



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

Multiple Sclerosis

Mechanisms of Disease and Strategies for Myelin and Axonal Repair



Hernan Nicolas Lemus, MD^a, Arthur E. Warrington, PhD^{a,*},
Moses Rodriguez, MD^{a,b}

KEY WORDS

- Multiple sclerosis • Autoimmune • Immune • Axon • Remyelination

KEY POINTS

- Multiple sclerosis is an inflammatory demyelinating disease of the central nervous system with a variety of presentations and unclear pathogenesis.
- Multiple sclerosis has been associated with the term *autoimmunity* as a surrogate for pathogenesis.
- Still, multiple sclerosis is an organ-specific disease with immune-mediated myelin destruction.
- Understanding the complex etiology of multiple sclerosis (autoimmune induced, virus induced, or immune mediated) and the importance of axon integrity is critical for clinicians who treat the disease.

INTRODUCTION

Multiple sclerosis (MS) is an inflammatory demyelinating disease of the central nervous system (CNS) with a variety of clinical presentations. The profound heterogeneity of MS is not limited to the symptoms but to neuroradiologic and histologic appearances of lesions and response to therapy.¹ As expected, the pathogenesis of MS is controversial, and there is no effective treatment that halts the neuro-axonal damage

Financial Support: Authors are supported by grants from the Minnesota Partnership Award for Biotechnology and Medical Genomics, Mayo Clinic Center for Multiple Sclerosis and Autoimmune Neurology (CMSAN) through a gift from Dr and Mrs Moon Park.

Competing Interests: Patents for human antibodies that promote remyelination and CNS repair are issued and owned by Mayo Clinic. A.E. Warrington and M. Rodriguez have a potential conflict of interest.

^a Department of Neurology, Mayo Clinic, 200 First Street Southwest, Rochester, MN 55905, USA; ^b Department of Immunology, Mayo Clinic, 200 First Street Southwest, Rochester, MN 55905, USA

* Corresponding author. Department of Neurology, Mayo Clinic, Guggenheim Building 401, 200 First Street Southwest, Rochester, MN 55905.

E-mail address: Warrington.Arthur@mayo.edu

or promotes remyelination. As a leading cause of disability, MS affects 400,000 people in the United States and 2.5 million of people worldwide.²

The demyelinating plaque, the main pathologic hallmark of MS, contains a prominent immunologic response dominated by CD8+ and CD4+ T cells.^{1,3} Moreover, the presence of oligoclonal bands in the cerebrospinal fluid of MS patients shows the presence of immunoglobulin-producing B cells suggesting their participation in the pathogenesis of the disease.⁴ These findings suggest that MS is an immune-mediated disorder involving multiple antigens of the CNS^{5,6} and, further, that MS is an autoimmune disease of the CNS. Understanding the mechanism of MS is essential to elucidate possible strategies to repair myelin and axonal structures.

AUTOIMMUNITY VERSUS IMMUNE-MEDIATED DEMYELINATION

Several criteria have been established to determine whether a disease can be classified as *autoimmune*.^{7,8} First, an autoantigen must be present in all patients with a proven immune response directed against it. Second, one must identify autoantibodies within a lesion or serum of patients with a direct correlation to disease activity or observed clinical improvement after immunosuppressive treatment. In systemic lupus erythematosus (SLE), a well-characterized autoimmune disorder, the presentation of autoantigens by T cells promotes antibody formation and hence, the clinical manifestations.⁹ It is also critical to reproduce the clinical and histopathologic aspects of the human disease after administration of the autoantibody or autoantigen within an animal. For example, when transferring anti-DNA antibodies to naïve recipient mice, there is an immunologic reaction against glomerular antigens leading to nephropathy similar to that seen in SLE.¹⁰ Diseases like SLE have an experimental-based extensive literature that meets all the criteria and proves the role of autoimmunity in the pathophysiology.

MS is also an organ-specific disease (the brain and spinal cord) with immune-mediated myelin destruction. Nevertheless, after extensive research, confirmation of a specific auto-antigen in MS is lacking. The absence/presence of an infectious agent in patients with MS has also not been proven. Other organ-specific immune-mediated diseases such as herpes encephalitis have a persistent exogenous antigen (in this case, the herpes virus) that resides in the CNS and drives the development of acute inflammation and necrotizing lesions.¹¹ However, the absence of any consistent viral or bacterial antigen in MS patients suggests the presence of an autoantigen that drives this disease. Antibodies directed against CNS myelin proteins, lipids, and carbohydrates (possible candidates as autoantigens) can be identified in the tissue and serum of patients with MS.

