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*CORRESPONDENCE Valter Aragão do Nascimento aragao60@hotmail.com

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Oxidative stability and elemental analysis of sunflower (*Helianthus annuus*) edible oil produced in Brazil using a domestic extraction machine

David Johane Machate¹, Elaine S. P. Melo², Lincoln Carlos Silva de Oliveira³, Danielle Bogo⁴, Flávio S. Michels⁵, Arnildo Pott⁶, Leandro F. Cavalheiro³, Rita de Cássia Avellaneda Guimarães⁴, Karine de Cássia Freitas⁴, Priscila Aiko Hiane⁴, Anderson R. L. Caires⁵, Marcelo Luiz Brandão Vilela², Rodrigo Juliano Oliveira⁴ and Valter Aragão do Nascimento^{1.2*}

¹Graduate Program in Materials Science, Federal University of Mato Grosso do Sul, Campo Grande, Brazil, ²Group of Spectroscopy and Bioinformatics Applied Biodiversity and Health (GEBABS), Graduate Program in Health and Development in the Central-West Region of Brazil, Federal University of Mato Grosso do Sul, Campo Grande, Brazil, ³Chemistry Institute, Federal University of Mato Grosso do Sul, Campo Grande, MS, Brazil, ⁴Post-graduate Program in Health and Development in the Mid-West Region, Federal University of Mato Grosso do Sul, Campo Grande, MS, Brazil, [§]Optics and Photonics Group, Institute of Physics, Federal University of Mato Grosso do Sul, Campo Grande, MS, Brazil, [§]Graduate Program in Biotechnology and Biodiversity in the Central-West Region, Federal University of Mato Grosso do Sul, Campo Grande, MS, Brazil

The consumption of regular vegetable oils has been linked to energy acquisition, nutritional benefits, health improvement, and the regulation of metabolic diseases. This study evaluated fatty acids composition, physicochemical, thermal, oxidative, and optical properties, and quantified trace elements in the sunflower oil extracted by a domestic cold-press machine. The oil presented linoleic (54.00%) and oleic (37.29%) primary unsaturated fatty acids (91.67%), in which atherogenic (0.05), thrombogenic (0.16), hypocholesterolemic/hypercholesterolemic (21.97), peroxide (16.16), saponification (141.80), and relative density indices (0.92) demonstrated to be suitable for human consumption and possible health promotion. In addition, the concentrations of trace elements by ICP OES were ordered Zn > Fe > Al > Cu > Mn > Cr. Concentrations of Zn, Fe, Al, Cu, and Mn were lower than FAO/WHO and DRI/AI limits, while Cr concentrations exceeded the FAO/WHO limits, which can be used as an indicator of the polluted ambiance. Sunflower oil quantities daily consumption were calculated by taking into account non-carcinogenic risk (CR $< 10^{-4}$), and total non-carcinogenic hazard index (HI < 1). Based on trace elements determined in this study, the suitable quantity of sunflower oil consumption varies according to individuals aged 8, 18, and 30 years and will be deemed 0.61, 1.46, and 1.65 g/kg, respectively, attending HI = 0.99 and $CR < 10^{-4}$.

KEYWORDS

cold-press, vegetable oil quality, trace elements, oil quantity consumption, non-carcinogenic indices

Introduction

One of the three main energy sources for human life activities is edible vegetable oil, whose stages (field production, harvesting, processing, storage) must be monitored (1). In addition, vegetable oils are strictly associated with health promotion and metabolic diseases prevalence such as obesity, diabetes mellitus, cardiovascular (CVD), coronary heart diseases (CHDs), systemic inflammation, atherosclerosis, cancers, etc. (2). Benefits of vegetable oils are associated with fatty acids composing them: saturated (C6:0-C20:0, no double bonds) monounsaturated (C16:1-C20:1, one double bond) and polyunsaturated (C18:2-C24:6), two up to six double bonds) (2, 3), essential components: macroand microelements, vitamins A, C, K, and E (tocopherols, phytosterols, phytostanols, tocotrienols), carotenoids, chlorophylls, polyphenols, flavonoids, acting as antioxidants and health promoting (4-6).

Most commercially available plant seed and fruit oils are extracted using petroleum solvents, with high toxicity levels for human health (7). Moreover, their essential components are removed during refining (chemical and physical) and bleaching processes, formatting short-chain compounds [esters, polymeric triacylglycerols, trans and free fatty acids, (hydro) peroxides, dienes and trienes, and others], which act as pro-oxidants (8). These substances reduce vegetable oil shelf life and vital properties, which decrease healthy and nutraceutical properties, and are associated with the occurrence of several metabolic diseases in consumers (9) due to the higher amount of free radicals increasing oxygen species (10).

Globally, the sunflower is the second most cultivated plant after maize, and its seeds are used to treat inflammatory, hypertensive, CVD, and other diseases (11). Additionally, its oil controls tumors, diabetes, cholesterol, cancers, hypertension, hypercholesterolemia, and CHD. These benefits are associated with the synergic action of tocopherols, phytosterols, terpenoids, polyphenols, carotenoids, alkaloids, amino acids, proteins, vitamins, oleic acid, antioxidants, and other anti-inflammatory substances (4, 12, 13). Due to reducing of the production of the reactive oxygen species (ROS) and reactive nitrogen species (RNS), consequently lowering disorder metabolites [superoxide ions ($*O_2^-$), hydroxyl (OH⁻), hydrogen peroxide (H₂O₂), nitrate oxide (NO), etc.], which are linked with pro-inflammatory several metabolites [interleukin-1, -1β ,-6,-8,-12 (IL-1, -1β ,-6,-8,-12), Toll-like receptor 4 (TLR4), lipopolysaccharides (LPS, microbial component), hepatic nuclear factor-kappa B (NF-kB), tumor necrosis factor-alpha (TNF- α), inducible nitric oxide synthase (INOS), cyclooxygenase-2 (COX-2), lipoxygenase (LOX), cytochrome P450, nitric oxide, G protein-coupled receptor 120, etc.] potentially associated with oxidative stress and metabolic diseases due to high damage of cellular protein, lipids, and DNA (9, 14, 15).

Nevertheless, it is remarkable that food frying (pan and deep) using sunflower oil rich in linoleic fatty acid reduces its oxidation and thermal stability, in which various hazardous substances to human health form (16). To reverse this scenario, blending cold-pressed oils rich in natural antioxidants or their natural antioxidants [thymoquinone and tocopherols from black cumin (*Nigella sativa*)] with sunflower oil is highly recommended for oils stability and for consumers' health improvement (16–19).

In addition, vegetable oils are a source of macroelements such as sodium (Na), potassium (K), calcium (Ca), magnesium (Mg), phosphorus (P), and microelements such as iron (Fe), selenium (Se), manganese (Mn), chromium (Cr), zinc (Zn), aluminum (Al), barium (Ba), strontium (Sr), tin (Sn), copper (Cu), cobalt (Co), and thallium (Ti), which are essential or toxic for human health when consumed in large quantities (20, 21). In addition, some chemical elements, such as arsenic (As), cadmium (Cd) nickel (Ni), lead (Pb), mercury (Hg), and Cu are found in edible vegetable oils, which are toxic (22-24) and carcinogenic for consumers even at a low amount (25). Moreover, the presence of Ca, Co, Mg, Fe, Zn, Cu, Mn, Sn, and Ni accelerate vegetable oils' oxidation affecting their flavor, freshness, storability, and toxicity (22, 26). Therefore, cold pressing and filtration are the alternatives of the several methods to obtain healthy edible vegetable oils associated with good fatty acids and antioxidants composition, and lower amounts of hazardous chemical elements (27-29).

Interestingly, it is increasing the consumption of the unrefined edible vegetable oils, including sunflower oils obtained by cold pressed due to the high amount of natural antioxidants (tocopherols, phytosterols, carotenoids, etc.), waxes, while presenting low content of free fatty acids and phospholipids (30, 31). In addition, some studies have reported nutritional qualities

correlated with physicochemical properties, trace elements, fatty acid composition, and essential components in sunflower oil obtained by cold pressing (32). Therefore, a domestic cold-press machine is mostly recommended to obtain healthy edible vegetable oils, due to their higher amount presence of antioxidants and fatty acids, moreover, with a lower amount of heavy metals contaminants (20).

Although some studies have been carried out using different types of oils and extraction conditions, there is scarce information in the literature on the quality and mineral composition of sunflower oil extracted by a domestic cold-press machine, as well as risk assessment for human health due to the ingestion of this type of oil containing metals.

Motivated by the manuscript published by Melo et al. (20), which demonstrated that cold-extracted oils maintain their quality and chemical composition, this study aimed to evaluate for the first time the fatty acids composition, physicochemical and optical properties, thermal and oxidative stability of sunflower oil samples cold-press extracted using a domestic machine. In addition, the chemical elements Mg, Cr, Mn, Fe, Co, Ni, Cu, Zn, Cd, Al, Pb, As, and Se in sunflower oils were quantified using inductively coupled plasma optical emission spectroscopy (ICP OES) and the results were compared with DRI/AI* and FAO/WHO parameters. The fatty acid composition, physicochemical and optical properties, and thermal and oxidative qualities evaluated in this oil were compared with previous results in the literature.

