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Plasma miR-21, miR-155, miR-10b, and Let-7a as the potential biomarkers for the monitoring of breast cancer patients

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There is a pressing need for further studies to categorize and validate circulating microRNAs (miRs) in breast cancer patients that can be one of the novel strategies for cancer screening and monitoring. The present study is aimed to investigate the expression of the circulating candidate microRNAs after the operation, chemotherapy, and radiotherapy in the non-metastatic breast cancer patients. Tumor tissue and plasma samples were collected from the 30 patients with recently diagnosed Luminal A breast cancer. Control plasma samples were collected from the 10 healthy subjects. A panel of four miRs including miR-21, miR-55, miR-10b, and Let-7a were selected and their expression levels were measured before and after the operation, chemotherapy, and radiotherapy by using Real-Time PCR technique. The plasma expression of the miR-21, miR-155, and miR-10b was significantly increased and the Let-7a plasma expression decreased in the breast cancer patients compromised to the control ones. There was a similar expression pattern of the miRs between the tissue and plasma samples. The plasma levels of the miR-21, miR-155, and miR-10b were significantly down-regulated and the Let-7a plasma level was up-regulated after the operation, chemotherapy, and radiotherapy compromised to the pre-treatment. There was a significant difference in the miR-155 plasma level after the operation, chemotherapy, and radiotherapy compromised with each other. Moreover, there was no significant difference between the plasma levels of the miRs after the radiotherapy compromised to the control cases. The operation, chemotherapy, and radiotherapy led to a more reduction in the oncomiRs and an increase in the tumor suppressor-miRs. It seems that monitoring miRs during treatment might be considered as a respectable diagnostic tool for monitoring of breast cancer patients.

Breast cancer (BC) is the second leading cause of gynecological cancer deaths¹. The diagnosis of BC in the early stages, as well as monitoring of the disease progression and response to treatment, could be made easy with the aims of the liquid biopsy in the neoadjuvant setting². In this respect, existing diagnostic tools and biomarkers for BC have many inherent deficiencies³. A number of the circulating tumor markers including carcinoembryonic and carbohydrate antigens are widely used in BC monitoring, but the sensitivity of these markers is low³. Therefore, they cannot be used as reliable screening tools, although they have long been in clinical approaches. An ideal biomarker should be easily accessible such that it can be sampled noninvasively and be sensitive enough to detect the early presence of tumors⁴. This new approach has the potential to revolutionize clinical management including determining cancer classification, estimating prognosis, predicting therapeutic efficacy, maintaining surveillance following surgery as well as forecasting disease recrudescence⁵.

MicroRNAs (miRs) are the short single-stranded RNAs that have known as important regulators of the various cellular processes⁶. An estimated 30–60% of the genome is regulated by miR-mediated silencing⁶, though the aberrant expression of the miRs is associated with many diseases such as cancer. Early studies showed that some miRs can regulate cellular differentiation, proliferation and apoptosis processes that can be important in the cancer aggravation. A number of the differentially derived miRs from the tissues have reported and their expression profiles may be used as the potential biomarkers for the diagnosis, prognosis, and therapy. In addition, the

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| MicroRNAs | Target genes |
|------------------------|--|
| Up-regulation | |
| Mir-181 | ATM |
| Mir-155 | SOCS1 |
| Mir-10b | HOXD10 |
| Mir-373 | CD44 |
| Mir-520 | CD44 |
| Mir-103 | DICER |
| Mir-107 | DICER |
| Mir-21 | MASPIN, TPM1, PDCD4 |
| Mir-31 | FZD3, ITGA5, RDX, RHOA |
| Mir-193b | UPA |
| Mir-221 | P27 |
| Mir-222 | P27 |
| Mir-125b | BAK1 |
| Down-regulation | |
| MIR-30e | ITGB3, UBC9 |
| Mir-200 | HAS, TUP, CFA |
| Let -7 | HAS, TUP, CFA, SSC, BTA, MMU |
| Mir-335 | SOX4, TNC |
| MIR-126 | SSC, BTA, MMU |
| Mir-206 | TUP, CFA, SSC, |
| MIR-451 | MDR1 |
| Mir-345 | MRP1 |
| Mir-7 | MRP1 |
| Mir-125b | EPO, EPOR, ENPEP, CK2- α , CCNJ, MEGF9, ERBB2 |
| Mir-205 | HMGB3 |
| Mir-17-92 | Mekk2 |
| Mir-146 | NFkB, STAT3 |
| Mir-31 | RhoA, WAVE3, RhoA, WAVE3 |

Table 1. Main oncogenic and tumor suppressive of miRNAs in Luminal A breast cancer.

discovery of the roles of the miRs in developing BC may provide new opportunities for the development of the novel strategies for diagnosing and treating this type of the malignancy.

To date, only a few studies have begun to profile the circulating miRs in blood⁷. The analysis of the circulating miRs is at an early stage of development, and there is a persuasive need for further studies to categorize and validate circulating miR biomarkers in BC. Circulating miRs have been found to be significantly elevated in the blood of the cancer patients compromised with the healthy controls⁸. Furthermore, the elimination of the primary tumor leads to the loss of raised circulating miRs; suggesting that many of the miRs are 'tumor-derived' and 'cancer-specific'. The current belief is that these 'tumor-derived' circulating miRs are released from the primary tumor via the exosome vesicles and apoptotic bodies, though the discoveries of the exact underlying mechanisms are still developing⁹. Blenkiron *et al.*¹⁰ detected the expression levels of the different miRs between the basal and Luminal subtypes of the 309 miRs in 93 breast cancer patients¹⁰. Miska *et al.* (2015) showed that the different molecular subtypes of the BC including Luminal A, Luminal B, Basal-like, HER2, and Normal-like present the expression profiles of the different miRs¹¹. Moreover, a set of miRs was able to classify Luminal A from Luminal B tumors with the high expression of the Let-7 family members with Luminal A tumor, ER-positive status, and low tumor grade (Table 1)¹².

