

PD-1 Immune Checkpoint Inhibitor Therapy Malignant Tumor Based on Monotherapy and Combined Treatment Research

Technology in Cancer Research & Treatment
Volume 20: 1-9
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DOI: 10.1177/15330338211004942
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Abstract

Recently, immunotherapy has become the fourth pillar of cancer treatment in addition to surgery therapy, chemotherapy, and radiation therapy. The inhibitors of programmed cell death protein 1 (PD-1) and its ligand PD-L1 are the new stars in immunotherapy, as they can overcome tumor immunosuppression. However, the efficacy of PD-1 inhibitors still needs to be further developed for clinical treatment. Therefore, research into treatment with anti-PD-1 drugs has emerged as a new development field. This review provides novel insights into the role and mechanism of PD-1 combination anti-tumor therapy, thereby promoting its clinical application in anti-tumor immunotherapy.

Keywords

PD-1, PD-L1, anti-tumor, mechanism, immunotherapy

Received: January 25, 2021; Revised: January 25, 2021; Accepted: February 25, 2021.

Introduction

Programmed cell death protein 1 (PD-1) is an important co-inhibitory signal on the T cells and plays a negative regulatory role in the immune response.¹ PD-1 bound to PD-L1, PD-L2, or the corresponding antibodies can play an immunosuppressive role around the tumor, resulting in the down-regulation of T cells activity and promoting tumor cells evasion of immune surveillance.^{2,3} Unlike traditional treatments that directly kill large areas of tumor tissue, anti-PD-1 immunotherapy blocks this signaling pathway and activates an immune response that inhibits tumor cell growth.⁴ According to anti-PD-1 clinical treatment data, this treatment is more effective for cancer patients whose level of PD-L1/PD-L2 expression is more than 50% on the tumor cell surface.^{5,6} To improve the clinical efficacy of anti-PD-1, researchers have turned their attention to the combination therapy.^{7,8} This article reviews the anti-tumor clinical effect and signaling mechanism of PD-1 and provides a theoretical reference for PD-1 combination as a clinical immunotherapy for anti-tumor.

PD-1 Is Expressed in Tumor Cells

PD-1 is mainly expressed on immune cells such as T cells, B cells, monocytes, and natural killer (NK) cells.⁹ The mature

form of the PD-1 protein contains 268 amino acids (aa) including the cytoplasmic domain (94 aa), hydrophobic transmembrane domain (27 aa), and extracellular domain (147 aa). The PD-1 protein has an immunoreceptor tyrosine-based switch motif (ITSM) and immunoreceptor tyrosine-based inhibitory

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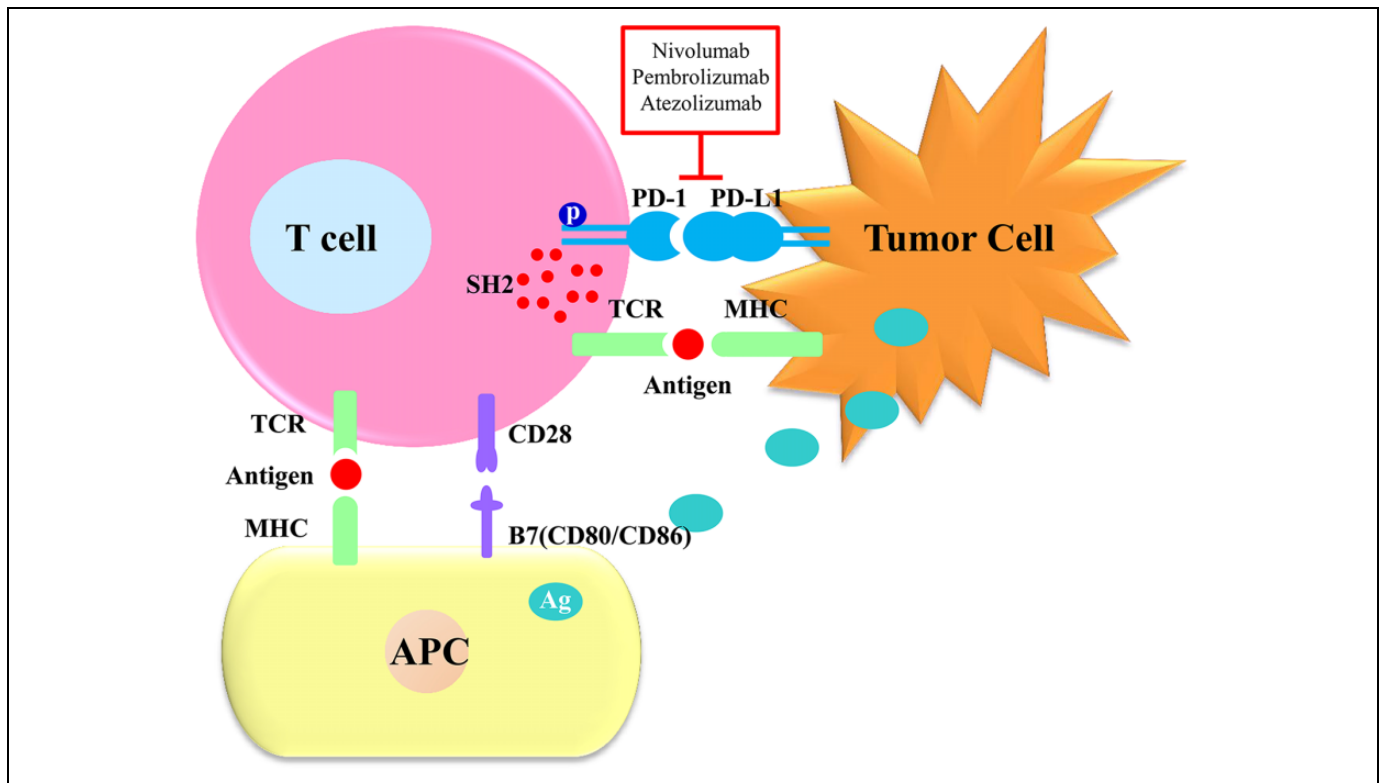


Figure 1. PD-1/PD-L1 signaling pathway. When PD-1 binds to PD-L1 that is expressed on the surface of tumor cells, it induced PD-1 phosphorylation of the intracellular N-terminal ITIM, thereby recruiting SH2 domain-containing protein tryrosine phosphatase-2 (SHP-2) to C-terminal ITSM tyrosine. The TCR of T cells binds to the MHC presented by APC to complete antigen recognition and can secrete antibodies to tumor cells. The above procedure can be reversed by PD-1 inhibitors (Nivolumab, Pembrolizumab and Atezolizumab). In addition, CD28 and ligands (CD80 and CD86) are exposed to the tumor microenvironment, T cells become unreactive or eliminated by programmed cell death.

