



Genome Sequence of *Acetobacter tropicalis* DmPark25_167, a Bacterium Isolated from a *Drosophila melanogaster* Genetic Model of Parkinson's Disease

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ABSTRACT Here, we report the genome of *Acetobacter tropicalis* DmPark25_167, a bacterial strain isolated from a *Drosophila melanogaster park²⁵* mutant. The *park²⁵* mutant is an established genetic model of Parkinson's disease. DmPark25_167 has duplicated methionine metabolism and type IV secretion gene alleles compared with another strain of *A. tropicalis*.

Acetobacter tropicalis is a species of acetic acid bacteria commonly associated with *Drosophila melanogaster* (1). Here, we report the genome sequence of *A. tropicalis* DmPark25_167, a strain cultured from homogenates of a *D. melanogaster park²⁵* mutant, which is a model of Parkinson's disease that was provided to the Call laboratory by Leo Pallanck (2). We also report genomic differences between DmPark25_167 and DmCS_006, another *A. tropicalis* strain. We present this genome in our continued efforts to understand the genomic basis for *Drosophila*-microbiota interactions.

The genome sequence of DmPark25_167 was obtained using the following approach. A pool of five whole-body *Drosophila park²⁵* mutants (reared on the following diet formulation per 1 liter food: 0.957 liter water, 11.8 g yeast, 5.9 g soy flour, 5.9 g agar, 58.8 g yellow cornmeal, 35.3 ml light corn syrup, 35.3 ml molasses, 4.4 ml propionic acid, and 10 ml 10% methylparaben in 95% ethanol) were homogenized and plated onto modified MRS medium (3) in a Glendale, AZ, laboratory. Copper and tan colonies that grew were streaked for isolation, and their taxonomic assignment as *Acetobacter* species was confirmed by Sanger sequencing using primers based on the universal 27F and 1492R primers (27F, 5'-AGAGTTTGATCMTGGCTCAG-3'; 1492R, 5'-TACGGYACTACCTTGTTACGACTT-3'). Following our previous methods (4), we prepared a genome sequencing library for an isolate with 100% 16S sequence identity to *A. tropicalis* DmCS_006, a strain of *Acetobacter* isolated from laboratory *D. melanogaster* Canton-S in Ithaca, NY (5). DNA was extracted using the Qiagen DNeasy Powerlyzer microbial kit, fragmented to 500 bp (NEBNext double-stranded DNA [dsDNA] Fragmentase [M0348L]), end repaired (NEBNext Ultra II end repair/dA-tailing module [E7546L]), ligated to Illumina sequencing adapters (NEBNext Ultra II ligation module [E7595L]), and size selected to ~500 bp. Ligated target sequences were enriched with the KAPA library amplification kit (KK2621), size selected to 500 bp, and normalized to 5 ng/ μ l using Qubit measurements. The library was sequenced by 250-bp paired-end sequencing on a partial lane of an Illumina HiSeq 2500 instrument. The 1,828,193 paired-end reads that passed Illumina default quality filtering were assembled with Velvet version 1.2.10 (6). A k-mer length of 151 was chosen by testing k-mer lengths between 135 and 231 in increments of 8 and selecting an assembly that maximized the N_{50} value at an estimated nucleotide coverage of 114 \times . The final assembly contained 3,722,678 bp in 138 contigs and had an N_{50} value of 105,815 and a 55.5% GC content. The taxonomic assignment as *A. tropicalis* was verified using ANIm analysis in JSpeciesWS (7). DmPark25_167 contains ~3,383 genes that code for 3,215

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proteins, as annotated by the NCBI Prokaryotic Genome Annotation Pipeline (8). Default parameters were used for all software unless otherwise specified.

We compared the gene contents of DmPark25_167 and DmCS_006 using RAST 2.0 annotations (9). Relative to DmCS_006, DmPark25_167 has perfect and lower-similarity (<89%) versions of single-copy DmCS_006 methionine metabolism and type IV secretion genes. The type IV secretion genes are on a contig with a plasmid replication initiation protein, suggesting that they might be on a plasmid.

Data availability. This whole-genome shotgun project and raw reads have been deposited at DDBJ/ENA/GenBank under accession numbers [JACAOJ000000000](https://doi.org/10.1093/nar/gkw569) and [SAMN15399467](https://doi.org/10.1093/nar/gkw569), respectively.

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REFERENCES

1. Staubach F, Baines JF, Künzel S, Bik EM, Petrov DA. 2013. Host species and environmental effects on bacterial communities associated with *Drosophila* in the laboratory and in the natural environment. *PLoS One* 8:e70749. <https://doi.org/10.1371/journal.pone.0070749>.
2. Greene JC, Whitworth AJ, Kuo I, Andrews LA, Feany MB, Pallanck LJ. 2003. Mitochondrial pathology and apoptotic muscle degeneration in *Drosophila parkin* mutants. *Proc Natl Acad Sci U S A* 100:4078–4083. <https://doi.org/10.1073/pnas.0737556100>.
3. Newell PD, Douglas AE. 2014. Interspecies interactions determine the impact of the gut microbiota on nutrient allocation in *Drosophila melanogaster*. *Appl Environ Microbiol* 80:788–796. <https://doi.org/10.1128/AEM.02742-13>.
4. Rudman SM, Greenblum S, Hughes RC, Rajpurohit S, Kiratli O, Lowder DB, Lemmon SG, Petrov DA, Chaston JM, Schmidt P. 2019. Microbiome composition shapes rapid genomic adaptation of *Drosophila melanogaster*. *Proc Natl Acad Sci U S A* 116:20025–20032. <https://doi.org/10.1073/pnas.1907787116>.
5. Newell PD, Chaston JM, Wang Y, Winans NJ, Sannino DR, Wong ACN, Dobson AJ, Kagle J, Douglas AE. 2014. In vivo function and comparative genomic analyses of the *Drosophila* gut microbiota identify candidate symbiosis factors. *Front Microbiol* 5:576. <https://doi.org/10.3389/fmicb.2014.00576>.
6. Zerbino DR, Birney E. 2008. Velvet: algorithms for *de novo* short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <https://doi.org/10.1101/gr.074492.107>.
7. Richter M, Rosselló-Móra R, Oliver Glöckner F, Peplies J. 2016. JSpeciesWS: a web server for prokaryotic species circumscription based on pairwise genome comparison. *Bioinformatics* 32:929–931. <https://doi.org/10.1093/bioinformatics/btv681>.
8. Tatusova T, DiCuccio M, Badretdin A, Vyacheslav C, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
9. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.