

p75 neurotrophin receptor: A potential surface marker of tongue squamous cell carcinoma stem cells

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Abstract. The present study detected p75 neurotrophin receptor (p75^{NTR}) expression in tongue squamous cell carcinoma (TSCC) cell lines, in order to define the biological properties of p75^{NTR+} cells and to confirm the use of p75^{NTR+} as a surface marker for TSCC stem cells. p75^{NTR+} cells were separated from Tca-8113 and CAL-27 TSCC cells by fluorescence-activated cell sorting. Colony formation, MTT and scratch assays, and a tumorigenicity analysis were performed to measure self-renewal and proliferation, multidirectional differentiation, and tumorigenicity of p75^{NTR+} cells. p75^{NTR+} cells comprised 3.1 and 1.9% of Tca-8113 and CAL-27 cells (mean of three experiments), respectively, and were more able to form colonies compared with non-sorted cells (P<0.01). In addition, the proportion of p75^{NTR+} cells generated from monoclonal p75^{NTR+} cells decreased to 14.5 (Tca-8113) and 5.8% (CAL-27) of cells within 2 weeks, thus suggesting that p75^{NTR+} cells are able to generate p75^{NTR+} and p75^{NTR-} cells. Furthermore, p75^{NTR+} cells exhibited increased proliferation, as evidenced by MTT assay (P<0.01) and had greater metastatic ability according to the scratch assay (P<0.01), compared with non-sorted cells. p75^{NTR+} cells also exhibited a greater tumorigenic capacity compared with non-sorted cells. In conclusion, p75^{NTR+} cells isolated from TSCC cell lines possess the characteristics of cancer stem cells; therefore, p75^{NTR} may be considered a useful surface marker for the identification of TSCC stem cells.

Introduction

Oral squamous cell carcinoma (OSCC) accounts for >90% of head and neck carcinomas; and, of these 90%, OSCCs of the tongue (TSCC) are reported to occur with rates of up to 40-50% (1). In order to enhance patient survival following TSCC, surgical techniques and diagnostic accuracy have improved; however, treatment failure persists due to local recurrence, and regional lymph node and distant metastases are common. It is well known that treatment failure may be associated with cancer stem cells (CSCs) or 'tumor-initiating cells' (2-4), which exhibit long-term self-renewal and a high migratory capacity, as well as the ability to generate phenotypically diverse tumor cells (5). This behavior is explained by the 'cancer stem cell' theory (6), which is a basis for oncology research (4,7,8). Previous studies in OSCC-derived cell lines have indicated that cell subpopulations with phenotypic and behavioral characteristics of normal epithelial stem cells may initiate tumorigenesis *in vivo* (9,10). CSCs rarely divide; however, they can produce fast-proliferating daughter cells. The majority of CSCs in various types of cancer have been isolated from tumor cells based on marker expression that characterizes stem cells in normal tissues (11). However, few studies have focused on the expression and function of a reliable marker to identify TSCC stem cells; therefore, at present, there is little understanding regarding their behavior and fate.

It has previously been reported that the p75 neurotrophin receptor (p75^{NTR}) may be involved in the invasion and poor prognosis of OSCC (12). As a member of the tumor necrosis factor superfamily, p75^{NTR} is a 75-kDa cell-surface receptor glycoprotein (13,14), which is involved in diverse cellular responses, including cell proliferation and survival, and apoptosis in neural and non-neural tissues (15,16) via unique pathways (17,18) or activation of the intrinsic caspase pathway (19). Furthermore, the expression and diverse function of p75^{NTR} has previously been reported in numerous types of cancer (20-22). Okumura *et al* (23) reported that p75^{NTR+} esophageal epithelial cells were actually stem cells, since they were able to proliferate, self-renew and undergo multidirectional differentiation. In addition, p75^{NTR} has been used to screen and identify mouse testis peritubular smooth muscle precursors (24), rat adipose multipotent stem cells (25) and human corneal epithelial progenitor cells (26).

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The present study detected p75^{NTR} expression in Tca-8113 and CAL-27 TSCC cell lines, and noted that p75^{NTR+} TSCCs exhibited CSC properties, particularly with regards to self-renewal and proliferation, multidirectional differentiation, and strong *in vivo* tumorigenic capacity.

Materials and methods

Cell source and culture conditions. Tca-8113 and CAL-27 TSCC cell lines were kindly provided by the Shanghai Key Laboratory of Stomatology (Department of Oral and Maxillofacial-Head Neck Oncology, Ninth People's Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China). The cell lines were originally purchased from the Shanghai Cell Biology Institute of the Chinese Academy of Sciences (Shanghai, China).

Tca-8113 cells were cultured in RPMI-1640 medium (Invitrogen; Thermo Fisher Scientific, Inc., Waltham, MA, USA) supplemented with 10% (v/v) fetal bovine serum (FBS; HyClone; GE Healthcare Life Sciences, Chalfont, UK), 100 IU/ml penicillin and 100 mg/ml streptomycin (Invitrogen; Thermo Fisher Scientific, Inc.). CAL-27 cells were cultured in Dulbecco's modified Eagle's medium (DMEM; HyClone; GE Healthcare Life Sciences) supplemented with 10% (v/v) FBS, 100 IU/ml penicillin and 100 mg/ml streptomycin. All cell cultures were maintained in a humidified incubator containing 5% CO₂/95% air at 37°C.

