



Review

Spoilage of Chilled Fresh Meat Products during Storage: A Quantitative Analysis of Literature Data

Ngoc-Du Martin Luong¹, Louis Coroller², Monique Zagorec¹, Jeanne-Marie Membre¹ and Sandrine Guillou^{1,*}

¹ SECALIM, INRAE, ONIRIS, Université Bretagne Loire, Route de Gachet, CS 40706, F-44307 Nantes, France; ngoc-du.luong@inrae.fr (N.-D.M.L.); monique.zagorec@oniris-nantes.fr (M.Z.); Jeanne-marie.membre@oniris-nantes.fr (J.-M.M.)

² Université de Brest, Laboratoire Universitaire de Biodiversité et Ecologie Microbienne, UMT Alter'ix, F-29334 Quimper, France; louis.coroller@univ-brest.fr

* Correspondence: sandrine.guillou@oniris-nantes.fr; Tel.: +33-(0)2-40-68-7763

Received: 24 July 2020; Accepted: 3 August 2020; Published: 6 August 2020



Abstract: A literature search was performed on spoilage of fresh meat products by combining keyword query, text mining and expert elicitation. From the 258 collected studies, a quantitative analysis was first performed to identify the methods which are the most used to evaluate spoilage beside the preservation strategies suggested. In a second step focusing on a subset of 24 publications providing quantitative data on spoilage occurrence time, associations between spoilage occurrence time of meat products and specific spoilage indicators were investigated. The analysis especially focused on factors well represented in the 24 publications, i.e., gas packaging (O₂ and CO₂) and storage temperature. Relationships between spoilage occurrence and several microbiological indicators were also sought. The results point out possible advantages of removing dioxygen in packaging to delay spoilage occurrence, whereas, in the presence of dioxygen, the carbon dioxide proportion in the gas mixtures was shown to influence spoilage occurrence. The collected data clearly reveal a potentially protective role of lactic acid bacteria. Besides, while a spoilage role could be attributed to *Pseudomonas* spp., the growth of mesophilic aerobic microbes, *Brochothrix* spp. and *Enterobacteriaceae* seemed independent of spoilage occurrence time.

Keywords: microbiological spoilage; modified atmosphere packaging; storage temperature; expert knowledge; text-mining; data analysis

1. Introduction

Meat products are an excellent source of valuable nutrients for human consumption, such as animal protein, phosphorus, zinc and iron [1,2]. The global consumption of meat products in 2015 was estimated at 41.3 kg per capita (bovine meat at 10.1 kg per capita, ovine and caprine meat at 2.1 kg per capita, pork meat at 15 kg per capita (excluding China) and poultry meat at 13.8 kg per capita) [3]. However, an important portion of meat and meat products are also lost or wasted every year. These annual losses represent approximately 20% of the initial meat production (304.2 million tons). These losses are essentially due to spoilage, characterized by a decrease in the sensory quality of the meat products [4]. There is a diversity of spoilage characteristics in meat products for consumers, such as changes in aspect (texture, slime or liquid production), various color deteriorations or off-odors [5,6]. These spoilage defects generally take place by different mechanisms. Meat spoilage can be caused by natural processes in meat, such as lipid oxidation [7,8] or autolytic enzymatic reactions in the muscle cells of the animal after slaughtering [9]. However, the major cause of spoilage is the unavoidable contamination by microorganisms (essentially bacteria) during the processing of animals into meat

products [10] and their subsequent growth and metabolic activities during storage. Each processing step can influence microbial contamination, and storage conditions can shape the structure of bacterial communities, consequently impacting the occurrence of microbial spoilage over time [11].

Spoilage may shorten meat product shelf-life and lead to their rejection by consumers because quality expectations are not met [12]. Spoilage of products of animal origin has been recently reviewed [13]. In the latter paper, microorganisms associated with meat and dairy product spoilage are reviewed and factors influencing spoilage such as temperature, pH, salt and packaging atmosphere are described. However, the effects of factors are sometimes difficult to interpret since experimental conditions and food matrices may differ tremendously between studies. The aim of the present study was to collect studies through a rigorous procedure to perform a review of meat spoilage, in an ultimate objective to provide a quantitative analysis of the effects of selected processing factors on spoilage occurrence. First, studies were gathered by using both keyword search, expert elicitation and text mining. Second, from relevant selected studies, a quantitative analysis of the literature was performed on the methods generally used to monitor meat spoilage and preservation strategies suggested. At last, from a subset of publications providing quantitative data on spoilage over time, associations between spoilage occurrence time of meat products and specific spoilage indicators were investigated, highlighting the effects of storage temperature, dioxygen and carbon dioxide in Modified Atmosphere Packaging (MAP) on spoilage.

2. Materials and Methods

2.1. Query Process of Literature Data

Our bibliographic search aimed to collect relevant data associated with microbiological spoilage of meat products from scientific literature. A preliminary search from the Science Direct database, based on three main keywords (“meat” AND “microbiology” AND “spoilage”) in full-text, provided more than 8000 records. However, these records were mostly not relevant, since many of them were studies on pathogens in meat or in other food products (seafood, vegetables, etc.). Thus, an alternative procedure, combining expert opinion and data mining, was adopted for identifying the relevant keywords for search query functions.

The procedure used to extract relevant keywords for query functions is detailed in Figure 1. First, 63 publications were initially provided by experts. Afterwards, the keyword extraction from the full text of these 63 publications was done using a text mining approach [14,15] using the tm R package [16]. This approach extracted the most relevant terms after different text treatment steps (filtering, categorization) based on correlations between terms, i.e., the occurrence frequency of multiple keywords in the same article [15]. The extraction was carried out following the procedure outlined by Williams (2016) [17]. The text mining of 63 publications provided an output of 18 relevant keywords. The data searches were carried out afterwards using advanced searches of full texts in two scientific platforms: Science Direct and Web of Science. While query functions required the “AND” function for the three principal keywords, (“meat”, “microbiology” AND “spoilage”), all supplementary extracted keywords were added with the “OR” option. The keyword and search function used were thus written as follows:

FULL-TEXT (meat * AND spoil * AND microbiology * AND (Brochothrix OR community OR data OR food OR lactate OR “lactic acid bacteria” OR “modified atmosphere packaging” OR odor OR psychrotroph * OR Pseudomonas OR volatile OR salt OR sodium OR refrigerated OR yeast)).

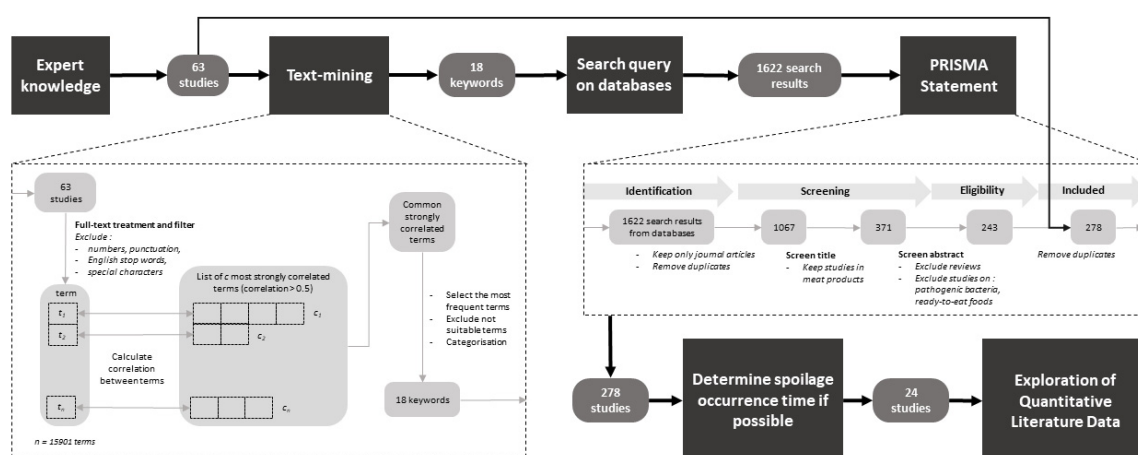


Figure 1. Flow chart outlining the procedure for quantitative data collection from the literature.

The latest query process using those above-mentioned keywords was performed on the 7 July 2019 and identified 1622 records. Afterwards, these records were filtered in accordance with the PRISMA Statement Guidelines [18] (Figure 1). Different criteria were used to exclude non-relevant publications after several stages of screening: (i) selecting from databases: only research journal articles were kept (book chapters, short communications and other article types were excluded) and duplicates from databases were removed; (ii) screening using titles: only studies on meat products in research articles were kept; (iii) screening using abstracts: review articles, studies exclusively on pathogenic bacteria in meat products and ready-to-eat foods were excluded; and (iv) removing duplicates already provided by experts.

After the screening using titles and abstracts, there were 243 research articles reporting microbiological spoilage in meat products, to which the 63 experts' articles were added. After the removal of 20 duplicates, the remaining 278 articles were filtered, while removing reviews and studies specifically concerning modeling strategies, giving in total 258 articles. The overview of methods of spoilage characterization and preservation strategies was performed based upon these 258 publications. Then, in a more quantitative part, a subset of publications was performed to calculate what we called the spoilage occurrence time in selected publications providing quantitative data related to spoilage over time. In this respect, only studies in which sensorial evaluations were performed on meat products stored at temperatures between 0 and 12 °C were kept. Next, we removed studies in which the time interval between two sensory evaluations exceeded three days. This interval was arbitrarily chosen to approximate the determination of spoilage occurrence time with a maximum error margin of three days.

2.2. Collection of Quantitative Data Associated with Spoilage Occurrence in Meat Products

2.2.1. Determination of Spoilage Occurrence Time

In each study, based on the characterization of organoleptic defects by scores (appearance, color, odor and flavor), the spoilage occurrence time was calculated as the time at which the considered scores exceeded a preliminary threshold value chosen by the authors. In cases where experimental sampling was carried out daily, we considered the observed spoilage occurrence time as the exact sampling time point. Otherwise, in cases where spoilage was observed after a two- or three-day interval between consecutive sampling times, spoilage was assumed to occur between the two sampling time points, and the spoilage occurrence time was considered as the median value of this interval (expressed in days).

2.2.2. Factors Considered as Influencing Spoilage

In the 24 selected studies, a multifactorial design, i.e., combination of several factors, was followed. The considered factors were the type of meat products, storage temperature, initial gas composition (percentage of dioxygen, carbon dioxide and inert gases such as nitrogen or argon) of the packaging headspace (air, modified atmosphere packaging and vacuum-packaging). Hence, combinations of factor levels constituted different initial experimental conditions. For example: a study i on two types of meat products stored under three different modified atmospheres constituted six initial experimental conditions ($n_i = 6$). In each study, the spoilage time was then estimated from multiple replicates, based on the criteria from sensory analyses detailed in the above section.

2.2.3. Microbiological Data

Microbiological data associated with spoilage were collected from the selected studies. High throughput sequencing data were provided in some studies, but this type of information was not available often enough to be considered for the present study. Therefore, we considered only enumeration data acquired by conventional cultural methods for the five most studied microbial indicators, i.e., total mesophilic aerobic counts, lactic acid bacteria (LAB), *Brochothrix* spp., *Pseudomonas* spp. and *Enterobacteriaceae*. For each initial experimental condition, we determined the count of each microbial indicator observed at the spoilage occurrence time. If experimental sampling was carried out daily, we considered the bacterial count as the value observed at the sampling time point. Otherwise, if spoilage was observed after a two- or three-day interval between consecutive sampling times, the bacterial count at the spoilage occurrence time was considered as the average count observed at the two sampling time points.

The gathered dataset used for quantitative analyses included the following for each initial experimental condition: meat type, storage temperature ($^{\circ}\text{C}$), initial composition of the atmosphere packaging (initial percentage of dioxygen and carbon dioxide), the corresponding spoilage occurrence time (days) and the bacterial count observed at the spoilage occurrence time (log CFU/g).

2.3. Statistical Analyses

The symmetrical Gaussian distribution of different studied variables, such as spoilage occurrence time, storage temperature or bacterial counts, were tested using the Shapiro–Wilk test. The evaluation of the difference between two groups following Gaussian distributions was performed using the Student's t -test; otherwise, the non-parametric Wilcoxon rank sum test for non-Gaussian distributions was used. The difference between more than two groups was tested using ANOVA and Kruskal–Wallis rank sum tests for variables following Gaussian and non-Gaussian distributions, respectively. Correlation between variables was evaluated using non-parametric Spearman's rank correlation test.

Pearson's chi-squared tests were performed to evaluate differences between the observed and expected frequencies in contingency tables summarizing relationships between different variables. All statistical analyses were performed with the R program [19]. All graphs were created using the ggplot2 R package [20].

3. Results and Discussion

3.1. Measurement of Spoilage in Meat Products and Control Strategies

3.1.1. Types of Meat and Meat Products Associated with Spoilage

As mentioned previously, the bibliographic search provided a set of 258 original research articles from 1984 to 2018 considered in our review on microbiological spoilage in meat products (Table S1). The most studied meat matrices corresponded to products from pork (117), beef (83) and poultry (54) (chicken or turkey) meats. The remaining meat matrices corresponded to lamb (21 studies), venison, goat, llama, foal or rabbit (fewer than five studies). Considering all meat matrices, the most

studied meat types corresponded to fresh meats such as fillets, ground meats, strips, legs, carcasses, etc. (175 studies). Processed meats such as fresh sausages, dry or fermented sausages, mixed sausages, carpaccio, etc. were studied in 62 studies. Only 40 studied considered cooked meat products such as cooked ham or smoked sausages.

3.1.2. Measurement of Spoilage in Meat Products

The exploration of all studies suggested a very important number of possible approaches for measuring spoilage. The most common methods of spoilage assessment were found to be classified in different categories: microbiological analysis, sensorial analysis and physicochemical analyses. Figure 2 illustrates the evolution of spoilage measurement among publications over the years.

