

Review

# A New and Integral Approach to the Etiopathogenesis and Treatment of Breast Cancer Based upon Its Hydrogen Ion Dynamics

Salvador Harguindey <sup>1,\*</sup><sup>(D)</sup>, Khalid Alfarouk <sup>2</sup>, Julián Polo Orozco <sup>1</sup>, Kévin Hardonnière <sup>3</sup><sup>(D)</sup>, Daniel Stanciu <sup>4</sup>, Stefano Fais <sup>5</sup> and Jesús Devesa <sup>6</sup><sup>(D)</sup>

- <sup>1</sup> Institute of Clinical Biology and Metabolism, Postas 13, 01004 Vitoria, Spain; polorozco@gmail.com
- <sup>2</sup> Al-Ghad International Colleges for Applied Medical Sciences, Al-Madinah Al-Munawarah, Saudi Arabia and Alfarouk Biomedical Research LLC, Tampa, FL 33617, USA; khaliomer@gmail.com
- <sup>3</sup> Université Paris-Saclay, Inserm, Inflammation, Microbiome and Immunosurveillance, 92290 Châtenay-Malabry, France; kevin.hardonniere@u-psud.fr
- <sup>4</sup> Scientific Direction, MCS Foundation For Life, 5623KR Eindhoven, The Netherlands; cd.stanciu@gmail.com
- <sup>5</sup> Department of Oncology and Molecular Medicine, Istituto Superiore di Sanità (National Institute of Health), Viale Regina Elena, 299, 00161 Rome, Italy; Stefano.fais@iss.it
- <sup>6</sup> Scientific Direction, Foltra Medical Centre, Travesía de Montouto 24, 15886 Teo, Spain; devesa.jesus@gmail.com
- \* Correspondence: salvaszh@telefonica.net

Received: 21 January 2020; Accepted: 6 February 2020; Published: 7 February 2020



MDF

**Abstract:** Despite all efforts, the treatment of breast cancer (BC) cannot be considered to be a success story. The advances in surgery, chemotherapy and radiotherapy have not been sufficient at all. Indeed, the accumulated experience clearly indicates that new perspectives and non-main stream approaches are needed to better characterize the etiopathogenesis and treatment of this disease. This contribution deals with how the new pH-centric anticancer paradigm plays a fundamental role in reaching a more integral understanding of the etiology, pathogenesis, and treatment of this multifactorial disease. For the first time, the armamentarium available for the treatment of the different types and phases of BC is approached here from a Unitarian perspective-based upon the hydrogen ion dynamics of cancer. The wide-ranged pH-related molecular, biochemical and metabolic model is able to embrace most of the fields and subfields of breast cancer etiopathogenesis and treatment. This single and integrated approach allows advancing towards a unidirectional, concerted and synergistic program of treatment. Further efforts in this line are likely to first improve the therapeutics of each subtype of this tumor and every individual patient in every phase of the disease.

**Keywords:** pH and breast cancer; breast cancer etiology; breast cancer pathogenesis; breast cancer treatment; pH-centric anticancer paradigm; hydrogen ion dynamics of cancer; cancer proton reversal; multiple drug resistance (MDR) integral approach

# 1. Introduction

According to The International Agency For Research on Cancer (IARC), breast cancer (BC) is the most common malignant tumor in humans and the secondary cause of mortality of cancer in women, just behind lung cancer [1]. BC-related morbidity is primarily due to a progressive metastatic process [2]. Many associated risk factors, either genetic, from *BRCA1* and *BRCA2* gene mutations, a wide array of other genetic derangements [3], and a multiplicity of environmental factors such as age, obesity and estrogens, among many others [4], are involved in the onset of BC. Despite significant advances in therapy, the overall results are not too successful, especially in advanced disease [5,6]. This may

indicate that a change towards a more comprehensive and perhaps radically different perspective is necessary in order to incorporate more rational and less toxic treatments, and at the same time foster a better understanding of this multifaceted disease.

A new and all-comprehensive pH-related paradigm has increasingly grown during the last few years, notably after the realization that the cancer-selective intracellular/extracellular pH deregulation is critical in the control of many cellular, both normal and pathological, processes [7]. One of the latest achievements of the new pH-paradigm has been to describe the integral and unitarian mechanism mediating the wide array of apparently unrelated factors involved in the etiopathogenesis of cancer, a finding that we can now be applied to BC [8].

The study of the abnormal hydrogen ion (H<sup>+</sup>) dynamics of cancer started almost five decades ago [9–13]. Since then, a rapid increase in the understanding of the deregulated H<sup>+</sup> dynamics in cancer and the cancer-specific proton gradient reversal (CPR) has resulted in a new and increasingly outreaching paradigm, known as the pH-centric anticancer paradigm. This perspective embraces many different aspects of basic, preclinical and clinical oncology, all derived from this H<sup>+</sup>-related energetic concept that has allowed an intimate acid-base approach to the inner nature of malignancy. Nowadays the pH or H<sup>+</sup>- related model is already able to unite different fields, from molecular biology to biochemistry and the metabolism of cancer, having already reached up to the clinical aspects of cancer cells and tissues [14,15]. This perspective has rapidly extended to other different collateral areas of oncological research, incorporating within its range areas previously far apart when approached by the old and reductionist model, such as angiogenesis, environmental carcinogenesis and cancer immunology, also including the initiation, progression, metastatic process and even the spontaneous regression of cancer [16–21]. In summary, this integral and comprehensive paradigm can embrace most, if not all, aspects of cancer, and also of BC, from etiopathogenesis to treatment.

The cancer-selective abnormalities of intracellular alkalization plus extracellular acidification of all types of solid tumors and leukemias (CPR) represent the mirror image of normality, an upside-down disruption and tip over from normal homeostasis and allostasis [8]. Normality is exactly the opposite: a more acidic medium inside than outside non-cancerous cells [22,23]. As a consequence of this acid-base homeostatic disruption and energetic failure of cellular hydrogen ion (H<sup>+</sup>) dynamics, attempts to induce intracellular acidification using proton transport and pump inhibitors (PTIs and PPIs), as well as other intracellular acidifiers of different origins and natures (repurposed drugs), has become a new and valuable therapeutic strategy in selective cancer treatment. In this vein, the wide-ranging applications and potential benefits of this approach to the therapeutics of solid tumors has been recently published in a full issue containing fourteen reviews addressing the different aspects of the new pH-centric anticancer paradigm [24].

In the same line, we recently published an original review dealing with the pH-related possibilities in the treatment of brain malignancies in humans [25]. To uncover further pathophysiological and therapeutic applications of this post-traditional and non-main stream approach to cancer, this basic to clinically oriented and translational review will also discuss the foundations of the H<sup>+</sup>-related paradigm now applied in this contribution to the etiopathogenesis and treatment of BC [8].

## 2. Cancer as an Acid-Base Disease

The highest and lowest limits of pH in cellular and human life are considered to range between 6.8 and 7.8, a difference of only one unit. The normal cellular pH (pHi) is accepted to be 7.0–7.1, while a normal interstitial and systemic pH (pHe) is considered to be 7.35–7.45. Otherwise, the pHi of malignant cells can be as high as 7.8 [8]. Contrariwise, during malignant cells apoptosis the pHi has been shown to be as low as to 5.0 [26]. Most importantly, seminal studies in this area demonstrated that malignant transformation takes place at a pHi > 7.2, that is, only 0.2 units above normality, the time period needed for the oncogenic effect of the acid-base change being inversely related to the pHi increase [27].

During the last few years the new pH-centric anticancer paradigm and the H<sup>+</sup> dynamics of cancer have helped to significantly increase the understanding of the intimate nature of human malignant tumors. Nowadays, it is agreed that all cancer cells and tissues have deeply rooted evolutionary and thermodynamic pH-related advantages over all normal tissues. This characteristic allows neoplastic cells first to survive in the most hostile conditions, then to grow locally, later to invade neighboring tissues to finally disseminate out of control, overwhelming all the defensive barriers and immune mechanisms of the host: "the neostrategy of cancer cells and tissues". These highly pathological energetic and metabolic disruptions are based on a pathognomonic intracellular alkalization of cancer cells and, secondarily, to an extracellular and intratumoral microenvironmental acidosis. Both coordinated phenomena are facilitated by overactive membrane-bound proton transporters (PT) and pumps (PP) extruding mechanisms, inducing an inversion of the normal cells/surrounding tissues pH gradient (pHi to pHe), or CPR, across cellular membranes. These deregulated pH dynamics also determine the cancerous effects on normal cells and tissues, from early tumorigenesis and transformation to proliferation, local growth and a metastatic process that usually ends up with the death of the host.

Such metabolic reprogramming confers to cancer cells and tissues other important thermodynamic advantages, such as enhancing their resistance to hypoxia and to cancer therapy (MDR), allowing them to survive under almost any conditions. Finally, these dynamic changes allow malignant cells and tissues to avoid the pro-apoptotic intracellular acidification (IA), which would result in a selective cancer cell death as the successful outcome of treatment. As previously mentioned, the new acid-base approach to cancer has extended side-ways to the point that it can now provide further meaning to most, if not all, the hallmarks previously described for cancer, and even envision new ones [28,29]. Such a conceptual deepening into the intimate nature of malignancy allows the new H<sup>+</sup>-related paradigm to cover under one single heading the many different areas and hallmarks of cancer research and treatment previously disconnected to a large extent, namely:

- (a) pH and the Warburg effect: Recent publications of our group and others have defended that the Warburg effect can be fully explained by the selective pH abnormalities of cancer cells and their effect on aerobic glycolysis [8,24,25,30–36].
- (b) pH abnormalities in the etiology and pathogenesis of BC and other tumors: Nowadays, intracellular (IC) alkalization is increasingly recognized as a fundamental and sine qua non factor in cellular transformation in BC and other malignant tumors [8,37–44].
- (c) pH and cancer molecular biology, biochemistry and metabolism: Recent experimental data have clearly shown that the pH-related paradigm can reinterpret the molecular biology, biochemistry and intermediary metabolism of cancer cells and tissues from an integral and Unitarian dynamic perspective [8,37–42,45,46].
- (d) pH, tumor growth and invadopodia: Local invasion is promoted by Na<sup>+</sup>/H<sup>+</sup> mediated low pH of invadopodia at the advancing edge of cancer cells [37,47,48].
- (e) pH and microenvironmental-intratumoral acidosis in cancer growth and dissemination: Through different mechanisms, from biochemical to immunological, the high extrusion of lactic acid and other metabolites from cancer cells creates a highly acidified extracellular media that stimulates different and coordinated mechanisms strategically organized to favor local growth, invasion and dissemination [8,22,49–54].
- (f) pH and the metastatic process: The pHi/pHe gradient reversal (CPR) is fundamental in all stages of cancer growth, from cell transformation and the initiation of tumor growth to the activity and progression of the metastatic process [23,35,48,55].
- (g) pH, proton transporters (PTs), proton pumps (PPs) and their inhibitors (PTIs and PPIs): During the last decade the increasing interest and knowledge of the different membrane-bound PTs and PPs in cancer pathogenesis, as well as their inhibitors in cancer treatment, has made it possible

to approach, from a different and non-mainstream perspective, the latest therapeutic efforts in cancer treatment, either in BC or in other human malignant tumors [15,36,40,42,54–70].

- (h) pHi acidification and reverting cancer proton reversal (CPR) in cancer treatment: Attempts to revert CPR in cancer are the fundamental therapeutic issue in the entire paradigm of the H<sup>+</sup>-related dynamics of cancer, BC included, from the initiation stage to angiogenesis and to the treatment of metastatic disease [8,16,23,55,63,71].
- pH and voltage-gated sodium channels (VGSC): The expression of Na+ channels synergically associated with Na<sup>+</sup>/H<sup>+</sup> antiporter activity and over-expression is important in BC pathogenesis by stimulating local invasion and the metastatic process, while its suppression is a valuable complementary therapeutic option [72–77].
- (j) pH and environmental carcinogenesis: An integral explanation of human environmental carcinogenesis has been recently advanced, proposing that the oncogenic activity of many different carcinogens induce the same cancer-specific effects on cellular H<sup>+</sup> dynamics (CPR). This recent and highly important integration into the H<sup>+</sup>-related paradigm strongly suggests the possibility of the existence of a universal mechanism responsible for environmental carcinogenesis [18,19].
- (k) pH and cancer immunity: The acidic pHe of tumors has been considered to be the ultimate mechanism allowing them to escape from the anti-tumor immunity of the parasitized human organism. The final result is that this microenvironmental-intratumoral-extracellular (EC) low pHe creates a protective shield around cancer tissues with the onset of a state of anergy and immunosuppression mediated by the EC acidification-induced loss of function of T and NK cells. It also helps to explain the limitations and failures of checkpoint blockade in immunotherapy. Contrariwise, counteracting microenvironmental tumor acidity improves the antitumor responses to immunotherapy [20,21,47,78,79].
- (l) pH and apoptosis: Malignant cell apoptosis is induced by pHi acidification while intracellular alkalization suppresses it [8,80–83].
- (m) pH and drug resistance (MDR): Beyond the fact that the expression of P-glycoprotein (P-gp) leads to an elevation of pHi in cancer cells, an integral mechanism that explains MDR-based upon the selective changes in pHi and pHe (CPR), has been recently developed [84–90].
- (n) pH, nanodrugs and liposomes: Systemic administration of nanoparticles disrupts microtubule dynamics and can be potentially useful for treatment on its own and in the overcoming of MDR. Some preparations of nanoparticles structure and delivery are highly pH-dependent [91–93].
- (o) pH and aquaporins: Environmental pH changes either facilitate or hinder water diffusion across membranes while a rapid drop of cytosolic pH due to anoxia leads to closure of aquaporins in the plasma membrane [94,95].
- (p) pH and autophagy: No matter that autophagy still appears to be a controversial issue, is considered a defensive survival mechanism of cancer cells in order to overcome drug-induced cellular stress and cytotoxicity. Acidic conditions increase autophagy in cancer cells suggesting that autophagy is a protective mechanism for tumor cells to survive under the microenvironmental acidic stress. Inhibiting autophagy may lead to a full cytotoxic effect [96–100] (for further details see the text).
- (q) pH and repurposed drugs in cancer treatment: There is a wide array of intracellular acidifiers unrelated to PTIs or PPIs effects from many different origins and natures that have shown anticancer properties and in many cases are minimally toxic or not toxic at all [8,101–103].
- (r) pH and photodynamic therapy in cancer: pHi acidification using photosensitizing agents leads to apoptosis and has been shown to suppress tumor growth and increase survival in animal tumors. Clinical studies in different solid tumors in humans are underway [104,105].
- (s) pH and the spontaneous regression of cancer (SRC): The association of spontaneous regression of different cancers in humans with deep-seated systemic acid-base changes has been recognized for more than half a century [106–112].

(t) pH, evolution and cancer metabolism: From the fields of physics and chemistry to carcinogenesis, a pH-related reverse evolutionary process have been considered even from the times of Albert Szent-Györgyi (personal communication) [113–117].

## 3. All Phases of Breast Cancer Are Weaved into Each Other to Conform a Single, All-Comprehensive and Progressive, Multistage Unity

All the above-mentioned oncological fields and subfields have in common a pivotal characteristic, namely, the aberrant pHi/pHe regulation of hydrogen (H<sup>+</sup>) ion dynamics [118], an abnormality that cannot be more opposed to the acid-base and energetic normality of non-cancerous cells and tissues. In a recent publication on brain malignancies, a large number of known etiopathogenic factors from many different natures and origins, all known to cause different human malignancies, were considered together [8,25]. Importantly, all the cancer-inducing factors listed in that review, but also in other previous reports of this group, act via a universal mediating mechanism, that is, an increase of cell pH secondary to the stimulation of the activity of NHE and/or other H<sup>+</sup>-related membrane-bound PTs and PPIs (Figure 1).



**Figure 1.** Proton transporters and proton pumps involved in the carcinogenicity of breast cancer and other malignant tumors. pHi: intracellular pH; CT: cell transformation; pHe: extracellular pH; PTs: proton transporters; PPs: Proton pumps; NHE-1: Na<sup>+</sup>/H<sup>+</sup> antiporter; H<sup>+</sup>: hydrogen ion; CAs: carbonic anhydrases; MCT: monocarboxylate transporters; P-gp: P-glycoprotein; V-H<sup>+</sup>-ATPase: Vacuolar ATPase; MDR: Multiple drug resistance. Blue arrows: Induction. Black arrows: Result. Yellow color: intracellular space of a tumor cell.

The nature and evolution of any solid tumor, and also of BC, makes it possible to accept that the deep-seated pH deregulations and/or highly disrupted H<sup>+</sup>-dynamics of malignancy are a fundamental factor behind a predetermined, progressive and staggered strategy of growth and dissemination ("the selective neostrategy of cancer cells and tissues"). This process begins with cell transformation and is closely followed by local growth and invasion under a highly hostile acid-base tumor microenvironment (TME). These initial phases are followed by neoangiogenesis, which favors metastatic dissemination and drug resistance. Each of these closely related periods of the natural evolution of cancer is not separated at all from the previous or subsequent ones but forms a Unitarian and dynamically active process that can even be considered a "conscious" and organized preprogrammed strategy. Despite this, the understanding raised by the pH-anticancer perspective has the potential to offer the possibility of applying therapeutic methods in a unique direction in all the phases of the malignant process, either in BC or in other solid tumors. This strategy is bound to interfere with each and all cancer hallmarks through different procedures acting from prevention to the treatment of advanced disease.

# 4. pH/NHE, Microenvironmental Acidosis and Immunity, Insulin, Prolactin, Estrogens, Genetic Abnormalities and Growth Factors in the Promotion of Breast Cancer

# 4.1. pH/NHE, H<sup>+</sup> Extrusion and/or Intracellular Alkalization in the Etiology and Pathogenesis of Breast Cancer

In addition to NHE overexpression, H<sup>+</sup> extrusion from cells can also be mediated by a cohort of other membrane-bound proton transporters, pumps and ion channels [8,15]. On one hand, these actors participate in keeping pHi normal to elevated, so preventing a low pHi-mediated therapeutic apoptosis. Such factors are carbonic anhydrases (CAs), mainly CAIX and CAXII, vacuolar H<sup>+</sup>-ATPase proton pumps, voltage gated sodium channels, sodium bicarbonate cotransporters, monocarboxylate transporters (MCTs), Cl<sup>-</sup>/HCO3<sup>-</sup> exchangers and ATP-Synthase [15,48] (Figure 1). Different types of acid extruders like NHE1, NBCn1 and MCT4 are expressed in human mammary tumors, promoting growth of at least triple negative BC (TNBC) through synergistic and different mechanisms of action [56]. Paradoxically, the Na<sup>+</sup>/H<sup>+</sup> exchanger regulatory factor 1 (NHERF1) presents a dual activity, either oncosuppressant or prooncogenic in invasive BC, depending of the cellular location of its activity [119]. Also, NHE1 and NBCn1 drive cell cycle progression in human BC cells, while their knocking down reduces proliferation and progression [70]. TME acidosis is also associated with pain in bone metastasis in BC [120]. Unfortunately, although one or more of these H<sup>+</sup>-extrusion systems substantially applies to all malignancies, a complete selective mapping of which PTs and PPs are overexpressed in each particular tumor is still missing. Therefore, at the moment the concerted use of all of them in pharmacological doses is recommended [53].

# 4.2. A Universal Mechanism as a Final Mediating Cause of Breast Cancer

It has been recently demonstrated that H<sup>+</sup> efflux alone is sufficient to induce dysplasia and potentiate cancer growth and invasion by oncogenic Ras and that inhibiting H<sup>+</sup> efflux induces cell death in invasive primary tumor mammary cells [38]. In the same line, the most striking results have been obtained by the group of Fliegel, showing that NHE-mediated H<sup>+</sup> extrusion alone has a direct carcinogenic effect on breast cells [40]. The same alteration also plays a fundamental role in the metastatic process and in multiple drug resistance (MDR) [41–44]. In these studies, NHE1 hyperactivity and/or a high pHi act as an early and decisive driver in BC carcinogenesis and also in most, if not all, other human malignancies [37,121]. Moreover, the elevated pHi is also the main responsible actor for the secondary acidification of the extracellular/intratumoral microenvironment (TME). Importantly, H<sup>+</sup> extrusion by itself has been implicated in the transition and progression from precancerous ductal carcinoma in situ to invasive BC. Of note, even the precancerous lesion already shows a higher than normal proton export rate [40,42,43]. Indeed, the invasive BC cells show a higher pHi and a higher production and exportation of H<sup>+</sup> into the TME than noninvasive cells [40,53,122]. (Table 1). All these are qualitative and highly specific changes in the etiopathogenesis of BC.

