

The impact of aging on regulatory T-cells

Johannes Fessler¹, Anja Ficjan¹, Christina Duftner² and Christian Dejaco¹*

¹ Department of Rheumatology and Immunology, Medical University Graz, Graz, Austria

² Department of Internal Medicine, General Hospital Kufstein, Kufstein, Austria

Edited by:

Dietmar Herndler-Brandstetter, Yale University School of Medicine, USA

Reviewed by:

Nikolai Petrovsky, Flinders Medical Centre, Australia Tyler Curiel, University of Texas Health Science Center at San Antonio, USA

*Correspondence:

Christian Dejaco, Department of Rheumatology and Immunology, Medical University Graz, Auenbruggerplatz 15, A-8036 Graz, Austria e-mail: christian.dejaco@gmx.net Age-related deviations of the immune system contribute to a higher likelihood of infections, cancer, and autoimmunity in the elderly. Senescence of T-lymphocytes is characterized by phenotypical and functional changes including the loss of characteristic T-cell surface markers, while an increase of stimulatory receptors, cytotoxicity as well as resistance against apoptosis is observed. One of the key mediators of immune regulation are naturally occurring regulatory T-cells (T_{regs}). T_{regs} express high levels of CD25 and the intracellular protein forkhead box P3; they exert their suppressive functions in contact-dependent as well as contact-independent manners. Quantitative and qualitative defects of Tregs were observed in patients with autoimmune diseases. Increased T_{reg} activity was shown to suppress antitumor and anti-infection immunity. The effect of aging on Tregs, and the possible contribution of age-related changes of the Treg pool to the pathophysiology of diseases in the elderly are still poorly understood. Trea homeostasis depends on an intact thymic function and current data suggest that conversion of non-regulatory T-cells into Tregs as well as peripheral expansion of existing T_{reas} compensates for thymic involution after puberty to maintain constant Treg numbers. In the conventional T-cell subset, peripheral proliferation of T-cells is associated with replicative senescence leading to phenotypical and functional changes. For Treas, different developmental stages were also described; however, replicative senescence of T_{reas} has not been observed yet.

Keywords: FOXP3, regulatory T-lymphocyte, aging, cellular senescence, thymus, suppressor cells

INTRODUCTION

The immune system combats against infectious agents and depletes damaged or transformed cells, whereas intact selfcomponents are usually ignored. Nevertheless, clinical manifestations of autoimmunity occur in at least 5% of the general population. The exact causes of autoimmune diseases are elusive; however, genetic and environmental risk factors as well as an insufficient elimination of cells bearing autoreactive T-cell receptors (TCRs) in the thymus contribute to the evolvement of disease (1, 2). To prevent autoimmunity, tolerance mechanisms including clonal deletion, induction of apoptosis, or anergy of self-reactive T-cells are essential. In addition, regulatory T-cells (Tregs) were identified as sentinels of the immune response keeping aberrant/exaggerated immune reactions in balance. Several distinct T-cell subsets with regulatory function have been identified so far including natural T_{regs}, adaptive or induced T_{regs} (iT_{reg}), type 1 regulatory T-cells (Tr1), T helper 3 cells (Th3), double-negative (dn) T-cells, yo T-cells, and iNKT cells. In a number of autoimmune diseases a diminished prevalence and/or impaired function of Tregs were observed (3). As several autoimmune disorders (such as rheumatoid arthritis or vasculitis) occur more frequently in the elderly, the question arises whether aging is linked to quantitative and/or qualitative defects of the T_{reg} pool (4-6).

In this review we summarize current data about the effects of aging on T_{regs} and highlight the possible mechanisms leading to senescence of T_{regs} .

CHARACTERIZATION OF TREGS

DEFINITION AND PHENOTYPE

Natural T_{regs} develop in the thymus through recognition of selfantigen presented by thymic epithelial or dendritic cells. For this process CD28 co-stimulation is required, whereas IL-2 and TGF- β are less important as indicated by knock-out mice models (7).

Today, there is still no consensus on the reliable identification of T_{regs} by flow cytometry. A variety of cell surface molecules have been proposed as specific T_{regs} markers such as glucocorticoid-induced tumor necrosis factor receptor (GITR), cytotoxic T-lymphocyte associated antigen-4 (CTLA-4), the coreceptors Neuropilin-1 and PD-1, the adhesion molecule CD62L, major histocompatibility complex (MHC) class II DR, or CD45 isoforms. The type I cytokine receptor CD127 is a negative marker of T_{regs} and the absence of this molecule is frequently used for T_{reg} identification (8).

The forkhead transcription factor FoxP3 was proposed as the most specific marker of T_{regs} as FoxP3 expression is essential for T_{reg} development and function (9): T_{regs} were unable to develop in a mouse receiving FoxP3-deficient progenitor cells from another animal (10) and retroviral expression of FoxP3 in human and murine T-cells enabled the conversion of non-regulatory naïve T-cells into a T_{reg} -like phenotype with suppressive activity and surface expression of CD25 (9). A mutation of the *FoxP3* gene in humans results in the fatal autoimmune syndrome IPEX (immune dysregulation, polyendocrinopathy, X-linked) (11). For experimental studies, however, FoxP3 appears not to be an optimal T_{reg}

marker because first, permeabilization of T-cells is necessary to stain FoxP3 and cells are thus not viable anymore and second, newer data indicate that human FoxP3 is up-regulated in activated T-cells without suppressive function as well (12).

