metabolism of cysteine; p. 121-97.
- 15. Anderson CM, Howard A, Walters JR, Ganapathy V, Thwaites DT. Taurine uptake across the human intestinal brush-border membrane is via two transporters: H+-coupled PAT1 (SLC36A1) and Na+- and Cl(-)-dependent TauT (SLC6A6). J Physiol 2009;587(Pt 4):731-44.
- 16. Lourenço R, Camilo ME. Taurine: a conditionally essential amino acid in humans?: an overview in health and disease. Nutr Hosp 2002;17:262-70.
- 17. van Stijn MF, Vermeulen MA, Siroen MP, Wong LN, van den Tol MP, Ligthart-Melis GC, et al. Human taurine metabolism: fluxes and fractional extraction rates of the gut, liver, and kidneys. Metabolism 2012;61:1036-40.
- Seidel U, Huebbe P, Rimbach G. Taurine: a regulator of cellular redox homeostasis and skeletal muscle function. Mol Nutr Food Res 2019;63:e1800569.
- Jong CJ, Azuma J, Schaffer S. Mechanism underlying the antioxidant activity of taurine: prevention of mitochondrial oxidant production. Amino Acids 2012;42:2223-32.
- Shimada K, Jong CJ, Takahashi K, Schaffer SW. Role of ROS production and turnover in the antioxidant activity of taurine. Adv Exp Med Biol 2015;803:581-96.
- 21. Maleki V, Mahdavi R, Hajizadeh-Sharafabad F, Alizadeh M. The effects of taurine supplementation on oxidative stress indices and inflammation biomarkers in patients with type 2 diabetes: a randomized, double-blind, placebo-controlled trial. Diabetol Metab Syndr 2020;12:9.
- 22. Weiss SJ, Klein R, Slivka A, Wei M. Chlorination of taurine by human neutrophils: evidence for hypochlorous acid gen-

- eration. J Clin Invest 1982;70:598-607.
- Kang IS, Kim C. Taurine chloramine administered in vivo increases NRF2-regulated antioxidant enzyme expression in murine peritoneal macrophages. Adv Exp Med Biol 2013;775:259-67.
- Schaffer SW, Shimada K, Jong CJ, Ito T, Azuma J, Takahashi K. Effect of taurine and potential interactions with caffeine on cardiovascular function. Amino Acids 2014;46: 1147-57.
- 25. Bkaily G, Jazzar A, Normand A, Simon Y, Al-Khoury J, Jacques D. Taurine and cardiac disease: state of the art and perspectives. Can J Physiol Pharmacol 2020;98:67-73.
- Ramila KC, Jong CJ, Pastukh V, Ito T, Azuma J, Schaffer SW. Role of protein phosphorylation in excitation-contraction coupling in taurine deficient hearts. Am J Physiol Heart Circ Physiol 2015;308:H232-9.
- 27. Dutka TL, Lamboley CR, Murphy RM, Lamb GD. Acute effects of taurine on sarcoplasmic reticulum Ca2+ accumulation and contractility in human type I and type II skeletal muscle fibers. J Appl Physiol (1985) 2014;117:797-805.
- 28. Terrill JR, Pinniger GJ, Graves JA, Grounds MD, Arthur PG. Increasing taurine intake and taurine synthesis improves skeletal muscle function in the mdx mouse model for Duchenne muscular dystrophy. J Physiol 2016;594:3095-110.
- 29. Wu H, Jin Y, Wei J, Jin H, Sha D, Wu JY. Mode of action of taurine as a neuroprotector. Brain Res 2005;1038:123-31.
- Jong CJ, Ito T, Prentice H, Wu JY, Schaffer SW. Role of mitochondria and endoplasmic reticulum in taurine-deficiency-mediated apoptosis. Nutrients 2017;9:795.
- 31. Roy A, Sil PC. Taurine protects murine hepatocytes against oxidative stress-induced apoptosis by tert-butyl hydroper-oxide via PI3K/Akt and mitochondrial-dependent pathways. Food Chem 2012;131:1086-96.
- 32. Sun G, Wang X, Li T, Qu S, Sun J. Taurine attenuates acrylamide-induced apoptosis via a PI3K/AKT-dependent manner. Hum Exp Toxicol 2018;37:1249-57.
- 33. Li K, Wang D, Zhou X, Shao J, Li Y, Liu X, et al. Taurine protects against arsenic-induced apoptosis via PI3K/Akt pathway in primary cortical neurons. Adv Exp Med Biol 2019;1155:747-54.