Materials and methods

Sunflower seed, oil preparation, seed and oil moisture, and lipid quantification

Sunflower seeds were obtained from 10 farms in Campo Grande, Mato Grosso do Sul state, Brazil, in September 2020. Seeds were mixed and immediately dried in an air circulation oven at 40°C for 48 h. We collected crude oil from dried hull seeds using a domestic cold-press machine extractor equipped with a stainless continuous screw and drainage hole with an internal filter (Yoda Nut & Seed Cold Press Oil), Extractor-Gourmet Extractor, oil Natural, Homeup, (Yoda Europe, Cluj-Napoca, Romania). Immediately, the oil was placed into an amber and hermetic glass bottle and then used for analysis.

Moisture content was measured using milled seeds (1.0 g) subjecting in the over at 105° C for 60 min, then the sample was rested in a desiccator, until achieving constant weight. For the filtered sunflower oil (1.4 g), relative humidity was determined

by the Karl Fischer technique (KEM MKC-610 Karl Fischer Moisture Titrator, Japan).

Free fatty acids (FFAs) were extracted using a Soxhlet extractor with petroleum ether as solvent at 60° C for 6 h (33).

Methylation and fatty acid profile

Fatty acid methyl esters (FAMEs) of the sunflower oil were obtained at ambient temperature. The samples (157 mg) were weighed into assay tubes, saponified with methanolic NaOH 0.5 N (4 ml), esterified with a mixture of H_2SO_4 and NH_4Cl in methanol (5 ml), then saturated with NaCl (4 ml), and finally dissolved in hexane (5 ml) (34).

FAMEs were analyzed using a gas chromatograph (model CP-3800, Varian, Santa Clara, CA, USA) equipped with a flame ionization detector, a split/splitless injector, and a stationary phase fused silica capillary column of polyethylene glycol (CARBOWAX 20 M, length 30 m × 0.25 mm, Quadrex, Santa Clara, CA, USA). Operational parameters were followed for chromatography: the injector and detector temperatures were 250°C. The column temperature was programmed to 80°C for 2 min, followed by a ramp of 4°C /min up to 220°C and kept for 13 min; hydrogen carrier gas with 1 mL min⁻¹ flow and injection volume 1 µL. Retention times were compared with the respective methyl ester standards (Supelco, F.A.M.E. mix C4:0 to C24:0, Sigma-Aldrich, Darmstadt, DA, Germany) (35).

Fatty acids nutritional quality indices

The nutritional quality of sunflower oil was evaluated according to its fatty acids composition assessed by three following indices

Atherogenicity index (AI) (36).

$$AI = \frac{C12:0 + (4 \times C14:0) + C16:0}{\sum MUFA + \sum PUFA}$$
(1)

Thrombogenicity index (TI) (36)

$$TI = \frac{C14:0 + C16:0 + C18:0}{(0.5 \times \sum MUFA) + (0.5 \times \sum \omega 6) + (3 \times \sum \omega 3)}$$
(2)

Fatty acids hypocholesterolemic/hypercholesterolemic (HH) ratio (37)

$$HH = \frac{C18:1\omega9 + C18:2\omega6 + C20:4\omega6 + C18:3\omega3 + C20:5\omega3 + C22:5\omega3 + C22:6\omega3}{C14:0 + C16:0}$$
(3)

Identity and quality characteristics of sunflower oil

Sunflower characterization conducted oil was according to the American Chemist's Society Oil (38), in triplicate, for qualification parameters: acidity (Ca 5a-40) and peroxide indices (Cd 8-53); identity relative density (Cc iodineparameters: 10a-25), Wijis method (d 1-25), and saponification values (Cd 3-25).

Determination of oxidative stability

Sunflower oil oxidative stability was determined by the Rancimat method (873 Metrohm Co, Basel, Switzerland) by accelerated oxidation according to the European Union standardized standard EN 14112 (39). The analyses were conducted subjecting 3.0 g of oil at a constant temperature of 110° C under an airflow rate of 10 L h⁻¹ through the samples, and then into a measuring vessel containing 50 ml ultrapure water Mill-Q in which the conductivity generated by volatile products during the oil decomposition was measured as a function of time.

Thermal analyses: Thermogravimetric analysis/derivative thermogravimetry, and differential scanning calorimetry

Thermogravimetric analysis (TGA)/derivative thermogravimetry (DTG) curves were obtained using TGA-Q50 (TA Instruments, New Castle, DE, USA). Samples of sunflower oil (6.0 mg) were added into a platinum pan from 10 to 550°C at a heating rate of 2° C min⁻¹ under inert nitrogen and synthetic air atmosphere gases at a flow rate of 60 ml min⁻¹. In addition, DSC curves were conducted with DSC-Q20 equipment with RCS90 coupled with a cooling system (TA Instruments). The DSC curves were obtained in a calorimeter model DSC-Q20 coupled with an RCS90 refrigeration system (TA Instruments). Approximately 3.0 mg of sunflower oil was used for analysis using aluminum crucibles (Tzero standard) as support and reference, at a heating rate of 10°C min⁻¹, heating cycle followed by cooling at temperatures between -80° C and 25° C, under an inert nitrogen atmosphere with a flow rate of 60 ml min⁻¹.

Curves TGA/DTG obtained were from and DSC data. which were generated by Universal Analyzes 2000 software 3.7A version (TA Instruments).

Optical molecular analyses: UV-Visible absorption and fluorescence spectroscopy

Sunflower oil was diluted in HPLC grade hexane (spectroscopic grade 99.9%) at concentration а 10^{-3} $g L^{-1}$. of $1 \times$ UV-Visible absorption measurements were made using a Lambda 265 UV/Vis Waltham, spectrophotometer (Perkin Elmer, MA. USA), and the spectra were collected in the 200 to 800 nm range.

Excitation-emission matrix fluorescence maps were obtained using a spectrofluorometer (FluoroMate FS-2, SCINCO). The excitation-emission maps were obtained by exciting the samples between 240 and 450 nm in steps of 5 nm and collecting the emission from 250 to 750 nm in 1 nm steps.

For the UV/Vis absorption and fluorescence measurements, the diluted sunflower oils were placed into a four-polished-sided quartz cuvette with a 10 mm optical path.

Extraction induced by emulsion breaking procedure and trace elements quantification

The extraction induced by emulsion breaking was performed according to Carneiro et al. (40) with adaptations. A falcon tube containing 3.0 ml of crude sunflower oil and 3.0 ml of ethanol was shaken for 20 s in a vertex shaker, then 3.0 ml of ultrapure water (conductivity 18.2 M Ω cm, Millipore, Biocel, Germany), 0.76 ml of Triton x-100, and 3.0 ml of HNO₃ were added, and then mixed with a vortex mixer for 20 min.

The concentration of Al, Cr, Cu, Fe, Mn, and Zn in sunflower oil samples was obtained using an ICP OES with an Axial Plasma (iCAP 6000 Series, Thermo Scientific, Cambridge, UK). Standard solutions were prepared by diluting a standard multielement stock solution (SpecSol, Quinlab, Jacarei, SP, Brazil) containing 1,000 mg L⁻¹ of each element. Nine concentrations were used to build calibration curves for the quantitative analyses of oil. The concentration for the elements was 0.01-5.0 mg L⁻¹ range. The setup of ICP OES instrumental conditions for elemental analyses was the same used by Melo et al. (20). Table 1 summarizes the operational condition used in the ICP OES apparatus analysis as wavelength, limit of detection (LOD), limit of quantification (LOQ), and correlation coefficient (R²) in the current study.

One blank and nine calibration curves were generated using the following concentrations: 0.005, 0.01, 0.025, 0.05, 0.1, 0.25, 0.5, 1.0, and 2.0 mg kg⁻¹ of each element standard.

An addition/recovery test for the elements under study was carried out in a sunflower oil sample by spiking 0.5 mg L^{-1} of each analyte. The method had a recovery interval of 80–110%

Elements	Wavelength (nm)	LOD (mg/kg)	LOQ (mg/Kg)	Correlation (R ²)		
Al	396.152	0.0114	0.0380	0.9995		
As	193.696	0.0048	0.0159	0.9996		
Cd	228.802	0.0007	0.0024	0.9994		
Со	238.892	0.0016	0.0054	0.9995		
Cr	425.435	0.0015	0.0049	0.9994		
Cu	327.396	0.0014	0.0048	0.9995		
Fe	259.940	0.0193	0.0644	0.9991		
Mg	285.213	0.0675	0.2251	0.9984		
Mn	257.610	0.0004	0.0012	0.9996		
Ni	221.647	0.0023	0.0077	0.9994		
Pb	220.353	0.0070	0.0233	0.9996		
Se	196.026	0.0080	0.0267	0.9997		
Zn	213.856	0.0018	0.0060	0.9991		

TABLE 1 The ICP OES operating conditions for analysis.

for the spike of 0.5 mg L^{-1} , to the established limit proposed by the Association of Official Analytical Chemists (41).