**Table 1.** pH-related and -unrelated mechanisms in the etiopathogenesis and progression of breast cancer. BC: Breast cancer; TME: Tumor microenvironment; MDR: Multidrug resistance; NHE: Na<sup>+</sup>/H<sup>+</sup> antiporter: CAs: Carbonic anhydrases; ER-: Estrogen negative cells; PTIs: Proton transport inhibitors; PPIs; Proton pump inhibitors.

Mechanisms	Summary	References
H <sup>+</sup> extrusion and/or elevated pHi	H <sup>+</sup> extrusion from cells is sufficient to induce transformation, growth and invasion in BC and other tumors. NHE-mediated H <sup>+</sup> extrusion by itself has a carcinogenic effect on breast cells and increases MDR.	[8,23,25,37–44,48,54,59, 122–125]
Tumor micro-environmental (TME) acidosis, immunity and MDR	Acidity of the TME disrupts the body immune defense mechanisms towards malignant tumors, locally and systemically. This allows a relentless and uncontrolled tumor progression. TME also has an essential role in the progression of inflammatory BC. Thus, TME is a novel therapeutic target in BC. TME acidity also induces MDR.	[20–22,47,49–51,78,79, 84,87,116,120,126–138]

Mechanisms	Summary	References
Insulin (INS) and insulin-like growth factor-1 (IGF-1)	INS and INS-resistance have a direct effect in raising pHi and are associated with breast cancer carcinogenicity and progression. Over-expression of insulin/insulin-like growth factor-1 is associated with a decrease in the life span of women with BC.	[3,35,39,139–149]
Prolactin (PRL)	PRL stimulates growth, motility and invasiveness of BC cells through NHE1 activation. In this way contributes to the metastatic process of human BC and becomes another therapeutic target.	[150–153]
Estrogens	Estrogens play a crucial role in breast tumorigenesis by promoting cell proliferation and decreasing apoptosis. ER-cells are considered to have a higher expression of NHE activity and are preferably associated with CA-IX over CA-XII. Inhibition of CA-IX improves the prognosis of the disease.	[4,154–157]
Ion channels	NaV1.5-Na <sup>+</sup> channels associated with NHE-1 are overexpressed in BC, stimulating the formation of invadopodia, facilitating local growth and the metastatic process.	[8,72,73,75–77,148]
PTs, PPs, and Growth factors	NHE1-overexpression is stimulated by a myriad of factors, which alone or in combination induce a carcinogenic elevation of pHi as the oncogenic response of normal cells of different origins and locations. Carbonic anhydrases (CAs) also have an important role in the pathogenesis of BC, mainly in hypoxic conditions. NHE1 levels are significantly higher in BC tissue than in adjacent normal tissue, and also in resistant BC cells when compared to sensitive cells.	[8,25,41,55–71]

Table 1. Cont.

#### 4.3. Tumor Microenvironmental (TME) Acidosis and Immunity

There is a direct effect of tumor interstitial acidosis in hindering the antitumor immune response of the organism, another negative effect of the CPR. A complete review of the mechanisms by which tumor acidity disrupts the body immune defenses, locally and systemically, have been published by Huber et al. [20] (Table 1). These authors have shown how the acidity of the TME disrupts the immune defense mechanisms against malignant tumors, locally and systemically (Figure 1), enabling a relentless and uncontrolled tumor progression. Similar conclusions have been reached by other groups, relating aerobic glycolysis and lactic acid production with tumor invasion and even with MDR [126]. For all these reasons, the TME has been targeted by different methods in order to decrease, control and, if at all possible, revert, tumor extracellular acidity, both in animals and humans, in different malignant tumors. To this end, dietary lipids, PPIs or large daily amounts of sodium bicarbonate or other buffers have been used [51,158], occasionally with good results. The positive and antimetastatic effects of this strategy are secondary to the fact that acidity blocks T-cell activation and impairs tumor immunity [78]. Therapeutically, controlling TME acidity corrects T-cell dysfunction and allows to improve the efficacy of many other T-cell-based anticancer treatments [21,127,129]. A similar situation arises in lymph nodes, where activated T-cells are inhibited by acidosis [47]. The different methods to counteract TME acidity have been recently reviewed [22].

Most importantly, seminal research in this area by Marches et al. demonstrated that the anti-IgM-mediated induction of cell death in human B lymphoma cells is dependent on NHE1 inhibition and subsequent intracellular acidification. These findings do not appear to have been properly followed, in spite that they represent a synthesis of three different fields of modern oncology research: biochemistry, molecular biology and cancer immunity, all under one wide-ranged embracing unit [79].

## 4.4. Insulin (INS) and Breast Cancer

After all the experience accumulated on the carcinogenic effects of Na<sup>+</sup>/H<sup>+</sup> overstimulation and/or an elevated pHi, it can be concluded that any factor that up-regulates this antiporter may have a carcinogenic activity on its target cells (Table 1). Through its stimulating effects on glycolysis INS is one of these metabolic factors [35,145]. INS presents a direct effect in raising pHi, which at the same time increases glycolysis, and probably these are the two reasons for its known tumor-stimulating properties [144]. This is reasonable too, since hyperinsulinemia and obesity have been associated with an increased incidence of BC [139,140] (Table 1).

Cancer cells are also associated with INS insensitivity (resistance), due to high oxidative stress, especially during malignant transformation, and this could be an earlier event of carcinogenesis [141]. Recently published data show that behind the effect of INS on resistance appears to be an abnormality of the pH/NHE-1 signaling pathway [142], with NHE-1 over-expression as the first known key event of transformation in carcinogenesis [23,37]. Moreover, microenvironmental acidification and even systemic metabolic acidosis in cancer are linked with INS resistance [159,160], both phenomena being a reflection of the metabolic complications of cancer, the latter in advanced and disseminated disease [161–163]. For these reasons, some antidiabetic drugs like sulfonylureas, known to act by stimulating the pancreatic secretion of insulin, may have a negative impact on cancer growth [164–167]. In contrast, other antidiabetic agents, like Rosiglitazone and Metformin, show promising anticancer properties as INS-sensitizing agents [168,169] (Table 2). From a clinical perspective, it has also been shown that the over-expression of INS or the IGF-1 gene is associated with a decrease in the life span of women with BC, while their deletion improves life span and may also decrease tumorigenesis [143,146,147]. Finally, a recently published and highly interesting contribution has shown that insulin resistance might be a secondary effect of an abnormal NHE-1 signaling pathway [142] (Table 1).

**Table 2.** pH-related drugs with present and potential benefit in the treatment of breast cancer. BC: Breast cancer; NHE: Na<sup>+</sup>/H<sup>+</sup> antiporter; MDR: Multiple drug resistance; P-gp: P-glycoprotein; CAs: Carbonic anhydrases; ER<sup>+</sup>: Estrogen positive cells; ER-: Estrogen negative cells; PTIs: Proton transport inhibitors; PPIs; Proton pump inhibitors. TME: tumor microenvironment.

Drug	Summary	References
Amiloride (AM) (and/or liposomal amiloride), proton transport inhibitors (PTIs) and intracellular (IC) acidifiers	AM is a non-specific NHE inhibitor first introduced for human use as a $K^+$ sparing diuretic. It works as an antiangiogenic agent and has proved to be most effective as an antimetastatic drug in transplanted breast tumors in animals. A positive clinical experience in an occasional patient has been reported with its chronic use when traditional chemotherapy had failed. Also, the many anti-cancer effects of AM have been fully described. However, its utilization has not entered clinical trials in bedside oncology. (For further details, see the text).	[8,16,25,26,32,53,54,60,63, 64,67–69,79,82,88,101,110, 118,129,158,165,170–183]
Proton pump inhibitors (PPIs) and TME alkalization	PPIs are useful in the prevention of BC. Besides, the clinical utilization of V-ATPase inhibitors is a novel therapeutic measure to counteract the abnormal proton dynamics of BC and other tumors. PPIs also benefit from the microenvironmental acidity of tumors. Preclinical and clinical studies also support a direct anti-tumor effect of PPIs in BC and other solid tumors.	[22,49,51,67,78,85,128,130, 177,184–196]
Monocarboxylate transport (MCT) inhibitors	Quercetin is a pan-monocarboxylate transporter (MCT) inhibitor and intracellular acidifier. Liposomal quercetin is also available, since gastrointestinal absorption is very limited in the non-liposomal drug form.	[101,102,137,197–199]
Acetazolamide (AZM)	AZM is a carbonic anhydrase (CA) pan-inhibitor and cell acidifier. CAIX inhibition significantly reduces invasion of BC cells and represents a most promising drug in the treatment of BC, alone or in combination with different NHE inhibitors.	[63,65,165,183,200–203]
Doxorubicin (DOXO)	There is a progressive increase in resistance to DOXO by increasing elevations of pHi, resistance that is suppressed by P-gp inhibitors, while P-gp also increases pHi. MDR is characterized by a reversal of the pH gradient (cancer proton reversal or CPR) across cancer cell membranes.	[8,25,84,88–90,101,179,204]
Paclitaxel	The inhibition of NHE1, which is fundamental in the chemotherapy of triple-negative BC metastasis, improves the efficacy of Paclitaxel and mediates in Paclitaxel-induced apoptosis of BC cells.	[40,42,44,179,180,205–208]
Cis-platinum (CDDP)	The first effect of CDDP is the induction of intracellular acidification by inhibiting H+ extrusion secondary to downregulation of NHE-1. Contrariwise, the activity of NHE-1 and its effect on elevating pHi increases CDDP resistance to treatment.	[206,209–211]

Drug	Summary	References
Antiestrogens	ER- breast cancer cells have a higher expression of NHE activity and are preferably associated with CA-IX over CA-XII. Inhibition of CA-IX improves the prognosis of the disease. Although the role of Tamoxifen and Letrozole is well established, no further connections among pH dynamics and these antiestrogens have been described.	[4,5,154–157,200,212,213]
Melatonin (MT)	MT has an antiestrogenic effect and only for this reason it should be contemplated in BC therapy. Treatment with MT modulates tumor aggressiveness and increases apoptosis n BC cell lines. MT also suppresses tumor aerobic metabolism (the Warburg effect) and decreases breast cancer angiogenesis and metastasis.	[214-225]
Cariporide (CP)	CP (HOE-642) is a powerful NHE1 inhibitor that, unfortunately, is not available for clinical use in bedside oncology. It induces apoptotic cells death in different malignant tumors.	[8,25,35,170]
Compound 9t (C9t) (Unavailable)	C9t is 500-fold more potent against NHE1 than cariporide and has a 1400-fold greater selectivity for NHE1 over NHE2. Besides, C9t is orally bioavailable, has low side-effects in mice and it presents a significantly improved safety profile over other NHE1inhibitors.	[8,35,178]
Phx-3	Phx-3 is a potent, selective and non-toxic NHE1 inhibitor that triggers apoptosis in a variety of cancer cell lines and is highly effective in some animal tumor models.	[8,32,182]
Repurposed drugs	Phloretin, Lonidamine, Niclosamide, Docosaexaenoic acid, Salinomycin and Simvastatin have been reported to be useful in the treatment of BC because of their pH-related effects. Resveratrol also has a role as an aromatase inhibitor. (For further details, see the text).	[8,103,226–234]
Metformin (MET)	MET has been introduced as an anticancer agent in BC. It induces intracellular hyperacidification in tumor xenograft models. MET has been reported to inhibit insulin and insulin/IGF-1, HIF-1 $\alpha$ , Warburg metabolism, gene expression, angiogenesis, cancer migration, invasion and metastasis, apart from reducing the side effects of doxorubicin. MET has also been reported to act synergistically with chemotherapy and decrease its dosages, thus, its side-effects. It has also been used to target resistant cells in BC and has been considered a radio-sensitizer.	[235–255]

Table 2. Cont.

### 4.5. Prolactin and Breast Cancer

The role of prolactin (PRL) in the pathogenesis and progression of human BC is generally accepted [151]. Through NHE1 activation, this hormone stimulates growth, motility and invasiveness of BC, in this way contributing to the progression of the disease in a similar fashion that estrogens do [48,150] (Table 1). We agree with these authors that because of the effects of PRL, its inhibition should play a preventive, therapeutic and adjuvant role in the treatment of BC, as has already been suggested for other tumors [8].

Furthermore, there seems to exist a protumoral and synergistic interaction between PRL and growth hormone (GH) in stimulating the growth of certain tumors, BC among them [152]. In this vein, Clevenger et al. advanced that antagonists of PRL/PRL receptor interaction can be useful in the treatment of human BC, either alone or in combination with traditional antiestrogenic agents like tamoxifen and letrozole [151]. For all these reasons, PRL inhibitors such as bromocriptine or cabergolide (dopaminergic agonists drugs) should be taken into account as part of the armamentarium of repurposed drugs in BC therapy, even as a drug sensitizer [153].

### 4.6. Estrogens and Breast Cancer

Human BC is a heterogeneous disease classified in three major subtypes based on the expression of estrogen and progesterone receptors and human epidermal growth factor receptor-2 [256,257]. Among these BC subtypes, triple-negative BC results in a higher risk of metastatic dissemination and early death (Table 1).

Estrogens frequently play a crucial role in breast tumorigenesis by promoting cellular proliferation and decreasing apoptosis [154]. Interestingly, a recent study discussed why some tumors express ER<sup>+</sup> (estrogen receptors) and not ER- [155]. One of the suggested explanations is that while ER<sup>+</sup> tumor cells are highly vascularized ER- cells are better characterized by a higher expression of:

- (a) NHE1 activity.
- (b) Hypoxia-inducible Factor activity (HIF).
- (c) Carbonic Anhydrases (CAs) activity: CA-XII expression relies on estradiol activity [156]. Therefore, ER<sup>+</sup> is more likely to be associated with CA-XII rather than with CA-IX, while CA-IX is more frequently associated with ER- cells [212]. It has been shown that the selective inhibition of CA-XI improves the prognosis of the disease [200]. Although estrogens are growth factors, their effects or relations with the H<sup>+</sup> dynamics of BC cells have not been well established. However, ER-BC cells seem to be associated with NHE1 activity [157] (Table 1). ERs show a high degree of heterogeneity in BC [156], as first reported by Puddefoot et al. in 1993 [258], and further confirmed by Leclercq some years later [259]. This heterogeneity implies, among other aspects, that at least four isoforms of ER alpha may exist, migrating to different isoelectric points in isoelectric focusing gels. Whether one or more of these different isoelectric points may be related or even contribute to changes in the pHi of mammary cells leading to BC remains to be established.

### 4.7. Ion Channels

It has been previously shown that ion channels are an important factor in the etiopathogenesis of cancer and neurodegenerative diseases, both pathologies staying at opposite ends of a pH-related metabolic spectrum [8]. It is also been demonstrated that NaV1.5-Na<sup>+</sup> channels are in a close association with NHE-1, both being overexpressed in BC, where they contribute together to degrade the tumoral microenvironment, stimulate the formation of invadopodia and foster the metastatic process in a similar manner that CPR does [54,74–76,149] (Table 1). Furthermore, ion channels are activated at low microenvironmental tumor pH in BC and other tumors, thus promoting cell proliferation and migration. In this context, ion channels become relevant therapeutic targets [148].

### 4.8. PTs, PPs and Growth Factors

NHE1-overexpression is stimulated by a myriad of factors, all of which induce a pathological and carcinogenic elevation of pHi as a response of cells of many different origins and locations [25] (Table 1). Hence, the possibility of a cause–effect relationship between *BRCA1* and *BRCA2* genetic mutations in BC and pH/NHE1 and/or other PTs upregulation, has been recently pointed out by these authors. Among hormones, growth factors and cytokines that have been shown to be protumoral, either in BC or in other solid tumors, apart from estrogens, human growth hormone (hGH), prolactin, insulin and EGF and its receptor, are VEGF, PDGF, certain interleukins and sex steroids, some of which up-regulate NHE1 (Figure 1) [8].

To this already extensive list, PPs should be added, as well as certain oncogenes, virus and gene products such as Bcl-2 [124], a dysfunctional p53 and many chemical products known to play a role in carcinogenesis [80,82]. Other carcinogenic NHE-related factors are chronic hypoxia and the hypoxia inducible factor (HIF) [260]. Even high glucose loads stimulate Na<sup>+</sup>/H<sup>+</sup> activity [261].

It can be concluded that if so many unrelated etiopathogenic factors are known to be carcinogenic, the up-regulation of any of them, or several ones at the same time in a synergistic combination with other stimulators in the same direction, indicates that the pHi/pHe abnormalities exert their carcinogenic effect through the same acid-base intracellular (IC)/extracellular (EC), or pHi/pHe, deregulated dynamics. In addition, this suggests the existence of a universality of phenomenon involved in human carcinogenesis and cancer etiopathogenesis, BC being no exception to this rule [25].

#### 4.9. NHE1-Related Genetics of Breast Cancer

The Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1 (NHE1) is nowadays increasingly recognized as one of the most important factors involved in the etiology and pathogenesis of BC [38,40–43]. NHE1 has been found to be produced from the *APNH* gene located on chromosome 1p35-36, whose deletion has been blamed to be involved in the development of different tumors, BC among them [26]. Other genes have also been described to have a role in the genetic abnormalities behind BC metastasis [3]. These

authors have screened 4200 target genes and discovered 133 and 113 migratory modulators of Hs578T and MDA-MB-231 cells, which are predictive of BC progression and bad prognosis. Other genetic mutations, like *BRCA1* and *BRCA2*, are known to be strongly associated with familial breast and ovarian cancers [262]. The possibility that these two genetic abnormalities can be dependent on NHE1 hyperactivity has recently been proposed. However, no factual evidence is available at the present time that can ascertain such cause–effect relationships [25]. Finally, other pathways known to be involved in the pathogenesis of BC seem to act via different mechanisms and linked to other genes [263]. It is important to realize, as we have previously suggested, that in order to exert their role on cellular metabolism, genetic aberrations do so through the mediation of the microenvironmental changes they secondarily induce, and not directly on their own [264].

Since there is no formal proof of a relationship between H<sup>+</sup> -dynamics and *BRCA1* and *BRCA2* genetic abnormalities in BC, this important issue remains to be investigated. In the same line, it is most surprising the total lack of information relating inflammatory breast cancer (IBC) with PTs, PPs and H<sup>+</sup>-dynamics. Despite the known importance of TME in this aggressive kind of BC, the few reviews available on TME in IBC completely ignore such more than possible relationships or any other reference to the H<sup>+</sup>-related paradigm in BC and IBC etiopathogenesis [265].

# 5. Hydrogen Ion Dynamics in Multiple Drug Resistance (MDR) in Breast Cancer and Other Malignant Tumors: An Integral Approach to Its Etiopathogenesis and Mediating Mechanisms

Resistance of BC cells to drugs like doxorubicin (DOXO), paclitaxel and cis-platinum depends on pH regulation [89,125,179] (Table 2). DOXO resistance is related to a progressive increase of pHi, and could be suppressed by the addition of P-glycoprotein inhibitors like verapamil [204]. These findings, apart from showing the close relationship of P-gp and pHi, have made it possible to conclude that P-gp behaves as a proton (H<sup>+</sup>) extrusion pump [84].

