

# Physiological Role of a Multigrain Diet in Metabolic Regulations of Lipid and Antioxidant Profiles in Hypercholesteremic Rats

## -Multigrain diet in hyperlipemia-

Rupal A. Vasant, Namrata D. Patel, Sanjay S. Karn, Amaravadi V. R. L. Narasimhacharya\*

Laboratory for Animal Sciences, Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, India

### Key Words

enzymatic antioxidants, fecal lipids, lipid metabolism, multigrain, non-enzymatic antioxidants, oxidative stress

### Abstract

**Objectives:** The objective of the present study was to investigate the lipid and the antioxidant regulatory potential of a multigrain diet in laboratory animals with reference to lipid profiles, tissue lipid peroxidation and antioxidant status.

**Methods:** Two types of diets, with or without addition of cholesterol, were used in the study – a commercial diet and a formulated multigrain diet (with *Sorghum vulgare*, *Avena sativa*, *Pennisetum typhoideum*, *Oryza sativa*, *Eleusine coracana* and *Zea mays* grains). After a 10-week period of feeding the diets to albino rats the plasma, liver and fecal lipid profiles and the hepatic and renal antioxidant status of the animals that were fed the commercial and the formulated diets (with and without cholesterol addition) were assessed.

**Results:** The commercial diet supplemented with cholesterol elevated the levels of plasma total lipids, to-

tal cholesterol, triglycerides, low-density lipoprotein cholesterol (LDL-C), and very low-density lipoprotein cholesterol (VLDL-C), as well as the atherogenic index (AI). The high-density lipoprotein cholesterol (HDL-C) content and the antioxidant profiles (total ascorbic acid, superoxide dismutase, catalase, glutathione peroxidase reduced glutathione) declined along with increases in lipid peroxidation. The formulated diet (with and without addition of cholesterol) was found to be more efficient than the commercial diet in controlling plasma, hepatic and fecal lipid profiles, as well as hepatic and renal lipid peroxidation and antioxidant status, than of the hypercholesteremic animals.

**Conclusion:** The multigrain diet used in the present study is effective in countering the hyperlipidemia and oxidative stress caused by high cholesterol intake.

## 1. Introduction

Defects in the cholesterol metabolism leading to hypercholesteremia are very important and recognized risk factors for atherosclerosis and cardiovascular diseases [1-3]. Abnormalities in lipid profiles associated with obesity are characterized by elevated triglycer-

Received: Jan 16, 2014 Accepted: Feb 18, 2014

© This is an Open-Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0/>) which permits unrestricted noncommercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

© This paper meets the requirements of KS X ISO 9706, ISO 9706-1994 and ANSI/NISO Z39.48-1992 (Permanence of Paper).

\*Corresponding Author

Amaravadi V. R. L. Narasimhacharya, Department of Biosciences, Sardar Patel Mайдan, Vadtal Road, Satellite Campus, Sardar Patel University, Vallabh Vidyanagar- 388 120, Gujarat, India.  
Tel: +91-2692-234412 Ext.308 Fax: +91-2692-231041  
E-mail: [narasimhacharya@yahoo.com](mailto:narasimhacharya@yahoo.com)

ides and low-density lipoproteins (LDLs) with abnormal compositions and structures [4]. Epidemiological studies have shown that high-density lipoprotein (HDL)-cholesterol levels could potentially contribute to anti-atherogenesis, including inhibition of LDL oxidation to protect the endothelial cells from the cytotoxic effects of oxidized LDL [5, 6]. Besides medication, concerted efforts to make changes in life styles and dietary regulations are being advised to control the epidemic of cardiovascular ailments affecting the populations across the world.

Cereals and millets are important components of the human diet throughout the world, particularly in tropical and subtropical regions [7]. Cereals constitute a major source of dietary carbohydrates, proteins, vitamins and minerals, especially for the vegetarians worldwide [8]. The present study deals with a formulated diet comprising of millets and cereals – *Pennisetum typhoideum* (Bajra), *Eleusine coracana* (Ragi), *Sorghum vulgare* (Jowar), *Avena sativa* (Oat), *Oryza sativa* (Rice) and *Zea mays* (Maize), all of which are used as foods in many parts of India, especially, in dry and drought prone areas. These are the main sources of energy and contain several nutrients such as protein, calcium, iron, vitamin B-complex and fibers in Indian diets [8]. The present study was undertaken to examine the role of a multigrain diet in controlling the lipid metabolism on the premise that in most cases, consumption of a diet with high fat content leads to obesity, atherosclerosis and cardio-vascular ailments. Because a high lipid profile and hypercholesterolemic conditions exist in clinical situations, the present experiment was designed in order to determine the efficacy of a multi grain diet in controlling the lipid and antioxidant metabolism in presence of cholesterol.

