RESEARCH ARTICLE

Widespread of non-typeable *Haemophilus influenzae* with high genetic diversity after two decades use of Hib vaccine in China

Qiaoli Dong^{1,2} | Wei Shi³ | Xiaoping Cheng⁴ | Changhui Chen⁴ | Qinghong Meng³ | Kaihu Yao³ | Suyun Qian¹

¹Pediatric Intensive Care Unit, Beijing Children's Hospital, Capital Medical University, Beijing, China

²Department of Pediatrics, Affiliated Hospital of Hebei University, Baoding, China

³Key Laboratory of Major Diseases in Children, Beijing Key Laboratory of Pediatric Respiratory Infection Diseases, National Key Discipline of Pediatrics (Capital Medical University), National Clinical Research Center for Respiratory Diseases, National Center for Children's Health, Beijing Pediatric Research Institute, Beijing Children's Hospital, Capital Medical University, Beijing, China

⁴Youyang Hospital, People's Hospital of Chongqing Youyang County, The First Affiliated Hospital of Chongqing Medical University, Chongqing, China

Correspondence

Kaihu Yao, Key Laboratory of Major Diseases in Children, Beijing Key Laboratory of Pediatric Respiratory Infection Diseases, National Key Discipline of Pediatrics (Capital Medical University), National Clinical Research Center for Respiratory Diseases, Beijing Pediatric Research Institute, Beijing Children's Hospital, Capital Medical University, National Center for Children's Health, 56 Nanlishi Road, Xicheng District, Beijing 100045 China. Email: jiuhu2655@sina.com

Suyun Qian, Pediatric Intensive Care Unit, Beijing Children's Hospital, Capital Medical University, 56 Nanlishi Road, Xicheng District, Beijing 100045, China. Email: syqian1211@163.com

Funding information

This study was supported by a national clinical center project from the Ministry of Science and Technology of the People's Republic of China (2013BAI09B11) and the Beijing Talents Fund (2015000021469G209).

Abstract

Background: The aim of this study was to analyze the microbiological characteristics of nasopharyngeal carriage *Haemophilus influenzae* isolates collected from children with respiratory infections in Beijing hospital and Youyang Hospital of China.

Methods: The serotypes of all isolates were determined using latex agglutinated antisera (a-f). The minimum inhibitory concentrations (MICs) of 11 antibiotics were determined using E-test strips. For the beta-lactamase-negative ampicillin-resistant (BLNAR) isolates, *ftsl* gene was sequenced based on fragments amplified by PCR. STs of *H influenzae* isolates were determined by multi-locus sequence typing.

Results: The overall carriage rate of *H* influenzae in the study population was 9.1% (362/3984). One hundred and ninety *H* influenzae isolates which were selected in our study were non-typeable (NTHi) and 44 (23.2%) of them were positive for β -lactamase. All isolates were susceptible to ceftriaxone and levofloxacin. Susceptibility rates to erythromycin and sulfamethoxazole-trimethoprim in Beijing were significantly higher than Youyang (*P* < .05). Thirty-six BLNAR isolates were identified. The MLST analysis showed 108 STs in 190 isolates, the most common of which were ST408 (11, 5.8%), ST914 (10, 5.3%), ST57 (9, 4.7%), and ST834 (6, 3.2%). Twelve STs were detected in both of the study sites, which covered 63 isolates.

Conclusions: All isolates in the present study were NTHi, which suggested widespread of this type in China. The BLNAR isolates were detected more frequently than before. Because high genetic diversity of NTHi isolates of *H influenzae* exists worldwide, it is important to continuously monitor these bacteria in the future.

Abbreviations: BLNAR, beta-lactamase-negative ampicillin-resistant; CC, clonal complex; MIC, minimal inhibitory concentration; MLST, multiple locus sequence types; NTHi, non-typeable Haemophilus influenzae; PCR, polymerase chain reaction.

