



Review

The Role of the Tumor Microenvironment (TME) in Advancing Cancer Therapies: Immune System Interactions, Tumor-Infiltrating Lymphocytes (TILs), and the Role of Exosomes and Inflammasomes

Atef M. Erasha ^{1,†}, Hanem EL-Gendy ^{2,†}, Ahmed S. Aly ³ , Marisol Fernández-Ortiz ^{4,*} and Ramy K. A. Sayed ⁵

¹ Department of Anatomy and Embryology, Faculty of Veterinary Medicine, Sadat City University, Sadat City 32897, Egypt; atef.areisha@vet.usc.edu.eg

² Department of Pharmacology, Faculty of Veterinary Medicine, Sadat City University, Sadat City 32897, Egypt; hanem.elgendy@vet.usc.edu.eg

³ Department of Animal Production, Faculty of Agriculture, Ain Shams University, Cairo 11241, Egypt; a_yousef129@agr.asu.edu.eg

⁴ Greehey Children's Cancer Research Institute, University of Texas Health Science Center San Antonio, San Antonio, TX 78229, USA

⁵ Department of Anatomy and Embryology, Faculty of Veterinary Medicine, Sohag University, Sohag 82524, Egypt; ramy.kamal@vet.sohag.edu.eg

* Correspondence: fernandezort@uthscsa.edu

† These authors contributed equally to this work.

Abstract: Understanding how different contributors within the tumor microenvironment (TME) function and communicate is essential for effective cancer detection and treatment. The TME encompasses all the surroundings of a tumor such as blood vessels, fibroblasts, immune cells, signaling molecules, exosomes, and the extracellular matrix (ECM). Subsequently, effective cancer therapy relies on addressing TME alterations, known drivers of tumor progression, immune evasion, and metastasis. Immune cells and other cell types act differently under cancerous conditions, either driving or hindering cancer progression. For instance, tumor-infiltrating lymphocytes (TILs) include lymphocytes of B and T cell types that can invade malignancies, bringing in and enhancing the ability of immune system to recognize and destroy cancer cells. Therefore, TILs display a promising approach to tackling the TME alterations and have the capability to significantly hinder cancer progression. Similarly, exosomes and inflammasomes exhibit a dual effect, resulting in either tumor progression or inhibition depending on the origin of exosomes, type of inflammasome and tumor. This review will explore how cells function in the presence of a tumor, the communication between cancer cells and immune cells, and the role of TILs, exosomes and inflammasomes within the TME. The efforts in this review are aimed at garnering interest in safer and durable therapies for cancer, in addition to providing a promising avenue for advancing cancer therapy and consequently improving survival rates.

Keywords: cancer therapy; exosomes; immune cells; inflammasomes; tumor infiltrating lymphocytes (TILs); tumor microenvironment (TME)



Academic Editor: Alessandro Poggi

Received: 21 February 2025

Revised: 10 March 2025

Accepted: 14 March 2025

Published: 18 March 2025

Citation: Erasha, A.M.; EL-Gendy, H.; Aly, A.S.; Fernández-Ortiz, M.; Sayed, R.K.A. The Role of the Tumor Microenvironment (TME) in Advancing Cancer Therapies: Immune System Interactions, Tumor-Infiltrating Lymphocytes (TILs), and the Role of Exosomes and Inflammasomes. *Int. J. Mol. Sci.* **2025**, *26*, 2716. <https://doi.org/10.3390/ijms26062716>

Copyright: © 2025 by the authors.

Licensee MDPI, Basel, Switzerland.

This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

According to the National Cancer Institute (NCI) and the Centers for Disease Control and Prevention (CDC), cancer represents a significant global health problem, encompassing over 100 different types that may originate in specific tissues and spread throughout the

body. It is among the leading causes of morbidity and death worldwide [1,2]. Scientists developed a variety of cancer therapies, such as chemotherapy, radiotherapy, surgical therapy, and immunotherapy, which are currently available. These treatments can harm healthy cells and may cause other adverse effects. Consequently, scientists are investigating innovative approaches to target and eliminate tumor cells while selectively minimizing side effects.

The immune system has a crucial defensive role against tumors and can inhibit their progression. Initially, immunotherapy exhibited limited effectiveness, causing a lack of interest in cancer immunotherapy. However, later advances revealed that immune treatments may result in durable responses in a specific subgroup of patients. For example, interleukin-2 (IL-2) and adoptive transfer of autologous tumor-infiltrating lymphocytes (TILs) were found to increase the chances of long-term survival, even in patients with a poor prognosis [3]. Additionally, TILs hold significant promise as a form of cancer therapy. These lymphocytes can recognize and destroy cancer cells (tumoricidal activity) and halt tumor progression (tumorostatic role). As a result, the level of these lymphocytes can be used as an indicator of an individual's immune response to tumors [4]. Identifying and quantifying TILs is possible through quantifying CD3 markers or employing techniques like gene expression microarray analysis or RNA sequencing. The presence of tumor-infiltrating T lymphocytes can be increased when anti-programmed cell death protein 1 (anti-PD1) therapy is utilized by using drugs like Pembrolizumab, Nivolumab, and Cemiplimab [5–7].

Cancer cells can employ different mechanisms to avoid their destruction by the immune system. They can stimulate immune cell suppressors, such as regulatory T cells (Tregs) and myeloid-derived suppressor cells, which inhibit the proliferation of T-cells and can even suppress the TILs [8,9]. Another mechanism includes the upregulation of programmed death ligand-1 (PD-L1), also called B7-H1, which can bind to the programmed death-1 receptor (PD-1) on the surface of T-cells, impairing their activity. In addition, tumor cells enhance the levels of reactive oxygen species and release substances like transforming growth factor- β (TGF- β), IL-10, exosomes, and nitric oxide, all of which display a direct and positive relationship with immunosuppression [10,11]. The heterogeneity of the tumor microenvironment (TME), composed of immune cells, stromal cells, blood vessels, extracellular matrix (ECM), and exosomes, contributes to this immune evasion, thereby impairing the efficacy of the immunotherapy [12–14]. The cellular components of the TME recruit and secrete protective cytokines, leading to treatment resistance. Non-cellular components of the TME, like ECM, as well as conditions that include hypoxia, high acidity and lactate, and the depletion of glucose and amino acids, act as a physical barrier to immunotherapy and impair the normal metabolism of lymphocytes. Exosomes produced by cancer cells in the TME facilitate inflammation, angiogenesis, tumor progression, and metastasis.

Exosomes are microscopic vesicles enclosed by a phospholipid bilayer, and they can be derived or secreted by most living cells. These membranous structures can transport various active biomolecules from one cell to another or from hosts to recipients. Recently, exosomes have emerged as potential vehicles for drug delivery, including both biological and non-biological medications. In this regard, exosomes offer several advantages over liposomes and nanoparticles, including low clearance rates, higher bioavailability (they can easily pass through biological barriers including the blood–brain barrier, placental barrier, and intestinal barrier), minimal immunogenicity, low cumulative toxicity in normal tissues, and the capacity to specifically deliver anti-cancer drugs to cancer cells by utilizing ligand–receptor interaction or endocytosis, thereby addressing drug resistance caused by P-glycoprotein or other multidrug resistance-associated problems [15,16].

Inflammasome-induced inflammation has been associated with the onset and progression of cancer. By activating caspase-1, inflammasomes initiate inflammatory responses that

result in the release of pro-inflammatory cytokines such as IL-1 β and IL-18 [17–19]. These cytokines impact tumor development by altering the TME, regulating immune responses, and facilitating cancer cell survival and metastasis. Inflammasomes can be activated in various cell subpopulations within the TME, including tumor cells, tumor-associated macrophages (TAMs), tumor-associated fibroblasts, and marrow-derived suppressive cells [20–23]. Recent studies highlight the dual role of inflammasomes in the TME, where they can either promote or inhibit tumor progression, depending on the specific inflammasome type and tumor context [24].

Exosomes play a crucial role as mediators of communication between tumor cells, immune cells, and other components of TME. Their influence on inflammasome activation varies, either enhancing or suppressing it, depending on the composition of the exosomal cargo and the cellular context. The impact of exosomes on inflammasome activation is highly complex, with studies indicating both pro-inflammatory and anti-inflammatory effects based on their cellular origin and the specific TME conditions. Exosomes derived from diverse cell types—including immune, epithelial, cartilage, and cancer cells—can modulate inflammasome activity, with immune cell-derived exosomes being the most extensively studied [25]. This review will provide comprehensive insights into the TME, focusing on the dynamic interplay between the immune system and tumors. Moreover, the importance of TILs and the role of exosomes as promising and safer strategies for cancer therapy will be explored. Moreover, this review will also highlight the role of the inflammasome and its activity modulation as a potential therapeutic strategy for cancer treatment [26].

2. Differences in Cell Behavior Between Normal and Cancerous Conditions

Table 1 summarizes the differences in the behavior of different immune cells under normal and tumor conditions and their impact on tumor progression.

Table 1. Differences in the behavior of different immune cells.

Immune Cell	Normal Conditions (Antitumor) (Tumor Regression, Rejection, Apoptosis, Cytotoxic Good Prognosis)	Tumor Conditions (Protumor) (Tumor Growth, Spread, Metastasis, Poor Prognosis)	References
Macrophages	M1 can produce cytokines and establish an environment that enhances immune defense in reaction to inflammation.	M1 can release M1-Th1, which plays a defensive role against tumor cells. M2 macrophages in tumor conditions can release IL-10, angiogenic factors, and tumor growth factor β (TGF- β).	[27–29]
Fibroblasts	Fibroblasts have an important role in the healing process following inflammation or injuries.	Fibroblasts can transform into cancer-associated fibroblasts, which can regulate cytokines, myeloid suppressor cells, and Tregs, thus contributing to the progression and dissemination of tumors.	[30,31]

Table 1. Cont.

Immune Cell	Normal Conditions (Antitumor) (Tumor Regression, Rejection, Apoptosis, Cytotoxic Good Prognosis)	Tumor Conditions (Protumor) (Tumor Growth, Spread, Metastasis, Poor Prognosis)	References
Endothelial cells	Endothelial cells have a role in inflammation, the process of regeneration, and healing through producing substances such as tumor necrosis factor α (TNF α) and other specific interleukins.	Endothelial cells exhibit an altered structure due to angiogenesis, resulting in impaired immune cell function.	[32]
Regulatory T cells (Tregs)	Tregs play a key role in regulating the immune system.	Tregs stimulate the release of IL-10 and TGF- β .	[33,34]
Neutrophils	Neutrophils participate in phagocytosis and generate cytokines.	Neutrophils can be categorized into two types: N1 and N2. N1 exerts an antitumoral effect by attracting IL-8 from tumor cells, and N2 activates a pro-tumoral effect by contributing to angiogenesis.	[35]
Eosinophils	Eosinophils, under normal circumstances, demonstrate antiparasitic actions and contribute to immune responses.	Eosinophils exhibit a tumoricidal role by releasing interleukins such as IL-2 and IL-4.	[36,37]
$\gamma\delta$ T-cells	These T-cells can respond to phosphor antigens and communicate antigens to CD8+ and CD4+ lymphocytes, besides collaborating with natural killer (NK) cells.	$\gamma\delta$ T-cells exhibit the strongest positive correlation with cancer prognosis. These cells also include IL-17-secreting cells, which can trigger the production of vascular endothelial growth factors and other angiogenesis-related factors.	[38,39]
Natural killer (NK) cells	NK cells are the primary antitumor defenders. They enhance the action of T-helper 1 lymphocytes (Th1) and stimulate CD8+ lymphocytes.	NK cells interacting with tumor cells usually express the two CD45 isoforms; CD45RA and CD45RO. The anti-tumor NK cells perform trogocytosis on tumor markers, facilitating their identification.	[40,41]
Dendritic cells (DCs)	DCs attract lymphocytes to antigen-presenting cells (APCs).	DCs draw lymphocytes to tumor-presenting cells, thus, their invasion of tumor cells is associated with delayed cancer progression and, in turn, a favorable prognosis.	[40,42]

Table 1. Cont.

Immune Cell	Normal Conditions (Antitumor) (Tumor Regression, Rejection, Apoptosis, Cytotoxic Good Prognosis)	Tumor Conditions (Protumor) (Tumor Growth, Spread, Metastasis, Poor Prognosis)	References
Type 1 CD8+ T cells	These cells are the primary defense against cancer in humans.	They become activated when tumor antigens are presented by a dendritic cell along with attracting M1 macrophages, T-helper 1 lymphocytes (Th1), and T-helper 9 lymphocytes (Th9) to the tumor cells. Their presence indicates a favorable prognosis in tumor cases.	[43,44]
CD4+ T cells, Th1, Th2	CD4+ T lymphocytes display diverse polarization based on the specific cytokine combinations influencing them. The Th1 polarization is driven by the presence of IFN and IL-12 from M1 macrophages, as well as IL18, IL-27, and IL1, all of which are involved in the anti-tumor defense.	Th2 polarization is driven by specific interleukins (ILs) released by mast cells, NK cells, and CD4+ memory. The Th2 cells respond by producing other cytokines, such as interleukins (ILs)- 4, 5, 10, 13, 25, and 33, impairing the effectiveness of the immune system in malignancies.	[45,46]
B cells	B cells act as a pro-tumoral in some malignancies by releasing IFN- γ and IL-12	B cells stimulate IL-10 and TGF- β release, which have a role in tumor progression.	[34]

3. Cancer Immunoediting

The interaction between the immune system and cancer cells is distinguished by three different phases (Figure 1). In the first phase, referred to as the elimination phase, cancer cells are actively targeted and removed through a process known as immunosurveillance. This phase involves the collaborative efforts of innate and adaptive immunity, working together to recognize and eliminate altered cells before they can manifest clinically. The equilibrium follows, during which transformed cells persist but are held in check and under the control of the immune system. During this stage, the immune system acts as a regulator, with adaptive immunity playing a crucial role in limiting the proliferation of clinically undetectable occult tumor cells while also modifying the immunogenic properties of tumor cells. The final phase (the escape phase) signifies the point at which malignant cells successfully evade immune control, leading to tumor progression. The regulation of these processes heavily relies on the involvement of leukocytes and cytokines. Within a tumor, a wide range of immune cell types can be found, involving neutrophil granulocytes, macrophages, mast cells, DC, NK cells, naive and memory lymphocytes, B cells, and effector T cells, including Th 1, 2, and 17 cells, Tregs, T follicular helper cells and cytotoxic T cells [47].