Flow cytometry and fluorescence-activated cell sorting (FACS). Tumor cells were harvested (final concentration, 1x10⁶ cells/ml) with Buffer 1 (PBS containing 0.5% bovine serum albumin (Sigma-Aldrich; Merck Millipore, Darmstadt, Germany) and 2 mM EDTA). Cells were then incubated with the primary antibody for 2 h at 4°C, washed twice in Buffer 1, and were resuspended in 500 µl Buffer 1, to which phycoerythrin (PE)-conjugated goat anti-mouse immunoglobulin G at a dilution of 1:100 (cat. no. 555749; BD Pharmingen, San Diego, CA, USA) was added. Cells were incubated in the dark for 15 min at 4°C. After staining, the samples were analyzed using a FACSCalibur flow cytometer with CellQuest software (version 5.1; BD Biosciences, San Jose, CA, USA). The primary antibody used was mouse anti-human p75^{NTR} at a dilution of 1:100 (cat. no. 557196; BD Pharmingen). FACS of p75^{NTR+} cells was performed using a Cytomation MoFlo[®] cytometer (Dako; Agilent Technologies, Santa Clara, CA, USA). The top 25% most brightly stained cells were isolated as p75^{NTR+} cells; cells incubated with PE-conjugated antibodies only were used as controls.

Colony formation assay. p75^{NTR+} single cell suspensions were prepared, diluted, and plated into a 96-well plate at various densities (1x10⁶/ml; 1x10⁵/ml; 1x10⁴/ml; 1x10³/ml; 1x10²/ml) (27). Cells were allowed 2 weeks to form colonies under standard conditions, and the rate at which this occurred was recorded. To assess p75^{NTR+} differentiation, colonies formed by one cell type were collected and incubated for another 2 weeks for p75^{NTR+} flow cytometric analysis.

MTT cell viability assay. Briefly, sorted p75^{NTR+} cells, and non-sorted Tca-8113 and CAL-27 cells (4x10³ cells/well)

were seeded in 96-well plates. After 1, 2, 3, 4, 5, 6 or 7 days, 100 µl MTT (5 mg/ml) was added, followed by incubation for a further 4 h at 37°C. The reaction was terminated by replacing MTT-containing medium with 100 µl acidic isopropanol (10% SDS, 5% isopropanol, 0.01 mol/l HCl); the resulting formazan crystals were dissolved by gentle agitation for ~10 min at room temperature. For colorimetric analysis, absorbance (490 nm) was measured on a microplate reader (Bio-Rad Laboratories, Inc., Hercules, CA, USA). The optical density (OD) values were analyzed using Quantity One analysis software (version 29.0; Bio-Rad Laboratories, Inc.). Each assay was repeated at least three times. Relative cell viability was compared to untreated (blanks) cells.

Scratch assay. Live tumor cells (Tca-8113 and CAL-27) were harvested from standard cultures and grown to confluence on 6-well Permaxox[™] plates. Consistently shaped wounds were made using a sterile 200 µl pipette tip across each well, creating a cell-free area, according to previously described methods (28). Cultures were gently washed with PBS to remove loose cells and adherent cells were maintained in culture medium supplemented with 1% FBS. Non sorted control cells were also scratched, washed, and maintained in culture medium supplemented with 1% FBS after scratching. Immediately after scratching and at 12 h, 3, 4, 5 and 8 d, images of the scraped areas were captured under phase contrast microscopy. The wounded areas and scratch widths were measured at six different points per image, and the same scratched area was used for every assessment.

Xenograft tumorigenicity assay. Throughout experiments, animals were maintained under the Guidelines for Animal Experimentation of the School and Hospital of Stomatology, Shandong University (Jinan, China). Experiments were conducted according to the National Institute of Health Guidelines for Research Involving Recombinant or Synthetic Nucleic Acid Molecules (November, 2013) the (regarding the care and use of animals for experimental procedures. In addition, the present study was independently reviewed and approved by the Animal Experimentation Committee of Shandong University, and was approved by the Medical Ethics Committee of the School of Medicine, Shandong University.