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KEYWORDS

antimicrobial susceptibility, children, Haemophilus influenzae, respiratory infection

1 | INTRODUCTION

Haemophilus influenzae (*H influenzae*), one of the most common bacterial pathogens causing respiratory tract infections in humans, can also cause bacterial meningitis and sepsis in children.¹ It is usually colonized in the nasopharynx, especially in children, which is the first step leading to a severe infectious diseases. Previous study has shown that the nasopharyngeal carriage rate of *H influenzae* in children under 5 years old with acute upper respiratory tract infections is as high as 26.3%.² It is also the second most common bacterial pathogen causing pneumonia in Chinese children.^{3,4}

In the past several decades, its resistance of *H* influenzae has increased in many countries, but there is a significant difference between geographic sites and monitoring time.⁵ A previous study in Beijing showed that the percentage of β -lactamase positive was 4% in 2000, 13% in 2002, 27% in 2010, and 31% in 2012, and few β -lactamase-negative ampicillin-resistant (BLNAR) isolates were detected.² Variations in penicillin-binding protein 3 (PBP3), which is encoded by the *ftsl* gene, have infrequently been analyzed in *H* influenzae isolates in China.

In the present study, *H influenzae* isolates collected from Chinese children with respiratory infections was monitored in Beijing and discrepancies in the epidemiology between two areas located far apart in China (Beijing City and Youyang County) were identified. The carriage rates, distribution of serotypes, drug resistance, multi-locus sequence types (MLSTs), and PBP3 of *H influenzae* isolates collected from children with respiratory infections in two hospitals were determined and compared.

2 | MATERIALS AND METHODS

2.1 | Study sites

A total of 3984 children in Beijing Children's Hospital affiliated with Capital Medical University [Beijing Children's Hospital] and People's Hospital of Youyang County [Youyang Hospital] were involved in this study. Beijing Children's Hospital is a specialized children's hospital located in northern China and is the largest children's hospital in China. The team at Beijing Children's Hospital has been monitoring the epidemiology of *H influenzae* for about 20 years. This study analyzed the data collected in Beijing Children's Hospital in 2014. Youyang Hospital is a general hospital in southern China. Since 2015, the Department of Pediatrics of Youyang Hospital has been conducting the pathogens investigation involving *H influenzae*. This study analyzed the data collected in Youyang Hospital in 2015.

2.2 | Study population and sample storage

Nasopharyngeal swabs were collected from outpatients with acute respiratory infections evaluated at Beijing Children's Hospital from January to December 2014. The swab samples were immediately placed into transfer tubes with culture medium composed of skimmed milk, glucose, and glycerin (skimmed milk-tryptone glucose-glycerol [STGG], Qingdao Tianqi Biotechnology Co., Ltd.), as previously reported.^{6,7} Samples were sent to the Microbiology Laboratory at Beijing Children's Hospital for culture within 2 hours after collection. Hospitalized patients with acute respiratory infections who were evaluated at Youyang Hospital were enrolled from January to December 2015. Because there is no chocolate culture substrate in Youyang Hospital, the STGG transfer tubes were incubated at 37°C for about 2 hours and then stored in a -60°C freezer immediately. The samples were transferred to Beijing Children's Hospital every 6 months. Finally, 362 *H influenzae* were isolated.

Inclusion criteria were as follows: (a) children of both sexes, aged 3 months to 14 years and (b) children with diagnosis of community-acquired upper or lower respiratory infections. Exclusion criteria were as follows: (a) children had received any immunotherapeutic agents within 2 months before enrollment; (b) children had received antibiotic treatment within 2 months before enrollment and were hypersensitive to the antibiotic prescribed; (c) children had severe underlying disease, such as leukemia; and (d) children had previously been included in the study.

Before enrollment and implementation of any study procedure, the parents and/or legal guardians of each participant signed a written informed consent document. This study was approved by the Ethics Committees of the two hospitals. No ethical issues were encountered in this study.

2.3 | Bacterial culture and identification

After incubated in the culture media at 37°C for 1 hour, 30 μ L of transfer medium was added to culture samples from Beijing Children's Hospital, which was cultured on chocolate medium plates (Guangdong HuanKai Microbial Technology Co., Ltd.). The samples from Youyang Hospital were transferred to Beijing for bacterial culture. The plates were incubated in a 37°C and 5% CO₂ incubator for 1 hour and examined after 24 and 48 hours the same methods as for the samples from Beijing Children's Hospital.