motif (ITIM) at the end of the cytoplasmic segment tail, which are important structural foundations for PD-1 to exert its immunosuppressive functions. The specific structure of PD-1 corresponds to its unique function.¹⁰

In normal cases, T-cell activation induces PD-1, initiating a feedback suppression mechanism to prevent excessive activation of the T-cell receptor (TCR).¹¹ In cancer patients, PD-1 binds to PD-L1/PD-L2 located on the signaling structure of tumor cells, inducing N-terminal ITIM phosphorylation in the cytoplasm of PD-1 and recruiting activated protein tyrosine phosphatase (SHP-2) turn to the C-terminal ITSM (Figure 1).¹² These activities lead to inhibition of the phosphatidylinositol-3-kinase/AKT/mammalian target of rapamycin (PI3K/AKT/mTOR) signaling pathway and enhancement of caspase activity, thereby activating PD-1-mediated immune cell cycle arrest, reducing cytokine production, and down-regulating immune cell metabolism and many other immunosuppressive responses.^{13,14} PD-1 interaction with PD-L1 can inhibit T cell proliferation and differentiation, resulting in decreased T cell function and even apoptosis, ultimately allowing tumor cells to escape the immune system.¹⁵ Therefore, PD-1/PD-L1 blockade therapies have garnered much attention, as they have proven efficacious for the treatment of many cancers. In a previous study, the PD-1 gene was knocked out in cytotoxic lymphocytes (CTLs) to evaluate its effects on the anti-tumor activity of

CTLs against multiple myeloma (MM). After knockdown, the secretion of cytokine tumor necrosis factor alpha (TNF- α) and interferon gamma (IFN- γ) was significantly increased, enhancing the cytotoxic effects of CTLs on tumor cells and ultimately inducing the apoptosis of tumor cells, further confirming the important role of anti-PD-1 in tumor suppression.¹⁶ Anti-PD-1 (nivolumab and pembrolizumab) or anti-PD-L1 drugs can significantly inhibit the invasion and migration ability of tumor cells, thereby enhancing TCR signal transduction and the function of T cells, which ultimately inhibits the growth of tumor cells. In clinical research, anti-PD-1 drugs are mainly used in solid tumors (e.g., lung cancer, metastatic melanoma), and their efficacy against other cancers is being studied in large-scale clinical trials in patients with cancers such as renal cell carcinoma, bladder cancer, and Hodgkin lymphoma.^{17,18}

Anti-PD-1 Prevent Malignant Tumors

Tumor immunotherapy is a current research hotspot. The successful development and clinical application of PD-1/PD-L1 inhibitors have elevated tumor immunotherapy to a new level.¹⁹ In 1992, the cDNA of PD-1 was discovered by the Japanese scientist Honjo; however, the structure and function of PD-1 were unknown. In 1994, cDNA encoding mouse PD-1

was isolated from apoptosis-cells by subtractive hybridization. The human T cell cDNA library was screened by mouse PD-1 probe, and the cDNA encoding human PD-1 protein was isolated, and then the human PD-1 gene was also mapped to 2q37.3.^{20,21} During the activation and differentiation of T lymphocytes, the B7 molecules interacting with PD-1 were reported in the next 2-5 years. Currently, anti-PD-1 (nivolumab, pembrolizumab) drugs have been used for the treatment of various cancers. In addition, new PD-1 such as Spartalizumab, Cemiplimab and MEDI0680 are also continuously entering clinical trials.

A related mechanism analysis has shown that anti-PD-1 activates dendritic cells (DCs) to present tumor-associated antigens to T cells, simultaneously blocking the release of NF- κ B-dependent cytokines and significantly mitigating tumor growth and prolonging the life of mice. Research has also shown that anti-PD-1 can enhance the killing effect of CIK cells on A520 lung cancer cell line.^{22,23} Anti-PD-1 significantly upregulates NK cell (CD3⁺CD56⁺) activity by cluster of differentiation 16 (CD16) and interleukin 2 (IL-2) and increases the migration of NK cells to MM target cells.^{24,25} In the tumor microenvironment, anti-PD-1 promotes the T-cell anti-tumor immune response during the effector phase. Anti-PD-1 reverses the function of CD8⁺ T cells, promoting intracellular IFN- γ accumulation and reducing IL-10 secretion. Accumulating research suggests that T cells expressing the PD-1 antigen chimeric receptor (ACR) in the signaling domain increases PI3K/AKT activity, increases cytokine secretion, and up-regulates B-cell lymphoma extra-large, which not only promotes T-cell activation but also prolongs the survival time of the T cells. A previous study also found that PD1-ACR can target U87 invasive tumor areas, significantly inhibiting the growth of metastatic tumors, thereby prolonging the survival of U87-bearing mice.²⁶

Nivolumab Prevents Tumor Growth and Metastasis

Nivolumab is an IgG4 monoclonal antibody and the first approved PD-1 inhibitor. It competes with PD-L1 ligand on the tumor surface for PD-1 on T cells, significantly suppressing the PD-1 pathway to produce immunological checkpoint inhibitors.²⁷ Previous studies have shown that nivolumab has a sustained tumor response rate of 20%-30% for a variety of cancers. Due to the effective results of nivolumab in Phase I trials, phase III evaluation was conducted in 3 different cancers (melanoma, renal cell carcinoma, and non-small cell lung cancer), an extraordinary moment in the development of an anti-cancer drug.²⁸ Topalian pointed out that this new drug has broken through the 10%-15% limit of sustained tumor response, which has hampered the development of cancer immunotherapy for the past 30 years. In a meta-analysis of 3404 patients from 20 studies, it was shown that nivolumab may cause a sustained response, with a median post-progression survival of approximately 1 year in patients with advanced non-small cell lung cancer (NSCLC).²⁹ Nivolumab is widely used for the clinical treatment of NSCLC. Yet, its therapeutic effect is unclear for lung cancer caused by interstitial

lung disease (ILD). Kanai counted the medical records of 216 patients with NSCLC who received nivolumab and found 26 with ILD. The effects of nivolumab were measured by response rate (RR), duration of progression-free survival (PFS), and lung toxicity (incidence, severity, and prognosis) in the ILD and non-ILD groups. The results showed that the overall incidence of NSCLC in the ILD group was higher than that in the non-ILD group after nivolumab treatment. Furthermore, more than 50% of patients with NSCLC in both groups had symptoms that improved over time.³⁰

