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Data Article

Surface characterization data for tethered polyacrylic acid layers synthesized on polysulfone surfaces



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ABSTRACT

The data presented are supplementary to an article [Kim et al., 2019] on synthesis and surface characterization of tethered polyacrylic acid (PAA) layers on polysulfone (PSf) film/membrane surfaces via atmospheric pressure plasma-induced graft polymerization (APPIGP). Data on surface characterization of the synthesized tethered PAA layers includes: AFM topographic surface images and height distributions of surface features, dry layer thickness, chain rupture length distributions determined via AFM based force spectroscopy (AFM-FS), in addition to measurements of water contact angles. Fouling propensity data for ultrafiltration of alginate acid as a model foulant are also provided for native and PAA grafted PSf ultrafiltration (UF) membranes.

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Specifications table

Subject area	Material science
More specific subject area	Graft polymerization, tethered polymer layers, surface hydrophilicity, surface fouling resistance
Type of data	Table, image (FIB-SEM, AFM), figure
How data were acquired	XPS (Kratos Axis Ultra DLD) FIB-SEM (FEI Nova 600 NanoLab DualBeam TM -SEM/FIB) AFM (surface topography: Bruker Dimension Icon SPM with a NanoScope V controller; AFM force spectroscopy: Bruker MultiMode 8-HR SPM with a PicoForce Spectroscopy Control Module)
Data format	Raw and analyzed
Experimental factors	Tethered polyacrylic acid chains were graft polymerized onto polysulfone surfaces post atmospheric pressure plasma surface activation.
Experimental features	Surface characterization was performed for tethered polyacrylic acid layers synthesized onto polysulfone surfaces.
Data source location	University of California, Los Angeles (UCLA), Los Angeles, California, USA.
Data accessibility	Data are within this article.
Related research article	S. Kim, Y. Cohen, K.J. Moses, S. Sharma, M. Bilal, Polysulfone surface nano-structured with tethered polyacrylic acid, <i>Applied Surface Science</i> , 470 (2019) 411–422.

Value of the data

- The data provide quantitative information regarding the characteristics (surface topography, wettability, chain length distributions) of tethered polyacrylic acid layer under various solvents. Such surfaces have potential applications as fouling resistant surfaces including in membrane filtration and in biomedical applications.
- The provided detailed AFM-FS data should be useful to researchers who may use the technique to evaluate surface tethered polymer chain lengths under different solvent conditions, as well as evaluate various algorithms for exploring AFM-FS data.
- Membrane fouling data should be useful to researchers who may conduct membrane fouling tests under similar experimental conditions.

1. Data

Tethered polyacrylic acid layers were synthesized via free-radical graft polymerization at initial monomer concentration ($[M]_0$) of 1–20 vol%. The resulting surfaces were characterized with respect to elemental surface composition determined via XPS (Table 1) and surface topography evaluated via AFM analysis (Fig. 1). Contact angles for the different surfaces (Fig. 9, Fig. 10, Table 2, Table 3) were evaluated via the sessile drop method in air and via the captive bubble method in DI water and saline water (35 g/L NaCl)), and the polar and dispersive components of the surface free energies are provided in Table 2. Typical AFM-FS retraction force curves for the native PSf film and PAA layer grafted PSf film surfaces are shown in Fig. 2 and Fig. 4, respectively. The AFM-FS determined chain rupture length and rupture force distribution data for the tethered PAA chains are presented in Figs. 5 and 6, respectively, and the rupture force distribution for the native PSf film in water is shown in Fig. 3. Equilibrium thicknesses for

Table 1
Elemental surface compositions for the native PSf-Si and PAA-PSf-Si^a.

Surface	Carbon%	Oxygen%	Sulfur%
Native PSf	85.2	11.3	3.5
<i>Tethered PAA layers synthesized at $[M]_0$:</i>			
1 vol%	80.9	17.0	2.1
5 vol%	80.4	17.5	2.1
10 vol%	80.1	17.7	2.1
15 vol%	79.1	19.0	1.9
20 vol%	75.4	23.4	1.2

^a AA graft polymerization was carried out for a period of 2 h following PSf surface activation by He/O₂ plasma for a period of 15 s.