Levels of the proton transporter (PT) NHE1 are significantly higher in BC and in resistant cancer cells when compared to adjacent normal tissues and selective cells [165]. A proton transport inhibitor (PTI) like cariporide (CP, HOE-642), induces apoptosis in MCF-7/ADR cells in vitro and in vivo and is associated with the intracellular accumulation of DOXO and G0/G1 cell cycle arrest. CP also induces tumor growth attenuation and diminishes tumor volume. This strongly suggests that NHE1 should be a promising adjuvant therapeutic target, not only in BC but also in a wide array of other malignant tumors [170]. Other PTs, like the  $HCO_3^-$ -cotransporter NBCn1 (Slc4a7), show similar effects, delaying BC development [58]. Finally, the association of PTIs with PPIs of the omeprazole family offers the possibility of further improving the effects of chemotherapy in metastatic BC [187], as well as in other malignant tumors [177]. Bringing all these findings together, it can be concluded that the association of PTIs and PPIs can have a synergistic effect in overcoming MDR in BC, apart from having a strong antitumoral effect on their own, from prevention to the treatment of advanced disease.

### 6. pH-Related Armamentarium in the Treatment of Breast Cancer

#### 6.1. NHE Inhibitors and/or Intracellular Acidifiers

A fundamental therapeutic aim of the pH-related anticancer paradigm is directed to the concerted inhibition of NHE1 and other PTs and PPs in order to induce a progressively deep intracellular acidification (IA), which first would decrease glycolysis, then lead to tumor cell growth arrest, and finally induce selective apoptosis [54,176,266,267].

Amiloride (AM) was the first Na<sup>+</sup>/H<sup>+</sup> antiporter introduced in medical oncology. However, AM is a weak and non-specific Na<sup>+</sup>/H<sup>+</sup> antiporter inhibitor, but at least it is still commercially available as a diuretic (Table 2). Seminal research with AM showed a complete inhibition of lung metastasis of BC in animals. Such an effect was initially reported to be secondary to its inhibitory effects on urokinase-type plasminogen activator (UPa) [175]. AM has clearly shown antitumoral, antiangiogenic and antimetastatic effects [268,269]. Also, when decreasing pHi values with AM, VEGF mRNA expression and tumor growth are inhibited, at least in gastric and leukemic cells [17,172]. An occasional patient with advanced cancer has benefited for the chronic utilization of AM after mainstream chemotherapy had failed [171]. Finally, for a review on the positive effects of AM on basic cell behavior, see [118].

Among the more modern, potent and selective NHE1 inhibitors, cariporide (CP, HOE-642) induces apoptosis through lowering pHi [174]. CP also decreases angiogenesis and induces selective apoptosis mediated through Na<sup>+</sup>/H<sup>+</sup> exchange inhibition [170]. Furthermore, CP induces apoptosis by overcoming paclitaxel resistance through NHE inhibition [180], and works synergistically with erlotinib in reducing metastasis in pancreatic cancer [208].

An even more potent NHE1 inhibitor than CP is the so called "Compound 9t" (C9t). C9t is five hundred times more potent against NHE1 than CP, apart from having a 1400-fold greater selectivity for NHE1 over NHE2 [178] (Table 2). Similarly, Phx-3, is a non-toxic NHE1 inhibitor that through selective apoptosis and intracellular acidification induces tumor growth regression after leukemic cell transplantation [32,182].

All available data indicate that these new and selective NHE1 inhibitors have a great potential as potent and selective anticancer agents in patients with different pathologies [170]. Additionally, it is difficult to understand why AM itself has not managed to find a place in bedside oncology, mainly in BC treatment, either as a preventive measure, as a complement of orthodox chemotherapy and/or as an antimetastatic factor. There can be two reasons for this: a) AM is not patentable, and b) in all areas of scientific research and medical life, sometimes the answers that are looked for in the future are waiting hiding in the past (Table 2).

### 6.2. Proton Pumps (PPs) and Their Inhibitors (PPIs) in Cancer and Breast Cancer

The over-expression of proton pump V-ATPases promote growth advantages to cancer cells of any origin, also disrupting pH homeostasis in the same CPR line of action [69,270]. Different publications have illuminated the crucial role of V-ATPases in tumor invasion and chemoresistance in several cancers, including BC. Therefore, as it happens with PTs inhibition, PPs inhibition of V-ATPases has recently become a novel therapeutic avenue for thwarting the highly abnormal H+ dynamics either in BC but also in other tumors of different origins and locations [15,49,194] (Table 2) Pretreatment with PPIs strongly enhances the in vitro efficacy of chemotherapeutic drugs against human BC cells and other malignancies [177,188,195,271,272]. Niikura showed that oral administration of a V-ATPase inhibitor to SCID mice carrying orthotropic BC xenografts resulted in delayed tumor growth and a decrease in bone metastasis [189]. Another in vitro report has shown the therapeutic effectiveness of PPIs in triple negative BC cell lines [69]. Finally, different preclinical studies support a direct anti-tumor effect of PPIs independently from cancer histology [190–192,273].

Two studies have been conducted in pets with spontaneous neoplasms using PPIs in combination with standard chemotherapy. The first one evaluated the ability of high dose lansoprazole to reverse chemoresistance in dogs and cats with cancers not responding to chemotherapy. In this study, the drug was used off-label with a three-day loading dosage of 5 mg/kg followed by a four-day maintenance regimen at 1 mg/kg as a chemosensitizer, combined with standard veterinary chemotherapy protocols. The results showed a reversal of chemoresistance in 23 out of 34 treated animals (67% response rate) (Table 2). A further study combined PPI with water alkalization and metronomic chemotherapy [184]. The cohort receiving alkalization showed enhanced tumor response, both in terms of the number of responders and the quality of response, when compared to the group receiving metronomic chemotherapy alone.

The application of this strategy to humans has led to the publication of two clinical trials. These studies evaluated the tolerability and effectiveness of high-dose PPIs in patients with osteosarcoma or metastatic BC [187,196]. One of these studies showed that the addition of PPIs to chemotherapy increased the effectiveness of chemotherapy in osteosarcoma patients. The second trial recruited women with metastatic BC that were randomized to receive either conventional chemotherapy or

chemotherapy with alkalization [187]. In the latter study, patients receiving high dosage PPIs obtained the highest response rates and the longest survivals. Furthermore, there is a statistically significant survival advantage for women who continued their proton pump inhibition therapy (PPIT) after the completion of chemotherapy [271]. This in vivo data are also supported by in vitro investigations showing the effectiveness of PPIs in triple negative BC cell lines [69].

An indirect confirmation of the validity of PPIs and alkaline therapy as antitumor agents was provided by a published meta-analysis in head and neck tumor patients that found a better outcome for patients receiving PPIs [274]. A more recent study describes the outcome of a few patients with metastatic colorectal cancer who were refractory to standard chemotherapy and targeted therapies. The combination of high-dose rabeprazole (a PPI inhibitor) and metronomic capecitabine overcame drug resistance [275]. Despite of the very small number of patients studied, the association of rabeprazole and capecitabine resulted in long-lasting stable disease with good quality of life and relatively minor side effects.

More recent reports have shown that women affected by medical conditions suitable for PPIT treatment, e.g., peptic diseases, have a reduced risk for developing BC [67,194]. Both studies were performed in a very large cohort of patients with very convincing results. The data obtained also show that the beneficial effects of PPIT increases with age, the BC risk being reduced to a greater extent in older PPIT users, getting to 83.0% in the 50 to 64 years old cohort. These data are of particular importance in women with a higher risk of developing BC, like those with a family history of BC, as well as women treated with long term hormone replacement therapy during and after menopause. These studies also support the results obtained in women with metastatic BC, particularly those with triple negative BC when exclusively treated with high dosage PPIs, while intermittent high dose PPIs also enhances the antitumor effects of chemotherapy in metastatic BC [187]. Another three articles recently suggested reconsidering the use of PPIs in cancer therapy [186,276,277]. Their conclusions are highly convincing and important for the treatment of a disease like BC that is becoming an increasing killer worldwide.

Finally, since PPIs are prodrugs requiring activation in the acidic microenvironment of solid tumors, they appear particularly suitable to be used as anticancer drugs in the very acidic tumor microenvironment [22], this obviously being the case of BC as well. Moreover, while it is not entirely scientifically supported, it appears conceivable that PPIs may also affect the systemic pH by buffering the stomach, that is their main target [185].

### 6.3. Melatonin (MT) in Breast Cancer

Melatonin (MT) has been shown to function as an antiestrogenic agent, and only for this reason should be strongly considered for BC treatment [215,219] (Table 2). Sonehara et al. showed that treatment with MT modulates tumor aggressiveness, increasing apoptosis under microenvironmental acidosis in BC cell lines [220]. Importantly, MT has been shown to be a significant antiangiogenic agent by downregulating VEGF expression in human BC cells [214,221,225]. Previous studies have also indicated that MT suppresses tumor aerobic metabolism (Warburg effect) and cell-signaling pathways that are key for the proliferation and survival of BC cells as well as for metastasis and resistance to anti-cancer drugs [215,222,223,278]. MT treatment of breast tumor cells decreases the HIF-1 $\alpha$  gene, HIF-1 $\alpha$  expression and regulates glucose metabolism as well as the expression of protumoral factors like GLUT1, GLUT3, CA-IX and CA-XII, indicating that MT controls hypoxia and tumor progression [279].

Otherwise, traditional oncology has clearly shown that antiestrogens are an effective measure in the treatment of ER + BC, using either Tamoxifen or Letrozole. In this vein, MT, as well as resveratrol, also appears to function as aromatase inhibitors, so becoming further candidates in the adjuvant treatment of ER + BC [226]. In addition to these mechanisms, MT shows many other anticancer and oncostatic effects in BC [216]. Among them, MT enhances the sensitivity to classical anti-cancer agents [215].

Interestingly, a recent in vitro study provides evidence about the positive effects of a novel MT-TMX drug combination in the treatment of BC. It is known that TMX use may eventually lead to resistance. However, this seems to be overcome by the novel MT-TMX conjugates [217]. Furthermore, an inverse correlation between nocturnal melatonin levels and the development of BC has been confirmed [213,219]. This appears to be related to the loss of the day/night MT circadian rhythm, increasing the risk of BC development in female night workers. In fact, it has been shown that women with BC had lower plasma levels of MT than normal women, and these levels are even lower in nurses working shifts [218].

In summary, it is beyond doubt that MT plays an important role in the prevention and treatment of BC. Although its primary effect seems to be exerted at a mitochondrial level by regulating aerobic metabolism, MT also decreases angiogenesis and proliferation while promoting apoptosis [214,215,220,221,224].

### 6.4. Cisplatin (CDDP) and pH/NHE in Breast Cancer

Cisplatin (CDDP) has been used in the treatment of BC and other malignant tumors for a long time, and it continues to be widely used nowadays [205,206,280]. From its first introduction in bedside oncology, different and unrelated mechanisms of action for CDDP have been described [210,281]. Most importantly, an almost completely disregarded issue has been the fact that CDDP can significantly modify the intracellular pH of cancer cells by inducing cytoplasmatic acidification through a CDDP-mediated inhibition of H<sup>+</sup> extrusion secondary to downregulation of NHE-1 [206,209–211] (Table 2). Most recently, this has been considered the first effect of CDDP [209]. Contrariwise, the activity of NHE-1 and its effect on elevating pHi increases CDDP resistance to treatment [209]. Apart from inducing pHi acidification, CDDP shifts cervical cancer cells from glycolysis to oxidative metabolism, and this is accompanied by inhibition of cancer cell growth. In these studies, cancer cells either recover, maintaining an alkaline pHi to survive and proliferate, although at reduced growth rates, or die [282].

### 6.5. pH and MDR in Breast Cancer: An Integrated Approach to Treatment

For many years, P-glycoprotein (P-gp) has been held to be the main responsible mechanism for multidrug resistance (MDR) in solid tumors. However, seminal research in this area initially showed that a progressive increase in pHi was correlated with the level of doxorubicin (DOXO) resistance in human lung tumor cells. In this case, drug resistance was counteracted upon the addition of verapamil, an inhibitor of P-gp activity [204]. At that time, the fact that P-gp affected pHi was already suggested. Furthermore, during the last two decades pH alterations have been shown to be behind the most fundamental aspects of MDR [84,90,130,283,284]. Indeed, it has been recently shown that P-gp needs a pH gradient in order to function [86,285] and an integrated mechanism to explain MDR has been developed based upon the H<sup>+</sup>-dynamics of the microenvironment of tumors (CPR) [49,84,86,89,130,131,179,286]. This new and integral model demonstrates that in MDR the CPR of cancer cells and tissues and the P-gp expression are related in a direct cause-effect relationship and cannot be separated from each other.

This integral approach to MDR has proposed that the therapeutic failure to induce cytoplasmic acidification and/or reverse CPR is the main underlying factor for drug resistance, which suggests that MDR and resistance to the induction of the low pHi-mediated therapeutic apoptosis are also one and the same thing. Furthermore, the expression of P-gp leads to an elevation of pHi [87,287], while intracellular acidification down-regulates the MDR transporter [87,288,289]. Finally, extracellular acidification increases the activity of P-gp, inducing MDR in different cancer cells and tissues [49,130,134]. In summary, all these findings perfectly fit into each other, meaning that the therapeutic induction of a reversion of CPR is also the key and fundamental target in overcoming MDR, probably in all malignant tumors, like it is in pH-related cancer treatment. This is in line with all the other integrations made possible when approached through the all-comprehensive pH-centric anticancer paradigm [8,24,133].

### 6.6. Repurposed Drugs in Breast Cancer Treatment

TME acidification makes BC more aggressive [135,290]. This intratumoral but extracellular acidosis, mainly caused by lactic acid production, is related to an increased aerobic glycolysis (Warburg effect) (Figure 1), being fundamental in promoting invasiveness of BC cells [136]. On the contrary, chronic administration of sodium bicarbonate to nude mice implanted with human BC reduces the number and size of metastases [51] (Table 2).

Among the many mechanisms responsible for the regulation of the protoplasmic acid-base balance (Carbonic anhydrases (CAs), Monocarboxylate Transporters (MCT), ATP synthase, V-ATPases and Na<sup>+</sup>/H<sup>+</sup> exchanger isoform 1(NHE1), CAIX appears to be a critical mediator for the expansion of BC in hypoxic niches, sustaining the mesenchymal and 'stemness' phenotypes of these cells [202]. CAIX activity affects the uptake and toxicity of anticancer drugs and is associated with a bad prognosis. Also, Erb-2 expression is associated with bad prognosis [201]. The CA inhibitor Acetazolamide (AZM) enhances DOXO toxicity but reduces Melphalan toxicity in BC cell lines that express CAIX, which is also a target for BC anticancer treatment [101]. In hypoxic BC tumor cells, the inhibition of different H+-extruding mechanisms has been proposed as a therapeutic strategy, while among them CAIX is considered to represent the most promising target [203].

Furthermore, V-ATPase and MCT4 are both major microenvironmental acidifying mechanisms in human BC cell lines [137]. Indeed, MCTs are often upregulated in BC tissue [197], while MCT4 is a clear therapeutic target, at least in certain subtypes of BC [59]. Thus, it is logical that targeting lactate transport with MCT inhibitors such as Quercetin suppresses BC growth and improves tumor immune response [102,181,198,291]. Other MCT inhibitors such as Simvastatin and Phloretin have also been found to be active against BC cells [103,227] (Table 2).

Lonidamine was first introduced in 1979 as an antispermatogenic agent. It inhibits L-lactate transport through activity on MCT1, MCT2 and MCT4, causing selective intracellular acidification of tumors. It has been active in metastatic BC patients, but is not commercialized any more [230].

The n-3 polyunsaturated Docosahexaenoic acid (DHA, 22:6n-3), is effective in increasing survival and chemotherapy efficacy in BC patients with metastasis [231], inhibiting NaV1.5 current and NHE-1 activity in human BC cells [232]. The daily doses used in the clinical trials were in the range of 1,800 mg DHA/day, while a single case report showed a positive result in a BC patient that only used 480 mg DHA/day as part of a more extensive supplementary cocktail using a multitargeted approach [228].

Drug screening has identified an FDA approved drug, Niclosamide as an inhibitor of BC stem-like cells [233]. Another group of compounds known to induce cytosolic acidification are the K+ ionophores. These compounds promote the outflow of K<sup>+</sup> from the mitochondria as well as from the cytoplasm, mediating an H<sup>+</sup>/K<sup>+</sup> exchange across lipid membranes. The result is the induction of an intracellular accumulation of protons [8]. One such K<sup>+</sup> ionophore is the antibiotic Salinomycin. Promising results from a few clinical pilot studies indicate that Salinomycin is able to induce partial clinical regression of heavily pretreated and therapy-resistant cancers, including BC [234] (Table 2). Finally, for a more complete exposure of repurposed drugs in preclinical and clinical oncology, see [8].

### 6.7. Metformin (MET) in Breast Cancer

Boosting glycolysis with mitochondria inhibitors such as Metformin (MET) have been proposed as a method to decrease pHi in various cancer cell lines, BC among them [52], alone and/or in combination with the MCT inhibitor Simvastatin [53] (Table 2). MET has been considered to be a viable anticancer drug since it induces intracellular hyperacidification in tumor xenograft models through inhibition of Wnt signaling, a feature found to be selective for cancer cells [52]. MET was introduced as an anticancer agent in clinical oncology after it was reported to decrease mortality of BC patients [235,236,245–247], increasing the survival of triple-negative BC patients [247]. MET can act as an anticancer drug through its activity on several glucose transporters [292] known to be associated with BC [237,238,293,294]. Some of the anticancer-related effects of MET are:

- (i) It reduces circulating insulin and insulin/IGF-1 receptor-mediated activation of the PI3K pathway [245].
- (ii) MET inhibits the expression of the Hypoxia Inducible Factor 1 alpha (HIF-1 $\alpha$ ) gene expression, increases Pyruvate Dehydrogenase (PDH) gene expression [249] and decreases Warburg metabolism [35]. Additionally, HIF-1 $\alpha$  is fundamental in tumoral angiogenesis and induces the expression of VEGF in BC [134,252,260,295]. Through this and other mechanisms MET also inhibits cancer growth, including triple-negative BC (TNBC) [254]. VC REMOVE:
- (iii) As an inhibitor of tumor angiogenesis [239,248,250,251], a recent study revealed the impact of MET inhibitory effect on microvasculature [296]. Through this antiangiogenic effect MET can also improve tumor prognosis [155,252].
- (iv) MET can reduce tumor progression through AMPK inactivation [297,298], although the opposite effect has also been reported [299].
- (v) MET can inhibit cancer migration, invasion and metastasis in BC and other tumors [240,300–303].
- (vi) MET is also active via the inhibition of the hedgehog signaling pathway in tumors like BC [241,242].

Finally, MET has not only been used as an anticancer agent by itself, but is also useful as an adjuvant to other cancer chemotherapy agents, particularly being able to reduce the side-effects of DOXO [304]. Moreover, MET has been reported to act synergistically with chemotherapy, also decreasing its dosages [243], and has even been used to target resistant cells in BC and other tumors [244]. MET has even been considered to be a radio-sensitizer agent [255].

### 6.8. Autophagy and Cannibalism in Breast Cancer

Autophagy; however still a controversial issue, has been shown to represent an adaptive survival mechanism to overcome drug-induced cellular stress and cytotoxicity. This was demonstrated using PPIs, which induced early accumulation of autophagosomes, also reducing the autophagic flux. Notably, the inhibition of autophagy by knockdown of Atg5 and Beclin-1 expression significantly increased PPIs cytotoxicity [305], suggesting that autophagy may exert a protective role in cancer cells treated with PPIs, and that inhibiting the autophagic process may lead to higher cytotoxic effects and improve therapeutic efficacy. Furthermore, TME acidic conditions increase autophagy in cancer, suggesting that it may protect tumor cells allowing them to survive under microenvironmental acidic stress [96]. This was shown in BC cell lines [97], suggesting that PPIs can help in improving anticancer therapeutic efficacy in a broad spectrum of cancers.