Pembrolizumab Prevents Tumor Growth and Metastasis

Pembrolizumab is a humanized anti-PD-1 antibody that has been extensively studied in a variety of malignancies. It recognizes PD-1 by intermolecular direct and water-mediated hydrogen bonds (WHBs), non-conventional hydrogen bonds (nHBs), hydrophobic contacts, and salt bridges (SBs).³¹ Pembrolizumab has a similar anti-tumor mode of action of nivolumab. In 2016, pembrolizumab was approved by the U.S Food and Drug Administration for untreated patients with metastatic NSCLC or those treated with platinum-based chemotherapy. Pembrolizumab induces an overall response rate (ORR) of 21%-34% in melanoma, and the root cause of the low survival rate is the difficulty of target therapy. In stage III/IV unresectable melanoma, pembrolizumab is superior to ipilimumab. Likewise, pemizumab induces an ORR of 19%-25% in NSCLC. Based on the above results, pembrolizumab has been approved for the treatment of advanced melanoma and NSCLC. Preliminary data have shown that the ORR of pembrolizumab is approximately 20%-50% in malignant tumors including lymphoma and other solid tumors.³² Pembrolizumab has been found to have durable responses in patients with advanced NSCLC through years of clinical research. These findings have changed the current therapeutic mode in advanced NSCLC, adding a new treatment option for patients.³³ In recent years, pembrolizumab has been approved for use in a growing number of cancer types. Frank examined the efficacy of pembrolizumab in the treatment of advanced melanoma and its associated clinical outcomes for more than 4 years in the United States. Pembrolizumab was given to 315 (59%), 152 (29%), and 65 (12%) patients. Overall, the 1- and 2-year survival rates of pembrolizumab were 61% and 48%, respectively; lactate dehydrogenase levels returned to normal (relative to elevation), and overall survival (OS) was significantly improved. These findings demonstrate the effectiveness of pembrolizumab in the actual clinical setting.³⁴ Another study found that pembrolizumab was used for intravenous infusion (escalating doses 2 or 10 mg/kg) until disease progression or severe toxicity. During the study, 80% of patients had nausea and fever, and it elevated aspartate aminotransferase/alanine aminotransferase. Collectively, the therapeutic safety profile of pembrolizumab in Japanese patients is similar to those previously reported for Caucasian patients.³⁵

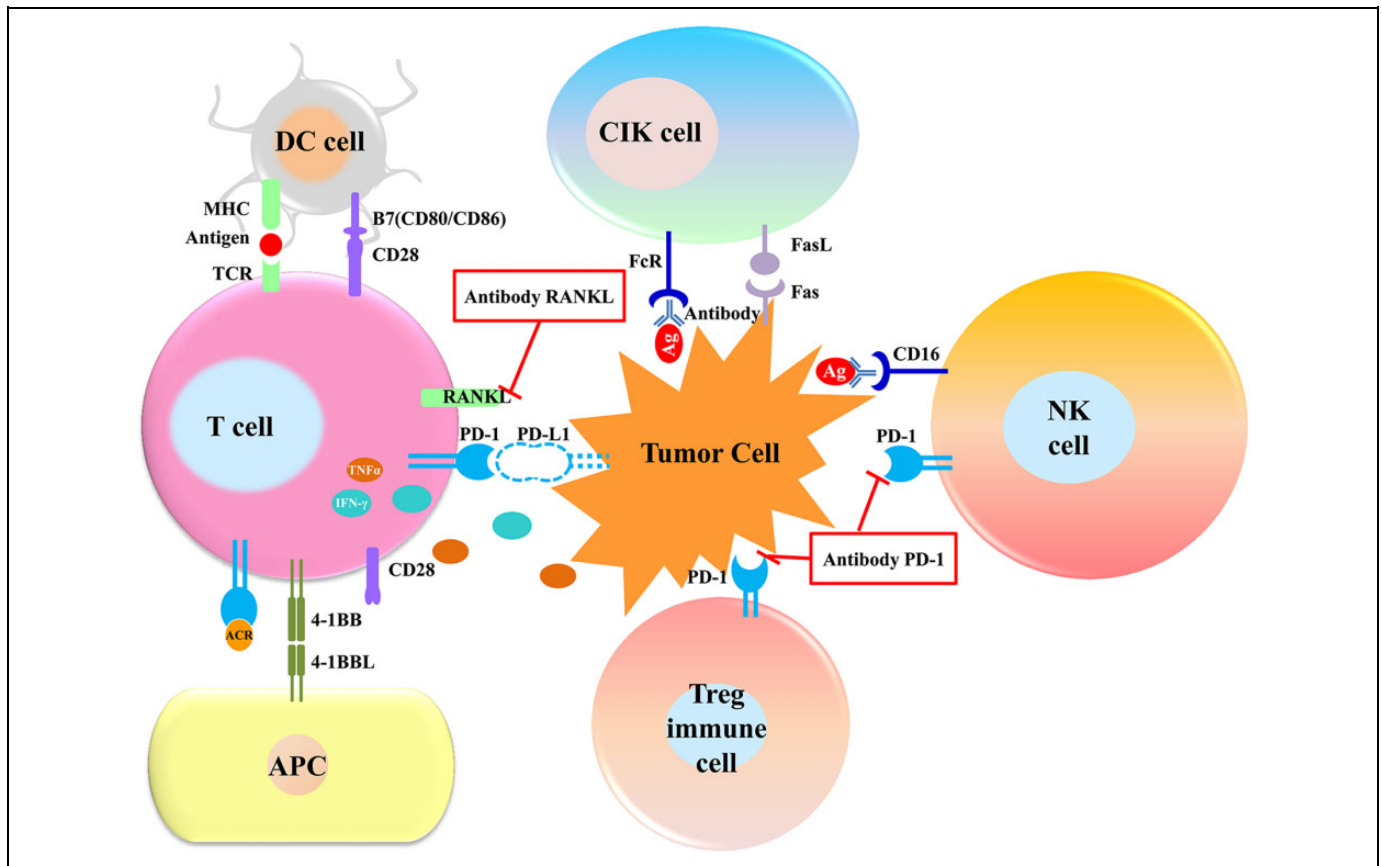


Figure 2. Anti-PD-1 combined with other cells inhibiting tumor growth and metastasis. The addition of anti-PD1 to TILs increased RANKL expression, and anti-RANKL directly reduced PD-L1 expression on the non-lymphoid component tumor microenvironment. Therefore, the addition of RANKL blocker can improve the efficacy of PD1, CTLA4 and RANKL inhibitors, and increase the proportion of IFN γ and TNF α in tumor-infiltrating CD4 $^{+}$ and CD8 $^{+}$ T cells. In addition, CD28 on T cells binds to ligands on DC cells to regulate tumor cells. T cells were activation of co-stimulatory molecules by 4-1BB and 4-1BBL enhances the survival of primary T cell responses and memory T cells. Besides, anti-PD-1 significantly inhibits tumor growth by activating DCs to present tumor-associated antigens to primary T cells. When CIK is activated by binding of the antigen-antibody complex to the FeR, toxic particles are excreted to the cells, and binding of FasL and Fas, the tumor cells undergo apoptosis. In addition to the release of toxic particles and Fas/FasL, CD16 expressed on the surface of NK cells binds to antibodies and kills tumor cells.