Human health risk assessment

The concentration of the chemical elements in sunflower oil was compared with the FAO/WHO recommended intake standards and hazards quotient. The non-carcinogenic was calculated according to the equation adopted by Machate et al. (42). Cancer risk is the probability of an individual developing any cancer type over a lifetime due to a specific exposure to a hazardous mineral. The chronic daily intake dose (CDI) of carcinogenic elements (mg/kg/day) and slope factor (SF) of Cr is 0.5 mg/kg/day, according to Equation (4).

$$Cancer Risk = CDI \times SF$$
(4)

Cancer risk is a sum of individual chemical elements in different exposure pathways to develop cancer in a person, which is the total cancer risk (CR). According to US UPA (43), acceptable values of cancer risk range from 10^{-6} to 10^{-4} , while values > 10^{-4} are considered inadmissible.

The human health risk of trace element consumption dose was calculated on the chronic daily intake dose (CDI, mg/kg/day) for a chemical contaminant element in the sunflower oil intake quantity CDI calculated according to Equation (5).

$$CDI_{oil} = \frac{C \times IR \times EF \times ED}{BW \times AT}$$
(5)

where CDI_{oil} – chronic daily oil intake dose; C – chemical element concentration in the crude sunflower oil sample (mg kg⁻¹); IR – ingestion rate g/day; EF – exposure frequency

(365 days/year); ED – exposure duration; BW – body weight (kg), estimated 26, 62, and 70 kg for 8, 18, and 30 years old, respectively. AT – average time (ED \times 365 days/year).

The risk to human health by the intake of trace element contaminated food was estimated using a hazard quotient (HQ), which is a ratio of CDI and chronic oral reference dose (RfD), determined by the following Equation (6):

$$HQ = \frac{CDI}{RfD}$$
(6)

The RfD values were previously established by the Joint Food and Agriculture Organization/World Health Organization Expert Committee on Food Additive (44) and the United States Environmental Protection Agency (45). The RfD (mg/kg/day) values are: Al = 1.0; Cr = 0.003; Cu = 0.04; Fe = 0.7; Mn = 0.14; and Zn = 0.3 (46). As shown in Equation (6), hazard quotient toxic risk on each trace element and their sum, Equation (7) (total non-carcinogenic hazard index) HI < 1, safe food consumption, while HI > 1, health risk food consumption.

$$HI = HQ_{Al} + HQ_{Cr} + HQ_{Cu} + HQ_{Fe} + HQ_{Mn} + HQ_{Zn}$$
(7)

Results and discussion

Oil preparation, sunflower seed, oil moisture, and lipid quantity

Sunflower seed yielded 260 mg g^{-1} (26%) of oil, lower than industrially obtained using petroleum solvents (36–50%) (47). However, oil extracted by cold-press yielded a higher amount of beneficial nutritional components such as tocopherols, phytosterols, terpenoids, phenolic acids, carotenoids, chlorophylls, antioxidants, waxes (C36–C48),

TABLE 2 Fatty acid composition of sunflower oil extracted by cold pressing.

Fatty acids	Mean ± Standard deviation (%)
Myristic (C14:0)	0.03 ± 0.00
Palmitic (C16:0)	4.13 ± 0.05
Palmitoleic (C16:1)	0.04 ± 0.00
Margaric (C17:0)	0.04 ± 0.00
Heptadecenoic (C17:1)	0.02 ± 0.00
Stearic (C18:0)	3.13 ± 0.08
Oleic (C18:1n9c)	37.29 ± 0.24
Linoleic (C18:2n6c)	54.00 ± 0.39
α-Linolenic (C18:3n3c)	0.05 ± 0.00
Araquidic (C20:0)	0.20 ± 0.00
Gondoic (C20:1)	0.13 ± 0.00
Eicosapentanoic (C20:5n3c)	0.06 ± 0.00
Behenic (C22:0)	0.53 ± 0.01
Docosadienoic (C22:2)	0.08 ± 0.00
Tricosylic (C23:0)	0.08 ± 0.00
Lignoceric (C24:0)	0.17 ± 0.01
\varSigma SFAs	8.33
\varSigma MUFAs	37.48
\varSigma PUFAs	54.19
\varSigma UFAs	91.67
Total FAs	100
Atherogenic index	0.05
Thrombogenic index	0.16
Hypocholesterolemic/hypercholesterolemic	21.97

 \varSigma SFAs, sum of saturated fatty acids; \varSigma MUFAs, sum of monounsaturated fatty acids; \varSigma PUFAs, sum of polyunsaturated fatty acids; FAs, fatty acids; ND, non-detectable; defined as <0.05%.

oleic and linoleic fatty acids, phospholipids, unsaponifiable matters, and others; in contrast, the hazardous components (trace metals) were extracted in lower amounts (29, 48, 49). The cold-pressing technique is widely recommended because it provides oils with healthy components, including their by-products (oil cake) (32), of which adequate consumption is correlated with regulating metabolic diseases (12).

The moisture determination in sunflower seeds yielded 72.76 mg g⁻¹ (7.28%) of dehydrated water, whereas dry matter was 927.24 mg g⁻¹ (92.72%), better for fatty acid and essential components integrity, which are relevant for nutrition functionality (50). Sunflower oil moisture was 998 μ g, (0.07%) of evaporated water, demonstrating the importance of this oil in food system quality, due to less susceptibility to oxidative stress and lipid oxidation by the action of prooxidants substances (51). Refined flour of sunflower seed (1.0 g) yielded 493.82 mg g⁻¹ (49.38%) of FFAs, demonstrating its relevance for nutritional value and a good source of vegetable oil (29).

TABLE 3 Physicochemical characteristics of sunflower oil compared with Codex Alimentarius parameters.

Parameters	Sunflower oil	Maximum values			
Peroxide index (mEq/kg)	16.61 ± 0.2	≤20 (55) (a)			
Iodine index (g $I_2/100$ g)	131.53 ± 0.1	118-141 (56) (b)			
Acidity index (mg KOH/g)	0.54 ± 0.0	4.0 (56) (a)			
Saponification index (mg KOH/g)	141.80 ± 1.92	184-196 (56) (b)			
Relative density (20 $^{\circ}$ C)	0.92 ± 0.1	0.92-0.93 (56) (c)			

(a) Parameter for cold-pressed and virgin oils; (b) parameters for refined sunflower oils; (c) parameter for crude sunflower oil.

Fatty acid profile

Table 2 presents the composition and quantity of fatty acids yielded in this study, with profiles in decreased order: linoleic (54.00%) > oleic (37.29%) > palmitic (4.13%) > stearic (3.17%) > behenic (0.55%) > araquidic (0.20%) > lignoceric (0.17%) > gondoic (0.13%) > docosadienoic = tricosylic (0.08%) > eicosapentanoic (0.06) > α -linolenic (0.05%) > palmitoleic = margaric (0.04%) > meristic (0.03%). In this study, most quantified fatty acids, linoleic/oleic ratios were proportionally shown at 1.44:1, better than oils extracted by n-hexane as solvent 3.24:1 (32, 52).

According to our results, the sunflower oil extracted using a domestic cold press machine yielded a higher amount of oleic acid and the highest hypocholesterolemic (HH) ratio (Table 2) compared with one that applied n-hexane (oleic, 19.81%; linoleic, 64.35%; HH, 0.16) (32), massively used by industry to obtain edible vegetable oils.

Therefore, long-term diets of vegetable oils rich in linoleic acid and the lowest hypocholesterolemic index are associated with the prevalence and incidence of several metabolic diseases (diabetes mellitus, coronary and inflammatory diseases, cancer, and obesity) (4, 6, 53). Moreover, in the human body, linoleic acid is converted into arachidonic acid (n-6 PUFAs), which is the pro-inflammatory precursor of prostaglandin and leukotriene synthesis at the cyclooxygenase (COX-2), lipoxygenase (LOX), and cytokines (TNF- α , IL-1, IL-1 β , IL-6, IL-8, IL-12, NF-kB, NO, LTs cytochrome P450, protein-coupled receptor 120), which compete with n-3 PUFAs enzymes during the biosynthesis pathway of long- (LC-PUFAs) and very-long-chain fatty acids (VLC-FAs) associated with anti-inflammatory effects (9, 14, 15).

Fatty acid nutritional quality indices and characteristics of sunflower oil

Table 2depictsnutritionalqualityindices:atherogenicityindex(AI),thrombogenicityindex(TI),andhypocholesterolemic/hypercholesterolemic(HH)



ratio calculated as 0.05, 0.16, and 21.97, respectively. Sunflower oil obtained by the cold-press machine presents better values associated with regulating several metabolic diseases for consumers than another extracted by petroleum solvent (54).