The Ikaros family transcription factor Helios was proposed as an alternative indicator of human T_{regs} with a higher specificity compared to FoxP3. Recent data, however indicate that Helios is also up-regulated in activated non-regulatory T-cells (13). In summary, there is currently no specific marker of human T_{regs} available limiting the validity of studies investigating qualitative and/or quantitative changes of the T_{reg} pool.

MECHANISM OF SUPPRESSION

The mechanisms of T_{reg} mediated immunosuppression are still unclear. Most likely, T_{regs} have multiple functions with direct and indirect inhibitory effects on antigen-presenting cells (APCs) and T-cells such as the following (14, 15): (a) expression of the surface molecule CTLA-4 directly suppressing the activity of Tcells, (b) indirect inhibition of effector cells by the induction of anti-inflammatory biochemical pathways in APC, (c) direct or indirect killing of effector cells and APCs, and/or (d) production of immunoregulatory cytokines such as TGF- β and IL-10 (16).

Interestingly, a recent study reported that human T_{regs} are able to induce senescence of naïve and memory responder T-cells *in vitro* and *in vivo*. The resulting senescent T-cell subset had an altered phenotype and revealed potent suppressive functions. The mechanisms leading to senescence of non-regulatory T-cells were not completely understood; however, the phosphorylation of p38 and ERK1/2 signaling pathways inhibiting naïve T-cell growth and cell-cycle regulation appeared to play a role (17).

THE EFFECT OF AGING ON T_{REG} PREVALENCES AND FUNCTION

A prevalence of approximately 0.6–15% out of the CD4⁺ T-cell pool has been reported for T_{regs} in healthy adults and mice (4, 18). The influence of aging on T_{reg} prevalence in humans has been rarely studied so far and available reports suggest only minor changes of the circulating T_{reg} pool through age (19). Higher proportions of T_{regs} were only found in cord blood samples suggesting a pivotal role of T_{regs} during homeostatic proliferation of naïve Tcells in the fetal life (20, 21). During the first 36 months of life T_{reg} levels decline rapidly (22) and remain relatively stable thereafter.

Mouse studies showed increased T_{reg} prevalences in lymphoid organs of aged compared to young animals, whereas frequencies in circulating blood and thymus were unchanged (23, 24). This finding led to the hypothesis that during aging T_{regs} accumulate in lymphoid tissues; hypothetically explaining the increased susceptibility to infections and reduced vaccine response in elderly animals. The accumulation of T_{regs} has further been observed in the skin of aged persons possibly resulting in a higher risk of skin cancer as T_{regs} reduce local anti-tumor immune responses (25–27).

In animals, T_{reg} function seems to decrease with advancing age. The transfer of CD25⁺ T_{regs} from aged mice into young animals for example resulted in a lower suppression of delayed type hypersensitivity responses compared to the infusion of young T_{reg} cells (23). Another study found that CD4⁺CD25^{high} T_{regs} from

aged animals less efficiently inhibited the proinflammatory activity of IL-17⁺ T-cells compared to T_{regs} from young mice (28). In human studies it was observed that T_{regs} from young and elderly individuals similarly inhibited the proliferation of responder cells whereas the production of the anti-inflammatory cytokine IL-10 was reduced in cells from the older group. The phenotype of T_{regs} including expression of CD25, FoxP3, IL-7R α , or chemokine receptor expression, however, was unchanged (29). In conclusion T_{regs} from aged individuals are less efficient in preventing the occurrence of autoimmunity, while their number remains unaltered.

On the other hand, cancer and infections occur more commonly in the elderly suggesting increased T_{reg} responses (see also above) (29–31). One mouse study found an increase of T_{reg} prevalences in aged animals correlating with a defective tumor clearance. CD25-depletion restored the anti-cancer immune response (32). Similarly, CD25-depletion in aged mice reduced the lesion size in a *Leishmania major* infection model (24). Others reported that the depletion of T_{regs} with denileukin diffitox improved tumorspecific immunity only in young mice whereas tumor growth was unaffected in aged mice. This was explained by increased numbers of myeloid-derived suppressor cell (MDSC) in aged animals, and upon depletion of these cells tumor-specific immunity was restored (33).

In summary, current data on age-related changes of T_{reg} prevalences and function are conflicting and do not completely explain the simultaneously increased risk of autoimmunity (suggesting lower T_{reg} function), cancer, and infections (indicating increased T_{reg} responses) in the elderly. Apart from the difficulty of a reliable identification of T_{regs} the possible accumulation of T_{regs} in lymphoid organs and/or tissues during aging might lead to an underestimation of the total T_{reg} pool in current human studies. Future studies investigating tissue samples from immune-organs of elderly individuals would be desirable to better understand the role of T_{regs} in the pathogenesis of age-related diseases.