In cancer, miRs can be classified as oncogenes (oncomiRs) or tumor-suppressor miRs¹³. Chromosomal regions of the encompassing oncogenic miRs may be amplified that result in the increased expression of the oncomiR such as miR-21, miR-155, and miR-10b and the decreased the tumor suppressors such as Let-7a¹⁴. The Let-7a displays a lower expression in cancer cells and can suppress oncogene expression, thereby, may control the cellular differentiation¹⁵. In this respect, the distinct biological properties of the miRs including the remarkable stability, accessibility for rapid and accurate quantification, and a direct link with disease states make them the ideally suitable to serve as the minimally invasive biomarkers to track the disease¹⁶. Based on these data, in the first section of the present study, the correlation of the expression level of several dis-regulated miRs was assayed between the tissue and plasma samples in the breast cancer patients. In the second section, we explored the influence of the tumor resection and chemo-radiation on the expression of the candidate miRs and presented discussions with an emphasis on their role in monitoring treatment responses.

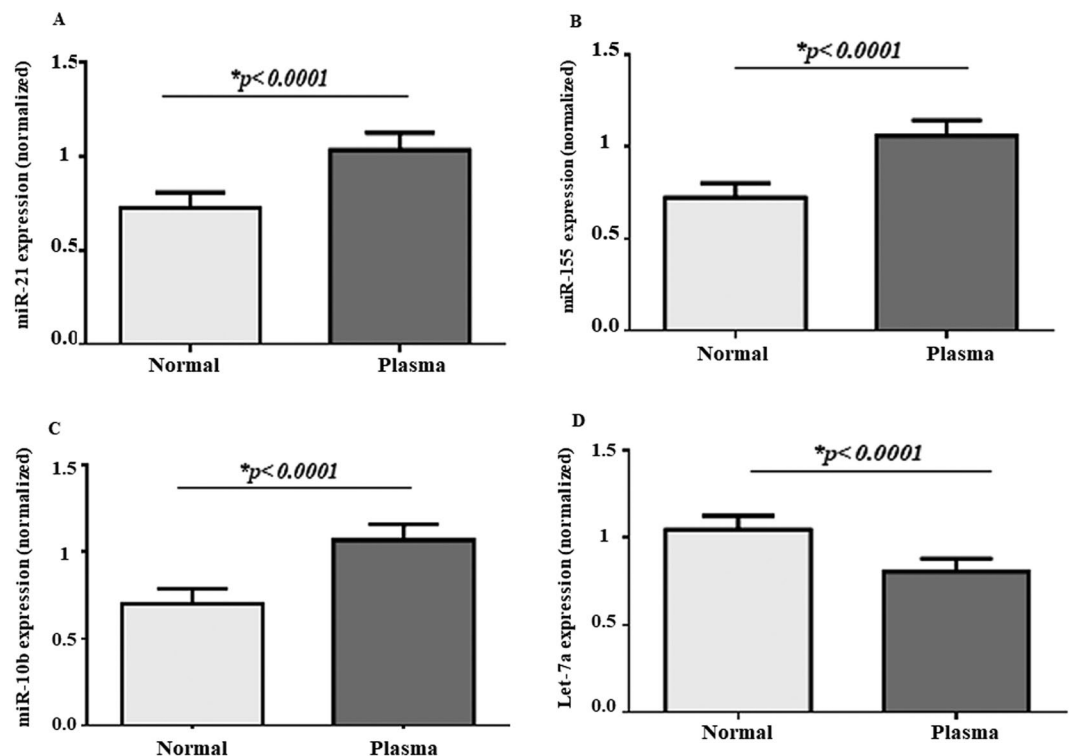


Figure 1. Comparison of the selected plasma miRs between the breast cancer patients and control. The relative expression level of the miR-21 (A), miR-155 (B), miR-10b (C), and Let-7a (D) were normalized using SNORD RNA as reference RNA.

Results

Pre-treatment analysis of the candidate miRs. We have determined the expression of miR-21, miR-10b, miR-155, and Let-7a in the plasma and the tissue of 40 samples (30 from patients with breast cancer and 10 from the controls). Except for three patients in the radiotherapy, all cases received cycles of the surgery, chemotherapy, and radiotherapy during the course of the study. Consequently, the relative abundance of the miR-21, miR-10b, and miR-155 was significantly up-regulated, and the Let-7a expression was significantly down-regulated in the plasma of the cases compromised with the control (Fig. 1).

Then, we have compromised the expression levels of the miR-21, miR-10b, miR-155, and Let-7a between the tissue and the plasma samples in 30 patients. The results showed a similar expression pattern between them and there were no significant changes (Fig. 2).

Moreover, we have stratified the patients to examine the associations between the plasma levels of the designated miRs and the stage II/III of the disease based on the TNM staging. Of the 30 cases of the breast cancer, the miR-21, miR-10b, miR-155, and Let-7a did show a significant difference in the staging compromised to the control (Fig. 3). Likewise, the expression of the miR-155 in the plasma showed a significant difference between stage II and III ($p = 0.0078$) (Fig. 3).

Furthermore, we have also compromised the expression levels of the miR-21, miR-10b, miR-155 and Let-7a between the tissue and plasma samples of the patients with the other clinical parameters including the nodal status (Table 2) and menopause status (Table 3). The results showed a similar expression pattern between the nodal status (-)/(+) of miR-21, but there was a significant change in the nodal status of the plasma expression of miR-155 and miR-10b and the tissue expression of Let-7a (Table 2).

Besides, our data showed a similar expression pattern between the menopause status of the miR-21, miR-10b, and miR-155 in both the plasma and tissue samples; nevertheless, there was a significant change in the menopause status of the Let-7a plasma expression (Table 3).