Table 3 presents physicochemical profiles revealing that this sunflower oil is suitable for human consumption, and its averages can be associated with oxidative stability, authenticity, quality, and identity (57). The iodine index (131.53) was found between the Codex Alimentarius parameters, which correspond to unsaturated fatty acids (91.67%). Moreover, this oil is characterized by the lowest acidity and peroxide indices, demonstrating its lipid stability against rancidity due to the lowest autoxidation products (ketones, aldehydes, hydroxyl alkenals, and dienes) formation which became off-flavor and toxic food for consumers associated with pro-oxidants action (processing manner, oxygen, heat, light, and metals) (9, 58). In addition, this oil presented a lower saponification index (141.80 mg KOH/g oil) compared with parameters and others obtained using petroleum solvent (188 and 189 mg KOH/g oil) (59), represented by the long-chain fatty acids (palmitic, stearic, oleic and linoleic acids), which can be used to identify these oils.

Relative density is the relevant parameter correlated with edible vegetable oil absorption and mass transfer rates during cooling or melting, better to lower values than the parameters (60). Sunflower oil obtained using a domestic machine presented a lower average (0.91) compared with that extracted using solvents (0.92–0.96) (61, 62). Although, these values represent a small difference among them, however, other parameters above referred can be used to identify the authenticity and origin of these oils.

Determination of oxidative stability

Rancimat data revealed that the crude sunflower oil (unsaturation 91.67%) presented an induction period (IP) of 5.06 h (Figure 1). This behavior can be attributed to natural antioxidants in this oil as well observed to refined sunflower oil (unsaturation 88.40%), which the IP shifted from 5.5 to 7.5 h, respectively, in control and sunflower oil added polyphenol, subjected under a temperature of 110°C with airflow 20 L h⁻¹ (63). Furthermore, the unsaturation amount of fatty acids is another relevant characteristic that is inversely proportional to oxidative stability. Sunflower hybrid oils (H) which presented higher iodine value (IV) showed lower IP, for instance, e.g., H19: (IV = 127, IP = 3.32 h) has lower IP compared with H21 (IV = 81, IP = 9.55 h) (64). Moreover, among unsaturation fatty acids, cold-pressed sunflower oil (SO) that presented a higher amount of oleic than linoleic fatty acids was more oxidative stably H20 (55), as well as SO1 (oleic, 86.52% and linoleic, 5.49%, IP = 19.87 h), while SO2 (oleic, 18.52% and linoleic, 66.02%, IP = 6.42 h) (65).

Therefore, to increase the stability of sunflower oil (rich in linoleic acid), it is recommended to blend it with oil rich in stable antioxidants such as tocopherols and thymoquinone or apply their natural antioxidants (17, 18).

Thermogravimetric analysis/derivative thermogravimetriy, and differential scanning calorimetry

Thermogravimetric analysis/derivative thermogravimetry curves of crude sunflower oil in the presence of synthetic air and nitrogen atmospheres are shown in Figure 2. The TGA/DTG curves of sunflower oil submitted under a synthetic air atmosphere show three steps of thermal decomposition that are illustrated in Figure 2A and summarized in Table 4. The first step can be attributed to moisture linked by natural antioxidants (polyphenols, carotenoids, stanols, and vitamins A and C), dimers, trimmers, polymers, PUFAs, and other compounds formed from PUFAs (54.19%), represented by linoleic acid (54.00%). In addition, this process can be influenced by Zn (6,6228 mg/kg) and Fe (1.6637 mg/kg), both metals that negatively influence on oxidation stability (22, 26). The first stage of thermal stability is the most important, as it demonstrates that this cold-pressed sunflower oil can be heated up to 125°C without undergoing oxidative degradation.

The second step of mass loss is attributed to products formed in the first step plus MUFAs (37.48%) decomposition represented by oleic fatty acid (37.29%), whose double bonds of MUFAs are broken and become SFAs of the oil. The third step of mass decomposition is attributed to the above fatty acids plus SFAs (7.26%) (palmitic (4.13%), stearic acids (3.13%), and



waxes (C36–C48), of which the majority are represented by C36, C37, C40, and C41, as previously reported in cold-pressed sunflower oil (49). The minor last mass loss is attributed to carbonaceous residue substances. This behavior was reported in commercial sunflower oil, although with more thermal stability compared with the current study (66–68). Moreover, cold press and unrefined oil present more oleic than refined one (10), natural antioxidants (69) and minerals, which are healthy and regulators of several metabolic diseases (9, 68, 70), and economic and ecological benefits from food by-products (71) than refined ones. Moreover, synthetic antioxidants, butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), tert-butylhydroquinone (TBHQ), and others (10, 72)

replace natural antioxidants in industrial edible vegetable oils, which are associated with the prevalence of metabolic diseases (cellular damage, cancers) and environmental contamination (73, 74).

In the nitrogen atmosphere, the oil showed one high step mass decomposition justified by a higher amount of unsaturated fatty acids (oleic and linoleic fatty acids) compared with SFAs. The minor last mass loss is attributed to formed carbonaceous residual substances.

In Figure 2B, DSC curves show two exothermic crystallization peaks, which the first attributed to SFAs and MUFAs, with T_{onset} at $-16.57^{\circ}C$ and enthalpy peak at 1.979 J/g, followed by a second peak representing PUFAs observed at

Sample	Atmosphere	Steps	Temperat	ure range (°C)	Event mass loss (%)	Residual mass (%)		
			Initial	Final				
Sunflower oil	Synthetic air	1 st	126.16	254.34	15.48	0.21		
		2 nd	254.34	392.17	49.74			
		3 rd	254.34	447.18	14.57			
	Nitrogen	1 st	236.19	465.90	99.09	0.33		

TABLE 4 TGA/DTG curves of sunflower oil obtained under synthetic air and nitrogen atmospheres.



 $-34.52^{\circ}\mathrm{C}$, with enthalpy peak at 1.466 J/g. Moreover, heating the sunflower oil, two peaks were also observed, the first peak corresponding to MUFAs, whose $\mathrm{T}_{\mathrm{onset}}$ was observed at $-36.27^{\circ}\mathrm{C}$ and enthalpy at 1.866 J/g. The $\mathrm{T}_{\mathrm{onset}}$ of the second peak appeared at $-28.07^{\circ}\mathrm{C}$ and the enthalpy peak at 24.59 J/g.

In the current study, DSC results were observed in lower temperatures than commercial sunflower oil due to their fatty acids and natural antioxidant composition, which influence its thermal oxidative levels (75). The TGA and DSC analyses are used to qualify, authenticate, and recognize vegetable oils, thus avoiding their adulteration, falsification, and consumption of improper products.

Optical molecular analyses: UV-Visible absorption and fluorescence spectroscopy

Figure 3A illustrates sunflower oil UV-Vis absorption spectrum with two major absorbance regions one in the 223– 236 nm range and the other from 257 to 452 nm (inset of Figure 3A). The first absorption band can be attributed to phytocholesterols (phytostanols), phytosterols, and tocopherols (vitamin E). The second region corresponds to carotenoids and fatty acids (linoleic, oleic acids) (6, 76). The consumption of oils rich in phytocholesterols, carotenoids, tocopherols and phytosterols, polyphenols, and unsaturated fatty acids are widely correlated with avoiding and regulating several metabolic diseases (3, 9).

The UV-Vis is used to monitor oxidation, identity, and authenticity, as the absorption in the 400–520 nm range appears higher in adulterated sunflower oil (77).

Figure 3B shows the excitation–emission map of sunflower oil exhibiting two intense bands. The first region presents emissions in the 297–327 nm range when excited between 290 and 310 nm. The second region of emission is observed between 360 and 445 nm due to excitation in the 360–400 nm range. These fluorescence bands can be correlated with the presence of vitamin E (tocopherols and tocotrienols), carotenoids, chlorophyll, and unsaturated fatty acids in sunflower oil (78).

The fluorescence emission wavelength ranging from 400 to 500 is used to identify, qualify, and authenticate original and adulterated vegetable edible oils due to the oxidation of fatty acids products and tocopherols (79).

TABLE 5 Trace elements in sunflower oil quantified by ICP OES (mg/kg \pm SD) compared with nutritional recommendations for adult, pregnancy, lactation, and children by RDA/AI and FAO/WHO.

Trace elements	Concentration (mg/kg)	FAO/WHO (mg/kg)	Dietary reference intakes (DRI) and adequate intake (AI*) (mg/day) (80)											
			Children	n Males		Females		Pregnancy		Lactation				
			8 y. old	18 y. old	30 y. old	18 y. old	30 y. old	18 y. old	30 y. old	18 y. old	30 y. old			
Cr	0.0242 ± 0.0049	0.002 (81)	0.015*	0.035*	0.035*	0.024*	0.025*	0.029*	0.030*	0.044*	0.045*			
Mn	0.2372 ± 0.0051	3.00 (82)	1.5*	2.2*	2.3*	1.6*	1.8	2.0*	2.0*	2.6*	2.6*			
Fe	1.6637 ± 0.0393	14.00 (82)	10	11	8	15	18	27	27	10	9			
Cu	0.2791 ± 0.0094	0.90 (82)	0.44	0.89	0.9	0.89	0.9	1	1	1.3	1.3			
Zn	6.6228 ± 0.0788	15.00 (82)	5	11	11	9	8	12	11	13	12			
Al	1.0401 ± 0.0199	5.00 (83)	ND	ND	ND	ND	ND	ND	ND	ND	ND			

Elements Mg, Co, Ni, Cd, Pb, As, and Se < LOD; NE, not established; ND, not determined; *The value for AI is used when there are no calculated values for the RDA.