Extensive literature is devoted to identifying antibodies against the myelin oligodendrocyte glycoprotein (MOG) in MS patients with inconsistent results. Enzyme-linked immunosorbent assay-based binding studies using a synthetic MOG peptide to identify antibodies found an increase in the frequency of MOG-binding IgGs in patients with MS compared with controls.^{12,13} In contrast, other studies using enzyme-linked immunosorbent assay binding to the recombinant human immunoglobulin domain of MOG showed no difference in the levels of bound antibodies in patients with MS or healthy controls.¹⁴ When using other techniques, such as immunoblot to detect antibodies directed against the recombinant human immunoglobulin domain of MOG in patients with MS, the results are inconsistent.^{15,16} Antibodies that bind to other antigens such as alpha-B-crystallin, alu repeats, myelin basic protein, and myelin-associated glycoprotein have been reported but not rigorously studied.^{17–20} Despite several published attempts to detect and quantify antibodies directed against

myelin and nonmyelin antigens in patients with MS, there is no consensus. This lack of a self-directed immune response producing antibodies argues against the hypothesis of MS as an autoimmune disease (**Fig. 1**).

On the other hand, neuromyelitis optica (NMO), an autoimmune astrocytopathy, is a CNS disease driven by the presence of antibodies against aquaporin 4 (AQP4), a plasma membrane-based water-transporting protein (see **Fig. 1**). AQP4 is the most abundant water channel in the brain, expressed primarily in astrocytes and highly involved in neuroexcitation.²¹ The discovery of pathogenic immunoglobulin G directed to AQP4 in almost 70% of the patients with NMO was the first evidence that this disease was an inflammatory autoimmune disease of the CNS.^{22,23} The pathogenicity of the anti-AQP4 IgG in NMO patients has been reproduced in vivo and their presence confirmed in pathologic lesions.^{24,25}

IS MULTIPLE SCLEROSIS AN AUTOIMMUNE DISEASE?

When comparing MS with other autoimmune diseases like neuromyelitis optica, it is clear that MS does not meet the full definition of autoimmune disease (**Table 1**). Although NMO meets 6 of 8 autoimmune disease criteria, MS meets only 2. Although there are some data for several of the other criteria, the evidence is controversial. For example, multiple studies focused on measuring the level of precursor T cells before and then during clinical exacerbations. However, none of these studies found a difference from the prevalence in healthy controls or are not confirmed using other patient populations.²⁶ It is clear that the data are weak to make a definitive conclusion that MS is an autoimmune disease.

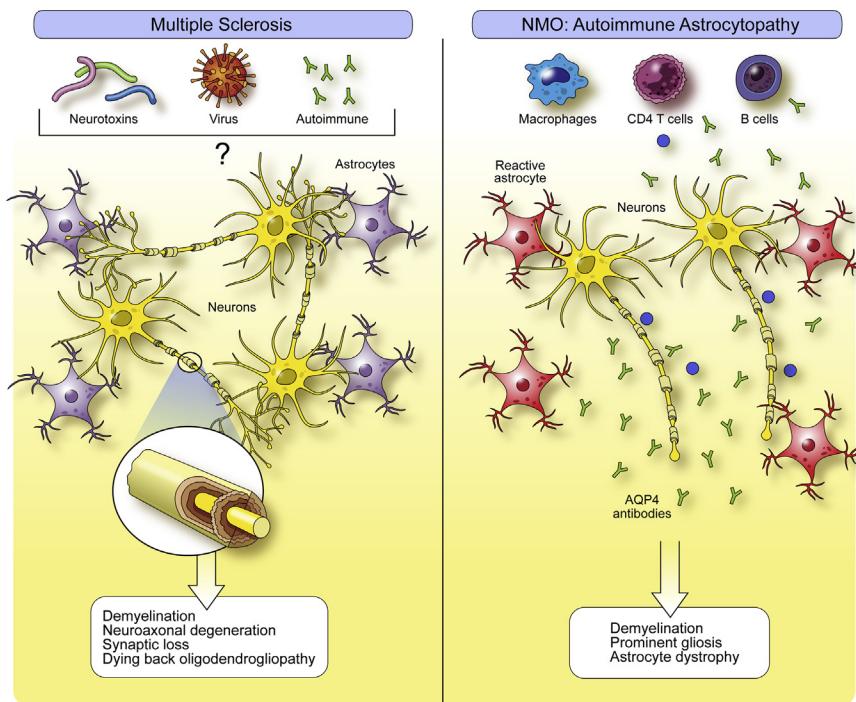


Fig. 1. Autoimmune response hypothesis for MS.