Monitoring of the miR-21, miR-10b, miR-155, and Let-7a during the course of the treatment.

Thirty patients with the non-metastatic breast cancer were monitored for changes in the plasma level of the miR-21, miR-10b, miR-155 and Let-7a before and after the surgery, chemotherapy, and radiotherapy. Then, we have compromised the expression levels of the miRs before and after the treatment. The miR-21 plasma level was significantly down-expressed after the operation (0.78 ± 0.09 , $p < 0.0001$), chemotherapy (0.71 ± 0.08 , $p < 0.0001$) and radiotherapy (0.70 ± 0.07 , $p < 0.0001$) than the pre-treatment (1.03 ± 0.09) (Fig. 4A). There was a significant difference in its plasma level after the chemotherapy and radiotherapy than the post-operation (1.03 ± 0.09), but there was no significant difference between the post-chemotherapy and post-radiotherapy (0.02347) (Fig. 4A).

Moreover, the miR-155 plasma level was significantly down-regulated after the operation (0.72 ± 0.07 , $p < 0.0001$), chemotherapy (0.66 ± 0.07 , $p < 0.0001$) and radiotherapy (0.62 ± 0.06 , $p < 0.0001$) than the

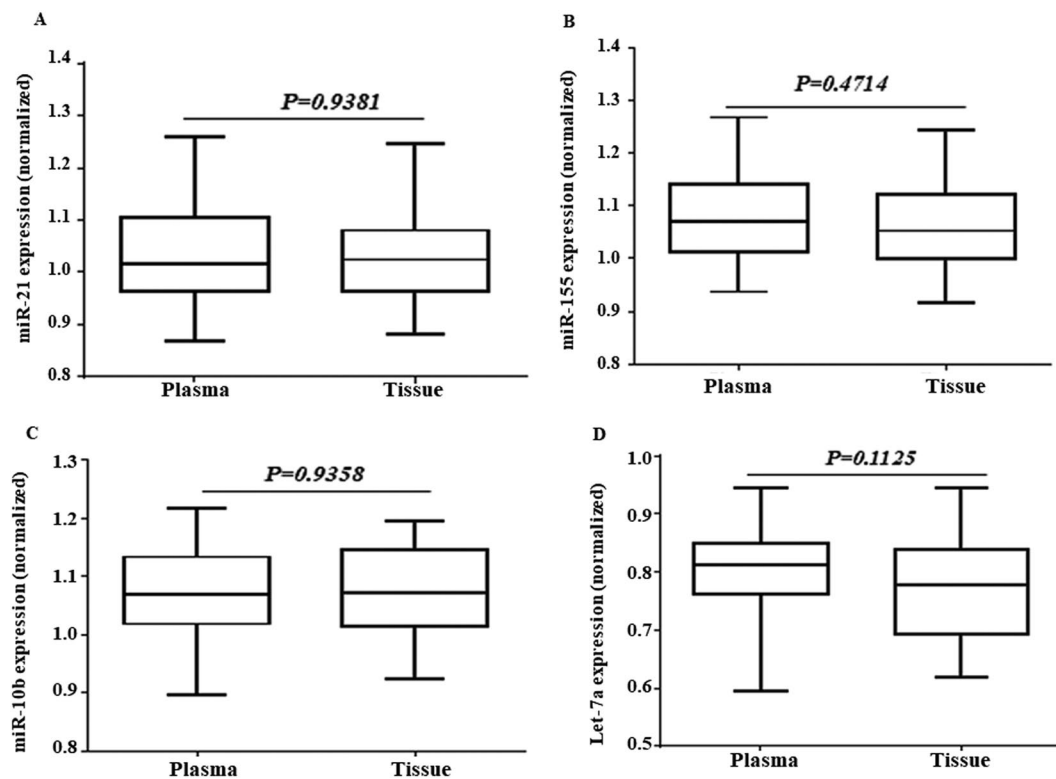


Figure 2. The differential expression of the selected miRNAs in both the tissue and plasma of the breast cancer patients. The relative expression level of the miR-21 (A), miR-155 (B), miR-10b (C), and Let-7a (D) were normalized using SNORD RNA as reference RNA. The line represents the median value.