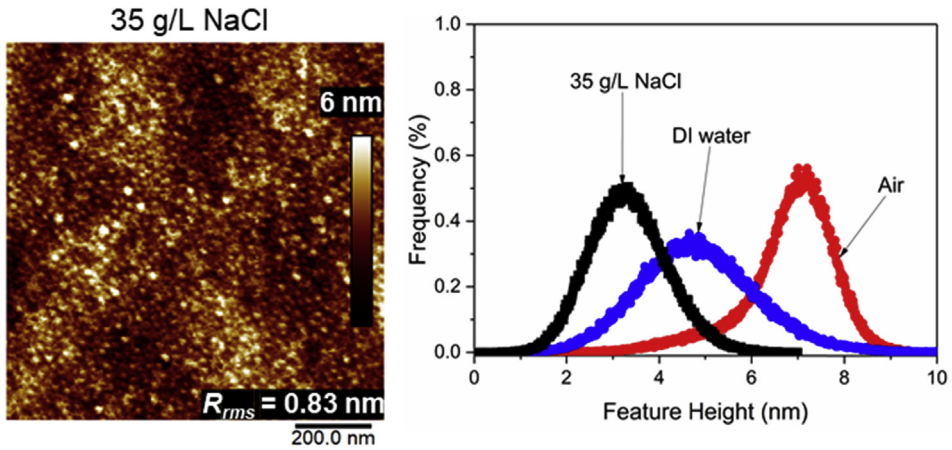


Fig. 1. AFM 2-D image obtained in 35 g/L NaCl solution (left) and the corresponding feature height distribution (FHD) (right) for the PAA-PSf-Si (the tethered PAA layer synthesized at $[M]_0 = 20$ vol% for 2 h following surface activation with He/O₂ APP) (plotted along with the FHD for the surface as determined in DI water and air). The feature heights are scaled to the lowest height determined by the AFM tip and R_{rms} is root-mean-square surface roughness.

Table 2

Sessile drop (SD) and captive bubble (CB) contact angles (CA) and surface free energy (SFE) data for the native PSf-Si and PAA-PSf-Si^a surfaces.

Surface	SD CA ^b (°)	CB CA ^c (°) (air)	CB CA ^c (°) (octane)	$\gamma_s^{p,d}$ (mJ m ⁻²)	$\gamma_s^{d,d}$ (mJ m ⁻²)	% Polar SFE component ^d
Native PSf	90.0	80.0	118.5	3.5	39.4	8.1
Tethered PAA layers synthesized at $[M]_0$:						
1 vol%	49.4	36.6	42.5	38.3	20.8	64.8
5 vol%	49.3	36.6	41.7	38.7	20.4	65.5
10 vol%	47.9	36.8	43.2	38.0	21.1	64.3
15 vol%	46.1	36.4	42.7	38.2	21.1	64.4
20 vol%	44.5	36.9	43.6	37.8	21.3	63.9

^a AA graft polymerization was carried out for a period of 2 h following PSf surface activation by He/O₂ plasma for a period of 15 s.

^b Sessile drop contact angles measured with 1 μ L DI water drops.

^c Captive bubble contact angles measured with 4 μ L air and octane bubbles.

^d Polar (γ_s^p) and dispersive (γ_s^d) components of the surface free energy.

Table 3

Captive bubble contact angles for PAA-PSf-Si (the tethered PAA layer was synthesized at $[M]_0 = 20$ vol% for 2 h post PSf activation with He/O₂ plasma) in DI water and NaCl solutions.

Solvent	Captive bubble contact angle (°)
DI water (pH 6)	38.4
DI water (pH 8)	29.7
35 g/L NaCl (pH 6)	29.3
35 g/L NaCl (pH 8)	29.1

the PAA layers in DI water and in saline water (35 g/L NaCl) are presented in Figs. 7 and 8, respectively, while dry layer thickness data (evaluated via FIB-SEM, Fig. 13) are shown in Fig. 10. Finally, permeate flux decline during filtration of saline alginate solution with PAA grafted PSf membrane and native PSf membrane at pH 6 and pH 8 are presented in Figs. 11 and 12, respectively.

