

Article

Using Bronson Equation to Accurately Predict the Dog Brain Weight Based on Body Weight Parameter

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Abstract: The study used 69 brains ($n = 69$) from adult dog cadavers, divided by their skull type into three groups, brachi (B), dolicho (D) and mesaticephalic (M) ($n = 23$ each), and aimed: (1) to determine whether the Bronson equation may be applied, without reservation, to estimate brain weight (BW) in brachy (B), dolicho (D), and mesaticephalic (M) dog breeds; and (2) to evaluate which breeds are more closely related to each other in an evolutionary scenario. All subjects were identified by sex, age, breed, and body weight (bw). An oscillating saw was used for a circumferential craniotomy to open the skulls; the brains were removed and weighed using a digital scale. For statistical analysis, p -values < 0.05 were considered significant. The work demonstrated a strong relationship between the observed and predicted BW by using the Bronson equation. It was possible to hypothesize that groups B and D present a greater encephalization level than M breeds, that B and D dog breeds are more closely related to each other than to M, and from the three groups, the D individuals presented the highest brain mass mean.

Keywords: dog; brain; brachycephalic; dolichocephalic; mesaticephalic; body weight

1. Introduction

Body weight influences almost every aspect of organism biology, including brain mass [1]. Variations in brain mass occur in proportion to the increase of body mass of the animal (allometric scaling); by increasing body size and weight, an increase of peripheral receptor conduction will promote a rise in the corresponding representation fields in the cerebral cortex (both motor and somatosensory cortices) [2–4]. Several studies demonstrated a strong positive correlation between brain weight (BW) and body weight (bw) in general and specifically in dogs [5–12]. The brain is at adult size when body growth is only 40 percent complete [13–17]. Bronson developed a mathematical equation to explain the encephalic allometry, demonstrating that the mean BW of dogs is related to mean bw and therefore BW could be calculated without previous models' inconsistencies. The evolution of diversity in brain size [3,18–24]. According to Bronson's dog equation, average BW (y) can be expressed by the following allometric function: $Y = 0.39X^{0.27}$, where 0.39 is the integration constant, X is the bw mean, and 0.27 is the allometric exponent or the potency associated with the intra-species ratio of BW and bw means [3,18,25–27]. This function may serve as a baseline for measuring increases or decreases in encephalization during species evolution [3]. Encephalization level (El) as a measure of relative brain size—defined as the ratio between the actual and the predicted brain mass for an individual El—is also used as a rough estimate of the intelligence of the animal and is useful for comparison within

species or between fairly closely related species [28]. Since the allometric coefficient often exhibits intraspecific variation, which can be heritable, some theories assume that greater BW differences are a result of selection for differences in bw, registered among more distantly than closely related species [5,15]. The present study was developed in dog specimens and aimed: (1) to determine whether the Bronson equation may be applied, without reservation, to estimate BW in all the brachy (B), dolicho (D), and mesaticephalic (M) dog breeds; and (2) to evaluate which breeds are more closely related to each other in an evolutionary scenario.

2. Materials and Methods

The study used 69 brains ($n = 69$) from adult dog cadavers, divided by their skull type into three groups, B, D, and M ($n = 23$ each), obtained at the teaching hospital of Faculty of Veterinary Medicine, University of Lisbon (FMV-ULisboa), Portugal, and Anjos of Assis Veterinary Medicine Centre (CMVAA), Barreiro, Portugal, after verification of death by a veterinary surgeon, authorization by the dog owners with signed consent forms, and approval by the ethical committee. All subjects were identified by sex, age, breed, and bw. An oscillating saw was used for a circumferential craniotomy to open the skulls; the brains were removed and weighed using a digital scale. The measurements were recorded onto recording forms, and the data were put into an SPSS® database (IBM, Armonk, North Castle, NY, USA). The Kolmogorov-Smirnov test (KS) was used to test for normality and ANOVA was used to test if BW differed among B, D, and M breeds. To test for pair-wise differences between groups, we used a post hoc Bonferroni correction. A p -value < 0.05 was considered statistically significant.

3. Results

Summary statistics for all considered variables (age, bw, and BW) are listed in Table 1. From all the evaluated parameters, only the BW of the brachycephalic group did not show data normality. The Bronson equation was used to calculate the expected BW from each specimen. Table 2 presents the one-way ANOVA results that tested if BW differed among B, D, and M breeds, and the post hoc Bonferroni correction to test for significant pair-wise differences between groups. Differences were registered for the bw and BW parameters. Statistically significant differences were registered for bw between the pair groups B-D ($p < 0.00$) and D-M ($p < 0.00$), and for BW only between the pair groups D-M ($p < 0.00$).