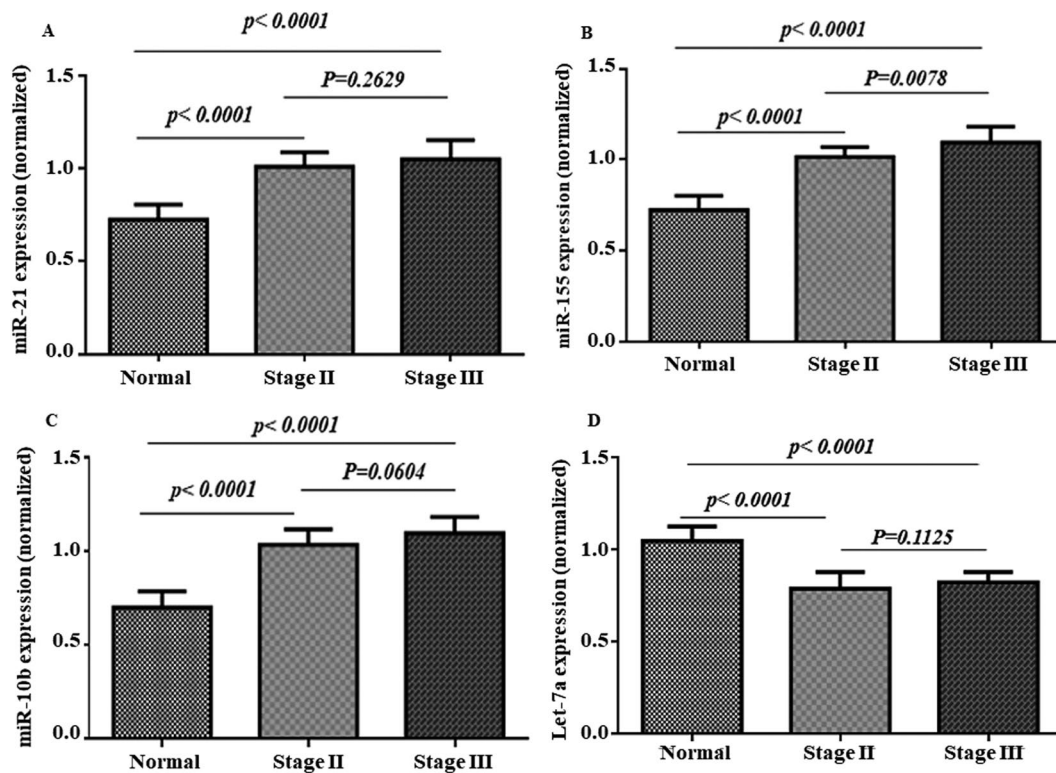


Figure 3. Comparison of stage II/III feature of the patients with the plasma relative expression of the selected miRNAs. The relative expression level of the miR-21 (A), miR-155 (B), miR-10b (C), and Let-7a (D) were normalized using SNORD RNA as reference RNA.

| Samples | Plasma | | P-value | Tissue | | P-value |
|---------|------------------|-----------------|---------|------------------|------------------|---------|
| | Nodal status (-) | Nodal status(+) | | Nodal status (-) | Nodal status (+) | |
| miR-21 | 1.0 ± 0.010 | 1.05 ± 0.02 | 0.1481 | 1.03 ± 0.03 | 1.03 ± 0.02 | 0.9148 |
| miR-155 | 1.01 ± 0.01 | 1.091 ± 0.02 | *0.0091 | 1.1 ± 0.03 | 1.07 ± 0.01 | 0.3637 |
| miR-10b | 1.02 ± 0.02 | 1.1 ± 0.02 | *0.0218 | 1.07 ± 0.02 | 1.07 ± 0.01 | 0.9512 |
| Let7a | 0.8 ± 0.02 | 0.9 ± 0.01 | 0.8519 | 0.8 ± 0.03 | 0.7 ± 0.02 | *0.0139 |

Table 2. The expression of the selected miRs and the nodal status (-)/(+) of the disease between the tissue and plasma samples.

| Samples | Plasma | | P-value | Tissue | | P-value |
|---------|-------------|-------------|---------|-------------|-------------|---------|
| | Pre | Post | | Pre | Post | |
| miR-21 | 1.04 ± 0.02 | 1.03 ± 0.02 | 0.7684 | 1.04 ± 0.02 | 1.03 ± 0.03 | 0.7516 |
| miR-155 | 1.06 ± 0.02 | 1.06 ± 0.02 | 0.8804 | 1.06 ± 0.02 | 1.1 ± 0.02 | 0.2644 |
| miR-10b | 1.1 ± 0.03 | 1.06 ± 0.02 | 0.5969 | 1.06 ± 0.02 | 1.08 ± 0.02 | 0.3689 |
| Let7a | 0.9 ± 0.01 | 0.8 ± 0.1 | *0.0215 | 0.8 ± 0.1 | 0.8 ± 0.02 | 0.3233 |

Table 3. Comparison of the menopause status of the breast cancer patients with the plasma relative expression of the selected miRs.

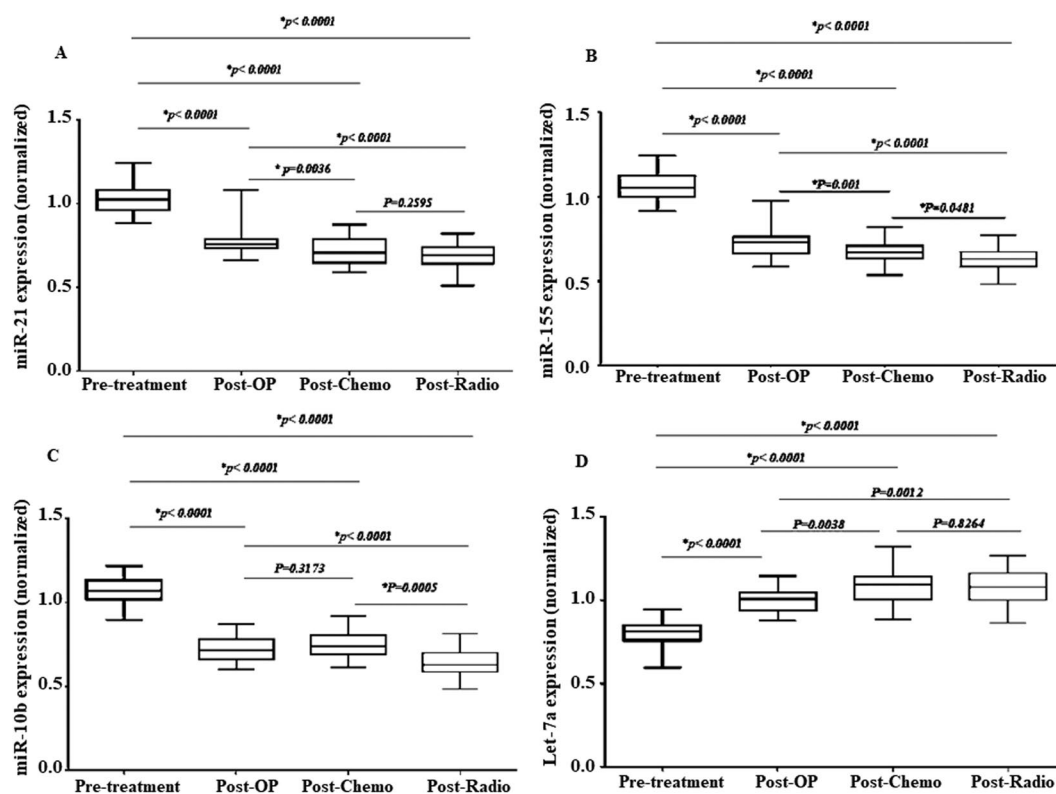


Figure 4. The expression of the selected miRs in the breast cancer patients before and after the treatment. The relative expression level of the miR-21 (A), miR-155 (B), miR-10b (C), and Let-7a (D) were normalized using SNORD RNA as reference RNA. The line represents the median value. Post-OP: Post-Operation, Post-Chemo: Post-chemotherapy, Post-Radio: Post-Radiotherapy.