## 2. Materials and Methods

Adult albino female rats (*Charles Foster*) weighing 200 – 280 g were used in the present study. The animals were maintained in polypropylene cages with *ad libitum* access to water in a well-ventilated room in the animal house facility (Department of Biosciences, Sardar Patel University Vallabh Vidyanagar, India) with constant temperature and humidity of  $26 \pm 2^\circ\text{C}$  and 60%, respectively. The care and procedures adopted for the present investigation were in accordance with the regulations of the Institutional Animal Ethics Committee (MoEF/CPCSEA/Reg. 337). After a 10-day adaptation period, 20 animals were randomly segregated into 4 groups of 5 animals each:

Group 1 (CD): Animals of this group were fed a commercial diet.

Group 2 (CDC): Animals of this group were fed a com-

mercial diet with 1.5% cholesterol.

Group 3 (FD): This group received a formulated diet.

Group 4 (FDC): Animals of this group received a formulated diet with 1.5% cholesterol

At the culmination of experiment, animals were fasted overnight and sacrificed under mild ether anesthesia. Blood was collected by retro-orbital puncture into ethylenediaminetetraacetic acid (EDTA)-coated tubes, and plasma was separated by centrifugation. Livers and kidneys were excised, and both plasma and tissues were kept frozen until analyzed. Fecal samples were also collected for biochemical analyses.

Plasma total lipid (TL) content was estimated by using the sulpho-phospho-vanillin method [10]. Plasma total cholesterol (TC), HDL cholesterol (HDL-C) and triglycerides (TG) were measured by using standard kits (Eve's Inn Diagnostics, Vadodara, India). The low-density lipoprotein cholesterol (LDL-C =  $\text{TC} - \text{HDL-C} - \text{TG}/5$ ) and very low density lipoprotein cholesterol (VLDL-C =  $\text{TG}/5$ ) levels and the atherogenic index ( $\text{AI} = \text{TC}/\text{HDL-C}$ ) were calculated [11]. The hepatic TL was extracted in a chloroform and methanol (2 : 1) mixture [12] and was estimated by using a gravimetric analysis. The same extract was used to estimate the TC and the TG contents by using the respective kits (Eve's Inn Diagnostics, Vadodara, India).

Hepatic HMG-CoA (3-hydroxy-3-methyl-glutaryl-CoA) reductase (EC 1.1.1.34) activity was measured in terms of the ratio of HMG-CoA to mevalonate [13]. Colorimetric assays were performed for both HMG-CoA and mevalonate by using hydroxylamine reagent at alkaline pH and acidic pH, respectively, and the ratio of HMG-CoA to mevalonate was determined, which is inversely proportional to the enzyme activity. The alkaline ethanolic extract of liver tissue was acidified, and the total bile acid content was estimated using vanillin-phosphoric acid reagent [14].

The fecal cholesterol was extracted using a chloroform and methanol (2 : 1) mixture [12], and the total cholesterol content was estimated by using standard kits (Eve's Inn Diagnostics, Vadodara, India). From the same extract, TL was estimated by using a gravimetric analysis. The alkaline ethanolic extract of fecal matter cholesterol was acidified and was used with vanillin-phosphoric acid reagent to estimate the total bile acid content [14].

The hepatic and renal lipid peroxidation (malondialdehyde concentration) was determined by using a thiobarbituric acid (TBA) assay [15]. Total ascorbic acid was estimated using 2,4-dinitrophenyl hydrazine reagent [16]. Superoxide dismutase (SOD; EC 1.15.1.1; nitroblue tetrazolium method) and catalase (EC 1.11.1.6; decomposition of  $\text{H}_2\text{O}_2$ ) activities were assayed following the methods of Kakkar *et al.* [17] and Aebi [18], respectively. The assay of glutathione peroxidase (GPx; EC 1.11.1.9) was based on

glutathione (GSH) consumption as described by Flohe and Gunzler [19]. Reduced GSH content was measured by reduction of DTNB, (5,5'-dithio-bis(2-nitrobenzoic acid) as prescribed by Jollow *et al.* [20].