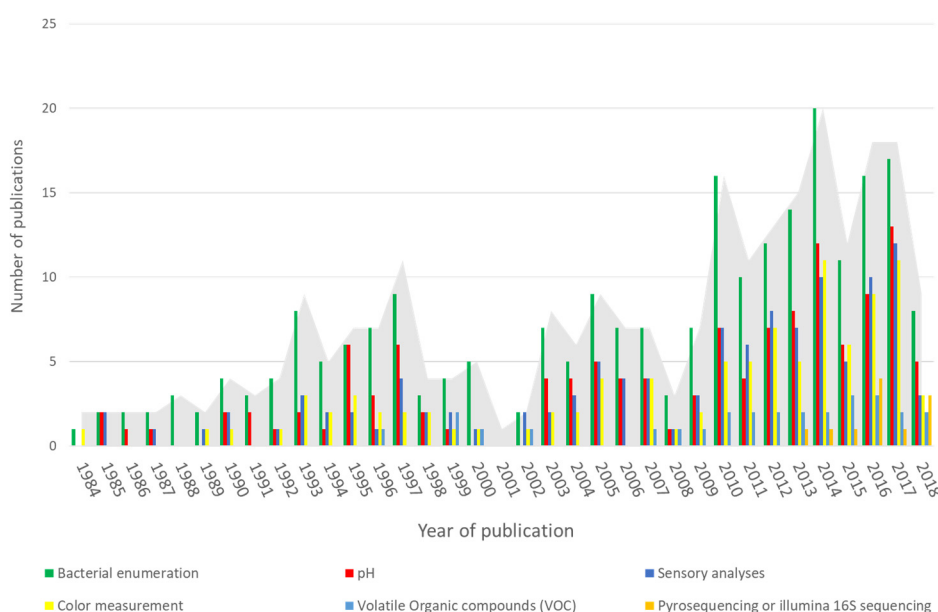


Figure 2. Publications considered in the review and evolution of approaches of spoilage assessment.

It appears that microbiological analyses remained the preferred strategy of spoilage assessment over the years. It especially concerns enumeration of bacterial groups. In recent years, recent high-throughput technologies involving 16S DNA amplicon sequencing has emerged but regarding our dataset, it still corresponds to a limited number of publications. Besides, sensory analyses have always represented a large part of the responses studied while considering spoilage.

Microbiological Analysis

Enumeration of microbial groups by using conventional cultural methods on selective or nonselective agar plates, has constituted the unmissable method for assessing food spoilage. Bacteria were more largely enumerated than molds and yeast (46 studies). The most studied bacterial groups corresponded to Lactic Acid Bacteria (173 studies), total mesophilic aerobic bacteria (TAMB) (162 studies), *Enterobacteriaceae* (113 studies), *Pseudomonas* spp. counts (89 studies), *Brochothrix* spp. (81 studies) and coliforms (46). Psychrotrophic aerobic counts, total anaerobic counts, psychrophilic bacteria or *staphylococci* were less frequently monitored (between 19 and 46 studies), whereas other specific bacterial groups such as proteolytic bacteria (2), *Leuconostoc* (3), *enterococci* (8), H₂S-producing bacteria (7), *Micrococcaceae* (6), or *Aeromonas hydrophila* (2) were scarcely monitored.

Isolate identification was performed in 70 studies plates mainly by genotyping methods such as molecular fingerprinting approaches using restriction enzymatic profiles such as Denaturing Gradient Gel Electrophoresis (DGGE); Restriction Fragment Length Polymorphism (RFLP) or ribotyping; or DNA amplification such as Random Amplified polymorphic DNA (RAPD) and Amplified Fragment Length

Polymorphism (AFLP). These methods have been gradually replaced by sequencing of the 16S gene. Only two studies performed in 2016 and 2018 used mass spectrometry identification MALDI-TOF for isolate identification [21,22]. At last, characterizing microbiological communities in meat products has been nowadays facilitated by several emerging high-throughput sequencing techniques. Eleven studies from 2013 used this technology to assess semi-quantitatively and identify the bacterial communities in meat products [22–32].

Phenotypic characteristics of isolate have been assessed in several studies either in pure culture or by re-inoculating the isolated strains in the products (41 studies). Apart from evaluating the organoleptic properties of inoculated products to confirm or invalidate the spoiler nature of the isolate, other more specific characteristics could be determined. Growth modeling was performed in 35 studies, mainly by using the Gompertz and Baranyi models. Acidification profiles of meat products were widely studied by pH monitoring or measurement of lactic acid [26,33]. Proteolytic, lipolytic or other potential metabolic activities assumed to be associated with a priori spoilage-responsible microorganisms were also measured in some studies [26,34]. Few studies also applied microbial Time Temperature Indicator (TTI) prototype for monitoring spoilage, based on the growth and metabolic activity of specific microorganisms [35,36]. The decomposition zone diameter (DZD) profiles of different microorganisms could be monitored to assess their potential spoilage ability [37]. Finally, some rare studies investigated other microbial abilities such as antibiotic resistance, salt tolerance, metabolite production (lactate, ethanol, glucose, etc.) as well as antibacterial activity towards pathogens [33,38].

Evaluation of the Organoleptic Quality

Based on our dataset, sensory analyses were identified as the most used approaches for evaluating the organoleptic quality of meat products (in 45% of all studies), probably because sensorial profiles are close to spoilage perception by consumers. In general, these approaches consist to determine the acceptability of food products by a panel constituted of several members. The number of members in the panel can vary from one study to another [39,40]. Depending on the types of meat (raw or cooked meats, red or white meats), the panel generally trained during preliminary sessions, determines different possible hedonic scales in order to evaluate organoleptic descriptors to be scored (intensity of off-odors or visible discoloration in fresh meats or general appearance as well as off-flavor for cooked products) [41,42]. A global score can be given afterwards to evaluate spoilage occurrence and intensity of the products using an acceptability rule, for example an arbitrary threshold value chosen by the panel in the used hedonic scale [43].

In addition to the use of sensory analyses, the organoleptic quality of meat products can also be evaluated using specific measurement instruments. For instance, we can cite the CIELAB color system which is widely used for measuring color of meat products [44–47]. Other measurements can also be carried out such as tenderness and texture (e.g., shear force) of beef steaks [48]; the weight loss during meat dry-aging, cooking or storage [49,50]; exudate volume in packages as well as package volume (blowing properties) [47,51,52]; or multispectral imaging assessment of meat surface for monitoring spoilage during storage [53].

Evaluation of Physicochemical Properties

The physicochemical response measured in half of studies is pH (Figure 2) since it reflects both bacterial growth and alteration of taste and flavor. Moreover, it is easy to measure and does not require any specific equipment. Activity water is more rarely measured (in 17 studies).

Other types of physicochemical responses can also be monitored in meat products in order to evaluate indirectly microbiological properties of meat products, assuming that microbiological spoilage results from metabolic activities of microorganisms. The most used responses corresponded to the identification and quantification of volatile organic compounds (VOCs) [54–56]. Volatile fingerprints from microbial activities in meat products could also be monitored using electronic noses for rapid assessment of meat quality [57,58]. Biogenic amines (BA) responsible for allergenic reactions were also

sometimes monitored since their production is due to bacteria producing amino acid decarboxylases likely to induce meat spoilage [26,59,60]. Other chemical indicators for predicting the microbial quality of meat may also be measured even if it is less frequent, for example volatile amines by the total volatile basic nitrogen (TVB-N) or trimethylamine nitrogen (TMA-N) methods [61,62]. Gas composition in products packaging such as O₂ and CO₂ was measured in 38 studies since these gases could be absorbed or produced by microorganisms, and also because they are added in Modified Atmosphere Packaging (MAP) [63,64]. Myoglobin oxygenation and redox state, assessment of lipid oxidation by the TBARS method (Thiobarbituric acid reactive substances) are other methods that are used because they may be associated with organoleptic properties such as taste, odor, flavor or color.

3.1.3. Preservation and Spoilage Control Strategies

Active and Modified Atmosphere Packaging

Figure 3 illustrates the preservation strategies which have been studied in the collected dataset. The improvement in the characteristics of packaging for meat products were identified as the most common used strategies for preventing spoilage during storage. According to our dataset, in more than 50% of publications, meat products were conditioned under MAP. Vacuum packaging was often the other type of packaging under investigation. MAP consists in the replacement of the atmosphere surrounding the product by mixed gas compositions of O₂, CO₂, N₂ and CO, as well as inert gases such as Ar. In general, the common gas compositions correspond to balances between O₂ or CO₂, since high O₂ concentrations are useful for red meat color stabilization [65–67], and increasing CO₂ in MAP is useful for inhibiting microbial growth [21,68]. In addition to MAP, some innovative methods of active packaging were also highlighted, either by injecting vapors of essential oils inside the package [69] or impregnating the film with essential oil [38,39,70–76]. The use of biodegradable nanocomposite packaging films containing TiO₂ and rosemary essential oil [77], the adjustment of oxygen permeability for the films [78], the supercritical carbon dioxide (SC-CO₂) treatment on the inactivation of the natural microbial growth [79] or chitosan/cyclodextrin treatment for packaging films [80] were also studied.

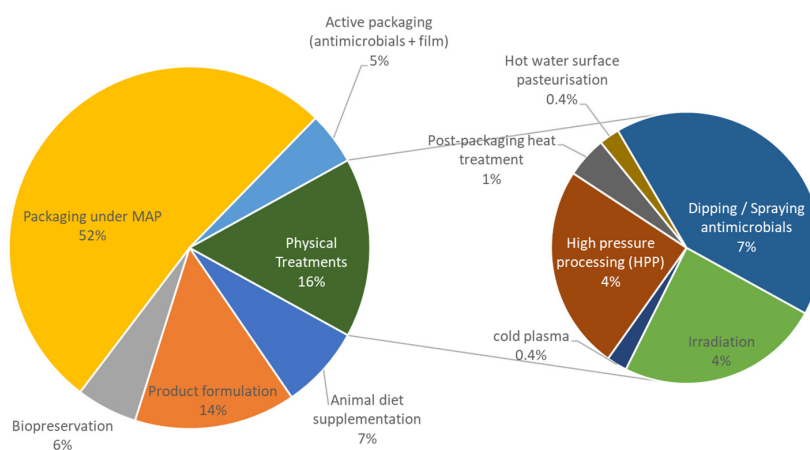


Figure 3. Main preservation strategies identified in collected studies.

Formulation

Formulations of meat products was identified as a commonly studied strategy to prevent spoilage (used in 37 of 258 studies). Antimicrobial compounds susceptible to influence positively spoilage-related responses were used, mainly organic acids such as lactate, marinades, preservatives such as nitrites and essential oils. Many studies focused on the effects induced by lowering the concentration of food additives and/or replacing them by “clean” compounds. Some examples of formulation applications for preserving meat products are shown in Table 1.

Table 1. Application of formulation for preserving fresh meat and processed meat products.

Formulations	Meat Products	Observed Effects	References
Oregano essential oil	Chicken liver meat	Maintenance of freshness and sensorial quality, limitation of lipid oxidation	[81]
Cinnamon essential oil	Pork meat	Increase of microbial shelf life but unacceptable discoloration in high-oxygen atmosphere	[82]
Rosemary extract or oil	Fresh pork sausages	In combination with chitosan, extension of shelf-life of meat products; inhibition of lipid oxidation and rancidity	[83]
Parsley extract	Mortadella-type sausages	Inhibition of <i>L. monocytogenes</i> ; improvement of overall appearance (color, cohesiveness, taste, aroma and saltiness)	[63]
Microencapsulated jaboticaba extract (MJE)	Fresh sausages	Natural dye used in replacement of commercial while maintaining antioxidant and antimicrobial activity and sensory acceptance	[84]
Satureja montana L. essential oil	Mortadella-type sausages	Antioxidant activity	[85]
Mixed spices and marinade (various ingredients and preservatives)	Beef (minced meats or steaks; pork meat)	Alteration of microbial counts due to preservative addition with decrease of microbial diversity dominated by <i>L. algidus</i> and <i>Leuconostoc</i> sp. irrespective of the preservative tested. Glucose and packaging under oxygen in favor of spoilage.	[25,86]
Chitosan	Pork (loins, burgers, sausages); turkey and chicken breasts	Extension of shelf-life of low-sulphite burgers and turkey fillets, with synergistic effect on low dose sulphite or rosemary oil on spoilage prevention.	[41,87]
Sodium nitrite	Minced beef	Combined with essential oil stabilization of red meat color even at low dose; inhibitory effect on bacterial growth, control of lipid oxidation.	[85,88]
NaCl	Pork (loins, ground meat, ham, sausages)	Acid and exudate production following salt reduction, combination of MAP and low salt concentration correlated with sulfurous off-odors and higher spoilage than under vacuum, Maintenance of bacterial richness, inhibition of pathogens. Inhibition of growth of <i>Aeromonas hydrophila</i> at 3%.	[27,89–91]
EDTA	Beefsteaks; chicken (breasts, liver meat)	Combined to oregano essential oil and MAP extended the shelf-life of fresh chicken liver.	[81]
Lysozyme	Synthetic media (target products: processed ham and bologna)	<i>B. thermosphacta</i> inhibited by 500 mg/l or less lysozyme. Lysozyme also effective against <i>P. acidilactici</i> , <i>En. faecalis</i> and <i>W. viridescens</i> .	[89]
Vinegar, acetic acid	Pork sausages, fresh pork	Bacteriostatic properties of vinegar/sodium lactate mixture with reduced bacterial growth. CO ₂ , citric acid and acetic acid reduced total growth.	[92,93]
Lactate	Pork sausages, pork meat, chicken fermented sausages	Lactate-diacetate altered the dynamics dramatically, yielding growth of a single species of <i>Lactobacillus</i> (<i>L. graminis</i>). Psychrotrophic, coliform and lactic acid bacteria retarded by lactate. No effect on sensory properties. Shelf-life extension. Synergistic effect between lactate and carbon dioxide. No or slight effect on color. Reduced effect with increased level of fat.	[24,94–99]
Polydextrose/glucose supplement	Hot-boned, mixed hindquarter cuts; minced beef	Increase of functionality properties of batters made with pre-rigor salted mince with added Polydextrose@. Similar composition and bacterial numbers in mince supplemented with glucose. Glucose, glucose 6-phosphate and lactic acid consumed at slower rates by the flora under MAP than in air.	[88,100]

Physical Treatments

Physical treatments corresponded to 16% of the studies, mainly the application of innovative treatments, such as High Pressure Processing (HPP) [95,101–103], treatments with aqueous ozone and electrolyzed water [104] as well as irradiation [105–114] and the use of cold plasma treatment [115]. Other treatments relative to addition of antimicrobials (urea, nisin, chitosan, oregano essential oil, lactate, etc.) by dipping or spraying the product were also studied as decontamination treatments or more generally as means of extending the shelf-life [41,43,116–128].

