

Draft Genome Sequence of Polychlorinated Biphenyl-Dechlorinating *Dehalococcoides mccartyi* Strain SG1, Which Carries a Circular Putative Plasmid

Shanquan Wang,^a Kern Rei Chng,^{a,b} Chen Wu,^a Andreas Wilm,^b Niranjana Nagarajan,^b Jianzhong He^a

Department of Civil and Environmental Engineering, National University of Singapore, Singapore^a; Computational and Systems Biology, Genome Institute of Singapore, Singapore^b

***Dehalococcoides mccartyi* strain SG1, isolated from digester sludge, dechlorinates polychlorinated biphenyls (PCBs) to lower congeners. Here we report the draft genome sequence of SG1, which carries a 22.65 kbp circular putative plasmid.**

Received 10 August 2014 Accepted 20 August 2014 Published 2 October 2014

Citation Wang S, Chng KR, Wu C, Wilm A, Nagarajan N, He J. 2014. Draft genome sequence of polychlorinated biphenyl-dechlorinating *Dehalococcoides mccartyi* strain SG1, which carries a circular putative plasmid. *Genome Announc.* 2(5):e00901-14. doi:10.1128/genomeA.00901-14.

Copyright © 2014 Wang et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](http://creativecommons.org/licenses/by/3.0/).

Address correspondence to Jianzhong He, jianzhong.he@nus.edu.sg.

Persistent and toxic polychlorinated biphenyls (PCBs) were massively produced as commercial mixtures for industrial uses, leading to the contamination of sediments of rivers, lakes, and harbors worldwide. The obligate organohalide-respiring *Dehalococcoides mccartyi* strains in both microcosms (1–4) and pure cultures (5–7) have been identified to be capable of dechlorinating PCBs in distinct patterns. Populations of this bacterial group have small circular chromosomes of around 1.5 Mbp, yet each carries a suite of 10 to 36 different *rdhA* genes encoding reductive dehalogenases (RDases) for catalyzing the chlorine removal (8). Most RDase genes are located in the high plasticity regions (HPRs) of chromosomes (7, 9, 10), suggesting that their horizontal acquisition is possibly through temperate bacteriophages or plasmids. To date, however, no plasmid has been found in *Dehalococcoides*.

Strain SG1 belonging to the *Dehalococcoides* Pinellas subgroup was isolated from a PCB-dechlorinating sediment-free culture originated from digester sludge of an industrial wastewater treatment plant in Singapore (2). The genome of strain SG1 was sequenced by using HiSeq 2000 from pair-end libraries with an average insert size of 300 bp and a read length of 76 bp. The reads were assembled using SOAPdenovo (11) and scaffolded was done with Opera (12). The coverage for the genome sequence assembly was 1,580×. GapCloser (11) was utilized for *in silico* closing of gaps between contigs, which generated 5 scaffolds. Finally, targeted PCR reactions and Sanger sequencing were used to confirm an independent circular piece of DNA, possibly a putative plasmid. Open reading frames (ORFs) were predicted using Prodigal (13). Functional annotations were assigned by screening predicted ORFs with entries in the KEGG database (14) using RapSearch (15).

The assembled draft genome of strain SG1 is 1,428,734 bp long, with a G+C content of 47.05%. It contains 1,486 protein-coding genes (including 28 RDase genes), which are similar to those of other sequenced *Dehalococcoides mccartyi* strains (7, 9, 10). The significant difference is that SG1 carries a 22.65 kbp circular putative plasmid. This putative plasmid is predicted to encode 26 proteins, including integrase, metallophosphoesterase, DNA pri-

mase, and a transcriptional regulator. Nine of the 26 protein coding sequences share 61% to 96% similarities with their homologues in the chromosome of *Dehalococcoides mccartyi* 195. The genome sequence of strain SG1 may provide new insights into the adaptation mechanisms of *Dehalococcoides* to the organohalide respiration of PCBs.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under accession no. [JPRE000000000](https://www.ncbi.nlm.nih.gov/nuccore/JPRE000000000). The version described in this paper is version JPRE01000000.

ACKNOWLEDGMENTS

This study was supported by the Science and Engineering Research Council, Agency for Science, Technology and Research under Project 102 101 0025, the Industrial Microbiology Genome Informatics platform, and by the National Research Foundation, Prime Minister's Office, Singapore, under its Competitive Research Programme (CRP Award NRF-CRP 5-2009-05).