pre-treatment (1.08 ± 0.08) (Fig. 4B). Interestingly, there was a significant difference (in its plasma level after the operation, chemotherapy, and radiotherapy than each other (Fig. 4B).

Furthermore, the miR-10b plasma level was significantly decreased after the operation (0.72 ± 0.07 , $p < 0.0001$), chemotherapy (0.75 ± 0.08 , $p < 0.0001$) and radiotherapy (0.64 ± 0.07 , $p < 0.0001$) than the pre-treatment (1.07 ± 0.09) (Fig. 4C). Additionally, there was no significant difference in its plasma level after the operation than the post-chemotherapy ($p = 0.317$), but there was a significant difference in its plasma level

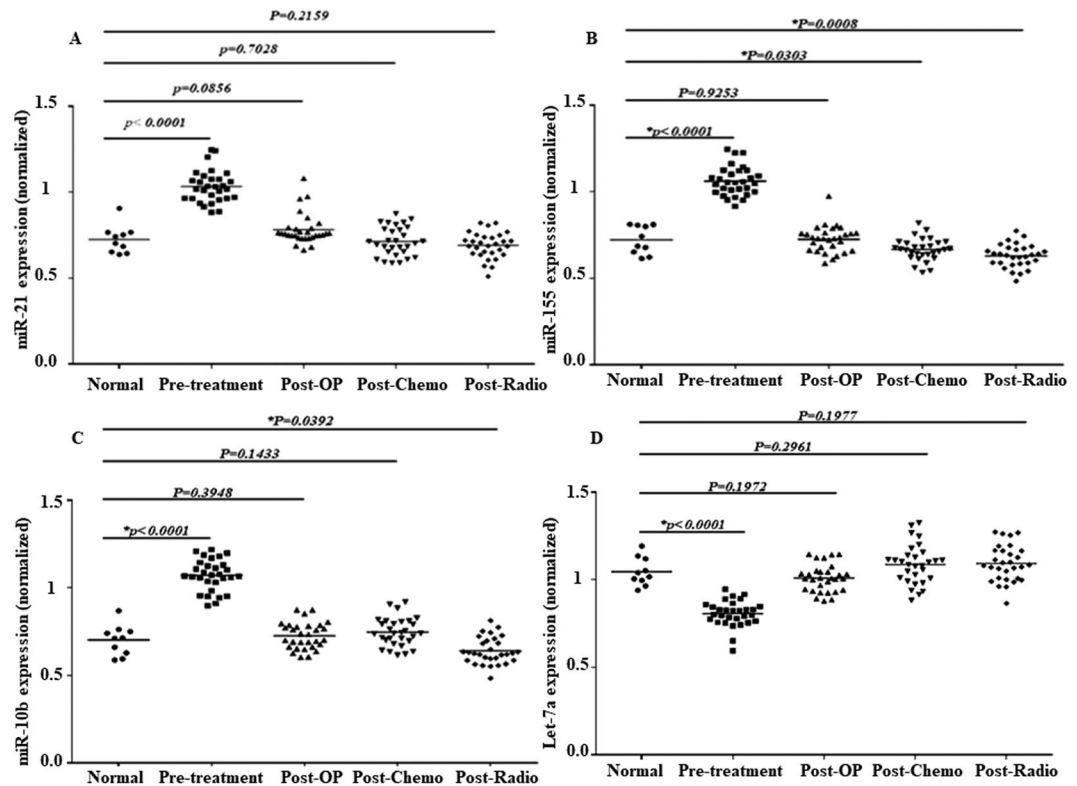


Figure 5. The expression of the selected miRs in the patients before and after the treatment compromised to the control. The relative expression of the miR-21 (A), miR-155 (B), miR-10b (C), and Let-7a (D) were normalized using SNORD RNA as reference RNA. The line represents the median value. Post-OP: Post-Operation, Post-Chemo: Post-chemotherapy, Post-Radio: Post-Radiotherapy.

after the radiotherapy than the post-operation (0.72 ± 0.07 , $p < 0.0001$) and post-chemotherapy (0.75 ± 0.08 , $p < 0.0001$) (Fig. 4C).

The Let-7a plasma level was significantly up-expressed after the operation (1.0 ± 0.07 , $p < 0.0001$), chemotherapy (1.08 ± 0.11 , $p < 0.0001$) and radiotherapy (1.1 ± 0.10 , $p < 0.0001$) than the pre-treatment (0.80 ± 0.07) (Fig. 4D). Nonetheless, there was no significant difference in its plasma level after the surgery, chemotherapy, and radiotherapy than each other (Fig. 4D).

Finally, we have compromised the plasma levels of the miRs between the control subjects and the course of treatments individually (Fig. 5). The miR-155 plasma level underwent more decrease post-chemotherapy ($p = 0.03$) and post-radiotherapy ($p = 0.0008$) than the control subjects. In addition, the miR-10b plasma level was done a more decrease post-radiotherapy ($p = 0.0392$) than the control subjects.