Analysis of Anti-PD-1 Combination Therapy to Inhibit Tumor Growth and Metastasis

Anti-PD-1 has anti-tumor effects, but its effective anti-cancer rate is not ideal. How to increasing the survival of patients is still a hot issue in current research. A great amount of data have shown PD-1 and B and T lymphocyte attenuator (CD272) simultaneously inhibit the signal at the interface of major effector T cells and T-cell antigen presenting cells, which provides a theoretical basis for the combination of blocking antibodies in cancer immunotherapy.^{36,37} Researchers have found through numerous studies that anti-PD-1 combination with CTLA4, lymphocyte activating gene 3 and T cell immunoglobulin and mucin domain-containing protein 3 antibodies significantly increase IFN- γ and TNF α .^{38,39} Besides, combined treatment restores activation and proliferation of CD4 $^{+}$ and CD8 $^{+}$ T cells, significantly enhancing the function of effector T cells and enhancing the tumor's immune inhibition response. At the same time, combined treatment prevents autoimmune diseases

by reducing regulatory T cell activity and upregulating IFN regulatory factor. In summary, anti-PD-1 combined treatment can inhibit the growth and metastasis of mouse ovarian cancer cells, H22 liver cancer cells, and MM cells, significantly improving the OS rate of mice, and has an effective therapeutic effect.⁴⁰⁻⁴² In addition, (S)-(-)-n-[2-(3-hydroxy-1H-indol-3-yl)-methyl]-acetamide (SNA), a specific inhibitor of PI3K δ , play an inhibitory on myeloid-derived suppressor cells (MDSCs), then trigger changes in the tumor microenvironment and enhance the activation and infiltration of T-cells. The combination of SNA with anti-PD1 can activate CD8 $^{+}$ T cell-selective infiltration by PI3K δ/γ , having good therapeutic effects.⁴³ Recently, irreversible electroporation combined with anti-PD-1 was found to activate CD8 $^{+}$ T cells by DCs to improve the tumor microenvironment. It also promotes micro-vascular production that in turn promotes the infiltration of T cells of tumor-associated antigens, significantly inhibiting tumor growth, but has no effect on normal tissues, prolonging the lifespan of orthotopic pancreatic tumor mouse models (Figure 2).^{44,45} The effects of the 2 PD-1

inhibitors will be described in detail. Reem et al⁴⁶ found that the co-blockade of PD-1 and PD-L1 further up-regulates the co-expression of TIM-3 and LAG-3 on CD4⁺CD25⁺ T cells and CD4⁺CD25⁺FoxP3⁺Helios⁺Tregs in the presence of TNBC cells, but not in non-TNBC cells. The results indicate that the emergence of compensatory inhibition mechanisms is most likely mediated by Tregs and activated non-Tregs. Our results indicate the emergence of compensatory inhibitory mechanisms, most likely mediated by Tregs and activated non-Tregs, which could lead to the development of TNBC resistance against PD-1/PD-L1 blockade.

At present, the sensitivity of chimeric antigen receptor-T (CAR-T) cell therapy is high, but damage caused by off-target effects can lead to serious side effects. Recent clinical results have shown that CAR-T therapy combined with anti-PD-1 is frequently used for solid tumor treatment.⁴⁷ However, it remains unknown whether it can truly treat cancer; more clinical data are needed for confirmation. Nevertheless, combination therapy is showing promise in treating cancer and producing durable responses.

Nivolumab Combined Cohort Analysis to Inhibit Tumor Growth and Metastasis

Data over the past several decades have shown that nivolumab-based immunotherapies have clinical efficacy, leading to approval of the combination of nivolumab and ipilimumab in treating patients. One study showed 64 patients treated with nivolumab, among which 26 patients discontinued the drug due to disease progression or AEs. The 26 patients were treated with nivolumab in combination with platinum doublets, monotherapy, with a disease response rate of 34.6% (9 patients) and 57.7% (15 patients) and disease control rate of 73.1% (19 patients) and 19.2% (5 patients). A multivariate regression analysis showed that nivolumab combined with platinum doublets is more effective than monotherapy.⁴⁸ In a trial of patients with late-phase melanoma, we randomly assigned patients who were not previously treated in a 1:1:1 ratio: ①Nivolumab+Ipilimumab group; ②Nivolumab+placebo group; ③Ipilimumab+placebo group. In subsequent follow-up visits, the overall 3-year survival rate of nivolumab combination with ipilimumab was 58%, in the nivolumab group was 52%, and in the ipilimumab group was 34%. In summary, the OS of nivolumab and ipilimumab was significantly longer compared to ipilimumab or nivolumab alone.⁴⁹

Pembrolizumab Combined Cohort Analysis to Inhibit Tumor Growth and Metastasis

With the widespread use of anti-PD-1 therapy in melanoma patients, problems have followed, of which brain metastasis is becoming more common.⁵⁰ From January 2014 to December 2015, Erik evaluated the safety of 21 patients receiving pembrolizumab combined with stereotactic radiosurgery (11 cases), hyperfractionated radiation (7 cases), and whole brain therapy (3 cases). In the initial response, all treatments were well

tolerated, and no grade 4 or 5 toxicity was observed. For brain metastases treated with combination therapy, 70% (16/23) showed complete remission (CR, n = 8) or partial remission (PR, n = 8) at the first scheduled follow-up (median 57 days post-treatment). The survey demonstrated that pembrolizumab combination with radiation therapy was safe for patients with metastatic melanoma, especially at the first visit, effectively reducing the size of brain metastases. In general, these results are superior to cases of individual treatment.⁵¹ In a previous study, EDP1503 was able to activate a variety of systemic immune pathways; therefore, it was hypothesized that EDP1503 could be combined with immunological checkpoint inhibitors to achieve anti-cancer goals. Recently, the monoclonal microbial candidate for drug EDP1503 combined with pembrolizumab has been officially enrolled in the I/II clinical trial, and tumor patients who have recurrent rectal cancer and triple-negative breast cancer have been formally enrolled. This clinical trial will assess the safety, tolerability, and overall response rate of the immune response in 120 patients after combination therapy. Besides, initial clinical data are expected to be tested in mid-2020.⁵² Dr. Humphrey Gardner affirmed the study and said that this clinical trial will explore the potential synergy between EDP1503 and pembrolizumab and provide the potential to treat multiple cancer types.