In the same line, Salinomycin exerts a potent inhibition activity of the autophagic flux, mainly when cells are cultured in acidic conditions. This has been shown in tumor cells obtained from BC patients [98]. Functioning as a potassium ionophore, Salinomycin has been shown to contribute in further inhibiting lysosomal degradation, supporting the idea of using a broad panel of other ion inhibitors in modulating autophagy in cancer [306]. From these studies it appears that H<sup>+</sup>/K<sup>+</sup> ATPases or V-ATPases may represent preferential targets to inhibit autophagy. However, from other studies it appears that the mechanism malignant tumor cells use to face off nutrient starvation, in acidic conditions as well, is cannibalism [307,308]. Cannibalism has been shown in BC as well [309], implying that autophagy may exert a different role in cancer cells than a mechanism of self-feeding in nutrient low supply conditions, inasmuch BC cells feed on other cells that within the tumor are in fact siblings cells, i.e., cells from the same individual. This implies the existence of an entirely new phenomenon than can be called self-cannibalism, or even auto-cannibalism, and that may be considered a specific characteristic of malignant tumors [308]. In fact, it has been shown that self-cannibalism is a feature of cells mainly derived from metastatic lesions rather than from the primary tumor [307].

# 7. Conclusions

If many unrelated etiopathogenic factors of so many different natures and origins cause cancer, the upregulation of any or several of them, alone or in a synergistic combination with other stimulators

of cellular hydrogen ion ( $H^+$ ) extrusion, indicates that the pHi/pHe abnormalities selectively induced in cancer cells and tissues mediate in the behavior of malignant tumors like breast cancer in all its phases of development. This also suggests the existence of a universality of phenomenon involved in the carcinogenesis of breast cancer and other human tumors. In this contribution, the multifactorial etiological and etiopathogenetic factors in breast cancer are considered all together, which allows to propose an integrated and unidirectional approach to the therapeutics of this deadly disease. It is also shown that all the areas and hallmarks of cancer research, perhaps with the exception of radiotherapy, can be integrated under the same Unitarian acid-base paradigm. Interestingly, there is a surprising lack of information relating to the pH/NHE-related paradigm and genetic abnormalities like *BRCA1* and *BRCA2* positive tumors and of H<sup>+</sup>-dynamics with inflammatory breast cancer, despite the known importance of tumor microenvironment in this aggressive form of the disease.

The ultimate goal of this integral approach to malignancy is to target the selective molecular and metabolic-dependent acid-base disruptions specific to cancer cells using different and concerted synergistic methods. The final aim of therapy is to take advantage of the H<sup>+</sup>-related selective abnormalities that cancer cells and tissues possess over their normal counterparts, in order to exploit such differences in treatment. It is concluded that any attempts to induce a low pHi-mediated apoptosis can be the cancer-specific and fundamental strategy to treat breast cancer as well as other human malignant tumors. The pH-centric anticancer paradigm recognizes that any attempts to selectively induce an intracellular hyperacidification incompatible with the life of cancer cells, and/or reverting cancer proton reversal (CPR), are its main therapeutic targets. The pending issue nowadays is to find that old Ehrlich's magic bullet that can selectively achieve that, if such a weapon exists. Otherwise, the concerted utilization of some of the measures conceptualized and described here, is likely to become a useful and integrated alternative, both nowadays and in the near future, to more efficiently treat all forms of breast cancer.

**Author Contributions:** Conceptualization, S.H., K.A., D.S., S.F. and J.D.; methodology, J.P.O.; validation, S.H., K.A., Y.Y., and J.D.; formal analysis, S.H.; investigation, S.H., J.P.O. and S.F.; writing—original draft preparation, S.H.; writing—review and editing, S.H., K.A., K.H., D.S., S.F. and J.D.; funding acquisition, J.P.O., S. H. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was funded by The Mercedes Castresana Foundation (Vitoria, Spain) (S.H.), and the Association for Proton Cancer Research and Treatment (APCRT), Madrid, Spain (J.P.O., S.H.).

**Conflicts of Interest:** The authors declare no conflict of interest. The funder had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

### References

- Evans, D.G.; Howell, A. Breast cancer risk-assessment models. *Breast Cancer Res.* 2007, 9, 213. [CrossRef]
  [PubMed]
- Redig, A.J.; McAllister, S.S. Breast cancer as a systemic disease: A view of metastasis. J. Intern. Med. 2013, 274, 113–126. [CrossRef] [PubMed]
- Koedoot, E.; Fokkelman, M.; Rogkoti, V.M.; Smid, M.; van de Sandt, I.; de Bont, H.; Pont, C.; Klip, J.E.; Wink, S.; Timmermans, M.A.; et al. Uncovering the signaling landscape controlling breast cancer cell migration identifies novel metastasis driver genes. *Nat. Commun.* 2019, 10, 2983. [CrossRef] [PubMed]
- 4. Samavat, H.; Kurzer, M.S. Estrogen metabolism and breast cancer. Cancer Lett. 2015, 356, 231–243. [CrossRef]
- Early Breast Cancer Trialists' Collaborative Group. Effects of chemotherapy and hormonal therapy for early breast cancer on recurrence and 15-year survival: An overview of the randomised trials. *Lancet* 2005, 365, 1687–1717. [CrossRef]
- 6. Egger, S.J.; Willson, M.L.; Morgan, J.; Walker, H.S.; Carrick, S.; Ghersi, D.; Wilcken, N. Platinum-containing regimens for metastatic breast cancer. *Cochrane Database Syst. Rev.* **2017**, *6*, CD003374. [CrossRef]
- Casey, J.R.; Grinstein, S.; Orlowski, J. Sensors and regulators of intracellular pH. *Nat. Rev. Mol. Cell Biol.* 2010, 11, 50–61. [CrossRef]

- 8. Harguindey, S.; Stanciu, D.; Devesa, J.; Alfarouk, K.; Cardone, R.A.; Polo Orozco, J.D.; Devesa, P.; Rauch, C.; Orive, G.; Anitua, E.; et al. Cellular acidification as a new approach to cancer treatment and to the understanding and therapeutics of neurodegenerative diseases. *Semin. Cancer Biol.* **2017**, *43*, 157–179. [CrossRef]
- 9. Harguindey, S. Hydrogen ion dynamics and cancer: An appraisal. *Med. Pediatr. Oncol.* **1982**, *10*, 217–236. [CrossRef]
- Pouyssegur, J.; Sardet, C.; Franchi, A.; L'Allemain, G.; Paris, S. A specific mutation abolishing Na+/H+ antiport activity in hamster fibroblasts precludes growth at neutral and acidic pH. *Proc. Natl. Acad. Sci. USA* 1984, *81*, 4833–4837. [CrossRef]
- 11. Moolenaar, W.H. Effects of growth factors on intracellular pH regulation. *Annu. Rev. Physiol.* **1986**, *48*, 363–376. [CrossRef]
- 12. L'Allemain, G.; Paris, S.; Pouyssegur, J. Growth factor action and intracellular pH regulation in fibroblasts. Evidence for a major role of the Na+/H+ antiport. *J. Biol. Chem.* **1984**, 259, 5809–5815. [PubMed]
- 13. Harguindey, S. The Hydrogen Ion (pH) and its Implications in Cancer. Ph.D. Thesis, University of The Basque Country, Bilbao, Spain, 1989.
- 14. Lagadic-Gossmann, D.; Hardonniere, K.; Mograbi, B.; Sergent, O.; Huc, L. Disturbances in H(+) dynamics during environmental carcinogenesis. *Biochimie* **2019**, *163*, 171–183. [CrossRef] [PubMed]
- Spugnini, E.P.; Sonveaux, P.; Stock, C.; Perez-Sayans, M.; De Milito, A.; Avnet, S.; Garcia, A.G.; Harguindey, S.; Fais, S. Proton channels and exchangers in cancer. *Biochim. Biophys. Acta* 2015, *1848*, 2715–2726. [CrossRef] [PubMed]
- 16. Orive, G.; Reshkin, S.J.; Harguindey, S.; Pedraz, J.L. Hydrogen ion dynamics and the Na+/H+ exchanger in cancer angiogenesis and antiangiogenesis. *Br. J. Cancer* **2003**, *89*, 1395–1399. [CrossRef] [PubMed]
- 17. He, B.; Deng, C.; Zhang, M.; Zou, D.; Xu, M. Reduction of intracellular pH inhibits the expression of VEGF in K562 cells after targeted inhibition of the Na+/H+ exchanger. *Leuk. Res.* **2007**, *31*, 507–514. [CrossRef]
- Hardonniere, K.; Huc, L.; Sergent, O.; Holme, J.A.; Lagadic-Gossmann, D. Environmental carcinogenesis and pH homeostasis: Not only a matter of dysregulated metabolism. *Semin. Cancer Biol.* 2017, 43, 49–65. [CrossRef]
- 19. Aravena, C.; Beltran, A.R.; Cornejo, M.; Torres, V.; Diaz, E.S.; Guzman-Gutierrez, E.; Pardo, F.; Leiva, A.; Sobrevia, L.; Ramirez, M.A. Potential role of sodium-proton exchangers in the low concentration arsenic trioxide-increased intracellular pH and cell proliferation. *PLoS ONE* **2012**, *7*, e51451. [CrossRef]
- 20. Huber, V.; Camisaschi, C.; Berzi, A.; Ferro, S.; Lugini, L.; Triulzi, T.; Tuccitto, A.; Tagliabue, E.; Castelli, C.; Rivoltini, L. Cancer acidity: An ultimate frontier of tumor immune escape and a novel target of immunomodulation. *Semin. Cancer Biol.* **2017**, *43*, 74–89. [CrossRef]
- 21. Lacroix, R.; Rozeman, E.A.; Kreutz, M.; Renner, K.; Blank, C.U. Targeting tumor-associated acidity in cancer immunotherapy. *Cancer Immunol. Immunother.* **2018**, *67*, 1331–1348. [CrossRef]
- 22. Pillai, S.R.; Damaghi, M.; Marunaka, Y.; Spugnini, E.P.; Fais, S.; Gillies, R.J. Causes, consequences, and therapy of tumors acidosis. *Cancer Metastasis Rev.* **2019**, *38*, 205–222. [CrossRef] [PubMed]
- 23. Cardone, R.A.; Alfarouk, K.O.; Elliott, R.L.; Alqahtani, S.S.; Ahmed, S.B.M.; Aljarbou, A.N.; Greco, M.R.; Cannone, S.; Reshkin, S.J. The Role of Sodium Hydrogen Exchanger 1 in Dysregulation of Proton Dynamics and Reprogramming of Cancer Metabolism as a Sequela. *Int. J. Mol. Sci.* **2019**, 20. [CrossRef] [PubMed]
- 24. Harguindey, S.; Reshkin, S.J. "The new pH-centric anticancer paradigm in Oncology and Medicine"; SCB, 2017. *Semin. Cancer Biol.* **2017**, *43*, 1–4. [CrossRef] [PubMed]
- 25. Harguindey, S.; Polo Orozco, J.; Alfarouk, K.O.; Devesa, J. Hydrogen Ion Dynamics of Cancer and a New Molecular, Biochemical and Metabolic Approach to the Etiopathogenesis and Treatment of Brain Malignancies. *Int. J. Mol. Sci.* **2019**, *20*. [CrossRef]
- 26. Rich, I.N.; Worthington-White, D.; Garden, O.A.; Musk, P. Apoptosis of leukemic cells accompanies reduction in intracellular pH after targeted inhibition of the Na(+)/H(+) exchanger. *Blood* **2000**, *95*, 1427–1434. [CrossRef]
- 27. Perona, R.; Serrano, R. Increased pH and tumorigenicity of fibroblasts expressing a yeast proton pump. *Nature* **1988**, *334*, 438–440. [CrossRef]
- 28. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. Cell 2011, 144, 646–674. [CrossRef]
- 29. Fouad, Y.A.; Aanei, C. Revisiting the hallmarks of cancer. Am. J. Cancer Res. 2017, 7, 1016–1036.
- 30. Warburg, O. On the origin of cancer cells. *Science* 1956, 123, 309–314. [CrossRef]
- 31. Warburg, O.H. The Prime Cause and Prevention of Cancer; Nobel Laureates: Lindau, Germany, 1969.

- 32. Nagata, H.; Che, X.F.; Miyazawa, K.; Tomoda, A.; Konishi, M.; Ubukata, H.; Tabuchi, T. Rapid decrease of intracellular pH associated with inhibition of Na+/H+ exchanger precedes apoptotic events in the MNK45 and MNK74 gastric cancer cell lines treated with 2-aminophenoxazine-3-one. *Oncol. Rep.* 2011, 25, 341–346. [CrossRef]
- Persi, E.; Duran-Frigola, M.; Damaghi, M.; Roush, W.R.; Aloy, P.; Cleveland, J.L.; Gillies, R.J.; Ruppin, E. Systems analysis of intracellular pH vulnerabilities for cancer therapy. *Nat. Commun.* 2018, *9*, 2997. [CrossRef] [PubMed]
- 34. Quach, C.H.; Jung, K.H.; Lee, J.H.; Park, J.W.; Moon, S.H.; Cho, Y.S.; Choe, Y.S.; Lee, K.H. Mild Alkalization Acutely Triggers the Warburg Effect by Enhancing Hexokinase Activity via Voltage-Dependent Anion Channel Binding. *PLoS ONE* **2016**, *11*, e0159529. [CrossRef] [PubMed]
- 35. Alfarouk, K.O.; Verduzco, D.; Rauch, C.; Muddathir, A.K.; Adil, H.H.; Elhassan, G.O.; Ibrahim, M.E.; David Polo Orozco, J.; Cardone, R.A.; Reshkin, S.J.; et al. Glycolysis, tumor metabolism, cancer growth and dissemination. A new pH-based etiopathogenic perspective and therapeutic approach to an old cancer question. *Oncoscience* **2014**, *1*, 777–802. [CrossRef] [PubMed]
- 36. Witkiewicz, A.K.; Whitaker-Menezes, D.; Dasgupta, A.; Philp, N.J.; Lin, Z.; Gandara, R.; Sneddon, S.; Martinez-Outschoorn, U.E.; Sotgia, F.; Lisanti, M.P. Using the "reverse Warburg effect" to identify high-risk breast cancer patients: Stromal MCT4 predicts poor clinical outcome in triple-negative breast cancers. *Cell Cycle* 2012, *11*, 1108–1117. [CrossRef] [PubMed]
- 37. Reshkin, S.J.; Bellizzi, A.; Caldeira, S.; Albarani, V.; Malanchi, I.; Poignee, M.; Alunni-Fabbroni, M.; Casavola, V.; Tommasino, M. Na+/H+ exchanger-dependent intracellular alkalinization is an early event in malignant transformation and plays an essential role in the development of subsequent transformation-associated phenotypes. *FASEB J.* 2000, *14*, 2185–2197. [CrossRef] [PubMed]
- 38. Grillo-Hill, B.K.; Choi, C.; Jimenez-Vidal, M.; Barber, D.L. Increased H(+) efflux is sufficient to induce dysplasia and necessary for viability with oncogene expression. *Elife* **2015**, *4*. [CrossRef]
- 39. Harguindey, S.; Orive, G.; Luis Pedraz, J.; Paradiso, A.; Reshkin, S.J. The role of pH dynamics and the Na+/H+ antiporter in the etiopathogenesis and treatment of cancer. Two faces of the same coin—One single nature. *Biochim. Biophys. Acta* **2005**, 1756, 1–24. [CrossRef]
- Amith, S.R.; Fliegel, L. Na(+)/H(+) exchanger-mediated hydrogen ion extrusion as a carcinogenic signal in triple-negative breast cancer etiopathogenesis and prospects for its inhibition in therapeutics. *Semin. Cancer Biol.* 2017, 43, 35–41. [CrossRef]
- 41. Amith, S.R.; Fong, S.; Baksh, S.; Fliegel, L. Na (+)/H (+)exchange in the tumour microenvironment: Does NHE1 drive breast cancer carcinogenesis? *Int. J. Dev. Biol.* **2015**, *59*, 367–377. [CrossRef]
- 42. Amith, S.R.; Fliegel, L. Regulation of the Na+/H+ Exchanger (NHE1) in Breast Cancer Metastasis. *Cancer Res.* **2013**, 73, 1259–1264. [CrossRef]
- Amith, S.R.; Wilkinson, J.M.; Fliegel, L. Assessing Na(+)/H(+) exchange and cell effector functionality in metastatic breast cancer. *Biochim. Open* 2016, 2, 16–23. [CrossRef] [PubMed]
- Amith, S.R.; Wilkinson, J.M.; Baksh, S.; Fliegel, L. The Na(+)/H(+) exchanger (NHE1) as a novel co-adjuvant target in paclitaxel therapy of triple-negative breast cancer cells. *Oncotarget* 2015, *6*, 1262–1275. [CrossRef] [PubMed]
- 45. Jang, M.; Kim, S.S.; Lee, J. Cancer cell metabolism: Implications for therapeutic targets. *Exp. Mol. Med.* **2013**, 45, e45. [CrossRef] [PubMed]
- 46. Parks, S.K.; Chiche, J.; Pouyssegur, J. Disrupting proton dynamics and energy metabolism for cancer therapy. *Nat. Rev. Cancer* **2013**, *13*, 611–623. [CrossRef] [PubMed]
- 47. Wu, H.; Estrella, V.; Enriquez-Navas, P.; El-Kenawi, A.; Russell, S.; Abrahams, D.; Ibrahim-Hashim, A.; Longo, D.; Reshetnyak, Y.; Luddy, K. Lymph Nodes Inhibit T-cell Effector Functions Locally by Establishing Acidic Niches. *bioRxiv* 2019. [CrossRef]
- 48. Cardone, R.A.; Casavola, V.; Reshkin, S.J. The role of disturbed pH dynamics and the Na+/H+ exchanger in metastasis. *Nat. Rev. Cancer* 2005, *5*, 786–795. [CrossRef] [PubMed]
- 49. Taylor, S.; Spugnini, E.P.; Assaraf, Y.G.; Azzarito, T.; Rauch, C.; Fais, S. Microenvironment acidity as a major determinant of tumor chemoresistance: Proton pump inhibitors (PPIs) as a novel therapeutic approach. *Drug Resist. Updat.* **2015**, *23*, 69–78. [CrossRef]
- 50. Corbet, C.; Feron, O. Tumour acidosis: From the passenger to the driver's seat. *Nat. Rev. Cancer* **2017**, *17*, 577–593. [CrossRef]