Data are presented as means  $\pm$  SEMs (Standard Error of the Mean). A one-way analysis of variance (ANOVA) with Tukey's significant difference post-hoc test was used to compare differences among groups. Data were statistically handled by Graph Pad Prism Version 3.0 statistical software. *P*-values  $< 0.05$  were considered significant.

### 3. Results

When the animals were fed commercial and formulated diets, the plasma lipid profiles (except HDL-C) in the FD group were significantly lower than those in the CD group. While the HDL-C content in the FD group was significantly elevated, the atherogenic index was substantially lower. With the addition of cholesterol to the diet of the CDC group, significant increases in plasma lipid profiles occurred, but the HDL-C content declined. On the other hand, although the FDC group exhibited elevated TL, TC, TG, and VLDL-C levels, the LDL-C level and the AI did not show any significant increases. Additionally, the HDL-C content in the FDC group was increased by 122%. The differences between the FD and the CD groups and between the FDC and the CDC groups clearly indicate the influence of a formulated (multigrain) diet, with or without cholesterol added, on lipid metabolism (Table 1).

The hepatic and the fecal lipid profiles of the CD, FD, CDC, and FDC groups essentially reflect the trends of the lipid profiles observed in the plasmas of these groups.

When compared to the CD group, the FD group exhibited declined hepatic TL, TC and TG levels with an increase in hepatic and fecal bile contents and fecal TL and TC.

However, lipid synthesis in the hepatic tissues of the CD and the FD groups did not vary significantly. Between the CDC and the FDC groups, the FDC group showed significant improvements in hepatic, as well as fecal, lipid profiles over the CDC group. The hepatic HMG-CoA: mevalonate ratio in the FDC group was found to be 20% lower than that in the CDC, indicating an increase in hepatic HMG-CoA activity (Table 2).

Between the CD and the FD groups, the FD group registered lowered hepatic and renal lipid peroxidation and increased antioxidant profiles, i.e., increases in SOD, catalase (CAT) and GPx activities; the total ascorbic acid (TAA) and glutathione content (in both hepatic and renal tissues) also increased in the FD group. A similar improvement was found in the FDC group, as compared to the CDC group, with reference to lipid peroxidation and to the non-enzymatic and the enzymatic antioxidant parameters (Tables 3 and 4).

### 4. Discussion

High plasma cholesterol represents a major risk factor for ischemic heart diseases. Defects in the cholesterol metabolism are major causes of cardiovascular disorders [21]. Chronic consumption of a diet with high fat content is well known to lead to an increase in plasma cholesterol, resulting in premature atherosclerosis [22]. The incidence of atherosclerosis and cardiovascular disease is related to high levels of plasma lipids, especially low density lipopro-

**Table 1** Effects of a multigrain-formulated diet on the plasma lipid profiles of the experimental animals

Parameters	CD	CDC	FD	FDC	Difference (%) FD vs. CD	Difference (%) FDC vs. CDC
TL*	685.2 $\pm$ 16.3	919.01 $\pm$ 34.85 <sup>†</sup>	671.30 $\pm$ 7.74 <sup>†</sup>	756.90 $\pm$ 10.61 <sup>†‡§</sup>	-2.03	-17.63
TC*	74.51 $\pm$ 1.90	118.30 $\pm$ 3.63 <sup>†</sup>	72.53 $\pm$ 1.60 <sup>†</sup>	87.90 $\pm$ 4.44 <sup>†‡§</sup>	-2.66	-25.70
TG*	71.63 $\pm$ 1.58	130.8 $\pm$ 8.05 <sup>†</sup>	61.34 $\pm$ 3.22 <sup>†</sup>	83.22 $\pm$ 3.87 <sup>†‡§</sup>	-14.36	-36.38
LDL-C*	23.52 $\pm$ 2.51	34.42 $\pm$ 2.75 <sup>†</sup>	21.40 $\pm$ 2.31 <sup>†</sup>	21.29 $\pm$ 1.85 <sup>†‡§</sup>	-9.01	-38.15
VLDL-C*	14.32 $\pm$ 0.37	26.56 $\pm$ 1.84 <sup>†</sup>	11.74 $\pm$ 0.43 <sup>†</sup>	16.20 $\pm$ 0.74 <sup>†‡§</sup>	-18.01	-39.00
HDL-C*	33.85 $\pm$ 0.51	26.37 $\pm$ 1.12 <sup>†</sup>	39.60 $\pm$ 1.08 <sup>†</sup>	58.62 $\pm$ 1.68 <sup>†‡§</sup>	+16.99	+122.30
AI	2.04 $\pm$ 0.09	2.07 $\pm$ 0.03 <sup>†</sup>	1.87 $\pm$ 0.09 <sup>†</sup>	1.77 $\pm$ 0.03 <sup>†‡§</sup>	-8.33	-14.49