Briefly, 5-6 week-old male BALB/c nude mice (~18±1.2 g; n=106) were obtained from Beijing Vital River Laboratory Animal Technology Co., Ltd. (Beijing, China), and were maintained in plastic cages (n=3-5/cage) under standard laboratory conditions with a 12 h dark, 12 h light cycle and a constant temperature of 23°C and humidity of 48%. All mice were fed a standard rodent diet *ad libitum*. After 1 week of acclimation, mice were randomly divided into four groups (named as follows: 75NTR⁺, Tca-8113 cells, non-sorted Tca-8113 cells, p75NTR⁺ CAL-27 cells and non-sorted CAL-27 cells injection; n=24/group) and each group was subdivided into four subgroups (n=6/subgroup). Four cell suspensions of p75^{NTR+} and non-sorted cells were prepared (1.0x10³/ml, 1.0x10⁴/ml, 1.0x10⁵/ml and 1.0x10⁶/ml) in 200 µl serum-free DMEM. These suspensions were injected subcutaneously into BALB/c nude mice, following anesthetization with 10% chloral hydrate (400 mg/100 g body weight) at room temperature. Tca-8113



Figure 1. Nude mice xenograft model formed using p75^{NTR+} Tca-8113 cells. (A) Inoculation of tumor cells into the backs of the mice. (B) Tumor formation after 4 weeks, and (C) removal of the tumor after 6 weeks. p75^{NTR}, p75 neurotrophin receptor.



Figure 2. Nude mice xenograft model formed using p75^{NTR+} CAL-27 cells. (A) Inoculation of tumor cells into the axilla. (B) Tumor formation after 2 weeks, and (C) removal of the tumor after 6 weeks. p75^{NTR}, p75 neurotrophin receptor.

cells were injected into the backs of the mice (Fig. 1), whereas CAL-27 cells were injected into the axilla of the mice (Fig. 2). Sterile PBS was injected into the contralateral side as a control.

At the end of the experiment, the mice were anesthetized as aforementioned (10% chloral hydrate, 400 mg/100 g) and subjected to transcardial perfusion with a fixative of 4% paraformaldehyde in 0.1 M phosphate buffer (pH 7.4) 6 weeks following tumor cell inoculation. Subsequently, tumors were removed and immersed in the same fixative for an additional 24 h at 4°C. Samples were dehydrated with graded ethanol and were embedded in paraffin according to standard procedures. Subsequently, 5- μ m serial sections were prepared for histological hematoxylin and eosin (H&E) staining (LEICA SM 2010R; Leica Microsystems GmbH, Wetzlar, Germany).

Histological examination and image analysis. H&E staining was performed to investigate the morphology. The prepared sections were immersed in Erthlich's haematoxylin for 15 min. Then the sections were washed with distilled water and differentiated in 1% HCl in 70% alcohol for 1 min and washed again for 2 min. After that, the sections were stained with 1% eosin for 10 min and washed with distilled water. Finally, all sections were dehydrated and mounted. The stained sections were observed and then digital images were taken with a light microscope (Olympus BX-53; Olympus Corporation, Tokyo, Japan).

Statistical analysis. For comparing distant metastases between p75^{NTR+} and negative groups, a χ^2 test was performed. Fisher's exact test was used to compare tumorigenicity between p75^{NTR+} and negative groups. Other experimental data are presented as the mean \pm standard deviation and were analyzed using Student's t-test. P<0.05 was considered to indicate a statistically significant difference. Statistical analyses were

conducted using SPSS 19.0 software (SPSS Inc., Chicago, IL, USA).

Results

Flow cytometry. p75^{NTR+} cells were detected in both TSCC cell lines (Fig. 3). p75^{NTR+} cells accounted for 3.1 and 1.9% of Tca-8113 and CAL-27 cells, respectively (an average of three experiments). After cell sorting, p75^{NTR+} cells accounted for 98.1 (Tca-8113) and 97.4% (CAL-27) of all sorted cells (Fig. 4).

Colony formation assay. The colony-forming ability of p75^{NTR+}, and non-sorted Tca-8113 and CAL-27 TSCC cells (Fig. 5) was assessed. The number of colonies formed by p75^{NTR+} cells from each cell line was greater compared with the number formed by non-sorted TSCC cells (Table I). Following 2 weeks of culture, monoclonal p75^{NTR+} cells were collected and cultured for a further 2 weeks, after which they were analyzed by flow cytometry. Data indicated that the colonies contained both p75^{NTR+} and p75^{NTR-} cells. The proportion of p75^{NTR+} cells in the Tca-8113 and CAL-27 cell populations was 14.5 and 5.8%, respectively (Fig. 6). These results indicated that p75^{NTR+} cells exhibit self-renewing and multidirectional differentiation properties.

MTT cell viability assay. To determine the proliferative ability of p75^{NTR+} and non-sorted cells *in vitro*, an MTT assay was conducted. The results revealed no differences in proliferative ability, according to OD values, on the first day (Fig. 7). After 3 days of culture, OD values for the p75^{NTR+} cells were greater compared with the non-sorted cells (P<0.05), indicating a stronger proliferative ability of p75^{NTR+} cells *in vitro*. Furthermore, proliferation was greater on days 5 and 7 (P<0.01; Fig. 7).