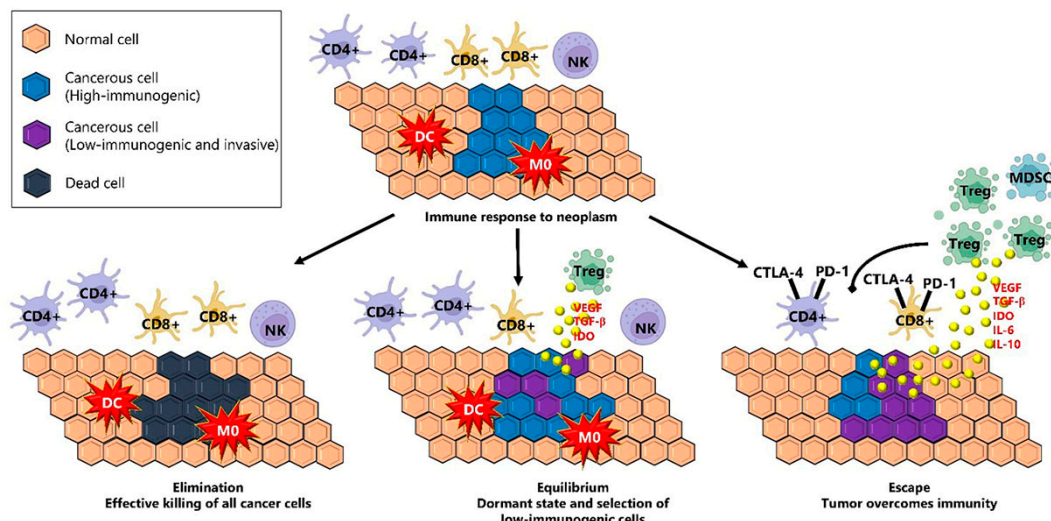


Figure 1. The immunoeediting process includes three stages: elimination, equilibrium, and escape, highlighting different subtypes of immune cells involved in tumors. The figure is adapted from De Mello et al. [47].

4. Phenotype of Tumor-Infiltrating Lymphocyte

Figure 2 displays the different phenotypes of TILs, including T-cells and B-lymphocytes. The clinical outcome is influenced by both the extent of lymphocytic infiltration and the specific infiltrate phenotype. A favorable prognosis is associated with type 1 T-cells, where CD4+ Th1 cells play an essential role in antigen presentation through cytokine secretion. Additionally, CD8+ cytotoxic T-helper (CTL) cells are crucial for tumor elimination [48]. In contrast, type 2, CD4+ Th2 can inhibit CTL activity, promote the proliferation of B-lymphocytes, and may activate an anti-inflammatory immune response that could potentially favor the progression of tumor [49,50].

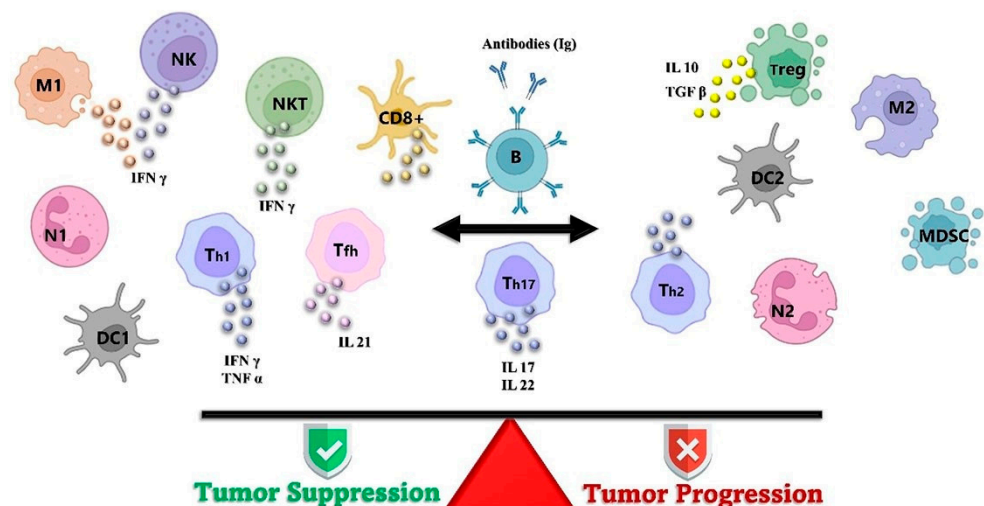


Figure 2. The intracellular interaction between different leukocyte subsets and their predominant roles in either stimulating or inhibiting tumor growth, including myeloid lineage leukocytes, tumor-associated macrophages (M) with both pro- and antitumor properties, various helper T-cell subsets (Th), cytotoxic T cells, regulatory T cells (Tregs), dendritic cells (DCs), natural killer cells (NK), neutrophils (N), B cells, and myeloid-derived suppressor cells (MDSC). The figure is adapted from Salgado et al. [51].

5. Tumor-Infiltrating Lymphocyte (TIL) Therapy

To date, there are three types of adoptive cell therapy (ACT): chimeric antigen receptor T-cell (CAR-T) therapy, TIL therapy, and T cell receptor (TRC) therapy [52]. The last one is the least used due to being restricted by the patient's major histocompatibility complex (MHC) type, high autoimmunity risk, and complexity of engineering [53]. Both CAR-T and TIL are used to treat patients in advanced stages of cancer, including those who are resistant to checkpoint inhibitor therapies. CAR-T therapy has been approved for hematological cancer like acute lymphoblastic leukemia, chronic lymphocytic leukemia, multiple myeloma, and different forms of lymphomas. TIL has shown promising results in treating certain solid tumors [54–58]. This review will focus on this last type of ACT for being directly connected with the TME.

TILs are immune cells, mostly T cells, that migrated from the bloodstream into the TME to recognize and eliminate tumor cells. In ACT, TILs are harvested from the patient's tumor, expanded ex vivo in the laboratory, and reinfused back into the patient. The equilibrium between pro-tumor and anti-tumor responses of TIL is predominantly influenced by the TME, which varies from patient to patient. Factors like immunosuppressive cytokines like TGF- β , immune checkpoint molecules as PD-1 and CTLA-4, TAMs, Tregs, or hypoxia in TME can impair the efficacy of TIL [59]. A key step in TIL therapy that eliminates some of these factors and increases the expansion and functionality of the infused TILs is lymphodepletion. Several findings suggest that lymphodepletion can enhance TIL effectiveness through multiple mechanisms that include the removal of Tregs elevation in host homeostatic cytokines such as IL-7 and IL-15 and reduction in endogenous lymphocytes. This reduction reduces competition for these critical trophic cytokines among the host's lymphocytes [60,61]. Additionally, the adoptively transplanted T lymphocytes are regulated by APCs that activate because of lymphodepletion.

A non-myeloablative (NMA) lymphodepletion regimen is normally used before TIL injection. This regimen typically involves lower doses of chemotherapy or low-dose total body irradiation (TBI). Ongoing research is actively refining lymphodepletion protocols to avoid toxicity and improve patient outcomes in different types of cancer [62–66]. An NMA lymphodepletion regimen with cyclophosphamide and fludarabine caused a 50% objective response rate in patients with metastatic melanoma treated with TIL. A strong long-term persistence of the adoptively transferred cells was also reported [67,68]. Although melanoma has been the main focus of research in this field, there is clinical evidence suggesting that TIL therapy with lymphodepletion may be beneficial for other types of cancer, among them non-small cell lung cancer (NSCLC), ovarian cancer, cervical cancer, colorectal cancer, and renal cell carcinoma (RCC). TIL therapy has also been combined in the clinic with immune checkpoint and BRAF inhibitors [69–74].

6. Combination Therapy with TIL

6.1. Immune Checkpoint Inhibitors

Recent trials have shown promising preliminary results with the combination of TIL treatment and anti-PD-1/PD-L1 antibody therapy [75]. T lymphocytes express immunological checkpoint receptors on their surface, including CTLA-4 (CD152) and PD-1/PD-L1. In cancer patients, CTLA-4 and PD-1 molecules on effector T cells are upregulated, binding to B7-1/B7-2 and PD-L1 on APCs or tumor cells. This binding leads to the inhibition of T cell function, a blockade that can be relieved through the use of anti-CTLA-4 and anti-PD-1 antibodies [76]. The first small molecule inhibitor based on the PD-1/PD-L1 axis, a derivative of the antibiotics sulfamethoxine and sulfamethimazole, was reported [77]. In advanced triple-negative breast cancer (TNBC) patients, the anti-PD-L1 drug atezolizumab has significantly reduced PD-L1-positive (PD-L1+) metastases and increased overall sur-

vival [78]. PD-1 inhibitors include pembrolizumab (Keytruda), nivolumab (Opdivo), and cemiplimab (Libtayo), while PD-L1 inhibitors include atezolizumab (Tecentriq), nivolumab (Bavencio), and durvalumab (Imfinzi). Both PD-1 and PD-L1 inhibitors are effective in treating various cancer types. Additionally, CTLA-4 inhibitors like ipilimumab (Yervoy) and tremelimumab (Imjuno) are monoclonal antibodies that bind to CTLA-4, inhibiting its activity and leading to the activation of TILs [79,80].

6.2. BRAF Inhibitor

The BRAF gene plays a crucial role in cell development and differentiation. In various malignancies, BRAF mutations disrupt the ERK/MAPK signaling pathway, leading to increased cell proliferation. Activating BRAF mutations, primarily V600E, can trigger immune-escape mechanisms, hindering cells less responsive to T-cell immune responses. Notably, the BRAF inhibitor vemurafenib can diminish linked immunosuppressive signals, enhance lymphocyte infiltration, and reduce the prevalence of immunosuppressive cells [81,82].

7. History of Exosomes

Exosomes are microscopic extracellular vesicles (EVs) that emerge from early endosomes (Figure 3). Exosomes were first found in the maturing mammalian reticulocyte by Stahl and their team in 1983 and subsequently by Johnstone and colleagues in the same year. Hence, initial reports on exosomes emerged in the mid-1980s [83,84]. In the beginning, researchers believed exosomes were only “garbage bags” used for disposing of undesired components [85]. However, accumulating evidence suggests that exosomes play a significant role in various cellular processes, providing a unique mode of intercellular interaction and influencing both pathological and physiological functions [86,87]. In 1991, Rose Johnstone demonstrated the presence of both the nucleoside transporter and the transferrin receptor in exosomes [88]. The researchers noticed that specific cellular stressors can lead to the internalization and shedding of these membrane components at different periods [89]. Several publications from the 1980s and 1990s focused on quantifying EVs and highlighted the changes in EV levels in various diseases. For example, Lee et al. [90] study on elevated microparticles in temporary brain ischemia and other infarctions marked the phenomena, which have since been examined in conditions such as angina [91] and Crohn’s disease [92]. Researchers also identified the capacity of immune cell EVs to display antigens [93], leading to the use of EVs as anti-tumor vaccines. Indeed, this work motivated the Amigorena lab to investigate whether DCs release EVs that, once loaded with tumor peptides, can effectively target tumors [94] paving the way for clinical trials in the subsequent decade [95]. This was a significant development as it highlighted the potential of EVs to participate in biological processes. Together, the recognition that EVs might have physiological roles, play as biomarkers, and hold therapeutic potential sparked a surge of interest in EVs in the early twenty-first century.

Early reviews on EV biology emerged in the decade following the 2000 [96]. Researchers delved deeper into EV nature, examining their proteome and lipidome across various cell types [97]. Mackenzie and colleagues [71] highlighted the crucial role of immune cell-produced EVs in immune system function [98]. The growing interest in EVs allowed for their exploration of anti-tumor therapies [99] and a better understanding of their immune system functions [100]. Remarkably, functional nucleic acid transfer was demonstrated [101], and EV-mediated communication was observed in plant cells as well [102]. With the rising interest in EV-based and stem cell therapies in 2009 [103], studies on vesicles derived from mesenchymal stem cells multiplied, broadening EVs’ therapeutic possibilities. In contrast, the signals received by the cell of origin can impact

exosome synthesis and content. One assumption proposes that tumor cells can adapt to a hypoxic microenvironment by releasing exosomes that stimulate angiogenesis or facilitate metastasis to more tumor-favorable environments. This assumption received subsequent support from various pieces of evidence [104].

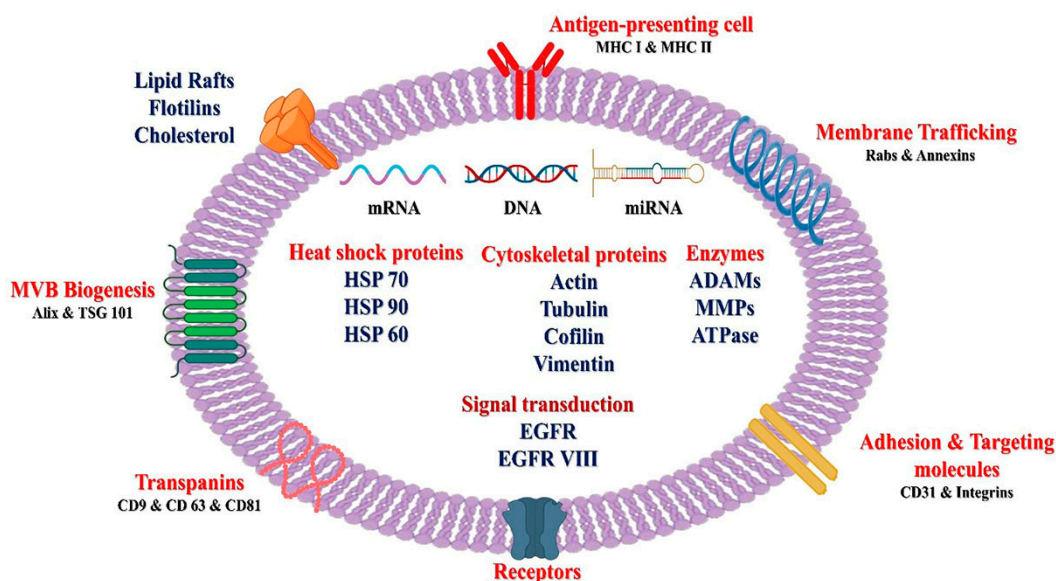


Figure 3. Typical structure of exosomes, nanosized extracellular vesicles enclosed by a phospholipid bilayer, encompassing various proteins (Integrins, TSG 101, Alix, HSP), nucleic acids (DNA, mRNA, miRNA), and other receptors. The figure is adapted from Kumar et al. [105].