Values are mean  $\pm$  SEM (n = 5); *P* < 0.05 was considered as statistically significant; \*mg/dL <sup>†</sup>compared with CD; <sup>‡</sup>compared with FD; <sup>§</sup>compared with CDC; CD, commercial diet; FD, formulated diet; CDC, commercial diet with cholesterol; FDC, formulated diet with cholesterol; TL, total lipid; TC, total cholesterol; TG, triglycerides; HDL-C, HDL cholesterol; VLDL-C, VLDL cholesterol; HDL-C, HDL cholesterol; AI = TC/HDL-C.

tein cholesterol (LDL-C), which is a predisposing factor for cardiovascular disorders [23, 24]. On the other hand, high density lipoprotein cholesterol (HDL-C) has a protective effect [25].

In the present study, we have examined the role of a multigrain diet in the regulation of body lipid and antioxidant metabolism. Since hyperlipidemia is often associated with high fat diet consumption, the animals (albino rats) were administered cholesterol along with two different diets: a commercial (basal) diet and a formulated diet. When the formulated diet alone was administered in the FD group, the formulated diet brought about a significant decline (2% – 18%) in the basal lipid profiles, an increase in HDL-C content (by 17%), and a decline in the atherogenic index (by 8%) when compared to the effects of a commercial diet in the CD group. With the addition of cholesterol to both the diets, the plasma TL, TC, TG and VLDL-C levels in both the CDC and the FDC groups exhibited significant increases compared to the levels in the CD and the FD groups. However, when the plasma lipid profiles of the FDC group were compared with those of the CDC group, the former registered significantly lower TL, TC, TG, LDL-C, and VLDL-C levels and atherogenic index. Further, the HDL-C content in the FDC group was increased by about 122% (Table 1). The hypolipidemic effect of the formulated diet was also seen in both the FD and the FDC groups, where the hepatic TL, TC and TG levels declined appreciably with improvements in fecal TL and TC contents. Concomitant increases in hepatic and fecal bile acid contents in the FDC group also confirmed the lipid regulatory role of the diet. Owing to an increased fecal

excretion of lipids in the group fed the formulated diet, the HMG-CoA: mevalonate ratio in hepatic tissue decreased (Table 2), indicating an increased lipid synthesis, perhaps as a result of a feedback mechanism in the cholesterol biosynthesis. Comparisons between the FD and the CD groups and between the FDC and the CDC groups with regard to hepatic and fecal lipid profiles clearly indicate that while hepatic TL, TC, and TG declined sharply in the FD and the FDC groups, the fecal excretion of TL and TC in these groups significantly increased compared to the CD and the CDC groups (Table 2).

As compared to the commercial diet, the formulated diet appeared to lower the lipid peroxidative processes and elevate the antioxidant status significantly in hepatic and renal tissues: particularly, the enzymatic activities of SOD, CAT, and GPx and the contents of TAA and GSH were increased significantly in the FDC group when compared to the CDC group. When the FD and the FDC groups were compared with their counterparts (the CD and the CDC groups), clear increases in the hepatic and renal enzymatic (7% – 30%) and non-enzymatic (5% – 27%) antioxidant contents were observed in the FD and the FDC groups (Tables 3 and 4). Concomitant increases in hepatic bile acid contents in the FD and the FDC groups (5.33% and 73%) over the CD and the CDC groups, along with elevated fecal bile acid contents in these groups (3.5% – 13%) over the CD and the CDC groups, also indicate the hypolipidemic activity of the formulated diet (Table 2).

