Severe Cystic Echinococcosis-Associated Immune Thrombocytopenic Purpura: A Case Report

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Abstract

We present a case of immune thrombocytopenic purpura (ITP), which leads to the diagnosis of severe cystic echinococcosis. Our patient presented with platelets of 5×10^9 /L, and investigations uncovered multiple large echinococcal hepatic cysts, the largest of which measured 19.4 × 15 × 12 cm, and peritoneal implants. While initially refractory to prednisone and immunoglobulins, the ITP responded to dexamethasone. The echinococcosis was treated with albendazole followed by surgical resection of all lesions. Our patient's disease course has evolved favorable since his initial treatment with an isolated episode of recurrent thrombocytopenia 2 years later, and has remained in remission for the past 2 years. While a causal association between echinococcosis and ITP cannot be confirmed, this case is a reminder of the importance of remaining inquisitive for atypical potential triggers of ITP. We also present a review of the limited literature on the association of parasitic infections and ITP.

Keywords: Immune thrombocytopenic purpura; Echinococcus; Hydatid cysts; Parasite; Thrombocytopenia

Introduction

Immune thrombocytopenic purpura (ITP) is an immune-mediated phenomenon of platelet destruction leading to profound thrombocytopenia and consequently a defective primary hemostasis of variable degree. While primary ITP is, by definition, not associated with a predisposing condition, secondary

Manuscript submitted February 2, 2021, accepted February 18, 2021 Published online April 27, 2021

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doi: https://doi.org/10.14740/jh789

ITP is triggered by an underlying disease. Of those, parasitic infections are not established culprits of ITP. We however present and discuss a first case of ITP in the context for severe cystic echinococcosis.

Case Report

Our patient was born in Algeria and immigrated to North America at the age of 40 years. He was diagnosed with mild hydatid disease at the age of 47 years upon travelling back to Algeria, which remained untreated. He was otherwise healthy and took no medication.

At the age of 59 years, he presented to our emergency department with a 3-day history of gradual onset of petechiae and bruising followed by gingival bleed and epistaxis. A complete review of systems was otherwise unremarkable apart from subjective fevers and chills. Physical examination revealed normal vital signs and was significant for abdominal distension, a palpable intra-abdominal mass in the right and left hypochondrium and wet purpura over the buccal mucosa and petechiae over the limbs and trunk. Initial investigation showed a platelet count of 5×10^{9} /L with a normal white blood cell differential and hemoglobin (Table 1). Creatinine and liver function tests were normal. The blood smear confirmed thrombocytopenia with giant platelets and no platelet clumping. White blood and red blood cell morphology was otherwise normal. Hepatitis B showed immunization from previous exposure, while hepatitis C and human immunodeficiency virus (HIV) serologies were negative. A bone marrow aspirate and biopsy were deemed unnecessary. Our patient was diagnosed with ITP and was given intravenous immunoglobulins at a dose of 1 g/kg daily for 2 days and prednisone 100 mg orally twice daily × 7 days. Given the absence of response by day 8, prednisone was replaced by dexamethasone 40 mg orally once a day for 4 days then stopped. On day 10 of treatment, platelet count rose to $36 \times$ 10^{9} /L and had normalized to 213×10^{9} /L by day 20 (Fig. 1).

Bedside ultrasound performed to investigate the abdominal distension revealed a large abdominal cystic mass. A computed tomography (CT) scan showed multiloculated hepatic cysts contained in an inferior left lobe lesion measuring 19.4 \times 15 \times 12 cm extending anteriorly into the abdominal fat and the proximal transverse colon. A smaller 5.8 \times 6.9 cm posterior hepatic lesion was also seen. Imaging findings were consistent with echinococcal hydatid disease (Fig. 2). There was

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White blood cells	$4.8 \times 10^{9}/L$
Hemoglobin	126 g/L
Platelets	$5 \times 10^{9}/L$
Ferritin	109 µg/L
HIV antibody/antigen	0.22 COI (non-reactive)
Hepatitis B surface antibody	901.900 IU/L (reactive)
Hepatitis B core antibody	0.262 COI (reactive)
Hepatitis B surface antigen	0.36 COI (non-reactive)
Hepatitis C antibody	0.08 COI (non-reactive)
Creatinine	66 μmol/L
Lactate dehydrogenase	268 U/L
Haptoglobin	1.73 g/L
Bilirubin	15 μmol/L
Alanine aminotransferase	29 U/L
Alkaline phosphatase	96 U/L
Prothrombin time	12.1 s
Activated partial thromboplastin time	25.3 s
C-reactive protein	13.7 mg/L

HIV: human immunodeficiency virus; COI: cut-off index.