T_{REG} DEVELOPMENT AND HOMEOSTASIS

Development of natural T_{regs} in the thymus depends on a positive selection process including high affinity interactions of the TCR to cortically expressed host antigens. Thymic stromal lymphopoietin activated CD11c-positive dendritic cells (34), co-stimulatory molecules including CD28, PD-1, CD40L (35) as well as the cytokine IL-2 were all shown to be crucial for thymic T_{reg} generation (36–38). Besides, the Nr4a nuclear receptors (involved in apoptosis, proliferation, DNA repair, inflammation, and others) were recently reported to contribute to T_{reg} development. Mice lacking these receptors in T-cells were unable to produce T_{regs} and died early from systemic autoimmunity (39).

During aging a progressive degeneration of the thymus occurs leading to a substantial loss of its capacity to generate and export new T-cells (40, 41). Throughout middle age thymic epithelial space and the functional unit of thymopoiesis (and thus the production of T-cells) decline by approximately 3% per year until the age of 45 when only an irrelevant level of functional thymic tissue remains. The total number of T-cells in the periphery nevertheless is unchanged and peripheral mechanisms of T-cell renewal have to compensate for progressive thymic failure (42–44). Parallel to the overall reduction of thymic T-cell output the production of thymically derived T_{regs} decreases with age (45). Alternative mechanisms such as increased surveillance of T_{regs} in the elderly (46) as well as peripheral T_{reg} generation may compensate for the loss of thymic function to maintain a sufficient T_{reg} pool (see **Figure 1**). Indeed, numerous studies indicate a possible conversion of non-regulatory CD4⁺CD25⁻ T-cells into T_{regs} *in vitro* and *in vivo* (47, 48). Moreover, mouse studies showed that peripheral self-antigen-driven proliferation of T_{regs} is a thymus-independent mechanism to maintain T_{regs} (49–51). The proportion of conventional T-cells differentiating into T_{regs} as well as the relative contribution of homeostatic T_{reg} proliferation to the overall T_{reg} pool in elderly individuals are unknown.

Peripheral mechanisms of T-cell renewal (particularly homeostatic expansion of existing T_{regs}) are probably not infinite. Normally, T-cells proliferate beyond the seventh decade of life. Thereafter, telomere lengths are usually contracted to levels known as the "Hayflick limit". At this stage, non-regulatory T-cells do not proliferate anymore and undergo phenotypical and functional changes such as down-regulation of CD28 and acquisition of cytotoxic potential (4, 52, 53). Due to the fact that T_{regs} display even shorter telomeres than non-regulatory T-cells, it is conceivable that peripherally proliferating T_{regs} reach the "Hayflick limit" even earlier (54). Impaired T_{reg} homeostasis may then result in immune dysfunction with increased risk of immune-mediated disorders. In addition to the shortened telomere length, TCR diversity is also contracted to at least 100-fold in elderly individuals (55). This has been explained by the observation that homeostatic proliferation of T-cells is antigen dependent. Thus, T-cells with a high affinity TCR to self-antigens or antigens deriving from chronic virus infections have a survival advantage over other T-cells (42, 56). Given that similar mechanisms drive peripheral proliferation of non-regulatory T-cells and T_{regs} , a reduction of T_{reg} TCR diversity (with a skew to certain antigens) can be expected in the elderly. Consequently, T_{regs} could mediate increased immunosuppression in response to specific self- (even if transformed) or viral antigens with increased incidence of malignancies and infections in the elderly. At the same time the reduced diversity of T_{regs} could result in decreased protection from autoimmunity (3).

DEVELOPMENT AND CELLULAR SENESCENCE OF T_{REGS} FROM NAÏVE TO MEMORY CELL STATUS

Similar to the developmental stages known for non-regulatory Tcells (development form CD45RA⁺ naïve to CD45RO⁺ memory and finally to CD28⁻ memory effector T-cells), different cellular subsets of T_{regs} were also observed. In humans, CD4⁺ foxP3⁺ T_{regs} may have either a "naïve-like" phenotype characterized by the expression of CD25⁺CD45RA⁺ or a CD25^{hi}CD45RO⁺ "memorylike" phenotype (54). In mice, naïve-like T_{regs} were characterized by the expression of CD25, CD62L, and CCR7 and by preferential homing to antigen-draining lymph nodes, where they

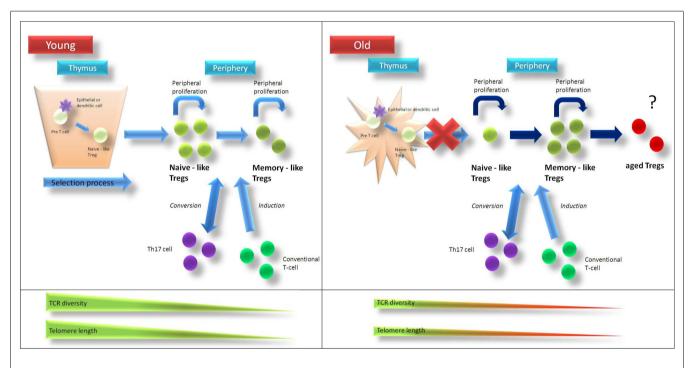


FIGURE 1 | Age-related changes of T_{reg} **homeostasis**. In young individuals T_{regs} are generated in the thymus and are released as "naïve-like" T_{regs} into circulation. After antigen-contact, T_{regs} develop into a "memory-like" phenotype. T_{reg} homeostasis is supported by homeostatic proliferation of "naïve-like" and "memory-like" T_{regs} as well as conversion of non-regulatory T-cells into T_{regs} . Telomere length and T-cell receptor diversity is higher in naïve-like compared to memory-like T_{regs} . After puberty thymic function is