Trace elements concentration in sunflower oil

Table 5 summarizes the data of six trace elements quantified in sunflower oil compared with Codex Alimentarius contents for oils and dietary reference intakes (DRIs).

Beyond the endogenous biological processes, plants acquire minerals from the soil, besides exogenous processes (environment pollution by industries, transports, mechanization, fertilizers, and pesticides used in agriculture), which influence raw material for edible plant production (42). Moreover, minerals can contaminate edible vegetable oils during production, refining, and storing processes.

Minerals (Ag, As, Be, Ca, Cu, Zn, Fe, Mg, Mn, Cd, Co, Na, K, Ni, Pb, and V) are used to monitor and qualify edible vegetable oils: corn (*Zea mays*), hazelnut (*Corylus avellana*), olive (*Olea europaea*), and sunflower in Turkey (84), pequi (*Caryocar brasiliense*), primrose (*Oenothera biennis*), avocado (*Persea americana*), coconut (*Cocos nucifera*), grape seed (*Vitis vinifera*), babassu (*Attalea speciosa*), and licuri (*Syagrus coronata*) in Brazil (40), and several varieties in China (26) and Iran (24).

Mineral concentrations can be used as a relevant fingerprint to distinguish the provenance, originality, and adulterated edible vegetable oils, promoting health benefits. The average concentrations of trace elements quantified in sunflower oil in decreased order are of Zn (6.6228 mg/kg) > Fe (1.6637 mg/kg) > Al (1.0401 mg/kg) > Cu (0.2791 mg/kg) > Mn (0.2372 mg/kg) > Cr (0.0242 mg/kg). Only Cr concentrations exceeded 1,210% of FAO/WHO limits, which can be influenced by the vehicle fumes, fertilizers, and pesticides used, as reported in the intensive agriculture modern farming, as well as the cultivated areas located near the road with high vehicle traffic (42, 85). In contrast, Zn, Fe, Al, Cu, and Mn concentrations were lower than FAO/WHO and DRI/AI* limits. Moreover, Cr and Zn concentrations were within DRI/AI* limits, while Cu, Fe, and Mn concentrations were quantified lower than DRI/AI* limits (Table 5).

Furthermore, Al concentration was higher than quantified in industrial refined oils of avocado, primrose, babassu, licuri, pequi, olive, and grape seeds $(0.04-0.52 \text{ mg kg}^{-1})$ (32). In contrast, Zn, Cu, and Mn amounts were reported between those found in commercially refined oils of sunflower, olive, canola, soybean, corn, and hazelnut corresponding to 1.03-9.54, 0.05-4.504, and $0.04-1.76 \text{ mg kg}^{-1}$, respectively. The concentrations of Cr and Fe were lower than reported in sunflower, olive, canola, soybean, corn, and hazelnut oils corresponding to 0.0126-7.106and $7.78-28.93 \text{ mg kg}^{-1}$, respectively (24, 84, 86).

Thus, in light of the acceptable concentrations of trace elements regarding referential parameters, the consumption of the sunflower oil herein studied can be beneficial, because Cr, Cu, Fe, Mn, and Zn play essential physiological roles in defense response, protein construction, enzymatic reactions, signaling pathways, regulation of oxidative stress and metabolic diseases, and others (87). However, lower Al concentrations are associated with reduced occurrence of metabolic diseases and dysfunctions, such as cancers, Alzheimer's and Parkinson's diseases, inhibited enzymatic, cytotoxic and neurotoxic reactions, gut imbalance, skeletal disorders, and others (42).

Health risk assessment

Most studies reported daily consumption of vegetable oils from 25 to 30 g kg⁻¹ (26, 88). Other studies demonstrated that quantities of cooking oils depend on the type of dishes: pure vegetables range from 9 to 167 g kg⁻¹, with a mean of 56 g kg⁻¹, pure meat from 4 to 353 g kg⁻¹ (142 g kg⁻¹), and mixed meat-vegetable 7 to 394 g kg⁻¹ (110 g kg⁻¹) (89).

However, the calculated quantities of the sunflower oil daily intake $(g kg^{-1})$ in comparison with other studies based on the concentration of trace elements regarding acceptable

TABLE 6 Non-carcinogenic risk (CR), hazard quotient (HQ), and total non-carcinogenic hazard index (HI) of trace elements on ingestion rate (IR g/kg) of sunflower oil obtained by cold-press (Brazil and Romania) and commercially available (China and Turkey).

Study	Years old	IR (g/day)	Index							Trace	elements							HI
				Со	Cr	Cu	Fe	Mn	Ni	Pb	Se	Zn	Li	Мо	Cd	As	Al	
ly)	8	0.61	CR	-	$2.8 imes 10^{-4}$	-	-	-	-	-	-	-	-	-	-	-	-	
stud			HQ	0	0.18926	0.1637	0.05576	0.03975	0	0	0	0.51794	-	-	0	0	0.0244	0.99
rrent	18	1.46	CR	-	2.9×10^{-4}	-	-	-	-	-	-	-	-	-	-	-	-	
(Cu			HQ	0	0.18996	0.16431	0.05597	0.0399	0	0	0	0.51985	-	-	0	0	0.02449	0.99
razil	30	1.65	CR	-	$2.9 imes 10^{-4}$	-	-	-	-	-	-	-	-	-	-	-	-	
<u>е</u>			HQ	0	0.19014	0.16447	0.05602	0.03994	0	0	0	0.52036	-	-	0	0	0.02452	0.99
	8	0.037	CR	-	$3.6 imes 10^{-4}$	-	-	-	-	-	-	-	-	-	-	-	-	
32)			HQ	-	0.2372	0.007	0.00002	0.00793	0.01352	-	0.333	0.00043	0.14231	0.25615	-	-	-	0.99
nia (18	0.08	CR	-	$3.2 imes 10^{-4}$	-	-	-	-	-	-	-	-	-	-	-	-	
Roma			HQ	-	0.21505	0.00677	0.00002	0.00719	0.01226	-	0.30194	0.00039	0.12903	0.23226	-	-	-	0.99
	30	0.099	CR	-	$3.5 imes 10^{-4}$	-	-	-	-	-	-	-	-	-	-	-	-	
			HQ	-	0.23571	0.00743	0.00002	0.00788	0.01344	-	0.33094	0.00042	0.14143	0.25457	-	-	-	0.99
	8	0.419	CR	-	-	-	-	-	-	$1.4 imes 10^{-6}$	-	-	-	-	2.1×10^{-5}	$2.7 imes 10^{-5}$	-	-
(9			HQ	-	-	0.02659	0.67224	0.05122	0.0274	0.04029	-	0.06607	-	-	0.056565	0.05909		0.99
1a (2	18	0.99	CR	-	-	-	-	-	-	1.4×10^{-6}	-	-	-	-	2.1×10^{-5}	2.6×10^{-5}	-	
Chir			HQ	-	-	0.02635	0.66608	0.05075	0.02715	0.03992	-	0.06547	-	-	0.05605	0.05855	-	0.99
	30	1.128	CR	-	-					1.4×10^{-6}	-	-	-	-	2.1×10^{-5}	2.7×10^{-5}	-	
			HQ	-	-	0.02659	0.67220	0.05122	0.02739	0.04029	-		-	-	0.05656	0.05909	-	0.99
	8	0.0093	CR	-	-	-	-	-	-	$3 imes 10^{-5}$	-	-	-	-	5.1×10^{-7}	-	-	
4)			HQ	0.0065	-	0.00099	0.05435	0.03097	-	0.90323	-	0.00132	-	-	0.00134	-	-	0.99
ey (8	18	0.0224	CR	-	-	-	-	-	-	$3.1 imes 10^{-5}$	-	-	-	-	$5.2 imes 10^{-7}$	-	-	
Furk			HQ	0.10074	-	0.01539	0.8419	0.0048	-	0.01399	-	0.00133	-	-	0.000000021	-	-	0.99
[30	0.0253	CR	-	-	-	-	-	-	$3.1 imes 10^{-5}$	-	-	-	-	$5.2 imes 10^{-7}$	-	-	
			HQ	0.00651	-	0.0099	0.05437	0.03098	-	0.90357	-	0.02043	-	-	0.001358971	-	-	0.99
	8	0.0097	CR	-	$1.3 imes 10^{-3}$	-	-	-	-	$3.2 imes 10^{-7}$	-	-	-	-	$8.1 imes 10^{-6}$	-	-	
(9)			HQ	-	0.88369	0.02766	0.00475	-	0.03231	0.00933	-	0.0079	-	-	0.02127	-	-	0.99
ey (8	18	0.0234	CR	-	$1.3 imes 10^{-3}$	-	-	-	-	$3.2 imes 10^{-7}$	-	-	-	-	$8.2 imes 10^{-6}$	-	-	
Turk			HQ	-	0.89398	0.02799	0.00481	-	0.03268	0.00944	-	0.00799	-	-	0.02151	-	-	0.99
-	30	0.0264	CR	-	$1.3 imes 10^{-3}$	-	-	-	-	$3.2 imes 10^{-7}$	-	-	-	-	8.2×10^{-6}	-	-	
			HQ	-	0.89333	0.02797	0.00481	-	0.03266	0.00943	-	0.00799	-	-	0.0215	-	-	0.99