Table 1. Descriptive statistics with mean, standard deviation, minimum and maximum values, sigma and t , regarding the parameters of age, living weight, brain weight, and breeds in brachy (B), dolicho (D) and mesaticephalic (M) dogs.

Group	Parameter	n	$\bar{x} \pm SD$	95%CI		σ for $p > 0.05$	t	
				Min (mm)	Max (mm)			
B	age	23	$8.00 \pm 1.65^{\ddagger}$	4.00	10.00	$\sigma > 0.10$	20.13	
	bw	23	$15.79 \pm 6.46^{\S}$	4.70	26.10	$\sigma > 0.10$	11.71	
	BW	23	$84.91 \pm 31.29^*$	26.62	147.27	$\sigma = 0.08$	13.02	
	Breed		8	French Bulldog				
			7	Boxer				
			4	Pekingese				
			2	Pug Carlin				
	2	Shitzu						
D	age	23	$8.70 \pm 2.40^{\ddagger}$	5.00	14.00	$\sigma > 0.10$	17.46	
	bw	23	$23.63 \pm 2.59^{\S}$	18.70	29.39	$\sigma > 0.10$	37.79	
	BW	23	$92.50 \pm 8.60^*$	71.60	102.10	$\sigma = 0.00$	50.60	
	Breed		11	Doberman Pinscher				
			5	Rough Collie				
			4	Whippet				
			3	Miniature Bull Terrier				

Table 1. Cont.

Group	Parameter	n	$\bar{x} \pm SD$	95%CI		σ for $p > 0.05$	t	
				Min (mm)	Max (mm)			
M	age	23	9.50 \pm 2.50 [‡]	5.00	15.00	$\sigma = 0.02$	17.70	
	bw	23	13.80 \pm 7.20 [§]	7.30	31.50	$\sigma < 0.01$	8.74	
	BW	23	69.90 \pm 28.70 *	35.70	143.0	$\sigma = 0.04$	11.68	
	Breed		7	Beagle				
			6	Golden Retriever				
			5	Yorkshire Terrier				
			3	Border Collie				
	2	Dalmatian						

Body weight (bw); brain weight (BW), minimum (min); maximum (max); confidence interval (CI). Measurements are in millimetres. [‡] Years, [§] kilograms, * grams.

Table 2. One-way ANOVA and post hoc Bonferroni corrected tests for differences in age, living weight, brain weight, maximum width, length and height between the brachy (B), dolicho (D) and mesaticephalic (M) dogs. The comparison between groups was made with an $n = 46$ (23 specimens within each group). Also, the comparison between observed and expected brain weight values obtained by using the Bronson equation in brachy (B), dolicho (D) and mesaticephalic (M) dogs. The correlation coefficient (Pearson’s product moment correlation) between both values in the three considered groups. The difference was significant at p -values < 0.05 .

Sample Characteristics												
Parameter	Type of Test	Differences Between Groups		n	DM (I–J)	EP	SS	MS	F	Sig.	CI 95%	
											Min	Max
Age	ANOVA	-	-	69	-	-	28.17	14.08	2.77	0.06	-	-
	Bonferroni	B	D	46	-0.78	1.17	-	-	-	1.00	-2.41	0.85
		B	M	46	-1.56	2.35	-	-	-	0.09	-3.19	0.06
bw	ANOVA	-	-	69	-	-	3303	447.7	12.27	0.00	-	-
	Bonferroni	B	D	46	-6.62 *	3.72	-	-	-	0.00	-11.00	2.25
		B	M	46	1.73	0.97	-	-	-	1.00	-2.64	6.10
BW	ANOVA	-	-	69	-	-	4739	3026	4.83	0.01	-	-
	Bonferroni	B	D	46	-7.62	1.03	-	-	-	1.00	-25.75	10.50
		B	M	46	14.92	2.02	-	-	-	>0.05	-3.20	33.06
Brain Weight	ANOVA	-	-	69	-	-	4739	3026	4.83	0.01	-	-
	Bonferroni	D	M	46	22.55 *	3.05	-	-	-	0.00	4.41	40.68
Group	n	Observed BW (g)			Expected BW (g)			p	r	χ^2 for $\chi^2_c > 5.99$		
		\bar{x}	min	max	\bar{x}	min	max					
B	23	84.91	53.60	116.20	82.20	71.30	90.10	0.68	0.99	2.47		
D	23	92.50	83.90	101.10	91.30	88.50	94.00	0.35	0.81	1.28		
M	23	69.90	41.20	98.60	79.20	64.60	88.80	0.17	0.96	5.85		

Body weight (bw); brain weight (BW); Mean Difference (I–J); confidence interval (CI); minimum (min); maximum (max); Pearson correlation coefficient (r); Chi-square (χ^2); Chi-square critical (χ^2_c); * statistically significant.