Table 1
Revised criteria for a disease to be considered autoimmune: neuromyelitis optica and multiple sclerosis

Criteria for Autoimmune Disease ⁷	NMO	MS
Immune response to a precise autoantigen in all patients	aquaporin 4 ^{22,23}	Multiple antigens have been described, not present in all patients ^{12,13,17–20}
Lesion reproducibility after administration of autoantibody or T cells	Exacerbation of EAE model after adoptive transfer of neuromyelitis optica Abs ²⁷	
Animal: induction of lesion by antigen immunization		EAE model: induced by myelin oligodendrocyte glycoprotein, proteolipid protein, myelin basic protein ²⁸ and reactivated CD4+ T cells ²⁹
Autoantibody or T cell isolation from lesion or serum	aquaporin 4 antibodies ^{27,30}	
Autoantibody titers or T-cell levels associated with disease activity	Higher antibody titers during relapse than during remission ³¹	
Autoimmune disorders or autoantigens associated with the disease	Sjogren syndrome, SLE ³²	No association in population-based cohort studies ³³
Immune absorption with purified autoantigen abrogates pathogenic autoantibody or T cell		
Reduction of autoantibody or T cell associated with clinical improvement	Plasma exchange ³⁴	Plasma exchange ^{35,36}

Abbreviations: EAE, experimental autoimmune encephalomyelitis; MBP, myelin basic protein; PLP, proteolipid protein.

Adapted from Paul WE, Schwartz RS, Datta SK. Autoimmunity and autoimmune diseases. In Fundamental immunology. New York: Raven Press; 1989. p. 819–66; and Rodriguez M, Warrington AE, Pease LR. Invited article: human natural autoantibodies in the treatment of neurologic disease. Neurology 2009;72(14):1269–76.

IS MULTIPLE SCLEROSIS A VIRUS-INDUCED DISEASE?

Viral microorganisms are postulated by many investigators to be causal agents of MS. The idea is plausible because other demyelinating diseases of the CNS like progressive multifocal leukoencephalopathy are caused by a virus: the JC virus in immune-compromised individuals.³⁷ Recently, much attention has focused on Epstein-Barr virus (EBV) because EBV antigens are expressed at significantly higher levels in the cerebrospinal fluid and serum of some MS patients compared with controls.³⁸ However, studies show an absence of intrathecal anti-EBV antibody synthesis in nearly 93% of patients with MS, which argues against this hypothesis.³⁹ Hence, the association of EBV with adult MS is not well established, and its role in pathogenesis remains to be determined. Other viral agents like varicella zoster and rubella are considered as possible risk factors but not causal agents.^{40,41} Of note, vitamin D

deficiency has been linked as a causal agent in relapsing remitting MS. Vitamin D has a role in regulating immune function. Low levels of this vitamin could produce an immune-deficient state against viral agents. Although clinical trials and observational studies using high levels of vitamin D have only shown modest reductions in the levels of interleukin 17, it has not been seen in other inflammatory markers.^{42,43} The fact that high dose treatment of vitamin D in MS patients has scant effect on the overall course of the disease does not support this theory.

Demyelinating disease nearly identical to human MS can be consistently established in animals using well-characterized viral agents. Theiler's murine encephalomyelitis virus (TMEV) is a mouse enteric pathogen that belongs to the picornavirus family. It produces a chronic progressive demyelinating condition similar to what it is seen in humans with progressive MS.⁴⁴ This model presents with an immune response to virally infected cells and an autoimmune response to CNS antigens.⁴⁵ Novel therapies like natural human antibodies that induce either remyelination or neuroprotection have been successfully evaluated in this model of virus-induced demyelination.⁴⁶⁻⁴⁸ Other virus-induced models with a persistent viral infection without dramatic animal mortality are coronavirus (JHM and MHV-4) and Semliki Forest affecting mice, distemper virus specific for canines, and visna virus in sheep and goats.