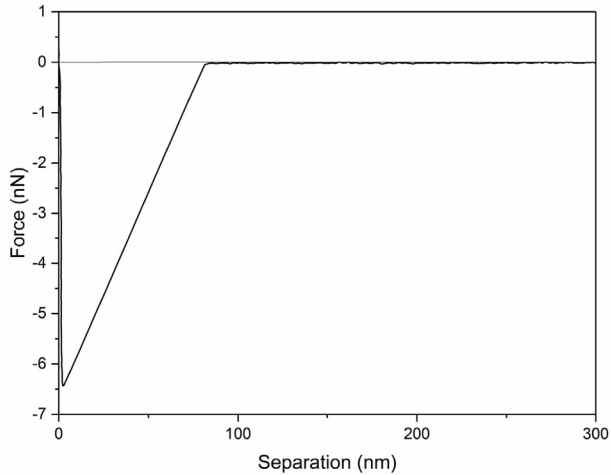


Fig. 2. A typical retraction force curve for the native PSf-Si surface in DI water.

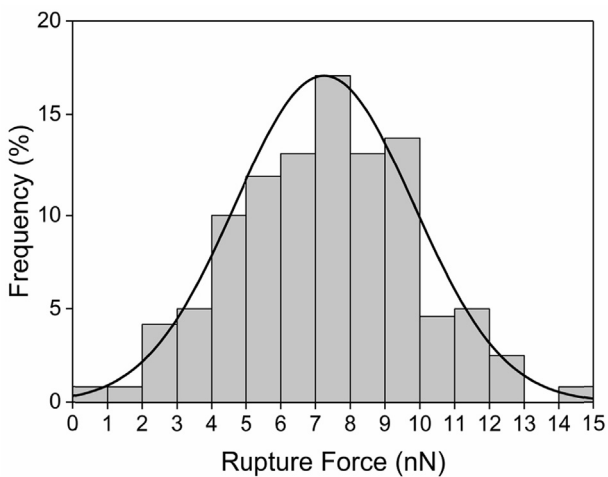


Fig. 3. AFM-FS determined rupture force distribution for the native PSf-Si surface in DI water.

2. Experimental design, materials, and methods

2.1. Materials

Helium (He), hydrogen (H₂) and oxygen (O₂) gases were used for plasma generation, and nitrogen (N₂) was utilized for drying and for degassing solvents. Poly (ethyleneimine) (PEI, M_w ~750,000) solution (50 wt% in water), PSf pellets (M_w ~35,000), chloroform and acrylic acid were all utilized in the synthesis of PAA tethered surfaces. Piranha solutions were prepared using sulfuric acid (H₂SO₄, 96%) and hydrogen peroxide (H₂O₂, 30% in water). Sodium chloride and deionized (DI) water (pH ~6) were used for preparation of the saline water (35 g/L NaCl). The model foulant [2] for fouling tests was alginic acid sodium salt, and the prepared alginic acid solution pH was adjusted with 50 wt% aqueous sodium hydroxide solution. Finally, membranes with tethered chains were prepared using PSf base membrane of 100 kDa molecular weight cutoff (MWCO).

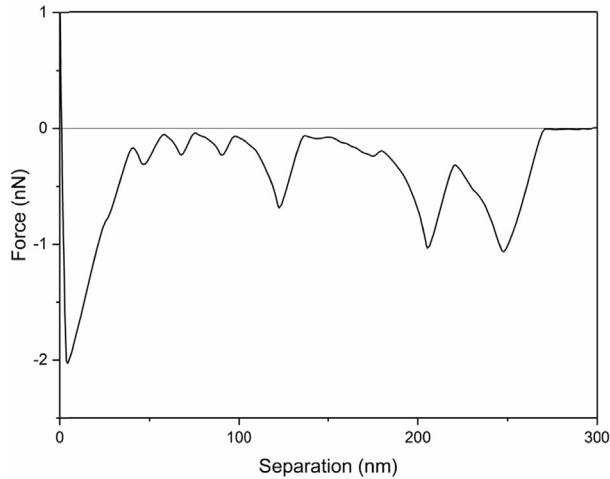


Fig. 4. A typical retraction force curve for PSf with the tethered PAA layer (PAA-PSf-Si) in DI water. The tethered PAA layer (synthesized onto PSf activated with He/O₂ plasma) was synthesized at $[M]_0 = 20$ vol% for 2 h.