Discussion

The aim of the present study is to evaluate the plasma levels of the miR-21, miR-10b, miR-155 and Let-7a in the non-metastatic BC patients following the common treatments such as the surgery, chemotherapy, and radiotherapy using RT qPCR. Our results showed that the expression levels of the oncomiRs such as the miR-21, miR-10b, and miR-155 were significantly increased, while the expression level of the tumor suppressor such as the Let-7a was significantly decreased in the plasma of the patients. Remarkably, using the common treatments has reversed these effects. In this context, many of the studies have been conducted on the expression of the miRs in the tumorigenesis processes¹⁷. The first report of the miRs related to cancer was shown in the patients with B cell chronic lymphocytic leukemia¹⁸. Blenkiron *et al.*¹⁰ have also identified 133 miRs that displayed the abnormal expression levels in the breast tumor tissues compromised to the normal breast tissues. Similar to our results, they showed a difference of 29 miRs which have a key role in breast cancer development¹⁰. Furthermore, we have compromised the expression levels of the miR-21, miR-10b, miR-155, and Let-7a between the tissue and the plasma samples of the patients. The results showed a similar expression pattern between them and there were no significant changes. This finding was not supported by the obtained results by Matamala *et al.*⁹. Nevertheless, Svoronos *et al.*¹³ have also shown that a high correlation between the miRNA expression level that was found between the breast tumor tissues and the serum level. Here, the miR-21, miR-106a, and miR-155 were significantly over-expressed in the tumor specimens compromised with the control, whereas miR-126, miR-199a, and miR-335 were significantly under-expressed. Furthermore, the relative expression of the miR-21, miR-126, miR-155, miR-199a, and miR-335 was closely associated with the clinicopathological features of the breast cancer such as the histological tumor grades and expression of sex hormone receptor^{13,19}.

We have also analyzed the results from the selected miRs expression in the plasma to evaluate whether there was a correlation between the expression level of the miRs and the various clinic-pathologic features or not. For example, the expression of the miR-21 and miR-155 is related to the expression of the estrogen receptor (ER) and progesterone receptor (PR)²⁰. Approximately 70% of breast tumors overexpress ER²⁰. The ER up-regulation during the early stages of tumorigenesis has been identified as an important factor in stimulating the mammary cell proliferation which can lead to tumor development²¹. Similarly, Al-Khanbashi *et al.*²² observed the expression patterns of the miRs associated with Her2/neu/ER/PR in the breast tumors²². In this context, two other independent studies have also shown an increase in the expression of the miR-21 in the breast cancer patients as compromised to the healthy ones²³. Furthermore, the plasma levels of the miR-10b and miR-155 can also be detected between the breast cancer patients and the healthy ones. Besides, the serum level of the miR-155 in PR+ tumors has shown a significant difference in compromised to PR- cases²⁰. Wang *et al.* (2015) showed that the expression of the miR-21 and miR-155 was associated with the clinical pathological features of the breast tumors such as the histological grades and the sex hormone receptor²⁴. The molecular subtypes of the ER are characterized by different responses to the therapy, differential course, and prognosis²⁰. The variances between the ER+ and ER- breast tumor not only relate to their morphology but also are mostly due to the alteration in their transcriptional reactions. Moreover, the expression of the oncomiRs can be due to the transcriptional exacerbation of their genes due to the availability of the transcription factors, hyper-methylation or placement in the intra-region or between the genes. MiRs are heavily linked to cancer and can play a role through the effects of the key points in cell cycle regulation, genome integrity, and response to stress, apoptosis, and metastasis. In this respect, the difference between the expressions of the miRs may be due to differences in the sample sources, different analytical methods, or different platform in the studies. Therefore, it seems that an increasing or decreasing expression of miRs can be related to various reasons.

Moreover, we have stratified the patients to examine the associations between the plasma levels of the designated miRs and the stage II/III of the disease based on the TNM staging. Of the 30 cases of the breast cancer, the miR-21, miR-10b, miR-155, and Let-7a did show a significant difference in the staging compromised to the control. Besides, the expression of the miR-155 in the plasma showed a significant difference between stage II and III. Wang *et al.* (2015) have shown that the miR-21, miR-106a, and miR-155 were significantly up-regulated in higher malignancy grades compromised to the control breast tissues¹³. The relative expression of the miR-21 was not altered in the benign tumors but increased 2 fold in the grade II tumors and 4.5 fold in the grade III tumors. MiR-155 expression was not altered in the benign tumors but was increased 2 fold and 5 fold in the grade II and III, respectively. In contrast, the miR-21, miR-126, miR-155, miR-199a, and miR-335 were highly correlated to ER or PR in both the grades. There was a greater difference in expression levels of samples with negative hormone receptor expression ($P = 0.05$), which was a predictive factor for prognosis of patients with breast cancer. The findings suggest that the miRs can be used to identify the different nature of breast tissues, and deregulation of the selected miRs may affect critical molecular events involved in tumor progression. Thus, the measurements of miRs as the biochemical markers can help to diagnose the different stages of cancer prior to clinical investigations on the samples.

Additionally, the present study investigates the changes in the expression level of the miRs after common treatments including operation, chemotherapy, and radiotherapy. Our results showed that the miR-21 plasma level was significantly down-expressed after the operation, chemotherapy, and radiotherapy than the pre-treatment. Moreover, there was a significant difference in its plasma level after chemotherapy and radiotherapy than post-operation. These results are consistent with Chang *et al.*²⁵, Badr *et al.*²⁶, and Farsinejad *et al.*²⁷. They found that patients with better miR-21 expression after treatment have better clinical outcomes that can increase survival^{25–27}. Studies have also shown that miR-21 can reduce the expression of the tumor suppressor proteins and increase the expression of the oncogene proteins²⁸. Likewise, the increase in the expression of the miR-21 in the breast tumor tissues has been shown to be directly related to the incidence of the disease, tumor size, and staging. The miR-21 can be an effect on tumor cells and may be of an anticoagulant and anti-apoptosis effect, and disrupts the pathway of apoptosis which is in the interest of cancer cell survival²⁹. Therefore, it seems that miR-21 can be a good alternative to monitoring cancer or metastasis.