MECHANISMS OF MULTIPLE SCLEROSIS: DEMYELINATION VERSUS AXONAL DAMAGE

Demyelination, the major pathologic hallmark of MS, is not sufficient to explain the deficits seen in these patients. In 1969, the first observation of demyelination and complete axonal dysfunction in the CNS was made,⁴⁹ and since then, many have considered it as a unique event. Recent studies found that in MS, damage to myelin is not enough to produce the spectrum of symptoms. In humans, levels of demyelination are not strictly correlated to disease stage, neurologic deficits, or lesion pathology.⁵⁰⁻⁵² Imaging studies in postmortem brains using MRI have shown how axonal injury is the primary event leading to clinical deficits (more than demyelination).^{53,54}

On the other hand, murine models of demyelination have also questioned the role of demyelination as the sole event in MS. Despite profound demyelination seen with TMEV murine model, in the absence of major histocompatibility complex (MHC) class I, there is no deficit in motor function.⁵⁵ Moreover, there is preservation of axonal transport in these mice despite demyelination.⁵⁶ CD8+ T cells direct recognition of MCH class I is a well-known mediator of axonal injury and dropout. In murine models of demyelination and murine culture of neurons, CD8+ T cells injure demyelinated axons selectively.^{57,58} CD8+ clones are the dominant cells in active MS lesions⁵⁹ and have direct correlation with accumulation of amyloid precursor protein, a marker of acute axonal damage.⁶⁰

Perforin, a critical mediator in cell cytotoxicity and apoptosis, is released by CD8+ T cells after recognition of the MCH class I complex.⁶¹ Once inside the cell, perforin creates a pore in the membrane and delivers granzymes that initiate a cascade of signals causing death of cells.⁶² During viral infection with TMEV and in electrically silent neurons in vitro, both have an increased expression of MCH class I complex.^{63,64} More importantly, axon injury secondary to demyelination is mediated by inflammatory factors, especially, perforin.⁶⁵ Perforin-deficient mice had an increased number of large-diameter axons and better functional motor abilities when compared with perforin-competent controls, despite having the same levels of demyelination.

Sodium channels are also a critical component in demyelinated axons, as changes in their number can influence impulse conduction.^{66,67} After an acute injury in peripheral axons, there is a high density of sodium channels.⁶⁸ Murine models of

demyelination have also shown how formation of nodes of Ranvier (where action potentials are generated) and saltatory conduction precedes remyelination in axons with demyelination.⁶⁹ Demyelinated mice with MHC class I deficiency and normal functional status have been previously described.⁵⁵ The normal motor function is caused by a preservation of axons and increased intensities of sodium channels suggesting an upregulation or redistribution of these molecules in the axons. Based on these observations, it can be concluded that axonal injury plays a critical role in the neurologic deficits seen in patients with MS and must be taken into account when considering strategies for MS therapies.

MYELIN AND AXONAL REPAIR STRATEGIES

Understanding the complex etiology of MS and the importance of axon integrity is critical for clinicians who expect halting of neuro-axonal damage. When a patient with newly diagnosed MS has the first neurologic symptom, there is already axonal loss. Hence, there is a need to treat early and use multiple strategies that target remyelination and preservation of axons and oligodendrocytes (OL).

The authors' laboratory has done extensive research in natural human recombinant antibodies formerly known as *rH IgM22* and *rH IgM12*. These molecules, part of human innate immunoglobulin repertoire, are able to bind to oligodendrocytes (*rH IgM22*) and neurons (*rH IgM12*).⁷⁰⁻⁷² In TMEV and lysolecithin-induced demyelination, *rH IgM22* promotes oligodendrocyte remyelination and protects spinal cord axon number.⁴⁷ Currently, this molecule is under a clinical trial to establish its tolerance in MS patients after an acute exacerbation. In addition, *rH IgM12* protects against axonal injury in TMEV and amyotrophic lateral sclerosis murine models.⁷² It also binds to PSA-NCAM and gangliosides of glia and neurons resulting in neurite extension in vitro and neurite outgrowth.⁷³ Recently, *rH IgM12* showed an immune-modulatory therapeutic effect in an MOG-induced experimental autoimmune encephalomyelitis (EAE).⁷⁴

Inhibition of the leucine rich repeat and immunoglobulinlike domain containing NOGO receptor interacting protein-1 (LINGO-1) increases differentiation of oligodendrocyte progenitor cells into mature oligodendrocytes.⁷⁵ In TMEV murine model of demyelination, inhibition of this molecule promoted remyelination leading to the discovery of anti-LINGO-1.⁷⁶ This was the first drug entering a clinical trial showing how 41% of patients had improvement of nerve signaling and possibly myelin repair.⁷⁷ Despite this, anti-LINGO-1 failed to improve disability and cognitive function after 72 weeks of follow-up.