Bacterial identification proceeded according to standard procedures, including colony morphologic features, gram staining, and requirement for both X and V factors (Oxoid, Basingstoke, UK). Detection of β -lactamase activity was determined using the chromogenic cephalosporin nitrocefin (BR66A; Oxoid) method with a known β -lactamase-positive isolate as a control. TABLE 1Carriage rates of Haemophilusinfluenzae isolates in different age groupin two sites

		Study sites				
Age (years)	Total (n = 362)	Beijing Children' Hospital (n = 271)	Youyang Hospital (n = 91)	χ ²	Р	
<1	34 (9.3%)	84 (9.6%)	55 (8.9%)	0.22	.17	
1-2	27 (7.5%)	44 (7.2%)	13 (8.8%)	0.45	.24	
2-3	30 (8.2%)	30 (7.9%)	9 (9.6%)	0.28	.19	
3-4	56 (15.4%)	47 (13.4%)	9 (12.7%)	0.02	.22	
4-5	38 (10.6%)	23 (11.2%)	2 (6.5%)	0.65	.35	
≥5	29 (7.7%)	43 (8.5%)	3 (3.3%)	3.02	.29	
Total	362 (9.1%)	271 (9.2%)	91 (8.6%)	0.36	.16	

2.4 | Molecular identification

All of the isolates were determined not only by regular microbiologic procedures, but also by present molecular identification methods. All *H* influenzae isolates were confirmed by genotype identification of the outer membrane protein P6 and 16S RNA genes, which are specific for *H* influenzae. A polymerase chain reaction (PCR) was performed to identify the genotype by using primers and the procedure as previous described with some modification.^{8,9}

2.5 | Serotyping

Serotypes of all isolates were determined using latex agglutination antisera (a-f; Statens Serum Institut). Non-typeable isolates (NTHis) were confirmed not only by negative reactions of antisera, but also by negative PCR amplification of the *bexA* gene, as described in previous reports.^{10,11}

2.6 | Antimicrobial susceptibility testing

The minimum inhibitory concentrations (MICs) of all isolates for ampicillin, amoxicillin-clavulanic acid, cefuroxime, ceftriaxone, cefepime, meropenem, levofloxacin, erythromycin, tetracycline, sulfamethoxazole-trimethoprim, and chloramphenicol were determined using E-test strips (AB Biodisk). MIC50/MIC90 was defined as the MIC of antibiotics that inhibit the growth of 50%/90% of isolates. *H influenzae* ATCC10211, ATCC49247, and ATCC49766 were used as quality control strains in each test batch. Breakpoints were determined in accordance with the Clinical and Laboratory Standards Institute (CLSI) 2016 criterion.¹²

2.7 | ftsl mutation in BLNAR isolates

The *ftsl* gene was sequenced based on fragments of BLNAR isolates amplified by PCR. The PCR procedure and the sequencing primers

were the same as previously reported.¹³ Sequences were analyzed using Lasergene software (DNASTAR) and compared with the *ftsl* gene sequence of the Rd KW 20 *H influenzae* strain to detect nucleotide substitutions. As previously described, different molecular groups were subsequently classified.^{14,15} According to the amino acid transposition point of pbp3, it can be divided into types I, II, III, and III-like.¹⁶ Group II isolates were typed based on other amino acid substitution patterns in the transpeptidase region of PBP3 (338-573).^{13,16} The other 13 substitutions used for typing in group II have been previously reported.¹⁶ Unreported types were defined as "Ch" plus capital letters to distinguish the unreported types from types reported in previous studies.

2.8 | MLST

The STs of *H influenzae* isolates were determined by multi-locus sequence typing as described in previous reports.^{17,18} Briefly, seven housekeeping genes (*adk*, *atpG*, *frdB*, *fucK*, *mdh*, *pgi*, and *recA*) were amplified from chromosomal DNA, and the products were sent to BGI Company for sequencing on both strands. The STs were determined by comparing the allelic profiles with recognized STs on the MLST website (https://pubmlst.org/hinfluenzae/). New alleles and allelic profiles identified in the present study were submitted to the MLST database for name assignment. eBURST software (version 3; available at http://haemophilus.mlst.net/eburst/) was used to estimate the relationships between the isolates and to assign strains to a clonal complex (CC) using the stringent group definition of six of seven shared alleles.