no splenomegaly. The patient was referred to the Departments of Infectious Diseases and General Surgery. Albendazole was started 3 months later. After 1 month of treatment, a CT scan with contrast was performed and showed similar findings with the addition of an anterior peritoneal extrahepatic mass that had enlarged to 1.7×1.4 cm suggesting active echinococcal disease. A few weeks later, given concerns for rupture and anaphylaxis, our patient underwent partial left hepatectomy to remove the anterior cyst, unroofing of the posterior cyst and removal of two hydatid omental implants (Fig. 3). Corticosteroids were not administered perioperatively. Both the large anterior cyst and the smaller omental cysts showed presence of

daughter cysts which suggested viability. Pathology confirmed the diagnosis of echinococcal cysts. Postoperative convalescence was uneventful without signs of sepsis or anaphylaxis. Albendazole was continued for a total of 6 months. Given the high risk of recurrence, our patient underwent close radiological follow-up and there has since been no evidence of echinococcal disease.

Nearly 2 years after initial presentation with ITP, our patient relapsed and presented with a platelet count of 19×10^{9} /L. He responded well to dexamethasone 40 mg orally once daily for 4 days, and has remained in complete remission, without any further treatment with a total follow-up of 4 years since diagnosis (Fig. 1).

Discussion

A new diagnosis of ITP requires careful investigation for possible underlying predisposing conditions, the management of which is paramount to the successful treatment. Common causes of secondary ITP include viral infections, lymphoproliferative disorders, *Helicobacter pylori* and systemic lupus erythematosus among others (Table 2 [1, 2]). This case of ITP leads to the diagnosis of severe cystic echinococcosis, and albeit impossible to ascertain, prompted questioning on whether a causal effect was at play. This report gives an overview of the published evidence available associating parasitic infections and ITP.

The impact on platelet function or number of many parasitic infections has been well documented, and a variety of pathogenic mechanisms have been hypothesized. In particular, the role of platelets in mediating infections from the Plasmodium species responsible for malaria has been extensively studied. Platelets have been shown to contribute to the innate immune response, have a direct cytotoxic effect and mediate cytotoxicity through platelet factor 4 against plasmodium [3]. In fact, animal studies showed that thrombocytopenic mice prior to infection had a worse outcome compared to those with normal platelet count. In humans thrombocytopenia is a poor



Figure 1. Platelet levels since diagnosis. Diagnosis was made at day 0. Albendazole was started at 3 months (blue line). The peak in platelet count at 5 months coincided with surgery (red line) and its associated inflammatory response. There was a period of loss to follow-up between 7 and 21 months after diagnosis. At 21 months, our patient had its first and only relapse.



Figure 2. Coronal images of the computed tomodensitometry scan performed at presentation. (a) Anterior cut demonstrating the large left lobe cystic lesion extending downwards towards the proximal transverse colon. (b) Middle view of the abdomen demonstrating the same lesion. (c) Posterior view of the abdomen demonstrating the posterior liver lesion.

prognostic marker [3]. Thrombocytopenia is a common finding in malaria and was described in 85% of *Plasmodium fal*- *ciparum* and 72% of *Plasmodium vivax* malaria cases [4]. A retrospective study on 28 patients with malaria dating back to



Figure 3. Postoperative partial hepatectomy specimen containing the large inferior left lobe hepatic cyst that was extending anteriorly into the abdominal fat and the proximal transverse colon. (a, b) View of the intact postoperative hepatic cyst and compared to a 15 cm (6 inches) ruler. (c, d) View of the inside of the large cyst following an excision of the cyst wall unveiling a multitude of intact smaller cysts of variable size containing a translucent fluid.

Table 2.	Causes	of	Secondary	Immune	Thrombocytopenic
Purpura					

Viruses and bacteria	
Human immunodeficiency virus	
Hepatitis B virus	
Hepatitis C virus	
Cytomegalovirus	
Varicella-zoster virus	
Epstein-Barr virus	
Helicobacter pylori	
Auto-immune disease	
Systemic lupus erythematosus	
Antiphospholipid syndrome	
Evan's syndrome	
Rheumatoid arthritis	
Inflammatory bowel disease	
Medication	
Pembrolizumab	
Nivolumab	
Alemtuzumab	
Hematological etiology	
Chronic lymphocytic leukemia	
Lymphoproliferative disorders	
Side effect of bone marrow transplantation	
Other	
Vaccines (i.e., MMR)	
Common variable immunodeficiency	
Pregnancy	

MMR: measles-mumps-rubella.

1983 demonstrated that 17 of those had thrombocytopenia, of which 16 had positive platelet-associated immunoglobulin G (PaIgG). Both platelet count and PaIgG levels normalized after malaria treatment. Results from *in vitro* experiments suggest that immune-induced thrombocytopenia may occur as a result of the action of specific immunoglobulins binding to the H-malarial antigen bound to platelets through the Fab terminus [5]. Lacerda et al did in fact describe a case of ITP responsive to steroids in the context of a *Plasmodium vivax* malaria infection which relapsed with recurrence of the infection [6]. Yamaguchi et al also described two cases of thrombocytopenia associated with positive PaIgG and thought to be of immunological etiology. The platelet count and PaIgG levels improved after treatment of the malaria [7].