- 34. Prideaux M, Kitase Y, Kimble M, O'Connell TM, Bonewald LF. Taurine, an osteocyte metabolite, protects against oxidative stress-induced cell death and decreases inhibitors of the Wnt/β-catenin signaling pathway. Bone 2020;137:115374.
- 35. Sun Q, Jia N, Yang J, Chen G. Nrf2 signaling pathway mediates the antioxidative effects of taurine against corticoste-

- rone-induced cell death in human SK-N-SH cells. Neurochem Res 2018:43:276-86.
- 36. Iezhitsa I, Agarwal R. New solutions for old challenges in glaucoma treatment: is taurine an option to consider? Neural Regen Res 2021;16:967-71.
- 37. Qiu T, Pei P, Yao X, Jiang L, Wei S, Wang Z, et al. Taurine attenuates arsenic-induced pyroptosis and nonalcoholic steatohepatitis by inhibiting the autophagic-inflammasomal pathway. Cell Death Dis 2018;9:946.
- 38. Gentile CL, Nivala AM, Gonzales JC, Pfaffenbach KT, Wang D, Wei Y, et al. Experimental evidence for therapeutic potential of taurine in the treatment of nonalcoholic fatty liver disease. Am J Physiol Regul Integr Comp Physiol 2011; 301:R1710-22.
- Wu G, Yang J, Lv H, Jing W, Zhou J, Feng Y, et al. Taurine prevents ethanol-induced apoptosis mediated by mitochondrial or death receptor pathways in liver cells. Amino Acids 2018;50:863-75.
- 40. Wen C, Li F, Zhang L, Duan Y, Guo Q, Wang W, et al. Taurine is involved in energy metabolism in muscles, adipose tissue, and the liver. Mol Nutr Food Res 2019;63:e1800536.
- 41. Liu Y, Li F, Zhang L, Wu J, Wang Y, Yu H. Taurine alleviates lipopolysaccharide induced liver injury by anti inflammation and antioxidants in rats. Mol Med Rep 2017;16: 6512-7.
- 42. De Luca A, Pierno S, Camerino DC. Taurine: the appeal of a safe amino acid for skeletal muscle disorders. J Transl Med 2015;13:243.
- 43. Kp AD, Martin A. Recent insights into the molecular regulators and mechanisms of taurine to modulate lipid metabolism: a review. Crit Rev Food Sci Nutr 2022 Jan 18 [Epub]. https://doi.org/10.1080/10408398.2022.2026873.
- 44. Midwinter RG, Cheah FC, Moskovitz J, Vissers MC, Winterbourn CC. IkappaB is a sensitive target for oxidation by cell-permeable chloramines: inhibition of NF-kappaB activity by glycine chloramine through methionine oxidation. Biochem J 2006;396:71-8.
- 45. Gomez-Chavez F, Correa D, Navarrete-Meneses P, Cancino-Diaz JC, Cancino-Diaz ME, Rodriguez-Martinez S. NF-κB and its regulators during pregnancy. Front Immunol 2021;12:679106.
- 46. Niu X, Zheng S, Liu H, Li S. Protective effects of taurine against inflammation, apoptosis, and oxidative stress in brain injury. Mol Med Rep 2018;18:4516-22.
- 47. Ahmadi S, Wang S, Nagpal R, Wang B, Jain S, Razazan A, et al. A human-origin probiotic cocktail ameliorates aging-

- related leaky gut and inflammation via modulating the microbiota/taurine/tight junction axis. JCI Insight 2020;5:e132055.
- 48. Kim SH, Yum HW, Kim SH, Kim SJ, Kim K, Kim C, et al. Topically applied taurine chloramine protects against UVB-induced oxidative stress and inflammation in mouse skin. Antioxidants (Basel) 2021;10:867.
- 49. Ramos CO, Campos KK, Costa GP, Cangussu SD, Talvani A, Bezerra FS. Taurine treatment decreases inflammation and oxidative stress in lungs of adult mice exposed to cigarette smoke. Regul Toxicol Pharmacol 2018;98:50-7.
- 50. Abd-Elhakim YM, Ghoneim MH, Ebraheim LL, Imam TS. Taurine and hesperidin rescues carbon tetrachloride-triggered testicular and kidney damage in rats via modulating oxidative stress and inflammation. Life Sci 2020;254:117782.