progressively lost and in aged individuals homeostatic proliferation of existing $T_{\rm regs}$ as well as conversion of non-regulatory T-cells into $T_{\rm regs}$ compensate for thymic failure to maintain $T_{\rm reg}$ pool. Due to ongoing homeostatic replication telomere length and T-cell receptor diversity of $T_{\rm regs}$ from elderly people are contracted compared to those from young individuals. Recurrent stimulation of $T_{\rm regs}$ might then lead to a status of "terminal-differentiation" with altered phenotype and function. $T_{\rm reg}$ *regulatory T-cell*, TCR \ldots *T-cell receptor*.

were able to inhibit the induction of inflammation (10, 57, 58). Memory/effector-like T_{regs} (characterized by expression of CD29, CD44, ICOS, and LFA-1) migrated into non-lymphogenic tissues and sites of inflammation; a local down-regulation of immune reactions was shown (57, 58).

In humans, the highest prevalence of naïve-like T_{regs} were found in cord blood and it was assumed that these naïve-like T_{regs} are produced in the thymus (20, 59). The prevalence of memory-like Tregs increases rapidly during childhood and it was demonstrated that these memory-like Tregs have shorter telomeres and a lower content of TCR excision circles (Trecs) compared to naïve-like T_{regs} reflecting a longer replicative history (54). The mechanisms mediating the transition of a naïve-like Treg into a memory-like phenotype still have to be explored; however, it is believed that antigen experienced dendritic cells migrating to secondary lymphoid tissues are involved. Tregs proliferate upon stimulation with autologous immature and mature dendritic cells (54, 60). A low surface expression of CD45RB on memory-like Tregs further supports the hypothesis of an antigen-driven development of naïve-like Tregs. CD45RB is normally down-regulated after repeated antigen-contact (61).

Human adult peripheral blood usually contains both, naïvelike and memory-like T_{regs} . Parallel to the reduction of total naïve T-cells, the quantity of naïve-like T_{regs} declines with age whereas the prevalence of memory-like T_{regs} increases (29, 62). The total pool of circulating T_{regs} ; however, remains unchanged as mentioned above (19). As naïve-like T_{regs} exhibit a higher proliferative potential *in vitro* compared to memory-like T_{regs} it can be expected that the capacity of the immune system to downregulate abnormal immune responses declines with age (54).

END-DIFFERENTIATED T_{REGS} AND ASPECTS OF T_{REG} SENESCENCE

Replicative senescence of T-cells is a prominent feature of aging resulting from homeostatic proliferation and repetitive antigen exposure (63). The most important phenotypic feature of senescent T-cells is the loss of the type I transmembrane protein CD28, a major co-stimulatory molecule (64). From the functional perspective, non-regulatory CD28⁻ T-cells produce large amounts of interferon γ , perforin, and granzyme B, providing them with the ability to lyse target cells (65). Another feature of CD28⁻ T-cells is their longevity and persistence that can be explained by defects in the apoptotic pathway with upregulation of bcl-2 and Fas-associated death domain like IL-2-converting enzymelike inhibitory protein (FLIP) (66, 67). Terminally differentiated T-cells also acquire new stimulatory receptors including killer cell immunoglobulin-like receptors (KIRs) and Toll-like receptors

REFERENCES

- Danke NA, Koelle DM, Yee C, Beheray S, Kwok WW. Autoreactive T cells in healthy individuals. J Immunol (2004) 172(10):5967–72.
- Stahl EA, Raychaudhuri S, Remmers EF, Xie G, Eyre S, Thomson BP, et al. Genome-wide association study meta-analysis identifies seven new rheumatoid arthritis risk loci. Nat Genet (2010) 42(6):508–14. doi:10. 1038/ng.582
- Dejaco C, Duftner C, Grubeck-Loebenstein B, Schirmer M. Imbalance of regulatory T cells in human autoimmune diseases. *Immunology* (2006) 117(3):289– 300. doi:10.1111/j.1365-2567.2005. 02317.x
- Dejaco C, Duftner C, Schirmer M. Are regulatory T-cells linked with aging? *Exp Gerontol* (2006) 41(4):339–45. doi:10.1016/j.exger. 2006.01.008

(TLRs) (68, 69). Thus, activation of CD4⁺CD28⁻ T-cells no longer depends on professional antigen-presenting cells, rather it is promoted by stress molecules as well as bacterial and/or viral products (65). The frequency of terminally differentiated CD4⁺CD28⁻ Tcells is increased in old individuals as well as in younger patients with autoimmune diseases such as rheumatoid arthritis or spondyloarthritis (70). Given that T_{regs} proliferate in the periphery to maintain the total T_{regs} may undergo terminal-differentiation as well.