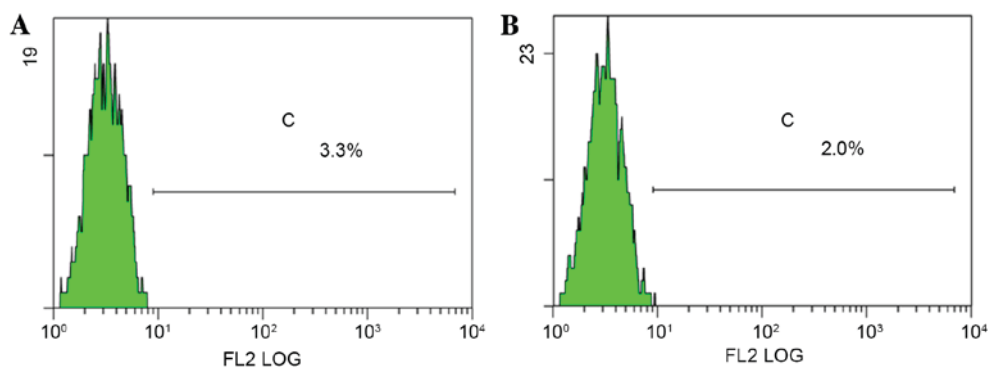


Figure 3. Flow cytometry. p75^{NTR+} cells (average of three experiments) in (A) Tca-8113 and (B) CAL-27 tongue squamous cell carcinoma cell lines. The average of three experiments indicated that the proportion of p75^{NTR+} cells was 3.1 and 1.9% in Tca-8113 and CAL-27 cells, respectively. p75^{NTR}, p75 neurotrophin receptor.

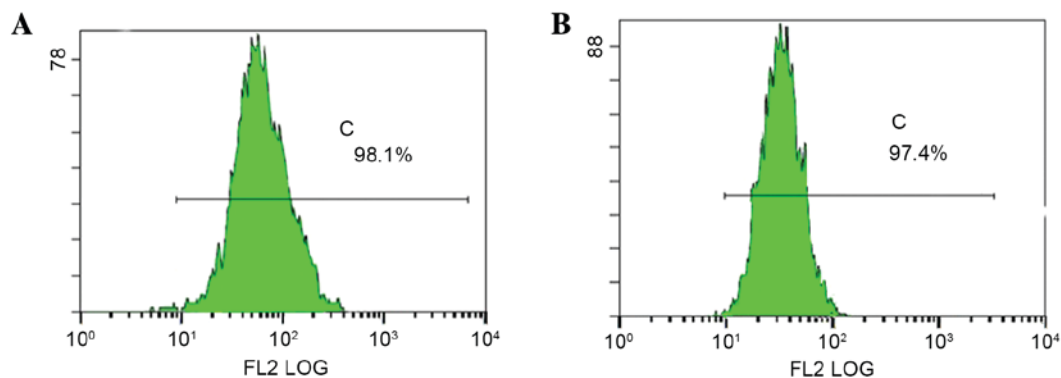


Figure 4. p75^{NTR+} sorted cells from (A) Tca-8113 and (B) CAL-27 tongue squamous cell carcinoma cell lines. p75^{NTR+} cells were sorted and accounted for 98.1 (Tca-8113) and 97.4% (CAL-27) of sorted cells. p75^{NTR}, p75 neurotrophin receptor.

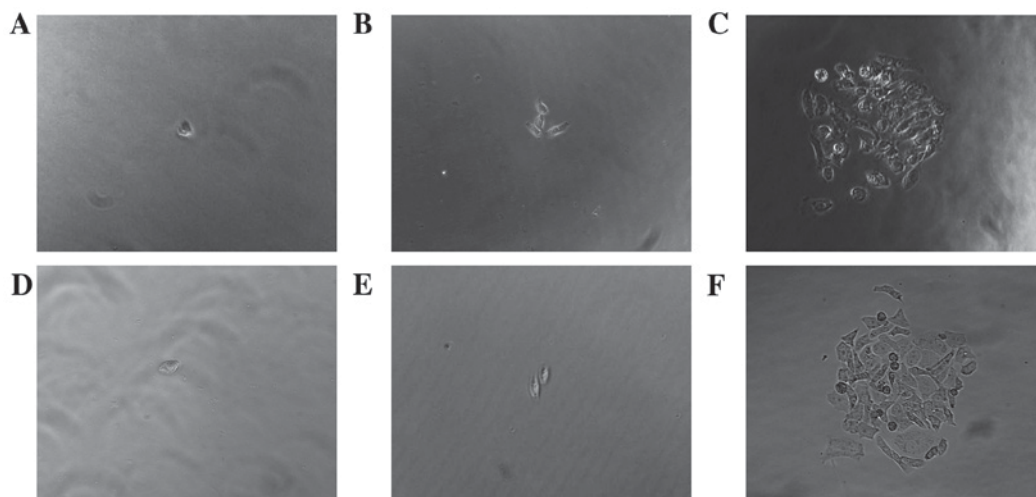


Figure 5. Colony formation assay for (A-C) p75^{NTR+} Tca-8113 and (D-F) p75^{NTR+} CAL-27 cells. Cell colonies formed by Tca-8113 TSCC single cells in culture after (A) 12 h, (B) 48 h and (C) 7 days of culture. Colonies formed by CAL-27 TSCC single cells after (D) 12 h, (E) 48 h and (F) 7 days of culture. Single TSCC cells proliferated in normal culture and formed colonies. A total of 2-7 days after plating, colony formation was observed. TSCC, tongue squamous cell carcinoma. A-F, x400 magnification.