More PD-1 Inhibitors Cohort Analysis to Prevent Tumor Growth and Metastasis

PD-1 inhibitors are still under continuous development in anti-tumor. In addition to Nivolumab and pembrolizumab, more PD-1 inhibitors are gradually being discovered. Two other PD-1 inhibitors are introduced below. Cemiplimab (Libtayo) is the third PD-1 monoclonal antibody approved for marketing in the United States, and the first and only drug specifically approved for skin squamous cell carcinoma (CSCC). At present, clinical studies of cemiplimab cover a variety of solid tumors, including non-small cell lung cancer (NSCLC), glioblastoma, prostate cancer, ovarian cancer, cervical cancer, and thyroid cancer. The results of the phase III of the PD-1 monoclonal antibody cemiplimab (Libtayo), the first-line treatment of NSCLC, were announced at the ESMO meeting in 2020.^{53,54} It is strong evidence that PD-1 monoclonal antibody has taken the first-line treatment of NSCLC. In the PD-L1 $\geq 50\%$ intention-to-treat (ITT) population, the median follow-up time is about 10 months. The median overall survival (OS) data of cemiplimab reached (17.9 months-not estimable), and the median OS data of the chemotherapy group was 14.2 months (11.2-17.5 months). The median progression-free survival (PFS) of the cemiplimab group was 8.2 months (6.1-8.8 months), and the chemotherapy group was 5.7 months (4.5-6.2 months). The objective response rate (ORR) of cemiplimab group was 39.2%, and the chemotherapy group was 20.4%. The median remission of the cemiplimab group was 16.7 months (12.5-22.8 months), and the chemotherapy group was 6.0 months (4.3-6.5 months). Cemiplimab reduces the risk of disease progression or death by 43%. The above data show that cemiplimab has

obvious advantages in the first-line treatment of NSCLC patients with PD-L1 $\geq$ 50%. CT-011, a new type of anti-PD-1 antibody, seems to be specific cytotoxicity to NK cells trafficking, immune complex formation, and cytotoxicity for PD-L1⁺ multiple myeloma (MM) tumor cells rather than normal cells. Jacalyn et al⁵⁵ show that lenalidomide down-regulates PD-L1 on primary MM cells and may enhance NK cell function through CT-011. CT-011 combined with lenalidomide should be considered for phase II clinical trials of MM patients. Studies have shown that 17 patients have received increasing doses of CT-011 (0.2-6 mg/kg). Blood samples were taken before treatment, immediately after treatment, 24 hours, 48 hours, 7th day, 14th day and 21st day. Within 21 days after CT-011 treatment, the percentage of CD4⁺ in peripheral blood was observed to continue to increase. Pharmacokinetic analysis showed that the serum C_{max} and AUC of CT-011 increased proportionally with the increase of dose. The results showed that the CT-011 is safe and well tolerated in this patient population and 33% clinical benefit was observed in patients.⁵⁶

Anti-PD-Ls Prevent Malignant Tumors

A prospective study observed that PD-Ls (PD-L1/PD-L2) can be overexpressed in tumor cells such as ovarian cancer, meningioma, and melanoma, as well as can directly promote tumor growth.⁵⁷ Another study showed that the overexpression of PD-Ls in tumor cells is closely related to the recurrence and metastasis of tumor cells, and the expression level in cancer tissues is significantly higher than that in normal tissues.⁵⁸ In addition, it has been reported that there is no significant correlation between PD-L1 and PD-1 expression in cancer and lymph node tissues. However, IL-2 and IL-10 are involved in the regulation of PD-L1 expression and induction of tumorigenesis with lymph node metastasis, which may be one of the factors affecting prognosis.⁵⁹ Therefore, it suggests that common anti-PD-L1 can improve the survival rate and remission rate in cancer patients, which has a positive impact. Until now, anti-PD-L1 (atezolizumab, durvalumab, and avelumab) has been approved for the treatment of urothelial carcinoma, and several other drugs are still in early clinical trials stage. The discovery of anti-PD-L1 provides a new solution for cancer therapy. In the same period, it was shown that blocking PD-L1 can enhance the expansion of TILs and its function, and reverse the tumor immune CD8⁺ T cells in the tumor microenvironment.^{60,61} Studies have also proposed the role of immune cell infiltration and immune cell function in predicting the efficacy of PD-1/PD-L1 blockade therapy. Based on these mechanisms, researchers have proposed combined treatment strategies, and the importance of patient-specific treatment plans to prolong the life of patients.⁶² To summarize, anti-PD-L1 can block the binding of PD-1 and PD-L1, up-regulate the growth and proliferation of T cells, enhance the recognition of tumor cells by T cells, which activate its attack and killing functions, and mobilize the body's own immune function to achieve resistance tumor effect.

Discussion

At present, immunological checkpoint inhibitors (ICPis) are mainly used in the treatment of anti-tumor immune response. After many years of research, PD-1/PD-Ls are an important immune-regulator factor. PD-1/PD-Ls inhibitors show target tumors with effects in blocking tumor growth and proliferation. Moreover, as the tumor evolves, the immune system of the body can be improved. Due to individual reasons and different types of cancer, 15%-40% of cancer patients show clinical responses. Other patients cannot benefit from anti-PD1/PD-L1 treatment. Some patients with clinical response will develop acquired resistance after the initial response. Therefore, understanding the mechanism of drug resistance is a necessary condition for improving the efficacy of anti-PD1/PD-L1. At present, researchers have identified the main drug resistance mechanisms for tumor, including insufficient tumor immunogenicity caused by loss of tumor antigens, cell and molecular suppression signals in the tumor microenvironment (TME), MHC dysfunction, and irreversible T cell failure, the immunosuppressive microenvironment, all of which may impair the durability of treatment. In this regard, researchers described a potential strategy for the combination of targeted Treg and ICPis to overcome this resistance and maximize the efficacy of treatment for cancer patients.⁶³⁻⁶⁵