2.9 | Statistical analysis

The antimicrobial susceptibility data and STs were analyzed with WHONET 5.6 software, as recommended by the World Health Organization. The chi-squared test and Fisher's exact test were used for statistical comparisons and performed with SPSS software (version 16.0; SPSS, Inc). A two-tailed cutoff of P < .05 was considered as statistical significance.

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		Study sites			
			Youvang		
β-lactamase/Antimicrobials	Total (n = 190)	Beijing (n = 99)	(n = 91)	χ^2	Р
β-lactamase-positive [n(%)]	44 (23%)	22 (22.2%)	22 (24.1%)	0.102	.750
<i>TEM</i> [n (%)]	42 (95%)	22 (100%)	20 (90.9%)		
ROB [n (%)]	0 (0%)	0 (0%)	0 (0%)		
Ampicillin				3.845	.050
Susceptible [n (%)]	112 (58.9%)	65 (65.7%)	47 (51.6%)		
Intermediate [n (%)]	25 (13.2%)	10 (10.1%)	15 (16.5%)		
Resistant [n (%)]	53 (27.9%)	24 (24.2%)	29 (31.9%)		
MIC50 (mg/L)	1	0.75	1		
MIC90 (mg/L)	≥256	12	≥256		
MIC range (mg/L)	0.094 to ≥256	0.094 to ≥256	0.125 to ≥256		
Amoxicillin-clavulanic acid				1.640	.200
Susceptible [n (%)]	177 (93.2%)	90 (90.9%)	87 (95.6%)		
Resistant [n (%)]	13 (6.8%)	9 (9.1%)	4 (4.4%)		
MIC50 (mg/L)	1	0.5	1		
MIC90 (mg/L)	3	4	3		
MIC range (mg/L)	0.023 to ≥256	0.023 to ≥256	0.25 to ≥256		
Cefuroxime				0.696	.404
Susceptible [n (%)]	151 (79.5%)	81 (81.8%)	70 (76.9%)		
Intermediate [n (%)]	16 (8.4%)	7 (7.1%)	9 (9.9%)		
Resistant [n (%)]	23 (12.1%)	11 (11.1%)	12 (13.2%)		
MIC50 (mg/L)	1	1	1		
MIC90 (mg/L)	16	16	24		
MIC range (mg/l)	0.064 to ≥256	0.064 to ≥256	0.19 to ≥256		
Ceftriaxone			0.17 10 -200	_	_
Susceptible [n (%)]	190 (100%)	99 (100%)	91 (100%)		
MIC 50 (mg/L)	0.016	0.016	0.016		
MIC90 (mg/L)	0.19	0.19	0.19		
MIC range (mg/L)	0.002-5	0.002-0.5	0.002-0.25		
Levofloxacin	0.002 5	0.002 0.0	0.002 0.20	_	_
Susceptible [n (%)]	190 (100%)	99 (100%)	91 (100%)		
MIC50 (mg/L)	0.016	0.016	0.016		
MIC90 (mg/L)	0 25-0 75	0.25	0.75		
MIC range (mg/l)	0.004-1	0.004-0.5	0.008-1		
Frythromycin			0.0001	13.970	<.001
Susceptible [n (%)]	104 (54,7%)	67 (67,7%)	37 (40,7%)	10070	
Intermediate [n (%)]	_	_	_		
Resistant In (%)]	_	_	_		
MIC50 (mg/L)	4	4	6		
MIC90 (mg/L)	256	64	>256		
MIC range (mg/L)	0.38 to >256	0.38 to >256	2 to >256		
Tetracycline	0.0010 -200	0.0010 =200	210-200	2 885	089
Suscentible [n (%)]	179 (94 2%)	96 (97%)	83 (91.2%)	2.005	.007
Intermediate [n (%)]	1 (2 1%)	2 (2%)	2 (2 2%)		
intermediate [h (%)]	4 (2.1%)	∠ (∠70)	∠ (∠.∠%)		

TABLE 2 (Continued)