Scratch assay. Cells in each group were observed 8 days after the scratch assay and the data indicated that p75^{NTR+} cells exhibited better wound healing compared with the controls (Figs. 8 and 9). By day 8, the wounded area in the Tca-8113 p75^{NTR+} cell group was covered, whereas non-sorted cells still

retained a denuded area (Fig. 8). In the CAL-27 p75^{NTR+} cell group, cells covered the wounded area by day 5 (Fig. 9). These results suggested that p75^{NTR+} cells exhibit better migratory and invasive characteristics compared with non-sorted cells (Tca-8113, $P < 0.05$; CAL-27, $P < 0.05$; Fig. 10).

Table I. Colony formation between p75^{NTR+} and non-sorted cells.

Cell line	p75 ^{NTR+} cell colony formation (%)	Non-sorted cell colony formation (%)	P-value
Tca-8113	32.92±1.91	7.09±1.20	P=0.0001
CAL-27	34.96±2.03	5.54±2.16	P=0.0001

p75^{NTR}, p75 neurotrophin receptor.

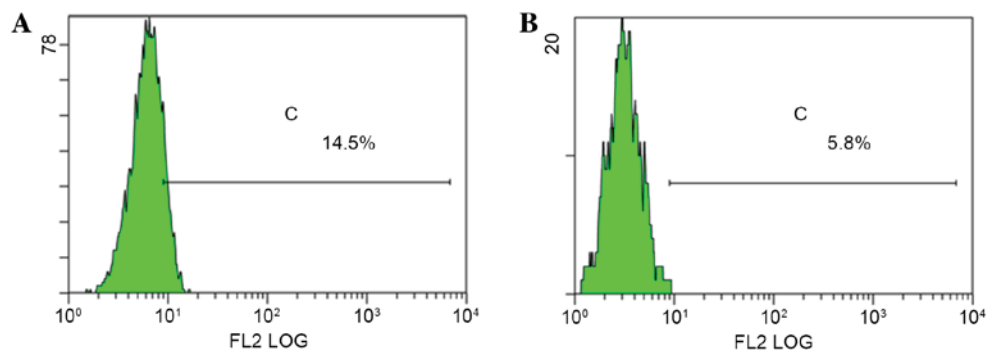


Figure 6. p75^{NTR+} cell colonies formed by sorted cells from (A) Tca-8113 and (B) CAL-27 tongue squamous cell carcinoma cell lines. Flow cytometry confirmed that colonies contained p75^{NTR+} and p75^{NTR} cells. p75^{NTR+} cells accounted for 14.5 and 5.8% of Tca-8113 and CAL-27 cells, respectively. p75^{NTR+} cells exhibited self-renewing and differentiation abilities. p75^{NTR}, p75 neurotrophin receptor.

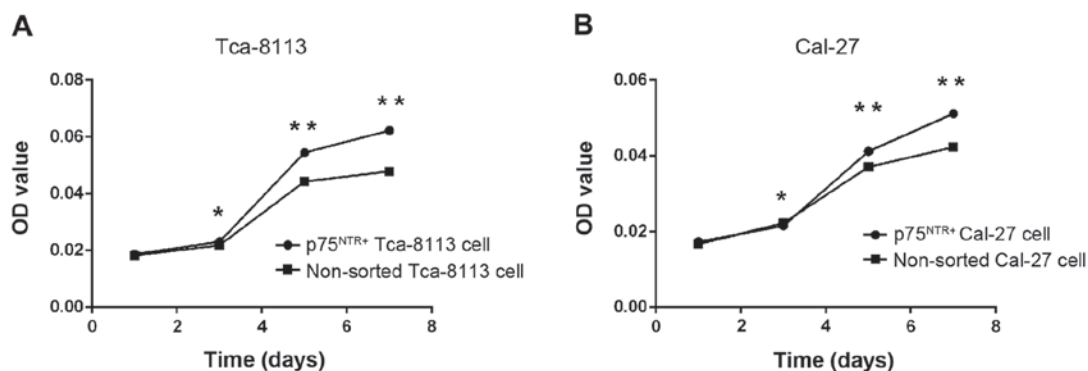


Figure 7. Growth curves of sorted p75^{NTR+} cells and non-sorted cells from the (A) Tca-8113 and (B) CAL-27 tongue squamous cell carcinoma cell lines. Cell growth curves were determined using MTT assay. No differences in proliferative ability (OD values) were detected on day 1. After 3 days of culture, OD values for p75^{NTR+} cells were greater compared with non-sorted cells (*P<0.05), indicating stronger proliferative ability of p75^{NTR+} cells *in vitro*. Proliferation was increased on days 5 and 7 (**P<0.01). *P<0.05 vs. non-sorted cells. p75^{NTR}, p75 neurotrophin receptor; OD, optical density.