non-carcinogenic risk (CR) $< 10^{-6}$ to 10^{-4} and total noncarcinogenic hazard index (HI) < 1 are summarized in Table 6. Based on the country of origin, it is remarkable that the quantity of sunflower oil daily intake is independent of the obtained by cold-pressed (Brazil and Romania) or petroleum solvent extraction (China and Turkey). Regarding trace elements concentrations in sunflower oil, the quantity of sunflower oil daily intake by individuals aged 8, 18, and 30 years old are, respectively, described in decreased order for Brazil (0.61, 1.46, 1.65 g/day) > China (0.41, 0.99, 1.12 g/day) > Romania (0.037, 0.08, 0.099 g/day) > Turkey (0.0097, 0.0234, 0.0264 g/kg) > Turkey (0.0093, 0.0224, 0.0253 g/kg).

Thus, given the findings of the current study (mineral concentrations), it seems relevant to explore the calculating amount of vegetable oil consumption per day based on trace elements quantified from different origins to be used for health promotion and regulation of several metabolic diseases.

Conclusion

The assessed sunflower oil obtained by a domestic cold-pressing demonstrates optimal qualitative properties for consumption, correlated with observed results on fatty acids composition, physicochemical optical features, thermal and oxidative qualities, and trace elements compared with DRI/AI* and FAO/WHO parameters herein evaluated. However, Cr concentration in sunflower oil was above FAO/WHO limits, which can be used as an indicator of ambient pollution.

However, to obtain values of HI < 1, and CR < 10^{-4} , the maximum sunflower oil daily consumption varies between 0.61, 1.46, and 1.65 g kg⁻¹, respectively, for individuals aged 8, 18, and 18 years. Moreover, the calculated results based on trace elements concentration regarding HI < 1 and CR < 10^{-4} indices of the sunflower oil previously qualitative approved show lower daily intake compared with prior daily consumption varying from 25 to 142 g kg⁻¹.

Thus, it is expected that this quantitative daily consumption of sunflower oil here presented can be used for other vegetable oils and several foodstuffs for health improvement and metabolic disease regulation.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Author contributions

DM: conceptualization of the topic, methodology, investigation, formal analysis, writing the original draft,

visualization, and data curation. EM, FM, LO, and AC: methodology. DB, AP, LC, KF, PH, MV, RO, and RG: investigation and formal analysis. VN and DM: writing, reviewing, and editing. VN: supervision, funding acquisition, and project administration. All authors have read and approved the final version of the manuscript, ensure the accuracy and integrity of the work, and agree to be accountable for all appearances.

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Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

1. Zhou Y, Zhao W, Lai Y, Zhang B, Zhang D. Edible Plant Oil: Global Status, Health Issues, and Perspectives. *Front Plant Sci.* (2020) 11:1315. doi: 10.3389/fpls.2020.01315

2. Mazzocchi A, de Cosmi V, Risé P, Milani GP, Turolo S, Syrén M-L, et al. Bioactive compounds in edible oils and their role in oxidative stress and inflammation. *Front Physiol.* (2021) 12:659551. doi: 10.3389/fphys.2021.659551

3. Machate DJ, Figueiredo PS, Marcelino G, Guimarães RCA, Hiane PA, Bogo D, et al. Fatty acid diets: Regulation of gut microbiota composition and obesity and its related metabolic dysbiosis. *Int J Mol Sci.* (2020) 21:4093. doi: 10.3390/ijms21114093

4. Kuo YS, Hu MH, Chan WH, Huang TY, Chou YC, Huang GS. Evaluation of the preventive effects of fish oil and sunflower oil on the pathophysiology of sepsis in endotoxemic rats. *Front Nutr.* (2022) 9:857255. doi: 10.3389/fnut.2022.857255

5. Zhao X, Xiang X, Huang J, Ma Y, Sun J, Zhu D. Studying the evaluation model of the nutritional quality of edible vegetable oil based on dietary nutrient reference intake. *ACS Omega.* (2021) 6:6691–8. doi: 10.1021/acsomega.0c05544

6. Machate DJ, Candido CJ, Inada AC, Franco BC, de Carvalho IRA, de Oliveira LCS, et al. Fatty acid profile and physicochemical, optical and thermal characteristics of *Campomanesia adamantium* (Cambess.) O. Berg seed oil. *Food Sci Tech.* (2020) 40:538–544. doi: 10.1590/fst.32719

7. Yara-Varón E, Li Y, Balcells M, Canela-Garayoa R, Fabiano-Tixier A-S, Chemat F. Vegetable oils as alternative solvents for green oleo-extraction, purification and formulation of food and natural products. *Molecules*. (2017) 22:1474. doi: 10.3390/molecules22091474

8. Gharby S. Refining vegetable oils: Chemical and physical refining. Sci World J. (2022) 2022:6627013. doi: 10.1155/2022/6627013

9. Siroma TK, Machate DJ, Zorgetto-Pinheiro VA, Figueiredo PS, Marcelino G, Hiane PA, et al. Polyphenols and ω -3 PUFAs: Beneficial outcomes to obesity and its related metabolic diseases. *Front Nutr.* (2022) 8:781622. doi: 10.3389/fnut.2021.781622

10. Carocho M, Ferreira ICFR, A. review on antioxidants, prooxidants and related controversy: Natural and synthetic compounds, screening and analysis methodologies and future perspectives. *Food Chem Toxicol.* (2012) 51:15–25. doi: 10.1016/j.fct.2012.09.021

11. Guo S, Ge Y, Jom KN. A review of phytochemistry, metabolite changes, and medicinal uses of the common sunflower seed and sprouts (*Helianthus annuss L*). *Chem Central J.* (2017) 11:95. doi: 10.1186/s13065-017-0328-7

12. Adeleke BS, Babalola OO. Oilseed crop sunflower (*Helianthus annuus*) as a source of food Nutritional and health benefits. *Food Sci Nutr.* (2020) 8:4666–84. doi: 10.1002/fsn3.1783

13. Zeb A. A comprehensive review on different classes of polyphenolic compounds present in edible oils. *Food Res Int.* (2021) 143:110312. doi: 10.1016/j.foodres.2021.110312

14. Zorgetto-Pinheiro VA, Machate DJ, Figueiredo OS, Marcelino G, Hiane PA, Pott A, et al. Omega-3 fatty acids and balanced gut microbiota on chronic inflammatory diseases: A close look at ulcerative colitis and rheumatoid arthritis pathogenesis. *J Med Food*. (2022) 25:341–54. doi: 10.1089/jmf.2021.0012

15. Rani V, Deep G, Singh RK, Palle K, Yadav UCS. Oxidative stress and metabolic disorders: Pathogenesis and therapeutic strategies. *Life Sci.* (2016) 148:183–93. doi: 10.1016/j.lfs.2016.02.002

16. Ramadan MF. Oxidation of β -sitosterol and campesterol in sunflower oil upon deep- and pan-frying of French fries. *J Food Sci Technol.* (2015) 52:6301–11. doi: 10.1007/s13197-015-1738-y

17. Ramadan MF. Healthy blends of high linoleic sunflower oil with selected cold pressed oils: Functionality, stability and antioxidative characteristics. *Ind Crops Prod.* (2013) 43:65–72. doi: 10.1016/j.indcrop.2012.07.013

18. Kiralan M, Ulas M, Ozaydin A, Ozdemir N, Ozkan G, Bayrak A, et al. Blends of cold pressed black cumin oil and sunflower oil with improved stability: A study based on changes in the levels of volatiles, tocopherols and thymoquinone during accelerated oxidation conditions. *J Food Biochem.* (2017) 41:e12272. doi: 10.1111/jfbc.12272

19. Yildiz S, Dilmen S, Turmen S, Kiralan M, Ramadan MF. Effects of natural phenolics and synthetic antioxidants on the oxidative thermal stability of refined and purified sunflower oils. *Riv Ital Sostanze Grasse*. (2021) 98:93–104. Available online at: https://www.innovhub-ssi.it/kdocs/2012984/2021_vol._ 982_-_art._02_-_yildiz.pdf (accessed September 20, 2022).

