

REFERENCES

1. Iba T, Levy JH, Levi M, Thachil J. Coagulopathy in COVID-19. *J Thromb Haemost.* 2020;18(9):2103-2109.
2. Zoccolella S, Martino D, Defazio G, Lamberti P, Livrea P. Hyperhomocysteinemia in movement disorders: current evidence and hypotheses. *Curr Vasc Pharmacol.* 2006;4(3):237-243.
3. Verdoia M, Rolla R, Negro F, et al. Homocysteine levels and platelet reactivity in coronary artery disease treated with ticagrelor. *Nutr Metab Cardiovasc Dis.* 2020;30:292-299.
4. Lupi-Herrera E, Soto-Lopez ME, Lugo-Dimas AJ, et al. Polymorphisms C667T and A1298C of MTHFR gene: Homocysteine levels and pro-thrombotic biomarkers in coronary and pulmonary thromboembolic disease. *Clin Appl Thromb Hemost.* 2019;25:1076029618780344.
5. Poissy J, Goutay J, Caplan M, et al. Pulmonary embolism in COVID-19 patients: awareness of an increased prevalence. *Circulation.* 2020;142(2):184-186.
6. Faggiano P, Bonelli A, Paris S, et al. Acute pulmonary embolism in COVID-19 disease: preliminary report on seven patients. *Int J Cardiol.* 2020;313:129-131.
7. Yang Z, Shi J, He Z, et al. Predictors for imaging progression on chest CT from coronavirus disease 2019 (COVID-19) patients. *Aging.* 2020;12(7):6037-6048.
8. Ponti G, Ruini C, Tomasi A. Homocysteine as a potential predictor of cardiovascular risk in patients with COVID-19. *Med Hypotheses.* 2020;143:109859.
9. B-Vitamin Treatment Trialists' Collaboration. Homocysteine-lowering trials for prevention of cardiovascular events: a review of the design and power of the large randomized trials. *Am Heart J.* 2006;151(2):282-287.
10. Tan SHS, Hong CC, Saha S, Murphy D, Hui JH. Medications in COVID-19 patients: summarizing the current literature from an orthopaedic perspective. *Int Orthopaedics.* 2020;44(8):1599-1603.
11. Ibrahimagić OČ, Smajlović D, Dostović Z, Pašić Z, Šehanović A, Hodžić R. Hyperhomocysteinemia and its treatment in patients with ischemic stroke. *Medicus.* 2012;21(2):267-272.
12. Ibrahimagić OČ, Smajlović D, Dostović Z, et al. Hyperhomocysteinemia and its treatment in patients with Parkinson's disease. *Mater Sociomed.* 2016;28(4):303-306.

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Letter in response to: Coagulation markers are independent predictors of increased oxygen requirements and thrombosis in COVID-19

We read with interest the recent publication by Rauche et al and congratulate them on highlighting the potentially significant role von Willebrand factor (VWF) may play in the progression and prognosis of COVID-19.¹ Since SARS-Cov2 first presented there has been an increasing recognition that the disease may trigger a widespread endotheliopathy. This fact could help to explain the highly varied presentation of patients with COVID-19 from bowel ischemia to large vessel occlusion causing strokes in addition to the more common presentation of respiratory symptoms. At the same time there has been an increased understanding that the infection may result in microthrombosis as well as macrothrombosis both on the arterial and venous side of the circulation. Contrast enhanced ultrasound was recently used to identify microthrombi and wedge-shaped perfusion defects in vivo in the pulmonary, renal, and gastrointestinal beds.^{2,3} Routine imaging methods, such as computed tomography (CT) and CT angiography, do not have the spatial or contrast resolution to detect microvascular thrombosis and therefore, the demonstration of perfusion defects in vivo has been delayed. Post mortem studies

have also shown microthrombosis within the lungs confirming the in vivo imaging findings. In the study of Carsana et al, microthrombi, in vessels < 1mm in diameter, were seen in 87% of cases.⁴ A further case series of COVID-19 pulmonary autopsies revealed that, alongside diffuse alveolar damage, numerous localized platelet-rich microthrombi, and foci of hemorrhage were present in the lungs.⁵ The authors posited a pulmonary-localised thrombotic microangiopathy as key to the pathogenesis of COVID-19 with others also suggesting the micro-thrombosis is a critical driver in the disease process.⁶