Other applications based on the influence of temperatures on spoilage have also been investigated such as rapid chilling, critical for meat hygiene, safety, shelf-life and nutritional quality [11,129], heat treatment at different processing steps [130] or hot water surface pasteurization [131].

Bio-Preservation

The use of controlled microbiota or antimicrobial flora for preserving meat during storage (principle of the bio-preservation) was reported in 14 studies [26,33,89,90,97,132–140]. These studies aimed generally to prevent spoilage occurrence or inhibit pathogenic bacteria to ensure safety by inoculating, during food processes, one or several microorganisms considered as protective cultures especially from LAB groups, or bacteriocins thereof. One can cite for example the use of *Lactobacillus rhamnosus* for inhibiting several strains of *L. monocytogenes* and *S. enterica* in fermented sausages [137], cultures of *Lactococcus lactis* for inhibiting pathogenic or spoilage spore-forming species such as *Bacillus cereus* or *Clostridium botulinum* in ham or Thai traditional fermented sausages [132,140], different mixed protective starters with *Lactobacillus sakei* for inhibiting *Staphylococcus carnosus* and *Staphylococcus xylosum* in dry fermented chicken sausages [97] or the use of *Lactobacillus plantarum* as efficient antimicrobial bio-preservatives on fresh chicken meat [139].

Animal Diet Supplementation

Some authors also studied the influence of the animal diet supplementation, especially in antioxidants in the improvement of the meat quality. These studies represented 7% of the collected data [47,141–152].

3.2. Spoilage Occurrence Time in Meat Products

3.2.1. Spoilage Occurrence Time in Red and White Meat Products

The second selection of publications providing quantitative data on spoilage over time resulted in 24 studies corresponding to data obtained from white (chicken or turkey) or red (beef, pork, ostrich, foal or lamb) meat stored under refrigerated temperatures between 0 and 12 °C. Several types of fresh products, such as raw fillets or fresh sausages, were included in the dataset (Table 2). In total, 84 initial experimental conditions ($N = \sum n_i = 84$) were considered from the 24 chosen studies. The whole dataset obtained provided a global distribution of spoilage occurrence time, which varied from 3 to 26 days under refrigerated temperatures, with a median spoilage time estimated at nine days (Figure 4). Since this distribution was asymmetrical, the median time of nine days was defined as a critical threshold characterizing two principal spoilage stages. Spoilage occurring before the ninth day was considered “early spoilage” while that occurring from the ninth day onward was considered “belated spoilage”. Among the 41 experimental conditions showing an early spoilage stage, no spoilage was observed before three days.

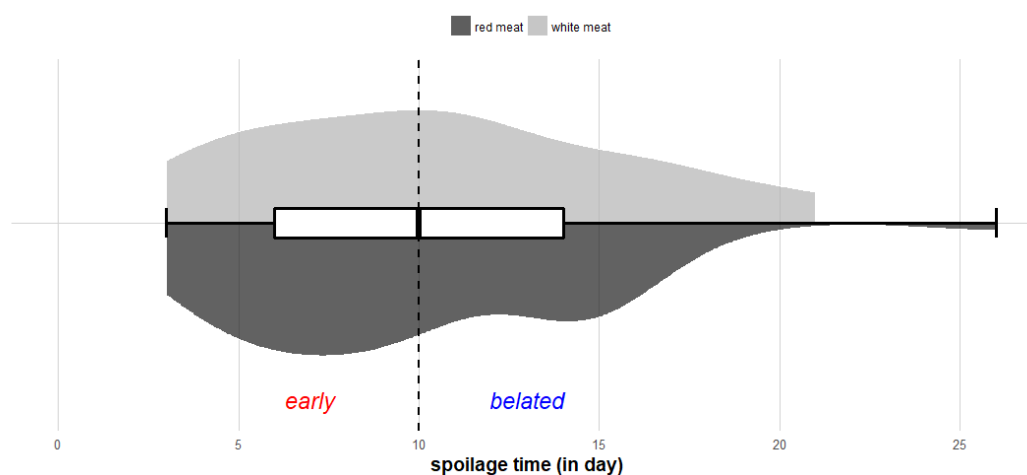


Figure 4. Distribution of spoilage occurrence time. The central boxplot represents the spoilage occurrence time obtained from all experimental conditions. The half violin plot represents spoilage time from studies involving red and white meat, respectively. The vertical dotted line represents the median spoilage time from all conditions, characterizing early and belated spoilage stages.

Spoilage occurred between the third and sixth day in 24 experimental conditions and between the sixth and ninth day in 17 conditions. For conditions corresponding to belated spoilage, the spoilage occurred between the 9th and 16th day in 32 of 43 experiments. Thus, the spoilage occurrence time of fresh meat reached more than 16 days in 11 of 84 experiments under refrigerated temperatures, corresponding to about 13% of cases.

The above variability in the distribution of spoilage time led us to assume, firstly, that spoilage might depend on the type of meat products, i.e., red or white. To consider this, the distributions of spoilage occurrence time for red and white meat products were separately studied (Figure 4). In the case of white meat, the distribution followed a rather symmetrical Gaussian distribution and varied around an average value of 9.9 days. The average spoilage occurrence time in red meat products was slightly lower and estimated at 9.2 days. Although spoilage occurrence in red meats was mainly quite early, the belated spoilage could exceed 21 days and also had a slightly greater variability in comparison with white meat. Surprisingly, our data did not show any significant difference in spoilage occurrence time between red and white meats ($p > 0.05$ —Wilcoxon test). This suggested that spoilage was observed at similar storage times in spite of the different characteristics of red and white meat products. However, the onset and kinetics of spoilage may depend on specific mechanisms, such as protein degradation or preferential metabolisms of spoilage-associated microorganisms, causing modification in color or off-odor [11,68,166].

3.2.2. Influence of Initial Gas Composition in the Packaging

The dynamics of gas composition in the packaging headspace was not available for the 24 chosen studies; therefore, we focused only on initial compositions of the gas mixtures. Besides air or vacuum packaging in which no additional gas was added, several types of gas mixtures were used with different proportions of O_2 and carbon dioxide, completed by inert gases (argon or nitrogen). The used gas mixtures were O_2 - CO_2 , O_2 - CO_2 - N_2 , O_2 - CO_2 -Ar, CO_2 - N_2 , CO_2 -Ar and 100% N_2 . The O_2 and CO_2 proportions varied from 5% to 80%. The distribution of the spoilage stages depending on the initial gas composition of O_2 and CO_2 and different types of packaging (vacuum, air and MAP) is represented in Figure 5. Plotting the spoilage stage as a function of the initial gas composition provided a global visualization of the effect of packaging. This graphical exploration enabled us to identify different visual patterns of spoilage, as well as advantageous options of MAP regarding spoilage delay.

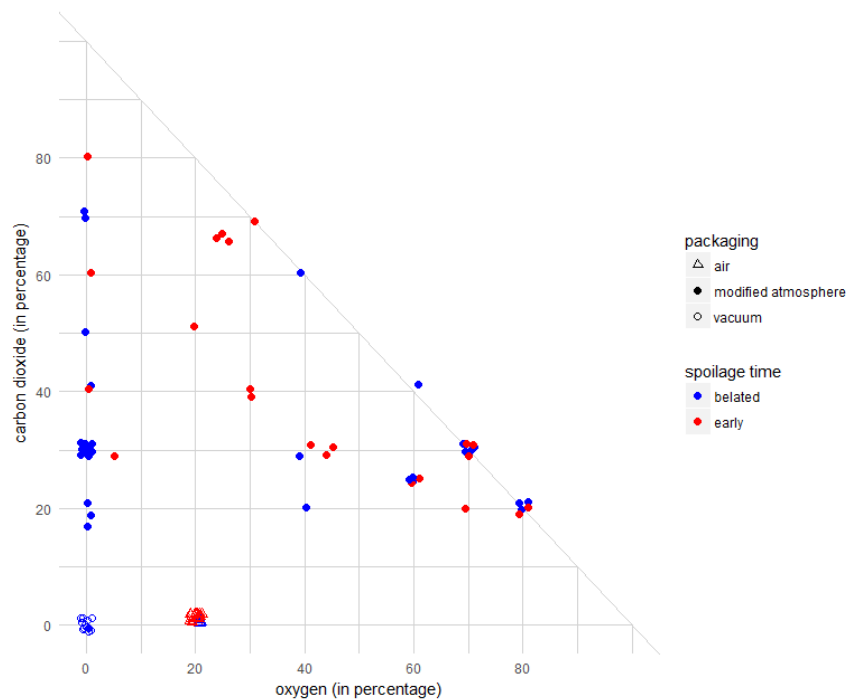


Figure 5. MAP triangular graph depicting the spoilage stage following different initial proportions of O₂ and carbon dioxide in the packaging. The data are represented as jitter points to avoid overplotting. The blue and red symbols correspond to belated and early spoilage, respectively. The shapes of the points correspond to different types of packaging.

Based on our data, removing O₂ in the packaging seemed to be relevant for meat preservation. Indeed, spoilage under anaerobic packaging was generally delayed (Figure 5, blue points on the vertical axis). The packaging containing neither O₂ nor CO₂ represented MAP advantageous options of spoilage delay: the products were spoiled later in seven out of nine experiments. These conditions corresponded to vacuum-packaging or atmosphere under 100% N₂ (rarely used, only in one study). Anaerobic MAP containing carbon dioxide (Figure 5) was observed as another MAP advantageous option of spoilage delay: spoilage was belated in 17 of 22 experiments. In all the above O₂-depleted conditions, no significant difference in the spoilage occurrence time was observed between vacuum and anaerobic atmosphere packaging. Thus, regardless of the packaging mode and carbon dioxide concentration, O₂ depletion appeared beneficial in terms of delaying spoilage occurrence.

Although spoilage occurrence was belated in most cases with anaerobic packaging, it was not possible to conclude with certainty that aerobic packaging accelerates spoilage. O₂ was generally present with other gases. Indeed, gas mixtures with 60–80% O₂ were used in 19 experiments (mostly for red meat products, such as raw pork and beef meat, probably to keep the red color). These compositions led to 7 early and 12 belated spoilages, and it is thus difficult to conclude the role of high O₂ content. Conversely, packaging containing intermediate levels (from 20% to 50%) of O₂ mostly led to early spoilage: 17 early spoilage events occurred in 20 cases under air and 10 events in 13 cases under MAP. Globally, regardless of O₂ content, the spoilage occurrence time was shorter for products under air than under MAP. This difference might be explained by the higher initial concentration of CO₂ in MAP. Finally, removing inert gases (Ar and N₂) from initial aerobic atmosphere did not seem to impact spoilage occurrence time. Indeed, gas mixtures encompassing exclusively O₂ and CO₂ led to five early and nine belated spoilages (Figure 5).

The above findings on various gas compositions (aerobic vs. anaerobic packaging and air vs. aerobic MAP completed with CO₂) revealed implicitly separate effects of the presence of O₂ and CO₂ on spoilage occurrence time. Thus, we found it interesting to study the spoilage time according to the presence of these gases in packaging. A threshold value was then chosen in order to define the limit

between what we considered the “presence” or “absence” of these gases in the packaging. In this study, this value was arbitrarily set at 5% of the gas proportion, since the minimal initial O₂ percentage was 5% among the 84 experimental conditions. The distribution of spoilage time in the groups containing “less than” or “more than” 5% of O₂ and CO₂, are represented in Figure 6.

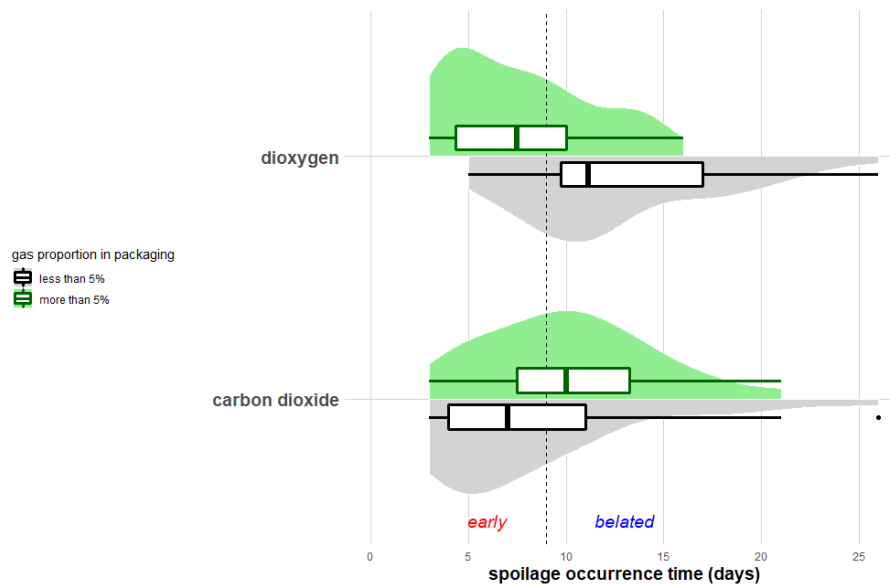


Figure 6. Spoilage occurrence time of meat products stored in packaging containing more or less than 5% of O₂ or carbon dioxide. The vertical dotted line corresponds to the median spoilage time characterizing early and belated spoilage stages.