20. Melo M, Michel F, Arakaki D, Lima N, Gonçalves D, Cavalheiro L, et al. First study on the oxidative stability and elemental analysis of Babassu (*Attalea speciosa*)

edible oil produced in Brazil using a domestic extraction machine. *Molecules*. (2019) 24:4235. doi: 10.3390/molecules24234235

21. Martinec N, Balbino S, Dobsa J, Simunic-Meznaric V. Macro- and microelements in pumpkin seed oils: Effect of processing, crop season, and country of origin. *Food Sci Nutr.* (2019) 7:1634–44. doi: 10.1002/fsn3.995

22. Astolfi ML, Marini F, Frezzini MA, Massimi L, Capriotti AL, Montone CM, et al. Multielement characterization and antioxidant activity of Italian extra-virgin olive oils. *Front Chem.* (2021) 9:769620. doi: 10.3389/fchem.2021.769620

23. Mukhametov AE, Yerbulekova MT, Dautkanova DR, Tuyakva GA, Aitkhozhayeva G. Heavy metal contentes in vegetable oils of Kazakhstan origin and life risk assessment. *Int J Sci Res Innov.* (2020) 14:163–7. Available online at: https://publications.waset.org/10011598/heavy-metal-contents-in-vegetable-oils-of-kazakhstan-origin-and-life-risk-assessment (accessed September 20, 2022).

24. Farzin L, Moassesi ME. Determination of metal contents in edible vegetable oils produced in Iran using microwave-assisted acid digestion. *J App Chem Res.* (2014) 8:35–43. Available online at: https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.850.5212&rep=rep1&type=pdf (accessed September 20, 2022).

25. Tchounwon PB, Yedjou CG, Patlolla AK, Sutton DJ. Heavy metals toxicity and the environment. EXS. (2012) 101:133-64. doi: 10.1007/978-3-7643-8340-4_6

 Zhu F, Fan W, Wang X, Qu L, Yao S. Health risk assessment of eight heavy metals in nine varieties of edible vegetable in oils consumed in China. *Food Chem Toxicol.* (2011) 49:3081–5. doi: 10.1016/j.fct.2011.09.019

27. Rounizi SK, Mohajeri FA, Broujeni HM, Pourramezani F, Jambarsang S, Kiani H, et al. The chemical composition and heavy metal content of sesame oil produced by different methods: A risk assessment study. *Food Sci Nutr.* (2021) 9:2886–93. doi: 10.1002/fsn3.2245

28. Fathollahy I, Pezeshki A, Sorouraddin SM. Effect of different extraction methods on trace metal elements content of Persian lime (*Citrus latifolia*) seed oil. *OCL*. (2021) 28:44. doi: 10.1051/ocl/2021031

29. Serra JL, Rodrigues AMC, Meirelles AJA, Darnet SH, Silva LHM. Alternative sources of oils and fats from Amazonian plants: Fatty acids, methyl tocols, total carotenoids and chemical composition. *Food Res Int.* (2019) 116:12–9. doi: 10.1016/j.foodres.2018.12.028

30. Romanić R. Cold pressed sunflower (Helianthus annuus L.) oil. In: Mohamed FR, editor. *Cold Pressed Oils*. Cambridge: Academic Press (2020). p. 197-218. 197-218. doi: 10.1016/B978-0-12-818188-1.00017-7

31. Mazaheri Y, Torbati M, Azadmard-Damirchi S, Savage GP. A comprehensive review of the physicochemical, quality and nutritional properties of *Nigella sativa* oil. *Food Rev Int.* (2019) 35:342–262. doi: 10.1080/87559129.2018.1563793

32. Petraru A, Ursachi F, Amariei S. Nutritional characteristics assessment of sunflower seeds, oil and cake. Perspective of using sunflower oilcake as a functional ingredient. *Plants.* (2021) 10:2487. doi: 10.3390/plants10112487

33. Bligh EG, Dyer WJ, A. rapid method of total lipid extraction and purification. *Can J Biochem Physiol.* (1959) 37:911–7. doi: 10.1139/o59-099

34. Maia EL, Rodriguez-Amaya DB. Avaliação de um método simples e econômico para a metilação de ácidos graxos com lipídios de diversas espécies de peixes. *Rev Inst Adolfo Lutz.* (1993) 53:27–35.

35. Dodds ED, McCoy MR, Rea LD, Kennish JM. Gas chromatographic quantification of fatty acid methyl esters: Flame ionization detection vs. electron impact mass spectrometry. *Lipids*. (2005) 40:419–428. doi: 10.1007/s11745-006-1399-8

36. Ulbricht TLV, Southgate DAT. Coronary heart disease: Seven dietary factors. Lancet. (1991) 338:985–92. doi: 10.1016/0140-6736(91)91846-M

37. Santos-Silva J, Bessa RJB, Santos-Silva F. Effect of genotype, feeding system and slaughter weight on the quality of light lambs. II Fatty acid composition of meat. *Livest Prod Sci.* (2002) 77:187–94. doi: 10.1016/S0301-6226(02)0 0059-3

38. American Oil Chemists Society. Official Methods and Recommended Practices of the AOCS. AOCS: Washington, DC, USA. (2004).

39. European Committee for Standardization – CEN. Method EN14112: fatty acid methyl esters (fame)-determination of oxidation stability (accelerated oxidation test). Brussels: CEN. (2003).

40. Carneiro AF, Carneiro CN, Pires LN, Teixeira LSG, Azcarate SM, Dias FS. D-optimal mixture design for the optimization of extraction induced by emulsion breaking for multielemental determination in edible vegetable oils by microwave-induced plasma optical emission spectrometry. *Talanta*. (2020) 219:121218. doi: 10.1016/j.talanta.2020.121218

41. Association of Official Analytical Chemists. (2002). Guidelines for Single Laboratory Validation of Chemical Methods for Dietary Supplements and Botanicals. Available online at: https://members.aoac.org/AOAC_Docs/ StandardsDevelopment/SLV_Guidelines_Dietary_Supplements.pdf (accessed March 21, 2021).

42. Machate DJ, Melo ESP, Arakaki DG, Guimarães RCA, Hiane PA, Bogo D, et al. High concentration of heavy metal and metalloid levels in edible campomanesia adamantium pulp from anthropic areas. *Int J Environ Res Public Health.* (2021) 18:5503. doi: 10.3390/ijerph18115503

43. US. EPA – United States Environmental Protection Agency. *Risk Assessment Guidance for Superfund, Volume I: Human Health Evaluation Manual.* Washington, DC, USA: Office of Emergency and Remedial Response. (1989).

44. JECFA WHO. Summary and Conclusions of the 61st Meeting of the Joint FAO/WHO Expert Committee on Food Additives. Rome, Italy: JECFA WHO. (2003).

45. US EPA IRIS. (2016). Program Information about the Integrated Risk Information System: Chronic Oral Reference Dose (RfD). Available online at: https://cfpub.epa.gov/ncea/iris/search/ (accessed April 30, 2022).

46. US EPA-RAIS. The Risk Assessment Information System. Available online at: https://cfpub.epa.gov/ncea/iris/search/index.cfm (accessed April 30, 2022).

47. Rauf S, Jamil N, Tariq SA, Khan M, Kausar M, Kaya Y. Progress in modification of sunflower oil to expand its industrial value. *J Sci Food Agr.* (2017) 97:1997–2006. doi: 10.1002/jsfa.8214

48. Gotor AA, Rhazi L. Effects of refining process on sunflower oil minor components: a review. OCL. (2016) 23:D207. doi: 10.1051/ocl/2016007

49. Carelli AA, Frizzera LM, Forbito PR, Crapiste GH. Wax composition of sunflower seed oils. JOACS. (2002) 79:763-8. doi: 10.1007/s11746-002-0556-9

50. Huang Y, Lu M, Wu H, Zhao T, Wu P, Cao D. High drying temperature accelerates sunflower seed determination by regulating the fatty acid metabolism, glycometabolism, and abscisic acid/gibberellin balance. *Front Plant Sci.* (2021) 12:628251. doi: 10.3389/fpls.2021.628251

51. Yun J-M, Surh J. Fatty acid composition as a predictor for the oxidation stability of Korean vegetable oils with or without induced oxidative stress. *Prev Nutr Food Sci.* (2012) 17:158–165. doi: 10.3746/pnf.2012.17.2.158

52. Nde DB, Foncha AC. Optimization methods for the extraction of vegetable oils: A review. *Process.* (2020) 8:209. doi: 10.3390/pr8020209

53. Thies F, Garry JMC, Yaqoob P, Rerkasem K, Williams J, Shearman C P, et al. Association of n-3 polyunsaturated fatty acids with stability of atherosclerotic plaques: a randomised controlled trial. *Lancet.* (2003) 361:477-85. doi: 10.1016/S0140-6736(03)12468-3

54. Chen J, Liu H. Nutritional indices for assessing fatty acids: A mini-review. Int J Mol Sci. (2020) 21:5695. doi: 10.3390/ijms21165695

55. Food and Agriculture Organization of the United Nations – FAO, Codex Alimentarius. *Standard for olive and olive pomace oils codex stan 33-1981 named vegetable oils codex stan 210-1999*. Rome: FAO/WHO. (2015).