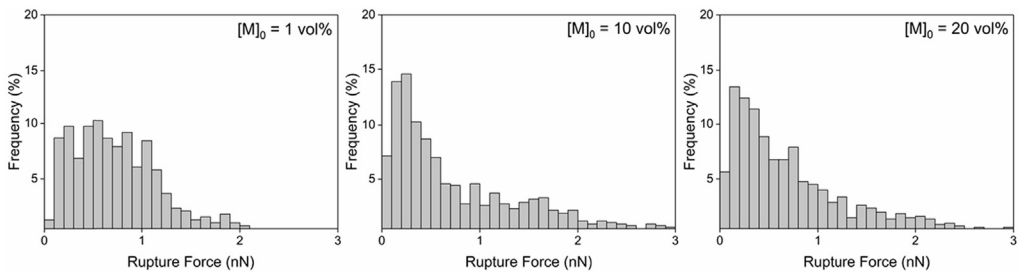


Fig. 5. AFM-FS determined rupture force distributions (evaluated under DI water) for PAA chains synthesized at $[M]_0$ of 1 vol%, 10 vol%, and 20 vol% for 2 h (Note: the PSf surfaces were activated with He/O₂ plasma; data include the surface adhesion peaks (i.e., the first rupture event in the retraction profiles, $L_R < 10$ nm)).

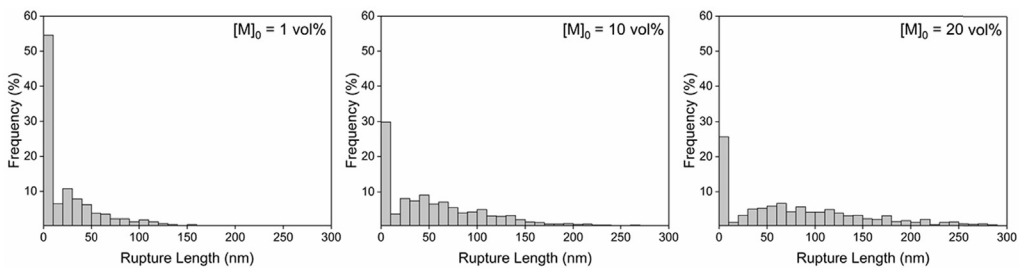


Fig. 6. AFM-FS determined rupture length distributions (evaluated under DI water) for PAA chains synthesized at $[M]_0$ of 1 vol%, 10 vol%, and 20 vol% for 2 h (Note: the PSf surfaces were activated with He/O₂ plasma; data include the surface adhesion peaks (i.e., the first rupture event in the retraction profiles, $L_R < 10$ nm)).

2.2. PSf film preparation

Smooth PSf substrate films were prepared on prime grade silicon wafers by spin-coating a PSf solution using a spin coater. A piranha solution (a 7:3 mixture of concentrated H₂SO₄ and 30% aqueous H₂O₂) was used to clean the silicon wafers at 90–100 °C for 10 minutes, followed by rinsing with DI

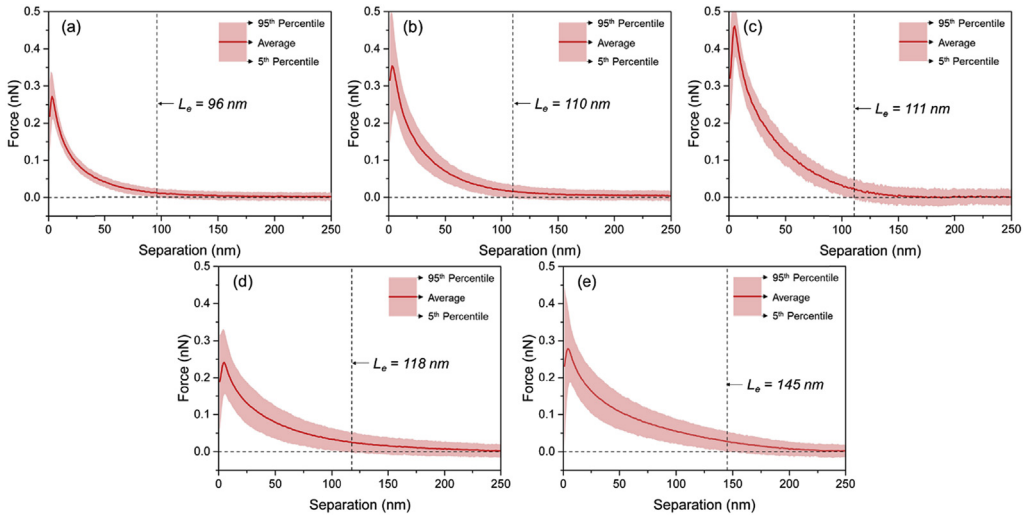


Fig. 7. Equilibrium thickness (L_e) in DI water estimated for PSf with the tethered PAA layers (PAA-PSf-Si) synthesized at $[M]_0$: (a) 1 vol%, (b) 5 vol%, (c) 10 vol%, (d) 15 vol%, and (e) 20 vol% for a reaction period of 2 h (Note: PSf surface was activated with He/O₂ plasma).