Spoilage occurrence varied differently in the presence and absence of O₂ (Figure 6). In packaging with less than 5% O₂, the spoilage occurrence time was rather distributed following a symmetrical distribution and varied around an average value of 12.7 days. In these almost anaerobic conditions, spoilage occurred mostly after the ninth day (in 25 of 32 cases). Conversely, with more than 5% O₂, the spoilage occurrence time was asymmetrically distributed, with early occurrence in most cases, especially at the fourth day (Figure 6). Early spoilage corresponded to various gas mixtures, including packaging under air and other MAP, with different O₂-CO₂ proportions. Finally, the comparison of spoilage occurrence time between the “more than 5% O₂” and the “less than 5% O₂” groups highlighted a significant difference (Wilcoxon and Pearson’s chi-squared tests, $p < 0.05$). Therefore, the presence of O₂ in packaging significantly impacted the spoilage occurrence time of fresh meat products.

The spoilage occurrence time seemed to also be impacted by the “more than 5%” or “less than 5%” carbon dioxide groups, since its distribution behaved differently in these conditions (Figure 6). In packaging with less than 5% carbon dioxide, the asymmetrical distribution indicated that there were more early spoiled products than belated ones. With less than 5% of carbon dioxide, most of the cases indicated early spoilage before the fifth day. These cases corresponded exclusively to packaging under air. The effect of CO₂ on spoilage time was also evaluated in the same way as described above for O₂. Surprisingly, our dataset did not show any significant statistical difference between packaging containing more than 5% or less than 5% CO₂.

Spoilage was significantly delayed in the absence of O₂; however, even under anaerobic conditions, the great variability and symmetrical distribution of spoilage occurrence time suggested that O₂ depletion from the packaging did not ensure a belated spoilage for all products. Otherwise, under packaging containing O₂, spoilage time was globally shortened, but also influenced by CO₂ percentage. Moreover, O₂ and CO₂ concentrations in packaging can vary over storage time, which can differentially impact bacterial metabolic activities and meat spoilage processes during storage [68].

3.2.3. Influence of Chilled Storage Temperature

The relationship between storage temperature and spoilage occurrence time is represented in Figure 7. Most experiments under refrigerated temperatures (between 0 and 12 °C) were carried out at 4 °C. The local regression showed a slight decrease of the smooth curve for early spoiled products (before the ninth day) and a stabilization phase at the belated stage (after nine days, inclusively). This trend could be explained by the fact that all products stored at higher than 8 °C spoiled very early. All products stored at temperatures between 2 and 6 °C did not necessarily spoil after our critical threshold of nine days. Indeed, in the case of products stored at 4 °C, the spoilage occurrence time showed great variability from 3 to 21 days. However, despite the observed trend from local regression, our data on refrigerated storage conditions did not enable us to highlight a significant correlation between temperature and spoilage time.

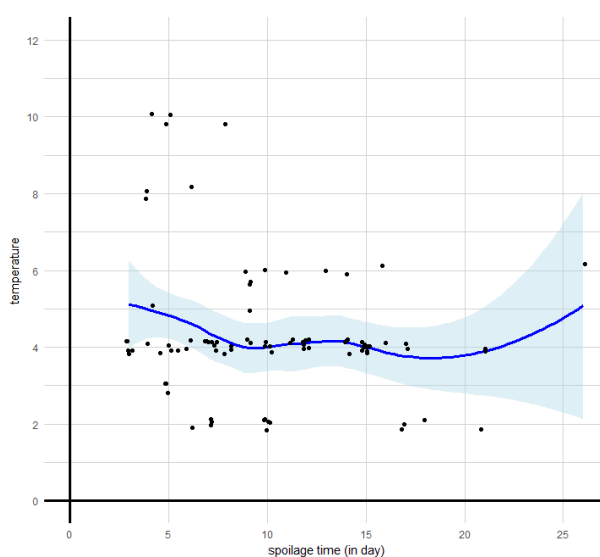


Figure 7. Spoilage occurrence time observed at different storage temperatures. The data are represented as jitter points to avoid overplotting. The thick black line represents the smooth curve obtained by LOESS local non-linear regression. The grey band corresponds to the 95% confidence interval around the smooth curve. The vertical dotted line corresponds to the median spoilage time characterizing early and belated spoilage stages.

In this study, the difficulties in analyzing effects of storage temperature on spoilage occurrence time might be mainly due to several statistical reasons. Since data were collected from experiments performed under different MAP, with various types of meat, temperatures, etc., the data analyses are thus multifactorial. Although effects of temperature on microbial growth in meat products are incontestable [167,168], the multifactorial data analyses may have allowed the effects of storage temperature to be covered up by other factors. Moreover, as seen in Figure 5, very few data points corresponding to a spoilage time greater than 15–16 days were reported, which led to a bad estimation of the confidence interval. Finally, since most experiments were conducted at 4 °C, the non-Gaussian distribution of storage temperature led us to assess correlation between temperature and spoilage occurrence time using the inevitably appropriate non-parametric test, which usually has less statistical power than its parametric equivalent [169].

3.2.4. Relationship between Spoilage Time and Microbiological Indicators

Five principal microbiological indicators were considered, i.e., total mesophilic aerobic counts, LAB, *Brochothrix* spp., *Pseudomonas* spp. and *Enterobacteriaceae*. The CFU enumerations are summarized

in Table 2. The distribution of the counts for each indicator is represented as a function of the spoilage stage (Figure 8).

At the spoilage time and regardless of spoilage stage (early or belated), total mesophilic aerobic counts were observed as the most abundant group (6.52 log CFU/g in average) and *Enterobacteriaceae* as the least abundant group (4.61 log CFU/g in average). However, *Enterobacteriaceae* counts presented the highest variability range in comparison with the other groups, as their values ranged from 0.8 to 9.8 log CFU/g. Mesophilic aerobic counts were observed as the most abundant group, but the least variable, as its distribution varied between 2.8 and 9.8 log CFU/g with the lowest standard deviation. The remaining bacterial indicators, including LAB, *Brochothrix* spp. and *Pseudomonas* spp., had overall average and standard deviation values of approximately the same level. Globally, the results from the whole dataset (84 initial experimental experiments) showed a significant difference in the bacterial counts of the five bacterial indicators observed at the spoilage occurrence time, regardless of the spoilage stage ($p < 0.05$, ANOVA).

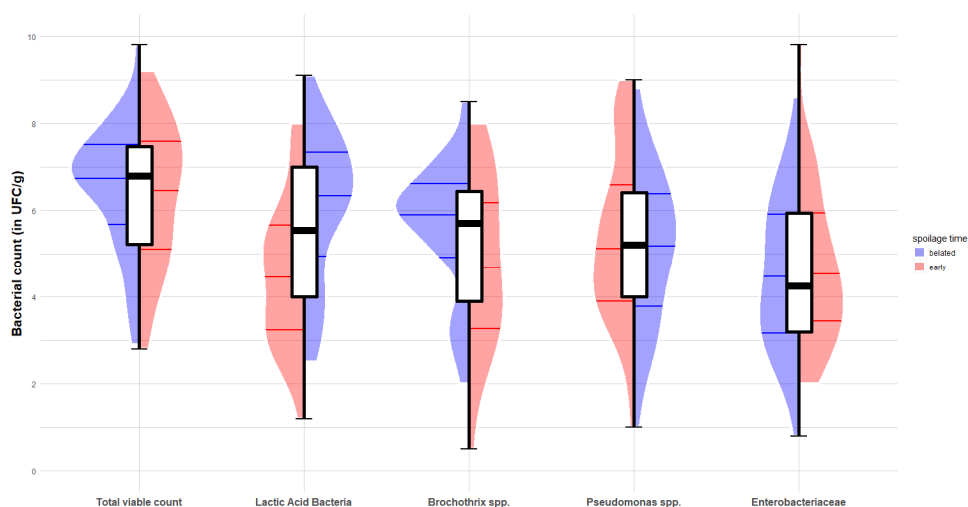


Figure 8. Distribution of bacterial count observed at spoilage occurrence time for different microbiological indicators in early and belated spoilage stages. The different plots correspond to different indicators. For each indicator, the central boxplot represents the overall distribution of the bacterial count. Half violin plots in red (or blue) represent the distribution of the bacterial count at the spoilage occurrence time in the cases of early (or belated) spoilage. Horizontal thick lines on half violin plots correspond to the first quartile, median value and third quartile of each distribution, respectively.

The comparison between early and belated spoilage suggested different distributions of the bacterial indicators in the two spoilage stages (Table 3 and Figure 8). Firstly, the distribution of total mesophilic aerobic counts was similar in early and belated spoiled products. The average counts were estimated, respectively, at 6.31 and 6.71 log CFU/g in early and belated spoilage stages. These results suggest that, regardless of the time at which spoilage occurred (before or after the ninth day), total mesophilic aerobic counts had grown in the same way as before spoilage. Hence, a low total mesophilic aerobic count may not necessarily trigger belated spoilage outcomes. The similarities in bacterial counts between early and belated spoils were also observed for the groups of *Brochothrix* spp., *Pseudomonas* spp. or *Enterobacteriaceae*. These indicators seemed independent of the spoilage stage. In contrast, LAB counts in the belated spoiled products was significantly higher than in the early ones. The observed average values of the LAB count in spoiled products before and after the critical threshold of nine days were, respectively, 4.46 and 6.05 log CFU/g, with more than 3 log CFU/g of variability.

The results suggest that relationships between observed counts and spoilage occurrence time might differ from one bacterial indicator to another. In other words, all the studied groups do not grow in the same way before the occurrence of spoilage, so they may contribute differently to the spoilage

process. To investigate this, for each indicator the correlation between their population level and the spoilage occurrence time was assessed (Table 3 and Figure 9).

For all indicators, spoilage occurrence after the 15th day was exceptional and associated with particularly low bacterial counts. In these conditions, the smooth curves and confidence intervals showed a decreasing trend, which corresponded to 11 experimental conditions from six different studies, mostly concerning raw chicken meat packed under anaerobic MAP or vacuum. Moreover, in two of the six studies, in addition to the use of MAP, meat products were also treated with different combinations of additives, such as oregano, rosemary oil or chitosan. These additives probably contributed to the exceptionally low bacterial population.

Table 3. Enumeration of different bacterial groups at the spoilage occurrence time (overall, early and belated spoilage). N, total number of experimental data collected from all studies.

	Mesophilic Aerobic Counts	Lactic Acid Bacteria	<i>Brochothrix</i> spp.	<i>Pseudomonas</i> spp.	<i>Enterobacteriaceae</i>
N	71	62	45	66	59
[min-max]	[2.8–9.8]	[1.2–9.1]	[0.5–8.5]	[1.0–9.0]	[0.8–9.8]
early	[2.8–9.2]	[1.2–8.0]	[0.5–8.0]	[1.0–9.0]	[2.0–9.8]
belated	[2.9–9.8]	[2.5–9.1]	[2.0–8.5]	[1.0–8.0]	[0.8–8.6]
Mean ± sd	6.52 ± 1.59	5.36 ± 1.89	5.18 ± 1.82	5.28 ± 1.89	4.61 ± 1.83
early	6.31 ± 1.70	4.46 ± 1.68	4.74 ± 2.08	5.43 ± 2.04	4.55 ± 1.90
belated	6.71 ± 1.43	6.05 ± 1.76	5.50 ± 1.56	5.13 ± 1.75	4.67 ± 1.80
Correlation with spoilage time	0.0279	0.3208 *	0.1042	−0.2046	−0.0762

* $p < 0.05$, non-parametric Spearman's rank correlation test.

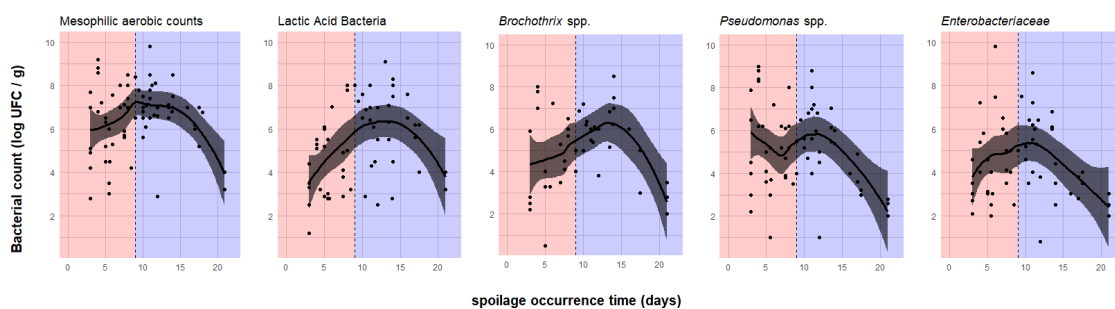


Figure 9. Enumeration of different microbiological indicators observed at spoilage occurrence times. The dotted vertical line corresponds to the median spoilage occurrence time characterizing early and belated spoilage stages, which are represented in red and blue backgrounds, respectively. The thick black curve represents the smooth curve obtained by LOESS local non-linear regression. The grey band corresponds to the 95% confidence interval around the smooth curve.

Considering all products spoiled within 15 days, the non-linear regression shown in Figure 9 suggested a clear ascending trend for the LAB and *Brochothrix* spp. count following spoilage occurrence time, a quite steady trend for the total mesophilic aerobic counts and *Enterobacteriaceae* and a slight decrease before the ninth day for the *Pseudomonas* spp. group. For LAB, a significant positive correlation was observed between the bacterial counts and the spoilage time. This correlation and the observed difference in LAB counts between early and belated spoiled products may, therefore, confirm a potential protective role of this group. Concerning *Brochothrix* spp., no significant statistical correlation between counts and spoilage time was observed. Figure 9 shows a confidence interval estimated much wider on the left side for this group, which means that the available data for *Brochothrix* spp. In early spoiled products are probably insufficient to describe correlation. For total mesophilic aerobic counts and *Enterobacteriaceae*, a steady regression curve and non-significant correlation coefficients (close to 0) were observed between counts and the spoilage occurrence time. Therefore, the development of mesophilic

aerobic counts, *Brochothrix* spp. and *Enterobacteriaceae* seemed to be independent of the spoilage time. Mesophilic aerobic counts encompass a very diverse population. This bacterial indicator is obviously not specific enough to find correlation between counts and spoilage occurrence. For *Pseudomonas* spp., a softly decreasing trend and a slight negative correlation between counts and the spoilage occurrence time supports the assumption of the potential spoiling abilities of these bacteria.