		Study sites			
β-lactamase/Antimicrobials	Total (n = 190)	Beijing (n = 99)	Youyang (n = 91)	χ ²	Р
Resistant [n (%)]	7 (3.7%)	1 (1%)	6 (6.6%)		
MIC50 (mg/L)	0.25	0.25	0.38		
MIC90 (mg/L)	0.5	0.38	0.5		
MIC range (mg/L)	0.047-12	0.047-12	0.19-8		
Sulfamethoxazole-trimethoprim				4.322	.038
Susceptible [n (%)]	73 (38.4%)	45 (45.5%)	28 (30.8%)		
Intermediate [(%)]	4.7%	7.1%	2.2%		
Resistant [(%)]	56.8%	47.5%	67%		
MIC50 (mg/L)	≥32	2	≥32		
MIC90 (mg/L)	≥32	≥32	≥32		
MIC range (mg/L)	0.008 to ≥32	0.008 to ≥32	0.047 to ≥32		
Chloramphenicol				2.885	.089
Susceptible [n (%)]	179 (94.2%)	96 (96%)	83 (91.2%)		
Intermediate [n (%)]	1 (0.5%)	0	1 (1.1%)		
Resistant [n (%)]	10 (5.3%)	4 (4%)	6 (6.6%)		
MIC50 (mg/L)	0.5	0.38	0.5		
MIC90 (mg/L)	0.75	0.5	1		
MIC range (mg/L)	0.094 to ≥256	0.094-8	0.25 to ≥256		

3 | RESULTS

3.1 | *H influenzae* strain carriage rates

A total of 362 isolates of *H* influenzae were identified. The proportion of *H* influenzae positive culture in the study population was 9.1% (362/3984). The carriage rate of *H* influenzae by age and study site was shown in Table 1. The results showed that the carriage rates in different age groups ranged from 7.5% to 15.4%, and the highest carriage rate was in "3-4" group. In addition, there was no significant difference in the carriage rates of any age group between the two study sites.

3.2 | Serotype distribution of *H* influenzae isolates

All of the *H* influenzae isolates tested in the present study could not be serotyped with latex agglutination antisera. In fact, no isolates of bexA and type b genes were successfully amplified.

3.3 | β-lactamase determination and antibiotic susceptibility pattern of *H influenzae* isolates

A total of 99 isolates were randomly chosen based on age from Beijing Children's Hospital, and a total of 91 isolates were randomly chosen from Youyang Hospital. All 190 isolates were tested for β -lactamase activity and antibiotic susceptibility patterns. The test results were shown in Table 2. It was showed that among the 190 isolates, 44 (23.2%) were positive for β -lactamase, 42 (22.1%) were positive for TEM, and none were positive for ROB. Although the susceptibility rates of isolates to ampicillin in the two study sites approached statistical significance, the susceptibility rates to other β -lactamase antibiotics were close. All isolates were susceptible to ceftriaxone and levofloxacin. Moreover, susceptibility rates of isolates to erythromycin and sulfamethoxazole-trimethoprim in Beijing Children's Hospital were significantly higher than susceptibility rates in Youyang Hospital (P < .05).

3.4 | Genotypes of BLNAR isolates

None of the TEM-positive isolates were susceptible to ampicillin. The 36 isolates without β -lactamase genes were not susceptible to ampicillin (ie, BLNAR strains), of which 14 (14.1%) were from Beijing Children's Hospital and 22 (24.2%) from Youyang Hospital. Among the 14 isolates of Beijing Children's Hospital, 10 isolates were in group III, and the others were in group II. Among the 22 isolates of Youyang Hospital, three isolates were in group III, four isolates were in group III-like, one isolate was in group I, and the others were in group II (Table 3).

The sequence analysis results revealed that 24 amino acid substitutions at 22 positions between amino acids 338 and 573 in PBP3 were associated with reference to *H* influenzae Rd KW20 (Table 3).