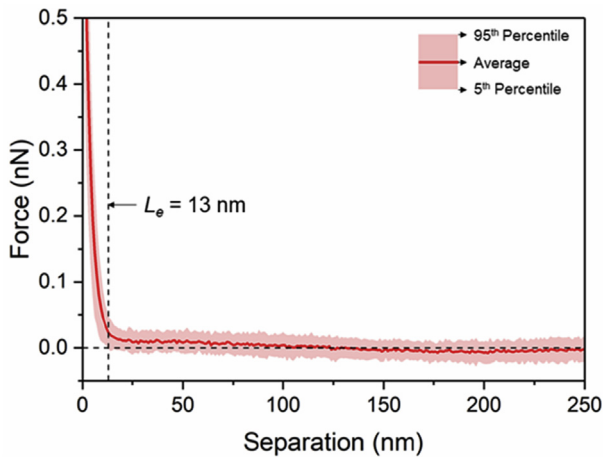


Fig. 8. Equilibrium thickness (L_e) in saline water (35 g/L NaCl) estimated for PSf with the tethered PAA layer (PAA-PSf-Si) synthesized at $[M]_0 = 20$ vol% for a reaction period of 2 h (Note: PSf surface was activated with He/O₂ plasma).

water. Silicon wafer samples (~ 1 cm \times 1 cm) were prepared and cleaned with isopropanol and then DI water, followed by blow drying with N₂. The wafer surface was first coated with PEI (using ~ 0.1 mL of a 0.3 wt% aqueous PEI solution at a spin rate of 2500 rpm for 30 seconds). Subsequently, PSf was coated onto the silicon wafer with PEI coating using ~ 0.1 mL of a 1 wt% PSf solution in chloroform that was spin-coated at 2500 rpm for 30 s. The produced PSf-Si substrates were then vacuum oven dried at ~ 75 °C prior to their subsequent use.

2.3. Plasma surface activation and graft polymerization

An impinging atmospheric pressure plasma (APP) jet [3] was used to activate the PSf substrates using He, He/H₂ or He/O₂ plasmas generated at the same He flow rate. All plasmas were generated at

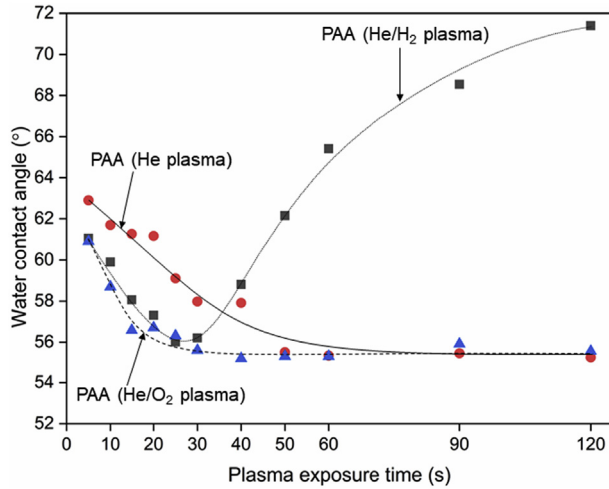


Fig. 9. Sessile drop water contact angles for PAA-PSf-Si surfaces (graft polymerized at $[M]_0 = 10$ vol% for 1 h) following PSf surface activation with He/H₂, He, and He/O₂ APP at various plasma exposure times (5–120 s). Water contact angles increased by ~7% after storage in air for ≥ 5 days for the tethered PAA layers regardless of the type of APP used for surface activation prior to graft polymerization.