As explained in the previous section, CD8+ T cells and the upregulation of MHC class I complex are critical in the pathogenesis of MS. Dimethyl fumarate (BG-12 or Tecfidera) is a molecule that showed remarkable efficacy in the treatment of relapsing remitting MS and exposure may result in reductions in CD8+ T-cell populations in certain patients.⁷⁸ Fingolimod, the first approved oral therapy for active relapsing remitting MS, modulates T-cell proliferation in vitro as well.⁷⁹ Together, these results suggest that an immunotherapy against active CD8+ cells using anti-CD8 antibodies⁸⁰ could suppress the immune-mediated reactions in patients with MS.

SUMMARY

MS is an immune-mediated disease that lacks the presence of a specific antigen that drives the inflammatory process. The extensive characterization of MS pathology, immunology, and serology has misled many investigators to conclude that MS is an autoimmune disease. However, there is a lack of scientific evidence supporting the role of active autoantigens and autoantibodies driving the inflammatory and

demyelinating cascades seen in patients with MS. Understanding the role of axonal injury and its relationship with clinical deficits is essential for a future drug. Future trials should aim a multifaceted approach with several reagents that target remyelination, protections of axons and oligodendrocyte progenitor cell, and suppressive therapies against active inflammatory cell populations.

ACKNOWLEDGMENTS

The authors acknowledge with many thanks support from the Applebaum, Hilton, Peterson and Sanford Foundations, and the McNeilus family.

REFERENCES

1. Lucchinetti C, Bruck W, Parisi J, et al. Heterogeneity of multiple sclerosis lesions: implications for the pathogenesis of demyelination. *Ann Neurol* 2000;47(6):707–17.
2. Pelletier D, Hafler DA. Fingolimod for multiple sclerosis. *N Engl J Med* 2012; 366(4):339–47.
3. Frohman EM, Racke MK, Raine CS. Multiple sclerosis—the plaque and its pathogenesis. *N Engl J Med* 2006;354(9):942–55.
4. Qin Y, Duquette P, Zhang Y, et al. Clonal expansion and somatic hypermutation of V(H) genes of B cells from cerebrospinal fluid in multiple sclerosis. *J Clin Invest* 1998;102(5):1045–50.
5. Ando DG, Clayton J, Kono D, et al. Encephalitogenic T cells in the B10.PL model of experimental allergic encephalomyelitis (EAE) are of the Th-1 lymphokine subtype. *Cell Immunol* 1989;124(1):132–43.
6. Pettinelli CB, McFarlin DE. Adoptive transfer of experimental allergic encephalomyelitis in SJL/J mice after in vitro activation of lymph node cells by myelin basic protein: requirement for Lyt 1+ 2- T lymphocytes. *J Immunol* 1981;127(4):1420–3.
7. Paul WE, Schwartz RS, Datta SK. Autoimmunity and autoimmune diseases. In: Fundamental Immunology. New York: Raven Press; 1989. p. 819–66.
8. Rodriguez M, Warrington AE, Pease LR. Invited article: human natural autoantibodies in the treatment of neurologic disease. *Neurology* 2009;72(14):1269–76.
9. Chan OT, Hannum LG, Haberman AM, et al. A novel mouse with B cells but lacking serum antibody reveals an antibody-independent role for B cells in murine lupus. *J Exp Med* 1999;189(10):1639–48.
10. Bagavant H, Fu SM. New insights from murine lupus: disassociation of autoimmunity and end organ damage and the role of T cells. *Curr Opin Rheumatol* 2005; 17(5):523–8.
11. Cinque P, Cleator GM, Weber T, et al. The role of laboratory investigation in the diagnosis and management of patients with suspected herpes simplex encephalitis: a consensus report. *J Neurol Neurosurg Psychiatry* 1996;61(4):339–45.
12. Kennel De March A, De Bouwerie M, Kolopp-Sarda MN, et al. Anti-myelin oligodendrocyte glycoprotein B-cell responses in multiple sclerosis. *J Neuroimmunol* 2003;135(1–2):117–25.
13. Khalil M, Reindl M, Lutterotti A, et al. Epitope specificity of serum antibodies directed against the extracellular domain of myelin oligodendrocyte glycoprotein: influence of relapses and immunomodulatory treatments. *J Neuroimmunol* 2006; 174(1–2):147–56.
14. Karni A, Bakimer-Kleiner R, Abramsky O, et al. Elevated levels of antibody to myelin oligodendrocyte glycoprotein is not specific for patients with multiple sclerosis. *Arch Neurol* 1999;56(3):311–5.