Moreover, the present study showed that the miR-155 plasma level was significantly down-regulated after the operation, chemotherapy, and radiotherapy than the pre-treatment. Interestingly, there was a significant difference in its plasma level after the operation, chemotherapy, and radiotherapy than each other. Similar to our results, Sochor *et al.*³⁰ showed that after surgery, the expression of the miR-155 was significantly decreased compromised to after chemotherapy. Their results have indicated a correlation between the removal of the breast tumors during the treatment and the miR-155 serum level³⁰. Nevertheless, Sun *et al.*³¹ have shown a short-term increase in the expression of the miR-155 after surgery, probably due to the removal of the tumor and the miR-155 release to the blood, although the decrease of the miR-155 expression after chemotherapy was similar to our results³¹.

Additionally, the miR-10b plasma level was significantly decreased after the operation, chemotherapy, and radiotherapy than the pre-treatment. There was no significant difference in its plasma level post-operation than post-chemotherapy, but there was a significant difference in its level of post-radiotherapy than post-operation and post-chemotherapy. Ma *et al.*³² showed that miR-10b level in the non-metastatic breast tumors was declined in the metastatic patients³². Similar to our results, Gee *et al.*³³ showed that the decreased levels of the miR-10b expression can occur in the early stages of the breast tumors³³. Moreover, our results showed a significant difference in the expression level of miR-10b after the operation, chemotherapy, and radiotherapy which is contrary to the results of Iorio and Croce *et al.*³⁴. In addition, a meta-analysis by Huang *et al.*³⁵ showed that the expression of the miR-10b is different in the tumor stages. These differences may be related to the cell type, the tumor stage or the tissue source³⁵. Thus, the miR-10b can be a prognosis for the early detection or a therapeutic goal for metastasis treatment.

| Characteristic | Patients (%) |
|---|--------------------------|
| Total | N = 30 |
| Age | |
| Mean | 45.52 years |
| Median (range) | 45.5 (range 26–70 years) |
| TNM stage | |
| II | 13 |
| III | 17 |
| T classification | |
| T1 | 7 |
| T2 | 11 |
| T3 | 12 |
| Nodal status | |
| Negative | 11 |
| Positive | 19 |
| ER⁺, PR⁺, Her2⁻ | |
| Negative | 0 |
| Positive | 30 |
| Menopause | |
| premenopausal | 14 |
| postmenopausal | 16 |
| Subtype | |
| Luminal A | 30 |

Table 4. The information of the patients.

In addition, our results showed that the Let-7a plasma level was significantly up-expressed after the operation, chemotherapy, and radiotherapy than the pre-treatment which is similar to Weidhaas *et al.*³⁶. Wang *et al.* (2013) have also shown that the high expression of the Let-7 can increase the sensitivity of the breast cancer cells to radiotherapy, which can help with treatment³⁷. Several studies have shown the Let-7 variations in the sensitivity of the breast cancer cells during radiotherapy³⁷. Surprisingly, in the present study after radiotherapy, the Let-7a level was increased and reached the level of the healthy ones.

Conclusion

In summary, the expression of the miR-21 and miR-155 was decreased, and the expression of Let-7a was increased at the end of the present study and reached the expression level of healthy individuals. The chemotherapy and radiotherapy were led to a more reduction in the oncomiRs and an increase of the tumor suppressor. Generally, in the absence of a recognizable tumor, an increase in the oncomiRs level or a decrease in the tumor suppressors may indicate a failure in treatment. As a result, measuring the expression of the miRs after the operation, chemotherapy, and radiotherapy can be considered as a recognizable marker for proper response of the patients to the treatment. It is worth noting that reducing oncomiRs and increasing tumor suppressors during treatment can be considered as a good diagnostic tool for the process of improvement and proper response to the standard treatments. Moreover, it should be noted that we will follow the patients annually for the first 5 years to relapse the disease or metastasis using the liquid biopsy and measure the candidate miRs.

Materials and Methods

Ethics statement. This study was approved by the Medical Ethics Committee of the Tehran University of Medical Sciences (NO: 23797) and Iranian Randomized Control Trial (IRCT) ethical board. In addition, the written informed consent was obtained from each participant prior to the sample collection.

Study design and sample collection. In the present study, the primary breast tumor samples were obtained from the 30 patients (Luminal A; ER⁺, PR⁺, Her2⁻) with informed consent approved, and the level of the plasma miR-155, miR-21, miR-10b, and Let-7a in the cases and 10 control plasma samples were screened. Then, we evaluated the changes in the levels of the miRs after the operation, chemotherapy, and radiotherapy. Pre-operative plasma from the patients with the histologically diagnosed breast cancer (n = 30) was drawn at Imam Khomeini Hospital, Cancer Institute of Iran from Oct 2016 to Sep 2017. The characteristic of the patients including age, T classification, nodal status, hormone-receptor positive, HER2 overexpression, and tumor subtype were retrospectively collected (Table 4). The patients with the severe infection, active clinical comorbidities, or a history of any other malignancy were excluded. For 30 patients who underwent treatment, the second sample was obtained one week before the chemotherapy, the third sample was collected at the periodical evaluation one week before the commencement of radiotherapy, and the fourth sample was collected at the periodical evaluation one month after radiotherapy. The applied adjuvant chemotherapies were epirubicin/cyclophosphamide, epirubicin/taxane, cyclophosphamide/pirarubicin or fluorouracil epirubicin/cyclophosphamide with and without taxane. In these patients, the response to therapy was assessed by the specialist doctors according to the World Organization

(WHO) guidelines³⁸. Additionally, the plasmas from a set of the 10 healthy females were collected from outpatients at Imam Khomeini Hospital. All participants were of Iranian ethnic. None of the healthy controls had previously diagnosed with any malignancies. The median age of these healthy cases was 45 (range from 26 to 70). There is no significant difference in age between the breast cancer patients and the controls ($p = 0.6999$, Mann-Whitney t-test). The blood sample from each participant was collected in the tube with EDTA (BD vacutainer SSTTM Tubes, Reference No. 367985). After exposure to the room temperature for 30 min to 2 hours, the specimens were centrifuged at 1,500 g for 20 min at 4 °C. The plasmas were aliquoted into microcentrifuge tubes and stored at -80°C before use.