- 51. Robey, I.F.; Baggett, B.K.; Kirkpatrick, N.D.; Roe, D.J.; Dosescu, J.; Sloane, B.F.; Hashim, A.I.; Morse, D.L.; Raghunand, N.; Gatenby, R.A.; et al. Bicarbonate increases tumor pH and inhibits spontaneous metastases. *Cancer Res.* **2009**, *69*, 2260–2268. [CrossRef]
- 52. Melnik, S.; Dvornikov, D.; Muller-Decker, K.; Depner, S.; Stannek, P.; Meister, M.; Warth, A.; Thomas, M.; Muley, T.; Risch, A.; et al. Cancer cell specific inhibition of Wnt/beta-catenin signaling by forced intracellular acidification. *Cell Discov.* **2018**, *4*, 37. [CrossRef]
- 53. Lee, Z.W.; Teo, X.Y.; Song, Z.J.; Nin, D.S.; Novera, W.; Choo, B.A.; Dymock, B.W.; Moore, P.K.; Huang, R.Y.; Deng, L.W. Intracellular Hyper-Acidification Potentiated by Hydrogen Sulfide Mediates Invasive and Therapy Resistant Cancer Cell Death. *Front. Pharmacol.* **2017**, *8*, 763. [CrossRef] [PubMed]
- 54. Harguindey, S.; Arranz, J.L.; Wahl, M.L.; Orive, G.; Reshkin, S.J. Proton transport inhibitors as potentially selective anticancer drugs. *Anticancer Res.* **2009**, *29*, 2127–2136. [PubMed]
- 55. Reshkin, S.J.; Cardone, R.A.; Harguindey, S. Na+-H+ exchanger, pH regulation and cancer. *Recent Pat. Anticancer Drug Discov.* **2013**, *8*, 85–99. [CrossRef] [PubMed]
- Andersen, A.P.; Samsoe-Petersen, J.; Oernbo, E.K.; Boedtkjer, E.; Moreira, J.M.A.; Kveiborg, M.; Pedersen, S.F. The net acid extruders NHE1, NBCn1 and MCT4 promote mammary tumor growth through distinct but overlapping mechanisms. *Int. J. Cancer* 2018, 142, 2529–2542. [CrossRef]
- Boedtkjer, E.; Moreira, J.M.; Mele, M.; Vahl, P.; Wielenga, V.T.; Christiansen, P.M.; Jensen, V.E.; Pedersen, S.F.; Aalkjaer, C. Contribution of Na+,HCO3(-)-cotransport to cellular pH control in human breast cancer: A role for the breast cancer susceptibility locus NBCn1 (SLC4A7). *Int. J. Cancer* 2013, *132*, 1288–1299. [CrossRef]
- Lee, S.; Axelsen, T.V.; Andersen, A.P.; Vahl, P.; Pedersen, S.F.; Boedtkjer, E. Disrupting Na(+), HCO(3)(-)-cotransporter NBCn1 (Slc4a7) delays murine breast cancer development. *Oncogene* 2016, 35, 2112–2122. [CrossRef]
- 59. Baenke, F.; Dubuis, S.; Brault, C.; Weigelt, B.; Dankworth, B.; Griffiths, B.; Jiang, M.; Mackay, A.; Saunders, B.; Spencer-Dene, B.; et al. Functional screening identifies MCT4 as a key regulator of breast cancer cell metabolism and survival. *J. Pathol.* **2015**, *237*, 152–165. [CrossRef]
- 60. Loo, S.Y.; Chang, M.K.; Chua, C.S.; Kumar, A.P.; Pervaiz, S.; Clement, M.V. NHE-1: A promising target for novel anti-cancer therapeutics. *Curr. Pharm. Des.* **2012**, *18*, 1372–1382. [CrossRef]
- 61. Pinheiro, C.; Sousa, B.; Albergaria, A.; Paredes, J.; Dufloth, R.; Vieira, D.; Schmitt, F.; Baltazar, F. GLUT1 and CAIX expression profiles in breast cancer correlate with adverse prognostic factors and MCT1 overexpression. *Histol. Histopathol.* **2011**, *26*, 1279–1286. [CrossRef]
- Hsieh, M.J.; Chen, K.S.; Chiou, H.L.; Hsieh, Y.S. Carbonic anhydrase XII promotes invasion and migration ability of MDA-MB-231 breast cancer cells through the p38 MAPK signaling pathway. *Eur. J. Cell Biol.* 2010, 89, 598–606. [CrossRef]
- 63. Neri, D.; Supuran, C.T. Interfering with pH regulation in tumours as a therapeutic strategy. *Nat. Rev. Drug Discov.* **2011**, *10*, 767–777. [CrossRef]
- 64. Mboge, M.Y.; Mahon, B.P.; McKenna, R.; Frost, S.C. Carbonic Anhydrases: Role in pH Control and Cancer. *Metabolites* **2018**, *8*. [CrossRef] [PubMed]
- 65. Berrino, E.; Supuran, C.T. Novel approaches for designing drugs that interfere with pH regulation. *Expert Opin. Drug Discov.* **2019**, *14*, 231–248. [CrossRef] [PubMed]
- Lauritzen, G.; Stock, C.M.; Lemaire, J.; Lund, S.F.; Jensen, M.F.; Damsgaard, B.; Petersen, K.S.; Wiwel, M.; Ronnov-Jessen, L.; Schwab, A.; et al. The Na+/H+ exchanger NHE1, but not the Na+, HCO3(-) cotransporter NBCn1, regulates motility of MCF7 breast cancer cells expressing constitutively active ErbB2. *Cancer Lett.* 2012, 317, 172–183. [CrossRef]
- Chen, C.H.; Lee, C.Z.; Lin, Y.C.; Kao, L.T.; Lin, H.C. Negative Association of Proton Pump Inhibitors With Subsequent Development of Breast Cancer: A Nationwide Population-Based Study. *J. Clin. Pharmacol.* 2019, 59, 350–355. [CrossRef] [PubMed]
- 68. Mihaila, R.G. A minireview on NHE1 inhibitors. A rediscovered hope in oncohematology. *Biomed. Pap. Med. Fac. Univ. Palacky Olomouc Czech. Repub.* **2015**, *159*, 519–526. [CrossRef]
- 69. Goh, W.; Sleptsova-Freidrich, I.; Petrovic, N. Use of proton pump inhibitors as adjunct treatment for triple-negative breast cancers. An introductory study. *J. Pharm. Pharm. Sci.* **2014**, *17*, 439–446. [CrossRef]
- 70. Flinck, M.; Kramer, S.H.; Schnipper, J.; Andersen, A.P.; Pedersen, S.F. The acid-base transport proteins NHE1 and NBCn1 regulate cell cycle progression in human breast cancer cells. *Cell Cycle* **2018**, *17*, 1056–1067. [CrossRef]

- 71. White, K.A.; Grillo-Hill, B.K.; Barber, D.L. Cancer cell behaviors mediated by dysregulated pH dynamics at a glance. *J. Cell Sci.* 2017, 130, 663–669. [CrossRef]
- 72. Brisson, L.; Gillet, L.; Calaghan, S.; Besson, P.; Le Guennec, J.; Roger, S.; Gore, J. NaV1. 5 enhances breast cancer cell invasiveness by increasing NHE1-dependent H+ efflux in caveolae. *Oncogene* **2011**, *30*, 2070–2076. [CrossRef]
- 73. Wang, Y.; Li, S.J.; Wu, X.; Che, Y.; Li, Q. Clinicopathological and biological significance of human voltage-gated proton channel Hv1 protein overexpression in breast cancer. *J. Biol. Chem.* **2012**, *287*, 13877–13888. [CrossRef]
- 74. Brisson, L.; Driffort, V.; Benoist, L.; Poet, M.; Counillon, L.; Antelmi, E.; Rubino, R.; Besson, P.; Labbal, F.; Chevalier, S.; et al. NaV1.5 Na(+) channels allosterically regulate the NHE-1 exchanger and promote the activity of breast cancer cell invadopodia. *J. Cell Sci.* **2013**, *126*, 4835–4842. [CrossRef] [PubMed]
- 75. Schwab, A.; Stock, C. Ion channels and transporters in tumour cell migration and invasion. *Philos. Trans. R. Soc. B Biol. Sci.* **2014**, *369*, 20130102. [CrossRef] [PubMed]
- 76. Fraser, S.P.; Diss, J.K.; Chioni, A.M.; Mycielska, M.E.; Pan, H.; Yamaci, R.F.; Pani, F.; Siwy, Z.; Krasowska, M.; Grzywna, Z.; et al. Voltage-gated sodium channel expression and potentiation of human breast cancer metastasis. *Clin. Cancer Res.* 2005, 11, 5381–5389. [CrossRef] [PubMed]
- 77. Yang, M.; Kozminski, D.J.; Wold, L.A.; Modak, R.; Calhoun, J.D.; Isom, L.L.; Brackenbury, W.J. Therapeutic potential for phenytoin: Targeting Na(v)1.5 sodium channels to reduce migration and invasion in metastatic breast cancer. *Breast Cancer Res. Treat.* **2012**, *134*, 603–615. [CrossRef]
- Pilon-Thomas, S.; Kodumudi, K.N.; El-Kenawi, A.E.; Russell, S.; Weber, A.M.; Luddy, K.; Damaghi, M.; Wojtkowiak, J.W.; Mule, J.J.; Ibrahim-Hashim, A.; et al. Neutralization of Tumor Acidity Improves Antitumor Responses to Immunotherapy. *Cancer Res.* 2016, *76*, 1381–1390. [CrossRef]
- 79. Marches, R.; Vitetta, E.S.; Uhr, J.W. A role for intracellular pH in membrane IgM-mediated cell death of human B lymphomas. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 3434–3439. [CrossRef]
- 80. Harguindey, S.; Pedraz, J.L.; Canero, R.G.; Katin, M. Edelfosine, apoptosis, MDR and Na+/H+ exchanger: Induction mechanisms and treatment implications. *Apoptosis* **2000**, *5*, 87–89. [CrossRef]
- 81. Perez-Sala, D.; Collado-Escobar, D.; Mollinedo, F. Intracellular alkalinization suppresses lovastatin-induced apoptosis in HL-60 cells through the inactivation of a pH-dependent endonuclease. *J. Biol. Chem.* **1995**, 270, 6235–6242. [CrossRef]
- 82. Lagadic-Gossmann, D.; Huc, L.; Lecureur, V. Alterations of intracellular pH homeostasis in apoptosis: Origins and roles. *Cell Death Differ.* 2004, *11*, 953–961. [CrossRef]
- Sergeeva, T.F.; Shirmanova, M.V.; Zlobovskaya, O.A.; Gavrina, A.I.; Dudenkova, V.V.; Lukina, M.M.; Lukyanov, K.A.; Zagaynova, E.V. Relationship between intracellular pH, metabolic co-factors and caspase-3 activation in cancer cells during apoptosis. *Biochim. Biophys. Acta Mol. Cell Res.* 2017, 1864, 604–611. [CrossRef] [PubMed]
- Daniel, C.; Bell, C.; Burton, C.; Harguindey, S.; Reshkin, S.J.; Rauch, C. The role of proton dynamics in the development and maintenance of multidrug resistance in cancer. *Biochim. Biophys. Acta* 2013, 1832, 606–617. [CrossRef] [PubMed]
- 85. Walsh, M.; Fais, S.; Spugnini, E.P.; Harguindey, S.; Abu Izneid, T.; Scacco, L.; Williams, P.; Allegrucci, C.; Rauch, C.; Omran, Z. Proton pump inhibitors for the treatment of cancer in companion animals. *J. Exp. Clin. Cancer Res.* **2015**, *34*, 93. [CrossRef] [PubMed]
- 86. Rauch, C. On the relationship between drug's size, cell membrane mechanical properties and high levels of multi drug resistance: A comparison to published data. *Eur. Biophys. J.* **2009**, *38*, 537–546. [CrossRef]
- Weisburg, J.H.; Roepe, P.D.; Dzekunov, S.; Scheinberg, D.A. Intracellular pH and multidrug resistance regulate complement-mediated cytotoxicity of nucleated human cells. *J. Biol. Chem.* 1999, 274, 10877–10888. [CrossRef]
- 88. Roepe, P.D. pH and multidrug resistance. Novartis Found. Symp. 2001, 240, 232–247. [CrossRef]
- Rauch, C.; Blanchard, A.; Wood, E.; Dillon, E.; Wahl, M.L.; Harguindey, S. Cell Membranes, Cytosolic pH and Drug Transport in Cancer and MDR: Physics, Biochemistry and Molecular Biology. *Mult. Drug Resist.* 2009, 1, 24.
- Roepe, P.D.; Wei, L.Y.; Cruz, J.; Carlson, D. Lower electrical membrane potential and altered pHi homeostasis in multidrug-resistant (MDR) cells: Further characterization of a series of MDR cell lines expressing different levels of P-glycoprotein. *Biochemistry* 1993, 32, 11042–11056. [CrossRef]

- 91. Ho, C.C.; Ding, S.J. The pH-controlled nanoparticles size of polydopamine for anti-cancer drug delivery. *J. Mater. Sci. Mater. Med.* **2013**, *24*, 2381–2390. [CrossRef]
- 92. Karanth, H.; Murthy, R.S. pH-sensitive liposomes–principle and application in cancer therapy. *J. Pharm. Pharmacol.* **2007**, *59*, 469–483. [CrossRef]
- Bariwal, J.; Kumar, V.; Chen, H.; Bhattarai, R.S.; Peng, Y.; Li, W.; Mahato, R.I. Nanoparticulate delivery of potent microtubule inhibitor for metastatic melanoma treatment. *J. Control. Release* 2019, 309, 231–243. [CrossRef] [PubMed]
- 94. Nemeth-Cahalan, K.L.; Kalman, K.; Hall, J.E. Molecular basis of pH and Ca2+ regulation of aquaporin water permeability. *J. Gen. Physiol.* **2004**, *123*, 573–580. [CrossRef] [PubMed]
- Mosca, A.F.; de Almeida, A.; Wragg, D.; Martins, A.P.; Sabir, F.; Leoni, S.; Moura, T.F.; Prista, C.; Casini, A.; Soveral, G. Molecular Basis of Aquaporin-7 Permeability Regulation by pH. *Cells* 2018, 7. [CrossRef] [PubMed]
- Marino, M.L.; Pellegrini, P.; Di Lernia, G.; Djavaheri-Mergny, M.; Brnjic, S.; Zhang, X.; Hagg, M.; Linder, S.; Fais, S.; Codogno, P.; et al. Autophagy is a protective mechanism for human melanoma cells under acidic stress. J. Biol. Chem. 2012, 287, 30664–30676. [CrossRef] [PubMed]
- Wojtkowiak, J.W.; Rothberg, J.M.; Kumar, V.; Schramm, K.J.; Haller, E.; Proemsey, J.B.; Lloyd, M.C.; Sloane, B.F.; Gillies, R.J. Chronic autophagy is a cellular adaptation to tumor acidic pH microenvironments. *Cancer Res.* 2012, 72, 3938–3947. [CrossRef] [PubMed]
- 98. Pellegrini, P.; Dyczynski, M.; Sbrana, F.V.; Karlgren, M.; Buoncervello, M.; Hagg-Olofsson, M.; Ma, R.; Hartman, J.; Bajalica-Lagercrantz, S.; Grander, D.; et al. Tumor acidosis enhances cytotoxic effects and autophagy inhibition by salinomycin on cancer cell lines and cancer stem cells. *Oncotarget* 2016. [CrossRef] [PubMed]
- 99. Pellegrini, P.; Strambi, A.; Zipoli, C.; Hagg-Olofsson, M.; Buoncervello, M.; Linder, S.; De Milito, A. Acidic extracellular pH neutralizes the autophagy-inhibiting activity of chloroquine: Implications for cancer therapies. *Autophagy* **2014**, *10*, 562–571. [CrossRef]
- Matsuo, T.; Sadzuka, Y. Extracellular acidification by lactic acid suppresses glucose deprivation-induced cell death and autophagy in B16 melanoma cells. *Biochem. Biophys. Res. Commun.* 2018, 496, 1357–1361. [CrossRef]
- 101. Gieling, R.G.; Parker, C.A.; De Costa, L.A.; Robertson, N.; Harris, A.L.; Stratford, I.J.; Williams, K.J. Inhibition of carbonic anhydrase activity modifies the toxicity of doxorubicin and melphalan in tumour cells in vitro. *J. Enzyme Inhib. Med. Chem.* 2013, 28, 360–369. [CrossRef]
- 102. Morais-Santos, F.; Granja, S.; Miranda-Goncalves, V.; Moreira, A.H.; Queiros, S.; Vilaca, J.L.; Schmitt, F.C.; Longatto-Filho, A.; Paredes, J.; Baltazar, F.; et al. Targeting lactate transport suppresses in vivo breast tumour growth. *Oncotarget* 2015, *6*, 19177–19189. [CrossRef]
- 103. Bai, F.; Yu, Z.; Gao, X.; Gong, J.; Fan, L.; Liu, F. Simvastatin induces breast cancer cell death through oxidative stress up-regulating miR-140-5p. *Aging* **2019**, *11*, 3198–3219. [CrossRef]
- 104. Iessi, E.; Logozzi, M.; Lugini, L.; Azzarito, T.; Federici, C.; Spugnini, E.P.; Mizzoni, D.; Di Raimo, R.; Angelini, D.F.; Battistini, L.; et al. Acridine Orange/exosomes increase the delivery and the effectiveness of Acridine Orange in human melanoma cells: A new prototype for theranostics of tumors. *J. Enzyme Inhib. Med. Chem.* 2017, *32*, 648–657. [CrossRef] [PubMed]
- 105. Gdovin, M.J.; Kadri, N.; Rios, L.; Holliday, S.; Jordan, Z. Focal photodynamic intracellular acidification as a cancer therapeutic. *Semin. Cancer Biol.* **2017**, *43*, 147–156. [CrossRef] [PubMed]
- 106. Cole, W.H.; Everson, T.C. Spontaneous regression of cancer: Preliminary report. *Ann. Surg.* **1956**, 144, 366–383. [CrossRef] [PubMed]
- Mahoney, E.M. Complete regression of vesical carcinoma following urinary diversion. *Am. J. Surg.* 1960, 100, 133–136. [CrossRef]
- 108. Papac, R.J. Spontaneous regression of cancer: Possible mechanisms. In Vivo 1998, 12, 571–578. [PubMed]
- Goldberg, L.G. Complete regression of carcinoma of the bladder following ureterosigmoidostomy. *J. Urol.* 1950, 63, 116–123. [CrossRef]
- 110. Harguindey, S.; Cragoe, E.J., Jr. The Na+/H+ antiporter in oncology in the light of the spontaneous regression of cancer and cell metabolism. *Med. Hypotheses* **1992**, *39*, 229–237. [CrossRef]
- 111. Oleksyszyn, J. Spontaneous Regression of Cancer, an Ever Actual Inspiration for the New Cancer Treatment. *Cell Biol.* **2016**, *5*. [CrossRef]

