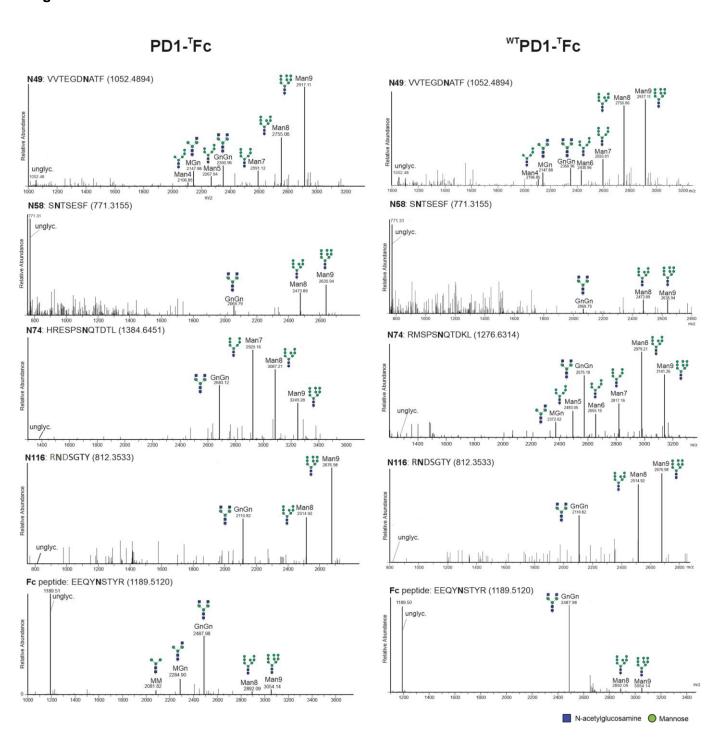
Fig. S1



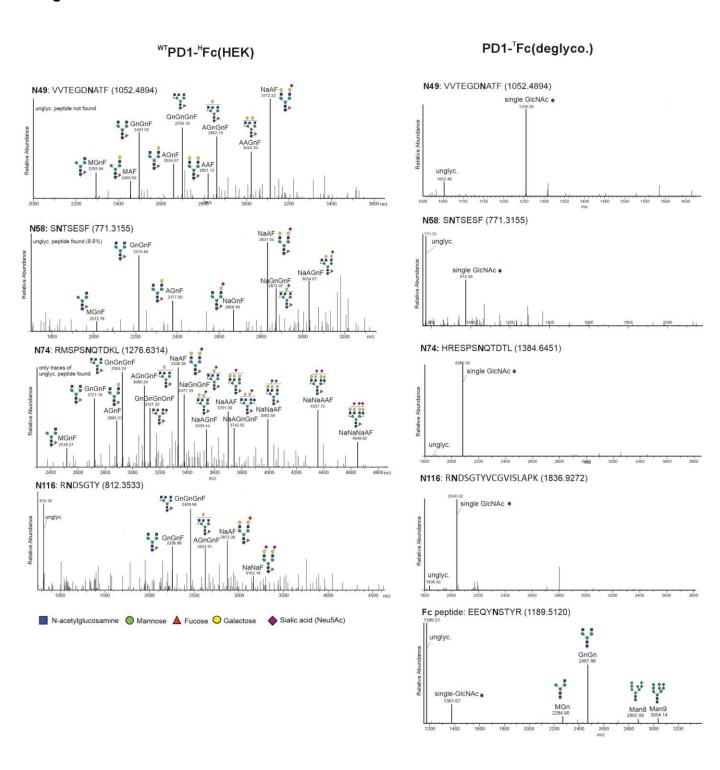
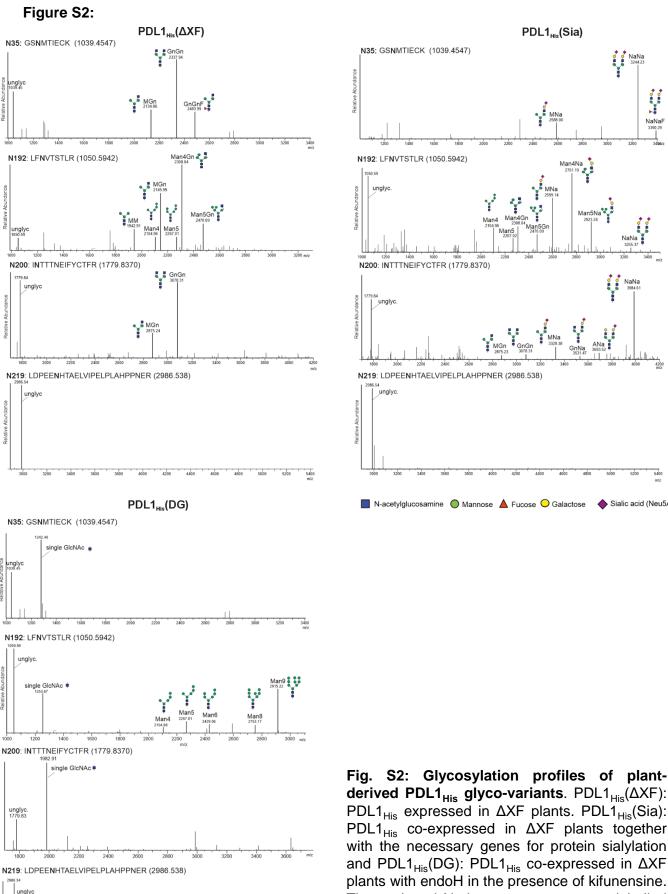