8. Exosomal Role in Cancer Therapy

Exosomes, nanometer-sized vesicles enclosed by a lipid bilayer (Figure 3), are released by various cell types and are present in most bodily fluids, including breast milk, blood, saliva, bile, urine, pancreatic juice, and peritoneal and cerebrospinal fluids [105,106]. Exosomes exhibit a tenfold greater propensity for internalization and adhesion to tumor cells in comparison to liposomes of corresponding size, which reinforces their enhanced suitability for cancer-targeting [107]. In addition, exosomes tend to accumulate more in tumor tissues with inadequately developed blood vessels than in healthy tissues due to elevated permeability and retention effects. This characteristic enables exosomes to distribute drugs more efficiently, effectively reaching most solid tumors. Moreover, exosomes can be modified to transport tumor-targeting peptides, antibodies, or proteins for the precise delivery of drugs and therapeutic nucleic acids. These qualities firmly establish exosomes as formidable candidates for targeted cancer therapy.

Recently, scientists improved the precision of targeting tumors by employing a magnetic method. They achieved this by connecting superparamagnetic-conjugated transferrin to blood exosomes with transferrin receptors along with applying an external magnet at the tumor site. In particular, Qi and his research team created magnetically guided exosomes, resulting in suppressing the tumor growth effectively [108].

In contrast to the administration of free drugs in animal tumor models, the delivery of chemotherapeutics-loaded exosomes has shown significant enhancements in their anti-tumor effects. For example, drugs such as Doxorubicin, Paclitaxel, and Withaferin, which are frequently utilized in cancer treatment, can lead to adverse effects on animal tissues. Nevertheless, when these drugs are encapsulated in exosomes (exosome-delivered chemotherapeutics), they mitigate the side effects and deliver a more potent and prolonged therapeutic impact [15,109,110].

9. Dualist Actions of Exosomes in Carcinogenesis

Exosomes serve as a natural transporter of a wide array of bioactive compounds derived from donor cells, involving lipids, proteins, and different types of RNAs such as long non-coding RNAs (lncRNA), microRNAs, inhibitors, and antibodies [106]. Exosomes can transmit these molecules to nearby as well as distant recipient cells. Consequently, exosomes garnered significant attention as a potential natural carrier of cancer therapies. The impact of exosomes on cancer growth and progression varies depending on their cellular source, influencing the course of the disease. Exosomes secreted by cancer cells are implicated in shaping the pre-metastatic microenvironment, facilitating tumor advancement, immune evasion, anti-apoptotic signaling, angiogenesis, treatment resistance, and various other processes [111–113]. In contrast, exosomes originating from normal cells, such as T and B lymphocytes and DCs, play a critical role in preventing tumor progression [114,115].

10. The Origin of Exosomes

The source of exosomes dictates whether they promote or inhibit tumor growth, as illustrated in Figure 4.

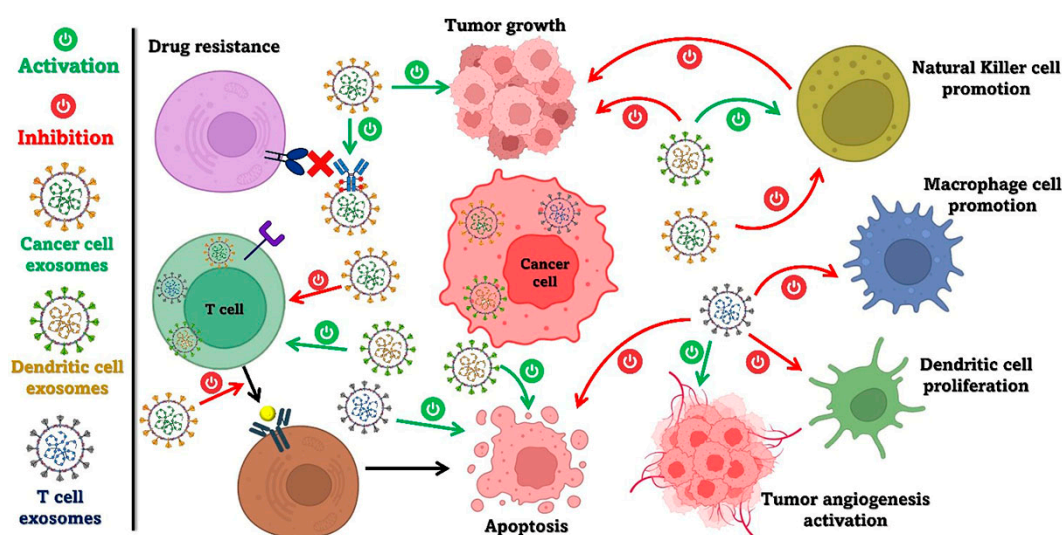


Figure 4. Origins of exosomes and their impact on tumor progression. The figure is adapted from Awadasseid et al. [116].

10.1. Exosomes Released from Tumor Cells

Exosomes originating from tumors can exhibit diverse functional roles, contingent upon their cellular origin and the surrounding environmental conditions. For instance, exosomes derived from rat pancreatic adenocarcinoma have been observed to activate cytotoxic T cell (CTL) responses specific to tumor antigens while simultaneously inhibiting leukocyte proliferation by downregulating the Zeta chain of T cell receptor-associated protein kinase 70 (ZAP70) and extracellular signal-regulated kinases 1,2 (ERK1,2) [117]. Additionally, exosome proteins depleted of miRNA can function as agonists, preferentially activating DCs and cytokine-induced killer cells [118]. The presence of leukemia cell-derived exosomes induces the secretion of TNF and IL-12p70 by DCs [119], promoting T cell proliferation and elevating IFN levels, thereby enhancing cytotoxic T lymphocyte (CTL) activity via the FasL/Fas signaling pathway in renal cancer [120]. On the contrary, exosomes derived from breast cancer can impair NK cell cytotoxicity and hinder the proliferation of CD8⁺ and CD4⁺ T cells, potentially diminishing the immune system's effectiveness in fighting cancer [121]. Moreover, tumor-released exosomes in head and neck cancer have been demonstrated to induce a suppressor phenotype in CD8⁺ T cells, primarily

through the cooperative actions of exosomal components like RNA and galectin1 [122]. Additionally, exosomes derived from melanoma cells alter the transcriptome of CTLs, ultimately diminishing cytotoxic immune responses [123].

10.2. Exosomes Derived from DCs

The DCs hold a pivotal role in cancer immunotherapy due to their capability to capture and display tumor-associated antigens, making them essential contributors to tumor immunity. However, their effectiveness in combating low-immunogenic tumors has remained unsatisfactory. Implications such as the activation of Tregs, limited antigen uptake efficiency, and antigen availability have posed challenges [124]. Recent research suggests that exosomes can serve as an ideal source of antigens for DC vaccinations, emphasizing the need to explore how exosome-based DC vaccines generate anti-tumor immunity before establishing their suitability as tumor antigens for DC vaccine-based immunotherapy [125].

The DCs are highly efficient APCs known to release exosomes that can trigger potent anti-cancer effects. Exosomes produced by DCs, containing chaperones like MHC I, MHC II, HSP70-90, and CD86, can stimulate CD4⁺ and CD8⁺ T cells [126,127]. The secretion of exosomal peptide MHC I is transferred to CD8⁺ T cells under the influence of released IL-2 and exosomal CD80, leading to increased proliferation of CD8⁺ T cells and the production of a more robust anti-tumor immune response in vivo [128]. Several studies have observed the activation of CD8⁺ and CD4⁺ T cells when exposed to DCs exosomes associated with the induction of an anti-tumor immune response in vivo through exosomal CD80 and endogenous IL-2 [129,130]. Furthermore, in mice with hepatic cell carcinoma, exosomes derived from alpha-fetoprotein-expressing DCs induced the production of more IFN-expressing CD8⁺ T cells, triggering higher levels of IFN and IL-2, with reduction in CD25⁺ Foxp3⁺ Tregs, TGF- β , and IL-10 levels [131].

10.3. Exosomes Derived from B Lymphoma Cell

Exosome-based DC vaccines can induce clonal proliferation of T cells through exposure to exosomes derived from diffuse large B cell lymphoma cells [132]. Conversely, exosomes from B cell lymphoma cells have been found to induce apoptosis in CD4⁺ T lymphocytes via MHC II [133]. B lymphoma cells induced by heat shock released exosomes with elevated levels of HSP90 and HSP60, along with heightened immunogenicity molecules like MHC I, MHC II, CD86, CD40, RANTES, and IL-1. These exosomes effectively stimulate CD8⁺ T cells, yielding an anti-cancer effect [134]. When DCs interact with exosomes from B cell lymphoma cells, they can enhance the activation of T cells, leading to the release of TNF- α and IL-6, while simultaneously decreasing the production of immunosuppressive cytokines like IL-4 and IL-10 [132].

10.4. Exosomes Derived from T Lymphocytes

Immunotherapy is a rapidly emerging and promising treatment approach that utilizes genetically modified T cells to express the chimeric antigen receptor [135,136]. There are two types of T cells, CD8⁺ cytotoxic T lymphocytes (CTLs) and CD4⁺ helper T cells. CD8⁺ CTLs play a crucial role in supporting the body's defense against intracellular infections and tumor cells by directly binding to antigens through MHC I. In addition to their direct killing of tumor cells, activated CD8⁺ T cells can indirectly destroy tumor cells by releasing exosomes [137]. In a mouse model of melanoma, intratumoral delivery of exosomes derived from activated CD8⁺ T cells effectively hindered tumor invasion and metastasis mediated by fibroblastic stroma [138]. It has been discovered that T cell exosomes expressing the CD63 protein contain specific miRNAs that regulate immune responses and immune system development, playing a pivotal role in enhancing interaction between antigen-presenting T cells [139]. CD8⁺ T cells and CD63⁺ exosomes produce similar anti-infective effects [140].

Most surface markers of CD4⁺ helper T cells can interact with MHC II on the surface of APCs to initiate, modify, or support immune responses. Exosomes produced by CD4⁺ T cells have been established as a fundamental pathogenic mechanism in several inflammatory diseases [141]. These exosomes can employ target cells through CD4–MHC interactions, ultimately leading to the elimination of immune-deficient cells [142]. Exosomes derived from activated CD4⁺ helper T cells can also act as potent inducers of phagocyte and B cell activation, which support the inflammatory response [143].

10.5. Exosomes Derived from NK Cells

NK cells function as the body's initial defense against various disorders. Exosomes released by NK cells are equipped with cytotoxic proteins such as FasL and perforin, besides characteristic NK markers like CD56 [144]. Additionally, NK exosomes possess the ability to infiltrate tumor tissues directly, enabling them to exert their cytolytic effects. This capability overcomes the challenge of NK cells not naturally homing in on tumor sites [145]. Furthermore, NK cells activate both caspase-independent and caspase-dependent cell death pathways [146].

10.6. Exosomes Derived from Myeloid-Derived Suppressor Cell

A diverse group of immature myeloid cells referred to as myeloid-derived suppressor cells (MDSCs) possesses a remarkable ability to inhibit the cytotoxicity of T/NK cells, rendering them a significant obstacle in cancer immunotherapy [147]. Activation of MDSCs under cancerous conditions through utilizing various pharmacological drugs has been extensively investigated. Recent studies have begun to shed light on the immunosuppressive functions of MDSC-derived exosomes within the microenvironment of both cancer and autoimmune disorders [148]. Exosomes derived from MDSCs have been found to carry cargo that matches their role in mediating immunosuppression [149]. In mice bearing breast tumors, augmented MDSC-derived miR-126a⁺ exosomes have been shown to potentially stimulate metastasis and confer resistance to therapy [150].

10.7. Exosomes Derived from Tumor-Associated Macrophage

Macrophages in the TME can diminish the activity of T cells, paving the way for cancer cells to escape immunity. Remarkably, TAMs often exhibit two competing phenotypes: the anti-tumorigenic M1 subtype and the pro-tumorigenic M2 subtype [151] as previously discussed in this review. Accumulating pieces of evidence suggest that TAMs release exosomes that influence various aspects of cancer biology and the immune response [152]. These exosomes, originated from TAMs, can create an immune-suppressive environment and enhance the progression of ovarian cancer by transferring miRNAs into CD4⁺ T cells. Moreover, exosomes derived from M2 macrophages transmit oncogenic miRNAs, promoting cancer cell invasion, migration, and resistance to chemotherapy [153]. These TAM-derived exosomes primarily function as markers for the polarization of Th1 and M1 subtypes, containing contents that enhance pro-inflammatory signaling and the immune response [154].

10.8. Exosomes Derived from Mast Cells (MCs)

Exosomes, with their essential roles in RNA and protein transfer, intercellular interaction, and immunoregulation, can also be released by MCs. Remarkably, MC-originated exosomes have been demonstrated to compromise intestinal barrier function, likely due to the delivery of miRNAs to targeted cells [155]. Recent research has revealed that lung cancer cells can internalize MC-derived exosomes, subsequently promoting cancer cell growth by transferring the KIT protein [156]. Additionally, the biological processes of DCs, T cells, and B cells can be influenced by exosomes produced by MCs. For instance, exosomes from MCs

that express CD63 and OX40L have been found to enhance the communication between OX40L and OX40, resulting in the proliferation and development of CD4⁺ Th2 cells [157]. Furthermore, MC-derived exosomes stimulate immature DCs to up-regulate molecules such as MHC II, CD40, CD80, and CD86, enabling T cells to present antigens and initiate the development of immune responses targeted against specific antigens. The potential anti-cancer properties of MC-derived exosomes are currently under investigation [98].

10.9. Exosomes Derived from Neutrophils

Exosomes released by neutrophils have been demonstrated to adhere to and degrade the extracellular matrix through the actions of neutrophil elastase (NE) and the integrin Mac-1, facilitating the progression of inflammatory diseases [158]. In contrast, Li and colleagues recently made a remarkable discovery, revealing that these exosomes strongly inhibit the proliferation and migration of endothelial cells, thereby hindering pathological angiogenesis in immunological diseases [159]. Additionally, Vargas and co-authors tentatively confirmed the presence of the tumor susceptibility gene 101 in neutrophil-derived exosomes [160]. However, to the best of our understanding, there is a lack of pertinent research to elucidate the underlying molecular mechanisms of neutrophil-derived exosomes in the regulation of anti-cancer immune responses.

Table 2 summarizes the various sources of exosomes.

Table 2. Summary of the origins of the exosomes and their influence on tumorigenesis.