Von Willebrand factor is synthesized only in megakaryocytes and endothelial cells (ECs). The vast majority of VWF found in the plasma is derived from the VWF synthesized within the ECs, where it is stored within the Weibel-Palade bodies (WPB). Although restricted to ECs there are differences in the synthesis of VWF within the different vascular beds of the body with the small vessels of the lung and brain expressing higher levels of VWF than similar sized vessels of the liver or kidney and higher levels in venous rather than arterial ECs.⁷ A major portion of the VWF stored in the WPBs of endothelial cells is made up of ultra-large VWF (ULVWF). These larger multimers are more adhesive than the smaller multimers in the circulation and upon secretion ULVWF can spontaneously bind

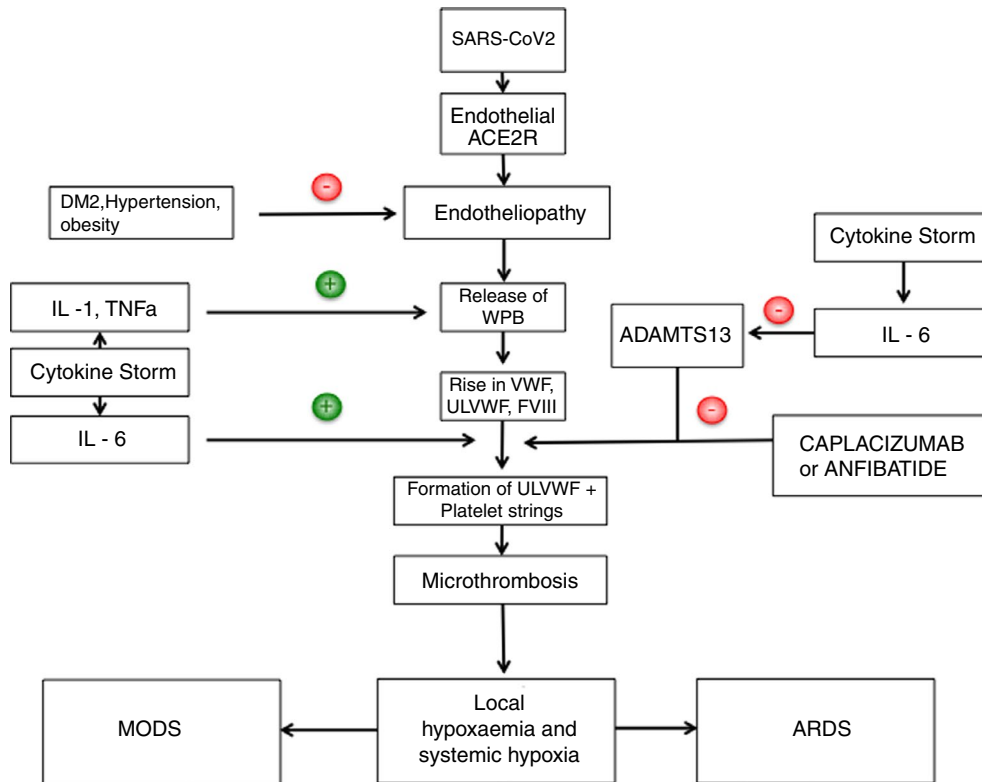


FIGURE 1 Pathways involved in the release of ULVWF, generation of microthrombosis and regulating factors. Both anfibatide and caplacizumab can inhibit platelet binding to VWF and therefore represent attractive options to prevent microthrombosis and subsequent hypoxia/ischaemia