4. Discussion

According to the results, BW mean differed little, with no statistical significant differences, between the three groups considered; however, from all the groups, the M group presented a greater mean age (9.5 years), followed by group D (8.7 years), and then B (8.0 years). This is a very important parameter, since aging is associated with decreased brain volume (about 0.29% per year) and weight

(about 5%–10% over the lifetime), due to atrophy of the gray and white matters, enhanced by multiple factors [17,29–32]. Aged dog brains show increased cortical atrophy, ventricular dilation, decreased total brain volume, and decreased frontal lobe volume, similar to humans [29,32]. In previous studies, age-related reduction in gray matter volume was observed bilaterally in the frontal gyrus, orbitalis gyrus, ectosylvius gyrus, olfactory bulb, superior olivaris nucleus in brainstem, and unilaterally in the posreal gyrus, sylvium gyrus, suprasylvian gyrus, cerebellum, and brainstem nuclei. Variations related to white matter loss were largely bilateral and included the internal capsula, tracts of anterior cingulate, and the alveus of the hippocampus [17,29,32]. Additionally, high body mass index, biochemical changes (in particular, those related to dopamine loss), and the loss of neurons and myelinated axons in different brain parenchyma regions decrease the brain mass with age promoting an enlargement of the ventricular system and the grooves on the brain surface [17,30–34]. In accordance with Schmidt et al. [35], the Bronson equation may serve as a baseline for measuring the EI of species during their evolution. According to Kruska [5] and Pagel and Harvey [15], greater BW differences are registered among distantly related species as a result of selection for differences in bw. The study results showed that the biggest differences were registered between the pair-wise groups M-B and M-D; thus we can conclude that B and D dog breeds are more closely related to each other than to M. This is supported by the fact that the B and D breeds were the result of natural and artificial selection over the M breeds, which are most closely related to the gray wolf—*Canis lupus*—and from which they were domesticated [36]. This selection acted directly on the brain itself, and also depended on the selective forces involved, promoting the decreasing size of one part of the brain in order to increase the size of another [35,37]. This is the result of simultaneous isocortical neurogenesis onset and terminal neurogenesis that varies substantially between the rostral and caudal poles in some species, allowing for disproportionate expansion of some encephalic cortical regions relative to others [38–43]. With the Bronson equation, it was possible to verify that D and B specimens presented the best ratio between the registered and the predicted BW, and according the study results, we found that D has the highest mean brain mass, heavier than the B and the M specimens.

5. Conclusions

In conclusion, the work demonstrated a strong relationship between the observed and predicted BW by using the Bronson equation. It was possible to hypothesize that groups B and D present a greater EI than M breeds, and from the three groups, the D individuals presented the highest brain mass mean.

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References

1. Lindstedt, S.L.; Schaeffer, P.J. Use of allometry in predicting anatomical and physiological parameters of mammals. *Lab. Anim.* **2002**, *36*, 1–19. [[CrossRef](#)] [[PubMed](#)]
2. Jerison, H.J. *Evolution of the Brain and Intelligence*; Academic: New York, NY, USA; London, UK, 1973; pp. 406–435.
3. Bronson, R.T. Brain weight-body weight scaling in breeds of dogs and cats. *Brain Behav. Evol.* **1979**, *16*, 227–236. [[CrossRef](#)] [[PubMed](#)]
4. Schoenemann, P.T. Brain size scaling and body composition in mammals. *Brain Behav. Evol.* **2004**, *63*, 47–60. [[CrossRef](#)] [[PubMed](#)]
5. Kruska, D. Effects of domestication on brain structure and behavior in mammals. *Hum. Evol.* **1988**, *3*, 473–485. [[CrossRef](#)]
6. Radinsky, L. Evolution of brain size in carnivores and ungulates. *Am. Nat.* **1978**, *112*, 815–831. [[CrossRef](#)]