Origin of Exosomes	Mechanism and Effect Observed	References
Tumor cells	Exosomes can exhibit diverse functional roles, contingent upon their cellular origin and the surrounding environmental conditions. They activate cytotoxic T cell (CTL) responses specific to tumor antigens while simultaneously inhibiting leukocyte proliferation by downregulating ZAP70 and ERK1, 2. Also, they can impair natural NK cell cytotoxicity and hinder the proliferation of CD8 ⁺ and CD4 ⁺ T cells, potentially diminishing the immune system's effectiveness in fighting cancer.	[117,120]
Dendritic cells (DCs)	Exosomes can trigger potent anti-cancer effects. Exosomes produced by DCs, containing chaperones like MHC I, MHC II, HSP70-90, and CD86, can stimulate CD4 ⁺ and CD8 ⁺ T cells. Exosomes derived from alpha-fetoprotein-expressing DCs induce the production of more IFN-expressing CD8 ⁺ T cells, triggering IFN and IL-2 levels, and reduce the levels of CD25 ⁺ Foxp3 ⁺ Tregs, TGF, and IL-10.	[126,127,131]
B-lymphoma cell	Exosomes induce apoptosis in CD4 ⁺ T lymphocytes via MHC II. B lymphoma cells induced by heat shock released exosomes with elevated levels of HSP90 and HSP60, along with heightened immunogenicity molecules, and these exosomes effectively stimulate CD8 ⁺ T cells, yielding an anti-cancer effect. When DCs interact with exosomes from B cell lymphoma cells, they can enhance the activation of T cells, leading to the release of TNF- α and IL-6 while simultaneously decreasing IL-4 and IL-10 production.	[132–134]

Table 2. Cont.

Origin of Exosomes	Mechanism and Effect Observed	References
T-lymphocytes	Exosomes derived from activated CD8+ T cells effectively hinder tumor invasion and metastasis mediated by fibroblastic stroma. Additionally, T cell exosomes express the CD63 protein and contain specific miRNAs that regulate immune responses and immune system development, playing a pivotal role in enhancing interaction between antigen-presenting T cells. Exosomes produced by CD4+ T cells can employ target cells through CD4–MHC interactions, ultimately leading to the elimination of immune-deficient cells	[138,139,142]
Natural killer (NK) cells	Exosomes are equipped with cytotoxic proteins such as FasL and perforin, besides characteristic NK markers like CD56. Additionally, NK-derived exosomes possess the ability to infiltrate tumor tissues directly, enabling them to exert their cytolytic effects.	[144,145]
Myeloid-derived suppressor cell (MDSC)	Exosomes derived from MDSCs carry cargo that matches their role in mediating immunosuppression. Additionally, MDSC-derived miR-126a+ exosomes have been shown to stimulate metastasis and confer resistance to therapy.	[149,150]
Tumor-associated macrophages (TAMs)	Exosomes originated from TAMs can create an immune-suppressive environment and enhance the progression of ovarian cancer by transferring miRNAs into CD4+ T cells. Moreover, M2 macrophage-derived exosomes transmit oncogenic miRNAs, promoting cancer cell invasion, migration, and resistance to chemotherapy.	[152,153]
Mast cells (MCs)	MC-derived exosomes promote cancer cell growth by transferring the KIT protein. Additionally, exosomes from MCs that express CD63 and OX40L result in the proliferation and development of CD4+ Th2 cells. Furthermore, MC-derived exosomes stimulate immature DCs to up-regulate molecules such as MHC II, CD40, CD80, and CD86, enabling T cells to present antigens and initiate the development of immune responses.	[156,157]
Neutrophils	Exosomes released by neutrophils adhere to and degrade the extracellular matrix through the actions of neutrophil elastase (NE) and the integrin Mac-1, facilitating the progression of inflammatory diseases. Furthermore, these exosomes inhibit the proliferation and migration of endothelial cells, thereby hindering pathological angiogenesis.	[158,159]

11. History of Inflammasomes

The term inflammasome was first introduced in 2002 by Dr. Jürg Tschopp [161]. This word describes an intracellular protein complex involved in immune responses. NLRP1 inflammasome was the first complex that was characterized, consisting of the Nod-like receptor (NLR) NLRP1, the adaptor protein ASC (Apoptosis-associated speck-like protein containing a CARD), and inflammatory caspases, for example, caspase-1 and -5. Inflammasomes belong to the Nod-like receptor (NLR) family. This group of intracellular pattern recognition receptors (PRRs) detects cellular stress, pathogens, and damage-associated molecular patterns (DAMPs). The inflammasomes cause inflammatory responses through two mechanisms: one, by activating caspases which process and release pro-inflammatory

cytokines, for example, IL-1 β and IL-18, and, two, by inducing the process of pyroptosis. The term pyroptosis describes a form of inflammatory cell death characterized both by plasma membrane rupture and cytokine release [162]. Since the discovery of the NLRP1 inflammasome, over twelve different inflammasomes have been characterized, including NLRP1, NLRP2, NLRP3, NLRP6, NLRP7, NLRP9, NLRP12, NAIP/NLRC4, AIM2, IFI16, CARD8, and PYRIN [163]. Most of these belong to the NLR family, with 22 members in humans and 34 in mice. They are characterized by a nucleotide-binding and oligomerization domain (NOD) and a C-terminal leucine-rich repeat (LRR). The family members are classified according to their N-terminal domains, which can include NLRA, BIR, CARD, or Pyrin domains.

NLRP1 and NLRP3 are among the most studied inflammasomes. NLRP3 is activated by a broad range of triggers, among which are MAMPs (microbe-associated molecular patterns) and DAMPs, such as particulate matter, cholesterol crystals, and cellular stress signals [164]. The precise ligand for NLRP3 remains difficult to characterize, although it is known that NLRP3 activation requires post-translational modifications such as ubiquitination, phosphorylation, and interactions with NEK7. On the other hand, the activation mechanisms of other inflammasomes, such as AIM2, NAIP/NLRC4, and Pyrin, are better understood. Cytosolic double-stranded DNA from both microbial and host origins activates AIM2 [165,166]. NAIP/NLRC4 responds to bacterial components such as flagellin and proteins of the bacterial type III secretion system [167,168]. Pyrin senses alterations in cellular dynamics caused by infection-induced modifications to Rho family small GTPases [169].

The study of inflammasomes has increased in recent years and findings have connected inflammasome dysregulation to various diseases, among them cancer. Inflammasomes play a wide range of roles in tumorigenesis and immune responses within the TME. These roles can be both pro-tumorigenic and anti-tumorigenic, depending on the type of cancer, cancer etiology, and cells activating the inflammasome pathway within the TME. This dual role highlights the complexity of inflammasomes in cancer biology, spurring a growing body of research into inflammasome-targeted therapies for cancer treatment [170,171].

12. Inflammasomes in Cancer Therapy

Inflammation caused by inflammasomes has been linked to the development and progression of cancer. Inflammasomes trigger inflammatory responses through caspase-1 activation, leading to the release of pro-inflammatory cytokines like IL-1 β and IL-18 [17–19]. These cytokines influence tumor development by modifying the TME, modulating immune responses, and promoting cancer cell survival and metastasis. Therefore, targeting inflammasomes with specific inhibitors or activators offers a promising therapeutic approach for treating cancer [26].

12.1. Inhibition of Inflammasomes

The most prevalent therapeutic approach against cancer consists of inhibiting inflammasome activation [172]. The principal target of these attempts to restrict inflammasome activation is NLRP3 [26,173]. Research has shown this inflammasome plays an important role in many cancer types. One of the main challenges researchers face when developing these therapies is determining whether to prioritize the upstream or downstream components of the inflammasomes [26,174–176]. The upstream targets, such as NLRP3, could allow for more precise interventions by specifically inhibiting certain inflammasome activations without affecting other sensors like AIM2, NLRP1, and NLRC4, which detect pathogens and other stimuli. On the other hand, downstream targeting of IL-1 β and IL-18 could be less selective overall but more effective in suppressing the pro-inflammatory signals that promote cancer progression.

One of the two most studied NLRP3 inhibitors are MCC950 and OLT1177, which selectively block the NACHT domain of NLRP3, avoiding its activation and subsequent inflammatory response. MCC950 has shown promising results in treating various cancer types, including pancreatic cancer, colorectal carcinoma, and head and neck squamous cell carcinoma [177–179]. OLT1177 has been used against melanoma [180,181]. Similarly, other inhibitors like CY-09, tranilast, and oridonin also target the NACHT domain to inhibit inflammasome activation. Some of these treatments also show anti-tumor effects in preclinical models [178,182–185]. Although NLRP3 inhibitors have worked to reduce inflammation, their combination with other therapeutic methods, such as immune checkpoint inhibitors, such as anti-PD-1, may enhance their anti-tumor effects by modulating the TME [186,187]. For example, OLT117 has been shown to disrupt the IL-1 β /IL-6/STAT3 axis in the TME, which decreases the immunosuppressive activity of MDSCs and improves anti-tumor immunity when combined with anti-PD-1 therapy [181].

Besides NLRP3 inhibitors, other strategies prioritize the targeting of other inflammasome components or proteins involved in their activation. For example, glycyrrhizin has been shown to inhibit both NLRP3 and AIM2 inflammasomes [188]. Methylene blue is a broad-spectrum inflammasome inhibitor that affects multiple inflammasome pathways, among them NLRP3, NLRC4, and AIM2 [189]. Additionally, andrographolide has shown promising results in inhibiting AIM2 in the prevention of colitis-associated cancer [190]. AIM2 and NLRC4 are involved in pathogen detection as well as immune cell activation. Targeting the NLRP3 inflammasome, therefore, could offer a more selective therapeutic approach, which reduces the risk of unwanted suppression of the immune system [191].

Alternatively, another approach to targeting inflammasomes consists of inhibiting caspase-1, the downstream effector that mediates pyroptosis. The caspase-1 inhibitor VX-765 has shown anti-tumor effects through the inhibition of pyroptosis and reduction in IL-1 β secretion in non-small cell lung cancer (NSCLC) models [192]. The effects of caspase-1 inhibition on tumor growth, however, are context-dependent. Some studies seem to indicate that inhibiting pyroptosis could allow cancer cells to evade immune surveillance. Others show potential benefits in specific immune subsets such MDSCs [193–196].

12.2. Activation of Inflammasomes for Cancer Immunotherapy

Although most inflammasome-targeting strategies have focused on inhibition, an alternative approach to cancer therapy may be the opposite method: the activation of inflammasomes. This approach may work because inflammasome activation can trigger pyroptosis in cancer cells, releasing pro-inflammatory cytokines such as IL-1 β and IL-18 that can aid in anti-tumor immune responses. The compounds polyphyllin VI and 17 β -estradiol have been shown to activate NLRP3 in cancer cells, inducing pyroptosis and improving the anti-tumoral immune response in NSCLC and hepatocellular carcinoma [192,197,198]. Furthermore, recombinant IL-18 has been used in clinical trials as an immunotherapy to enhance the activation of NK cells and T cells, which are critical for the recognition and elimination of tumor cells [191]. The development of specific inflammasome activators, however, remains an area of active research.

A promising strategy consists of using a combination of inflammasome activators with immune checkpoint inhibitors. For example, the addition of inflammasome activation may improve the efficacy of treatments like anti-PD-1 therapies by increasing the anti-tumor immune response. This combination approach attempts to boost the immune system's ability to recognize and destroy cancer cells simultaneously while also triggering pyroptosis in tumor cells [199].

13. Dual Actions of Inflammasomes in TME

Inflammasomes can be activated in the diverse subgroups of cells in TME, including tumor cells, TAMs, tumor-associated fibroblasts, and marrow-derived suppressive cells [20–23]. Recent findings have revealed the dual role of inflammasomes in TME, promoting or inhibiting tumor progression depending on different inflammasomes and tumors. Inflammasomes are mainly involved in tumorigenesis, metastasis, and immune evasion of malignant tumors [24].

Some inflammasome mutations have shown a tumorigenesis effect. Gain-of-function mutations in NLRP1 are associated with multiple self-healing palmoplantar carcinoma [200]. Individuals with NLRP1 variant rs12150220 or NLRP3 variant rs35829419 are more susceptible to nodular melanoma [201]. The NLRP3 variants rs10754558 and rs4612666 are linked with gastric cancer, and the amino acid mutation Q705K is related to pancreatic cancer [202,203]. It remains to be determined whether the pro-tumorigenic effects of these inflammasome mutations are driven by chronic inflammation. Transgenic mice carrying the relevant mutations could provide valuable insights into the underlying mechanisms. Suppression of inflammasomes has proved to attenuate tumorigenesis. For instance, in glycoprotein 130 (gp130)^{F/F} mice with spontaneous intestinal-type gastric cancer, the absence of ASC prevents tumor formation. This ASC knockout leads to decreased levels of mature IL-18 in the gastric tumor epithelium, which in turn enhances caspase-8-mediated apoptosis [204]. Likewise, knocking out ASC, inhibiting caspase-1, or deleting germ cells all reduce the occurrence of spontaneous cecal carcinogenesis in AhR-knockout (AhR^{-/-}) mice. These studies indicate that inflammation triggered by bacteria and inflammasome activation play harmful roles in tumor development [194]. Supporting this, mice with overexpressed IL-1 β in the stomach develop spontaneous gastric inflammation and cancer [205].

In contrast, other research has proposed that inflammasomes may play a protective role in tumor development. For example, the reduction in NLRP3 inflammasome components has been observed during the progression of multistage hepatocarcinogenesis [206]. ASC and caspase-1 recruited immune cells during tumorigenesis of chemically induced squamous cell carcinoma [195]. Mice without ASC, caspase-1, or NLRP3 had more severe colitis and tumorigenesis in colitis-associated cancer models [207,208]. The attenuated hematopoietic cell-derived IL-1 β and IL-18 at the tumor site of Nlrp3-knockout (Nlrp3^{-/-}) mice are found to be the key to inflammation and tumorigenesis. These studies align with results from pyrin knockout mice, which also exhibited more serious colitis and an increased tumor burden [209]. Mice lacking caspase-1 or NLRC4 exhibited greater proliferation of colonic epithelial cells and decreased apoptosis of tumor cells, which led to an increase in tumor formation in colitis-associated colorectal cancer models [193].

The role of inflammasomes in tumorigenesis may vary as malignant tumors progress. While NLRP3 inflammasome components are upregulated in tissues affected by hepatitis and cirrhosis, their expression is reduced in hepatocellular carcinoma [206]. ASC knockdown has opposite effects on tumor development in melanoma, suppressing tumorigenesis in metastatic melanoma and promoting it in primary melanoma. This contrasting behavior can be attributed to divergent NF- κ B activity downstream of ASC, where it is inhibited in primary melanoma but enhanced in metastatic melanoma [210]. The effect of inflammasomes in tumorigenesis not only depends on the clinical stage but also the cell class in TME. Deleting ASC conditionally in myeloid cells reduces the incidence of chemical-induced skin cancer, whereas eliminating ASC specifically in keratinocytes leads to an increase in tumor formation [211].