Interestingly, a proportion of T_{regs} from aged mice showed decreased expression of CD25 (46, 71). These CD25^{low} T_{regs} occurred predominantly in the spleen (24) but had comparable functional properties to CD25⁺ T_{regs}. A similar CD4⁺CD25⁻foxP3⁺ T_{reg} population has been observed in SLE patients. SLE patients are known to have a prematurely aged immune system (72) with accumulation of CD28- T-cells. A detailed characterization of CD4+CD25-FoxP3+ Trees regarding the expression of naïve/memory T-cell markers or determination of telomere lengths was unfortunately not performed. Further evidence for the occurrence of Treg senescence was found in a study on healthy aged individuals reporting the occurrence of a CD8⁺CD25⁺ T_{reg} population lacking CD28 expression. These regulatory cells shared phenotypic and functional features with $CD4^+$ T_{regs} from the same population (73). The occurrence and possible characteristics of terminally differentiated CD4⁺ T_{regs} is an interesting issue that has to be investigated by future studies.

CONCLUSION

Accumulating evidence suggests age-associated changes of T_{reg} prevalence and/or T_{reg} function. Due to involution of thymus after puberty peripheral mechanisms including homeostatic proliferation of T_{regs} or conversion of non-regulatory T-cells into T_{regs} compensate for the decreasing generation of new T_{reg} cells. However, these peripheral mechanisms are limited; this leads to altered composition of the T_{reg} pool. Age-related changes of T_{regs} are suspected to increase the risk of autoimmunity, cancer, and infections in the elderly; however, the exact mechanisms are still poorly understood. Current studies are limited by the difficult identification of human T_{regs} and the uncertainty whether circulating T_{regs} reflect the total T_{reg} pool or a cellular subset only. Future studies are required to investigate cellular senescence of T_{regs} and possible therapeutic approaches targeting T_{regs} in aged individuals.

ACKNOWLEDGMENTS

We thank Prof. Dr. Winfried Graninger (Medical University of Graz) for his critical review of the manuscript.

- Goronzy JJ, Weyand CM. Immune aging and autoimmunity. *Cell Mol Life Sci* (2012) 69(10):1615–23. doi:10.1007/ s00018-012-0970-0
- Hohensinner PJ, Goronzy JJ, Weyand CM. Telomere dysfunction, autoimmunity and aging. *Aging Dis* (2011) 2(6):524–37.
- Piccirillo CA, Letterio JJ, Thornton AM, McHugh RS, Mamura M, Mizuhara H, et al. CD4(+)CD25(+)

regulatory T cells can mediate suppressor function in the absence of transforming growth factor beta1 production and responsiveness. *J Exp Med* (2002) **196**(2):237–46. doi:10.1084/jem. 20020590

 Shevach EM. Regulatory/suppressor T cells in health and disease. Arthritis Rheum (2004) 50(9):2721–4. doi:10.1002/art.20500

- 9. Sakaguchi S. Naturally arising Foxp3-expressing CD25+CD4+ regulatory T cells in immunological tolerance to self and non-self. *Nat Immunol* (2005) **6**(4):345–52. doi:10.1038/ni1178
- Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4+CD25+ regulatory T cells. *Nat Immunol* (2003) 4(4):330–6. doi:10.1038/ ni904
- Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, Whitesell L, et al. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat Genet* (2001) 27(1):20– 1. doi:10.1038/83713
- 12. Tran DQ, Ramsey H, Shevach EM. Induction of FOXP3 expression in naive human CD4+FOXP3 T cells by T-cell receptor stimulation is transforming growth factor-beta dependent but does not confer a regulatory phenotype. *Blood* (2007) **110**(8):2983–90. doi: 10.1182/blood-2007-06-094656
- Fessler J, Felber A, Duftner C, Dejaco C. Therapeutic potential of regulatory T cells in autoimmune disorders. *BioDrugs* (2013) 27(4):281–91. doi:10.1007/s40259-013-0026-5
- Thornton AM, Piccirillo CA, Shevach EM. Activation requirements for the induction of CD4+CD25+ T cell suppressor function. *Eur J Immunol* (2004) **34**(2):366–76. doi: 10.1002/eji.200324455
- Qin S, Cobbold SP, Pope H, Elliott J, Kioussis D, Davies J, et al. "Infectious" transplantation tolerance. *Science* (1993) 259(5097):974–7. doi:10.1126/science.8094901
- Brusko TM, Putnam AL, Bluestone JA. Human regulatory T cells: role in autoimmune disease and therapeutic opportunities. *Immunol Rev* (2008) 223:371–90. doi:10.1111/j. 1600-065X.2008.00637.x
- Ye J, Huang X, Hsueh EC, Zhang Q, Ma C, Zhang Y, et al. Human regulatory T cells induce Tlymphocyte senescence. *Blood* (2012) **120**(10):2021–31. doi:10. 1182/blood-2012-03-416040
- Ohkura N, Sakaguchi S. Regulatory T cells: roles of T cell receptor for their development and function. *Semin Immunopathol* (2010) 32(2):95–106. doi:10.1007/s00281-010-0200-5
- Raynor J, Lages CS, Shehata H, Hildeman DA, Chougnet CA. Homeostasis and function of regulatory T cells in aging. *Curr*