Fig. S1: Glycosylation profiles of WTPD1-Fc(HEK) and plant-derived PD1-Fc variants expressed in *N. benthamiana* ΔXF plants. The assigned N-glycan structures were labelled according to the ProGlycAn nomenclature. A cartoon illustration highlights the main glycan structures detected for each peptide. For details see http://www.functionalglycomics.org/.



derived PDL1_{His} glyco-variants. PDL1_{His}(ΔXF): PDL1_{His} expressed in ΔXF plants. PDL1_{His}(Sia): PDL1_{His} co-expressed in ΔXF plants together with the necessary genes for protein sialylation and PDL1_{His}(DG): PDL1_{His} co-expressed in ΔXF plants with endoH in the presence of kifunensine. The assigned N-glycan structures were labelled according to the ProGlycAn nomenclature. A cartoon illustration highlights the main glycan structures detected for each peptide. For details see http://www.functionalglycomics.org/.

Sialic acid (Neu5Ac)

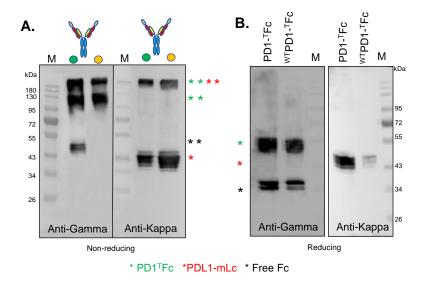
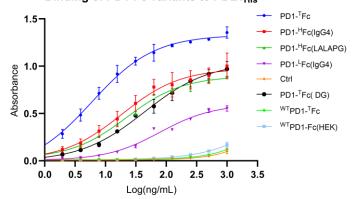
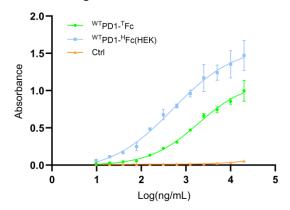


Fig. S3: In planta interaction of PDL1 and PD1-Fc fusions. (A) Western blot analysis of proteins purified with protein A (green dot) and KappaSelect (orange dot) after co-expression of PDL1-mLc and PD1-TFc. Co-purified proteins were analysed in non-reducing conditions with anti-gamma (left) and anti-kappa (right) antibodies. (B) Western blot analysis of proteins purified with protein A after co-expression of PDL1-mLc with PD1-TFc and WTPD1-TFc. Co-purified proteins were analysed under reducing conditions with anti-gamma (left) and anti-kappa (right) antibodies. The apparent molecular mass of marker proteins (M) is shown in kilo Dalton (kDa).