Tumor growth can be influenced by GSDMD, alongside IL-1 family members. Elevated GSDMD levels are associated with advanced TNM stages in NSCLC patients, and its knockdown inhibits tumor growth by activating the mitochondrial apoptotic pathway and

suppressing the EGFR/AKT signaling pathway [212]. In contrast, GSDMD expression is lower in gastric cancer cells and tissues, where reduced levels of GSDMD promote tumor cell proliferation by accelerating the S/G2 phase transition of the cell cycle. Moreover, GSDMD expression negatively correlates with the activation of STAT3, ERK, and PI3K/AKT pathways [213]. The differences in how GSDMD affects tumor pathways in various cancers may clarify these conflicting results.

Myeloid cells have been the main focus of research to study the connection between inflammasome activation and metastasis since IL-1 β is mostly produced by myeloid cells [214–216]. This connection has also been found in cancer-associated fibroblasts and tumor cells [22,217]. Even though IL-1 β is considered a marker of M-1-like macrophages that activates a tumor-targeted immune response, anomalous inflammasome activation in TAMs causes metastasis in different tumors. A direct relationship between the activation of inflammasomes, in particular NLRP3, and the late clinical stages of metastasis has been proved through clinical data. These data indicate a poor survival rate in patients with breast cancer and lung cancer [21,218]. The use of anakinra or canakinumab blocked the IL-1 signal and inhibited the metastasis of breast cancer [219].

Inflammasome activation can also inhibit metastasis [220]. Interestingly, NLRP3-produced IL-1 β induces the migration of colorectal cancer cells, and its activation in liver macrophages (Kupfer cells) decreases the metastasis of colorectal cancer cells. The secretion of IL-18 by NLRP3 in Kupffer cells plays a key role in promoting NK cell maturation and enhancing their tumor-killing activity [221,222]. These results indicate that the NLRP3 inflammasome signaling may either promote or inhibit tumor metastasis, depending on the tumor type and tissue involved. Variations in IL-1 β and IL-18 production across different cell subsets could potentially explain the differences observed in downstream events [191].

Immune evasion is quintessential for tumorigenesis and metastasis. Tumor cells can disrupt the Fas receptor, increase the expression of PD-L1, and decrease the expression of MHC-I. In the TME, immunosuppressive factors such as M2-like macrophages, MDSCs, and Tregs are key players that promote immune evasion by tumor cells [223,224]. Various stimulators can trigger the expression and activation of inflammasome components in cancer cells, fibroblasts, and macrophages [22,23,225–228]. This activation leads to the release of IL-1 β and IL-18, which, in turn, influences the expression of PD-L1 on tumor cells and contributes to the recruitment of immune-suppressive cells within the TME [191].

The activation of NLRP3 is associated with immunosuppressive cells such as Tregs, MDSCs, and TAMs. This process can be inhibited with NLRP3 inhibitors. The production of IL-18 in the TME of myeloma multiple has been linked to an increase in MDSCs, decreased T cell activity, and worse survival [229]. In other scenarios, IL-1 β and IL-18 are known to enhance T cell-mediated anti-cancer immunity [230,231]. Specifically, IL-18 derived from CD4⁺ T cells or introduced exogenously has been shown to promote the proliferation and anti-tumor activity of CD8⁺ T cells and CAR-T cells. These conflicting observations may be clarified by measuring the actual concentrations of IL-1 β and IL-18 in the TME rather than simply noting their relative changes. The hypothesis is that IL-1 β and IL-18 may induce different immune responses at varying concentrations. Moreover, understanding the specific locations of inflammasome, IL-1 β , and IL-18 expression could be crucial. Investigating inflammasome activation across different cell subsets might provide deeper insight into the inflammatory network and immune regulation within the TME, paving the way for targeted therapeutic strategies [191].

Both tumor-derived and fibroblast-derived inflammasome activation allows for an immune-suppressive environment in most cases, while myeloid cell-derived inflammasome activation seems to be favorable for anti-tumor immunity [227,230,232]. One potential explanation could lie in the varying size and duration of inflammasome activation between

myeloid cells and other cell types within the TME [191]. Studies have shown differences in inflammasome activation between macrophages and neutrophils, suggesting that inflammasomes can be activated to different extents, leading to distinct downstream effects. Differences in the signals exchanged between myeloid cells and other cell types could also play a role. For example, in DCs and macrophages, the release of IL-1 β and IL-18 facilitates antigen presentation and T-cell recruitment. Conversely, in tumor cells, inflammasome activation is frequently associated with the secretion of immunosuppressive signals like PD-1/PD-L1 [199,230,233]. Further research is needed to explore how inflammasome activation and its effects differ among cell subsets and to better understand the interactions between IL-1 family signals and other signaling pathways.

14. Connection Between Exosomes and Inflammasome Roles in TME

The interplay between exosomes and inflammasomes in the TME is a subject of growing interest due to its implications in cancer progression, inflammation, and immune response regulation. Exosomes have been identified as key mediators in the communication between tumor cells, immune cells, and other components of the TME. This communication can enhance or suppress inflammasome activation depending on the nature of the exosome cargo and the cellular context. The role of exosomes in inflammasome activation is complex, with evidence supporting both pro-inflammatory and anti-inflammatory effects depending on the origin of the exosomes and the specific context of the TME. Exosomes derived from various cell types, including immune, epithelial, cartilage, and cancer cells, have been shown to modulate inflammasome activity, with immune cell-derived exosomes being the most extensively studied [25].

14.1. Exosome-Mediated Inflammasome Activation in Immune Cells

Exosomes derived from immune cells, particularly macrophages, have been shown to significantly influence inflammasome activation within the TME [234]. These exosomes carry a variety of bioactive molecules, such as cytokines, RNA, and signaling proteins, that can activate inflammasomes in recipient cells. Upon uptake by recipient cells, these immune cell-derived exosomes trigger NF- κ B activation, leading to the upregulation of inflammasome components like NLRP3, pro-IL-1 β , and pro-IL-18. The dissociation of NF- κ B from I κ B in these cells promotes its nuclear translocation, where it initiates the transcription of these pro-inflammatory molecules, thus enhancing the inflammatory response in the TME [235]. Furthermore, exosomes from immune cells promote inflammasome complex formation through the interaction of PRRs, ASC, and pro-caspase-1, leading to the activation of caspase-1 and the subsequent secretion of pro-inflammatory cytokines such as IL-1 β , IL-18, and TNF- α .

In the context of cancer, immune cell-derived exosomes are particularly potent in inducing inflammasome activation. Studies have demonstrated that exosomes from macrophages in glioblastoma multiforme (GBM) patients contain miRNA-21, which inhibits the expression of the tumor suppressor gene PDCD4 in recipient GBM cells. This inhibition contributes to tumor cell proliferation and promotes chemoresistance, especially to treatments such as temozolomide [236]. Moreover, exosomes from immune cells also facilitate tumor immune evasion by modulating inflammasome activation, which can shift the immune response toward a more suppressive state. The capacity of immune cell-derived exosomes to modulate both inflammation and immune tolerance is a critical factor in shaping the TME and influencing cancer progression [237–239].

14.2. Cancer Cell-Derived Exosomes and Inflammasome Modulation

Exosomes derived from cancer cells have also been implicated in inflammasome activation. For instance, exosomes released from the HepG2 cell line, a hepatocellular carcinoma model, in response to palmitate fatty acid treatment, promote pro-IL-1 β expression and the release of mature IL-1 β from THP-1 monocytic cells [240]. Similar findings have been reported with exosomes derived from malignant ascites and amniotic fluid [241]. These findings suggest that exosomes from cancer cells not only enhance inflammasome activation but also propagate a pro-inflammatory environment, contributing to the inflammatory milieu that promotes tumor growth and metastasis.

Cancer-derived exosomes can also orchestrate the polarization of tumor-associated macrophage TAMs, further contributing to the inflammatory response in the TME [242–244]. M1-like TAMs, induced by pro-inflammatory signals, are associated with tumor growth and angiogenesis. M2-like TAMs are typically involved in immune suppression and tissue remodeling. Cancer-derived exosomes can shift TAMs toward the M1-like phenotype, enhancing the pro-inflammatory state of the TME and supporting tumor progression [245]. This polarization is linked to poor clinical prognosis [246–248]. A recent study showed that exosomes derived from glioma cells, enriched with HMGB3, promote M2 polarization in macrophages and activate the NLRP3 inflammasome, inducing pyroptosis. This mechanism plays a critical role in establishing an immunosuppressive and inflammatory TME in glioma [249]. The NLRP3 inflammasome is pivotal in TAM polarization since its activation is essential for IL-1 β production in macrophages and other immune cells. Cancer cells can influence key molecules in the NLRP3 pathway, resulting in TAM reprogramming toward a pro-inflammatory state that supports tumor progression. For instance, exosomes derived from murine lung cancer cells express the tripartite motif-containing protein TRIM59, which promotes the ubiquitination and degradation of ABHD5. This, in turn, activates the NLRP3 inflammasome pathway, leading to IL-1 β release from macrophages and supporting the growth and metastasis of lung cancer [250]. ABHD5 plays a crucial role in lipid metabolism, and its absence has been found to enhance macrophage reprogramming and inflammasome activation, further highlighting the connection between metabolic alterations in the TME and inflammasome activation [251]. Additionally, less studied inflammasomes like NLRP6 have been identified as critical for the M2 polarization of macrophages induced by small cell lung cancer (SCLC)-derived exosomes. This process facilitates SCLC metastasis both in vitro and in vivo [252]. These findings provide new insights into the role of novel inflammasome proteins in cancer progression and suggest their potential as therapeutic targets for managing cancer metastasis.

Beyond their role in TAM polarization, cancer-derived exosomes can also influence inflammasome activation in response to external factors such as viral infections. Exosomes secreted by virus-infected cells can carry viral proteins that impact inflammasome activation in neighboring cells. For example, Epstein–Barr virus (EBV)-infected B-cells and nasopharyngeal carcinoma (NPC) cells secrete exosomes containing latent membrane protein 1 (LMP1), a viral protein that enhances B-cell proliferation, tumor growth, and radioresistance. Upon uptake by non-infected cells, LMP1-containing exosomes contribute to the inflammatory microenvironment and promote tumor progression by facilitating immune evasion mechanisms that are characteristic of cancer [253,254]. These findings highlight the role of viral infections in modulating exosome–inflammasome interactions and influencing cancer-related inflammation.

Additionally, cancer therapies can influence exosome–inflammasome interactions. For instance, exosomes released from HepG2 cells treated with ezetimibe do not induce inflammasome activation or IL-1 β secretion, suggesting that exosomal cargo composition is sensitive to the metabolic state of the tumor cells [240]. On the other hand, doxorubicin,

a commonly used chemotherapeutic agent, has been shown to induce an inflammatory response in cardiomyocytes by activating TLR4, NLRP3, caspase-1, and IL-1 β . Exosomes derived from embryonic stem cells (ESCs) were found to mitigate the inflammasome-activating effects of doxorubicin, while exosomes from mouse embryonic fibroblasts (MEFs) had no such effect. This observation suggests that the type of exosomes and their cargo can influence inflammasome activation, indicating a potential therapeutic avenue for modulating the inflammatory responses induced by chemotherapy [255]. The differential effects of exosomes from various cell types underscore the cell-specific nature of exosome cargo and its impact on inflammasome signaling.

14.3. Exosome Cargo and Its Role in Inflammasome Activation

The cargo contained within exosomes is a critical determinant of their impact on inflammasome activation and their role in cancer biology. Exosomes derived from different cell types carry distinct biomolecules, including proteins, miRNAs, and lipids, which can influence the inflammatory response in recipient cells [256–258]. For example, exosomes from colorectal cancer cells, particularly those harboring KRAS mutations, exhibit distinct miRNA profiles linked to the KRAS gene's mutation status. These mutations in KRAS have been shown to influence the sorting of miRNAs into exosomes, which then modulate gene expression in recipient cells, contributing to inflammation and tumor progression [259].

Specific proteins involved in exosome biogenesis and cargo sorting, such as CD63 and RILP, play a significant role in determining the molecular content of exosomes [256,259,260]. In the case of EBV-infected cells, CD63 has been shown to mediate the loading of LMP1 into exosomes, which is essential for the viral protein's pro-inflammatory and tumorigenic effects [261]. Similarly, RILP regulates the packaging of miR-155 into exosomes derived from hepatitis C virus (HCV)-infected hepatoma cells, suggesting that cellular factors can selectively control the contents of exosomes, influencing their ability to modulate inflammasome activation and inflammatory pathways [262].

The selective packing of miRNAs and proteins into exosomes is not fully understood, but several studies have provided insights into the mechanisms behind this process. Exosome secretion and cargo sorting are influenced by specific cellular conditions such as hypoxia, extracellular acidification, and changes in intracellular calcium levels [263,264]. These conditions are commonly found in the TME and can stimulate exosome release, thereby facilitating the transfer of inflammatory signals between cells. Exosome release can also be triggered by the activation of immune pathways, such as those mediated by LPS-stimulated TLR4, which further underscores the link between immune responses and exosome-mediated inflammasome activation in the TME [265].

14.4. Exosomal Secretion and TME

The TME, characterized by its inflammatory and immune landscape, is crucial in modulating exosome secretion. Several factors inherent to the TME, such as hypoxia, acidity, and elevated calcium levels, have been shown to enhance exosome release. In particular, the acidic environment of the TME, which results from increased metabolic activity in tumor cells, has been demonstrated to promote exosome secretion from metastatic melanoma cells [266]. This observation suggests that the TME is not a passive environment but actively participates in shaping the nature of exosome-mediated communication, particularly concerning inflammasome activation and the inflammatory response.

The role of exosome secretion in cancer inflammation is further emphasized by the activation of inflammasomes in response to tumor-associated signals. For example, elevated extracellular ATP levels in the TME can activate the P2X7 receptor, leading to an increase in cytoplasmic calcium levels that subsequently promote exosome release. This pathway

provides an additional link between inflammation, immune activation, and exosome secretion, highlighting the complex interplay between these processes in the TME [267]. Moreover, inflammasome activation, whether through exosomal delivery of inflammasome components or as a consequence of TLR4 signaling, may further enhance exosome secretion, creating a feedback loop that perpetuates inflammation within the tumor.