Blocking PD-1/PD-L1 signal or co-processing other cooperative stimulation signals has shown amazing effects in inhibiting tumor growth. Based on the search of PD-1/PD-L1 inhibitors, the combination blocking forms are diverse, such as PD-1 inhibitors with immunosuppressive factors can strengthening prevent tumor growth and metastasis, and it with other killer cells can also enhance the anti-tumor effects achieved. The objective response of ICPis is different in each type of cancers. In addition, ICPis is usually accompanied with the risk of immune-related adverse events (irAEs) such as dermatitis and enterocolitis, if the treatment is not adjusted in time, it may be life-threatening. This indicates that patients need to be monitored in real time. Therefore, developing reliable biomarkers to predict patients with irAEs remains a challenge. But the structure of the human body is complicated. After the drug enters the body, it is affected by various factors such as bacterial flora and virus in the body. Even though PD-1 mono or combination options are increasingly used, response rate and duration of response are still where we to understand. Moreover, the regulatory factors of combined treatment and its downstream signaling pathways need to be further identified. Based on this, combined with relevant clinical practice theory, the PD-1 combination therapy is studied in depth from animal (such as the construction of nude mouse tumor model and zebrafish model), cells (the construction of various cell experimental models), and molecular level, is needed to promote the development of PD-1 as a target molecule for immunotherapy tumors and lay a new cornerstone to improve the quality of life of cancer patients.

Authors' Note

Cheng-Hao Jin and Ying-Hua Luo conceived and designed the review. Yu Zhang and Guang-Ze Mou drafted the manuscript. Tian-Zhu Li and Wan-Ting Xu drew figures of the review. Tong Zhang, Hui Xue, Wen-Bo Zuo, and Yan-Nan Li revise it critically for important intellectual content. All authors have read and approved the final version of the manuscript. Yu Zhang and Guang-Ze Mou contributed equally to this work. The authors declared no animal and human studies of the research.

Declaration of Conflicting Interests

The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The author(s) disclosed receipt of the following financial support for the research and/or authorship of this article: This work was supported by the central government supports local college reform and development fund talent training projects, the Heilongjiang Touyan Innovation Team Program (2019HTY078), the Project for Heilongjiang Bayi Agricultural University (XDB202012), the Heilongjiang Farms & Land Reclamation Administration Support Project for Key Scientific Research (HKKYZD190705), the Heilongjiang Bayi Agricultural University Support Program for "San Zong" (TDJH201905), Heilongjiang Province College Student Innovation and Entrepreneurship Training Program Project (202010223001), and Heilongjiang Province College Student Innovation and Entrepreneurship Training Program Project (202010223004).

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References

1. Elena G, Domenico VD, Alessandra F. Recent insights into the role of the PD-1/PD-L1 pathway in immunological tolerance and autoimmunity. *Autoimmun Rev*. 2013;12(11):1091-1100. doi:10.1016/j.autrev.2013.05.003
2. Sponaa A, Yang R, Rustad EH, et al. PD1 is expressed on exhausted T cells as well as virus specific memory CD8⁺ T cells in the bone marrow of myeloma patients. *Oncotarget*. 2018;9:32024-32035. doi:10.18632/oncotarget.25882
3. Okazaki T, Honjo T. PD-1 and PD-1 ligands: from discovery to clinical application. *Int Immunol*. 2007;19(7):813-824. doi:10.1093/intimm/dxm057
4. Zi JYX, Zhang MZ, Li JY, et al. PD-1/PD-L1 blockade: have we found the key to unleash the antitumor immune response? *Front Immunol*. 2017;8:1597-1614. doi:10.3389/fimmu.2017.01597
5. Bai J, Gao ZT, Li X, et al. Regulation of PD-1/PD-L1 pathway and resistance to PD-1/PDL1 blockade. *Oncotarget*. 2017;8(66):110693-110707. doi:10.18632/oncotarget.22690
6. Timothy JD, Arie B. Words of wisdom. re: safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *Eur Urol*. 2015;67(4):816-817. doi:10.1016/j.eururo.2014.12.05.
7. Liu J, Yuan Y, Chen W, et al. Immune-checkpoint proteins VISTA and PD-1 nonredundantly regulate murine T-cell responses. *Proc Natl Acad Sci U S A*. 2015;112(21):6682-6687. doi:10.1073/pnas.1420370112
8. Daniel AS, Ana M, Paloma R, et al. Analysis of the PD-1/PD-L1 axis in human autoimmune thyroid disease: insights into pathogenesis and clues to immunotherapy associated thyroid autoimmunity. *J Autoimmun*. 2019;19:30156-30172. doi:10.1016/j.jaut.2019.05.013
9. Sun X, Yan X, Zhuo W, et al. PD-L1 nanobody competitively inhibits the formation of the PD-1/PD-L1 complex: comparative molecular dynamics simulations. *Int J Mol Sci*. 2018;19(7):1984-2007. doi:10.3390/ijms19071984
10. Andrew MI, Craig BT. At the bench: preclinical rationale for CTLA-4 and PD-1 blockade as cancer immunotherapy. *J Leukoc Biol*. 2013;94(1):25-39. doi:10.1189/jlb.1212621
11. Takayuki K, George CT, Vaishali RM. Aberrant T cell signaling and subsets in systemic lupus erythematosus. *Front Immunol*. 2018;9:1088-1103. doi:10.3389/fimmu.2018.01088
12. Li J, Jie HB, Lei Y, et al. PD-1/SHP-2 inhibit Tc1/Th1 phenotypic responses and the activation of T cells in the tumor microenvironment. *Cancer Res*. 2015;75(3):508-518. doi:10.1158/0008-5472.can-14-1215
13. Ishida Y, Agata Y, Shibahara K, et al. Induced expression of PD-1, a novel member of the immunoglobulin gene superfamily. *EMBO J*. 1992;11(11):3887-3896. doi:10.1002/j.1460-2075.1992.tb05481.x
14. Raissa F, Rafael MS, Henrique B, et al. Programmed cell death protein 1-PDL1 interaction prevents heart damage in chronic Trypanosoma cruzi infection. *Front Immunol*. 2018;9:997-1018. doi:10.3389/fimmu.2018.00997
15. Svetlana PS, Edward AC. The dual-function CD150 receptor subfamily: the viral attraction. *Nat Immunol*. 2003;4(1):19-24. doi:10.1038/ni0103-19
16. Elizabeth B, Anupam D. CTLA-4 and PD-1 pathways: similarities, differences, and implications of their inhibition. *Am J Clin Oncol*. 2016;39(1):98-106. doi:10.1097/COC.0000000000000239.
17. Anna S, Laurie M, Katharina B, et al. Negative immune checkpoints on T lymphocytes and their relevance to cancer immunotherapy. *Mol Oncol*. 2015;9(10):1936-1965. doi:10.1016/j.molonc.2015.10.008
18. Dong YN, Sun Q, Zhang XW. PD-1 and its ligands are important immune checkpoints in cancer. *Oncotarget*. 2017;8(2):2171-2186. doi:10.18632/oncotarget.13895
19. Shinohara T, Taniwaki M, Ishida Y, et al. Structure and chromosomal localization of the human PD-1 gene (PDCD1). *Genomics*. 1994;23(3):704-706. doi:10.1006/geno.1994.1562
20. Vibhakkar R, Juan G, Traganos F, et al. Activation-induced expression of human programmed death-1 gene in T-lymphocytes. *Exp Cell Res*. 1997;232(1):25-28. doi:10.1006/excr.1997.3493
21. Dai CQ, Lin FJ, Geng RX, et al. Implication of combined PD-L1/PD-1 blockade with cytokine-induced killer cells as a synergistic immunotherapy for gastrointestinal cancer. *Oncotarget*. 2016;7(9):10332-10344. doi:10.18632/oncotarget.7243
22. Lavakumar K, Purushottam L, James K, et al. PD-1 blunts the function of ovarian tumor-infiltrating dendritic cells by inactivating NF-κB. *Cancer Res*. 2016;76(2):239-250. doi:10.1158/0008-5472.CAN-15-0748
23. Lin WM, Chen M, Le H, et al. Crosstalk between PD-1/PD-L1 blockade and its combinatorial therapies in tumor immune