- 51. Wang G, Ma N, He F, Kawanishi S, Kobayashi H, Oikawa S, et al. Taurine attenuates carcinogenicity in ulcerative colitis-colorectal cancer mouse model. Oxid Med Cell Longev 2020;2020:7935917.
- 52. Wu L, Zhang Q, Li Y, Song W, Chen A, Liu J, et al. Collagen sponge prolongs taurine release for improved wound healing through inflammation inhibition and proliferation stimulation. Ann Transl Med 2021;9:1010.
- 53. Faghfouri AH, Seyyed Shoura SM, Fathollahi P, Shadbad MA, Papi S, Ostadrahimi A, et al. Profiling inflammatory and oxidative stress biomarkers following taurine supplementation: a systematic review and dose-response meta-analysis of controlled trials. Eur J Clin Nutr 2022;76:647-58
- 54. Pasantes-Morales H. Taurine homeostasis and volume control. Adv Neurobiol 2017;16:33-53.
- 55. Shennan DB. Swelling-induced taurine transport: relationship with chloride channels, anion-exchangers and other swelling-activated transport pathways. Cell Physiol Biochem 2008;21:15-28.
- 56. Netti V, Pizzoni A, Perez-Dominguez M, Ford P, Pasantes-Morales H, Ramos-Mandujano G, et al. Release of taurine and glutamate contributes to cell volume regulation in human retinal Muller cells: differences in modulation by calcium. J Neurophysiol 2018;120:973-84.
- Pasantes-Morales H, Quesada O, Moran J. Taurine: an osmolyte in mammalian tissues. Adv Exp Med Biol 1998; 442:209-17.
- 58. Morgan EF, Unnikrisnan GU, Hussein AI. Bone mechanical properties in healthy and diseased states. Annu Rev Biomed Eng 2018;20:119-43.
- 59. Allen MR, Burr DB. Basic and applied bone biology. San

- Diego: Elsevier; 2019. Chapter 1, Bone morphology and organization; p. 3-26.
- 60. Allen MR, Burr DB. Basic and applied bone biology. San Diego: Elsevier; 2019. Chapter 5, Bone growth, modeling, and remodeling; p. 85-100.
- 61. Aghajanian P, Mohan S. The art of building bone: emerging role of chondrocyte-to-osteoblast transdifferentiation in endochondral ossification. Bone Res 2018;6:19.
- 62. Suzuki A, Iwata J. Amino acid metabolism and autophagy in skeletal development and homeostasis. Bone 2021;146: 115881.
- 63. Kim SJ, Lee HW, Gupta RC. Taurine, bone growth and bone development. Curr Nutr Food Sci 2008;4:135-44.
- 64. Roman-Garcia P, Quiros-Gonzalez I, Mottram L, Lieben L, Sharan K, Wangwiwatsin A, et al. Vitamin B12-dependent taurine synthesis regulates growth and bone mass. J Clin Invest 2014;124:2988-3002.
- 65. Shari FH, Dawood H, Hassan JK, Aljazeari QA, Najm MA, Salahuddin A, et al. To study the effect of taurine on the effects of vital bones and regulate the level of glucose in type II diabetes. Int J Res Pharm Sci 2019;10:2545-51.
- 66. Moon PD, Kim MH, Lim HS, Oh HA, Nam SY, Han NR, et al. Taurine, a major amino acid of oyster, enhances linear bone growth in a mouse model of protein malnutrition. Biofactors 2015;41:190-7.
- 67. Choi MJ, Seo JN. Effect of taurine feeding on bone mineral density and bone markers in rats. Adv Exp Med Biol 2013; 776:51-8.
- 68. Modupe OS, Adijat BR, Salau AK. Immune response and bone marker enzyme activities of broiler birds fed graded level taurine-supplemented-diets. Ann Sci Technol 2021;6: 16-25.
- 69. Martiniakova M, Sarocka A, Babosova R, Galbavy D, Kapusta E, Goc Z, et al. Bone microstructure of mice after prolonged taurine treatment. Physiol Res 2019;68:519-23.
- 70. Rutkovskiy A, Stenslokken KO, Vaage IJ. Osteoblast differentiation at a glance. Med Sci Monit Basic Res 2016;22: 95-106.
- 71. Long F. Building strong bones: molecular regulation of the osteoblast lineage. Nat Rev Mol Cell Biol 2011;13:27-38.