4. Conclusions

The first step of the analysis highlighted the main responses which are generally monitored for spoilage evaluation and preservation strategies suggested. Among 258 studies, only 24 publications provided quantitative data enabling a quantitative analysis of the effects of gas packaging on spoilage occurrence time. It highlights that, in this field, studies are often descriptive or provide end-point characterization of spoilage which does not enable to perform kinetic analysis. Our results give a large overview of the relationships between spoilage occurrence and various indicators. The role of each bacterial group revealed from this study seems consistent with prior knowledge in the literature. However, our analyses were mostly based on cultural enumeration data, since the data issued from literature using high throughput sequencing, which is more informative about the bacterial groups (species or genera), unfortunately reported only bacterial relative abundance. The text-mining approach was also useful to identify relevant literature data, since it identified relevant keywords from articles provided by experts, thus enabling better targeting of further searches on scientific databases. Nevertheless, the quantitative analysis of spoilage-related data was, unfortunately, limited by the data available from research articles. Although the searches were performed on full texts to avoid omission of important studies, the thorough “manual” filtering and required selection criterion regarding the quantitative sensorial data over time led to the ultimate selection of only 24 studies to use for quantitative data analyses. The identification of influencing factors on spoilage occurrence time was consequently performed from quite a small dataset (84 initial experimental conditions in multifactorial dataset). The effects of these factors could not all be evaluated by hypothesis testing because of statistical significance problems. Indeed, using a multitude of hypothesis tests (and *p* values) on the same dataset may lead to uninterpretable test statistics, increasing the “false positive” effect and otherwise covering up the effects of other factors [170,171]. Therefore, for some factors, we preferred avoiding the hypothesis significance testing and using only graph interpretation instead.

We aimed to study the effects of several biological and physicochemical factors on spoilage occurrence time of meat products using a novel method for analyzing literature data. This method of combining experts’ opinions and the text mining approach for extracting relevant information enabled us to give an overview of multiple different studies. It also pointed out the deficit of quantitative data in research articles, which would be beneficial to dispose for further thorough studies relative to spoilage. Within the food microbiology frameworks, such methods could be profitably implemented, for instance for other food products, for which existing data are also available.

Supplementary Materials: Supplementary materials can be found at <http://www.mdpi.com/2076-2607/8/8/1198/s1>, Table S1: Data table of the 258 publications, highlighting the main responses measured to assess spoilage and preservations technologies (excel file).

Author Contributions: Conceptualization, S.G., J.-M.M., L.C. and N.-D.M.L.; methodology, N.-D.M.L.; investigation, N.-D.M.L.; data curation, S.G. and N.-D.M.L.; writing—original draft preparation, N.-D.M.L. and S.G.; writing—review and editing, N.-D.M.L., S.G., J.-M.M., L.C. and M.Z.; supervision, S.G.; project administration, M.Z.; and funding acquisition, M.Z. All authors have read and agreed to the published version of the manuscript.

Funding: This work was funded by the French Research National Agency (ANR) REDLOSSES project (ANR-16-CE21-0006).

Acknowledgments: The authors are grateful to collaborators of the REDLOSSES consortium for their contribution to the data collection and the expert analysis of literature that participated to generate relevant keywords (Émilie Cauchie, Stéphane Chaillou, Marie-Christine Champomier-Vergès, Marie-Hélène Desmonts, Carole Feurer,

Rodéric Gohier, Erwann Hamon, Emmanuel Jaffrès, Sabine Jeuge and Dalal Werner). We also thank Simon de Givry for his advice on text mining approach.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Bohrer, B.M. Review: Nutrient density and nutritional value of meat products and non-meat foods high in protein. *Trends Food Sci. Technol.* **2017**, *65*, 103–112. [[CrossRef](#)]
- Henchion, M.; McCarthy, M.; Resconi, V.C.; Troy, D. Meat consumption: Trends and quality matters. *Meat Sci.* **2014**, *98*, 561–568. [[CrossRef](#)] [[PubMed](#)]
- FAO. *Faostat. Food and Agriculture Organisation of the United Nations*; FAO: Roma, Italy, 2015.
- Gram, L.; Ravn, L.; Rasch, M.; Bruhn, J.B.; Christensen, A.B.; Givskov, M. Food spoilage—interactions between food spoilage bacteria. *Int. J. Food Microbiol.* **2002**, *78*, 79–97. [[CrossRef](#)]
- Borch, E.; Kant-Muermans, M.L.; Blixt, Y. Bacterial spoilage of meat and cured meat products. *Int. J. Food Microbiol.* **1996**, *33*, 103–120. [[CrossRef](#)]
- Dainty, R.H.; Mackey, B.M. The relationship between the phenotypic properties of bacteria from chill-stored meat and spoilage processes. *J. Appl. Bacteriol.* **1992**, *73*, 103s–114s. [[CrossRef](#)]
- Gray, J.I. Measurement of lipid oxidation: A review. *J. Am. Oil Chem. Soc.* **1978**, *55*, 539–546. [[CrossRef](#)]
- Love, J.D.; Pearson, A.M. Lipid oxidation in meat and meat products: A review. *J. Am. Oil Chem. Soc.* **1971**, *48*, 547–549. [[CrossRef](#)]
- Tauro, P.; Kapoor, K.K.; Yadav, K.S. *An Introduction to microbiology*; Wiley Eastern: New Delhi, India, 1986; p. 412.
- Cervený, J.; Meyer, J.D.; Hall, P.A. Microbiological spoilage of meat and poultry products. In *Compendium of the Microbiological Spoilage of Foods and Beverages*; Sperber, W.H., Doyle, M.P., Eds.; Springer: New York, NY, USA, 2009; pp. 69–86. [[CrossRef](#)]
- Dave, D.; Ghaly, A.E. Meat spoilage mechanisms and preservation techniques: A critical review. *Am. J. Agric. Biol. Sci.* **2011**, *6*, 486–510. [[CrossRef](#)]
- Dousset, X.; Jaffrès, E.; Zagorec, M. *Spoilage: Bacterial Spoilage*, 1st ed.; Elsevier Ltd.: Amsterdam, the Netherlands, 2016; Volume 5, pp. 106–112.
- Odeyemi, O.A.; Alegbeleye, O.O.; Strateva, M.; Stratev, D. Understanding spoilage microbial community and spoilage mechanisms in foods of animal origin. *Compr. Rev. Food Sci. Food Saf.* **2020**, *19*, 311–331. [[CrossRef](#)]
- Huang, C.-C.; Lu, Z. Community challenges in biomedical text mining over 10 years: Success, failure and the future. *Brief. Bioinform.* **2016**, *17*, 132–144. [[CrossRef](#)]
- Onan, A.; Korukoğlu, S.; Bulut, H. Ensemble of keyword extraction methods and classifiers in text classification. *Expert Syst. Appl.* **2016**, *57*, 232–247. [[CrossRef](#)]
- Feinerer, I.; Hornik, K. Tm: Text Mining Package. R Package Version 0.7–4. 2008. Available online: <http://tm.r-forge.r-project.org/> (accessed on 23 July 2020).
- Williams, G. Hands-on data science with R text mining. Available online: <https://fr.scribd.com/doc/252462619/Hands-on-Data-Science-with-R-Text-Mining> (accessed on 23 July 2020).
- Moher, D.; Liberati, A.; Tetzlaff, J.; Altman, D.G.; Prisma Group. Preferred reporting items for systematic reviews and meta-analyses: The prisma statement. *Ann. Intern. Med.* **2009**, *151*, 264–269. [[CrossRef](#)] [[PubMed](#)]
- R. Core Team, D.T. *R: A language and environment for statistical computing*; R Foundation for Statistical Computing: Vienna, Austria, 2017.
- Wickham, H. *Ggplot2: Elegant Graphics for Data Analysis (Use R!)*; Springer: Berlin, Germany, 2009.
- Höll, L.; Behr, J.; Vogel, R.F. Identification and growth dynamics of meat spoilage microorganisms in modified atmosphere packaged poultry meat by MALDI-TOF MS. *Food Microbiol.* **2016**, *60*, 84–91. [[CrossRef](#)] [[PubMed](#)]
- Hilgarth, M.; Fuertes, S.; Ehrmann, M.; Vogel, R.F. *Photobacterium carnosum* sp. nov., isolated from spoiled modified atmosphere packaged poultry meat. *Syst. Appl. Microbiol.* **2018**, *41*, 44–50. [[CrossRef](#)] [[PubMed](#)]
- Kiermeier, A.; Tamplin, M.; May, D.; Holds, G.; Williams, M.; Dann, A. Microbial growth, communities and sensory characteristics of vacuum and modified atmosphere packaged lamb shoulders. *Food Microbiol.* **2013**, *36*, 305–315. [[CrossRef](#)]

24. Benson, A.K.; David, J.R.; Gilbreth, S.E.; Smith, G.; Nietfeldt, J.; Legge, R.; Kim, J.; Sinha, R.; Duncan, C.E.; Ma, J.; et al. Microbial successions are associated with changes in chemical profiles of a model refrigerated fresh pork sausage during an 80-day shelf life study. *Appl. Environ. Microbiol.* **2014**, *80*, 5178–5194. [[CrossRef](#)]
25. Stoops, J.; Ruyters, S.; Busschaert, P.; Spaepen, R.; Verreth, C.; Claes, J.; Lievens, B.; Van Campenhout, L. Bacterial community dynamics during cold storage of minced meat packaged under modified atmosphere and supplemented with different preservatives. *Food Microbiol.* **2015**, *48*, 192–199. [[CrossRef](#)]
26. Fontana, C.; Bassi, D.; López, C.; Pisacane, V.; Otero, M.C.; Puglisi, E.; Rebecchi, A.; Cocconcelli, P.S.; Vignolo, G. Microbial ecology involved in the ripening of naturally fermented llama meat sausages. A focus on lactobacilli diversity. *Int. J. Food Microbiol.* **2016**, *236*, 17–25. [[CrossRef](#)]
27. Fougy, L.; Desmots, M.-H.; Coeuret, G.; Fassel, C.; Hamon, E.; Hézard, B.; Champomier-Vergès, M.-C.; Chaillou, S. Reducing salt in raw pork sausages increases spoilage and correlates with reduced bacterial diversity. *Appl. Environ. Microbiol.* **2016**, *82*, 3928–3939. [[CrossRef](#)]
28. Mann, E.; Wetzels, S.U.; Piniór, B.; Metzler-Zebeli, B.U.; Wagner, M.; Schmitz-Esser, S. Psychrophile spoilers dominate the bacterial microbiome in musculature samples of slaughter pigs. *Meat Sci.* **2016**, *117*, 36–40. [[CrossRef](#)]
29. Stellato, G.; La Stora, A.; Filippis, F.D.; Borriello, G.; Villani, F.; Ercolini, D. Overlap of spoilage microbiota between meat and meat processing environment in small-scale vs large-scale retail distribution. *Appl. Environ. Microbiol.* **2016**, *82*, 4045–4054. [[CrossRef](#)] [[PubMed](#)]
30. Såde, E.; Penttinen, K.; Björkroth, J.; Hultman, J. Exploring lot-to-lot variation in spoilage bacterial communities on commercial modified atmosphere packaged beef. *Food Microbiol.* **2017**, *62*, 147–152. [[CrossRef](#)] [[PubMed](#)]
31. Raimondi, S.; Nappi, M.R.; Sirangelo, T.M.; Leonardi, A.; Amaretti, A.; Ulrici, A.; Magnani, R.; Montanari, C.; Tabanelli, G.; Gardini, F.; et al. Bacterial community of industrial raw sausage packaged in modified atmosphere throughout the shelf life. *Int. J. Food Microbiol.* **2018**, *280*, 78–86. [[CrossRef](#)] [[PubMed](#)]
32. Rouger, A.; Moriceau, N.; Prévost, H.; Remenant, B.; Zagorec, M. Diversity of bacterial communities in French chicken cuts stored under modified atmosphere packaging. *Food Microbiol.* **2018**, *70*, 7–16. [[CrossRef](#)]
33. Vermeiren, L.; Devlieghere, F.; Debevere, J. Evaluation of meat born lactic acid bacteria as protective cultures for the biopreservation of cooked meat products. *Int. J. Food Microbiol.* **2004**, *96*, 149–164. [[CrossRef](#)]
34. Ercolini, D.; Casaburi, A.; Nasi, A.; Ferrocino, I.; Di Monaco, R.; Ferranti, P.; Mauriello, G.; Villani, F. Different molecular types of *Pseudomonas fragi* have the same overall behaviour as meat spoilers. *Int. J. Food Microbiol.* **2010**, *142*, 120–131. [[CrossRef](#)]
35. Ellouze, M.; Augustin, J.C. Applicability of biological time temperature integrators as quality and safety indicators for meat products. *Int. J. Food Microbiol.* **2010**, *138*, 119–129. [[CrossRef](#)]
36. Vaikousi, H.; Biliaderis, C.G.; Koutsoumanis, K.P. Applicability of a microbial Time Temperature Indicator (TTI) for monitoring spoilage of modified atmosphere packed minced meat. *Int. J. Food Microbiol.* **2009**, *133*, 272–278. [[CrossRef](#)]
37. Wang, G.-y.; Wang, H.-h.; Han, Y.-w.; Xing, T.; Ye, K.-p.; Xu, X.-L.; Zhou, G.-H. Evaluation of the spoilage potential of bacteria isolated from chilled chicken in vitro and in situ. *Food Microbiol.* **2017**, *63*, 139–146. [[CrossRef](#)]
38. Nowak, A.; Kalembe, D.; Krala, L.; Piotrowska, M.; Czyzowska, A. The effects of thyme (*Thymus vulgaris*) and rosemary (*Rosmarinus officinalis*) essential oils on *Brochothrix thermosphacta* and on the shelf life of beef packaged in high-oxygen modified atmosphere. *Food Microbiol.* **2012**, *32*, 212–216. [[CrossRef](#)]
39. Ntzimani, A.G.; Giatrakou, V.I.; Savvaidis, I.N. Combined natural antimicrobial treatments (EDTA, lysozyme, rosemary and oregano oil) on semi cooked coated chicken meat stored in vacuum packages at 4 °C: Microbiological and sensory evaluation. *Innov. Food Sci. Emerg. Technol.* **2010**, *11*, 187–196. [[CrossRef](#)]
40. Triki, M.; Herrero, A.M.; Jiménez-Colmenero, F.; Ruiz-Capillas, C. Effect of preformed konjac gels, with and without olive oil, on the technological attributes and storage stability of merguez sausage. *Meat Sci.* **2013**, *93*, 351–360. [[CrossRef](#)] [[PubMed](#)]
41. Vasilatos, G.C.; Savvaidis, I.N. Chitosan or rosemary oil treatments, singly or combined to increase turkey meat shelf-life. *Int. J. Food Microbiol.* **2013**, *166*, 54–58. [[CrossRef](#)] [[PubMed](#)]
42. Vestergaard, J.S.; Martens, M.; Turkki, P. Analysis of sensory quality changes during storage of a modified atmosphere packaged meat product (pizza topping) by an electronic nose system. *Lebensm.-Wiss. Technol.* **2007**, *40*, 1083–1094. [[CrossRef](#)]