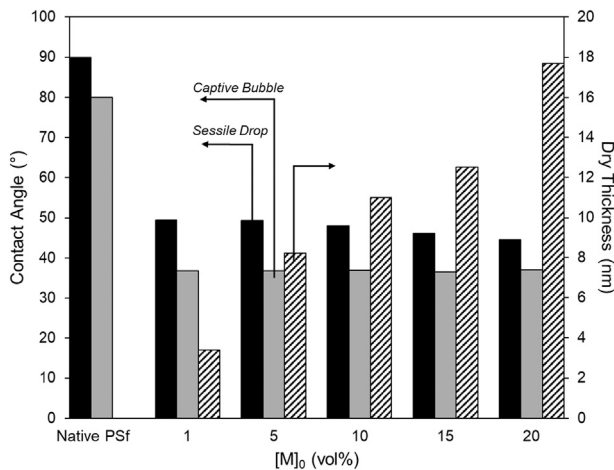


Fig. 10. Sessile drop and captive bubble contact angles for the native PSf-Si and for PSf with the tethered PAA layer (PAA-PSf-Si) synthesized at $[M]_0 = 1$ –20 vol% for 2 h post-PSf surface activation with He/O₂ plasma. Dry thickness is also plotted for the tethered PAA layers.

60 W. Graft polymerization of acrylic acid onto the activated surfaces was carried out at monomer concentration of 1–20 vol% over a period of up to 2 h at 70 °C. The DI water rinsed PAA-PSf-Si samples were vacuum dried at 40 °C prior to surface characterization. Tethered PAA layers were synthesized onto a plasma activated base PSf membrane having 100 kDa MW cutoff. Following graft polymerization, the PAA-PSf-Membrane samples were rinsed and stored in DI water prior to ultrafiltration fouling tests.

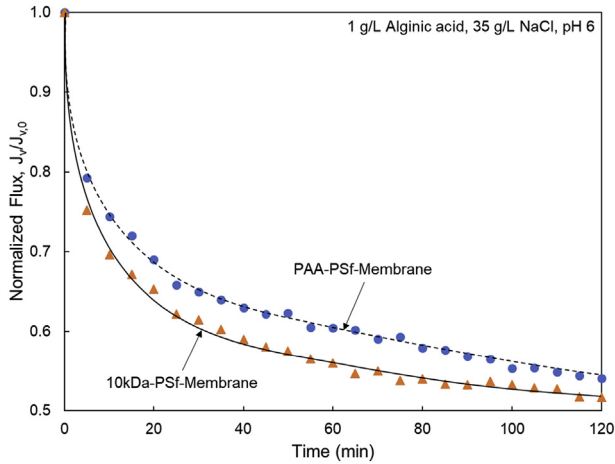


Fig. 11. Permeate flux decline during filtration of saline alginate acid solution at pH 6 with (a) PAA-PSF-Membrane (synthesized at $[M]_0 = 20$ vol% for 1 h post PSf surface activation with He/O₂ plasma) and (b) 10kDa-PSF-Membrane.

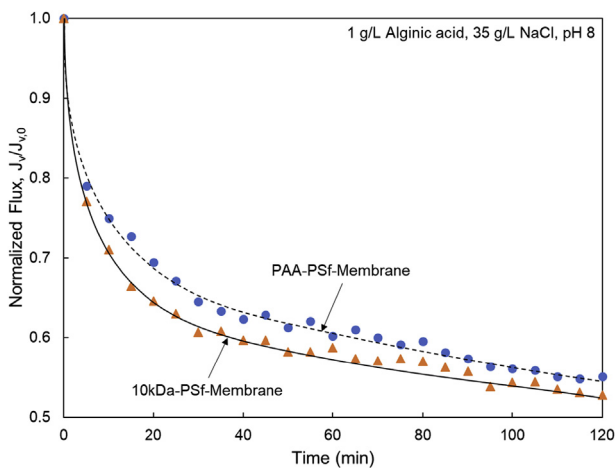


Fig. 12. Permeate flux decline during filtration of saline alginate acid solution at pH 8 with (a) PAA-PSF-Membrane (synthesized at $[M]_0 = 20$ vol% for 1 h post PSf surface activation with He/O₂ plasma) and (b) 10kDa-PSF-Membrane.

2.4. Surface characterization

Confirmation of the presence of tethered PAA chains on the PSf surface was obtained by X-ray photo electron spectroscopy (XPS). Survey spectra were obtained for both the native PSf-Si and PAA-PSf-Si surfaces at a pass energy of 160 eV. Contact angles for the different surfaces were measured by both the sessile drop (SD) and captive bubble (CB) methods. CB contact angle measurements were for sample immersed in water or saline water at 20–22 °C [4–6] allowing for ~30 min equilibration prior to taking measurements [7]. All reported contact angles are averages based on measurements at five different locations on each sample. The surface free energy (γ_s , expressed as the sum of dispersive (γ_s^d) and polar (γ_s^p) components, $\gamma_s = \gamma_s^d + \gamma_s^p$) was determined from the average air and *n*-octane CB contact angles (θ) following the method described in Refs. [8,9], which required the dispersive, polar, and total surface tension of liquids (i.e., water and *n*-octane) reported in Ref. [6].