15. Conclusions, Challenges, and Future Directions

Exosomes have emerged as promising natural carriers for cancer therapies. Their impact on cancer growth and progression varies depending on their cellular origin, shaping the trajectory of the disease by influencing key processes within the TME. Exosomes can be used to enhance TIL function by reversing T-cell exhaustion or delivering immune-stimulating molecules. Furthermore, exosome-based strategies may help improve TIL persistence and infiltration into tumors, making TIL therapy more effective. While TILs and exosome therapy offer exciting opportunities to modulate the TME for improved cancer immunotherapy, several challenges remain. Addressing these hurdles will be critical for translating these strategies into effective clinical applications.

Exosomes reflect their parent cells and the conditions under which they were formed, but identifying their origins *in vivo* is challenging. They carry diverse molecular cargo, including proteins, RNAs, and lipids, but pinpointing the specific effects of miRNA clusters and individual miRNAs is complex. The mechanisms behind exosome targeting, uptake, gene expression alterations, and physiological effects remain unclear. Moreover, determining their specific effects and dominant mechanisms is difficult due to the presence of multiple molecular components that may contribute to overlapping or distinct functions [268].

Furthermore, exosomes are quickly cleared by macrophages, reducing their circulation time and therapeutic efficacy, while extending their half-life may cause unforeseen side effects. Their poor zeta potential leads to aggregation, impacting delivery, triggering immune responses, and shortening circulation time. Determining the optimal dosage is challenging due to factors like delivery methods, short half-life, and variations in parent cell origins. Additionally, it remains uncertain whether exosomes or specific types, engineered versions, or alternatives like exosome-mimicking liposomes are the best therapeutic option [268]. Further research is essential to better understand how tumor-derived exosomes contribute to immune evasion, angiogenesis, metastasis, and therapy resistance. Specifically, exploring the role of exosomes in immune modulation, particularly in the regulation of immune checkpoints, and engineering exosomes for targeted drug delivery, especially for chemotherapy and gene therapy, could pave the way for novel immunotherapeutic strategies.

Inflammasomes play a dual role in cancer progression, both promoting and suppressing tumor growth by regulating inflammation and immune responses. While inflammasome-targeted therapies hold significant promise in cancer treatment, several challenges persist. The specificity of inflammasome inhibitors and activators is critical to prevent unintended disruptions in immune pathways. For instance, indiscriminate IL-1 β blockade may impair host defenses against infections, whereas widespread inflammasome activation could trigger excessive inflammation and tissue damage. Additionally, the heterogeneity of the TME complicates the development of universal inflammasome-targeting strategies, as their roles vary across cell types. In TAMs and DCs, inflammasomes may enhance anti-tumor immunity, whereas in tumor cells, they can promote malignancy. A deeper understanding of inflammasome functions in specific TME cell subsets will be essential for designing precise and effective therapies.

Furthermore, while combining inflammasome-targeting agents with immune checkpoint inhibitors shows promise, further research is needed to determine the most effective

combinations and assess their long-term impact on immune function and tumor progression. Ultimately, inflammasome-targeted therapies could become a powerful tool in cancer immunotherapy, but their success will hinge on enhancing selectivity, deciphering the intricate immune dynamics within tumors, and developing personalized treatment strategies.

Recent advancements in research techniques, particularly in bioinformatics, have provided powerful tools for elucidating inflammasome expression, function, and their associations with clinical outcomes. For instance, a pan-cancer study demonstrated that NLRP3 expression varies across tumor types, with its activity either elevated or suppressed depending on the cancer type [269]. Notably, this analysis highlighted a strong correlation between NLRP3 expression and patient survival, especially in melanoma and hepatocellular carcinoma, where higher NLRP3 levels were linked to improved survival rates, a more favorable prognosis, and enhanced responses to immunotherapy. Additionally, another study developed a risk-scoring system based on inflammasome-related genes to predict clinicopathologic features, prognosis, and immune response patterns in kidney renal clear cell carcinoma [270]. These findings underscore the need for further research to clarify the role of inflammasomes in cancer progression and their potential clinical applications.

Exosomes play a multifaceted role in regulating inflammasome activation within the TME. By carrying and transferring bioactive molecules, they serve as key mediators of inflammation and immune responses in cancer. Exosomes derived from immune cells, epithelial cells, and cancer cells have been shown to enhance inflammasome activation, triggering the release of pro-inflammatory cytokines such as IL-1 β and IL-18. Moreover, the selective packaging of biomolecules into exosomes is highly regulated and varies based on the cell of origin, further shaping their impact on recipient cells. While growing evidence underscores the link between exosomes and inflammasome activation, several key questions remain about the underlying mechanisms of these interactions. Although inflammasome activation is known to promote exosome release, its precise impact on exosome secretion is not yet fully understood. Discrepancies in research findings may stem from differences in the types of PRRs involved, the specific activators used, and the targeted effector cells. To clarify these processes, further studies are needed to examine how these factors influence exosome–inflammasome dynamics and to explore the therapeutic potential of targeting these pathways in cancer treatment.

Author Contributions: Conceptualization and supervision, A.M.E.; literature search and screening of articles, A.M.E., H.E.-G., A.S.A., M.F.-O. and R.K.A.S.; writing—original draft preparation, H.E.-G., A.S.A. and M.F.-O.; writing—review and editing, A.M.E. and R.K.A.S. All authors have read and agreed to the published version of the manuscript.

Funding: No external funding.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Chhikara, B.S.; Parang, K. Global Cancer Statistics 2022: The trends projection analysis. *Chem. Biol. Lett.* **2023**, *10*, 451.
2. Siegel, R.L.; Miller, K.D.; Wagle, N.S.; Jemal, A. Cancer statistics, 2023. *CA Cancer J. Clin.* **2023**, *73*, 17–48. [[CrossRef](#)]
3. Buchbinder, E.; Hodi, F.S. Cytotoxic T lymphocyte antigen-4 and immune checkpoint blockade. *J. Clin. Investig.* **2015**, *125*, 3377–3383. [[CrossRef](#)]
4. Heppner, B.I.; Loibl, S.; Denkert, C. Tumor-Infiltrating Lymphocytes: A Promising Biomarker in Breast Cancer. *Breast Care* **2016**, *11*, 96–100. [[CrossRef](#)] [[PubMed](#)]

5. Topalian, S.L.; Hodi, F.S.; Brahmer, J.R.; Gettinger, S.N.; Smith, D.C.; McDermott, D.F.; Powderly, J.D.; Carvajal, R.D.; Sosman, J.A.; Atkins, M.B.; et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N. Engl. J. Med.* **2012**, *366*, 2443–2454. [\[CrossRef\]](#)
6. El Bairi, K.; Haynes, H.R.; Blackley, E.; Fineberg, S.; Shear, J.; Turner, S.; de Freitas, J.R.; Sur, D.; Amendola, L.C.; Gharib, M.; et al. The tale of TILs in breast cancer: A report from The International Immuno-Oncology Biomarker Working Group. *NPJ Breast Cancer* **2021**, *7*, 150. [\[CrossRef\]](#)
7. Sezer, A.; Kilickap, S.; Gümüş, M.; Bondarenko, I.; Özgüroğlu, M.; Gogishvili, M.; Turk, H.M.; Cicin, I.; Bentsion, D.; Gladkov, O.; et al. Cemiplimab monotherapy for first-line treatment of advanced non-small-cell lung cancer with PD-L1 of at least 50%: A multicentre, open-label, global, phase 3, randomised, controlled trial. *Lancet* **2021**, *397*, 592–604. [\[CrossRef\]](#)
8. Fong, L.; Small, E.J. Anti-Cytotoxic T-Lymphocyte Antigen-4 Antibody: The First in an Emerging Class of Immunomodulatory Antibodies for Cancer Treatment. *J. Clin. Oncol.* **2008**, *26*, 5275–5283. [\[CrossRef\]](#)
9. Mocellin, S.; Nitti, D. CTLA-4 blockade and the renaissance of cancer immunotherapy. *Biochim. Biophys. Acta (BBA) Rev. Cancer* **2013**, *1836*, 187–196. [\[CrossRef\]](#)
10. Bronte, V.; Mocellin, S. Suppressive Influences in the Immune Response to Cancer. *J. Immunother.* **2009**, *32*, 1–11. [\[CrossRef\]](#)
11. Poschke, I.; Mougiakakos, D.; Kiessling, R. Camouflage and sabotage: Tumor escape from the immune system. *Cancer Immunol. Immunother.* **2011**, *60*, 1161–1171. [\[CrossRef\]](#)
12. Anderson, N.M.; Simon, M.C. The tumor microenvironment. *Curr. Biol.* **2020**, *30*, R921–R925. [\[CrossRef\]](#)
13. Bai, R.; Cui, J. Development of Immunotherapy Strategies Targeting Tumor Microenvironment Is Fiercely Ongoing. *Front. Immunol.* **2022**, *13*, 890166. [\[CrossRef\]](#)
14. Kim, S.K.; Cho, S.W. The Evasion Mechanisms of Cancer Immunity and Drug Intervention in the Tumor Microenvironment. *Front. Pharmacol.* **2022**, *13*, 868695. [\[CrossRef\]](#)
15. Kim, M.S.; Haney, M.J.; Zhao, Y.; Mahajan, V.; Deygen, I.; Klyachko, N.L.; Inskoe, E.; Piroyan, A.; Sokolsky, M.; Okolie, O.; et al. Development of exosome-encapsulated paclitaxel to overcome mdr in cancer cells. *Nanomed. Nanotechnol. Biol. Med.* **2016**, *12*, 655–664. [\[CrossRef\]](#)
16. Chen, L.; Wang, L.; Zhu, L.; Xu, Z.; Liu, Y.; Li, Z.; Zhou, J.; Luo, F. Exosomes as Drug Carriers in Anti-Cancer Therapy. *Front. Cell Dev. Biol.* **2022**, *10*, 728616. [\[CrossRef\]](#)
17. Fabbi, M.; Carbotti, G.; Ferrini, S. Context-dependent role of IL-18 in cancer biology and counter-regulation by IL-18BP. *J. Leukoc. Biol.* **2014**, *97*, 665–675. [\[CrossRef\]](#)
18. Tas, F.; Yasasever, C.T.; Karabulut, S.; Tastekin, D.; Duranyildiz, D. Clinical significance of serum interleukin-18 (IL-18) levels in patients with gastric cancer. *Biomed. Pharmacother.* **2015**, *70*, 19–23. [\[CrossRef\]](#)
19. Hu, Z.; Chai, J. Structural Mechanisms in NLR Inflammasome Assembly and Signaling. In *Inflammasome Signaling and Bacterial Infections*; Backert, S., Ed.; Springer: Cham, Switzerland, 2016; Volume 397, pp. 23–42. [\[CrossRef\]](#)
20. Bruchard, M.; Mignot, G.; Derangère, V.; Chalmin, F.; Chevriaux, A.; Végran, F.; Boireau, W.; Simon, B.; Ryffel, B.; Connat, J.L.; et al. Chemotherapy-triggered cathepsin B release in myeloid-derived suppressor cells activates the Nlrp3 inflammasome and promotes tumor growth. *Nat. Med.* **2012**, *19*, 57–64. [\[CrossRef\]](#)
21. Weichand, B.; Popp, R.; Dziubla, S.; Mora, J.; Strack, E.; Elwakeel, E.; Frank, A.C.; Scholich, K.; Pierre, S.; Syed, S.N.; et al. S1PR1 on tumor-associated macrophages promotes lymphangiogenesis and metastasis via NLRP3/IL-1v. *J. Exp. Med.* **2017**, *214*, 2695–2713. [\[CrossRef\]](#)
22. Ershaid, N.; Sharon, Y.; Doron, H.; Raz, Y.; Shani, O.; Cohen, N.; Monteran, L.; Leider-Trejo, L.; Ben-Shmuel, A.; Yassin, M.; et al. NLRP3 inflammasome in fibroblasts links tissue damage with inflammation in breast cancer progression and metastasis. *Nat. Commun.* **2019**, *10*, 4375. [\[CrossRef\]](#)
23. Das, S.; Shapiro, B.; Vucic, E.A.; Vogt, S.; Bar-Sagi, D. Tumor Cell-Derived IL1 β Promotes Desmoplasia and Immune Suppression in Pancreatic Cancer. *Cancer Res.* **2020**, *80*, 1088–1101. [\[CrossRef\]](#)
24. Cao, X.; Xu, J. Insights into inflammasome and its research advances in cancer. *Tumori J.* **2019**, *105*, 456–464. [\[CrossRef\]](#)
25. Noonin, C.; Thongboonkerd, V. Exosome-inflammasome crosstalk and their roles in inflammatory responses. *Theranostics* **2021**, *11*, 4436–4451. [\[CrossRef\]](#)
26. Gu, Q.; Zou, J.; Zhou, Y.; Deng, Q. Mechanism of inflammasomes in cancer and targeted therapies. *Front. Oncol.* **2023**, *13*, 1133013. [\[CrossRef\]](#)
27. Kawai, O.; Ishii, G.; Kubota, K.; Murata, Y.; Naito, Y.; Mizuno, T.; Aokage, K.; Saijo, N.; Nishiwaki, Y.; Gemma, A.; et al. Predominant infiltration of macrophages and CD8⁺ T Cells in cancer nests is a significant predictor of survival in stage IV nonsmall cell lung cancer. *Cancer* **2008**, *113*, 1387–1395. [\[CrossRef\]](#)
28. Xue, J.; Schmidt, S.V.; Sander, J.; Draffehn, A.; Krebs, W.; Quester, I.; De Nardo, D.; Gohel, T.D.; Emde, M.; Schmidleithner, L.; et al. Transcriptome-based network analysis reveals a spectrum model of human macrophage activation. *Immunity* **2014**, *40*, 274–288. [\[CrossRef\]](#)