43. del Río, E.; Panizo-Morán, M.; Prieto, M.; Alonso-Calleja, C.; Capita, R. Effect of various chemical decontamination treatments on natural microflora and sensory characteristics of poultry. *Int. J. Food Microbiol.* **2007**, *115*, 268–280. [[CrossRef](#)]
44. Lorenzo, J.M.; Gómez, M. Shelf life of fresh foal meat under MAP, overwrap and vacuum packaging conditions. *Meat Sci.* **2012**, *92*, 610–618. [[CrossRef](#)]
45. Luño, M.; Beltrán, J.A.; Roncalés, P. Shelf-life extension and colour stabilisation of beef packaged in a low O₂ atmosphere containing CO: Loin steaks and ground meat. *Meat Sci.* **1998**, *48*, 75–84. [[CrossRef](#)]
46. Muermans, M.L.T.; Stekelenburg, F.K.; Zwietering, M.H.; Huis in 't Veld, J.H.J. Modelling of the microbiological quality of meat. *Food Control* **1993**, *4*, 216–221. [[CrossRef](#)]
47. Ortuño, J.; Serrano, R.; Bañón, S. Antioxidant and antimicrobial effects of dietary supplementation with rosemary diterpenes (carnosic acid and carnosol) vs vitamin E on lamb meat packed under protective atmosphere. *Meat Sci.* **2015**, *110*, 62–69. [[CrossRef](#)] [[PubMed](#)]
48. Murphy, K.M.; O'Grady, M.N.; Kerry, J.P. Effect of varying the gas headspace to meat ratio on the quality and shelf-life of beef steaks packaged in high oxygen modified atmosphere packs. *Meat Sci.* **2013**, *94*, 447–454. [[CrossRef](#)]
49. Ahnström, M.L.; Seyfert, M.; Hunt, M.C.; Johnson, D.E. Dry aging of beef in a bag highly permeable to water vapour. *Meat Sci.* **2006**, *73*, 674–679. [[CrossRef](#)] [[PubMed](#)]
50. Torrieri, E.; Russo, F.; Di Monaco, R.; Cavella, S.; Villani, F.; Masi, F. Shelf life prediction of fresh Italian pork sausage modified atmosphere packed. *Food Sci. Technol. Int.* **2011**, *17*, 223–232. [[CrossRef](#)] [[PubMed](#)]
51. Bell, R.G.; Moorhead, S.M.; Broda, D.M. Influence of heat shrink treatments on the onset of clostridial “blown pack” spoilage of vacuum packed chilled meat. *Food Res. Int.* **2001**, *34*, 271–275. [[CrossRef](#)]
52. Broda, D.M.; Delacy, K.M.; Bell, R.G.; Braggins, T.J.; Cook, R.L. Psychrotrophic *Clostridium* spp. associated with ‘blown pack’ spoilage of chilled vacuum-packed red meats and dog rolls in gas-impermeable plastic casings. *Int. J. Food Microbiol.* **1996**, *29*, 335–352. [[CrossRef](#)]
53. Tsakanikas, P.; Pavlidis, D.; Panagou, E.; Nychas, G.-J. Exploiting multispectral imaging for non-invasive contamination assessment and mapping of meat samples. *Talanta* **2016**, *161*, 606–614. [[CrossRef](#)] [[PubMed](#)]
54. Argyri, A.A.; Mallouchos, A.; Panagou, E.Z.; Nychas, G.-J.E. The dynamics of the HS/SPME-GC/MS as a tool to assess the spoilage of minced beef stored under different packaging and temperature conditions. *Int. J. Food Microbiol.* **2015**, *193*, 51–58. [[CrossRef](#)]
55. Casaburi, A.; Piombino, P.; Nychas, G.-J.; Villani, F.; Ercolini, D. Bacterial populations and the volatilome associated to meat spoilage. *Food Microbiol.* **2015**, *45*, 83–102. [[CrossRef](#)]
56. Holm, E.S.; Schäfer, A.; Koch, A.G.; Petersen, M.A. Investigation of spoilage in saveloy samples inoculated with four potential spoilage bacteria. *Meat Sci.* **2013**, *93*, 687–695. [[CrossRef](#)]
57. Blixt, Y.; Borch, E. Using an electronic nose for determining the spoilage of vacuum-packaged beef. *Int. J. Food Microbiol.* **1999**, *46*, 123–134. [[CrossRef](#)]
58. Papadopoulou, O.S.; Tassou, C.C.; Schiavo, L.; Nychas, G.-J.E.; Panagou, E.Z. Rapid Assessment of Meat Quality by Means of an Electronic Nose and Support Vector Machines. *Procedia Food Sci.* **2011**, *1*, 2003–2006. [[CrossRef](#)]
59. Li, M.; Tian, L.; Zhao, G.; Zhang, Q.; Gao, X.; Huang, X.; Sun, L. Formation of biogenic amines and growth of spoilage-related microorganisms in pork stored under different packaging conditions applying PCA. *Meat Sci.* **2014**, *96*, 843–848. [[CrossRef](#)]
60. Pellissery, A.J.; Vinayamohan, P.G.; Amalaradjou, M.A.R.; Venkitanarayanan, K. Chapter 17 - Spoilage bacteria and meat quality. In *Meat Quality Analysis*; Biswas, A.K., Mandal, P.K., Eds.; Academic Press: Cambridge, MA, USA, 2020; pp. 307–334. [[CrossRef](#)]
61. Balamatsia, C.C.; Patsias, A.; Kontominas, M.G.; Savvaidis, I.N. Possible role of volatile amines as quality-indicating metabolites in modified atmosphere-packaged chicken fillets: Correlation with microbiological and sensory attributes. *Food Chem.* **2007**, *104*, 1622–1628. [[CrossRef](#)]
62. Kuswandi, B.; Nurfawaidi, A. On-package dual sensors label based on pH indicators for real-time monitoring of beef freshness. *Food Control* **2017**, *82*, 91–100. [[CrossRef](#)]
63. Riel, G.; Boulaaba, A.; Popp, J.; Klein, G. Effects of parsley extract powder as an alternative for the direct addition of sodium nitrite in the production of mortadella-type sausages—Impact on microbiological, physicochemical and sensory aspects. *Meat Sci.* **2017**, *131*, 166–175. [[CrossRef](#)]

64. Rodrigues, J.B.M.; Sarantópoulos, C.I.G.L.; Bromberg, R.; Andrade, J.C.; Brunelli, K.; Miyagusku, L.; Marquezini, M.G.; Yamada, E.A. Evaluation of the effectiveness of non-irradiated and chlorine-free packaging for fresh beef preservation. *Meat Sci.* **2017**, *125*, 30–36. [[CrossRef](#)]
65. Ercolini, D.; Russo, F.; Blaiotta, G.; Pepe, O.; Mauriello, G.; Villani, F. Simultaneous detection of *Pseudomonas fragi*, *P. lundensis*, and *P. putida* from meat by use of a multiplex PCR assay targeting the *carA* gene. *Appl. Environ. Microbiol.* **2007**, *73*, 2354–2359. [[CrossRef](#)] [[PubMed](#)]
66. Jakobsen, M.; Bertelsen, G. The use of CO₂ in packaging of fresh red meats and its effect on chemical quality changes in the meat: A review. *J. Muscle Foods* **2002**, *13*, 143–168. [[CrossRef](#)]
67. Tremonte, P.; Sorrentino, E.; Succi, M.; Reale, A.; Maiorano, G.; Coppola, R. Shelf life of fresh sausages stored under modified atmospheres. *J. Food Prot.* **2005**, *68*, 2686–2692. [[CrossRef](#)] [[PubMed](#)]
68. McMillin, K.W. Where is MAP Going? A review and future potential of modified atmosphere packaging for meat. *Meat Sci.* **2008**, *80*, 43–65. [[CrossRef](#)] [[PubMed](#)]
69. Skandamis, P.N.; Nychas, G.-J.E. Preservation of fresh meat with active and modified atmosphere packaging conditions. *Int. J. Food Microbiol.* **2002**, *79*, 35–45. [[CrossRef](#)]
70. Sirocchi, V.; Devlieghere, F.; Peelman, N.; Sagratini, G.; Maggi, F.; Vittori, S.; Ragaert, P. Effect of *Rosmarinus officinalis* L. essential oil combined with different packaging conditions to extend the shelf life of refrigerated beef meat. *Food Chem.* **2017**, *221*, 1069–1076. [[CrossRef](#)] [[PubMed](#)]
71. Ammor, M.S.; Argyri, A.A.; Nychas, G.-J.E. Rapid monitoring of the spoilage of minced beef stored under conventionally and active packaging conditions using Fourier transform infrared spectroscopy in tandem with chemometrics. *Meat Sci.* **2009**, *81*, 507–514. [[CrossRef](#)] [[PubMed](#)]
72. Doulgeraki, A.I.; Paramithiotis, S.; Kagkli, D.M.; Nychas, G.-J.E. Lactic acid bacteria population dynamics during minced beef storage under aerobic or modified atmosphere packaging conditions. *Food Microbiol.* **2010**, *27*, 1028–1034. [[CrossRef](#)] [[PubMed](#)]
73. Argyri, A.A.; Doulgeraki, A.I.; Blana, V.A.; Panagou, E.Z.; Nychas, G.-J.E. Potential of a simple HPLC-based approach for the identification of the spoilage status of minced beef stored at various temperatures and packaging systems. *Int. J. Food Microbiol.* **2011**, *150*, 25–33. [[CrossRef](#)]
74. Doulgeraki, A.I.; Paramithiotis, S.; Nychas, G.-J.E. Characterization of the *Enterobacteriaceae* community that developed during storage of minced beef under aerobic or modified atmosphere packaging conditions. *Int. J. Food Microbiol.* **2011**, *145*, 77–83. [[CrossRef](#)] [[PubMed](#)]
75. Doulgeraki, A.I.; Nychas, G.-J.E. Monitoring the succession of the biota grown on a selective medium for pseudomonads during storage of minced beef with molecular-based methods. *Food Microbiol.* **2013**, *34*, 62–69. [[CrossRef](#)]
76. Blana, V.A.; Nychas, G.-J.E. Presence of quorum sensing signal molecules in minced beef stored under various temperature and packaging conditions. *Int. J. Food Microbiol.* **2014**, *173*, 1–8. [[CrossRef](#)]
77. Alizadeh Sani, M.; Ehsani, A.; Hashemi, M. Whey protein isolate/cellulose nanofibre/TiO₂ nanoparticle/rosemary essential oil nanocomposite film: Its effect on microbial and sensory quality of lamb meat and growth of common foodborne pathogenic bacteria during refrigeration. *Int. J. Food Microbiol.* **2017**, *251*, 8–14. [[CrossRef](#)]
78. Cayré, M.E.; Garro, O.; Vignolo, G.; Cayre, M.E.; Garro, O.; Vignolo, G.; Cayré, M.E.; Garro, O.; Vignolo, G.; Cayre, M.E.; et al. Effect of storage temperature and gas permeability of packaging film on the growth of lactic acid bacteria and *Brochothrix thermosphacta* in cooked meat emulsions. *Food Microbiol.* **2005**, *22*, 505–512. [[CrossRef](#)]
79. Ferrentino, G.; Balzan, S.; Spilimbergo, S. Optimization of supercritical carbon dioxide treatment for the inactivation of the natural microbial flora in cubed cooked ham. *Int. J. Food Microbiol.* **2013**, *161*, 189–196. [[CrossRef](#)]
80. Higuera, L.; López-Carballo, G.; Hernández-Muñoz, P.; Catalá, R.; Gavara, R. Antimicrobial packaging of chicken fillets based on the release of carvacrol from chitosan/cyclodextrin films. *Int. J. Food Microbiol.* **2014**, *188*, 53–59. [[CrossRef](#)] [[PubMed](#)]
81. Hasapidou, A.; Savvaidis, I.N. The effects of modified atmosphere packaging, EDTA and oregano oil on the quality of chicken liver meat. *Food Res. Int.* **2011**, *44*, 2751–2756. [[CrossRef](#)]
82. Van Haute, S.; Raes, K.; Devlieghere, F.; Sampers, I. Combined use of cinnamon essential oil and MAP/vacuum packaging to increase the microbial and sensorial shelf life of lean pork and salmon. *Food Packag. Shelf Life* **2017**, *12*, 51–58. [[CrossRef](#)]