Opin Immunol (2012) **24**(4):482–7. doi:10.1016/j.coi.2012.04.005

- 20. Wing K, Ekmark A, Karlsson H, Rudin A, Suri-Payer E. Characterization of human CD25+ CD4+ T cells in thymus, cord and adult blood. *Immunology* (2002) 106(2):190–9. doi:10.1046/j.1365-2567.2002.01412.x
- Schonland SO, Zimmer JK, Lopez-Benitez CM, Widmann T, Ramin KD, Goronzy JJ, et al. Homeostatic control of Tcell generation in neonates. *Blood* (2003) 102(4):1428–34. doi:10.1182/blood-2002-11-3591
- 22. Sullivan KE, McDonald-McGinn D, Zackai EH. CD4(+) CD25(+) T-cell production in healthy humans and in patients with thymic hypoplasia. *Clin Diagn Lab Immunol* (2002) 9(5):1129–31.
- Zhao L, Sun L, Wang H, Ma H, Liu G, Zhao Y. Changes of CD4+CD25+Foxp3+ regulatory T cells in aged Balb/c mice. J Leukoc Biol (2007) 81(6):1386–94. doi:10. 1189/jlb.0506364
- 24. Lages CS, Suffia I, Velilla PA, Huang B, Warshaw G, Hildeman DA, et al. Functional regulatory T cells accumulate in aged hosts and promote chronic infectious disease reactivation. *J Immunol* (2008) 181(3):1835–48.
- 25. Agius E, Lacy KE, Vukmanovic-Stejic M, Jagger AL, Papageorgiou AP, Hall S, et al. Decreased TNFalpha synthesis by macrophages restricts cutaneous immunosurveillance by memory CD4+ T cells during aging. J Exp Med (2009) 206(9):1929–40. doi:10.1084/jem. 20090896
- 26. Green VL, Michno A, Stafford ND, Greenman J. Increased prevalence of tumour infiltrating immune cells in oropharyngeal tumours in comparison to other subsites: relationship to peripheral immunity. *Cancer Immunol Immunother* (2013) 62(5):863–73. doi:10.1007/s00262-013-1395-9
- 27. Ino Y, Yamazaki-Itoh R, Shimada K, Iwasaki M, Kosuge T, Kanai Y, et al. Immune cell infiltration as an indicator of the immune microenvironment of pancreatic cancer. *Br J Cancer* (2013) 108(4):914–23. doi:10.1038/bjc. 2013.32
- 28. Sun L, Hurez VJ, Thibodeaux SR, Kious MJ, Liu A, Lin P, et al. Aged regulatory T cells protect from autoimmune inflammation despite reduced STAT3 activation and decreased constraint of IL-17 producing T cells. Aging Cell

(2012) **11**(3):509–19. doi:10.1111/j. 1474-9726.2012.00812.x

- Hwang KA, Kim HR, Kang I. Aging and human CD4(+) regulatory T cells. *Mech Ageing Dev* (2009) 130(8):509–17. doi:10.1016/j.mad
- 30. Tsaknaridis L, Spencer L, Culbertson N, Hicks K, LaTocha D, Chou YK, et al. Functional assay for human CD4+CD25+ Treg cells reveals an age-dependent loss of suppressive activity. J Neurosci Res (2003) 74(2):296–308. doi:10.1002/ jnr.10766
- 31. Gregg R, Smith CM, Clark FJ, Dunnion D, Khan N, Chakraverty R, et al. The number of human peripheral blood CD4+ CD25high regulatory T cells increases with age. *Clin Exp Immunol* (2005) 140(3):540–6. doi:10.1111/j.1365-2249.2005.02798.x
- 32. Sharma S, Dominguez AL, Lustgarten J. High accumulation of T regulatory cells prevents the activation of immune responses in aged animals. J Immunol (2006) 177(12):8348–55.
- 33. Hurez V, Daniel BJ, Sun L, Liu AJ, Ludwig SM, Kious MJ, et al. Mitigating age-related immune dysfunction heightens the efficacy of tumor immunotherapy in aged mice. *Cancer Res* (2012) 72(8):2089–99. doi:10.1158/0008-5472.CAN-11-3019
- 34. Watanabe N, Wang YH, Lee HK, Ito T, Wang YH, Cao W, et al. Hassall's corpuscles instruct dendritic cells to induce CD4+CD25+ regulatory T cells in human thymus. *Nature* (2005) **436**(7054):1181–5. doi:10.1038/nature03886
- Paust S, Cantor H. Regulatory T cells and autoimmune disease. *Immunol Rev* (2005) **204**:195–207. doi:10. 1111/j.0105-2896.2005.00247.x
- 36. Nakamura K, Kitani A, Fuss I, Pedersen A, Harada N, Nawata H, et al. TGF-beta 1 plays an important role in the mechanism of CD4+CD25+ regulatory T cell activity in both humans and mice. *J Immunol* (2004) 172(2):834–42.
- 37. Jordan MS, Boesteanu A, Reed AJ, Petrone AL, Holenbeck AE, Lerman MA, et al. Thymic selection of CD4+CD25+ regulatory T cells induced by an agonist self-peptide. *Nat Immunol* (2001) 2(4):301–6. doi:10.1038/86302
- Ouyang W, Beckett O, Ma Q, Li MO. Transforming growth factorbeta signaling curbs thymic negative selection promoting regulatory T cell development. *Immunity* (2010) 32(5):642–53. doi:10.1016/j. immuni.2010.04.012