29. Aras, S.; Zaidi, M.R. TAMEless traitors: Macrophages in cancer progression and metastasis. *Br. J. Cancer* **2017**, *117*, 1583–1591. [\[CrossRef\]](#)
30. Dumont, N.; Liu, B.; DeFilippis, R.A.; Chang, H.; Rabban, J.T.; Karnezis, A.N.; Tjoe, J.A.; Marx, J.; Parvin, B.; Tlsty, T.D. Breast Fibroblasts Modulate Early Dissemination, Tumorigenesis, and Metastasis through Alteration of Extracellular Matrix Characteristics. *Neoplasia* **2013**, *15*, 249–262. [\[CrossRef\]](#)
31. Gascard, P.; Tlsty, T.D. Carcinoma-associated fibroblasts: Orchestrating the composition of malignancy. *Genes Dev.* **2016**, *30*, 1002–1019. [\[CrossRef\]](#)
32. A Konerding, M.; Malkusch, W.; Klapthor, B.; van Ackern, C.; Fait, E.; A Hill, S.; Parkins, C.; Chaplin, D.J.; Presta, M.; Denekamp, J. Evidence for characteristic vascular patterns in solid tumours: Quantitative studies using corrosion casts. *Br. J. Cancer* **1999**, *80*, 724–732. [\[CrossRef\]](#)
33. Goswami, T.K.; Singh, M.; Dhawan, M.; Mitra, S.; Bin Emran, T.; Rabaan, A.A.; Al Mutair, A.; Al Alawi, Z.; Alhumaid, S.; Dhama, K. Regulatory T cells (Tregs) and their therapeutic potential against autoimmune disorders—Advances and challenges. *Hum. Vaccines Immunother.* **2022**, *18*, 2035117. [\[CrossRef\]](#)
34. Peña-Romero, A.C.; Orenes-Piñero, E. Dual Effect of Immune Cells within Tumour Microenvironment: Pro- and Anti-Tumour Effects and Their Triggers. *Cancers* **2022**, *14*, 1681. [\[CrossRef\]](#)
35. Fridlender, Z.G.; Sun, J.; Kim, S.; Kapoor, V.; Cheng, G.; Ling, L.; Worthen, G.S.; Albelda, S.M. Polarization of tumor-associated neutrophil phenotype by TGF- β : “N1” versus “N2” TAN. *Cancer Cell* **2009**, *16*, 183–194. [\[CrossRef\]](#)
36. Tepper, R.I.; Pattengale, P.K.; Leder, P. Murine interleukin-4 displays potent anti-tumor activity in vivo. *Cell* **1989**, *57*, 503–512. [\[CrossRef\]](#)
37. Shamri, R.; Xenakis, J.J.; Spencer, L.A. Eosinophils in innate immunity: An evolving story. *Cell Tissue Res.* **2010**, *343*, 57–83. [\[CrossRef\]](#)
38. Gentles, A.J.; Newman, A.M.; Liu, C.L.; Bratman, S.V.; Feng, W.; Kim, D.; Nair, V.S.; Xu, Y.; Khuong, A.; Hoang, C.D.; et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat. Med.* **2015**, *21*, 938–945. [\[CrossRef\]](#)
39. Patil, R.S.; Shah, S.U.; Shrikhande, S.V.; Goel, M.; Dikshit, R.P.; Chiplunkar, S.V. IL17 producing $\gamma\delta$ T cells induce angiogenesis and are associated with poor survival in gallbladder cancer patients. *Int. J. Cancer* **2016**, *139*, 869–881. [\[CrossRef\]](#)
40. Mellman, I. Dendritic Cells: Master Regulators of the Immune Response. *Cancer Immunol. Res.* **2013**, *1*, 145–149. [\[CrossRef\]](#)
41. Krzywinska, E.; Allende-Vega, N.; Cornillon, A.; Vo, D.-N.; Cayrefourcq, L.; Panabieres, C.; Vilches, C.; Déchanet-Merville, J.; Hicheri, Y.; Rossi, J.-F.; et al. Identification of Anti-tumor Cells Carrying Natural Killer (NK) Cell Antigens in Patients With Hematological Cancers. *EBioMedicine* **2015**, *2*, 1364–1376. [\[CrossRef\]](#)
42. Lijun, Z.; Xin, Z.; Danhua, S.; Xiaoping, L.; Jianliu, W.; Huilan, W.; Lihui, W. Tumor-Infiltrating Dendritic Cells May Be Used as Clinicopathologic Prognostic Factors in Endometrial Carcinoma. *Int. J. Gynecol. Cancer* **2012**, *22*, 836–841. [\[CrossRef\]](#)
43. Ostroumov, D.; Fekete-Drimusz, N.; Saborowski, M.; Kühnel, F.; Woller, N. CD4 and CD8 T lymphocyte interplay in controlling tumor growth. *Cell. Mol. Life Sci.* **2017**, *75*, 689–713. [\[CrossRef\]](#)
44. Ziai, J.; Gilbert, H.N.; Foreman, O.; Eastham-Anderson, J.; Chu, F.; Huseni, M.; Kim, J.M. CD8+ T cell infiltration in breast and colon cancer: A histologic and statistical analysis. *PLoS ONE* **2018**, *13*, e0190158. [\[CrossRef\]](#)
45. Carretero, R.; Sektioglu, I.M.; Garbi, N.; Salgado, O.C.; Beckhove, P.; Hämmerling, G.J. Eosinophils orchestrate cancer rejection by normalizing tumor vessels and enhancing infiltration of CD8+ T cells. *Nat. Immunol.* **2015**, *16*, 609–617. [\[CrossRef\]](#)
46. Akbulut, G.D.; Özkazanc, D.; Esendağlı, G. Th1 cells in cancer-associated inflammation. *Turk. J. Biol.* **2017**, *41*, 20–30. [\[CrossRef\]](#)
47. De Mello, R.A.; Veloso, A.F.; Catarina, P.E.; Nadine, S.; Antoniou, G. Potential role of immunotherapy in advanced non-small-cell lung cancer. *OncoTargets Ther.* **2017**, *10*, 21. [\[CrossRef\]](#)
48. Zitvogel, L.; Galluzzi, L.; Kepp, O.; Smyth, M.J.; Kroemer, G. Type I interferons in anticancer immunity. *Nat. Rev. Immunol.* **2015**, *15*, 405–414. [\[CrossRef\]](#)
49. Tan, A.H.-M.; Goh, S.Y.-P.; Wong, S.-C.; Lam, K.-P. T Helper Cell-specific Regulation of Inducible Costimulator Expression via Distinct Mechanisms Mediated by T-bet and GATA-3. *J. Biol. Chem.* **2008**, *283*, 128–136. [\[CrossRef\]](#)
50. Stanton, S.E.; Disis, M.L. Clinical significance of tumor-infiltrating lymphocytes in breast cancer. *J. Immunother. Cancer* **2016**, *4*, 59. [\[CrossRef\]](#)
51. Salgado, R.; Denkert, C.; Demaria, S.; Sirtaine, N.; Klauschen, F.; Pruneri, G.; Wienert, S.; Van den Eynden, G.; Baehner, F.L.; Penault-Llorca, F.; et al. The evaluation of tumor-infiltrating lymphocytes (TILs) in breast cancer: Recommendations by an International TILs Working Group 2014. *Ann. Oncol.* **2015**, *26*, 259–271. [\[CrossRef\]](#)
52. June, C.H. Adoptive T cell therapy for cancer in the clinic. *J. Clin. Investig.* **2007**, *117*, 1466–1476. [\[CrossRef\]](#) [\[PubMed\]](#)
53. Tsimberidou, A.-M.; Van Morris, K.; Vo, H.H.; Eck, S.; Lin, Y.-F.; Rivas, J.M.; Andersson, B.S. T-cell receptor-based therapy: An innovative therapeutic approach for solid tumors. *J. Hematol. Oncol.* **2021**, *14*, 102. [\[CrossRef\]](#)
54. Zhao, Z.; Chen, Y.; Francisco, N.M.; Zhang, Y.; Wu, M. The application of CAR-T cell therapy in hematological malignancies: Advantages and challenges. *Acta Pharm. Sin. B* **2018**, *8*, 539–551. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Sterner, R.C.; Sterner, R.M. CAR-T cell therapy: Current limitations and potential strategies. *Blood Cancer J.* **2021**, *11*, 69. [\[CrossRef\]](#)

56. Zhao, Y.; Deng, J.; Rao, S.; Guo, S.; Shen, J.; Du, F.; Wu, X.; Chen, Y.; Li, M.; Chen, M.; et al. Tumor Infiltrating Lymphocyte (TIL) Therapy for Solid Tumor Treatment: Progressions and Challenges. *Cancers* **2022**, *14*, 4160. [\[CrossRef\]](#)
57. Sengsayadeth, S.; Savani, B.N.; Oluwole, O.; Dholaria, B. Overview of approved CAR-T therapies, ongoing clinical trials, and its impact on clinical practice. *EJHaem*. **2022**, *3*, 6–10. [\[CrossRef\]](#)
58. Dagar, G.; Gupta, A.; Masoodi, T.; Nisar, S.; Merhi, M.; Hashem, S.; Chauhan, R.; Dagar, M.; Mirza, A.; Bagga, P.; et al. Harnessing the potential of CAR-T cell therapy: Progress, challenges, and future directions in hematological and solid tumor treatments. *J. Transl. Med.* **2023**, *21*, 449. [\[CrossRef\]](#)
59. Cohen, I.J.; Blasberg, R. Impact of the Tumor Microenvironment on Tumor-Infiltrating Lymphocytes: Focus on Breast Cancer. *Breast Cancer Basic Clin. Res.* **2017**, *11*, 1178223417731565. [\[CrossRef\]](#)
60. Antony, P.A.; Piccirillo, C.A.; Akpınarlı, A.; Finkelstein, S.E.; Speiss, P.J.; Surman, D.R.; Palmer, D.C.; Chan, C.-C.; Klebanoff, C.A.; Overwijk, W.W.; et al. CD8+ T Cell Immunity Against a Tumor/Self-Antigen Is Augmented by CD4+ T Helper Cells and Hindered by Naturally Occurring T Regulatory Cells. *J. Immunol.* **2005**, *174*, 2591–2601. [\[CrossRef\]](#)
61. Gattinoni, L.; Finkelstein, S.E.; Klebanoff, C.A.; Antony, P.A.; Palmer, D.C.; Spiess, P.J.; Hwang, L.N.; Yu, Z.; Wrzesinski, C.; Heimann, D.M.; et al. Removal of homeostatic cytokine sinks by lymphodepletion enhances the efficacy of adoptively transferred tumor-specific CD8+ T cells. *J. Exp. Med.* **2005**, *202*, 907–912. [\[CrossRef\]](#)
62. Bechman, N.; Maher, J. Lymphodepletion strategies to potentiate adoptive T-cell immunotherapy—what are we doing; where are we going? *Expert Opin. Biol. Ther.* **2021**, *21*, 627–637. [\[CrossRef\]](#)
63. Nissani, A.; Lev-Ari, S.; Meirson, T.; Jacoby, E.; Asher, N.; Ben-Betzalel, G.; Itzhaki, O.; Shapira-Frommer, R.; Schachter, J.; Markel, G.; et al. Comparison of non-myeloablative lymphodepleting preconditioning regimens in patients undergoing adoptive T cell therapy. *J. Immunother. Cancer* **2021**, *9*, e001743. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Demin, O.; Kolesova, G.; Ramos-Hernandez, N.; Faltg, T.; Zajic, S.; Cucurull-Sanchez, L. 338 Comparison of lymphodepleting chemotherapy regimens as preconditioning for T-cell therapies. In Proceedings of the SITC 38th Annual Meeting (SITC 2023) Abstracts, San Diego, CA, USA, 3–5 November 2023; p. A387.
65. Lickfett, B.; Chu, L.; Ortiz-Maldonado, V.; Warmuth, L.; Barba, P.; Doglio, M.; Henderson, D.; Hudecek, M.; Kremer, A.; Markman, J.; et al. Lymphodepletion—An essential but undervalued part of the chimeric antigen receptor T-cell therapy cycle. *Front. Immunol.* **2023**, *14*, 1303935. [\[CrossRef\]](#) [\[PubMed\]](#)
66. Gautama, B.; Duncan, N.; Fletcher, P.; Besley, C. The Impact of Lymphodepleting Chemotherapy Dose Modifications on Axicabtagene Ciloleucel Expansion and Treatment Outcome in Patients with Relapsed or Refractory CD19-Positive Diffuse Large B-Cell Lymphoma. *Blood* **2024**, *144*, 3729. [\[CrossRef\]](#)
67. Dudley, M.E.; Wunderlich, J.R.; Robbins, P.F.; Yang, J.C.; Hwu, P.; Schwartzentruber, D.J.; Topalian, S.L.; Sherry, R.; Restifo, N.P.; Hubicki, A.M.; et al. Cancer Regression and Autoimmunity in Patients After Clonal Repopulation with Antitumor Lymphocytes. *Science* **2002**, *298*, 850–854. [\[CrossRef\]](#)
68. Hughes, M.S.; Yu, Y.Y.; Dudley, M.E.; Zheng, Z.; Robbins, P.F.; Li, Y.; Wunderlich, J.; Hawley, R.G.; Moayeri, M.; Rosenberg, S.A.; et al. Transfer of a TCR Gene Derived from a Patient with a Marked Antitumor Response Conveys Highly Active T-Cell Effector Functions. *Hum. Gene Ther.* **2005**, *16*, 457–472. [\[CrossRef\]](#)
69. Aoki, Y.; Takakuwa, K.; Kodama, S.; Tanaka, K.; Takahashi, M.; Tokunaga, A.; Takahashi, T. Use of adoptive transfer of tumor-infiltrating lymphocytes alone or in combination with cisplatin-containing chemotherapy in patients with epithelial ovarian cancer. *Cancer Res.* **1991**, *51*, 1934–1939.
70. Stevanović, S.; Draper, L.M.; Langhan, M.M.; Campbell, T.E.; Kwong, M.L.; Wunderlich, J.R.; Dudley, M.E.; Yang, J.C.; Sherry, R.M.; Kammula, U.S.; et al. Complete Regression of Metastatic Cervical Cancer After Treatment with Human Papillomavirus–Targeted Tumor-Infiltrating T Cells. *J. Clin. Oncol.* **2015**, *33*, 1543–1550. [\[CrossRef\]](#)
71. Tran, E.; Robbins, P.F.; Lu, Y.-C.; Prickett, T.D.; Gartner, J.J.; Jia, L.; Pasetto, A.; Zheng, Z.; Ray, S.; Groh, E.M.; et al. T-Cell Transfer Therapy Targeting Mutant KRAS in Cancer. *N. Engl. J. Med.* **2016**, *375*, 2255–2262. [\[CrossRef\]](#)
72. Saberian, C.; Amaria, R.N.; Najjar, A.M.; Radvanyi, L.G.; Haymaker, C.L.; Forget, M.-A.; Bassett, R.L.; Faria, S.C.; Glitza, I.C.; Alvarez, E.; et al. Randomized phase II trial of lymphodepletion plus adoptive cell transfer of tumor-infiltrating lymphocytes, with or without dendritic cell vaccination, in patients with metastatic melanoma. *J. Immunother. Cancer* **2021**, *9*, e002449. [\[CrossRef\]](#)
73. Rohaan, M.W.; Borch, T.H.; Berg, J.H.v.D.; Met, Ö.; Kessels, R.; Foppen, M.H.G.; Granhøj, J.S.; Nuijen, B.; Nijenhuis, C.; Jedema, I.; et al. Tumor-Infiltrating Lymphocyte Therapy or Ipilimumab in Advanced Melanoma. *N. Engl. J. Med.* **2022**, *387*, 2113–2125. [\[CrossRef\]](#)
74. Hu, W.; Bian, Y.; Ji, H. TIL Therapy in Lung Cancer: Current Progress and Perspectives. *Adv. Sci.* **2024**, *11*, e2409356. [\[CrossRef\]](#)
75. Creelan, B.C.; Wang, C.; Teer, J.K.; Toloza, E.M.; Yao, J.; Kim, S.; Landin, A.M.; Mullinax, J.E.; Saller, J.J.; Saltos, A.N.; et al. Tumor-infiltrating lymphocyte treatment for anti-PD-1-resistant metastatic lung cancer: A phase 1 trial. *Nat. Med.* **2021**, *27*, 1410–1418. [\[CrossRef\]](#)
76. Yang, Y. Cancer immunotherapy: Harnessing the immune system to battle cancer. *J. Clin. Investig.* **2015**, *125*, 3335–3337. [\[CrossRef\]](#)
77. Sharpe, A.H.; Pauken, K.E. The diverse functions of the PD1 inhibitory pathway. *Nat. Rev. Immunol.* **2017**, *18*, 153–167. [\[CrossRef\]](#)