The antioxidant status was elevated significantly in the FD and the FDC groups when compared to that in the CD and the CDC groups as can be seen from the differences

**Table 2** Effects of a multigrain-formulated diet on the hepatic and the fecal lipid profiles and on the HMG-CoA reductase activity of the experimental animals

Parameters	CD	CDC	FD	FDC	Difference (%) FD vs. CD	Difference (%) FDC vs. CDC
Hepatic TL*	18.67 ± 2.45	34.67 ± 2.86 <sup>†</sup>	14.25 ± 0.27 <sup>†</sup>	16.00 ± 1.03 <sup>§  </sup>	-23.67	-53.85
Fecal TL*	5.33 ± 1.33	21.33 ± 1.33 <sup>†</sup>	6.62 ± 0.08 <sup>†</sup>	38.67 ± 1.33 <sup>§  </sup>	+24.20	+81.29
Hepatic TC*	21.31 ± 0.79	66.92 ± 1.42 <sup>†</sup>	15.50 ± 0.36 <sup>†</sup>	24.25 ± 0.46 <sup>§  </sup>	-27.26	-63.76
Fecal TC*	53.05 ± 3.15	129.30 ± 6.83 <sup>†</sup>	61.34 ± 0.34 <sup>†</sup>	142.92 ± 5.26 <sup>§  </sup>	+15.63	+10.53
Hepatic TG*	1.70 ± 0.026	3.79 ± 0.29 <sup>†</sup>	1.68 ± 0.014 <sup>†</sup>	2.07 ± 0.22 <sup>§  </sup>	-1.17	-45.38
Hepatic bile acid*	2.44 ± 0.02	2.71 ± 0.25 <sup>†</sup>	2.31 ± 0.01 <sup>†</sup>	4.69 ± 0.07 <sup>§  </sup>	+5.33	+73.06
Fecal bile acid*	6.57 ± 0.89	7.20 ± 0.69 <sup>†</sup>	6.8 ± 0.83 <sup>¶</sup>	8.16 ± 1.00 <sup>§  </sup>	+3.50	+13.33
Hepatic HMG- CoA <sup>†</sup>	5.75 ± 0.47	8.02 ± 0.07 <sup>†</sup>	5.73 ± 0.23 <sup>†</sup>	6.40 ± 0.11 <sup>§  </sup>	-0.35	-20.20

Values are means ± SEMs (n = 5); P < 0.05 was considered as statistically significant; \*mg/g <sup>†</sup>HMG-CoA reductase activity is inversely proportional to the ratio of HMG-CoA to mevalonate; <sup>†</sup>compared with CD; <sup>‡</sup>compared with FD; <sup>§</sup>compared with CDC; CD, commercial diet; FD: formulated diet; CDC, commercial diet with cholesterol; FDC, formulated diet with cholesterol; <sup>¶</sup>not significant; TL, total lipid; TC, total cholesterol; TG, triglycerides; HMG-CoA, 3-hydroxy-3-methyl-glutaryl-CoA.

found between the FD and the CD and between the FDC and the CDC groups. Among the enzymatic activities of the antioxidant enzymes in hepatic and renal tissues, the SOD activity in both the FD and the FDC groups was highest (9.94% and 30% higher than in the CD and the CDC groups) as compared to CAT and GPx activities. While the increases in non-enzymatic antioxidants, TAA and GSH, contents in the hepatic and renal tissues of the FD and the FDC groups were of the order of 2.27% to 11.66% (TAA) and 6.39% to 27.68% (GSH), the lipid peroxidation (formation of thiobarbituric acid reactive substances) in both the liver and kidneys of the FD and the FDC groups was found to be significantly lower (ranging from 6% to 27%) than it was in the CD and the CDC groups (Tables 3 and 4).

The results of the present study clearly demonstrate the physiological role of a multigrain diet in metabolic regu-

lations of lipid and antioxidant profiles in hypercholesteremic rats. The presently observed hypolipidemic and antioxidant effects of a multigrain diet could be attributable to the dietary components and to the phytochemical/nutrient composition of the formulated diet. While the commercial diet contained wheat and soy as the principal components, the formulated diet was made up of *Sorghum vulgare*, *Avena sativa*, *Pennisetum typhoideum*, *Oryza sativa*, *Eleusine coracana* and *Zea mays* grains. Predictably, the phytochemical and nutrient constitutions of the two diets being different, notably the formulated diet was superior, having higher quantities of phytochemicals – polyphenols, flavonoids, saponins, phytosterols, ascorbic acid and fiber than the commercial (basal) diet [26-33]. All these phytochemicals are well-known antihyperlipemic and antioxidant agents and help reduce body cholesterol-

**Table 3** Effects of a multigrain-formulated diet on the hepatic lipid peroxidation and antioxidant status of the experimental animals