7. Röhrs, M. Cephalisation bei Caniden. *Z. Zool. Syst. Evolutionsforsch.* **1986**, *24*, 300–307. [[CrossRef](#)]
8. Carrera, I.; Dennis, R.; Mellor, D.J.; Penderis, J.; Sullivan, M. Use of magnetic resonance imaging for morphometric analysis of the caudal cranial fossa in Cavalier King Charles Spaniels. *Am. J. Vet. Res.* **2009**, *70*, 340–345. [[CrossRef](#)] [[PubMed](#)]
9. Carreira, L.M. Anatomotopography Cranioencephalic Relationships for Intracranial Neurosurgery Procedures in Dog. The Ultrasound as Neuronavigation Technique for Brain Surgery in Real Time. Ph.D. Thesis, Technical University of Lisbon (FMV/UTL), Lisbon, Portugal, 2011.
10. Carreira, L.M.; Ferreira, A. Reference values for dog sagittal and transverse cephalic indices in different skull types and their importance. *J. Anim. Vet. Adv.* **2015**, *14*, 91–94.
11. Hart, B.L.; Hart, L.A.; McCoy, M.; Sarath, C.R. Cognitive behaviour in Asian elephants: Use and modification of branches for fly switching. *Anim. Behav.* **2011**, *62*, 839–847. [[CrossRef](#)]
12. Seid, M.A.; Castillo, A.; Wcislo, W.T. The allometry of brain miniaturization in ants. *Brain Behav. Evol.* **2011**, *77*, 5–13. [[CrossRef](#)] [[PubMed](#)]
13. Armstrong, E. Relative brain size and metabolism in mammals. *Science* **1983**, *4603*, 1302–1304. [[CrossRef](#)]
14. Martin, R.D.; Harvey, P. Brain size allometry: Ontogeny and phylogeny. In *Size and Scaling in Primate Biology*; Jungers, W., Ed.; Plenum Press: New York, NY, USA, 1985.
15. Pagel, M.D.; Harvey, P.H. Taxonomic differences in the scaling of brain on body weight among mammals. *Science* **1989**, *30*, 1589–1593. [[CrossRef](#)]
16. Wikswo, J.P.; Curtis, E.L.; Eagleton, Z.E.; Evans, B.C.; Kole, A.; Hofmeister, L.H.; Matloff, W.J. Scaling and systems biology for integrating multiple organs-on-a-chip. *Lab Chip* **2013**, *13*, 3496–3511. [[CrossRef](#)] [[PubMed](#)]
17. Conn, P.M. *Handbook of Models for Human Aging*; Academic Press: Cambridge, MA, USA, 2006.
18. Gould, S.J. *Ontogeny and Phylogeny*; Harvard University Press: Estados Unidos, MA, USA, 1977.
19. Gould, S.J. An allometric interpretation of species-area curves: The meaning of the coefficient. *Am. Nat.* **1979**, *114*, 335–343. [[CrossRef](#)]
20. Jerison, H.J. Allometry, brain size, cortical surface, and convolutedness. In *Primate Brain Evolution*; Armstrong, E., Falk, D., Eds.; Plenum Press: New York, NY, USA, 1982.
21. Jerison, H.J. The evolutionary biology of intelligence: Afterthoughts. In *Intelligence and Evolutionary Biology*; Jerison, H., Jerison, I., Eds.; Springer: Berlin, Germany, 1988.
22. Jerison, H.J. The theory of Encephalization. *Ann. N. Y. Acad. Sci.* **1977**, *299*, 146–160. [[CrossRef](#)] [[PubMed](#)]
23. Gayon, J. History of the concept of allometry. *Integr. Comp. Biol. (ICB)* **2000**, *40*, 748–758. [[CrossRef](#)]
24. Williams, M.F. Primate Encephalization and intelligence. *Med. Hypotheses* **2002**, *58*, 284–290. [[CrossRef](#)] [[PubMed](#)]
25. Tan, U.; Caliskan, S. Allometry and asymmetry in the dog brain: The right hemisphere is heavier regardless of paw preference. *Int. J. Neurosci.* **1987**, *35*, 189–194. [[CrossRef](#)] [[PubMed](#)]
26. Rushton, J.P.; Ankney, C.A. Whole brain size and general mental ability: A review. *Int. J. Neurosci.* **2009**, *119*, 692–732. [[CrossRef](#)] [[PubMed](#)]
27. Bayat, P.D.; Ghanbari, A.; Sohoul, P.; Amiri, S.; Sari-aslani, P. Correlation of skull size and brain volume, with age, weight, height and body mass index of Arak Medical Sciences students. *Int. J. Morphol.* **2012**, *30*, 157–161. [[CrossRef](#)]
28. Marino, L.; Sol, D.; Toren, K.; Lefebvre, L. Does diving limit brain size in cetaceans? *Mar. Mammal Sci.* **2006**, *22*, 413–425. [[CrossRef](#)]
29. Su, M.Y.; Head, E.; Brooks, W.M.; Wang, Z.; Muggenburg, B.A.; Adam, G.E. Magnetic resonance imaging of anatomic and vascular characteristics in a canine model of human aging. *Neurobiol. Aging* **1988**, *19*, 479–485. [[CrossRef](#)]
30. Anderton, B.H. Ageing of the brain. *Mech. Ageing Dev.* **2002**, *123*, 811–817. [[CrossRef](#)]
31. Enzinger, C.; Fazekas, F.; Matthews, P.M.; Ropele, S.; Schmidt, H.; Smith, S.; Schmidt, R. Risk factors for progression of brain atrophy in aging: Six-year follow-up of normal subjects. *Neurology* **2005**, *64*, 1704–1711. [[CrossRef](#)] [[PubMed](#)]
32. Tapp, P.D.; Siwak, C.T.; Gao, F.Q.; Chiou, J.Y.; Black, S.E.; Head, E.; Muggenburg, B.A.; Cotman, C.W.; Milgram, N.W.; Su, M.Y. Frontal lobe volume, function, and beta-amyloid pathology in a canine model of aging. *J. Neurosci.* **2004**, *24*, 8205–8213. [[CrossRef](#)] [[PubMed](#)]