REFERENCES

- Bedard DL, Ritalahti KM, Löffler FE. 2007. The *Dehalococcoides* population in sediment-free mixed cultures metabolically dechlorinates the commercial polychlorinated biphenyl mixture aroclor 1260. *Appl. Environ. Microbiol.* 73:2513–2521. <http://dx.doi.org/10.1128/AEM.02909-06>.
- Wang S, He J. 2013. Phylogenetically distinct bacteria involve extensive dechlorination of aroclor 1260 in sediment-free cultures. *PLoS One* 8:e59178. <http://dx.doi.org/10.1371/journal.pone.0059178>.
- Wang S, He J. 2013. Dechlorination of commercial PCBs and other multiple halogenated compounds by a sediment-free culture containing *Dehalococcoides* and *Dehalobacter*. *Environ. Sci. Technol.* 47:10526–10534. <http://dx.doi.org/10.1021/es4017624>.
- Zhen H, Du S, Rodenburg LA, Mainelis G, Fennell DE. 2014. Reductive dechlorination of 1,2,3,7,8-pentachlorodibenzo-p-dioxin and aroclor 1260, 1254 and 1242 by a mixed culture containing *Dehalococcoides mccartyi* strain 195. *Water Res.* 52:51–62. <http://dx.doi.org/10.1016/j.watres.2013.12.038>.
- Adrian L, Dudková V, Demnerová K, Bedard DL. 2009. “*Dehalococcoides*” sp. strain CBDB1 extensively dechlorinates the commercial polychlorinated biphenyl mixture aroclor 1260. *Appl. Environ. Microbiol.* 75:4516–4524. <http://dx.doi.org/10.1128/AEM.00102-09>.
- LaRoe SL, Fricker AD, Bedard DL. 2014. *Dehalococcoides mccartyi* strain

- JNA in pure culture extensively dechlorinates aroclor 1260 according to polychlorinated biphenyl (PCB) Dechlorination Process N. Environ. Sci. Technol. 48:9187–9196. <http://dx.doi.org/10.1021/es500872t>.
7. Wang S, Chng KR, Wilm A, Zhao S, Yang KL, Nagarajan N, He J. 2014. Genomic characterization of three unique *Dehalococcoides* that respire on persistent polychlorinated biphenyls. Proc. Natl. Acad. Sci. U. S. A. 111: 12103–12108. <http://dx.doi.org/10.1073/pnas.1404845111>.
 8. Löffler FE, Yan J, Ritalahti KM, Adrian L, Edwards EA, Konstantinidis KT, Muller JA, Fullerton H, Zinder SH, Spormann AM. 2013. *Dehalococcoides mccartyi* gen. nov., sp. nov., obligately organohalide-respiring anaerobic bacteria relevant to halogen cycling and bioremediation, belong to a novel bacterial class, *Dehalococcoidia* classis nov., order *Dehalococcales* ord. nov. and family *Dehalococcoidaceae* fam. nov., within the phylum *Chloroflexi*. Int. J. Syst. Evol. Microbiol. 63:625–635. <http://dx.doi.org/10.1099/ijs.0.034926-0>.
 9. Seshadri R, Adrian L, Fouts DE, Eisen JA, Phillippy AM, Methe BA, Ward NL, Nelson WC, Deboy RT, Khouri HM, Kolonay JF, Dodson RJ, Daugherty SC, Brinkac LM, Sullivan SA, Madupu R, Nelson KE, Kang KH, Impraim M, Tran K, Robinson JM, Forberger HA, Fraser CM, Zinder SH, Heidelberg JF. 2005. Genome sequence of the PCE-dechlorinating bacterium *Dehalococcoides ethenogenes*. Science 307: 105–108. <http://dx.doi.org/10.1126/science.1102226>.
 10. McMurdie PJ, Behrens SF, Müller JA, Göke J, Ritalahti KM, Wagner R, Goltsman E, Lapidus A, Holmes S, Löffler FE, Spormann AM. 2009. Localized plasticity in the streamlined genomes of vinyl chloride respiring *Dehalococcoides*. PLoS Genet. 5:e1000714. <http://dx.doi.org/10.1371/journal.pgen.1000714>.
 11. Li R, Zhu H, Ruan J, Qian W, Fang X, Shi Z, Li Y, Li S, Shan G, Kristiansen K, Li S, Yang H, Wang J, Wang J. 2010. De novo assembly of human genomes with massively parallel short read sequencing. Genome Res. 20:265–272. <http://dx.doi.org/10.1101/gr.097261.109>.
 12. Gao S, Sung WK, Nagarajan N. 2011. Opera: reconstructing optimal genomic scaffolds with high-throughput paired-end sequences. J. Comput. Biol. 18:1681–1691. <http://dx.doi.org/10.1089/cmb.2011.0170>.
 13. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. <http://dx.doi.org/10.1186/1471-2105-11-119>.
 14. Kanehisa M, Goto S, Sato Y, Furumichi M, Tanabe M. 2012. KEGG for integration and interpretation of large-scale molecular data sets. Nucleic Acids Res. 40:D109–D114. <http://dx.doi.org/10.1093/nar/gkr988>.
 15. Zhao Y, Tang H, Ye Y. 2012. RAPSearch2: a fast and memory-efficient protein similarity search tool for next-generation sequencing data. Bioinformatics 28:125–126. <http://dx.doi.org/10.1093/bioinformatics/btr595>.