- 112. Harguindey, S.S.; Kolbeck, R.C.; Bransome, E.D., Jr. Letter: Ureterosigmoidostomy and cancer: New observations. *Ann. Intern. Med.* **1975**, *83*, 833. [CrossRef]
- Gillies, R.J.; Gatenby, R.A. Metabolism and its sequelae in cancer evolution and therapy. *Cancer J.* 2015, 21, 88–96. [CrossRef] [PubMed]
- 114. Harguindey, S.; Gillis, M. Evolution and cancer: Possible relationships to changes in environmental hydrogen ion concentration. *J. Theor. Biol.* **1980**, *86*, 487–492. [CrossRef]
- Alfarouk, K.O.; Shayoub, M.E.; Muddathir, A.K.; Elhassan, G.O.; Bashir, A.H. Evolution of Tumor Metabolism might Reflect Carcinogenesis as a Reverse Evolution process (Dismantling of Multicellularity). *Cancers (Basel)* 2011, 3, 3002–3017. [CrossRef] [PubMed]
- 116. Alfarouk, K.O.; Muddathir, A.K.; Shayoub, M.E. Tumor acidity as evolutionary spite. *Cancers (Basel)* **2011**, *3*, 408–414. [CrossRef]
- 117. Gatenby, R.A.; Vincent, T.L. An evolutionary model of carcinogenesis. Cancer Res. 2003, 63, 6212–6220.
- 118. Harguindey, S.; Pedraz, J.L.; Garcia Canero, R.; Perez de Diego, J.; Cragoe, E.J., Jr. Hydrogen ion-dependent oncogenesis and parallel new avenues to cancer prevention and treatment using a H(+)-mediated unifying approach: pH-related and pH-unrelated mechanisms. *Crit. Rev. Oncog.* **1995**, *6*, 1–33. [CrossRef]
- 119. Saponaro, C.; Vagheggini, A.; Scarpi, E.; Centonze, M.; Catacchio, I.; Popescu, O.; Pastena, M.I.; Giotta, F.; Silvestris, N.; Mangia, A. NHERF1 and tumor microenvironment: A new scene in invasive breast carcinoma. *J. Exp. Clin. Cancer Res.* 2018, *37*, 96. [CrossRef]
- 120. Di Pompo, G.; Lemma, S.; Canti, L.; Rucci, N.; Ponzetti, M.; Errani, C.; Donati, D.M.; Russell, S.; Gillies, R.; Chano, T.; et al. Intratumoral acidosis fosters cancer-induced bone pain through the activation of the mesenchymal tumor-associated stroma in bone metastasis from breast carcinoma. *Oncotarget* 2017, *8*, 54478–54496. [CrossRef]
- 121. Gonzalez, J.F.; Madamba, A.; Antón, L.A.; Jurado, M.C.; Harguindey, S. Cervical mosaic and an integrated pathophysiological approach to early cervical neoplasia. *J. Biol. Response Mod.* **1987**, *6*, 473–483.
- 122. Lobo, R.C.; Hubbard, N.E.; Damonte, P.; Mori, H.; Penzvalto, Z.; Pham, C.; Koehne, A.L.; Go, A.C.; Anderson, S.E.; Cala, P.M.; et al. Glucose Uptake and Intracellular pH in a Mouse Model of Ductal Carcinoma In situ (DCIS) Suggests Metabolic Heterogeneity. *Front. Cell Dev. Biol.* **2016**, *4*, 93. [CrossRef]
- 123. Cong, D.; Zhu, W.; Shi, Y.; Pointer, K.B.; Clark, P.A.; Shen, H.; Kuo, J.S.; Hu, S.; Sun, D. Upregulation of NHE1 protein expression enables glioblastoma cells to escape TMZ-mediated toxicity via increased H(+) extrusion, cell migration and survival. *Carcinogenesis* **2014**, *35*, 2014–2024. [CrossRef] [PubMed]
- 124. Shimizu, S.; Eguchi, Y.; Kamiike, W.; Funahashi, Y.; Mignon, A.; Lacronique, V.; Matsuda, H.; Tsujimoto, Y. Bcl-2 prevents apoptotic mitochondrial dysfunction by regulating proton flux. *Proc. Natl. Acad. Sci. USA* 1998, 95, 1455–1459. [CrossRef] [PubMed]
- 125. Murakami, T.; Shibuya, I.; Ise, T.; Chen, Z.S.; Akiyama, S.; Nakagawa, M.; Izumi, H.; Nakamura, T.; Matsuo, K.; Yamada, Y.; et al. Elevated expression of vacuolar proton pump genes and cellular PH in cisplatin resistance. *Int. J. Cancer* 2001, *93*, 869–874. [CrossRef] [PubMed]
- 126. Dumas, J.F.; Brisson, L.; Chevalier, S.; Maheo, K.; Fromont, G.; Moussata, D.; Besson, P.; Roger, S. Metabolic reprogramming in cancer cells, consequences on pH and tumour progression: Integrated therapeutic perspectives with dietary lipids as adjuvant to anticancer treatment. *Semin. Cancer Biol.* 2017, 43, 90–110. [CrossRef] [PubMed]
- 127. Roma-Rodrigues, C.; Mendes, R.; Baptista, P.V.; Fernandes, A.R. Targeting Tumor Microenvironment for Cancer Therapy. *Int. J. Mol. Sci.* 2019, 20. [CrossRef] [PubMed]
- 128. Fais, S.; Venturi, G.; Gatenby, B. Microenvironmental acidosis in carcinogenesis and metastases: New strategies in prevention and therapy. *Cancer Metastasis Rev.* **2014**, *33*, 1095–1108. [CrossRef]
- 129. Calcinotto, A.; Filipazzi, P.; Grioni, M.; Iero, M.; De Milito, A.; Ricupito, A.; Cova, A.; Canese, R.; Jachetti, E.; Rossetti, M.; et al. Modulation of microenvironment acidity reverses anergy in human and murine tumor-infiltrating T lymphocytes. *Cancer Res.* **2012**, *72*, 2746–2756. [CrossRef]
- 130. Thews, O.; Gassner, B.; Kelleher, D.K.; Schwerdt, G.; Gekle, M. Impact of extracellular acidity on the activity of P-glycoprotein and the cytotoxicity of chemotherapeutic drugs. *Neoplasia* **2006**, *8*, 143–152. [CrossRef]
- 131. Thiebaut, F.; Currier, S.J.; Whitaker, J.; Haugland, R.P.; Gottesman, M.M.; Pastan, I.; Willingham, M.C. Activity of the multidrug transporter results in alkalinization of the cytosol: Measurement of cytosolic pH by microinjection of a pH-sensitive dye. *J. Histochem. Cytochem.* **1990**, *38*, 685–690. [CrossRef]

- 132. Fais, S. Proton pump inhibitor-induced tumour cell death by inhibition of a detoxification mechanism. *J. Intern. Med.* **2010**, 267, 515–525. [CrossRef]
- 133. Alfarouk, K.O. Tumor metabolism, cancer cell transporters, and microenvironmental resistance. *J. Enzyme Inhib. Med. Chem.* **2016**, *31*, 859–866. [CrossRef] [PubMed]
- 134. Thews, O.; Nowak, M.; Sauvant, C.; Gekle, M. Hypoxia-induced extracellular acidosis increases p-glycoprotein activity and chemoresistance in tumors in vivo via p38 signaling pathway. In *Oxygen Transport to Tissue XXXII*; Springer: Boston, MA, USA, 2011; pp. 115–122. [CrossRef]
- 135. Damaghi, M.; Gillies, R. Phenotypic changes of acid-adapted cancer cells push them toward aggressiveness in their evolution in the tumor microenvironment. *Cell Cycle* **2017**, *16*, 1739–1743. [CrossRef] [PubMed]
- 136. Gupta, S.C.; Singh, R.; Pochampally, R.; Watabe, K.; Mo, Y.Y. Acidosis promotes invasiveness of breast cancer cells through ROS-AKT-NF-kappaB pathway. *Oncotarget* **2014**, *5*, 12070–12082. [CrossRef] [PubMed]
- 137. Montcourrier, P.; Silver, I.; Farnoud, R.; Bird, I.; Rochefort, H. Breast cancer cells have a high capacity to acidify extracellular milieu by a dual mechanism. *Clin. Exp. Metastasis* **1997**, *15*, 382–392. [CrossRef] [PubMed]
- Rothberg, J.M.; Bailey, K.M.; Wojtkowiak, J.W.; Ben-Nun, Y.; Bogyo, M.; Weber, E.; Moin, K.; Blum, G.; Mattingly, R.R.; Gillies, R.J.; et al. Acid-mediated tumor proteolysis: Contribution of cysteine cathepsins. *Neoplasia* 2013, 15, 1125–1137. [CrossRef]
- 139. Wani, B.; Aziz, S.A.; Ganaie, M.A.; Mir, M.H. Metabolic Syndrome and Breast Cancer Risk. *Indian J. Med. Paediatr. Oncol.* 2017, 38, 434–439. [CrossRef]
- Tsujimoto, T.; Kajio, H.; Sugiyama, T. Association between hyperinsulinemia and increased risk of cancer death in nonobese and obese people: A population-based observational study. *Int. J. Cancer* 2017, 141, 102–111. [CrossRef]
- 141. Evans, J.L.; Maddux, B.A.; Goldfine, I.D. The molecular basis for oxidative stress-induced insulin resistance. *Antioxid. Redox Signal.* **2005**, *7*, 1040–1052. [CrossRef]
- 142. Ramirez, M.A.; Beltran, A.R.; Araya, J.E.; Cornejo, M.; Toledo, F.; Fuentes, G.; Sobrevia, L. Involvement of Intracellular pH in Vascular Insulin Resistance. *Curr. Vasc. Pharmacol.* **2019**, *17*, 440–446. [CrossRef]
- 143. Boyd, D.B. Insulin and cancer. Integr. Cancer Ther. 2003, 2, 315–329. [CrossRef]
- 144. Moore, R.D.; Gupta, R.K. Effect of insulin on intracellular pH as observed by 31P NMR spectroscopy. *Int. J. Quantum Chem.* **1980**, *18*, 83–92. [CrossRef]
- 145. Kaloyianni, M.; Bourikas, D.; Koliakos, G. The effect of insulin on Na+-H+ antiport activity of obese and normal subjects erythrocytes. *Cell. Physiol. Biochem.* **2001**, *11*, 253–258. [CrossRef] [PubMed]
- 146. Arcidiacono, B.; Iiritano, S.; Nocera, A.; Possidente, K.; Nevolo, M.T.; Ventura, V.; Foti, D.; Chiefari, E.; Brunetti, A. Insulin resistance and cancer risk: An overview of the pathogenetic mechanisms. *Exp. Diabetes Res.* 2012, 2012, 789174. [CrossRef] [PubMed]
- 147. Ruiz-Narvaez, E.A.; Lunetta, K.L.; Hong, C.C.; Haddad, S.; Yao, S.; Cheng, T.D.; Bensen, J.T.; Bandera, E.V.; Haiman, C.A.; Troester, M.A.; et al. Genetic variation in the insulin, insulin-like growth factor, growth hormone, and leptin pathways in relation to breast cancer in African-American women: The AMBER consortium. *NPJ Breast Cancer* **2016**, 2. [CrossRef] [PubMed]
- 148. Wu, Y.; Gao, B.; Xiong, Q.J.; Wang, Y.C.; Huang, D.K.; Wu, W.N. Acid-sensing ion channels contribute to the effect of extracellular acidosis on proliferation and migration of A549 cells. *Tumor Biol.* 2017, 39. [CrossRef] [PubMed]
- 149. Bellizzi, A.; Greco, M.R.; Rubino, R.; Paradiso, A.; Forciniti, S.; Zeeberg, K.; Cardone, R.A.; Reshkin, S.J. The scaffolding protein NHERF1 sensitizes EGFR-dependent tumor growth, motility and invadopodia function to gefitinib treatment in breast cancer cells. *Int. J. Oncol.* **2015**, *46*, 1214–1224. [CrossRef]
- Pedraz-Cuesta, E.; Fredsted, J.; Jensen, H.H.; Bornebusch, A.; Nejsum, L.N.; Kragelund, B.B.; Pedersen, S.F. Prolactin Signaling Stimulates Invasion via Na(+)/H(+) Exchanger NHE1 in T47D Human Breast Cancer Cells. *Mol. Endocrinol.* 2016, 30, 693–708. [CrossRef]
- 151. Clevenger, C.V.; Furth, P.A.; Hankinson, S.E.; Schuler, L.A. The role of prolactin in mammary carcinoma. *Endocr. Rev.* **2003**, *24*, 1–27. [CrossRef]
- 152. Xu, J.; Sun, D.; Jiang, J.; Deng, L.; Zhang, Y.; Yu, H.; Bahl, D.; Langenheim, J.F.; Chen, W.Y.; Fuchs, S.Y.; et al. The role of prolactin receptor in GH signaling in breast cancer cells. *Mol. Endocrinol.* 2013, 27, 266–279. [CrossRef]
- 153. Seo, E.J.; Sugimoto, Y.; Greten, H.J.; Efferth, T. Repurposing of Bromocriptine for Cancer Therapy. *Front. Pharmacol.* **2018**, *9*, 1030. [CrossRef]

- 154. Yager, J.D.; Davidson, N.E. Estrogen carcinogenesis in breast cancer. *N. Engl. J. Med.* **2006**, 354, 270–282. [CrossRef] [PubMed]
- 155. Lloyd, M.C.; Alfarouk, K.O.; Verduzco, D.; Bui, M.M.; Gillies, R.J.; Ibrahim, M.E.; Brown, J.S.; Gatenby, R.A. Vascular measurements correlate with estrogen receptor status. *BMC Cancer* 2014, 14, 279. [CrossRef] [PubMed]
- 156. Gruvberger, S.; Ringner, M.; Chen, Y.; Panavally, S.; Saal, L.H.; Borg, A.; Ferno, M.; Peterson, C.; Meltzer, P.S. Estrogen receptor status in breast cancer is associated with remarkably distinct gene expression patterns. *Cancer Res.* 2001, *61*, 5979–5984. [PubMed]
- 157. Mouridsen, H.T. Letrozole in advanced breast cancer: The PO25 trial. *Breast Cancer Res. Treat.* 2007, 105, 19–29. [CrossRef]
- 158. Ibrahim-Hashim, A.; Wojtkowiak, J.W.; de Lourdes Coelho Ribeiro, M.; Estrella, V.; Bailey, K.M.; Cornnell, H.H.; Gatenby, R.A.; Gillies, R.J. *Free Base Lysine Increases Survival and Reduces Metastasis in Prostate Cancer Model*; OMICS Publishing Group: Hyderabad, India, 2011.
- 159. Harguindey, S.; DeCastro, L.; Barcos, M.; Getaz, E.P.; Henderson, E.S.; Freeman, A. Hypercalcemia complicating childhood malignancies: A report of seven cases with some pathophysiological considerations. *Cancer* **1979**, *44*, 2280–2290. [CrossRef]
- 160. Della Guardia, L.; Thomas, M.A.; Cena, H. Insulin Sensitivity and Glucose Homeostasis Can Be Influenced by Metabolic Acid Load. *Nutrients* **2018**, *10*. [CrossRef]
- 161. Harguindey, S. Homeostasis, diabetes and cancer. An. Med. Interna 1993, 10, 611-616.
- 162. Munoz, J.; Mohd, K.; Amr, H.; Stoltenberg, M. Severe lactic acidosis in a patient with metastatic prostate cancer. *J. Cancer Res. Ther.* **2011**, *7*, 201–202. [CrossRef]
- 163. Van der Mijn, J.C.; Kuiper, M.J.; Siegert, C.E.H.; Wassenaar, A.E.; van Noesel, C.J.M.; Ogilvie, A.C. Lactic Acidosis in Prostate Cancer: Consider the Warburg Effect. *Case Rep. Oncol.* **2017**, *10*, 1085–1091. [CrossRef]
- 164. Tuccori, M.; Wu, J.W.; Yin, H.; Majdan, A.; Azoulay, L. The use of glyburide compared with other sulfonylureas and the risk of cancer in patients with type 2 diabetes. *Diabetes Care* **2015**, *38*, 2083–2089. [CrossRef]
- 165. Chen, Q.; Liu, Y.; Zhu, X.L.; Feng, F.; Yang, H.; Xu, W. Increased NHE1 expression is targeted by specific inhibitor cariporide to sensitize resistant breast cancer cells to doxorubicin in vitro and in vivo. *BMC Cancer* 2019, 19, 211. [CrossRef] [PubMed]
- Pasello, G.; Urso, L.; Conte, P.; Favaretto, A. Effects of sulfonylureas on tumor growth: A review of the literature. *Oncologist* 2013, *18*, 1118–1125. [CrossRef] [PubMed]
- 167. Gao, R.; Yang, T.; Xu, W. Enemies or weapons in hands: Investigational anti-diabetic drug glibenclamide and cancer risk. *Expert Opin. Investig. Drugs* **2017**, *26*, 853–864. [CrossRef] [PubMed]
- 168. Tang, T.; Lord, J.M.; Norman, R.J.; Yasmin, E.; Balen, A.H. Insulin-sensitising drugs (metformin, rosiglitazone, pioglitazone, D-chiro-inositol) for women with polycystic ovary syndrome, oligo amenorrhoea and subfertility. *Cochrane Database Syst. Rev.* 2012. [CrossRef] [PubMed]
- Belli, S.H.; Graffigna, M.N.; Oneto, A.; Otero, P.; Schurman, L.; Levalle, O.A. Effect of rosiglitazone on insulin resistance, growth factors, and reproductive disturbances in women with polycystic ovary syndrome. *Fertil. Steril.* 2004, *81*, 624–629. [CrossRef]
- 170. Harguindey, S.; Arranz, J.L.; Polo Orozco, J.D.; Rauch, C.; Fais, S.; Cardone, R.A.; Reshkin, S.J. Cariporide and other new and powerful NHE1 inhibitors as potentially selective anticancer drugs–an integral molecular/biochemical/metabolic/clinical approach after one hundred years of cancer research. *J. Transl. Med.* 2013, 11, 282. [CrossRef]
- Harguindey, S.; Orive, G.; Pedraz, J.L.; Bello, G.; Arranz, J.L.; Samaniego, J.M. Apparent cure of a case of metastatic ovarian carcinoma after the chronic treatment with Na+H+ antiport inhibitors. *Oncologia* 2002, 25, 62–66.
- 172. He, B.; Zhang, M.; Zhu, R. Na+/H+ exchanger blockade inhibits the expression of vascular endothelial growth factor in SGC7901 cells. *Oncol. Rep.* **2010**, *23*, 79–87. [CrossRef]
- Evans, D.M.; Sloan-Stakleff, K.; Arvan, M.; Guyton, D.P. Time and dose dependency of the suppression of pulmonary metastases of rat mammary cancer by amiloride. *Clin. Exp. Metastasis* 1998, 16, 353–357. [CrossRef]

- 174. Di Sario, A.; Bendia, E.; Omenetti, A.; De Minicis, S.; Marzioni, M.; Kleemann, H.W.; Candelaresi, C.; Saccomanno, S.; Alpini, G.; Benedetti, A. Selective inhibition of ion transport mechanisms regulating intracellular pH reduces proliferation and induces apoptosis in cholangiocarcinoma cells. *Dig. Liver Dis.* 2007, 39, 60–69. [CrossRef]
- 175. Kellen, J.A.; Mirakian, A.; Kolin, A. Antimetastatic effect of amiloride in an animal tumour model. *Anticancer Res.* **1988**, *8*, 1373–1376.
- 176. Porporato, P.E.; Dhup, S.; Dadhich, R.K.; Copetti, T.; Sonveaux, P. Anticancer targets in the glycolytic metabolism of tumors: A comprehensive review. *Front. Pharmacol.* **2011**, *2*, 49. [CrossRef] [PubMed]
- 177. Luciani, F.; Spada, M.; De Milito, A.; Molinari, A.; Rivoltini, L.; Montinaro, A.; Marra, M.; Lugini, L.; Logozzi, M.; Lozupone, F.; et al. Effect of proton pump inhibitor pretreatment on resistance of solid tumors to cytotoxic drugs. *J. Natl. Cancer Inst.* **2004**, *96*, 1702–1713. [CrossRef] [PubMed]
- 178. Atwal, K.S.; O'Neil, S.V.; Ahmad, S.; Doweyko, L.; Kirby, M.; Dorso, C.R.; Chandrasena, G.; Chen, B.C.; Zhao, R.; Zahler, R. Synthesis and biological activity of 5-aryl-4-(4-(5-methyl-1H-imidazol-4-yl)piperidin-1-yl)pyrimidine analogs as potent, highly selective, and orally bioavailable NHE-1 inhibitors. *Bioorg. Med. Chem. Lett.* 2006, 16, 4796–4799. [CrossRef] [PubMed]
- Tavares-Valente, D.; Baltazar, F.; Moreira, R.; Queiros, O. Cancer cell bioenergetics and pH regulation influence breast cancer cell resistance to paclitaxel and doxorubicin. *J. Bioenerg. Biomembr.* 2013, 45, 467–475. [CrossRef]
- 180. Reshkin, S.J.; Bellizzi, A.; Cardone, R.A.; Tommasino, M.; Casavola, V.; Paradiso, A. Paclitaxel induces apoptosis via protein kinase A- and p38 mitogen-activated protein-dependent inhibition of the Na+/H+ exchanger (NHE) NHE isoform 1 in human breast cancer cells. *Clin. Cancer Res.* 2003, *9*, 2366–2373.
- 181. Granja, S.; Tavares-Valente, D.; Queiros, O.; Baltazar, F. Value of pH regulators in the diagnosis, prognosis and treatment of cancer. *Semin. Cancer Biol.* **2017**, *43*, 17–34. [CrossRef]
- 182. Xiao-Fang, C.; Zheng, C.-L.; Akiyama, S.-I.; Tomoda, A. 2-Aminophenoxazine-3-one and 2-amino-4, 4α-dihydro-4α, 7-dimethyl-3H-phenoxazine-3-one cause cellular apoptosis by reducing higher intracellular pH in cancer cells. *Proc. Jpn. Acad. Ser. B Phys. Biol. Sci.* 2011, *87*, 199. [CrossRef]
- 183. Supuran, C.T. Carbonic Anhydrases and Metabolism. Metabolites 2018, 8. [CrossRef]
- 184. Spugnini, E.P.; Buglioni, S.; Carocci, F.; Francesco, M.; Vincenzi, B.; Fanciulli, M.; Fais, S. High dose lansoprazole combined with metronomic chemotherapy: A phase I/II study in companion animals with spontaneously occurring tumors. *J. Transl. Med.* **2014**, *12*, 225. [CrossRef]
- 185. Spugnini, E.; Fais, S. Proton pump inhibition and cancer therapeutics: A specific tumor targeting or it is a phenomenon secondary to a systemic buffering? *Semin. Cancer Biol.* **2017**, *43*, 111–118. [CrossRef] [PubMed]
- 186. Lu, Z.N.; Tian, B.; Guo, X.L. Repositioning of proton pump inhibitors in cancer therapy. *Cancer Chemother. Pharmacol.* **2017**, *80*, 925–937. [CrossRef]
- 187. Wang, B.Y.; Zhang, J.; Wang, J.L.; Sun, S.; Wang, Z.H.; Wang, L.P.; Zhang, Q.L.; Lv, F.F.; Cao, E.Y.; Shao, Z.M.; et al. Intermittent high dose proton pump inhibitor enhances the antitumor effects of chemotherapy in metastatic breast cancer. *J. Exp. Clin. Cancer Res.* **2015**, *34*, 85. [CrossRef] [PubMed]
- 188. Azzarito, T.; Venturi, G.; Cesolini, A.; Fais, S. Lansoprazole induces sensitivity to suboptimal doses of paclitaxel in human melanoma. *Cancer Lett.* **2015**, *356*, 697–703. [CrossRef] [PubMed]
- Niikura, K. Effect of a V-ATPase inhibitor, FR202126, in syngeneic mouse model of experimental bone metastasis. *Cancer Chemother. Pharmacol.* 2007, 60, 555–562. [CrossRef] [PubMed]
- 190. De Milito, A.; Iessi, E.; Logozzi, M.; Lozupone, F.; Spada, M.; Marino, M.L.; Federici, C.; Perdicchio, M.; Matarrese, P.; Lugini, L.; et al. Proton pump inhibitors induce apoptosis of human B-cell tumors through a caspase-independent mechanism involving reactive oxygen species. *Cancer Res.* 2007, 67, 5408–5417. [CrossRef] [PubMed]
- 191. De Milito, A.; Canese, R.; Marino, M.L.; Borghi, M.; Iero, M.; Villa, A.; Venturi, G.; Lozupone, F.; Iessi, E.; Logozzi, M.; et al. pH-dependent antitumor activity of proton pump inhibitors against human melanoma is mediated by inhibition of tumor acidity. *Int. J. Cancer* **2010**, *127*, 207–219. [CrossRef]
- 192. Zhang, S.; Wang, Y.; Li, S.J. Lansoprazole induces apoptosis of breast cancer cells through inhibition of intracellular proton extrusion. *Biochem. Biophys. Res. Commun.* **2014**, 448, 424–429. [CrossRef]
- 193. Spugnini, E.P.; Baldi, A.; Buglioni, S.; Carocci, F.; de Bazzichini, G.M.; Betti, G.; Pantaleo, I.; Menicagli, F.; Citro, G.; Fais, S. Lansoprazole as a rescue agent in chemoresistant tumors: A phase I/II study in companion animals with spontaneously occurring tumors. *J. Transl. Med.* **2011**, *9*, 221. [CrossRef]