- 72. Park M, Choi HK, An JH. Taurine activates BMP-2/Wnt3amediated osteoblast differentiation and mineralization via Akt and MAPK signaling. Iran J Public Health 2019;48: 1960-70.
- 73. Zhou C, Zhang X, Xu L, Wu T, Cui L, Xu D. Taurine promotes human mesenchymal stem cells to differentiate into

- osteoblast through the ERK pathway. Amino Acids 2014; 46:1673-80.
- 74. Lou J, Han D, Yu H, Yu G, Jin M, Kim SJ. Cytoprotective effect of taurine against hydrogen peroxide-induced oxidative stress in UMR-106 cells through the Wnt/β-catenin signaling pathway. Biomol Ther (Seoul) 2018;26:584-90.
- 75. Zhang LY, Zhou YY, Chen F, Wang B, Li J, Deng YW, et al. Taurine inhibits serum deprivation-induced osteoblast apoptosis via the taurine transporter/ERK signaling pathway. Braz J Med Biol Res 2011;44:618-23.
- 76. Yuan LQ, Liu W, Cui RR, Wang D, Meng JC, Xie H, et al. Taurine inhibits osteoclastogenesis through the taurine transporter. Amino Acids 2010;39:89-99.
- 77. Kang YS, Kim SJ. The change of taurine transport in osteocytes by oxidative stress, hypertonicity and calcium channel blockers. Biomol Ther 2008:16:219-25.
- 78. Ono T, Nakashima T. Recent advances in osteoclast biology. Histochem Cell Biol 2018;149:325-41.
- 79. Park-Min KH. Mechanisms involved in normal and pathological osteoclastogenesis. Cell Mol Life Sci 2018;75:2519-28.
- 80. Lee NK, Choi YG, Baik JY, Han SY, Jeong DW, Bae YS, et al. A crucial role for reactive oxygen species in RANKL-induced osteoclast differentiation. Blood 2005;106:852-9.
- 81. Jang HJ, Kim SJ. Taurine exerts anti-osteoclastogenesis activity via inhibiting ROS generation, JNK phosphorylation and COX-2 expression in RAW264.7 cells. J Recept Signal Transduct Res 2013;33:387-91.
- 82. Kang IS, Kim C. Taurine chloramine inhibits osteoclastic differentiation and osteoclast marker expression in RAW 264.7 cells. Adv Exp Med Biol 2019;1155:61-70.
- 83. Kim C, Kang IS. Taurine chloramine, a taurine metabolite from activated neutrophils, inhibits osteoclastogenesis by suppressing NFATc1 expression. Adv Exp Med Biol 2015; 803:99-107.
- 84. Goldring SR. The osteocyte: key player in regulating bone turnover. RMD Open 2015;1(Suppl 1):e000049.
- 85. Robling AG, Bonewald LF. The osteocyte: new insights. Annu Rev Physiol 2020;82:485-506.
- 86. Einhorn TA, Gerstenfeld LC. Fracture healing: mechanisms and interventions. Nat Rev Rheumatol 2015;11:45-54.
- 87. Kaya Bicer E, Yener N, Doganavsargil B, Argin M, Kapubagli A. Does taurine improve fracture healing? An experimental study. J Ist Faculty Med 2018;81:91-8.
- 88. Hirata H, Ueda S, Ichiseki T, Shimasaki M, Ueda Y, Kaneuji A, et al. Taurine inhibits glucocorticoid-induced bone mi-

- tochondrial injury, preventing osteonecrosis in rabbits and cultured osteocytes. Int J Mol Sci 2020;21:6892.
- Yang SS, Oh JM, Chun S, Kim BS, Kim CS, Lee J. Tauroursodeoxycholic acid induces angiogenic activity in endothelial cells and accelerates bone regeneration. Bone 2020;130: 115073.
- 90. Samadian H, Farzamfar S, Vaez A, Ehterami A, Bit A, Alam M, et al. A tailored polylactic acid/polycaprolactone biodegradable and bioactive 3D porous scaffold containing gelatin nanofibers and taurine for bone regeneration. Sci Rep 2020;10:13366.
- 91. Kanis JA. Assessment of fracture risk and its application to screening for postmenopausal osteoporosis: synopsis of a WHO report. WHO Study Group. Osteoporos Int 1994;4:368-81.
- 92. Hershman SH, Razi AE. Vertebral compression fractures in osteoporotic and pathologic bone. Cham: Springer; 2020. Chapter 3, The economic burden of osteoporosis; p. 21-9.