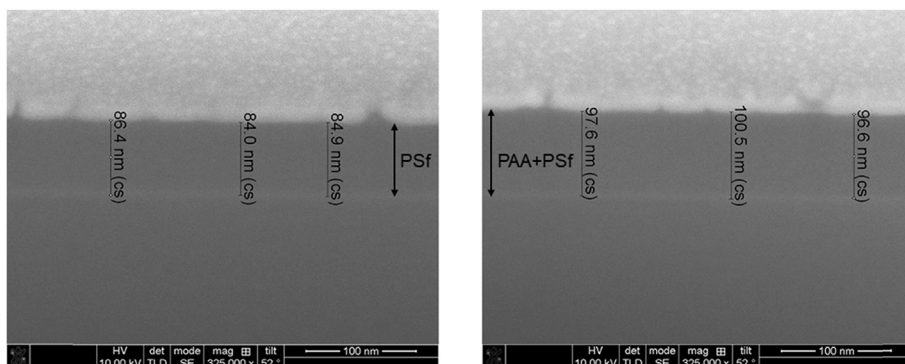


Fig. 13. Examples of FIB-SEM images obtained for PSf-Si (left) and PAA-PSf-Si (right) (Note: the tethered PAA layer synthesized at $[M]_0 = 20$ vol% for 2 h following surface activation with He/O₂ APP) used to estimate dry thickness of the PAA layer (by subtracting thickness for the native PSf layer (left) from PAA layer grafted PSf (right)). Note: prior to FIB, a gold layer was deposited onto the samples using a Denton Desk II Sputter Coater (Denton Vacuum, Inc., Moorestown, NJ), followed by coating a thin strip of platinum above the cross-sectioned regions.

Atomic force microscopy (AFM) (Bruker Dimension Icon Scanning Probe Microscope with a NanoScope V Controller, Bruker, Santa Barbara, CA) was utilized to obtain surface feature height data in PeakForce Tapping mode, under air, DI water and saline water (35 g/L NaCl), using ScanAsyst-Air and ScanAsyst-Fluid + probes (Bruker AFM Probes, Camarillo, CA). Cantilever deflection sensitivity and spring constant were determined following the approaches specified in Refs. [10,11], respectively. All AFM scans were obtained for areas of $1 \mu\text{m} \times 1 \mu\text{m}$ at 512×512 -pixel resolution and 0.5–0.8 Hz scan rates using a relatively small loading force (i.e., ~500 pN).

AFM force spectroscopy (AFM-FS; [12–14]) was accomplished using a Bruker MultiMode 8-HR Scanning Probe Microscope with a PicoForce Spectroscopy Control Module (Bruker, Santa Barbara, CA) under DI water and 35 g/L NaCl solution. A silicon nitride probe of a nominal tip radius of 20 nm (MLCT-D, Bruker AFM Probes, Camarillo, CA) was used, and force measurements were taken in contact mode at $1 \mu\text{m}$ ramp size and 500 nm/s tip velocity. The maximum surface applied force prior to tip retraction (i.e., trigger force) was 1 nN, and the approach and retraction force-distance profiles were obtained from 200 randomly selected locations for each sample. The rupture force and rupture length distributions as well as the equilibrium thickness [1] were determined from the retraction force profiles.

Sectioned substrate images were obtained via Focused Ion Beam (FIB) – Scanning Electron Microscopy (SEM) (Nova 600 NanoLab DualBeamTM-SEM/FIB; FEI company, Hillsboro, OR) [15]. The dry PAA layer thickness was calculated as the difference between the film thickness above the silicon wafer substrate before and after graft polymerization (an example of the thickness analysis is illustrated in Fig. 13).