Xenograft tumorigenicity assay. Various numbers of sorted p75^{NTR+} and non-sorted TSCC cells were injected into nude mice, as aforementioned. The results indicated that at least 5x10³ p75^{NTR+} cells or 5x10⁴ non-sorted cells were required to generate tumors. Tumorigenicity of the p75^{NTR+} and non-sorted cell groups are presented in Table II. Tumorigenicity of p75^{NTR+} cells was significantly higher compared with non-sorted cells according to Fisher's exact test.

H&E staining was used to identify morphological differences between the treatment groups. Stained sections were observed and digital images were captured under a light microscope (Olympus BX-53; Olympus Corporation). Both tumor types appeared to be squamous cell carcinomas with no intercellular bridges and undetectable mitosis. Tumors originating from Tca-8113 cells had larger necrotic areas

compared with those generated from CAL-27 cells. The tumor cell lines generated abundant cancer cell nests, thus resembling normal human, moderately differentiated TSCC (Fig. 11).

Discussion

p75^{NTR} expression was present in Tca-8113 and CAL-27 TSCC cell lines, as indicated in the results of the present study; these data were similar to those of previous studies, which detected the presence of cancer stem cells (CSCs) (29,30). p75^{NTR+} cells isolated from TSCC cell lines exhibited characteristics of CSCs, and were able to self-renew, proliferate and undergo multidirectional differentiation. Furthermore, p75^{NTR+} cells exhibited strong tumorigenic capacity *in vivo*. These results

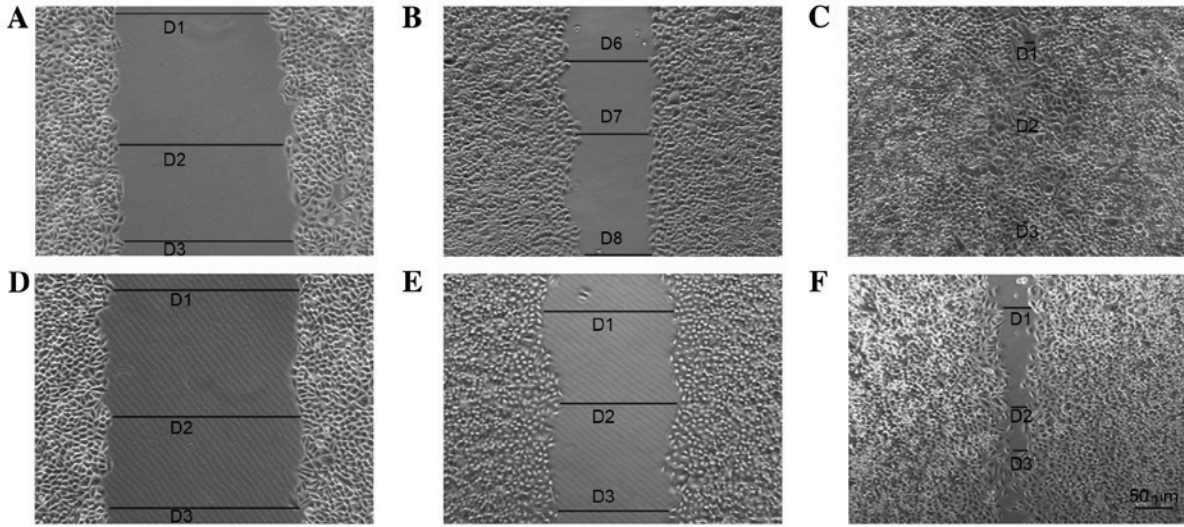


Figure 8. Scratch assay of Tca-8113 tongue squamous cell carcinoma cells. p75^{NTR+} cell group at (A) the time of scratching, (B) after 4 days and (C) after 8 days. Non-sorted cells at (D) the time of scratching, (E) after 4 days and (F) after 8 days. p75^{NTR}, p75 neurotrophin receptor. A-F, x40 magnification.