- microenvironment: a focus on HNSCC. *Front Oncol.* 2018;8: 532-548. doi:10.3389/fonc.2018.00532
24. Wong PF, Wei W, Gupta S, et al. Multiplex quantitative analysis of cancer-associated fibroblasts and immunotherapy outcome in metastatic melanoma. *J Immunother Cancer.* 2019;7(1):194-204. doi:10.1186/s40425-019-0675-0
 25. Guo YN, Feng XL, Jiang Y, et al. PD1 blockade enhances cytotoxicity of in vitro expanded natural killer cells towards myeloma cells. *Oncotarget.* 2016;7(30):48360-48374. doi:10.18632/oncotarget.10235
 26. Tang XL, Li QG, Zhu YQ, et al. The advantages of PD1 activating chimeric receptor (PD1-ACR) engineered lymphocytes for PDL1⁺ cancer therapy. *Am J Transl Res.* 2015;7(3):460-473. doi:ajtr.org/ISSN:1943-8141/AJTR0002962
 27. Borghaei H, Paz L, Horn L, et al. Nivolumab versus docetaxel in advanced nonsquamous non-small-cell lung cancer. *N Engl J Med.* 2015;373(17):1627-1639. doi:10.1056/NEJMoa1507643
 28. Wang CY, Thudium KB, Han M, et al. In vitro characterization of the anti-PD-1 antibody Nivolumab, BMS-936558, and in vivo toxicology in non-human primates. *Cancer Immunol Res.* 2014; 2(9):846-856. doi:10.1158/2326-6066.cir-14-0040
 29. Zhao BH, Zhang WX, Yu DL, et al. The benefit and risk of nivolumab in non-small-cell lung cancer: a single-arm meta-analysis of noncomparative clinical studies and randomized controlled trials. *Cancer Med.* 2018;7(5):1642-1659. doi:10.1002/cam4.1387
 30. Osamu K, Young HK, Yoshiki D, et al. Efficacy and safety of nivolumab in non-small cell lung cancer with preexisting interstitial lung disease. *Thorac Cancer.* 2018;9(7):847-855. doi:10.1016/j.jtho.2017.11.072
 31. Ana BM, José XL, Umberto LF, et al. Inhibition of the checkpoint protein PD-1 by the therapeutic antibody pembrolizumab outlined by quantum chemistry. *Sci Rep.* 2018;8(1):1840-1853. doi:10.1038/s41598-018-20325-0
 32. Gerry K, Thomas CC, Joanne WC, et al. Pembrolizumab (Keytruda). *Hum Vacc Immunother.* 2016;12(11):2777-2789. doi:10.1080/21645515.2016.1199310
 33. Smita SJ, Steven BM, Daniel VC. Pembrolizumab for treatment of advanced gastric and gastroesophageal junction adenocarcinoma. *Future Oncol.* 2018;14(5):417-430. doi:10.2217/fon-2017-0436
 34. Liu FX, Ou W, Diede SJ, et al. Real-world experience with pembrolizumab in patients with advanced melanoma a large retrospective observational study. *Medicine (Baltimore).* 2019; 98(30):e16542. doi:10.1097/MD.00000000000016542
 35. Toshio S, Takashi S, Fumihiko H, et al. Phase 1 study of pembrolizumab (MK-3475; anti-PD-1 monoclonal antibody) in Japanese patients with advanced solid tumors. *Invest New Drugs.* 2016;34:347-354. doi:10.1007/s10637-016-0347-6
 36. Tang LX, Bai JW, Chung CS, et al. Programmed cell death receptor ligand 1 modulates the regulatory T cells capacity to repress Shock/Sepsis-induced indirect acute lung injury by recruiting phosphatase SRC homology region 2 domain-containing phosphatase 1. *Shock.* 2016;43(1):47-54. doi:10.1097/SHK.0000000000000247
 37. Javier CG, Peter B, Zhai Y, et al. Quantitative interactomics in primary T cells provides a rationale for concomitant PD-1 and BTLA coinhibitor blockade in cancer immunotherapy. *Cell Rep.* 2019;27(11):315-3330. doi:10.1016/j.celrep.2019.05.041
 38. Elizabeth A, Heidi H, Jake S, et al. RANKL blockade improves efficacy of PD1-PD-L1 blockade or dual PD1-PD-L1 and CTLA4 blockade in mouse models of cancer. *Oncoimmunology.* 2018; 7(6):e1431088. doi:10.1080/2162402X.2018.1431088
 39. Liang LL, Ge K, Zhang F, et al. The suppressive effect of co-inhibiting PD-1 and CTLA-4 expression on H22 hepatomas in mice. *Cell Mol Biol Lett.* 2018;23(58):1392-1689. doi:10.1186/s11658-018-0122-0
 40. Zheng ZX, Bu ZD, Liu XJ, et al. Level of circulating PD-L1 expression in patients with advanced gastric cancer and its clinical implications. *Chin J Cancer Res.* 2014;26(1):104-112. doi:10.3978/j.issn.1000-9604.2014.02.08
 41. Jung HI, Jeong D, Ji S, et al. Overexpression of PD-L1 and PD-L2 is associated with poor prognosis in patients with hepatocellular carcinoma. *Cancer Res Treat.* 2017;49(1):246-254. doi:10.4143/crt.2016.066
 42. Liu B, Song Y, Liu D. Recent development in clinical applications of PD-1 and PD-L1 antibodies for cancer immunotherapy. *J Hematol Oncol.* 2017;10(1):174-182. doi:10.4143/crt.2016.066
 43. Shi X, Li X, Wang H, et al. Specific inhibition of PI3K δ/γ enhances the efficacy of anti-PD1 against osteosarcoma cancer. *J Bone Oncol.* 2019;16:100206-100214. doi:10.1016/j.jbo.2018.11.001
 44. Zhao J, Wen XF, Tian L, et al. Irreversible electroporation reverses resistance to immune checkpoint blockade in pancreatic cancer. *Nat Commun.* 2019;10(1):899-913. doi:10.1038/s41467-019-08782-1
 45. Huang RY, Cheryl E, Shashikant LL, et al. LAG3 and PD1 co-inhibitory molecules collaborate to limit CD8⁺ T cell signaling and dampen antitumor immunity in a murine ovarian cancer model. *Oncotarget.* 2015;6(29):27359-27377. doi:10.18632/oncotarget.4751
 46. Reem S, Salman MT, Sarah K, et al. Breast cancer cells and PD-1/PD-L1 blockade upregulate the expression of PD-1, CTLA-4, TIM-3 and LAG-3 immune checkpoints in CD4⁺ T Cells. *Vaccines (Basel).* 2019;7(4):149-162. doi:10.3390/vaccines7040149
 47. Tessa G, Wenbo Y, Gianpietro D, et al. GD2-specific CAR T Cells undergo potent activation and deletion following antigen encounter but can be protected from activation-induced cell death by PD-1 blockade. *Mol Ther.* 2016;24(6):1135-1149. doi:10.1038/mt.2016.63
 48. Yukihiro Y, Hiroyuki K. Post-progression survival after cessation of treatment with nivolumab for advanced non-small cell lung cancer: a retrospective study. *PLoS One.* 2018;13(8):e0203070. doi:10.1371/journal.pone.0203070
 49. Jedd DW, Vanna C, Rene G, et al. Overall survival with combined Nivolumab and Ipilimumab in advanced melanoma. *N Engl J Med.* 2017;377(14):1345-1356. doi:10.1056/NEJMoa1709684
 50. Huang M, Gilberto L, Ralph P, et al. Cost-effectiveness of pembrolizumab versus chemotherapy as first-line treatment in PD-L1-positive advanced non-small-cell lung cancer in the