15. Reindl M, Linington C, Brehm U, et al. Antibodies against the myelin oligodendrocyte glycoprotein and the myelin basic protein in multiple sclerosis and other neurological diseases: a comparative study. *Brain* 1999;122(Pt 11):2047–56.
16. Lutterotti A, Reindl M, Gassner C, et al. Antibody response to myelin oligodendrocyte glycoprotein and myelin basic protein depend on familial background and are partially associated with human leukocyte antigen alleles in multiplex families and sporadic multiple sclerosis. *J Neuroimmunol* 2002;131(1–2):201–7.
17. Agius MA, Kirvan CA, Schafer AL, et al. High prevalence of anti-alpha-crystallin antibodies in multiple sclerosis: correlation with severity and activity of disease. *Acta Neurol Scand* 1999;100(3):139–47.
18. Archelos JJ, Trotter J, Previtali S, et al. Isolation and characterization of an oligodendrocyte precursor-derived B-cell epitope in multiple sclerosis. *Ann Neurol* 1998;43(1):15–24.
19. Cruz M, Olsson T, Ernerudh J, et al. Immunoblot detection of oligoclonal anti-myelin basic protein IgG antibodies in cerebrospinal fluid in multiple sclerosis. *Neurology* 1987;37(9):1515–9.
20. Moller JR, Johnson D, Brady RO, et al. Antibodies to myelin-associated glycoprotein (MAG) in the cerebrospinal fluid of multiple sclerosis patients. *J Neuroimmunol* 1989;22(1):55–61.
21. Papadopoulos MC, Verkman AS. Aquaporin water channels in the nervous system. *Nat Rev Neurosci* 2013;14(4):265–77.
22. Hinson SR, Pittock SJ, Lucchinetti CF, et al. Pathogenic potential of IgG binding to water channel extracellular domain in neuromyelitis optica. *Neurology* 2007;69(24):2221–31.
23. Hinson SR, Roemer SF, Lucchinetti CF, et al. Aquaporin-4-binding autoantibodies in patients with neuromyelitis optica impair glutamate transport by down-regulating EAAT2. *J Exp Med* 2008;205(11):2473–81.
24. Bennett JL, Lam C, Kalluri SR, et al. Intrathecal pathogenic anti-aquaporin-4 antibodies in early neuromyelitis optica. *Ann Neurol* 2009;66(5):617–29.
25. Lucchinetti CF, Mandler RN, McGavern D, et al. A role for humoral mechanisms in the pathogenesis of Devic's neuromyelitis optica. *Brain* 2002;125(Pt 7):1450–61.
26. Steinman L. Immunology of relapse and remission in multiple sclerosis. *Annu Rev Immunol* 2014;32:257–81.
27. Bradl M, Misu T, Takahashi T, et al. Neuromyelitis optica: pathogenicity of patient immunoglobulin in vivo. *Ann Neurol* 2009;66(5):630–43.
28. Ollitsky PK, Yager RH. Experimental disseminated encephalomyelitis in white mice. *J Exp Med* 1949;90(3):213–24.
29. Swanson RH. Experimental autoimmune encephalomyelitis in the rat: lessons in T-cell immunology and autoreactivity. *Immunol Rev* 2001;184:129–35.
30. Lennon VA, Kryzer TJ, Pittock SJ, et al. IgG marker of optic-spinal multiple sclerosis binds to the aquaporin-4 water channel. *J Exp Med* 2005;202(4):473–7.
31. Jarius S, Wildemann B. AQP4 antibodies in neuromyelitis optica: diagnostic and pathogenetic relevance. *Nat Rev Neurol* 2010;6(7):383–92.
32. Pittock SJ, Lennon VA, de Seze J, et al. Neuromyelitis optica and non organ-specific autoimmunity. *Arch Neurol* 2008;65(1):78–83.
33. Wynn DR, Rodriguez M, O'Fallon WM, et al. A reappraisal of the epidemiology of multiple sclerosis in Olmsted County, Minnesota. *Neurology* 1990;40(5):780–6.
34. Bonnan M, Valentino R, Olindo S, et al. Plasma exchange in severe spinal attacks associated with neuromyelitis optica spectrum disorder. *Mult Scler* 2009;15(4):487–92.