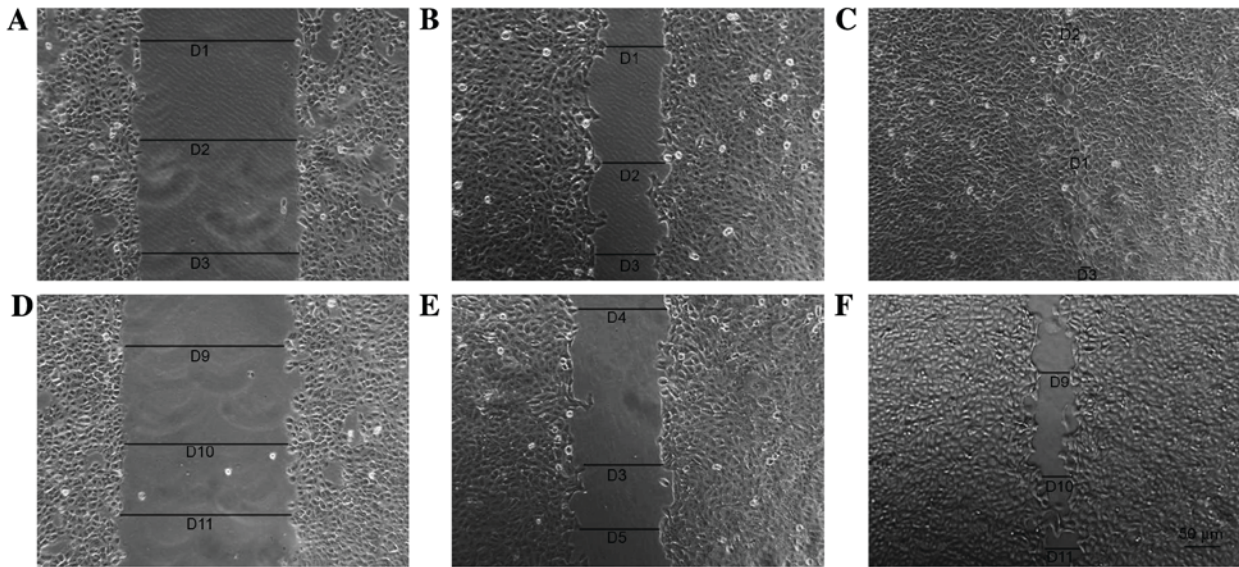


Figure 9. Scratch assay of CAL-27 tongue squamous cell carcinoma cells. p75^{NTR+} cell group at (A) the time of scratching, (B) after 3 days later and (C) after 5 days. Non-sorted cells at (D) the time of scratching, (E) after 3 days and (F) after 5 days. p75^{NTR}, p75 neurotrophin receptor. A-F, x40 magnification.

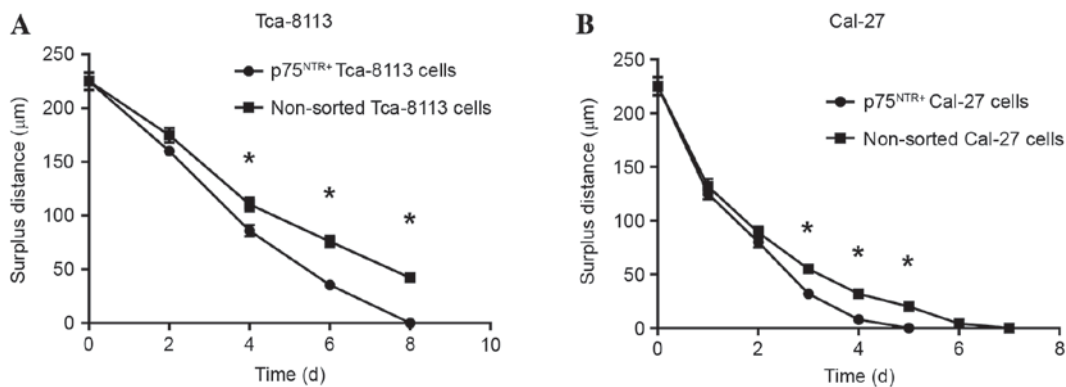


Figure 10. Surplus distance curve of sorted p75^{NTR+} and non-sorted (A) Tca-8113 and (B) CAL-27 tongue squamous cell carcinoma cells. After the scratch assay, cells from each group were observed for 8 days; p75^{NTR+} cells healed better than control cells. By day 8, the wounded area of the Tca-8113 p75^{NTR+} group was covered, whereas the non-sorted group retained a denuded area. By day 5, the wounded area of the CAL-27 p75^{NTR+} group was covered. p75^{NTR+} cells exhibit better migratory and invasive characteristics compared with non-sorted cells (Tca-8113, P<0.05; CAL-27, P<0.05). *P<0.05 vs. non-sorted cells. p75^{NTR}, p75 neurotrophin receptor.

Table II. Tumorigenicity of p75^{NTR+} and non-sorted tongue squamous cell carcinoma cells in a nude mouse xenograft model.

Cell group	Cell orders of magnitude/tumor numbers				P-value
	1.0x10 ³	1.0x10 ⁴	1.0x10 ⁵	1.0x10 ⁶	
p75 ^{NTR+} Tca-8113 cells	1/6	5/6	6/6	6/6	P=0.0176
Non-sorted Tca-8113 cells	0/6	2/6	2/6	6/6	
p75 ^{NTR+} CAL-27 cells	2/6	5/6	5/6	6/6	P=0.0084
Non-sorted CAL-27 cells	0/6	1/6	2/6	5/6	

p75^{NTR}, p75 neurotrophin receptor.