- Pontes TA, Barbosa AD, Silva RD, Melo-Junior MR, Silva RO. Osteopenia-osteoporosis discrimination in postmenopausal women by 1H NMR-based metabonomics. PLoS One 2019;14:e0217348.
- 94. Zhao Q, Shen H, Su KJ, Zhang JG, Tian Q, Zhao LJ, et al. Metabolomic profiles associated with bone mineral density in US Caucasian women. Nutr Metab (Lond) 2018;15:57.
- 95. Miyamoto T, Hirayama A, Sato Y, Koboyashi T, Katsuyama E, Kanagawa H, et al. Metabolomics-based profiles predictive of low bone mass in menopausal women. Bone Rep 2018;9:11-8.
- 96. Cheong SH, Chang KJ. The preventive effect of fermented milk supplement containing tomato (lycopersion esculentum) and taurine on bone loss in ovariectomized rats. Adv Exp Med Biol 2009;643:333-40.
- 97. Yu L, Qi H, An G, Bao J, Ma B, Zhu J, et al. Association between metabolic profiles in urine and bone mineral density of pre- and postmenopausal Chinese women. Menopause 2019;26:94-102.
- 98. Berry TM, Moustafa AA. Osteoporosis and the effect of dysregulation of the transsulfuration pathway via taurine on intracellular calcium homeostasis, vitamin D absorption and vitamin K absorption. Clin Nutr ESPEN 2021;43:191-6.
- 99. Ye M, Zhang C, Jia W, Shen Q, Qin X, Zhang H, et al. Metabolomics strategy reveals the osteogenic mechanism of yak (Bos grunniens) bone collagen peptides on ovariectomy-induced osteoporosis in rats. Food Funct 2020;11:1498-

- 512.
- 100. Chen SY, Yu HT, Kao JP, Yang CC, Chiang SS, Mishchuk DO, et al. An NMR metabolomic study on the effect of alendronate in ovariectomized mice. PLoS One 2014;9: e106559.
- 101. Liu S, Yuan X, Ma C, Zhao J, Xiong Z. 1H-NMR-based urinary metabolomic analysis for the preventive effects of gushudan on glucocorticoid-induced osteoporosis rats. Anal Biochem 2020;610:113992.
- 102. Ahmed HH, Hamza AH. Potential role of arginine, glutamine and taurine in ameliorating osteoporotic biomarkers in ovariectomized rats. Rep Opin 2009;1:24-35.
- 103. Paulos J, Poitout DG. Bone tumors. London: Springer; 2021. Chapter 2, Osteomas; p. 19-20.
- 104. Lv D, Zou Y, Zeng Z, Yao H, Ding S, Bian Y, et al. Comprehensive metabolomic profiling of osteosarcoma based on UHPLC-HRMS. Metabolomics 2020;16:120.
- 105. Lopez-Garrido L, Banuelos-Hernandez AE, Perez-Hernandez E, Tecualt-Gomez R, Quiroz-Williams J, Ariza-Castolo A, et al. Metabolic profiling of serum in patients with cartilage tumours using 1 H-NMR spectroscopy: a pilot study. Magn Reson Chem 2020;58:65-76.
- 106. Duarte IF, Lamego I, Marques J, Marques MP, Blaise BJ, Gil AM. Nuclear magnetic resonance (NMR) study of the effect of cisplatin on the metabolic profile of MG-63 osteosarcoma cells. J Proteome Res 2010;9:5877-86.
- 107. Marley K, Helfand SC, Edris WA, Mata JE, Gitelman AI, Medlock J, et al. The effects of taurolidine alone and in combination with doxorubicin or carboplatin in canine osteosarcoma in vitro. BMC Vet Res 2013;9:15.
- 108. Pilz M, Holinka J, Vavken P, Marian B, Krepler P. Taurine chloramine induces apoptosis in human osteosarcoma cell lines. J Orthop Res 2012;30:2046-51.
- 109. Green JD, Tollemar V, Dougherty M, Yan Z, Yin L, Ye J, et al. Multifaceted signaling regulators of chondrogenesis: implications in cartilage regeneration and tissue engineering. Genes Dis 2015;2:307-27.
- 110. Yao X, Huang H, Li Z, Liu X, Fan W, Wang X, et al. Taurine promotes the cartilaginous differentiation of human umbilical cord-derived mesenchymal stem cells in vitro. Neurochem Res 2017;42:2344-53.