Parameters	CD	CDC	FD	FDC	Difference (%) FD vs. CD	Difference (%) FDC vs. CDC
TBARS <sup>1</sup>	187.11 ± 1.69	211.51 ± 0.90*	167.90 ± 1.24*	194.60 ± 0.88 <sup>††</sup>	-10.27	-7.85
SOD <sup>2</sup>	1.61 ± 0.17	1.16 ± 0.12*	1.77 ± 0.03*	1.51 ± 0.02 <sup>††</sup>	+9.94	+30.17
TAA <sup>3</sup>	181.91 ± 1.21	152.70 ± 1.05*	199.11 ± 1.64*	164.4 ± 0.77 <sup>††</sup>	+9.45	+7.66
CAT <sup>4</sup>	62.37 ± 0.93	55.21 ± 0.48*	66.96 ± 0.19*	61.65 ± 0.53 <sup>††</sup>	+7.36	+11.66
GSH <sup>5</sup>	70.69 ± 4.17	47.00 ± 1.00*	75.21 ± 1.15*	60.01 ± 1.12 <sup>††</sup>	+6.39	+27.68
GPx <sup>2</sup>	14.02 ± 0.48	11.82 ± 0.32*	14.73 ± 0.47*	12.86 ± 0.22 <sup>††</sup>	+5.06	+8.80

Values are means ± SEMs (n = 5); P < 0.05 was considered as statistically significant; <sup>1</sup>nM MDA/g; <sup>2</sup>U/mg protein; <sup>3</sup>µg/g; <sup>4</sup>nM H<sub>2</sub>O<sub>2</sub> decomposed/s/g; <sup>5</sup>mg/100 g; \*compared with CD; <sup>†</sup>compared with FD; <sup>††</sup>compared with CDC; CD, commercial diet; FD, formulated diet; CDC, commercial diet with cholesterol; FDC, formulated diet with cholesterol; TBARS, Thiobarbituric acid reactive substances; SOD, superoxide dismutase; TAA, total ascorbic acid; CAT, catalase; GSH, glutathione; GPx, glutathione peroxidase.

**Table 4** Effects of a multigrain-formulated diet on the renal lipid peroxidation and antioxidant status of the experimental animals

Parameters	CD	CDC	FD	FDC	Difference (%) FD vs. CD	Difference (%) FDC vs. CDC
TBARS <sup>1</sup>	180.76 ± 0.98	283.40 ± 3.42*	169.56 ± 0.55*	205.52 ± 0.52 <sup>††</sup>	-6.20	-27.48
SOD <sup>2</sup>	1.38 ± 0.04	1.11 ± 0.92*	1.49 ± 0.01*	1.37 ± 0.017 <sup>††</sup>	+7.97	+23.42
TAA <sup>3</sup>	244.80 ± 2.13	224.80 ± 3.79*	261.41 ± 2.72*	229.90 ± 1.11 <sup>††</sup>	+6.78	+2.27
CAT <sup>4</sup>	27.52 ± 0.76	25.65 ± 1.55*	30.17 ± 0.20*	27.60 ± 0.33 <sup>††</sup>	+9.63	+7.60
GSH <sup>5</sup>	46.22 ± 2.52	35.46 ± 2.20*	50.30 ± 2.20*	42.96 ± 1.05 <sup>††</sup>	+8.83	+21.15
GPx <sup>2</sup>	11.82 ± 0.49	10.32 ± 0.13*	12.09 ± 0.16*	11.67 ± 0.14 <sup>††</sup>	+2.28	+13.08

Values are means ± SEMs (n = 5); P < 0.05 was considered as statistically significant; <sup>1</sup>nM MDA/g; <sup>2</sup>U/mg protein; <sup>3</sup>µg/g; <sup>4</sup>nM H<sub>2</sub>O<sub>2</sub> decomposed/s/g; <sup>5</sup>mg/100 g; \*compared with CD; <sup>†</sup>compared with FD; <sup>††</sup>compared with CDC; CD, commercial diet; FD, formulated diet; CDC, commercial diet with cholesterol; FDC, formulated diet with cholesterol; TBARS, Thiobarbituric acid reactive substances; SOD, superoxide dismutase; TAA, total ascorbic acid; CAT, catalase; GSH, glutathione; GPx, glutathione peroxidase.



ol content through its increased excretion [34-39]. In addition, the formulated diet, owing to the presence of five different grains, is also rich in nutrients, such as carbohydrates, proteins, vitamins and minerals.