83. Georgantelis, D.; Ambrosiadis, I.; Katikou, P.; Blekas, G.; Georgakis, S.A. Effect of rosemary extract, chitosan and alpha-tocopherol on microbiological parameters and lipid oxidation of fresh pork sausages stored at 4 °C. *Meat Sci.* **2007**, *76*, 172–181. [[CrossRef](#)] [[PubMed](#)]
84. Baldin, J.C.; Michelin, E.C.; Polizer, Y.J.; Rodrigues, I.; de Godoy, S.H.S.; Fregonesi, R.P.; Pires, M.A.; Carvalho, L.T.; Fávoro-Trindade, C.S.; de Lima, C.G.; et al. Microencapsulated jabuticaba (*Myrciaria cauliflora*) extract added to fresh sausage as natural dye with antioxidant and antimicrobial activity. *Meat Sci.* **2016**, *118*, 15–21. [[CrossRef](#)] [[PubMed](#)]
85. Coutinho de Oliveira, T.L.; Malfitano de Carvalho, S.; de Araújo Soares, R.; Andrade, M.A.; Cardoso, M.d.G.; Ramos, E.M.; Piccoli, R.H. Antioxidant effects of *Satureja montana* L. essential oil on TBARS and color of mortadella-type sausages formulated with different levels of sodium nitrite. *Lebensm-Wiss. Technol.* **2012**, *45*, 204–212. [[CrossRef](#)]
86. Vihavainen, E.J.; Björkroth, J. Spoilage of value-added, high-oxygen modified-atmosphere packaged raw beef steaks by *Leuconostoc gasicomitatum* and *Leuconostoc gelidum*. *Int. J. Food Microbiol.* **2007**, *119*, 340–345. [[CrossRef](#)]
87. Serrano, R.; Bañón, S. Reducing SO₂ in fresh pork burgers by adding chitosan. *Meat Sci.* **2012**, *92*, 651–658. [[CrossRef](#)]
88. Sadler, D.N.; Swan, J.E. Effect of NaCl, polydextrose, and storage conditions on the functional characteristics and microbial quality of pre- and post-rigor salted beef. *Meat Sci.* **1997**, *46*, 329–338. [[CrossRef](#)]
89. Gill, A.O.; Holley, R.A. Interactive inhibition of meat spoilage and pathogenic bacteria by lysozyme, nisin and EDTA in the presence of nitrite and sodium chloride at 24 [deg]C. *Int. J. Food Microbiol.* **2003**, *80*, 251–259. [[CrossRef](#)]
90. Samapundo, S.; Ampofo-Asiama, J.; Anthierens, T.; Xhaferi, R.; Van Bree, I.; Szczepaniak, S.; Goemaere, O.; Steen, L.; Dhooze, M.; Paelinck, H.; et al. Influence of NaCl reduction and replacement on the growth of *Lactobacillus sakei* in broth, cooked ham and white sauce. *Int. J. Food Microbiol.* **2010**, *143*, 9–16. [[CrossRef](#)] [[PubMed](#)]
91. Palumbo, S.A. The growth of *Aeromonas hydrophila* K144 in ground pork at 5 °C. *Int. J. Food Microbiol.* **1988**, *7*, 41–48. [[CrossRef](#)]
92. Crist, C.A.; Williams, J.B.; Schilling, M.W.; Hood, A.F.; Smith, B.S.; Campano, S.G. Impact of sodium lactate and vinegar derivatives on the quality of fresh Italian pork sausage links. *Meat Sci.* **2014**, *96*, 1509–1516. [[CrossRef](#)] [[PubMed](#)]
93. Schirmer, B.C.; Langsrud, S. A dissolving CO₂ headspace combined with organic acids prolongs the shelf-life of fresh pork. *Meat Sci.* **2010**, *85*, 280–284. [[CrossRef](#)] [[PubMed](#)]
94. Bradford, D.D.; Huffman, D.L.; Egbert, W.R.; Jones, W.R. Low fat fresh pork sausage patty stability in refrigerated storage with potassium lactate. *J. Food Sci.* **1993**, *58*, 488–491. [[CrossRef](#)]
95. Guillou, S.; Lerasle, M.; Simonin, H.; Anthoine, V.; Chéret, R.; Federighi, M.; Membre, J.M. Multi-criteria framework as an innovative tradeoff approach to determine the shelf-life of high pressure-treated poultry. *Int. J. Food Microbiol.* **2016**, *233*, 60–72. [[CrossRef](#)]
96. Devlieghere, F.; Geeraerd, A.H.; Versyck, K.J.; Bernaert, H.; Van Impe, J.F.; Debevere, J. Shelf life of modified atmosphere packed cooked meat products: Addition of Na-lactate as a fourth shelf life determinative factor in a model and product validation. *Int. J. Food Microbiol.* **2000**, *58*, 93–106. [[CrossRef](#)]
97. Deumier, F.; Collignan, A. The effects of sodium lactate and starter cultures on pH, lactic acid bacteria, *Listeria monocytogenes* and *Salmonella* spp. levels in pure chicken dry fermented sausage. *Meat Sci.* **2003**, *65*, 1165–1174. [[CrossRef](#)]
98. Friedrich, L.; Siró, I.; Dalmadi, I.; Horváth, K.; Ágoston, R.; Balla, C. Influence of various preservatives on the quality of minced beef under modified atmosphere at chilled storage. *Meat Sci.* **2008**, *79*, 332–343. [[CrossRef](#)]
99. Magrinya, N.; Terjung, N.; Loeffler, M.; Gibis, M.; Bou, R.; Weiss, J. Influence of fat addition on the antimicrobial activity of sodium lactate, lauric arginate and methylparaben in minced meat. *Int. J. Food Microbiol.* **2015**, *215*, 86–94. [[CrossRef](#)]
100. Lambropoulou, K.A.; Drosinos, E.H.; Nychas, G.J.E. The effect of glucose supplementation on the spoilage microflora and chemical composition of minced beef stored aerobically or under a modified atmosphere at 4 °C. *Int. J. Food Microbiol.* **1996**, *30*, 281–291. [[CrossRef](#)]

101. Lerasle, M.; Federighi, M.; Simonin, H.; Anthoine, V.; Rezé, S.; Chéret, R.; Guillou, S. Combined use of modified atmosphere packaging and high pressure to extend the shelf-life of raw poultry sausage. *Innov. Food Sci. Emerg. Technol.* **2014**, *23*, 54–60. [[CrossRef](#)]
102. Masana, M.O.; Barrio, Y.X.; Palladino, P.M.; Sancho, A.M.; Vaudagna, S.R. High pressure treatments combined with sodium lactate to inactivate *Escherichia coli* O157:H7 and spoilage microbiota in cured beef carpaccio. *Food Microbiol.* **2015**, *46*, 610–617. [[CrossRef](#)] [[PubMed](#)]
103. Patterson, M.F.; McKay, A.M.; Connolly, M.; Linton, M. Effect of high pressure on the microbiological quality of cooked chicken during storage at normal and abuse refrigeration temperatures. *Food Microbiol.* **2010**, *27*, 266–273. [[CrossRef](#)] [[PubMed](#)]
104. Botta, C.; Ferrocino, I.; Cavallero, M.C.; Riva, S.; Giordano, M.; Cocolin, L. Potentially active spoilage bacteria community during the storage of vacuum packaged beefsteaks treated with aqueous ozone and electrolyzed water. *Int. J. Food Microbiol.* **2018**, *266*, 337–345. [[CrossRef](#)] [[PubMed](#)]
105. Farkas, J.; Andrásy, É. Interaction of ionising radiation and acidulants on the growth of the microflora of a vacuum-packaged chilled meat product. *Int. J. Food Microbiol.* **1993**, *19*, 145–152. [[CrossRef](#)]
106. Grandison, A.S.; Jennings, A. Extension of the shelf life of fresh minced chicken meat by electron beam irradiation combined with modified atmosphere packaging. *Food Control* **1993**, *4*, 83–88. [[CrossRef](#)]
107. Gamage, S.D.; Faith, N.G.; Luchansky, J.B.; Buege, D.R.; Ingham, S.C. Inhibition of microbial growth in chub-packed ground beef by refrigeration (2 °C) and medium-dose (2.2 to 2.4 kGy) irradiation. *Int. J. Food Microbiol.* **1997**, *37*, 175–182. [[CrossRef](#)]
108. Kamat, A.S.; Khare, S.; Doctor, T.; Nair, P.M. Control of *Yersinia enterocolitica* in raw pork and pork products by γ -irradiation. *Int. J. Food Microbiol.* **1997**, *36*, 69–76. [[CrossRef](#)]
109. Sørheim, O.; Nissen, H.; Nesbakken, T. The storage life of beef and pork packaged in an atmosphere with low carbon monoxide and high carbon dioxide. *Meat Sci.* **1999**, *52*, 157–164. [[CrossRef](#)]
110. Lacroix, M.; Smoragiewicz, W.; Jobin, M.; Latreille, B.; Krzystyniak, K. Protein quality and microbiological changes in aerobically- or vacuum-packaged, irradiated fresh pork loins. *Meat Sci.* **2000**, *56*, 31–39. [[CrossRef](#)]
111. Lacroix, M.L.; Smoragiewicz, W.; Jobin, M.; Latreille, B.; Krzystyniak, K. The effect of irradiation of fresh pork loins on the protein quality and microbiological changes in aerobically—or vacuum-packaged. *Radiat. Phys. Chem.* **2002**, *63*, 317–322. [[CrossRef](#)]
112. Nortjé, K.; Buys, E.M.; Minnaar, A. Effect of γ -irradiation on the sensory quality of moist beef biltong. *Meat Sci.* **2005**, *71*, 603–611. [[CrossRef](#)] [[PubMed](#)]
113. Min, J.-S.; Lee, S.-O.; Jang, A.; Jo, C.; Lee, M. Irradiation and organic acid treatment for microbial control and the production of biogenic amines in beef and pork. *Food Chem.* **2007**, *104*, 791–799. [[CrossRef](#)]
114. Xavier, M.d.l.P.; Dauber, C.; Mussio, P.; Delgado, E.; Maquieira, A.; Soria, A.; Curuchet, A.; Márquez, R.; Méndez, C.; López, T. Use of mild irradiation doses to control pathogenic bacteria on meat trimmings for production of patties aiming at provoking minimal changes in quality attributes. *Meat Sci.* **2014**, *98*, 383–391. [[CrossRef](#)]
115. Wang, J.; Zhuang, H.; Hinton, A.; Zhang, J. Influence of in-package cold plasma treatment on microbiological shelf life and appearance of fresh chicken breast fillets. *Food Microbiol.* **2016**, *60*, 142–146. [[CrossRef](#)] [[PubMed](#)]
116. Narasimha Rao, D.; Sreenivasamurthy, V. A preliminary study of the effect of urea in the preservation of meat. *Meat Sci.* **1986**, *17*, 251–265. [[CrossRef](#)]
117. Zeitoun, A.A.M.; Debevere, J.M. The effect of treatment with buffered lactic acid on microbial decontamination and on shelf life of poultry. *Int. J. Food Microbiol.* **1990**, *11*, 305–311. [[CrossRef](#)]
118. Zeitoun, A.A.M.; Debevere, J.M. Inhibition, survival and growth of *Listeria monocytogenes* on poultry as influenced by buffered lactic acid treatment and modified atmosphere packaging. *Int. J. Food Microbiol.* **1991**, *14*, 161–169. [[CrossRef](#)]
119. Greer, G.G.; Dilts, B.D. Lactic acid inhibition of the growth of spoilage bacteria and cold tolerant pathogens on pork. *Int. J. Food Microbiol.* **1995**, *25*, 141–151. [[CrossRef](#)]
120. Guerrero, I.; Mendiola, R.; Ponce, E.; Prado, A. Inoculation of lactic acid bacteria on meat surfaces as a means of decontamination in semitropical conditions. *Meat Sci.* **1995**, *40*, 397–411. [[CrossRef](#)]
121. Djenane, D.; Sánchez-Escalante, A.; Beltrán, J.A.; Roncalés, P. The shelf-life of beef steaks treated with dl-lactic acid and antioxidants and stored under modified atmospheres. *Food Microbiol.* **2003**, *20*, 1–7. [[CrossRef](#)]