platelets and cause occlusion of small vessels. The secreted VWF, which partly enters the circulation and partly binds to the endothelium, is sensitive to shear stress. This shear stress unfolds the VWF and exposes sites for platelet binding and self-association as well as for cleavage via the enzyme ADAMTS13. It has previously been shown these VWF molecules can self-associate into long “strings” in the direction of flow, both arterial and venous, that bind to platelets and are adherent to the endothelium.⁸⁻¹⁰ The ULVWF multimers released from the WPBs unfold at lower levels of shear stress and therefore may represent the initiating molecules for this self-assembly process, which leads to hyper-adhesive strings capturing platelets. Preventing the binding of platelets to ULVWF may represent an attractive target to prevent microthrombosis formation. Inflammatory cytokines, such as interleukin (IL)-1 and tumor necrosis factor (TNF)-alpha, as well as hypoxia, can trigger the exocytosis of WPBs with release of their contents while IL-6 can inhibit the cleavage of ULVWF–platelet strings.¹¹ Furthermore, the synthesis of ADAMTS13, at least in cultured cells, is dramatically inhibited by a variety of cytokines including IL-6 and TNF-alpha.¹² This suggests that hypoxia, the cytokine storm, and particularly IL-6, may help to propagate the microthrombosis.

One of the most interesting findings of the published work was that the 10 patients presenting with normal VWF levels were all discharged from the emergency department with out-patient follow-up. Although only a very small cohort, we found it interesting

that the group with normal VWF levels did not require admission or treatment with oxygen and we believe that this supports the concept that VWF plays a critical role in the pathogenesis. Furthermore, it is possible that VWF levels and the VWF:factor VIII (FVIII) ratio could act as biomarkers for determining which patients are likely to progress as well as those that are responding to treatment. The recent finding of ABO blood type and disease severity also points to a potential link with VWF with those patients with type O blood group being at lowest risk and those with type A being at highest risk.¹³ Patients with type O have lower levels of VWF compared to those with non-O blood group and those with non-O blood group are at higher risk of arterial thromboembolic disease such as a ischemic heart disease and peripheral vascular disease. Although the exact nature of the interaction remains unknown it has been suggested that ABO blood group determinants may be important in influencing the susceptibility of plasma VWF to proteolysis by ADAMTS13.¹⁴ Therefore, it is possible patients more susceptible to severe disease are already predisposed to thromboembolic diseases and that the sudden surge in VWF, particularly ULVWF, triggered by a widespread endotheliopathy overwhelms the body's innate ability to break down VWF via ADAMTS13. Further evidence for this comes from the recent work of Ladikou et al,¹⁵ who are the first to show a marked drop in circulating ADAMTS13 levels (49.7% of normal) with the authors stating “we speculate that excess release of VWF seen in COVID-19

patients leads to depletion of ADAMTS13 and contributes to the prothrombotic state.”

If thromboembolic complications play a significant role in the pathogenesis of the disease as it is now widely accepted, then targeting this seems a natural course of action. In the recent propensity score-matched study by Tremblay et al¹⁶ there was no difference in the outcome of patients who were on prior anti-platelet or anti-coagulant medication compared to those who were not. This may suggest that that standard approaches to anti-aggregation and or anti-coagulation are insufficient. This could be explained by the fact that the ULVWF strings mentioned earlier are sufficient to occlude vessels alone and this will not be inhibited by standard treatments.

Taking all of this into account we believe that there is now compelling evidence for a critical role of VWF in COVID-19. Rather than merely representing a biomarker we believe it plays a pivotal role in the pathogenesis and prognosis of the disease. Targeting VWF, particularly early in the disease course, through agents such as caplacizumab or anfibatide, both of which inhibit binding of platelets to the VWF at the glycoprotein IX-Ib receptor, presents an attractive opportunity that is yet to be explored (Figure 1).