35. Rodriguez M, Karnes WE, Bartleson JD, et al. Plasmapheresis in acute episodes of fulminant CNS inflammatory demyelination. *Neurology* 1993;43(6):1100–4.
36. Magana SM, Keegan BM, Weinshenker BG, et al. Beneficial plasma exchange response in central nervous system inflammatory demyelination. *Arch Neurol* 2011;68(7):870–8.
37. SantaCruz KS, Roy G, Spigel J, et al. Neuropathology of JC virus infection in progressive multifocal leukoencephalopathy in remission. *World J Virol* 2016;5(1):31–7.
38. Cepok S, Zhou D, Srivastava R, et al. Identification of Epstein-Barr virus proteins as putative targets of the immune response in multiple sclerosis. *J Clin Invest* 2005;115(5):1352–60.
39. Villegas E, Santiago O, Carrillo JA, et al. Low intrathecal immune response of anti-EBNA-1 antibodies and EBV DNA from multiple sclerosis patients. *Diagn Microbiol Infect Dis* 2011;70(1):85–90.
40. Eftekharian MM, Ghannad MS, Taheri M, et al. Frequency of viral infections and environmental factors in multiple sclerosis. *Hum Antibodies* 2016;24(1–2):17–23.
41. Shaygannejad V, Rezaie N, Paknahad Z, et al. The environmental risk factors in multiple sclerosis susceptibility: a case-control study. *Adv Biomed Res* 2016;5:98.
42. Toghianifar N, Ashtari F, Zarkesh-Esfahani SH, et al. Effect of high dose vitamin D intake on interleukin-17 levels in multiple sclerosis: a randomized, double-blind, placebo-controlled clinical trial. *J Neuroimmunol* 2015;285:125–8.
43. Golan D, Halhal B, Glass-Marmor L, et al. Vitamin D supplementation for patients with multiple sclerosis treated with interferon-beta: a randomized controlled trial assessing the effect on flu-like symptoms and immunomodulatory properties. *BMC Neurol* 2013;13:60.
44. Theiler M. Spontaneous encephalomyelitis of mice—a new virus disease. *Science* 1934;80(2066):122.
45. dal Canto MC, Lipton HL. A new model of persistent viral infection with primary demyelination. *Neurol Neurocir Psiquiatr* 1977;18(2–3 Suppl):455–67.
46. Warrington AE, Bieber AJ, Ceric B, Pease LR, et al. A recombinant human IgM promotes myelin repair after a single, very low dose. *J Neurosci Res* 2007;85(5):967–76.
47. Wootla B, Denic A, Watzlawik JO, et al. Antibody-mediated oligodendrocyte remyelination promotes axon health in progressive demyelinating disease. *Mol Neurobiol* 2016;53(8):5217–28.
48. Wootla B, Denic A, Warrington AE, et al. A monoclonal natural human IgM protects axons in the absence of remyelination. *J Neuroinflammation* 2016;13(1):94.
49. McDonald WI, Sears TA. Effect of demyelination on conduction in the central nervous system. *Nature* 1969;221(5176):182–3.
50. Stevens JC, Farlow MR, Edwards MK, et al. Magnetic resonance imaging. Clinical correlation in 64 patients with multiple sclerosis. *Arch Neurol* 1986;43(11):1145–8.
51. Bruck W, Bitsch A, Kolenda H, et al. Inflammatory central nervous system demyelination: correlation of magnetic resonance imaging findings with lesion pathology. *Ann Neurol* 1997;42(5):783–93.
52. Mews I, Bergmann M, Bunkowski S, et al. Oligodendrocyte and axon pathology in clinically silent multiple sclerosis lesions. *Mult Scler* 1998;4(2):55–62.
53. van Waesberghe JH, Kamphorst W, De Groot CJ, et al. Axonal loss in multiple sclerosis lesions: magnetic resonance imaging insights into substrates of disability. *Ann Neurol* 1999;46(5):747–54.