- USA. *Immunotherapy*. 2019;11(17):1463-1478. doi:10.2217/imt-2019-0178
51. Anderson ES, Michael A, Jedd D, et al. Melanoma brain metastases treated with stereotactic radiosurgery and concurrent pembrolizumab display marked regression; efficacy and safety of combined treatment. *J Immunother Cancer*. 2017;5(1):76-84. doi:10.1186/s40425-017-0282-x
 52. Nanda R, Chow L, Dees E, et al. Pembrolizumab in patients with advanced triple-negative breast cancer: phase Ib KEYNOTE-012 Study. *J Clin Oncol*. 2016;34(21):2460-2467. doi:10.1200/jco.2015.64.8931
 53. Don M, Bakan E, Anjali M. The PD-1/PD-L1 axis modulates the natural killer cell versus multiple myeloma effect: a therapeutic target for CT-011, a novel monoclonal anti-PD-1 antibody. *Blood*. 2010;116(13):2286-2294. doi:10.1182/blood-2010-02-271874
 54. Mkrtichyan M, Najjar YG, Raulfs EC, et al. Anti-PD-1 synergizes with cyclophosphamide to induce potent anti-tumor vaccine effects through novel mechanisms. *Eur J Immunol*. 2011;41(10):2977-2986. doi:10.1002/eji.201141639
 55. Jacalyn R, Brett G, Heidi M, et al. PD-1 blockade by CT-011, anti-PD-1 antibody, enhances ex vivo T-cell responses to autologous dendritic cell/myeloma fusion vaccine. *J Immunother*. 2011;34(5):409-418. doi:10.1097/CJI.0b013e31821ca6ce
 56. Benson D, Bakan CE, Mishra A, et al. The PD-1/PD-L1 axis modulates the natural killer cell versus multiple myeloma effect: a therapeutic target for CT-011, a novel monoclonal anti-PD-1 antibody. *Blood*. 2010;116(13):2286-2294. doi:10.1182/blood-2010-02-271874
 57. Margaret KC, Michael AP, Jedd DW. Targeting T cell co-receptors for cancer therapy. *Immunity*. 2016;44(5):1069-1078. doi:10.1016/j.immuni.2016.04.023
 58. Keir ME, Butte MJ, Freeman GJ, et al. PD-1 and its ligands in tolerance and immunity. *Annu Rev Immunol*. 2008;26(1):677-704. doi:10.1146/annurev.immunol.26.021607.090331
 59. Liu C, Sun B, Xu B, et al. A panel containing PD-1, IL-2R α , IL-10, and CA15-3 as a biomarker to discriminate breast cancer from benign breast disease. *Cancer Manag Res*. 2018;10:1749-1761. doi:10.2147/CMAR.S160452
 60. Peter TS, Loise MF, Christopher VC, et al. The receptor PD-1 controls follicular regulatory T cells in the lymph nodes and blood. *Nat Immunol*. 2013;14(2):152-161. doi:10.1038/ni.2496
 61. Fujii T, Naing A, Rolfo, et al. Biomarkers of response to immune checkpoint blockade in cancer treatment. *Crit Rev Oncol Hematol*. 2018;130:108-120. doi:10.1016/j.critrevonc.2018.07.010
 62. Ren DX, Hua YZ, Yu BY. Predictive biomarkers and mechanisms underlying resistance to PD1/PD-L1 blockade cancer immunotherapy. *Mol Cancer*. 2020;19(1):1-19. doi:10.1186/s12943-020-1144-6
 63. Lei QY, Wang D, Sun K, et al. Resistance mechanisms of Anti-PD1/PDL1 Therapy in Solid Tumors. *Front Cell Dev Biol*. 2020;21(8):672-688. doi:10.3389/fcell.2020.00672
 64. Reem S, Eyad E. Acquired resistance to cancer immunotherapy: role of tumor-mediated immunosuppression. *Semin Cancer Biol*. 2020;65:13-27. doi:10.1016/j.semcancer
 65. Reem S, Eyad E. Treg-mediated acquired resistance to immune checkpoint inhibitors. *Cancer Lett*. 2019;457(10):168-179. doi:10.1016/j.canlet.2019.05.003