CONFLICTS OF INTEREST

PB, DH, and OJ have approached Sanofi to trial caplacizumab in COVID-positive patients. MM, GC, OS, and LM have no conflicts to disclose.

AUTHOR CONTRIBUTIONS

PB: originator of concept, manuscript drafting, editing. MM, GC, OS, LM, DH: critical review of manuscript, editing. OJ: guarantor.

KEYWORDS

COVID 19, anfibatide, caplacizumab, endotheliopathy, microthrombosis

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REFERENCES

1. Rauch A, Labreuche J, Lassalle F, et al. Coagulation biomarkers are independent predictors of increased oxygen requirements in COVID-19. *J Thromb Haemost*. 2020. <https://doi.org/10.1111/jth.15067>
2. Tee A, Wong A, Yusuff T, et al. Contrast-enhanced ultrasound (CEUS) of the lung reveals multiple areas of microthrombi in a COVID-19 patient. *Intens Care Med*. 2020;46(8):1660-1662.
3. Jung EM, Stroszczyński C, Jung F. Contrast enhanced ultrasonography (CEUS) to detect abdominal microcirculatory disorders in severe cases of COVID-19 infection: First experience. *Clin Hemorheol Microcirc*. 2020;74(4):353-361.
4. Carsana L, Sonzogni A, Nasr A, et al. Pulmonary post-mortem findings in a series of COVID-19 cases from northern Italy: a two-centre descriptive study. *Lancet Infect Dis*. 2020.
5. Fox SE, Akmatbekov A, Harbert JL, et al. Pulmonary and Cardiac Pathology in Covid-19: The First Autopsy Series from New Orleans. *medRxiv*. 2020;2020.04.06.20050575.
6. Ciceri F, Beretta L, Scandroglio AM, et al. Microvascular COVID-19 lung vessels obstructive thromboinflammatory syndrome (MicroCLOTS): an atypical acute respiratory distress syndrome working hypothesis. *Crit Care Resusc*. 2020.
7. Yamamoto K, de Waard V, Fearn C, et al. Tissue distribution and regulation of murine von Willebrand factor gene expression in vivo. *Blood*. 1998;92:2791-2801.
8. Dong J, Moake JL, Nolasco L, et al. ADAMTS-13 rapidly cleaves newly secreted ultralarge von Willebrand factor multimers on the endothelial surface under flowing conditions. *Blood*. 2002;100:4033-4039.
9. Padilla A, Moake JL, Bernardo A, et al. P-selectin anchors newly released ultralarge von Willebrand factor multimers to the endothelial cell surface. *Blood*. 2004;103:2150-2156.
10. Huang J, Roth R, Heuser JE, et al. Integrin alpha(v)beta(3) on human endothelial cells binds von Willebrand factor strings under fluid shear stress. *Blood*. 2009;113:1589-1597.
11. Bernardo A, Ball C, Nolasco L, et al. Effects of inflammatory cytokines on the release and cleavage of the endothelial cell-derived ultralarge von Willebrand factor multimers under flow. *Blood*. 2004;104:100-106.
12. Westwood J-P, Langley K, Heelas E, et al. Complement and cytokine response in acute Thrombotic Thrombocytopenic Purpura. *Br J Haematol*. 2014;164:858-866.
13. Ellinghaus D, Degenhardt F, Bujanda L, et al. Genomewide association study of severe Covid-19 with respiratory failure. *N Engl J Med*. 2020.
14. Jenkins PV, O'Donnell JS. ABO blood group determines plasma von Willebrand factor levels: a biologic function after all? *Transfusion*. 2006;46:1836-1844.
15. Ladikou EE, Sivaloganathan H, Milne KM, et al. Von Willebrand factor (vWF): marker of endothelial damage and thrombotic risk in COVID-19? *Clin Med*. 2020.
16. Tremblay D, van Gerwen M, Alsen M, et al. Impact of anticoagulation prior to COVID-19 infection: a propensity score-matched cohort study. *Blood*. 2020;136(1):144-147.