33. Conde, J.R.; Streit, W.J. Microglia in the aging brain. *J. Neuropathol. Exp. Neurol.* **2006**, *65*, 199–203. [[CrossRef](#)] [[PubMed](#)]
34. Raz, N.; Rodrigue, K.M.; Head, D.; Kennedy, K.M.; Acker, J.D. Differential aging of the medial temporal lobe: A study of a five-year change. *Neurology* **2004**, *62*, 433–438. [[CrossRef](#)] [[PubMed](#)]
35. Schmidt, M.J.; Amort, K.H.; Failing, K.; Klingler, M.; Kramer, M.; Ondreka, N. Comparison of the endocranial- and brain volumes in brachycephalic dogs, mesaticephalic dogs and Cavalier King Charles spaniels in relation to their body weight. *Acta Vet. Scand.* **2014**, *56*, 30. [[CrossRef](#)] [[PubMed](#)]
36. Hemmer, H. *Domestication: The Decline of Environmental Appreciation*, 2nd edition; Cambridge University Press: Cambridge, UK, 1990.
37. Healy, S.D.; Harvey, P.H. Comparative studies of the brain and its components. *Neth. J. Zool.* **1990**, *40*, 203–214. [[CrossRef](#)]
38. Rakic, P. Neurons in rhesus monkey visual cortex: Systematic relation between time of origin and eventual disposition. *Science* **2002**, *183*, 425–427. [[CrossRef](#)]
39. Bush, E.C.; Allman, J.M. The scaling of frontal cortex in primates and carnivores. *Proc. Natl. Acad. Sci. USA* **2004**, *101*, 3962–3966. [[CrossRef](#)] [[PubMed](#)]
40. Hutsler, J.J.; Lee, D.G.; Porter, K.K. Comparative analysis of cortical layering and supragranular layer enlargement in rodent carnivore and primate species. *Brain Res.* **2005**, *1052*, 71–81. [[CrossRef](#)] [[PubMed](#)]
41. Carreira, L.M.; Ferreira, A.; Burilo, F.L. The dorsal sagittal venous sinus anatomical variations in brachycephalic, dolichocephalic, and mesocephalic dogs and their significance for brain surgery. *Anat. Rec. Adv. Integr. Anat. Evol. Biol. (Hoboken)* **2011**, *294*, 1920–1929. [[CrossRef](#)] [[PubMed](#)]
42. Carreira, L.M.; Ferreira, A. The importance of the longitudinal cerebral fissure anatomy variations in brachy-, dolicho- and mesaticephalic dogs and their importance to brain surgery. *Anat. Rec. Adv. Integr. Anat. Evol. Biol. (Hoboken)* **2015**, *298*, 1612–1621. [[CrossRef](#)] [[PubMed](#)]
43. Workman, A.D.; Charvet, C.J.; Clancy, B.; Darlington, R.B.; Finlay, B.L. Modeling transformations of neurodevelopmental sequences across mammalian species. *J. Neurosci.* **2013**, *33*, 7368–7383. [[CrossRef](#)] [[PubMed](#)]



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