56. Codex Alimentarius Commission. *Standard for Named Vegetable Oils*. CXS 210-1999. Adopted in 1999. Revised in 2001, 2003, 2009, 2017, 2019. Amended in 2005, 2011, 2013, 2015, 2019, 2021. (1999).

57. Mengistie T, Alemu A, Mekonnen A. Comparison of physicochemical properties of edible vegetable oils commercially available in Bahir Dar, Ethiopia. *Chem Int.* (2018) 4:130–5. doi: 10.5281/zenodo.1475356

58. Tao L. Oxidation of polyunsaturated fatty acids and its impact on food quality and human health. *Adv Food Technol Nutr Sci Open J.* (2015) 1:135-42. doi: 10.17140/AFTNSOJ-1-123

59. Ivanova M, Hanganu A, Dumitriu R, Tociu M, Ivanov G, Stavarache C, et al. Saponification value of fats and oils as determined from ¹H-NMR data: The case of dairy fats. *Foods.* (2022) 11:1466. doi: 10.3390/foods11101466

60. Muñoz AM, Casimiro-Gonzales S, Gómez-Coca RB, Moreda W, Best I, Cajo-Pinche M I, et al. Comparison of four oil extraction methods for sinami fruit (*Oenocarpus mapora* H. Karst): Evaluation quality, polyphenol content and antioxidant activity. *Foods.* (2022) 11:1518. doi: 10.3390/foods11101518

61. Mabaleha MB, Mitei YC, Yeboah SO. A comparative study of the properties of selected melon seed oils as potential candidates for development into commercial edible vegetable oils. *J Amer Oil Chem Soc.* (2007) 84:31–36. doi: 10.1007/s11746-006-1003-7

62. Segatin N, Zontar TP, Ulrih NP. Dielectric properties and dipole moment of edible oils subjected to 'frying' thermal treatment. *Foods.* (2020) 9:900. doi: 10.3390/foods9070900

63. Gharby S, Harhar H, Bouzoubaa Z, Roudani A, Chafchaouni I, Kartah B, et al. Effect of polyphenols extracts from margins on the stability of sunflower oil. *J Mater Environ Sci.* (2014) 5:464–9.

64. Lužaić TZ, Grahovac NL, Hladni NT, Romanić RS. Evaluation of oxidative stability of new cold-pressed sunflower oils during accelerated thermal stability tests. *Food Sci Technol.* (2022) 42:e67320. doi: 10.1590/fst.67320

65. Symoniuk E, Ratusz K, Ostrowska-Ligeza E, Krygier K. Impact of selected chemical characteristics of cold-pressed oils on their oxidative stability determined using the Rancimat and pressure differential scanning Calorimetry method. *Food Anal Methods.* (2018) 11:1095–104. doi: 10.1007/s12161-017-1081-1

66. Santos JCO, dos Santos IMG, de Souza AG, Prasad S, dos Santos AV. Thermal stability and kinetic study on thermal decomp osition of commercial edible oils by thermogravimetry. *Food Eng Phys Prop.* (2002) 67:1393–8. doi: 10.1111/j.1365-2621.2002.tb10296.x

67. Dweck J, Sampaio CMS. Analysis of the thermal decomposition of commercial vegetable oils in air by simultaneous TG/DTA. *J Therm Anal Calorim.* (2004) 75:385–91. doi: 10.1023/B:JTAN.0000027124.96546.0f

68. Correia IMS, Souza MJB, Araújo AS, Sousa EMBD. Thermal stability during pyrolysis of sunflower oil produced in the northeast of Brazil. *J Therm Anal Calorim.* (2012) 109:967–74. doi: 10.1007/s10973-011-1773-5

69. Zhang ZS, Wang S, Liu H, Li BZ, Che L. Constituents and thermal properties of milk thistle seed oils extracted with three methods. *Food Sci Tech.* (2020) 126:109282. doi: 10.1016/j.lwt.2020.109282

70. Petkova Z, Antova G, A. comparative study on quality parameters of pumpkin, melon and sunflower oils during thermal treatment. OCL. (2019) 26:32. doi: 10.1051/ocl/2019028

71. Lourenço SC, Moldão-Martins M, Alves VD. Antioxidants of natural plant origin: from sources to food industry applications. *Molecules*. (2019) 24:4132. doi: 10.3390/molecules24224132

72. Sahin S, Elhussein E, Gulmez O, Kurtulbas E, Yazar S. Improving the quality of vegetable oils treated with phytochemicals: a comparative study. *J Food Sci Technol.* (2020) 57:3980–7. doi: 10.1007/s13197-020-04428-z

73. Wang W, Xiong P, Zhang H, Zhu Q, Liao C. Analysis, occurrence, toxicity and environmental health risks of synthetic phenolic antioxidants: A review. *Environ Res.* (2021) 201:111531. doi: 10.1016/j.envres.2021.111531

74. Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn Rev.* (2010) 4:118–26. doi: 10.4103/0973-7847.70902

75. Calligaris S, Manzocco L, Nicoli MC. Modelling the temperature dependence of oxidation rate in water-in-oil emulsions stored at sub-zero temperature. *Food Chem.* (2007) 101:1019–24. doi: 10.1016/j.foodchem.2006.02.056

76. Gonçalves RP, Março PH, Valderrama P. Thermal edible oil evaluation by UV-Vis spectroscopy and chemometrics. *Food Chem.* (2014) 163:83– 6. doi: 10.1016/j.foodchem.2014.04.109

77. Popa S, Milea MS, Boran S, Nitu SV, Mosoarca GE, Vancea C, et al. Rapid adulteration detection of cold pressed oils with their refined versions by UV–Vis spectroscopy. *Sci Rep.* (2020) 10:16100. doi: 10.1038/s41598-020-72558-7

78. Wu XJ, Pan Z, Zhao YP, Liu HL, Zheng LJ. Application of fluorescence spectra and parallel factor analysis in the classification of edible vegetable oils. *Guang Pu.* (2014) 34:2137–42. doi: 10.3964/j.issn.1000-0593(2014)08-2137-06

79. Li B, Wang H, Zhao Q, Ouyang J, Wu Y. Rapid detection of authenticity and adulteration of walnut oil by FTIR and fluorescence spectroscopy: A comparative study. *Food Chem.* (2015) 181:25–30. doi: 10.1016/j.foodchem.2015.02.079

80. National Institutes of Health. Office of Dietary Supplements. Dietary Reference Intakes (DRI): Recommended Dietary Allowances and Adequate Intakes, Elements food and Nutrion Board, National Academies. Washington, DC. National Academies Press (US). (2019).

81. Food and Agriculture Organization and Word Health Organization (FAO/WHO). Food Contaminants. In *Codex Alimentarius Commission*; XVII. Rome, Italy: FAO/WHO. (1984).

82. Lewis J. Codex Nutrient Reference Values. Rome, Italy: Food and Agriculture Organization of the United Nations and World Health Organization (FAO/WHO). (2019).

83. WHO. Aluminium in drinking-water. Background document for development of WHO Guidelines for Drinking-water quality. (2010).

84. Mendil D, Uluözlü OD, Tüzen M, Soylak M. Investigation of the levels of some element in edible oil samples produced in Turkey by atomic absorption spectrometry. *J Hazard Mater.* (2009) 165:724–8. doi: 10.1016/j.jhazmat.2008.10.046

85. Rosa ACG, Melo ESP, Junior ASA, Gondim JMS, Sousa AG, Cardoso CAL, et al. Transfer of metal(loid)s from soil to leaves and trunk xylem sap of medicinal plants and possible health risk assessment. Int J Environ Res Public Health. (2022) 19:660. doi: 10.3390/ijerph19 020660 86. Bakircioglu D, Kurtulus YB, Yurtsever S. Comparison of extraction induced by emulsion breaking, ultrasonic extraction and wet digestion procedures for determination of metals in edible oil samples in Turkey using ICP–OES. *Food Chem.* (2014) 138:770–775. doi: 10.1016/j.foodchem.2012. 10.089

87. Goldhaber SB. Trace element risk assessment: essentiality vs. *toxicity RTP.* (2003) 38:232–42. doi: 10.1016/S0273-2300(02) 00020-X

88. Rong S, Liao Y, Zhou J, Yang W, Yang Y. Comparison of dietary guidelines among 96 countries worldwide. *Trends Food Sci Technol.* (2021) 109:219–29. doi: 10.1016/j.tifs.2021. 01.009

89. Pu G, Lu S, Zheng M, Huang J, Cheng G. Study on the use of cooking oil in Chinese dishes. *Int J Environ Res Public Health.* (2019) 16:3367. doi: 10.3390/ijerph161 83367