FABLE 3	Categorizatio	n of PBP3 su	bstitutic	in types ac	cording to a	amino acid sı	ubstitution p	oattern in t	he transpep	tidase regic	n of PBP3	(338-573)					
				PBP3 su	bstitutions f	for typing acc	cording to pr	evious refe	rences ¹³ and	d ¹⁶							
Patterns	Groups	Types	No.	D350	S357	M377	S385	1449	G490	A502	R517	N526	A530	T532	V547	N569	•
01	=	ш	1									¥					V I I
02	=	0	2							F		¥			_	s	
03	=	chc	1						ш			¥	S				1
04	=	ChA	9						ш	F		¥			_	S	
05	=	ChD	1					>				¥					
90	=	В	2					>				¥			_	S	
07	III-like		1			_	н				Т			S	_	S	
08	=	ChE	1	z						F		\mathbf{x}					
60	=	D	ю	z					ш			¥	S				
10	=	ChB	2	z	z							\mathbf{x}					
11	_		1	z	z	_					н				_	S	
12	≡		8	z	z	_	н					\mathbf{x}			_	S	
13	≡		1	z	z	_	н					¥	S		_	S	
14	III-like		1	z	z	_	F				Н				_		
15	III-like		1	z	z	_	н				Т				_	S	
16	III-like		1	z	z	_	μ				Т			S	_	S	
17	≡		1	z	z	_	н			μ		¥			_	S	
18	≡		1	z	z	_	⊢			>		¥					
19	≡		1	z	z	_	н		ш			¥	S				
				0	ther PBP3 s	ubstitutions	found only i	n the prese	nt study								
Patterns	Groups	Types	Ż	د ا د	389	G489	T512	A520	G555	Y557	Y558	V56	52 N	1568	A586	A587	
01	=	ш	1														
02	=	0	7			ш											
07	III-like		1	ш				×			Т						
08	=	ChE	1					⊢									
10	=	ChB	2									_					
11	_		1	ш			S			т							
12-1	≡		7	ш								_					
12-2	≡		1	ш											S	S	
13	≡		Ч	ш					ш	т			S				
															9	continues)	

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Previously reported substitutions (A368T, A437S, and A554D) were not detected in present BLNAR isolates. Four types which were previously referred to as B, D, F, and O were detected in present study, and five new types were determined in group II isolates. Finally, 19 patterns were identified in the present research (Table 4). Although amino acid substitutions at the other nine positions between amino acids 338 and 573 in PBP3 were also detected, no additional patterns were revealed. Two additional substitutions at amino acids 586 and 587 were existed in an isolate of pattern 12. In group II, type ChA, identified in six isolates from Youyang Hospital, was the most prevalent type. In group III, pattern 12 was the most common type, and this pattern was found in six isolates from Beijing Children's Hospital and in two isolates from Youyang Hospital. Other patterns found in both Beijing Children's Hospital and Youyang Hospital were pattern 09 (type D) and pattern 10 (type ChB).

3.5 | Multi-locus sequence typing of *H influenzae* isolates

The multi-locus sequence typing analysis revealed that 108 STs, the most common of which were ST408 (11, 5.8%) and ST914 (10, 5.3%), followed by ST57 (9, 4.7%) and ST834 (6, 3.2%). eBURST analysis identified 13 CCs and 68 singletons (Figure 1). Four CCs, each included 10 or more isolates, were as follows: CC1497 (21 isolates), CC259 (15 isolates), CC57 (11 isolates), and CC107 (10 isolates). Each of the following CC/STs contained five isolates: CC11, ST113, ST474, ST486, ST1556, and ST1558. Twelve STs were determined in both of the study sites, which covered 63 isolates.

4 | DISCUSSION

The present study revealed that the overall *H influenzae* carriage rate in children with respiratory tract infections was 9.1%, and the most common age was 3-4 years old, which was similar to a report from Shanghai, China.¹⁹

In our study, none of the isolates could be typed by antisera, which was further confirmed by negative nucleic acid amplification of *bexA* and type b genes. This result was different from a study from Shanghai, China,¹⁹ in which all isolates were identified as type b. In most recent epidemiologic studies, isolates of *H influenzae* type b have been rarely detected. Jiang et al²⁰ reported that Hib (2.7% of *H influenzae* isolates) was rare among Chinese children in Beijing, Shanghai, and Guangzhou. The proportion of Hib in *H influenzae* isolates was also very low in other studies (0%-4.7%).²¹⁻²⁴ Since the typing method was not described in the Shanghai study,¹⁹ which reported 100% of Hib, it was possible that all *H influenzae* isolates were mistaken for type b.