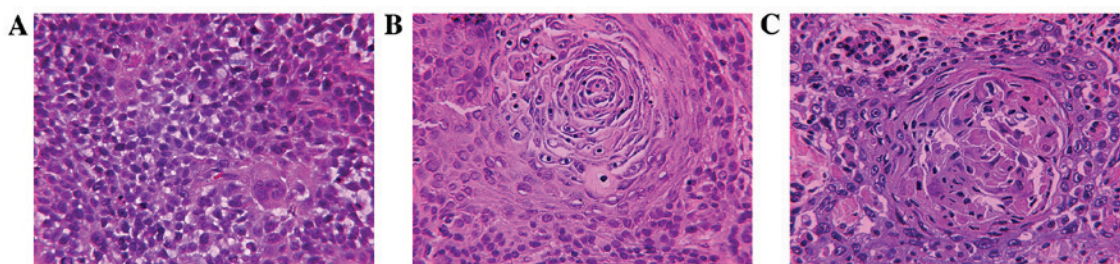


Figure 11. Tumor formation following inoculation with p75^{NTR+} TSCC cells (hematoxylin and eosin staining). (A) Tca-8113 and (B) CAL-27 cells. (C) Moderately differentiated human TSCC, used as a positive control. Squamous cell carcinomas with no intercellular bridges and undetectable mitosis were detected. Tumors originating from Tca-8113 cells exhibited larger necrotic areas compared with those derived from CAL-27 cells. Inoculation with both cell lines resulted in the generation of abundant cancer cell nests, which resemble normal human, moderately differentiated TSCC. p75^{NTR}, p75 neurotrophin receptor; TSCC, tongue squamous cell carcinoma. A-F, x 400.

indicated that p75^{NTR} may be considered a useful surface marker of TSCC stem cells.

Previous studies have reported on the existence of CSCs in brain tumors (4,6), liver (31,32), lung (33), breast (34), colorectal (35) and pancreatic carcinoma (36), and thyroid tumors (37). Bao *et al* (38) reported that cluster of differentiation (CD)133⁺ CSCs contributed to glioma radioresistance via preferential activation of the DNA damage checkpoint response and increased DNA repair capacity. In addition, the proportion of CD133⁺ cells in these tumors was 2-3%. Liu *et al* (39) reported that CD133⁺ CSCs, which accounted for 10.2% of brain glioblastoma cells, exhibited chemoresistance due to their increased expression of ATP-binding cassette sub-family G member 2 and O-6-methylguanine-DNA methyltransferase.

The results of the present study are consistent with those of previous reports, which demonstrated that the expression of p75^{NTR} is associated with the occurrence and development of numerous types of cancer, including stomach cancer (40), retinal neuroblastoma (41), prostatic carcinoma (42), pancreatic cancer (43) and melanoma (44). The present results suggested that the CSC theory is valid; and a small population of CSCs stimulate tumor recurrence and metastases.

Self-renewal is a hallmark of CSCs, since they are able to form new, identical stem cells that can proliferate, expand and differentiate. Colony-forming ability reflects self-renewal capacity, and previous studies regard colony-forming cells as CSCs (34,45). In the present study, a colony formation assay confirmed that p75^{NTR+} cells exhibited a better colony-forming ability compared with non-sorted cells, and these data agree with the fact that CSCs are capable of self-renewal. In addition,

flow cytometry confirmed that the proportion of p75^{NTR+} cells was significantly decreased after 4 weeks of colonization *in vitro*. Therefore, p75^{NTR+} cells may generate p75^{NTR+} and p75^{NTR-} progenies, thus suggesting they possess multidirectional differentiation ability.

The results of an MTT assay revealed that the viability of p75^{NTR+} cells was significantly higher compared with non-sorted cells. Invasion and metastasis are important biological characteristics of malignant tumors, which are associated with patient mortality and chemoresistance. According to the results of a scratch assay, p75^{NTR+} cells possess migratory ability. Kiyosue *et al* (12) reported a significant association between the magnitude of p75^{NTR} positivity and the mode of tumor invasion. Furthermore, Soland *et al* (46) reported that the average risk of local recurrence was increased by ~17-fold in OSCC when p75^{NTR} expression was positive, and marked tumor cell dissociation was detected at the invasive front. Therefore, p75^{NTR+} may be involved in the invasive pattern of cancer cells.

In the present study nude mice were inoculated with various numbers of p75^{NTR+} cells isolated from Tca-8113 and CAL-27 TSCC cell lines, or non-sorted cells, in order to determine whether p75^{NTR+} cells exhibit greater tumorigenicity compared with non-sorted cells. The results indicated that p75^{NTR+} cells possess stronger tumorigenicity. H&E staining revealed that the inoculated mice possessed squamous cell carcinoma with no intercellular bridges and undetectable mitosis. In addition, Tca-8113 cell tumors manifested larger necrotic areas compared with those derived from CAL-27 cells. Both tumor types possessed abundant cancer cell nests,

which resembled normal human, moderately differentiated TSCC. Therefore, the present study suggested that p75^{NTR+} cells may be useful for the surface marker identification of TSCC stem cells.

In conclusion, p75 neurotrophin receptor (p75^{NTR}) expression was detected in TSCC cell lines and found that p75^{NTR+} cells isolated from TSCC cell lines possess the characteristics of cancer stem cells. These findings suggest that p75^{NTR} may be considered a useful surface marker for the identification of TSCC stem cells, providing a potential target for novel therapies.

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