- 39. Sekiya T, Kashiwagi I, Yoshida R, Fukaya T, Morita R, Kimura A, et al. Nr4a receptors are essential for thymic regulatory T cell development and immune homeostasis. *Nat Immunol* (2013) 14(3):230–7. doi: 10.1038/ni.2520
- Simpson JG, Gray ES, Beck JS. Age involution in the normal human adult thymus. *Clin Exp Immunol* (1975) 19(2):261–5.
- Berzins SP, Uldrich AP, Sutherland JS, Gill J, Miller JF, Godfrey DI, et al. Thymic regeneration: teaching an old immune system new tricks. *Trends Mol Med* (2002) 8(10):469–76. doi:10.1016/ S1471-4914(02)02415-2
- Goronzy JJ, Weyand CM. Thymic function and peripheral T-cell homeostasis in rheumatoid arthritis. *Trends Immunol* (2001) 22(5):251–5. doi:10.1016/S1471-4906(00)01841-X
- 43. Mackall CL, Bare CV, Granger LA, Sharrow SO, Titus JA, Gress RE. Thymic-independent T cell regeneration occurs via antigen-driven expansion of peripheral T cells resulting in a repertoire that is limited in diversity and prone to skewing. J Immunol (1996) 156(12):4609–16.
- 44. Steinmann GG, Klaus B, Muller-Hermelink HK. The involution of the ageing human thymic epithelium is independent of puberty. A morphometric study. *Scand J Immunol* (1985) 22(5):563–75. doi:10.1111/j.1365-3083.1985. tb01916.x
- 45. Chiu BC, Stolberg VR, Zhang H, Chensue SW. Increased Foxp3(+) Treg cell activity reduces dendritic cell co-stimulatory molecule expression in aged mice. *Mech Ageing Dev* (2007) **128**(11–12):618–27. doi:10.1016/j.mad.2007.09.002
- 46. Chougnet CA, Tripathi P, Lages CS, Raynor J, Sholl A, Fink P, et al. A major role for Bim in regulatory T cell homeostasis. *J Immunol* (2011) **186**(1):156–63. doi:10.4049/ jimmunol.1001505
- Curotto de Lafaille MA, Lino AC, Kutchukhidze N, Lafaille JJ. CD25-T cells generate CD25+Foxp3+ regulatory T cells by peripheral expansion. *J Immunol* (2004) 173(12):7259–68.
- 48. Wan YY, Flavell RA. Identifying Foxp3-expressing suppressor T cells with a bicistronic reporter. *Proc Natl Acad Sci U S A* (2005) **102**(14):5126–31. doi:10. 1073/pnas.0501701102
- 49. Walker LS, Chodos A, Eggena M, Dooms H, Abbas AK.

Antigen-dependentprolifera-tionofCD4+CD25+regu-latoryTcellsinvivo.JExpMed(2003)198(2):249-58.doi:10.1084/jem.20030315

- Cozzo C, Larkin J III, Caton AJ. Cutting edge: self-peptides drive the peripheral expansion of CD4+CD25+ regulatory T cells. *J Immunol* (2003) 171(11):5678–82.
- Fisson S, Darrasse-Jeze G, Litvinova E, Septier F, Klatzmann D, Liblau R, et al. Continuous activation of autoreactive CD4+ CD25+ regulatory T cells in the steady state. J Exp Med (2003) 198(5):737–46. doi:10. 1084/jem.20030686
- Effros RB. From Hayflick to Walford: the role of T cell replicative senescence in human aging. *Exp Gerontol* (2004) **39**(6):885–90. doi: 10.1016/j.exger.2004.03.004
- Goronzy JJ, Lee WW, Weyand CM. Aging and T-cell diversity. *Exp Gerontol* (2007) 42(5):400–6. doi: 10.1016/j.exger.2006.11.016
- 54. Valmori D, Merlo A, Souleimanian NE, Hesdorffer CS, Ayyoub M. A peripheral circulating compartment of natural naive CD4 Tregs. J Clin Invest (2005) 115(7):1953–62. doi:10.1172/JCI23963
- Naylor K, Li G, Vallejo AN, Lee WW, Koetz K, Bryl E, et al. The influence of age on T cell generation and TCR diversity. *J Immunol* (2005) 174(11):7446–52.
- 56. Weiss L, Donkova-Petrini V, Caccavelli L, Balbo M, Carbonneil C, Levy Y. Human immunodeficiency virus-driven expansion of CD4+CD25+ regulatory T cells, which suppress HIV-specific CD4 T-cell responses in HIV-infected patients. *Blood* (2004) **104**(10):3249–56. doi:10.1182/blood-2004-01-0365
- 57. Huehn J, Siegmund K, Lehmann JC, Siewert C, Haubold U, Feuerer