## 5. Conclusion

In conclusion, the formulated multigrain diet used in the present investigation is found to be superior to a basal (commercial) diet in controlling both the lipid and the antioxidant profiles of the animals exposed to a cholesterol-containing diet. Therefore, mixed diets (using different grains) could be useful as they can improve and maintain health while obviating the necessity for drug intake over long periods.

## Acknowledgement

This study is supported by a grant from the Department of Biosciences, Sardar Patel University, Vallabh Vidyanagar, India. SSK and RAV are thankful to UGC, New Delhi, India and DST-PURSE Programme (Sardar Patel University) respectively, for financial assistance in the form of Research Fellowships.

## Conflict of interest

The authors declare that there are no conflicts of interest.

## References

1. Descamps OS, Gilbeau JP, Luwaert R, Heller FR. Impact of genetic defects on coronary atherosclerosis in patients suspected of having familial hypercholesterolemia. *Eur J Clin Invest.* 2003;33(1):1-9.
2. Vogel RA, Corretti MC, Gellman J. Cholesterol, cholesterol lowering, and endothelial function. *Prog Cardiovasc Dis.* 1998;41(2):117-36.
3. Gylling H. Cholesterol metabolism and its implications for therapeutic interventions in patients with hypercholesterolaemia. *Int J Clin Pract.* 2004;58(9):859-66.
4. Ruotolo G, Howard BV. Dyslipidemia of the metabolic syndrome. *Curr Cardiol Rep.* 2002;4(6):494-500.
5. Assmann G, Nofer JR. Atheroprotective effects of high-density lipoproteins. *Ann Rev Med.* 2003;54:321-41.
6. Rigotti A, Miettinen HE, Krieger M. The role of the high-density lipoprotein receptor SR-BI in the lipid metabolism of endocrine and other tissues. *Endocr Rev.* 2003;24(3):357-87.
7. Tharanathan RN, Muralikrishna GM, Salimath PV, Raghavendra RMR. Plant carbohydrates-an overview. *Proc Indian Acad Sci (Plant Science).* 1987;97(2):81-155.
8. Gopalan C, Sastri BVR, Balasubramanian SC. In *Nutritive Value of Indian Foods*. Hyderabad (India): National Institute of Nutrition, ICMR Press; 2004. p. 156.
9. Raghuramulu N, Madhavan NK, Kalyanasundaram S, *A Manual of Laboratory Techniques* 2nd ed. Hyderabad (India): National Institute of Nutrition Press; 2003. Section-E, Animal Experimentation; p. 343-68.
10. Frings CS, Fendley TW, Dunn RT, Queen CA. Improved determination of total serum lipids by the sulfo-phospho-vanillin reaction. *Clin Chem.* 1972;18(7):673-4.
11. Friedewald WT, Levy RI, Fredrickson DS. Estimation of the concentration of low-density lipoprotein cholesterol in plasma, without use of the preparative ultracentrifuge. *Clin Chem.* 1972;18(6):499-502.
12. Folch J, Lees M, Sloane Stanley GH. A simple method for the isolation and purification of total lipids from animal tissues. *J Biol Chem.* 1957;226(1):497-509.
13. Rao AV, Ramakrishnan S. Indirect assessment of hydroxymethylglutaryl CoA reductase (NADPH) activity in liver tissue. *Clin Chem.* 1975;21(10):1523-5.
14. Snell FD, Snell CT. *Colorimetric Methods of Analysis* Volume 3. 3rd ed. New York (United States): D Van Nostrand Company; 1953. Chapter 14, Substituted Monobasic Aliphatic acids and their Esters; p. 328-77.
15. Ohkawa H, Ohishi N, Yagi K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem.* 1979;95(2):351-8.
16. Schaffert RR, Kingsley GR. A rapid, simple method for the determination of reduced, dehydro-, and total ascorbic acid in biological material. *J Biol Chem.* 1955;212(1):59-68.
17. Kakkar P, Das B, Viswanathan PN. A modified spectrophotometric assay of superoxide dismutase. *Indian J Biochem Biophys.* 1984;21(2):130-2.
18. Aebi H. Catalase,. In: Bergmeyer HU (Ed.), *Methods of Enzymatic Analysis*, Vol 2. 2nd ed. New York (United States): Verlag Chemie-Academic Press; 1974. p. 673-84.
19. Flohe L, Gunzler WA. Assay of glutathione peroxidase. In: Lester Packer (Ed) *Methods in Enzymology-Oxygen Radicals in Biological Systems*, Vol. 105. New York (United States): Academic Press; 1984. p. 114-21.
20. Jollow DJ, Mitchell JR, Zampaglione N, Gillette JR. Bromobenzene-induced liver necrosis. Protective role of glutathione and evidence for 3,4-bromobenzene oxide as the hepatotoxic metabolite. *Pharmacology.*