A. Binding of PD1-Fc variants to PDL1_{His}



B. Binding of WTPD1-Fc variants to PDL1His



C. Binding of PD1-^TFc to PDL1His glycovariants

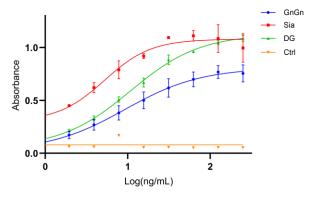


Fig. S4: ELISA binding assays. (**A**) Binding of plant-produced PD1-Fc fusion variants to recombinant PDL1_{His}. Serial dilutions of PD1-Fc variants (1000 to 0.48 ng/mL) were added to plates coated with 200 ng of PDL1_{His} and detected with anti-human IgG-HRP. (**B**) Binding of WTPD1-TFc and WTPD1-Fc(HEK) to recombinant PDL1_{His}. Serial dilutions of PD1-Fc variants (20000 to 10 ng/mL) were added to plates coated with 1 μg of PDL1_{His} and detected with anti-human IgG-HRP. An unrelated Fc fusion was used as negative control. Data represent the mean values of triplicates. (**C**) Binding of plant-produced PD1-TFc fusion to PDL1_{His} glycovariants: PDL1_{His} expressed in ΔXF plants. PDL1_{His}(Sia): PDL1_{His} co-expressed in ΔXF plants together with the necessary genes for protein sialylation and PDL1_{His}(DG): PDL1_{His} co-expressed in ΔXF plants with endoH in the presence of kifunensine. An unrelated His-tagged protein was used as negative control. Data represent the mean values of triplicates. Bars represent SD.

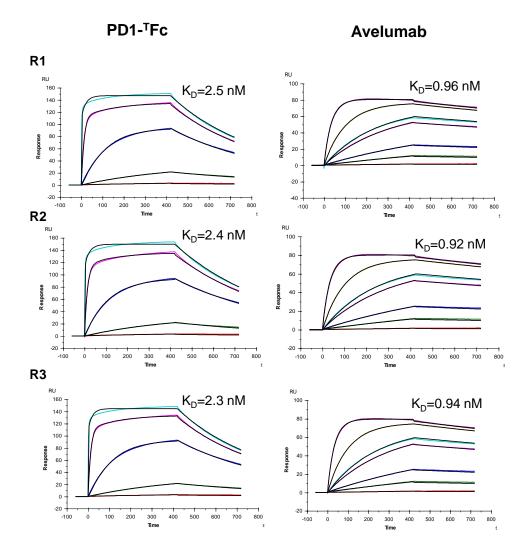


Fig. S5: SPR sensorgrams for the binding of PD1-TFc, WTPD1-Fc(HEK) and Avelumab to PDL1_{His.} Analyte serial dilutions of 250 nM-4000 nM, 0.1 nM-1000 nM and 0.1 nM-75 nM were used to probe affinity of PDL1_{His} to WTPD1-HFc(HEK), PD1-TFc and Avelumab, respectively. Experiments were done in three runs (R1-R3). K_D values are given for each sensorgram.

Blocking PD1/PDL1 interaction

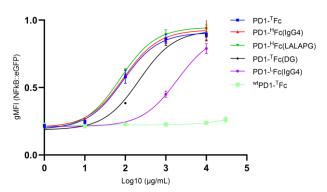


Fig. S6: Blocking PD1/PDL1 interaction by PD1-Fc variants. Inhibition curves used to determine functional half maximum effective concentrations (EC_{50}). PD1-Fc variants were evaluated at different concentrations (ranging from 10 to 0.001 µg/mL) using PD1+NF- κ B::eGFP T-cell reporters co-cultured with T-cell stimulator cells expressing PDL1. gMFI: geometric mean of fluorescence intensity. Data are derived from two independent experiments performed in triplicates (n=6). Bars represent SD.