- 194. Ding, D.C.; Sung, F.C.; Chen, W.; Wang, J.H.; Lin, S.Z. Proton pump inhibitors reduce breast cancer risk in gastric ulcer patients: A population-based cohort study. *Breast J.* **2019**. [CrossRef]
- 195. Lee, Y.Y.; Jeon, H.K.; Hong, J.E.; Cho, Y.J.; Ryu, J.Y.; Choi, J.J.; Lee, S.H.; Yoon, G.; Kim, W.Y.; Do, I.G.; et al. Proton pump inhibitors enhance the effects of cytotoxic agents in chemoresistant epithelial ovarian carcinoma. *Oncotarget* 2015, *6*, 35040–35050. [CrossRef] [PubMed]
- 196. Ferrari, S.; Perut, F.; Fagioli, F.; Brach Del Prever, A.; Meazza, C.; Parafioriti, A.; Picci, P.; Gambarotti, M.; Avnet, S.; Baldini, N.; et al. Proton pump inhibitor chemosensitization in human osteosarcoma: From the bench to the patients' bed. *J. Transl. Med.* 2013, *11*, 268. [CrossRef] [PubMed]
- Payen, V.L.; Mina, E.; Van Hee, V.F.; Porporato, P.E.; Sonveaux, P. Monocarboxylate transporters in cancer. *Mol. Metab.* 2019. [CrossRef] [PubMed]
- 198. Long, Y.; Gao, Z.; Hu, X.; Xiang, F.; Wu, Z.; Zhang, J.; Han, X.; Yin, L.; Qin, J.; Lan, L. Downregulation of MCT 4 for lactate exchange promotes the cytotoxicity of NK cells in breast carcinoma. *Cancer Med.* 2018, 7, 4690–4700. [CrossRef]
- 199. Nath, K.; Guo, L.; Nancolas, B.; Nelson, D.S.; Shestov, A.A.; Lee, S.C.; Roman, J.; Zhou, R.; Leeper, D.B.; Halestrap, A.P.; et al. Mechanism of antineoplastic activity of lonidamine. *Biochim. Biophys. Acta* 2016, 1866, 151–162. [CrossRef]
- 200. Mboge, M.Y.; Chen, Z.; Wolff, A.; Mathias, J.V.; Tu, C.; Brown, K.D.; Bozdag, M.; Carta, F.; Supuran, C.T.; McKenna, R.; et al. Selective inhibition of carbonic anhydrase IX over carbonic anhydrase XII in breast cancer cells using benzene sulfonamides: Disconnect between activity and growth inhibition. *PLoS ONE* 2018, 13, e0207417. [CrossRef]
- 201. Bartosova, M.; Parkkila, S.; Pohlodek, K.; Karttunen, T.J.; Galbavy, S.; Mucha, V.; Harris, A.L.; Pastorek, J.; Pastorekova, S. Expression of carbonic anhydrase IX in breast is associated with malignant tissues and is related to overexpression of c-erbB2. *J. Pathol.* 2002, *197*, 314–321. [CrossRef]
- Lock, F.E.; McDonald, P.C.; Lou, Y.; Serrano, I.; Chafe, S.C.; Ostlund, C.; Aparicio, S.; Winum, J.Y.; Supuran, C.T.; Dedhar, S. Targeting carbonic anhydrase IX depletes breast cancer stem cells within the hypoxic niche. *Oncogene* 2013, *32*, 5210–5219. [CrossRef]
- 203. Meehan, J.; Ward, C.; Turnbull, A.; Bukowski-Wills, J.; Finch, A.J.; Jarman, E.J.; Xintaropoulou, C.; Martinez-Perez, C.; Gray, M.; Pearson, M.; et al. Inhibition of pH regulation as a therapeutic strategy in hypoxic human breast cancer cells. *Oncotarget* **2017**, *8*, 42857–42875. [CrossRef]
- 204. Keizer, H.G.; Joenje, H. Increased cytosolic pH in multidrug-resistant human lung tumor cells: Effect of verapamil. *J. Natl. Cancer Inst.* **1989**, *81*, 706–709. [CrossRef]
- 205. Sohn, J.H.; Kim, Y.T.; Rha, S.Y.; Yoo, N.C.; Roh, J.K.; Kim, B.S.; Suh, C.O.; Kim, G.E.; Jang, W.I.; Chung, H.C. Paclitaxel and Cisplatin Combination Chemotherapy in Pretreated Breast Cancer. *Cancer Res. Treat.* 2003, 35, 267–273. [CrossRef] [PubMed]
- 206. Elserafi, M.M.; Zeeneldin, A.A.; Abdelsalam, I.M.; Nassar, H.R.; Moneer, M.M.; Buhoush, W.H. First-line paclitaxel and cisplatin used sequentially or in combination in metastatic breast cancer: A phase II randomized study. *J. Egypt. Natl. Cancer Inst.* **2018**, *30*, 13–20. [CrossRef] [PubMed]
- 207. Shetti, D.; Zhang, B.; Fan, C.; Mo, C.; Lee, B.H.; Wei, K. Low Dose of Paclitaxel Combined with XAV939 Attenuates Metastasis, Angiogenesis and Growth in Breast Cancer by Suppressing Wnt Signaling. *Cells* 2019, 8. [CrossRef] [PubMed]
- 208. Cardone, R.A.; Greco, M.R.; Zeeberg, K.; Zaccagnino, A.; Saccomano, M.; Bellizzi, A.; Bruns, P.; Menga, M.; Pilarsky, C.; Schwab, A.; et al. A novel NHE1-centered signaling cassette drives epidermal growth factor receptor-dependent pancreatic tumor metastasis and is a target for combination therapy. *Neoplasia* 2015, 17, 155–166. [CrossRef]
- Raudenska, M.; Balvan, J.; Fojtu, M.; Gumulec, J.; Masarik, M. Unexpected therapeutic effects of cisplatin. *Metallomics* 2019, 11, 1182–1199. [CrossRef]
- Rosenberg, B. Cisplatin: Its history and possible mechanisms of action. In *Cisplatin*; Elsevier: Cambridge, MA, USA, 1980; pp. 9–20. [CrossRef]
- 211. Makovec, T. Cisplatin and beyond: Molecular mechanisms of action and drug resistance development in cancer chemotherapy. *Radiol. Oncol.* **2019**, *53*, 148–158. [CrossRef]
- 212. Chen, Z.; Ai, L.; Mboge, M.Y.; Tu, C.; McKenna, R.; Brown, K.D.; Heldermon, C.D.; Frost, S.C. Differential expression and function of CAIX and CAXII in breast cancer: A comparison between tumorgraft models and cells. *PLoS ONE* **2018**, *13*, e0199476. [CrossRef]

- 213. Dauchy, R.T.; Xiang, S.; Mao, L.; Brimer, S.; Wren, M.A.; Yuan, L.; Anbalagan, M.; Hauch, A.; Frasch, T.; Rowan, B.G.; et al. Circadian and melatonin disruption by exposure to light at night drives intrinsic resistance to tamoxifen therapy in breast cancer. *Cancer Res.* 2014, 74, 4099–4110. [CrossRef]
- 214. Lacerda, J.Z.; Ferreira, L.C.; Lopes, B.C.; Aristizabal-Pachon, A.F.; Bajgelman, M.C.; Borin, T.F.; Zuccari, D. Therapeutic Potential of Melatonin in the Regulation of MiR-148a-3p and Angiogenic Factors in Breast Cancer. *Microrna* **2019**, *8*, 237–247. [CrossRef]
- 215. Hill, S.M.; Belancio, V.P.; Dauchy, R.T.; Xiang, S.; Brimer, S.; Mao, L.; Hauch, A.; Lundberg, P.W.; Summers, W.; Yuan, L.; et al. Melatonin: An inhibitor of breast cancer. *Endocr. Relat. Cancer* **2015**, *22*, R183–R204. [CrossRef]
- 216. Mediavilla, M.D.; Sanchez-Barcelo, E.J.; Tan, D.X.; Manchester, L.; Reiter, R.J. Basic mechanisms involved in the anti-cancer effects of melatonin. *Curr. Med. Chem.* **2010**, *17*, 4462–4481. [CrossRef] [PubMed]
- 217. Hasan, M.; Marzouk, M.A.; Adhikari, S.; Wright, T.D.; Miller, B.P.; Matossian, M.D.; Elliott, S.; Wright, M.; Alzoubi, M.; Collins-Burow, B.M.; et al. Pharmacological, Mechanistic, and Pharmacokinetic Assessment of Novel Melatonin-Tamoxifen Drug Conjugates as Breast Cancer Drugs. *Mol. Pharmacol.* 2019, *96*, 272–296. [CrossRef] [PubMed]
- 218. De Castro, T.B.; Bordin-Junior, N.A.; de Almeida, E.A.; de Campos Zuccari, D.A.P. Evaluation of melatonin and AFMK levels in women with breast cancer. *Endocrine* **2018**, *62*, 242–249. [CrossRef] [PubMed]
- 219. Nooshinfar, E.; Safaroghli-Azar, A.; Bashash, D.; Akbari, M.E. Melatonin, an inhibitory agent in breast cancer. *Breast Cancer* 2017, 24, 42–51. [CrossRef] [PubMed]
- 220. Sonehara, N.M.; Lacerda, J.Z.; Jardim-Perassi, B.V.; de Paula, R., Jr.; Moschetta-Pinheiro, M.G.; Souza, Y.S.T.; de Andrade, J.C.J.; De Campos Zuccari, D.A.P. Melatonin regulates tumor aggressiveness under acidosis condition in breast cancer cell lines. *Oncol. Lett.* **2019**, *17*, 1635–1645. [CrossRef] [PubMed]
- 221. Alvarez-García, V.; González, A.; Alonso-González, C.; Martínez-Campa, C.; Cos, S. Regulation of vascular endothelial growth factor by melatonin in human breast cancer cells. *J. Pineal Res.* 2013, 54, 373–380. [CrossRef] [PubMed]
- 222. Reiter, R.J.; Rosales-Corral, S.A.; Tan, D.X.; Acuna-Castroviejo, D.; Qin, L.; Yang, S.F.; Xu, K. Melatonin, a Full Service Anti-Cancer Agent: Inhibition of Initiation, Progression and Metastasis. *Int. J. Mol. Sci.* 2017, 18. [CrossRef]
- 223. Menéndez-Menéndez, J.; Hermida-Prado, F.; Granda-Díaz, R.; González, A.; García-Pedrero, J.M.; Del-Río-Ibisate, N.; González-González, A.; Cos, S.; Alonso-González, C.; Martínez-Campa, C. Deciphering the molecular basis of melatonin protective effects on breast cells treated with doxorubicin: TWIST1 a transcription factor involved in EMT and metastasis, a novel target of melatonin. *Cancers (Basel)* 2019, *11*, 1011. [CrossRef]
- 224. Borin, T.F.; Arbab, A.S.; Gelaleti, G.B.; Ferreira, L.C.; Moschetta, M.G.; Jardim-Perassi, B.V.; Iskander, A.S.; Varma, N.R.; Shankar, A.; Coimbra, V.B.; et al. Melatonin decreases breast cancer metastasis by modulating Rho-associated kinase protein-1 expression. *J. Pineal Res.* **2016**, *60*, 3–15. [CrossRef]
- 225. Jardim-Perassi, B.V.; Lourenco, M.R.; Doho, G.M.; Grigolo, I.H.; Gelaleti, G.B.; Ferreira, L.C.; Borin, T.F.; Moschetta, M.G.; Pires de Campos Zuccari, D.A. Melatonin Regulates Angiogenic Factors under Hypoxia in Breast Cancer Cell Lines. *Anticancer Agents Med. Chem.* 2016, 16, 347–358. [CrossRef]
- 226. Chottanapund, S.; Van Duursen, M.B.; Navasumrit, P.; Hunsonti, P.; Timtavorn, S.; Ruchirawat, M.; Van den Berg, M. Anti-aromatase effect of resveratrol and melatonin on hormonal positive breast cancer cells co-cultured with breast adipose fibroblasts. *Toxicol. In Vitro* **2014**, *28*, 1215–1221. [CrossRef] [PubMed]
- 227. Wu, K.H.; Ho, C.T.; Chen, Z.F.; Chen, L.C.; Whang-Peng, J.; Lin, T.N.; Ho, Y.S. The apple polyphenol phloretin inhibits breast cancer cell migration and proliferation via inhibition of signals by type 2 glucose transporter. *J. Food Drug Anal.* **2018**, *26*, 221–231. [CrossRef] [PubMed]
- 228. Meiners, C. Clinical Response of Metastatic Breast Cancer to Multi-targeted Therapeutic Approach: A Single Case Report. *Cancers (Basel)* **2011**, *3*, 1454–1466. [CrossRef] [PubMed]
- 229. Wang, Y.C.; Chao, T.K.; Chang, C.C.; Yo, Y.T.; Yu, M.H.; Lai, H.C. Drug screening identifies niclosamide as an inhibitor of breast cancer stem-like cells. *PLoS ONE* **2013**, *8*, e74538. [CrossRef]
- 230. Pronzato, P.; Amoroso, D.; Bertelli, G.; Conte, P.F.; Cusimano, M.P.; Ciottoli, G.B.; Gulisano, M.; Lionetto, R.; Rosso, R. Phase II study of lonidamine in metastatic breast cancer. *Br. J. Cancer* **1989**, *59*, 251–253. [CrossRef]
- Bougnoux, P.; Hajjaji, N.; Ferrasson, M.N.; Giraudeau, B.; Couet, C.; Le Floch, O. Improving outcome of chemotherapy of metastatic breast cancer by docosahexaenoic acid: A phase II trial. *Br. J. Cancer* 2009, 101, 1978–1985. [CrossRef]

- 232. Wannous, R.; Bon, E.; Gillet, L.; Chamouton, J.; Weber, G.; Brisson, L.; Goré, J.; Bougnoux, P.; Besson, P.; Roger, S. Suppression of PPARβ, and DHA treatment, inhibit NaV1. 5 and NHE-1 pro-invasive activities. *Pflug. Arch.* 2014, 467, 1249–1259. [CrossRef]
- 233. Li, Y.; Li, P.K.; Roberts, M.J.; Arend, R.C.; Samant, R.S.; Buchsbaum, D.J. Multi-targeted therapy of cancer by niclosamide: A new application for an old drug. *Cancer Lett.* **2014**, *349*, 8–14. [CrossRef]
- Naujokat, C.; Steinhart, R. Salinomycin as a drug for targeting human cancer stem cells. J. Biomed. Biotechnol. 2012, 2012, 950658. [CrossRef]
- Lega, I.C.; Austin, P.C.; Gruneir, A.; Goodwin, P.J.; Rochon, P.A.; Lipscombe, L.L. Association between metformin therapy and mortality after breast cancer: A population-based study. *Diabetes Care* 2013, 36, 3018–3026. [CrossRef]
- Roshan, M.H.; Shing, Y.K.; Pace, N.P. Metformin as an adjuvant in breast cancer treatment. SAGE Open Med. 2019, 7, 2050312119865114. [CrossRef]
- 237. Grover-McKay, M.; Walsh, S.A.; Seftor, E.A.; Thomas, P.A.; Hendrix, M.J. Role for glucose transporter 1 protein in human breast cancer. *Pathol. Oncol. Res.* **1998**, *4*, 115–120. [CrossRef] [PubMed]
- Garrido, P.; Osorio, F.G.; Moran, J.; Cabello, E.; Alonso, A.; Freije, J.M.; Gonzalez, C. Loss of GLUT4 induces metabolic reprogramming and impairs viability of breast cancer cells. *J. Cell. Physiol.* 2015, 230, 191–198. [CrossRef] [PubMed]
- 239. Wang, J.; Li, G.; Wang, Y.; Tang, S.; Sun, X.; Feng, X.; Li, Y.; Bao, G.; Li, P.; Mao, X. Suppression of tumor angiogenesis by metformin treatment via a mechanism linked to targeting of HER2/HIF-1α/VEGF secretion axis. *Oncotarget* 2015, *6*, 44579. [CrossRef] [PubMed]
- 240. Schexnayder, C.; Broussard, K.; Onuaguluchi, D.; Poche, A.; Ismail, M.; McAtee, L.; Llopis, S.; Keizerweerd, A.; McFerrin, H.; Williams, C. Metformin Inhibits Migration and Invasion by Suppressing ROS Production and COX2 Expression in MDA-MB-231 Breast Cancer Cells. *Int. J. Mol. Sci.* 2018, *19.* [CrossRef] [PubMed]
- 241. Fan, C.; Wang, Y.; Liu, Z.; Sun, Y.; Wang, X.; Wei, G.; Wei, J. Metformin exerts anticancer effects through the inhibition of the Sonic hedgehog signaling pathway in breast cancer. *Int. J. Mol. Med.* **2015**, *36*, 204–214. [CrossRef]
- 242. Riobo-Del Galdo, N.A.; Lara Montero, A.; Wertheimer, E.V. Role of Hedgehog Signaling in Breast Cancer: Pathogenesis and Therapeutics. *Cells* **2019**, *8*. [CrossRef]
- Iliopoulos, D.; Hirsch, H.A.; Struhl, K. Metformin decreases the dose of chemotherapy for prolonging tumor remission in mouse xenografts involving multiple cancer cell types. *Cancer Res.* 2011, 71, 3196–3201. [CrossRef] [PubMed]
- 244. Vazquez-Martin, A.; Oliveras-Ferraros, C.; Del Barco, S.; Martin-Castillo, B.; Menendez, J.A. The anti-diabetic drug metformin suppresses self-renewal and proliferation of trastuzumab-resistant tumor-initiating breast cancer stem cells. *Breast Cancer Res. Treat.* **2011**, *126*, 355–364. [CrossRef]
- 245. Leone, A.; Di Gennaro, E.; Bruzzese, F.; Avallone, A.; Budillon, A. New perspective for an old antidiabetic drug: Metformin as anticancer agent. *Cancer Treat. Res.* **2014**, *159*, 355–376. [CrossRef]
- 246. De, A.; Kuppusamy, G. Metformin in breast cancer: Preclinical and clinical evidence. *Curr. Probl. Cancer* 2019. [CrossRef] [PubMed]
- 247. Bayraktar, S.; Hernadez-Aya, L.F.; Lei, X.; Meric-Bernstam, F.; Litton, J.K.; Hsu, L.; Hortobagyi, G.N.; Gonzalez-Angulo, A.M. Effect of metformin on survival outcomes in diabetic patients with triple receptor-negative breast cancer. *Cancer* **2012**, *118*, 1202–1211. [CrossRef] [PubMed]
- 248. Qu, H.; Yang, X. Metformin inhibits angiogenesis induced by interaction of hepatocellular carcinoma with hepatic stellate cells. *Cell Biochem. Biophys.* **2015**, *71*, 931–936. [CrossRef] [PubMed]
- 249. Guimaraes, T.A.; Farias, L.C.; Santos, E.S.; de Carvalho Fraga, C.A.; Orsini, L.A.; de Freitas Teles, L.; Feltenberger, J.D.; de Jesus, S.F.; de Souza, M.G.; Santos, S.H.; et al. Metformin increases PDH and suppresses HIF-1alpha under hypoxic conditions and induces cell death in oral squamous cell carcinoma. *Oncotarget* 2016, 7, 55057–55068. [CrossRef]
- 250. Han, J.; Li, Y.; Liu, X.; Zhou, T.; Sun, H.; Edwards, P.; Gao, H.; Yu, F.S.; Qiao, X. Metformin suppresses retinal angiogenesis and inflammation in vitro and in vivo. *PLoS ONE* **2018**, *13*, e0193031. [CrossRef]
- 251. Ni, H.Z.; Liu, Z.; Sun, L.L.; Zhou, M.; Liu, C.; Li, W.D.; Li, X.Q. Metformin inhibits angiogenesis of endothelial progenitor cells via miR-221-mediated p27 expression and autophagy. *Future Med. Chem.* 2019, 11, 2263–2272. [CrossRef]