M, et al. Developmental stage, phenotype, and migration distinguish naive- and effector/memorylike CD4+ regulatory T cells. *J Exp Med* (2004) **199**(3):303–13. doi:10. 1084/jem.20031562

- Fontenot JD, Rasmussen JP, Williams LM, Dooley JL, Farr AG, Rudensky AY. Regulatory T cell lineage specification by the forkhead transcription factor foxp3. *Immunity* (2005) 22(3): 329–41. doi:10.1016/j.immuni. 2005.01.016
- Takahata Y, Nomura A, Takada H, Ohga S, Furuno K, Hikino S, et al. CD25+CD4+ T cells in human cord blood: an immunoregulatory subset with naive phenotype and specific expression of forkhead box p3 (Foxp3) gene. *Exp Hematol* (2004) 32(7):622–9. doi:10.1016/j.exphem. 2004.03.012
- Cupedo T, Nagasawa M, Weijer K, Blom B, Spits H. Development and activation of regulatory T cells in the human fetus. *Eur J Immunol* (2005) 35(2):383–90. doi:10.1002/ eij.200425763
- 61. Salmon M, Pilling D, Borthwick NJ, Viner N, Janossy G, Bacon PA, et al. The progressive differentiation of primed T cells is associated with an increasing susceptibility to apoptosis. *Eur J Immunol* (1994) 24(4):892–9. doi:10.1002/ eji.1830240417
- 62. Seddiki N, Santner-Nanan B, Tangye SG, Alexander SI, Solomon M, Lee S, et al. Persistence of naive CD45RA+ regulatory T cells in adult life. *Blood* (2006) **107**(7):2830–8. doi:10.1182/blood-2005-06-2403
- 63. Vallejo AN. CD28 extinction in human T cells: altered functions and the program of T-cell senescence. *Immunol Rev* (2005) **205**:158–69. doi:10.1111/j.0105-2896.2005. 00256.x

- 64. Linsley PS, Ledbetter JA. The role of the CD28 receptor during T cell responses to antigen. Annu Rev Immunol (1993) 11:191–212. doi:10.1146/annurev. iy.11.040193.001203
- Nakajima T, Schulte S, Warrington KJ, Kopecky SL, Frye RL, Goronzy JJ, et al. T-cell-mediated lysis of endothelial cells in acute coronary syndromes. *Circulation* (2002) 105(5):570–5. doi:10.1161/hc0502. 103348
- 66. Schirmer M, Vallejo AN, Weyand CM, Goronzy JJ. Resistance to apoptosis and elevated expression of Bcl-2 in clonally expanded CD4+CD28-T cells from rheumatoid arthritis patients. *J Immunol* (1998) 161(2):1018–25.
- Vallejo AN, Schirmer M, Weyand CM, Goronzy JJ. Clonality and longevity of CD4+CD28null T cells are associated with defects in apoptotic pathways. *J Immunol* (2000) 165(11):6301–7.
- 68. Namekawa T, Snyder MR, Yen JH, Goehring BE, Leibson PJ, Weyand CM, et al. Killer cell activating receptors function as costimulatory molecules on CD4+CD28null T cells clonally expanded in rheumatoid arthritis. J Immunol (2000) 165(2):1138–45.
- 69. Raffeiner B, Dejaco C, Duftner C, Kullich W, Goldberger C, Vega SC, et al. Between adaptive and innate immunity: TLR4mediated perforin production by CD28null T-helper cells in ankylosing spondylitis. *Arthritis Res Ther* (2005) 7(6):R1412–20. doi:10.1186/ ar1840
- 70. Dejaco C, Duftner C, Klauser A, Schirmer M. Altered T-cell subtypes in spondyloarthritis, rheumatoid arthritis and polymyalgia rheumatica. *Rheumatol Int* (2010) **30**(3):297–303. doi:10.1007/s00296-009-0949-9

- Nishioka T, Shimizu J, Iida R, Yamazaki S, Sakaguchi S. CD4+CD25+Foxp3+ T cells and CD4+CD25-Foxp3+ T cells in aged mice. *J Immunol* (2006) 176(11):6586–93.
- 72. Bonelli M, Savitskaya A, Steiner CW, Rath E, Smolen JS, Scheinecker C. Phenotypic and functional analysis of CD4+ CD25- Foxp3+ T cells in patients with systemic lupus erythematosus. J Immunol (2009) 182(3):1689–95.
- 73. Simone R, Zicca A, Saverino D. The frequency of regulatory CD3+CD8+CD28- CD25+ T lymphocytes in human peripheral blood increases with age. J Leukoc Biol (2008) 84(6):1454–61. doi:10. 1189/jlb.0907627

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 28 March 2013; accepted: 22 July 2013; published online: 06 August 2013.

Citation: Fessler J, Ficjan A, Duftner C and Dejaco C (2013) The impact of aging on regulatory T-cells. Front. Immunol. 4:231. doi: 10.3389/fimmu.2013.00231 This article was submitted to Frontiers in T Cell Biology, a specialty of Frontiers in Immunology.

Copyright © 2013 Fessler, Ficjan, Duftner and Dejaco. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.