



Draft Genome Sequence of the Entomopathogenic Bacterium *Bacillus* pumilus 15.1, a Strain Highly Toxic to the Mediterranean Fruit Fly *Ceratitis capitata*

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We present the draft whole-genome sequence of the entomopathogenic *Bacillus pumilus* 15.1 strain that consists of 3,795,691 bp and 3,776 predicted protein-coding genes. This genome sequence provides the basis for understanding the potential mechanism behind the toxicity and virulence of *B. pumilus* 15.1 against the Mediterranean fruit fly.

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Bacillus pumilus strain 15.1 has been recently described to be toxic against larvae of the Mediterranean fruit fly, *Ceratitis capitata*, one of the most damaging pests for fruits and vegetables worldwide (1). Strain 15.1 was isolated from a partially decomposed reed plant and has recently been characterized microbiologically and biochemically (D. C. Garcia-Ramon, C. A. Molina, A. Osuna, S. Vilchez, submitted for publication). *B. pumilus* strain 15.1 was deposited in the Spanish Type Culture Collection as CECT 7462.

B. pumilus strains are known to have very interesting properties, such as being highly resistant to environmental stresses (2) and producing a wide range of metabolites of industrial value. Nevertheless, only a few genomes from this species have been published. We report the first genome sequence of an entomopathogenic *B. pumilus* strain.

DNA from *B. pumilus* 15.1 was sequenced on a HiSeq 2000 sequencing system (Illumina Sequencing) in a single-read mode with a read length of 50 bases (GATC Biotech, Constance, Germany). The whole-genome sequencing yielded 26,322,535 reads. The reads were assembled in contigs by using Velvet software (3) with the *de novo* assembly tool and default parameters and then by iterative mapping using Geneious Pro R8 software, obtaining 63 contigs. The genome size of *B. pumilus* was 3,795,691, with a G+C content of 41.3%. Genome annotation was added by the NCBI Prokaryotic Genome Annotation Pipeline (PGAP), although it was also analyzed with BLAST (4) using a custom insecticidal toxin database (5, 6). A total of 3,776 protein-coding genes, 45 pseudogenes, 2 rRNA genes, 16 tRNA genes, and 1 noncoding RNA gene were predicted in the *B. pumilus* 15.1 genome.

B. pumilus 15.1 bears at least two extrachromosomal elements, one plasmid and one megaplasmid (D. Garcia-Ramon, M. J. Luque-Navas, C. A. Molina, C. del Val, A. Osuna, and S. Vilchez, submitted for publication). The plasmid sequence (7,785 bp, 35.7% G+C content) corresponds to contig 38 in the assembly. The megaplasmid has not yet been assigned to any contig or contig

sequences, but it has been detected in total DNA extractions on agarose gels (D. Garcia-Ramon et al., submitted). The *B. pumilus* 15.1 genome also contains prophage elements, even though a genome analysis revealed the presence of a clustered regularly interspaced short palindromic repeat (CRISPR)/Cas system (bacterial innate immune mechanism for protection from foreign DNA) (7). Interestingly, during sporulation this strain forms parasporal crystals that morphologically resemble those produced by *Bacillus thuringiensis* Cry proteins (D. C. Garcia-Ramon, C. A. Molina, A. Osuna, S. Vilchez, submitted for publication). The role of these crystalline structures has not yet been elucidated, and we have detected no *cry*-like gene coding sequences in the genome. However, the *B. pumilus* 15.1 genome harbors other genes encoding well-known entomopathogenic factors, such as chitinases (8), metalloproteases (9), and cytolysins (10).

The analysis and characterization of the *B. pumilus* 15.1 genome will provide new insights to allow the elucidation of the pathogenic mechanism of *B. pumilus* 15.1 against the Mediterranean fruit fly, providing useful information for the development of novel biotechnological products suitable for the control of this pest and the discovery of unknown virulence factors of this entomopathogenic bacterium.

Nucleotide sequence accession numbers. This whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number LBDK00000000. The version described in this paper is version LBDK00000000.1.

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REFERENCES

- 1. Molina CA, Caña-Roca JF, Osuna A, Vilchez S. 2010. Selection of a *Bacillus pumilus* strain highly active against *ceratitis capitata* (Wiedemann) larvae. Appl Environ Microbiol 76:1320–1327. http://dx.doi.org/10.1128/AEM.01624-09.
- 2. Gioia J, Yerrapragada S, Qin X, Jiang H, Igboeli OC, Muzny D, Dugan-Rocha S, Ding Y, Hawes A, Liu W, Perez L, Kovar C, Dinh H, Lee S, Nazareth L, Blyth P, Holder M, Buhay C, Tirumalai MR, Liu Y, Dasgupta I, Bokhetache L, Fujita M, Karouia F, Eswara Moorthy P, Siefert J, Uzman A, Buzumbo P, Verma A, Zwiya H, McWilliams BD, Olowu A, Clinkenbeard KD, Newcombe D, Golebiewski L, Petrosino JF, Nicholson WL, Fox GE, Venkateswaran K, Highlander SK, Weinstock GM. 2007. Paradoxical DNA repair and peroxide resistance gene conservation in *Bacillus pumilus* SAFR-032. PLoS One 2:e928. http://dx.doi.org/10.1371/journal.pone.0000928.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res 18:821–829. http:// dx.doi.org/10.1101/gr.074492.107.
- Altschul ŠF, Gish W, Miller W, Myers EW, Lipman DJ. 1990. Basic local alignment search tool. J Mol Biol 215:403–410. http://dx.doi.org/10.1016/ S0022-2836(05)80360-2.
- Palma L, Muñoz D, Berry C, Murillo J, Caballero P. 2014. Draft genome sequences of two *Bacillus thuringiensis* strains and characterization of a

- putative 41.9-kDa insecticidal toxin. Toxins 6:1490-1504. http://dx.doi.org/10.3390/toxins6051490.
- Palma L, Muñoz D, Murillo J, Caballero P. 2014. Draft genome sequence of bacillus thuringiensis serovar Tolworthi strain Na205-3, an isolate toxic for Helicoverpa armigera. Genome Announc 2(2):e00187-14. http:// dx.doi.org/10.1128/genomeA.00187-14.
- Barrangou R, Fremaux C, Deveau H, Richards M, Boyaval P, Moineau S, Romero DA, Horvath P. 2007. CRISPR provides acquired resistance against viruses in prokaryotes. Science 315:1709–1712. http://dx.doi.org/ 10.1126/science.1138140.
- 8. Cai Y, Yan J, Hu X, Han B, Yuan Z. 2007. Improving the insecticidal activity against resistant *Culex quinquefasciatus* mosquitoes by expression of chitinase gene chiAC in *Bacillus sphaericus*. Appl Environ Microbiol 73:7744–7746. http://dx.doi.org/10.1128/AEM.01510-07.
- 9. Fedhila S, Nel P, Lereclus D. 2002. The InhA2 metalloprotease of *Bacillus thuringiensis* strain 407 is required for pathogenicity in insects infected via the oral route. J Bacteriol 184:3296–3304. http://dx.doi.org/10.1128/JB.184.12.3296-3304.2002.
- Nishiwaki H, Nakashima K, Ishida C, Kawamura T, Matsuda K. 2007. Cloning, functional characterization, and mode of action of a novel insecticidal pore-forming toxin, sphaericolysin, produced by *Bacillus sphaericus*. Appl Environ Microbiol 73:3404–3411. http://dx.doi.org/10.1128/AEM.00021-07.