Identification of the breast cancer-related miRs. The Gene Expression Omnibus database (GEO, <http://www.ncbi.nlm.nih.gov/geo/>) in the National Center for Biotechnology Information (NCBI) is the largest fully public gene expression resource and includes 214,268 samples and 4,500 platforms³⁹. The selected miRs were mapped into the human miR disease database (HMDD; <http://cmbi.bjmu.edu.cn/hmdd> and <http://202.38.126.151/hmdd/tools/hmdd2.html>) to further select the differentially expressed miRs related to BC. As a database for experimentally supported human miRs and disease associations, HMDD serves as a valuable resource for studying the roles of miRs in human disease⁴⁰. Furthermore, the target genes of the differentially expressed miRs of breast cancer-related tissues were predicted by five miR databases, named miRanda (<http://microrna.sanger.ac.uk>)⁴¹, MirTarget2 (<http://nar.oxfordjournals.org/cgi/content/abstract/34/5/1646>)⁴², PicTar (<http://pictar.bio.nyu.edu>)⁴³, PITA (<http://genie.weizmann.ac.il/pubs/mir07>)⁴⁴ and TargetScan (<http://targets-can.org>)⁴⁵. Additionally, the published oncogenes and suppressors of breast cancer were selected from TSGene (<http://bioinfo.mc.vanderbilt.edu/TSGene/>)⁴⁵ and Tumor-Associated Gene (TAG; <http://www.binfo.ncku.edu.tw/TAG/>) databases⁴⁶. In the present study, DAVID has applied to conduct Kyoto encyclopedia of genes and genomes (KEGG) pathway and gene ontology (GO) enrichment analyses for the identified target genes. KEGG is a knowledge base for systematic analysis of gene functions⁴⁷.

Quantitative Real-Time PCR Analysis. Total RNA was extracted from 100 μl plasma samples and 50 μg tumor samples using 1 ml trizol reagent according to the manufacturer's instructions (Sinagene, Tehran, Iran). The trizol reagent is used for isolating both enriched miRNAs and larger RNA species. Qualitative and quantitative assessment of the isolated RNA was carried out by the electrophoresis and spectrometric methods⁴⁸. The RNA was stored at -80°C for later analysis. For miRs quantification by Real-Time PCR in all samples, 10 μl of the total RNA were reverse-transcribed in a 20 μl reaction mix using the BONmiR 1st-strand cDNA synthesis kit (stem cell technology research center, Tehran, Iran) following the manufacturer's recommendations. Then, cDNA was used in each of the real-time PCR assays with the BONmiR qPCR Kit (stem cell technology research center, Tehran, Iran) based on the manufacturer's instructions. Real-Time PCR analyses of the miRs were carried out in triplicate. The levels of miRs were normalized using SNORD RNA as reference RNA. MicroRNA gene expression was analyzed by means of the Step-One system (ABI, Massachusetts, USA) and the expression levels were evaluated using $2^{(-\Delta\Delta\text{ct})}$.

Relative expression of the miRNA was normalized to SNORD and was calculated using the $2^{(-\Delta\Delta\text{ct})}$ method. ΔCT was calculated by subtracting the CT values of SNORD from the CT values of the target miRs. $\Delta\Delta\text{CT}$ was then determined by subtracting average ΔCT of the control from the ΔCT of cases. The fold changes of candidate miRNA expression were calculated by the equation $2^{(-\Delta\Delta\text{ct})}$ ^{49,50}.

The expression levels were compromised with the healthy ones and expressed as fold change. Sequences of the used forward primers are as below:

MiR-21 Forward primer: ACGTGTTAGCTTATCAGACTG
 MiR-155 Forward primer: CCGTTAATGCTAATCGTG
 MiR-10b Forward primer: TAAGCACGAGACTTACGGAGGA
 Let-7a Forward primer: GGCTGAGGTAGTAGGTTGTATAG
 Snord Forward primer: ATCACTGTAACCGTTCCA

Universal Reverse Primers were obtained from Bonyakhteh Company (Bonyakhteh, Tehran, Iran)

Data analysis. All data presented as mean \pm SD. The statistical analyses were performed using the SPSS 16.0 software (SPSS) and the GraphPad Prism 5.0, GraphPad. Kolmogorov-Smirnov was used to evaluate the natural distribution of data. For inferential analysis of data in two groups, parametric data from the t-test and non-parametric data from the Mann-Whitney method were used. The repaid measured ANOVA test was used to evaluate the intra-group treatment trend. P values less than 0.05 was considered to be statistically significant.

Ethical approval. All procedures performed in studies involving human participants were in accordance with the ethical standards of the institutional and/or national research committee and with the 1964 Helsinki declaration and its later amendments or comparable ethical standards.

Informed consent. Informed consent was obtained from all individual participants included in the study.

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Author Contributions

S.K.: manuscript preparation, sample collection, sample processing, and data analysis. A.M.A.: study conception and design, and article revision. S.I.: sample processing. R.O.: clinical annotation.

Additional Information

Competing Interests: The authors declare no competing interests.

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