Cases of ITP in other parasitic infections are limited to case reports or animal studies. A case of babesiosis suspected to have triggered ITP in an otherwise healthy woman was described. Her ITP remained in complete remission following treatment of the parasitemia [8]. This clinical response resembles the favorable clinical course of our patient. A study in mice infected with *Schistosoma mansoni* has demonstrated the appearance of specific antibodies against mouse platelets and even human platelets causing a seemingly immune-dependent thrombocytopenia [9]. In a study on dogs with *Leishmania infantum* and *Ehrlichia canis* infections, platelet-bound antibodies were detected in 80% of co-infected dogs, 50% of leishmaniotic dogs and 60% of ehrlichiotic dogs, despite thrombocytopenia being a rare finding. According to the authors these results support the hypothesis of immune dysregulation of platelet function in these diseases often associated with hemorrhagic mortality [10].

Cystic echinococcosis is a zoonotic parasitic infection of which dogs are the definitive hosts and is caused by larvae of the Echinococcus granulosus tapeworms. Eggs gain access to circulation by hatching and penetrating the bowel wall and develop into metacestodes in affected organs leading to growth of large cysts [11]. While frequently asymptomatic, this disease can lead to organ failure and death. Surgical resection and anthelmintic medication such as albendazole are the mainstay of treatment of advanced disease [12]. Risks of the surgical resection include secondary echinococcosis from spillage of cyst content and anaphylaxis from exposure to high plasma concentration of echinococcal antigen [13]. Echinococcal disease has never been described as a cause of secondary ITP. Recent data have shown that ITP may be associated with complement activation [14], and that complement activating anti-Echinococcus specific antibodies may be found in echinococcosis [15]. There has however been no published case report or in vitro study on the association between thrombocytopenia and echinococcosis in the literature. In the case of our patient, PaIgG was not measured given their lack of sensitivity and specificity, and the diagnosis of ITP was based on the presence of profound thrombocytopenia of otherwise unclear cause and responsiveness to steroids.

In cases such as ours, other causes of thrombocytopenia must be investigated and ruled out. Splenomegaly is a common cause of thrombocytopenia in parasitic infections. Hepatosplenic *Schistosomiasis mansoni* has been shown to cause platelet pooling in the spleen of infected dogs [16]. Babesiosis, a tick-borne zoonotic parasite, has been associated with cytopenia and disseminated intravascular coagulation. Medicationinduced thrombocytopenia must also be recognized, such as in the case of quinine for the treatment of malaria, which may induce immunologic thrombocytopenia or disseminated intravascular coagulopathy for the treatment of malaria [6, 17].

Whether the echinococcal infection in our otherwise healthy patient triggered the episode of ITP remains indeterminate, especially given the paucity of published data on non-malarial parasitic infections and ITP, and the fact the ITP relapsed 2 years later without evidence of recurrent echinococcosis. Nevertheless, this case is a reminder to the importance of remaining inquisitive for atypical potential triggers of ITP.

In conclusion, we present a case of ITP in the context of severe echinococcosis. Thrombocytopenia associated with parasitic infections must be thoroughly investigated, and a broad differential diagnosis should be considered which should include splenomegaly and platelet consumption. In cases where thrombocytopenia is severe and remains unexplained, the diagnosis of ITP could be entertained as a possible consequence from the parasitic infection. Albeit rare, ITP has been described in the literature in the context of some parasitic infections. In general, the pathogenesis of those cases remains elusive and more studies of such cases are warranted for further characterization. However, published data on the association between Plasmodium parasites and ITP has shed some light on some of the possible mechanisms of parasite-associated immune responses against platelets. In many cases of the literature as well as ours, management of both the parasitic infection and the ITP has led to favorable outcome.

Acknowledgments

The authors would like to thank the patient presented in this report for accepting and consenting to the publication of this manuscript.

Financial Disclosure

No funding was received. None of the authors have disclosures relevant to this manuscript.

Conflict of Interest

The authors declare no conflict of interest.

Informed Consent

The manuscript has been sufficiently de-identified to protect the patient. Moreover, written consent was obtained.

Author Contributions

M. Bienz was responsible of the original draft preparation. S. Morin-Ben Abdallah was involved in initial treating and reviewing the manuscript. C. Greenaway, J-S Pelletier and S. Caplan were involved in the management of the case and reviewed the manuscript. H. Knecht is involved in the continued follow-up of the patient and supervised and reviewed the manuscript.

Data Availability

The authors declare that data supporting the findings of this study are available within this article.

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