54. Fisher E, Chang A, Fox RJ, et al. Imaging correlates of axonal swelling in chronic multiple sclerosis brains. *Ann Neurol* 2007;62(3):219–28.
55. Rivera-Quinones C, McGavern D, Schmelzer JD, et al. Absence of neurological deficits following extensive demyelination in a class I-deficient murine model of multiple sclerosis. *Nat Med* 1998;4(2):187–93.
56. Ure DR, Rodriguez M. Preservation of neurologic function during inflammatory demyelination correlates with axon sparing in a mouse model of multiple sclerosis. *Neuroscience* 2002;111(2):399–411.
57. Medana I, Martinic MA, Wekerle H, et al. Transection of major histocompatibility complex class I-induced neurites by cytotoxic T lymphocytes. *Am J Pathol* 2001;159(3):809–15.
58. Johnson AJ, Upshaw J, Pavelko KD, et al. Preservation of motor function by inhibition of CD8+ virus peptide-specific T cells in Theiler's virus infection. *FASEB J* 2001;15(14):2760–2.
59. Babbe H, Roers A, Waisman A, et al. Clonal expansions of CD8(+) T cells dominate the T cell infiltrate in active multiple sclerosis lesions as shown by micromanipulation and single cell polymerase chain reaction. *J Exp Med* 2000;192(3):393–404.
60. Bitsch A, Schuchardt J, Bunkowski S, et al. Acute axonal injury in multiple sclerosis. Correlation with demyelination and inflammation. *Brain* 2000;123(Pt 6):1174–83.
61. Lieberman J. The ABCs of granule-mediated cytotoxicity: new weapons in the arsenal. *Nat Rev Immunol* 2003;3(5):361–70.
62. Metkar SS, Wang B, Aguilar-Santelises M, et al. Cytotoxic cell granule-mediated apoptosis: perforin delivers granzyme B-serglycin complexes into target cells without plasma membrane pore formation. *Immunity* 2002;16(3):417–28.
63. Lindsley MD, Patick AK, Prayoonwiwat N, et al. Coexpression of class I major histocompatibility antigen and viral RNA in central nervous system of mice infected with Theiler's virus: a model for multiple sclerosis. *Mayo Clin Proc* 1992;67(9):829–38.
64. Neumann H, Cavalie A, Jenne DE, et al. Induction of MHC class I genes in neurons. *Science* 1995;269(5223):549–52.
65. Howe CL, Adelson JD, Rodriguez M. Absence of perforin expression confers axonal protection despite demyelination. *Neurobiol Dis* 2007;25(2):354–9.
66. Waxman SG, Black JA, Kocsis JD, et al. Low density of sodium channels supports action potential conduction in axons of neonatal rat optic nerve. *Proc Natl Acad Sci U S A* 1989;86(4):1406–10.
67. Johnston WL, Dyer JR, Castellucci VF, et al. Clustered voltage-gated Na⁺ channels in Aplysia axons. *J Neurosci* 1996;16(5):1730–9.
68. Foster RE, Whalen CC, Waxman SG. Reorganization of the axon membrane in demyelinated peripheral nerve fibers: morphological evidence. *Science* 1980;210(4470):661–3.
69. Smith KJ, Bostock H, Hall SM. Saltatory conduction precedes remyelination in axons demyelinated with lysophosphatidyl choline. *J Neurol Sci* 1982;54(1):13–31.
70. Avrameas S. Natural autoantibodies: from 'horror autotoxicus' to 'gnothi seauton'. *Immunol Today* 1991;12(5):154–9.
71. Watzlawik J, Holicky E, Edberg DD, et al. Human remyelination promoting antibody inhibits apoptotic signaling and differentiation through Lyn kinase in primary rat oligodendrocytes. *Glia* 2010;58(15):1782–93.

72. Xu X, Denic A, Jordan LR, et al. A natural human IgM that binds to gangliosides is therapeutic in murine models of amyotrophic lateral sclerosis. *Dis Model Mech* 2015;8(8):831–42.
73. Watzlawik J, Kahoud RJ, Ng S, et al. Polysialic acid as an antigen for monoclonal antibody HlgM12 to treat multiple sclerosis and other neurodegenerative disorders. *J Neurochem* 2015;134(5):865–78.
74. Lemus HN, Warrington AE, Denic A, et al. Treatment with a recombinant human IgM that recognizes PSA-NCAM preserves brain pathology in MOG-induced experimental autoimmune encephalomyelitis. *Hum Antibodies* 2017;25(3–4):121–9.
75. Mi S, Miller RH, Lee X, et al. LINGO-1 negatively regulates myelination by oligodendrocytes. *Nat Neurosci* 2005;8(6):745–51.
76. Mi S, Miller RH, Tang W, et al. Promotion of central nervous system remyelination by induced differentiation of oligodendrocyte precursor cells. *Ann Neurol* 2009;65(3):304–15.
77. MS Society 2016. 2016. Available at: <https://www.mssociety.org.uk/node/690821>. Accessed September 26, 2017.
78. Wu Q, Wang Q, Mao G, et al. Dimethyl fumarate selectively reduces memory Tcells and shifts the balance between Th1/Th17 and Th2 in multiple sclerosis patients. *J Immunol* 2017;198(8):3069–80.
79. Thomas K, Sehr T, Proschmann U, et al. Fingolimod additionally acts as immunomodulator focused on the innate immune system beyond its prominent effects on lymphocyte recirculation. *J Neuroinflammation* 2017;14(1):41.
80. Clement M, Pearson JA, Gras S, et al. Targeted suppression of autoreactive CD8+ T-cell activation using blocking anti-CD8 antibodies. *Sci Rep* 2016;6:35332.