2.5. Ultrafiltration fouling tests

Membrane fouling tests for the PSf (10 kDa MWCO) and PAA-PSf membranes were performed in a dead-end filtration mode (at 20 °C) using a small (50 mL) stirred cell system (MilliporeSigma, Burlington, MA) of 13.4 cm² membrane filtration area. Fouling challenge tests were conducted with a 1 g/L alginate solution at pH 8 and at pH 6 in high salinity water (35 g/L NaCl). All filtration tests were carried out at 22 L m⁻² h⁻¹ initial permeate flux which is well within range of permeate flux for UF seawater pretreatment [16].

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Transparency document

Transparency document associated with this article can be found in the online version at <https://doi.org/10.1016/j.dib.2019.103728>.

References

- [1] S. Kim, Y. Cohen, K.J. Moses, S. Sharma, M. Bilal, Polysulfone surface nano-structured with tethered polyacrylic acid, *Appl. Surf. Sci.* 470 (2019) 411–422.
- [2] A. Resosudarmo, Y. Ye, P. Le-Clech, V. Chen, Analysis of UF membrane fouling mechanisms caused by organic interactions in seawater, *Water Res.* 47 (2013) 911–921.
- [3] A. Schütze, J.Y. Jeong, S.E. Babayan, J. Park, G.S. Selwyn, R.F. Hicks, The atmospheric-pressure plasma jet: a review and comparison to other plasma sources, *IEEE Trans. Plasma Sci.* 26 (1998) 1685–1694.
- [4] J.D. Andrade, R.N. King, D.E. Gregonis, D.L. Coleman, Surface characterization of poly(hydroxyethyl methacrylate) and related polymers. 1. contact angle methods in water, *J. Polym. Sci., Polym. Symp.* 66 (1979) 313–336.
- [5] H.A. Güleş, K. Sarıoğlu, M. Mutlu, Modification of food contacting surfaces by plasma polymerisation technique. part I: determination of hydrophilicity, hydrophobicity and surface free energy by contact angle method, *J. Food Eng.* 75 (2006) 187–195.
- [6] A.R. Roudman, F.A. DiGiano, Surface energy of experimental and commercial nanofiltration membranes: effects of wetting and natural organic matter fouling, *J. Membr. Sci.* 175 (2000) 61–73.
- [7] M.A.C. Stuart, W.M. de Vos, F.A.M. Leermakers, Why surfaces modified by flexible polymers often have a finite contact angle for good solvents, *Langmuir* 22 (2006) 1722–1728.
- [8] D.K. Owens, R.C. Wendt, Estimation of the surface free energy of polymers, *J. Appl. Polym. Sci.* 13 (1969) 1741–1747.
- [9] D.H. Kaelble, Dispersion-polar surface tension properties of organic solids, *J. Adhes.* 2 (1970) 66–81.
- [10] A.X. Cartagena-Rivera, W.-H. Wang, R.L. Geahlen, A. Raman, Fast, multi-frequency, and quantitative nanomechanical mapping of live cells using the atomic force microscope, *Sci. Rep.* 5 (2015) 11692.
- [11] R. Lévy, M. Maaloum, Measuring the spring constant of atomic force microscope cantilevers: thermal fluctuations and other methods, *Nanotechnology* 13 (2002) 33–37.
- [12] L. Sonnenberg, J. Parvole, O. Borisov, L. Billon, H.E. Gaub, M. Seitz, AFM-based single molecule force spectroscopy of end-grafted poly(acrylic acid) monolayers, *Macromolecules* 39 (2006) 281–288.
- [13] A. AL-Baradi, M.R. Tomlinson, Z.Y.J. Zhang, M. Geoghegan, Determination of the molar mass of surface-grafted weak polyelectrolyte brushes using force spectroscopy, *Polymer* 67 (2015) 111–117.
- [14] M.A. Nash, H.E. Gaub, Single-molecule adhesion of a stimuli-responsive oligo(ethylene glycol) copolymer to gold, *ACS Nano* 6 (2012) 10735–10742.
- [15] J. Thompson, A. Rahardianto, S. Kim, M. Bilal, R. Breckenridge, Y. Cohen, Real-time direct detection of silica scaling on RO membranes, *J. Membr. Sci.* 528 (2017) 346–358.
- [16] A. Masse, O. Arab, V. Sechet, P. Jaouen, M. Pontie, N.E. Sabiri, S. Plantier, Performances of dead-end ultrafiltration of seawater: from the filtration and backwash efficiencies to the membrane fouling mechanisms, *Separ. Purif. Technol.* 156 (2015) 512–521.