122. Nattress, F.M.; Baker, L.P. Effects of treatment with lysozyme and nisin on the microflora and sensory properties of commercial pork. *Int. J. Food Microbiol.* **2003**, *85*, 259–267. [[CrossRef](#)]
123. Economou, T.; Pournis, N.; Ntzimani, A.; Savvaidis, I.N. Nisin–EDTA treatments and modified atmosphere packaging to increase fresh chicken meat shelf-life. *Food Chem.* **2009**, *114*, 1470–1476. [[CrossRef](#)]
124. Petrou, S.; Tsiraki, M.; Giatrakou, V.; Savvaidis, I.N. Chitosan dipping or oregano oil treatments, singly or combined on modified atmosphere packaged chicken breast meat. *Int. J. Food Microbiol.* **2012**, *156*, 264–271. [[CrossRef](#)]
125. Latou, E.; Mexis, S.F.; Badeka, A.V.; Kontakos, S.; Kontominas, M.G. Combined effect of chitosan and modified atmosphere packaging for shelf life extension of chicken breast fillets. *Lebensm-Wiss. Technol.* **2014**, *55*, 263–268. [[CrossRef](#)]
126. Pavelková, A.; Kačániová, M.; Horská, E.; Rovná, K.; Hleba, L.; Petrová, J. The effect of vacuum packaging, EDTA, oregano and thyme oils on the microbiological quality of chicken's breast. *Anaerobe* **2014**, *29*, 128–133. [[CrossRef](#)] [[PubMed](#)]
127. Paparella, A.; Mazzarrino, G.; Chaves-López, C.; Rossi, C.; Sacchetti, G.; Guerrieri, O.; Serio, A. Chitosan boosts the antimicrobial activity of Origanum vulgare essential oil in modified atmosphere packaged pork. *Food Microbiol.* **2016**, *59*, 23–31. [[CrossRef](#)] [[PubMed](#)]
128. Geeraerts, W.; Pothakos, V.; De Vuyst, L.; Leroy, F. Diversity of the dominant bacterial species on sliced cooked pork products at expiration date in the Belgian retail. *Food Microbiol.* **2017**, *65*, 236–243. [[CrossRef](#)] [[PubMed](#)]
129. Zhou, G.H.; Xu, X.L.; Liu, Y. Preservation technologies for fresh meat—A review. *Meat Sci.* **2010**, *86*, 119–128. [[CrossRef](#)] [[PubMed](#)]
130. Saucier, L. Microbial spoilage, quality and safety within the context of meat sustainability. *Meat Sci.* **2016**. [[CrossRef](#)] [[PubMed](#)]
131. Hauge, S.J.; Wahlgren, M.; Røtterud, O.-J.; Nesbakken, T. Hot water surface pasteurisation of lamb carcasses: Microbial effects and cost-benefit considerations. *Int. J. Food Microbiol.* **2011**, *146*, 69–75. [[CrossRef](#)] [[PubMed](#)]
132. Noonpakdee, W.; Santivarangkna, C.; Jumriangrit, P.; Sonomoto, K.; Panyim, S. Isolation of nisin-producing *Lactococcus lactis* WNC 20 strain from nham, a traditional Thai fermented sausage. *Int. J. Food Microbiol.* **2003**, *81*, 137–145. [[CrossRef](#)]
133. Papamanoli, E.; Tzanetakis, N.; Litopoulou-Tzanetaki, E.; Kotzekidou, P. Characterization of lactic acid bacteria isolated from a Greek dry-fermented sausage in respect of their technological and probiotic properties. *Meat Sci.* **2003**, *65*, 859–867. [[CrossRef](#)]
134. Vermeiren, L.; Devlieghere, F.; Vandekinderen, I.; Debevere, J. The interaction of the non-bacteriocinogenic *Lactobacillus sakei* 10A and lactocin S producing *Lactobacillus sakei* 148 towards *Listeria monocytogenes* on a model cooked ham. *Food Microbiol.* **2006**, *23*, 511–518. [[CrossRef](#)] [[PubMed](#)]
135. Vasilopoulos, C.; De Mey, E.; Dewulf, L.; Paelinck, H.; De Smedt, A.; Vandendriessche, F.; De Vuyst, L.; Leroy, F. Interactions between bacterial isolates from modified-atmosphere-packaged artisan-type cooked ham in view of the development of a bioprotective culture. *Food Microbiol.* **2010**, *27*, 1086–1094. [[CrossRef](#)]
136. Kalschne, D.L.; Geitenes, S.; Veit, M.R.; Sarmiento, C.M.P.; Colla, E. Growth inhibition of lactic acid bacteria in ham by nisin: A model approach. *Meat Sci.* **2014**, *98*, 744–752. [[CrossRef](#)]
137. Rubio, R.; Martín, B.; Aymerich, M.T.; Garriga, M. The potential probiotic *Lactobacillus rhamnosus* CTC1679 survives the passage through the gastrointestinal tract and its use as starter culture results in safe nutritionally enhanced fermented sausages. *Int. J. Food Microbiol.* **2014**, *186*, 55–60. [[CrossRef](#)]
138. Osés, S.M.; Diez, A.M.; Gómez, E.M.; Wilches-Pérez, D.; Luning, P.A.; Jaime, I.; Rovira, J. Control of *Escherichia coli* and *Listeria monocytogenes* in suckling-lamb meat evaluated using microbial challenge tests. *Meat Sci.* **2015**, *110*, 262–269. [[CrossRef](#)]
139. da Silva Sabo, S.; Pérez-Rodríguez, N.; Domínguez, J.M.; de Souza Oliveira, R.P. Inhibitory substances production by *Lactobacillus plantarum* ST16Pa cultured in hydrolyzed cheese whey supplemented with soybean flour and their antimicrobial efficiency as biopreservatives on fresh chicken meat. *Food Res. Int.* **2017**, *99*, 762–769. [[CrossRef](#)]
140. Ramarason, M.; Guillou, S.; Rossero, A.; Rezé, S.; Anthoine, V.; Moriceau, N.; Martin, J.-L.; Durantou, F.; Zagorec, M. Selection procedure of bioprotective cultures for their combined use with High Pressure Processing to control spore-forming bacteria in cooked ham. *Int. J. Food Microbiol.* **2018**, *276*, 28–38. [[CrossRef](#)] [[PubMed](#)]

141. Mäkelä, P.; Korkeala, H. *Lactobacillus* contamination of cooked ring sausages at sausage processing plants. *Int. J. Food Microbiol.* **1987**, *5*, 323–330. [[CrossRef](#)]
142. Gill, C.O.; Bryant, J. The contamination of pork with spoilage bacteria during commercial dressing, chilling and cutting of pig carcasses. *Int. J. Food Microbiol.* **1992**, *16*, 51–62. [[CrossRef](#)]
143. Gustavsson, P.; Borch, E. Contamination of beef carcasses by psychrotrophic *Pseudomonas* and *Enterobacteriaceae* at different stages along the processing line. *Int. J. Food Microbiol.* **1993**, *20*, 67–83. [[CrossRef](#)]
144. Nerbrink, E.; Borch, E. Evaluation of bacterial contamination at separate processing stages in emulsion sausage production. *Int. J. Food Microbiol.* **1993**, *20*, 37–44. [[CrossRef](#)]
145. Holley, R.A.; Doyon, G.; Fortin, J.; Rodrigue, N.; Carbonneau, M. Post-process, packaging-induced fermentation of delicatessen meats. *Food Res. Int.* **1996**, *29*, 35–48. [[CrossRef](#)]
146. Greer, G.G.; Jones, S.D.M. Quality and bacteriological consequences of beef carcass spray-chilling: Effects of spray duration and boxed beef storage temperature. *Meat Sci.* **1997**, *45*, 61–73. [[CrossRef](#)]
147. Voidarou, C.; Vassos, D.; Rozos, G.; Alexopoulos, A.; Plessas, S.; Tsinas, A.; Skoufou, M.; Stavropoulou, E.; Bezirtzoglou, E. Microbial challenges of poultry meat production. *Anaerobe* **2011**, *17*, 341–343. [[CrossRef](#)]
148. Bañón, S.; Méndez, L.; Almela, E. Effects of dietary rosemary extract on lamb spoilage under retail display conditions. *Meat Sci.* **2012**, *90*, 579–583. [[CrossRef](#)]
149. Ortuño, J.; Serrano, R.; Jordán, M.J.; Bañón, S. Shelf life of meat from lambs given essential oil-free rosemary extract containing carnosic acid plus carnosol at 200 or 400 mgkg⁻¹. *Meat Sci.* **2014**, *96*, 1452–1459. [[CrossRef](#)]
150. Serrano, R.; Jordán, M.J.; Bañón, S. Use of dietary rosemary extract in ewe and lamb to extend the shelf life of raw and cooked meat. *Small Rumin. Res.* **2014**, *116*, 144–152. [[CrossRef](#)]
151. Chulayo, A.Y.; Bradley, G.; Muchenje, V. Effects of transport distance, lairage time and stunning efficiency on cortisol, glucose, HSPA1A and how they relate with meat quality in cattle. *Meat Sci.* **2016**, *117*, 89–96. [[CrossRef](#)] [[PubMed](#)]
152. Leticia, M.; Paola, D.; Jordi, O.; Julio, O.; Sancho, B. Effects of sage distillation by-product (*Salvia lavandulifolia* Vahl.) dietary supplementation in light lambs fed on concentrates on meat shelf life and fatty acid composition. *Meat Sci.* **2017**, *134*, 44–53. [[CrossRef](#)]
153. Balamatsia, C.C.; Paleologos, E.K.; Kontominas, M.G.; Savvaidis, I.N. Correlation between microbial flora, sensory changes and biogenic amines formation in fresh chicken meat stored aerobically or under modified atmosphere packaging at 4 degrees C: Possible role of biogenic amines as spoilage indicators. *Antonie Leeuwenhoek* **2006**, *89*, 9–17. [[CrossRef](#)] [[PubMed](#)]
154. Capita, R.; Álvarez-González, T.; Alonso-Calleja, C. Effect of several packaging conditions on the microbiological, physicochemical and sensory properties of ostrich steaks during refrigerated storage. *Food Microbiol.* **2017**, *72*, 146–156. [[CrossRef](#)]
155. Chouliara, E.; Karatapanis, A.; Savvaidis, I.N.N.; Kontominas, M.G.G. Combined effect of oregano essential oil and modified atmosphere packaging on shelf-life extension of fresh chicken breast meat, stored at 4 degrees C. *Food Microbiol.* **2007**, *24*, 607–617. [[CrossRef](#)]
156. Herbert, U.; Rossaint, S.; Khanna, M.A.M.A.; Kreyenschmidt, J. Comparison of argon-based and nitrogen-based modified atmosphere packaging on bacterial growth and product quality of chicken breast fillets. *Poult. Sci.* **2013**, *92*, 1348–1356. [[CrossRef](#)]
157. Jääskeläinen, E.; Johansson, P.; Kostiaainen, O.; Nieminen, T.T.; Schmidt, G.; Somervuo, P.; Mohsina, M.; Vanninen, P.; Auvinen, P.; Björkroth, J. Significance of heme-based respiration in meat spoilage caused by *Leuconostoc gasicomitatum*. *Appl. Environ. Microbiol.* **2013**, *79*, 1078–1085. [[CrossRef](#)]
158. Jääskeläinen, E.; Hultman, J.; Parshintsev, J.; Riekkola, M.-L.; Björkroth, J.; Jaaskelainen, E.; Hultman, J.; Parshintsev, J.; Riekkola, M.-L.; Björkroth, J.; et al. Development of spoilage bacterial community and volatile compounds in chilled beef under vacuum or high oxygen atmospheres. *Int. J. Food Microbiol.* **2016**, *223*, 25–32. [[CrossRef](#)]
159. Kapetanakou, A.E.; Agathaggelou, E.I.; Skandamis, P.N. Storage of pork meat under modified atmospheres containing vapors from commercial alcoholic beverages. *Int. J. Food Microbiol.* **2014**, *178*, 65–75. [[CrossRef](#)]
160. Liu, F.; Yang, R.Q.; Li, Y.F. Correlations between growth parameters of spoilage micro-organisms and shelf-life of pork stored under air and modified atmosphere at -2, 4 and 10 °C. *Food Microbiol.* **2006**, *23*, 578–583. [[CrossRef](#)] [[PubMed](#)]

161. Martinez, L.; Djenane, D.; Cilla, I.; Beltran, J.A.; Roncalés, P. Effect of varying oxygen concentrations on the shelf-life of fresh pork sausages packaged in modified atmosphere. *Food Chem.* **2006**, *94*, 219–225. [[CrossRef](#)]
162. Miks-Krajnik, M.; Yoon, Y.J.; Ukuku, D.O.; Yuk, H.G. Identification and quantification of volatile chemical spoilage indexes associated with bacterial growth dynamics in aerobically stored chicken. *J. Food Sci.* **2016**, *81*, M2006–M2014. [[CrossRef](#)] [[PubMed](#)]
163. Nieminen, T.T.; Dalgaard, P.; Björkroth, J. Volatile organic compounds and *Photobacterium phosphoreum* associated with spoilage of modified-atmosphere-packaged raw pork. *Int. J. Food Microbiol.* **2016**, *218*, 86–95. [[CrossRef](#)]
164. Rahkila, R.; Nieminen, T.T.; Johansson, P.; Säde, E.; Björkroth, J. Characterization and evaluation of the spoilage potential of *Lactococcus piscium* isolates from modified atmosphere packaged meat. *Int. J. Food Microbiol.* **2012**, *156*, 50–59. [[CrossRef](#)]
165. Rossaint, S.; Klausmann, S.; Kreyenschmidt, J. Effect of high-oxygen and oxygen-free modified atmosphere packaging on the spoilage process of poultry breast fillets. *Poult. Sci.* **2015**, *94*, 96–103. [[CrossRef](#)]
166. Pin, C.; García de Fernando, G.D.; Ordóñez, J.A. Effect of modified atmosphere composition on the metabolism of glucose by *Brochothrix thermosphacta*. *Appl. Environ. Microbiol.* **2002**, *68*, 4441–4447. [[CrossRef](#)]
167. Senter, S.D.; Arnold, J.W.; Chew, V. APC values and volatile compounds formed in commercially processed, raw chicken parts during storage at 4 and 13 °C and under simulated temperature abuse conditions. *J. Sci. Food Agric.* **2000**, *80*, 1559–1564. [[CrossRef](#)]
168. Zhang, Q.Q.; Han, Y.Q.; Cao, J.X.; Xu, X.-L.; Zhou, G.-H.; Zhang, W.Y. The spoilage of air-packaged broiler meat during storage at normal and fluctuating storage temperatures. *Poult. Sci.* **2012**, *91*, 208–214. [[CrossRef](#)]
169. Grech, V.; Calleja, N. WASP (Write a Scientific Paper): Parametric vs. non-parametric tests. *Early Hum. Dev.* **2018**, *123*, 48–49. [[CrossRef](#)]
170. Gill, J. The insignificance of null hypothesis significance testing. *Polit. Res. Quart.* **1999**, *52*, 647–674. [[CrossRef](#)]
171. Gliner, J.A.; Morgan, G.A.; Leech, N.L.; Harmon, R.J. Problems with null hypothesis significance testing. *J. Experim. Educ.* **2002**, *71*, 83–92. [[CrossRef](#)]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).