





Draft Genome Sequence of *Pseudomonas putida* CBF10-2, a Soil Isolate with Bioremediation Potential in Agricultural and Industrial Environmental Settings

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Pseudomonas putida CBF10-2 is a microorganism isolated from farmland soil in Fairchild, TX, found to degrade high-impact xenobiotics, including organophosphate insecticides, petroleum hydrocarbons, and both monocyclic and polycyclic aromatics. The versatility of CBF10-2 makes it useful for multipurpose bioremediation of contaminated sites in agricultural and industrial environments.

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seudomonas putida is a Gram-negative gammaproteobacterium ubiquitous to contaminated soil environments and is well known for its tolerance and degradation capacity for organic solvents (1). Here, we report a draft genome sequence of an organophosphate-degrading strain of P. putida, isolated from ranch soil in Fairchild, TX, through an Environmental Sampling Research Module undertaken by University of Houston biotechnology undergraduates (Houston, TX) (2). The closest relative to CBF10-2 is the naphthalene degrader P. putida CVS86, known for its preference for the polycyclic aromatic naphthalene as a carbon source over glucose (3). While appearing to lack a clearly defined naphthalene operon itself, CBF10-2 shares its species' proclivity for aromatic hydrocarbon substrates and possesses an assortment of degradation enzymes with broad activity against petroleum hydrocarbons as well as organophosphate phosphotriesters and phosphorothioates. The genome sequencing of CBF10-2 was performed through Illumina MiSeq paired-end sequencing (35 to 251 bp in each read), with a final sequencing coverage of $61\times$. Sequence reads were assessed for quality using FastQC (4) and filtered using BBTools (5), with a minimum Phred score of 20. Paired-end reads were assembled into 73 contigs with the SPAdes 3.7 program (6). Preliminary reference-based annotation using PATRIC (7) Web resources was carried out to identify conserved pathways. Final de novo annotation was performed with Prokka (8) and the NCBI Prokaryotic Genome Annotation Pipeline (http://www.ncbi.nlm.nih.gov/genomes/static/Pipeline .html). The metabolic pathways of aromatic and heterocyclic compounds were examined using the KEGG databases (9). This draft genome of strain CBF10-2 consists of a total of 6,120,625 bp. CBF10-2 contains 5,449 total genes, of which 64 are pseudogenes, 1,140 represent hypothetical proteins, and 4,171 form known functional proteins. The genome has a G+C content of 63.72% and contains nine rRNA (five complete and four partial), 61 tRNA, and four noncoding RNA (ncRNA) loci.

Nucleotide sequence accession numbers. The *Pseudomonas putida* CBF10-2 whole-genome shotgun (WGS) project has the project accession no. LUCV00000000. This version of the project (01) has the accession number LUCV01000000 and consists of sequences LUCV01000001 to LUCV0100073.

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REFERENCES

- Iyer R, Smith K, Kudrle B, Leon A. 2015. Detection and location of OP-degrading activity: a model to integrate education and research. N Biotechnol 32:403–411. http://dx.doi.org/10.1016/j.nbt.2015.03.010.
- Wu X, Monchy S, Taghavi S, Zhu W, Ramos J, van der Lelie D. 2011. Comparative genomics and functional analysis of niche-specific adaptation in *Pseudomonas putida*. FEMS Microbiol Rev 35:299–323. http:// dx.doi.org/10.1111/j.1574-6976.2010.00249.x.
- Phale PS, Paliwal V, Raju SC, Modak A, Purohit HJ. 2013. Genome sequence of naphthalene-degrading soil bacterium *Pseudomonas putida* CSV86. Genome Announc 1(1):e00234-12. http://dx.doi.org/10.1128/ genomeA.00234-12.
- 4. Bioinformatics B. 2011. FastQC a quality control tool for high throughput sequence data. Babraham Institute, Cambridge, United Kingdom. http://www.bioinformatics.babraham.ac.uk/projects/fastqc/.
- 5. Bushnell B. BBMap short read aligner. http://sourceforge.net/projects/bbmap.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/cmb.2012.0021.
- 7. Wattam AR, Abraham D, Dalay O, Disz TL, Driscoll T, Gabbard JL,

Gillespie JJ, Gough R, Hix D, Kenyon R, Machi D, Mao C, Nordberg EK, Olson R, Overbeek R, Pusch GD, Shukla M, Schulman J, Stevens RL, Sullivan DE, Vonstein V, Warren A, Will R, Wilson MJC, Seung YH, Zhang C, Zhang Y, Sobral BW. 2014. PATRIC, the bacterial bioinformatics database and analysis resource. Nucleic Acids Res 42:D581–D591. http://dx.doi.org/10.1093/nar/gkt1099.

- 8. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. Bioinformatics 30:2068–2069. http://dx.doi.org/10.1093/bioinformatics/btu153.
- Kanehisa M, Goto S. 2000. KEGG: Kyoto Encyclopedia of Genes and Genomes. Nucleic Acids Res 28:27–30. http://dx.doi.org/10.1093/nar/ 28.1.27.