- 252. Blagosklonny, M.V. Hypoxia-inducible factor: Achilles' heel of antiangiogenic cancer therapy (review). *Int. J. Oncol.* **2001**, *19*, 257–262. [CrossRef]
- 253. De Francesco, E.M.; Lappano, R.; Santolla, M.F.; Marsico, S.; Caruso, A.; Maggiolini, M. HIF-1alpha/GPER signaling mediates the expression of VEGF induced by hypoxia in breast cancer associated fibroblasts (CAFs). *Breast Cancer Res.* **2013**, *15*, R64. [CrossRef]
- 254. Briggs, K.J.; Koivunen, P.; Cao, S.; Backus, K.M.; Olenchock, B.A.; Patel, H.; Zhang, Q.; Signoretti, S.; Gerfen, G.J.; Richardson, A.L.; et al. Paracrine Induction of HIF by Glutamate in Breast Cancer: EglN1 Senses Cysteine. *Cell* **2016**, *166*, 126–139. [CrossRef]
- 255. Song, C.W.; Lee, H.; Dings, R.P.; Williams, B.; Powers, J.; Santos, T.D.; Choi, B.H.; Park, H.J. Metformin kills and radiosensitizes cancer cells and preferentially kills cancer stem cells. *Sci. Rep.* **2012**, *2*, 362. [CrossRef]
- 256. Turashvili, G.; Brogi, E. Tumor Heterogeneity in Breast Cancer. Front. Med. 2017, 4, 227. [CrossRef] [PubMed]
- 257. Waks, A.G.; Winer, E.P. Breast Cancer Treatment: A Review. JAMA 2019, 321, 288-300. [CrossRef]
- 258. Puddefoot, J.; Baker, V.; Bakkers, B.; Marsigliante, S.; Barker, S.; Panahy, C.; Goode, A.; Carpenter, R.; Vinson, G. The nature and significance of multiple isoforms of the oestrogen receptor in breast tumours. *J. Mol. Endocrinol.* **1993**, *11*, 83–90. [CrossRef] [PubMed]
- 259. Leclercq, G. Molecular forms of the estrogen receptor in breast cancer. J. Steroid Biochem. Mol. Biol. 2002, 80, 259–272. [CrossRef]
- 260. Brahimi-Horn, C.; Pouyssegur, J. The role of the hypoxia-inducible factor in tumor metabolism growth and invasion. *Bull. Cancer* **2006**, *93*, E73–E80.
- Williams, B.; Howard, R.L. Glucose-induced changes in Na+/H+ antiport activity and gene expression in cultured vascular smooth muscle cells. Role of protein kinase C. J. Clin. Invest. 1994, 93, 2623–2631. [CrossRef]
- 262. Hawsawi, Y.M.; Al-Numair, N.S.; Sobahy, T.M.; Al-Ajmi, A.M.; Al-Harbi, R.M.; Baghdadi, M.A.; Oyouni, A.A.; Alamer, O.M. The role of BRCA1/2 in hereditary and familial breast and ovarian cancers. *Mol. Genet. Genomic Med.* 2019, 7, e879. [CrossRef]
- 263. Taherian-Esfahani, Z.; Taheri, M.; Dashti, S.; Kholghi-Oskooei, V.; Geranpayeh, L.; Ghafouri-Fard, S. Assessment of the expression pattern of mTOR-associated lncRNAs and their genomic variants in the patients with breast cancer. J. Cell. Physiol. 2019, 234, 22044–22056. [CrossRef]
- 264. Nijhout, H.F. Metaphors and the role of genes in development. *Bioessays* 1990, 12, 441–446. [CrossRef]
- 265. Lim, B.; Woodward, W.A.; Wang, X.; Reuben, J.M.; Ueno, N.T. Inflammatory breast cancer biology: The tumour microenvironment is key. *Nat. Rev. Cancer* **2018**, *18*, 485. [CrossRef]
- 266. Barriere, H.; Poujeol, C.; Tauc, M.; Blasi, J.M.; Counillon, L.; Poujeol, P. CFTR modulates programmed cell death by decreasing intracellular pH in Chinese hamster lung fibroblasts. *Am. J. Physiol. Cell Physiol.* 2001, 281, C810–C824. [CrossRef]
- 267. Harguindey, S.; Gonzalez Molinillo, J.; Chinchilla, D.; Reshkin, S.; Tomoda, A. Further Along a Clinical Protocol Using a Cocktail of PTIs in Human Cancer. In Proceedings of the 2nd ISPD Meeting, Nice, France, 18–19 November 2011.
- 268. Matthews, H.; Ranson, M.; Kelso, M.J. Anti-tumour/metastasis effects of the potassium-sparing diuretic amiloride: An orally active anti-cancer drug waiting for its call-of-duty? *Int. J. Cancer* 2011, 129, 2051–2061. [CrossRef] [PubMed]
- Harguindey, S. Use of Na+/H+ antiporter inhibitors as a novel approach to cancer treatment. In *Amiloride and its Analogs: Unique Cation Transport Inhibitors*; VCH Publishers Inc.: New York, NY, USA, 1992; pp. 317–334.
- 270. Rath, S.; Liebl, J.; Furst, R.; Vollmar, A.M.; Zahler, S. Regulation of endothelial signaling and migration by v-ATPase. *Angiogenesis* **2014**, *17*, 587–601. [CrossRef] [PubMed]
- 271. Wang, X.; Liu, C.; Wang, J.; Fan, Y.; Wang, Z.; Wang, Y. Proton pump inhibitors increase the chemosensitivity of patients with advanced colorectal cancer. *Oncotarget* **2017**, *8*, 58801–58808. [CrossRef] [PubMed]
- 272. Avnet, S.; Lemma, S.; Cortini, M.; Pellegrini, P.; Perut, F.; Zini, N.; Kusuzaki, K.; Chano, T.; Grisendi, G.; Dominici, M.; et al. Altered pH gradient at the plasma membrane of osteosarcoma cells is a key mechanism of drug resistance. *Oncotarget* 2016, *7*, 63408–63423. [CrossRef] [PubMed]
- 273. Canitano, A.; Iessi, E.; Spugnini, E.P.; Federici, C.; Fais, S. Proton pump inhibitors induce a caspase-independent antitumor effect against human multiple myeloma. *Cancer Lett.* **2016**, *376*, 278–283. [CrossRef]

- 274. Papagerakis, S.; Bellile, E.; Peterson, L.A.; Pliakas, M.; Balaskas, K.; Selman, S.; Hanauer, D.; Taylor, J.M.; Duffy, S.; Wolf, G. Proton pump inhibitors and histamine 2 blockers are associated with improved overall survival in patients with head and neck squamous carcinoma. *Cancer Prev. Res.* 2014, 7, 1258–1269. [CrossRef]
- 275. Falcone, R.; Roberto, M.; D'Antonio, C.; Romiti, A.; Milano, A.; Onesti, C.E.; Marchetti, P.; Fais, S. High-doses of proton pump inhibitors in refractory gastro-intestinal cancer: A case series and the state of art. *Dig. Liver Dis.* 2016, 48, 1503–1505. [CrossRef]
- 276. Ikemura, K.; Hiramatsu, S.; Okuda, M. Drug Repositioning of Proton Pump Inhibitors for Enhanced Efficacy and Safety of Cancer Chemotherapy. *Front. Pharmacol.* 2017, *8*, 911. [CrossRef]
- 277. Fako, V.E.; Wu, X.; Pflug, B.; Liu, J.Y.; Zhang, J.T. Repositioning proton pump inhibitors as anticancer drugs by targeting the thioesterase domain of human fatty acid synthase. *J. Med. Chem.* 2015, *58*, 778–784. [CrossRef]
- 278. Sanchez-Sanchez, A.M.; Antolin, I.; Puente-Moncada, N.; Suarez, S.; Gomez-Lobo, M.; Rodriguez, C.; Martin, V. Melatonin cytotoxicity is associated to warburg effect inhibition in ewing sarcoma cells. *PLoS ONE* 2015, 10, e0135420. [CrossRef] [PubMed]
- 279. De Lima Mota, A.; Jardim-Perassi, B.V.; de Castro, T.B.; Colombo, J.; Sonehara, N.M.; Nishiyama, V.K.G.; Pierri, V.A.G.; de Campos Zuccari, D.A.P. Melatonin modifies tumor hypoxia and metabolism by inhibiting HIF-1α and energy metabolic pathway in the in vitro and in vivo models of breast cancer. *Melatonin Res.* 2019, 2, 83–98. [CrossRef]
- Turner, N.C.; Tutt, A.N. Platinum chemotherapy for BRCA1-related breast cancer: Do we need more evidence? Breast Cancer Res. 2012, 14, 115. [CrossRef]
- Dasari, S.; Tchounwou, P.B. Cisplatin in cancer therapy: Molecular mechanisms of action. *Eur. J. Pharmacol.* 2014, 740, 364–378. [CrossRef]
- 282. Shirmanova, M.V.; Druzhkova, I.N.; Lukina, M.M.; Dudenkova, V.V.; Ignatova, N.I.; Snopova, L.B.; Shcheslavskiy, V.I.; Belousov, V.V.; Zagaynova, E.V. Chemotherapy with cisplatin: Insights into intracellular pH and metabolic landscape of cancer cells in vitro and in vivo. *Sci. Rep.* 2017, 7, 8911. [CrossRef]
- 283. Boscoboinik, D.; Gupta, R.S.; Epand, R.M. Investigation of the relationship between altered intracellular pH and multidrug resistance in mammalian cells. *Br. J. Cancer* **1990**, *61*, 568–572. [CrossRef]
- 284. Epand, R.F.; Epand, R.M.; Gupta, R.S.; Cragoe, E.J., Jr. Reversal of intrinsic multidrug resistance in Chinese hamster ovary cells by amiloride analogs. *Br. J. Cancer* **1991**, *63*, 247–251. [CrossRef]
- 285. Omran, Z.; Scaife, P.; Stewart, S.; Rauch, C. Physical and biological characteristics of multi drug resistance (MDR): An integral approach considering pH and drug resistance in cancer. *Semin. Cancer Biol.* 2017, 43, 42–48. [CrossRef]
- 286. Alfarouk, K.O.; Stock, C.M.; Taylor, S.; Walsh, M.; Muddathir, A.K.; Verduzco, D.; Bashir, A.H.; Mohammed, O.Y.; Elhassan, G.O.; Harguindey, S.; et al. Resistance to cancer chemotherapy: Failure in drug response from ADME to P-gp. *Cancer Cell Int.* 2015, *15*, 71. [CrossRef]
- 287. Wei, L.Y.; Roepe, P.D. Low external pH and osmotic shock increase the expression of human MDR protein. *Biochemistry* **1994**, *33*, 7229–7238. [CrossRef]
- 288. Jin, W.; Li, Q.; Lin, Y.; Lu, Y.; Li, H.; Wang, L.; Hu, R.; Ma, L.; Wang, J.; Pang, T. Reversal of Imatinib resistance in BCR-ABL-positive leukemia after inhibition of the Na+/H+ exchanger. *Cancer Lett.* 2011, 308, 81–90. [CrossRef] [PubMed]
- 289. Li, Q.H.; Lu, Y.; Jin, W.N.; Lin, Y.N.; Hu, R.H.; Zhu, X.F.; Wang, J.X.; Pang, T.X. Effect of intracellular acidification on drug resistance of leukemia cells with high P-glycoprotein expression. *Zhonghua Xue Ye Xue Za Zhi* **2009**, *30*, 605–609. [PubMed]
- 290. Gallagher Ferdia, A.; Kettunen Mikko, I.; Day Sam, E. Magnetic resonance imaging of pH in vivo using hyperpolarized 13C labelled bicarbonate. *Nature* **2008**, *453*, 940–944. [CrossRef]
- 291. Lemma, S.; Di Pompo, G.; Porporato, P.E.; Sboarina, M.; Russell, S.; Gillies, R.J.; Baldini, N.; Sonveaux, P.; Avnet, S. MDA-MB-231 breast cancer cells fuel osteoclast metabolism and activity: A new rationale for the pathogenesis of osteolytic bone metastases. *Biochim. Biophys. Acta Mol. Basis Dis.* 2017, 1863, 3254–3264. [CrossRef] [PubMed]
- 292. Lee, J.O.; Lee, S.K.; Kim, J.H.; Kim, N.; You, G.Y.; Moon, J.W.; Kim, S.J.; Park, S.H.; Kim, H.S. Metformin regulates glucose transporter 4 (GLUT4) translocation through AMP-activated protein kinase (AMPK)-mediated Cbl/CAP signaling in 3T3-L1 preadipocyte cells. *J. Biol. Chem.* 2012, 287, 44121–44129. [CrossRef] [PubMed]

- 293. Barron, C.; Tsiani, E.; Tsakiridis, T. Expression of the glucose transporters GLUT1, GLUT3, GLUT4 and GLUT12 in human cancer cells. *BMC Proc.* **2012**. [CrossRef]
- 294. Adekola, K.; Rosen, S.T.; Shanmugam, M. Glucose transporters in cancer metabolism. *Curr. Opin. Oncol.* 2012, 24, 650–654. [CrossRef]
- 295. Chen, C.; Lou, T. Hypoxia inducible factors in hepatocellular carcinoma. *Oncotarget* 2017, *8*, 46691–46703. [CrossRef]
- 296. Dallaglio, K.; Bruno, A.; Cantelmo, A.R.; Esposito, A.I.; Ruggiero, L.; Orecchioni, S.; Calleri, A.; Bertolini, F.; Pfeffer, U.; Noonan, D.M.; et al. Paradoxic effects of metformin on endothelial cells and angiogenesis. *Carcinogenesis* 2014, 35, 1055–1066. [CrossRef]
- 297. Zheng, L.; Yang, W.; Wu, F.; Wang, C.; Yu, L.; Tang, L.; Qiu, B.; Li, Y.; Guo, L.; Wu, M.; et al. Prognostic significance of AMPK activation and therapeutic effects of metformin in hepatocellular carcinoma. *Clin. Cancer Res.* **2013**, *19*, 5372–5380. [CrossRef]
- 298. Incio, J.; Suboj, P.; Chin, S.M.; Vardam-Kaur, T.; Liu, H.; Hato, T.; Babykutty, S.; Chen, I.; Deshpande, V.; Jain, R.K.; et al. Metformin Reduces Desmoplasia in Pancreatic Cancer by Reprogramming Stellate Cells and Tumor-Associated Macrophages. *PLoS ONE* **2015**, *10*, e0141392. [CrossRef] [PubMed]
- 299. Phoenix, K.N.; Vumbaca, F.; Claffey, K.P. Therapeutic metformin/AMPK activation promotes the angiogenic phenotype in the ERalpha negative MDA-MB-435 breast cancer model. *Breast Cancer Res. Treat.* **2009**, *113*, 101–111. [CrossRef] [PubMed]
- 300. Sun, X.J.; Zhang, P.; Li, H.H.; Jiang, Z.W.; Jiang, C.C.; Liu, H. Cisplatin combined with metformin inhibits migration and invasion of human nasopharyngeal carcinoma cells by regulating E-cadherin and MMP-9. *Asian Pac. J. Cancer Prev.* 2014, 15, 4019–4023. [CrossRef] [PubMed]
- 301. Han, B.; Cui, H.; Kang, L.; Zhang, X.; Jin, Z.; Lu, L.; Fan, Z. Metformin inhibits thyroid cancer cell growth, migration, and EMT through the mTOR pathway. *Tumor Biol.* **2015**, *36*, 6295–6304. [CrossRef]
- 302. Son, T.X.; Huyen, N.T.B.; Saimuang, K.; Prachayasittikul, V.; On, W.C. Metformin inhibits migration and invasion of cholangiocarcinoma cells. *Asian Pac. J. Cancer Prev.* **2017**, *18*, 473. [CrossRef]
- 303. Ferretti, A.C.; Hidalgo, F.; Tonucci, F.M.; Almada, E.; Pariani, A.; Larocca, M.C.; Favre, C. Metformin and glucose starvation decrease the migratory ability of hepatocellular carcinoma cells: Targeting AMPK activation to control migration. *Sci. Rep.* **2019**, *9*, 2815. [CrossRef]
- 304. Biondo, L.A.; Batatinha, H.A.; Souza, C.O.; Teixeira, A.A.S.; Silveira, L.S.; Alonso-Vale, M.I.; Oyama, L.M.; Alves, M.J.; Seelaender, M.; Neto, J.C.R. Metformin Mitigates Fibrosis and Glucose Intolerance Induced by Doxorubicin in Subcutaneous Adipose Tissue. *Front. Pharmacol.* 2018, *9*, 452. [CrossRef]
- 305. Marino, M.L.; Fais, S.; Djavaheri-Mergny, M.; Villa, A.; Meschini, S.; Lozupone, F.; Venturi, G.; Della Mina, P.; Pattingre, S.; Rivoltini, L.; et al. Proton pump inhibition induces autophagy as a survival mechanism following oxidative stress in human melanoma cells. *Cell Death Dis.* **2010**, *1*, e87. [CrossRef]
- 306. Cecconi, F.; Jaattela, M. Targeting ions-induced autophagy in cancer. Cancer Cell 2014, 26, 599-600. [CrossRef]
- 307. Lugini, L.; Matarrese, P.; Tinari, A.; Lozupone, F.; Federici, C.; Iessi, E.; Gentile, M.; Luciani, F.; Parmiani, G.; Rivoltini, L.; et al. Cannibalism of live lymphocytes by human metastatic but not primary melanoma cells. *Cancer Res.* 2006, *66*, 3629–3638. [CrossRef]
- 308. Fais, S.; Overholtzer, M. Cell-in-cell phenomena in cancer. Nat. Rev. Cancer 2018, 18, 758–766. [CrossRef] [PubMed]
- Haupt, S.; Keam, S.P.; Haupt, Y. Cannibalism in Breast Cancer: The Dangers of Overeating. *Trends Cancer* 2019, 5, 761–762. [CrossRef] [PubMed]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).