In China, the Hib vaccines have been on the market in the private sector for nearly 20 years, and the coverage rate of vaccines are close to 50% in infants,²⁵ which plays an important role in preventing Hib infection and transmission in China. After nearly 20 years of Hib immunization, NTHis have been identified more and more

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Patterns	Groups	Types	No.	Ampicillin	Amoxicillin/clavulanate	Cefuroxime	Ceftriaxone
01	II	F	1	1.5	2	2	0.064
02	II	0	2	1.5	3	4-8	0.016-0.032
03	II	ChC	1	1.5	1.5	1.5	0.047
04	Ш	ChA	6	1.5-3	2-4	4-12	0.016-0.023
05	II	ChD	1	1.5	1.5	8	0.023
06	Ш	В	2	1.5	2	2-4	0.016
07	III-like		1	16	8	64	0.25
08	Ш	ChE	1	1.5	6	6	0.016
09	Ш	D	3	1.5	2-4	3-6	0.032-0.064
10	Ш	ChB	2	1.5-2	3-12	4-24	0.064-0.125
11	I		1	3	3	1.5	0.25
12-1	III		7	1-6	0.125-48	6->256	0.19-0.5
12-2	III		1	1.5	8	24	0.38
13	III		1	1.5	4	64	0.094
14	III-like		1	12	4	>256	0.25
15	III-like		1	2	2	>256	0.125
16	III-like		1	6	4	>256	0.25
17	III		1	1.5	>256	>256	0.38
18	III		1	0.75	8	32	0.125
19	Ш		1	6	4	>256	0.19

TABLE 4 The MICs (mg/L) distribution of BLNAR isolates by different PBP3 groups and substitution types

Note: The MIC50s of group II type ChB against ampicillin, amoxicillin/clavulanate, cefuroxime, and ceftriaxone were 2, 3, 6, and 0.023 mg/L, respectively.

The MIC50s against ampicillin, amoxicillin/clavulanate, cefuroxime, and ceftriaxone were 2, 6, >256, and 0.25 mg/L, respectively.



FIGURE 1 Population snapshot of 190 *H influenzae* strains revealed by eBURST analysis. The lines indicated the presence of single locus variant links in particular sequence types (STs), which were indicated by circles. The size of the circle corresponded to the number of isolates belonging to the ST. The green ST numbers indicated isolates from Youyang Hospital, the black numbers indicated isolates from Beijing Children's Hospital, and the pink numbers indicates isolates from Youyang Hospital and Beijing Children's Hospital

frequently. Hu^{26} and Hu^{27} reported that NTHis are responsible for 6.17% and 10.08% of pediatric lower respiratory tract infections, respectively. NTHis represent the major proportion of *H* influenzae

isolates cultured in nasopharyngeal specimens from healthy individuals, and the proportion is usually more than 90% except the Hua study (80.7%).²² One study in Italy reported that after 15 years of introduction of Hib vaccine, 97.0% (98/101) of the isolates were NTHis in the oropharyngeal carriage rate of *H influenzae* in young children in two Italian cities.²⁸ A study from Spain showed that with the introduction of the Hib vaccine, NTHis have become more and more common in invasive isolates.²⁹ The NTHis can even cause infectious outbreaks in nursing homes with the spread of some clones. Andersson et al³⁰ reported that an outbreak of the beta-lactam-resistant NTHi and ST14 was associated with severe clinical outcomes in Sweden. A corollary study showed that ST14 clone was associated with increased clinical virulence and resistance to several antimicrobial agents.³¹ ST14 has also been identified in one of the present isolates, which was included the most common CC1497. Therefore, CC1497 should be monitored in the future.

The β -lactamase-positive rate and the antimicrobial resistance pattern of the 190 isolates tested were similar to our previous report.³ BLNAR isolates were previously considered rare in China (0%-1.3%).^{32,33} However, in the current study, 36 isolates of BLNAR were identified from 190 test isolates (18.9%). The BLNAR proportion in Beijing Children's Hospital (14.1%) was clearly higher than that in previous investigation, and the BLNAR proportion in Youyang Hospital (24.2%) was more impressive.³ In Japan, where Hib vaccine had not been introduced in 2006, the BLNAR was determined in more than 50% of the *H influenzae* isolates.³⁴ The increase in BLNAR isolates resulted in more discrepancies between the β -lactamase-positive rate no longer represented the ampicillin-resistant rate. Therefore, the resistant rates of amoxicillin/clavulanate, cefuroxime, and ceftriaxone were increased to varying degrees.