- 1974;11(3):151-69.
21. Ikonen E. Mechanisms for cellular cholesterol transport: defects and human disease. *Physiol Rev.* 2006;86(4):1237-61.
  22. Steinberg D. Lipoproteins and the pathogenesis of atherosclerosis. *Circulation.* 1987;76(3):508-14.
  23. Steinberg D. Lipoprotein and atherosclerosis. A look back and look ahead. *Arteriosclerosis.* 1983;3(4):283-301.
  24. Goldstein JL, Brown MS. The low-density lipoprotein pathway and its relation to atherosclerosis. *Annu Rev Biochem.* 1977;46:897-930.
  25. Mattson FH, Grundy SM. Comparison of effects of dietary saturated, monounsaturated, and polyunsaturated fatty acids on plasma lipids and lipoproteins in man. *J Lipid Res.* 1985;26(2):194-202.
  26. Riedl KM, Lee JH, Renita M, St Martin SK, Schwartz SJ, Vodovotz Y. Isoflavone profiles, phenol content, and antioxidant activity of soybean seeds as influenced by cultivar and growing location in Ohio. *J Sci Food Agric.* 2007;87(7):1197-206.
  27. Malencic D, Popovic M, Miladinovic J. Phenolic content and antioxidant properties of soybean (*Glycine max* (L.) Merr.) seeds. *Molecules.* 2007;12(3):576-81.
  28. Yun S, Liang J, Peng X, Yan L, Jinsong B. Total phenolics, flavonoids, antioxidant capacity in rice grain and their relations to grain color, size and weight. *J Cereal Sci.* 2009;49(1):106-11.
  29. Awika JM, Rooney LW. Sorghum phytochemicals and their potential impact on human health. *Phytochemistry.* 2004;65(9):1199-221.
  30. Lim TK. Edible Medicinal and Non-Medicinal Plants: Volume 5, Fruits. Springer; 2013. *Zea mays*; p. 416-47.
  31. Palanisamy BD, Rajendran V, Sathyaseelan S, Nagappa GM, Venkatesan BP. Health benefits of finger millet (*Eleusine coracana* L.) polyphenols and dietary fiber: a review. *J Food Sci Technol.* 2011; <http://dx.doi.org/10.1007/s13197-011-0584-9>.
  32. Nambiar VS, Sareen N, Daniel M, Gallego EB. Flavonoids and phenolic acids from pearl millet (*Pennisetum glaucum*) based foods and their functional implications. *Funct Food Health Dis.* 2012;2(7):251-64.
  33. Peterson DM. Oat antioxidants. *J Cereal Sci.* 2001;33(2):115-29.
  34. Pandey KB, Rizvi SI. Current understanding of dietary polyphenols and their role in health and disease. *Curr Nutr Food Sci.* 2009;5(4):249-63.
  35. Yao LH, Jiang YM, Shi J, Tomás-Barberán FA, Datta N, Singanusong R, *et al.* Flavonoids in food and their health benefits. *Plant Foods Hum Nutr.* 2004;59(3):113-22.
  36. Kritchevsky D, Chen SC. Phytosterols-health benefits and potential concerns: a review. *Nutr Res.* 2005;25(5):413-28.
  37. Francis G, Kerem Z, Makkar HP, Becker K. The biological action of saponins in animal systems: a review. *Br J Nutr.* 2002;88(6):587-605.
  38. Oguntibeju OO. The biochemical, physiological and therapeutic roles of ascorbic acid. *Afr J Biotechnol.* 2008;7(25):4700-5.
  39. Arjmandi BH, Ahn J, Nathani S, Reeves RD. Dietary soluble fiber and cholesterol affect serum cholesterol concentration, hepatic portal venous short-chain fatty acid concentrations and fecal sterol excretion in rats. *J Nutr.* 1992;122(2):246-53.