- 111. Liu Q, Lu Z, Wu H, Zheng L. Chondroprotective effects of taurine in primary cultures of human articular chondrocytes. Tohoku J Exp Med 2015;235:201-13.
- 112. Karjalainen HM, Qu C, Leskela SS, Rilla K, Lammi MJ. Chondrocytic cells express the taurine transporter on their

- plasma membrane and regulate its expression under anisotonic conditions. Amino Acids 2015;47:561-70.
- 113. Peat G, Thomas MJ. Osteoarthritis year in review 2020: epidemiology & therapy. Osteoarthritis Cartilage 2021;29:180-
- 114. Mobasheri A, Rayman MP, Gualillo O, Sellam J, van der Kraan P, Fearon U. The role of metabolism in the pathogenesis of osteoarthritis. Nat Rev Rheumatol 2017;13:302-11.
- 115. Chen R, Han S, Liu X, Wang K, Zhou Y, Yang C, et al. Perturbations in amino acids and metabolic pathways in osteoarthritis patients determined by targeted metabolomics analysis. J Chromatogr B Analyt Technol Biomed Life Sci 2018;1085:54-62.
- 116. Yang G, Zhang H, Chen T, Zhu W, Ding S, Xu K, et al. Metabolic analysis of osteoarthritis subchondral bone based on UPLC/Q-TOF-MS. Anal Bioanal Chem 2016;408:4275-86.
- 117. Anderson JR, Chokesuwattanaskul S, Phelan MM, Welting TJM, Lian LY, Peffers MJ, et al. 1H NMR metabolomics identifies underlying inflammatory pathology in osteoarthritis and rheumatoid arthritis synovial joints. J Proteome Res 2018;17:3780-90.
- 118. Xu Z, Chen T, Luo J, Ding S, Gao S, Zhang J. Cartilaginous metabolomic study reveals potential mechanisms of osteophyte formation in osteoarthritis. J Proteome Res 2017;16: 1425-35.
- 119. Saraswati AAEW, Sunardi D, Lubis AMT, Heru F, Mudjihartini N. Taurine intakes increase superoxide dismutase activity in knee osteoarthritis. IOP Conf Ser Earth Environ Sci 2019;217:012054.
- 120. Bian Y, Zhang M, Wang K. Taurine protects against knee osteoarthritis development in experimental rat models. Knee 2018;25:374-80.

- 121. Bian Y, Wang H, Sun S. Taurine alleviates endoplasmic reticulum stress in the chondrocytes from patients with osteoarthritis. Redox Rep 2018;23:118-24.
- 122. Almutairi K, Nossent J, Preen D, Keen H, Inderjeeth C. The global prevalence of rheumatoid arthritis: a meta-analvsis based on a systematic review. Rheumatol Int 2021;41: 863-77.
- 123. Liu Y, Wei M, Yue K, Wang R, Ma Y, Men L, et al. Nontarget metabonomic method provided new insights on the therapeutical mechanism of Gancao Fuzi decoction on rheumatoid arthritis rats. J Chromatogr B Analyt Technol Biomed Life Sci 2019;1105:93-103.
- 124. Cheng B, Zheng H, Wu F, Wu J, Liu X, Tang C, et al. Metabolomics analysis of Danggui Sini decoction on treatment of collagen-induced arthritis in rats. J Chromatogr B Analyt Technol Biomed Life Sci 2017;1061-1062:282-91.
- 125. Yun X, Dong S, Hu Q, Dai Y, Xia Y. 1H NMR-based metabolomics approach to investigate the urine samples of collagen-induced arthritis rats and the intervention of tetrandrine. J Pharm Biomed Anal 2018:154:302-11.
- 126. Sweeney SR, Kavanaugh A, Lodi A, Wang B, Boyle D, Tiziani S, et al. Metabolomic profiling predicts outcome of rituximab therapy in rheumatoid arthritis. RMD Open 2016;2:e000289.
- 127. Wang Z, Chen Z, Yang S, Wang Y, Yu L, Zhang B, et al. (1) H NMR-based metabolomic analysis for identifying serum biomarkers to evaluate methotrexate treatment in patients with early rheumatoid arthritis. Exp Ther Med 2012;4:165-71.
- 128. Malek Mahdavi A, Javadivala Z. A systematic review of preclinical studies on the efficacy of taurine for the treatment of rheumatoid arthritis. Amino Acids 2021;53:783-800.