Compared with previous reports, known substitutions of PBP3 amino acid for grouping could be determined in each of the tested BLNAR isolate. Groups I, II, III, and III-like were identified, and group III was the most common. Isolates in group III showed higher maximum MICs against ampicillin, amoxicillin/clavulanate, cefuroxime, and ceftriaxone than that in groups I and II. Characteristic alterations of PBP3 in group III usually cause cephalosporin resistance. The BLNAR isolates in groups III and III-like were all determined as the third-stage L389F substitution (first stage was R517H or N526K substitution, and second stage was S385T substitution).³⁵ Interestingly, one isolate in group I was shown to have L389F and R517H substitutions, without S385T substitution. Most of the isolates in group II in the present study could not be shown to have any reported types based on the amino acid substitutions from 338 to 573 of the transpeptidase region of PBP3. A few isolates in group II showed MICs as high as group III, and some isolates in group III showed low MICs. It suggested that other amino acid substitutions might play a role in the resistance pattern. The variations in amino acid substitution and MIC values showed the complexity of resistance mechanism of BLNAR isolates.

The homology of present isolates was very weak. Among the typed 190 isolates, 108 STs were determined. Although some shared CCs/ STs were identified in both Beijing Children's Hospital and Youyang Hospital, most singletons were found in only one site. High heterogeneity was also found in BLNAR isolates, which was in accordance with the variations in the *ftsl* amino acid substitutions. High heterogeneity of NTHi or BLNAR was also revealed in other cities or countries. Tian et al³⁶ typed 273 isolates of *H* influenzae from pediatric pneumonia patients in Chengdu, China, and 50 different STs (including 39 novel STs) were identified by multi-locus sequence typing. No ST was correlated with isolates from Korea, which is geographically adjacent to China. A study involving 316 invasive NTHi isolates from children in England and Wales during 2003-2010 revealed that a genetically heterogeneous population (155 STs) had different biotypes.³⁷ A report from Italy showed that 98 of 101 were NTHi isolates, and 76 STs were identified among the 98 NTHi strains, of which only four STs (ST12, ST57, ST238, and ST1238) comprised of more than three isolates.²⁸ Among 18 BLNAR NTHis collected in Spain. 15 different genetically diverse STs were identified.²⁹ Unlike high genetic variability of NTHis, encapsulated isolates express widely clonal characteristics.³⁸ The high heterogeneity of NTHis is a block to the development of vaccines and may also be the reason for rare outbreaks of NTHi infections worldwide.

However, there were also some limitations in this study. Firstly, the sample from Youyang Hospital was stored at -60°C for 6 months. We assumed that the bacteria would not change in this process. Secondly, the sample size was too small for two hospitals to represent all hospitals in China. In future study, more hospitals located in different regions of China should be involved. It will be better to culture the bacteria in local laboratories than to do it after samples storage and transport.

5 | CONCLUSION

In conclusion, all isolates in the present study were NTHi, which suggested widespread of this type in China. The BLNAR isolates were detected more frequently than before. Because high genetic diversity of NTHi isolates of *H influenzae* exist worldwide, it is important to continuously monitor these bacteria in the future.

AUTHORS' CONTRIBUTIONS

QD, WS, XC, CC, QM, KY, and SQ designed the study. XC and CC collected the specimens from Youyang. QD, WS, and QM identified the bacteria and performed the antimicrobial susceptibility tests. QD, WS, and KY performed the molecular studies. QD, WS, KY, and SQ collected the data, analyzed them, interpreted the results, and drafted the manuscript. All authors reviewed and revised the manuscript and approved the final version.

ETHICAL APPROVAL

Before enrollment and implementation of any study procedure, the parents and/or legal guardians of each participant signed the written informed consent document. This study was approved by the Ethics Committee of the two hospitals. No ethical problems were encountered in this study.

ORCID

Kaihu Yao (D) https://orcid.org/0000-0002-4747-8920

DATA AVAILABILITY STATEMENT

The datasets analyzed during the present study are available from the corresponding authors Kaihu Yao (email address: jiuhu2655@ sina.com) and Suyun Qian (email address: syqian1211@163.com) on reasonable request.

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2016;35(3):e76-e84.

How to cite this article: Dong Q, Shi W, Cheng X, et al. Widespread of non-typeable *Haemophilus influenzae* with high genetic diversity after two decades use of Hib vaccine in China. *J Clin Lab Anal*. 2020;34:e23145. <u>https://doi.</u> org/10.1002/jcla.23145