78. Schmid, P.; Adams, S.; Rugo, H.S.; Schneeweiss, A.; Barrios, C.H.; Iwata, H.; Diéras, V.; Hegg, R.; Im, S.-A.; Shaw Wright, G.; et al. Atezolizumab and Nab-Paclitaxel in Advanced Triple-Negative Breast Cancer. *N. Engl. J. Med.* **2018**, *379*, 2108–2121. [\[CrossRef\]](#)
79. He, R.; Zhao, X.; Liu, J.; Zhou, Y.; Zhang, X.; Cheng, F. PD-1 and CTLA-4 inhibitors in combination vs. alone for the treatment of advanced melanoma: A systematic review and meta-analysis. *Medicine* **2022**, *101*, e30561. [\[CrossRef\]](#) [\[PubMed\]](#)
80. Zhao, B.-W.; Zhang, F.-Y.; Wang, Y.; Chen, G.-M.; Nie, M.; Zhao, Z.-K.; Chen, X.-J.; Jiang, K.-M.; Nie, R.-C.; Chen, Y.-B. LAG3-PD1 or CTLA4-PD1 Inhibition in Advanced Melanoma: Indirect Cross Comparisons of the CheckMate-067 and RELATIVITY-047 Trials. *Cancers* **2022**, *14*, 4975. [\[CrossRef\]](#)
81. Kuske, M.; Westphal, D.; Wehner, R.; Schmitz, M.; Beissert, S.; Praetorius, C.; Meier, F. Immunomodulatory effects of BRAF and MEK inhibitors: Implications for Melanoma therapy. *Pharmacol. Res.* **2018**, *136*, 151–159. [\[CrossRef\]](#)
82. Subbiah, V.; Baik, C.; Kirkwood, J.M. Clinical Development of BRAF plus MEK Inhibitor Combinations. *Trends Cancer* **2020**, *6*, 797–810. [\[CrossRef\]](#)
83. Harding, C.; Stahl, P. Transferrin recycling in reticulocytes: pH and iron are important determinants of ligand binding and processing. *Biochem. Biophys. Res. Commun.* **1983**, *113*, 650–658. [\[CrossRef\]](#)
84. Pan, B.-T.; Johnstone, R.M. Fate of the transferrin receptor during maturation of sheep reticulocytes in vitro: Selective externalization of the receptor. *Cell* **1983**, *33*, 967–978. [\[CrossRef\]](#)
85. Lin, J.; Li, J.; Huang, B.; Liu, J.; Chen, X.; Chen, X.-M.; Xu, Y.-M.; Huang, L.-F.; Wang, X.-Z. Exosomes: Novel Biomarkers for Clinical Diagnosis. *Sci. World J.* **2015**, *2015*, 657086. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Yamashita, T.; Takahashi, Y.; Nishikawa, M.; Takakura, Y. Effect of exosome isolation methods on physicochemical properties of exosomes and clearance of exosomes from the blood circulation. *Eur. J. Pharm. Biopharm.* **2016**, *98*, 1–8. [\[CrossRef\]](#)
87. Janouskova, O.; Herma, R.; Semeradtova, A.; Poustka, D.; Liegertova, M.; Malinska, H.A.; Maly, J. Conventional and Nonconventional Sources of Exosomes—Isolation Methods and Influence on Their Downstream Biomedical Application. *Front. Mol. Biosci.* **2022**, *9*, 846650. [\[CrossRef\]](#) [\[PubMed\]](#)
88. Johnstone, R.M. Maturation of reticulocytes: Formation of exosomes as a mechanism for shedding membrane proteins. *Biochem. Cell Biol.* **1992**, *70*, 179–190. [\[CrossRef\]](#)
89. Couch, Y.; Buzàs, E.I.; Di Vizio, D.; Gho, Y.S.; Harrison, P.; Hill, A.F.; Lötvall, J.; Raposo, G.; Stahl, P.D.; Théry, C.; et al. A brief history of nearly EV-erything—The rise and rise of extracellular vesicles. *J. Extracell. Vesicles* **2021**, *10*, e12144. [\[CrossRef\]](#)
90. Lee, Y.J.; Jy, W.; Horstman, L.L.; Janania, J.; Reyes, Y.; Kelley, R.E.; Ahn, Y.S. Elevated platelet microparticles in transient ischemic attacks, lacunar infarcts, and multiinfarct dementias. *Thromb. Res.* **1993**, *72*, 295–304. [\[CrossRef\]](#)
91. Singh, N.; Gemmell, C.H.; A Daly, P.; Yeo, E.L. Elevated platelet-derived microparticle levels during unstable angina. *Can. J. Cardiol.* **1995**, *11*, 1015–1021.
92. Powell, J.J.; Harvey, R.; Thompson, R. Microparticles in Crohn's disease—has the dust settled? *Gut* **1996**, *39*, 340. [\[CrossRef\]](#)
93. Raposo, G.; Nijman, H.W.; Stoorvogel, W.; Liejendekker, R.; Harding, C.V.; Melief, C.J.; Geuze, H.J. B lymphocytes secrete antigen-presenting vesicles. *J. Exp. Med.* **1996**, *183*, 1161–1172. [\[CrossRef\]](#)
94. Zitvogel, L.; Regnault, A.; Lozier, A.; Wolfers, J.; Flament, C.; Tenza, D.; Ricciardi-Castagnoli, P.; Raposo, G.; Amigorena, S. Eradication of established murine tumors using a novel cell-free vaccine: Dendritic cell derived exosomes. *Nat. Med.* **1998**, *4*, 594–600. [\[CrossRef\]](#)
95. Escudier, B.; Dorval, T.; Chaput, N.; André, F.; Caby, M.-P.; Novault, S.; Flament, C.; Leboulleire, C.; Borg, C.; Amigorena, S.; et al. Vaccination of metastatic melanoma patients with autologous dendritic cell (DC) derived-exosomes: Results of the first phase I clinical trial. *J. Transl. Med.* **2005**, *3*, 10. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Scharltz, N.E.C.; Chaput, N.; André, F.; Zitvogel, L. From the antigen-presenting cell to the antigen-presenting vesicle: The exosomes. *Curr. Opin. Mol. Ther.* **2002**, *4*, 372–381.
97. Bard, M.P.; Hegmans, J.P.; Hemmes, A.; Luidier, T.M.; Willemsen, R.; Severijnen, L.-A.A.; van Meerbeeck, J.P.; Burgers, S.A.; Hoogsteden, H.C.; Lambrecht, B.N. Proteomic Analysis of Exosomes Isolated from Human Malignant Pleural Effusions. *Am. J. Respir. Cell Mol. Biol.* **2004**, *31*, 114–121. [\[CrossRef\]](#)
98. Skokos, D.; Botros, H.G.; Demeure, C.; Morin, J.; Peronet, R.; Birkenmeier, G.; Boudaly, S.; Mécheri, S. Mast Cell-Derived Exosomes Induce Phenotypic and Functional Maturation of Dendritic Cells and Elicit Specific Immune Responses In Vivo. *J. Immunol.* **2003**, *170*, 3037–3045. [\[CrossRef\]](#)
99. Wolfers, J.; Lozier, A.; Raposo, G.; Regnault, A.; Théry, C.; Masurier, C.; Flament, C.; Pouzieux, S.; Faure, F.; Tursz, T.; et al. Tumor-derived exosomes are a source of shared tumor rejection antigens for CTL cross-priming. *Nat. Med.* **2001**, *7*, 297–303. [\[CrossRef\]](#)
100. Chaput, N.; Scharltz, N.; Andre, F.; Zitvogel, L. Exosomes for immunotherapy of cancer. In *New Trends in Cancer for the 21st Century, Proceedings of the International Symposium on Cancer: New Trends in Cancer for the 21st Century, Valencia, Spain, 10–13 November 2002*; Springer: Cham, Switzerland, 2003.

101. Skog, J.; Würdinger, T.; Van Rijn, S.; Meijer, D.H.; Gainche, L.; Curry, W.T., Jr.; Carter, B.S.; Krichevsky, A.M.; Breakefield, X.O. Glioblastoma microvesicles transport RNA and proteins that promote tumour growth and provide diagnostic biomarkers. *Nat. Cell Biol.* **2008**, *10*, 1470–1476. [\[CrossRef\]](#)
102. An, Q.; van Bel, A.J.; Hükelhoven, R. Do plant cells secrete exosomes derived from multivesicular bodies? *Plant Signal. Behav.* **2007**, *2*, 4–7. [\[CrossRef\]](#)
103. Bruno, S.; Grange, C.; Deregibus, M.C.; Calogero, R.A.; Saviozzi, S.; Collino, F.; Morando, L.; Busca, A.; Falda, M.; Bussolati, B.; et al. Mesenchymal Stem Cell-Derived Microvesicles Protect Against Acute Tubular Injury. *J. Am. Soc. Nephrol.* **2009**, *20*, 1053–1067. [\[CrossRef\]](#)
104. Park, J.E.; Tan, H.S.; Datta, A.; Lai, R.C.; Zhang, H.; Meng, W.; Lim, S.K.; Sze, S.K. Hypoxic Tumor Cell Modulates Its Microenvironment to Enhance Angiogenic and Metastatic Potential by Secretion of Proteins and Exosomes. *Mol. Cell. Proteom.* **2010**, *9*, 1085–1099. [\[CrossRef\]](#) [\[PubMed\]](#)
105. Kumar, D.N.; Chaudhuri, A.; Aqil, F.; Dehari, D.; Munagala, R.; Singh, S.; Gupta, R.C.; Agrawal, A.K. Exosomes as Emerging Drug Delivery and Diagnostic Modality for Breast Cancer: Recent Advances in Isolation and Application. *Cancers* **2022**, *14*, 1435. [\[CrossRef\]](#)
106. Von Schulze, A.; Deng, F. A review on exosome-based cancer therapy. *J. Cancer Metastasis Treat.* **2020**, *6*, 42. [\[CrossRef\]](#)
107. Smyth, T.J.; Redzic, J.S.; Graner, M.W.; Anchordoquy, T.J. Examination of the specificity of tumor cell derived exosomes with tumor cells in vitro. *Biochim. Biophys. Acta (BBA) Biomembr.* **2014**, *1838*, 2954–2965. [\[CrossRef\]](#)
108. Qi, H.; Liu, C.; Long, L.; Ren, Y.; Zhang, S.; Chang, X.; Qian, X.; Jia, H.; Zhao, J.; Sun, J.; et al. Blood Exosomes Endowed with Magnetic and Targeting Properties for Cancer Therapy. *ACS Nano* **2016**, *10*, 3323–3333. [\[CrossRef\]](#)
109. Jang, S.C.; Kim, O.Y.; Yoon, C.M.; Choi, D.-S.; Roh, T.-Y.; Park, J.; Nilsson, J.; Lötvall, J.; Kim, Y.-K.; Gho, Y.S. Bioinspired Exosome-Mimetic Nanovesicles for Targeted Delivery of Chemotherapeutics to Malignant Tumors. *ACS Nano* **2013**, *7*, 7698–7710. [\[CrossRef\]](#)
110. Munagala, R.; Aqil, F.; Jeyabalan, J.; Gupta, R.C. Bovine milk-derived exosomes for drug delivery. *Cancer Lett.* **2016**, *371*, 48–61. [\[CrossRef\]](#) [\[PubMed\]](#)
111. Sun, W.; Luo, J.-D.; Jiang, H.; Duan, D.D. Tumor exosomes: A double-edged sword in cancer therapy. *Acta Pharmacol. Sin.* **2018**, *39*, 534–541. [\[CrossRef\]](#)
112. Osaki, M.; Okada, F. Exosomes and Their Role in Cancer Progression. *Yonago Acta Medica* **2019**, *62*, 182–190. [\[CrossRef\]](#)
113. Dilsiz, N. Role of Exosomes and Exosomal microRNAs in Cancer. *Futur. Sci. OA* **2020**, *6*, FSO465. [\[CrossRef\]](#)
114. Hao, S.; Bai, O.; Li, F.; Yuan, J.; Laferte, S.; Xiang, J. Mature dendritic cells pulsed with exosomes stimulate efficient cytotoxic T-lymphocyte responses and antitumour immunity. *Immunology* **2007**, *120*, 90–102. [\[CrossRef\]](#)
115. Pitt, J.M.; Charrier, M.; Viaud, S.; André, F.; Besse, B.; Chaput, N.; Zitvogel, L. Dendritic Cell-Derived Exosomes as Immunotherapies in the Fight against Cancer. *J. Immunol.* **2014**, *193*, 1006–1011. [\[CrossRef\]](#) [\[PubMed\]](#)
116. Awadasseid, A.; Wu, Y.; Zhang, W. Extracellular Vesicles (Exosomes) as Immunosuppressive Mediating Variables in Tumor and Chronic Inflammatory Microenvironments. *Cells* **2021**, *10*, 2533. [\[CrossRef\]](#)
117. Zech, D.; Rana, S.; Büchler, M.W.; Zöller, M. Tumor-exosomes and leukocyte activation: An ambivalent crosstalk. *Cell Commun. Signal.* **2012**, *10*, 37. [\[CrossRef\]](#)
118. Que, R.-s.; Lin, C.; Ding, G.; Wu, Z.; Cao, L. Increasing the immune activity of exosomes: The effect of miRNA-depleted exosome proteins on activating dendritic cell/cytokine-induced killer cells against pancreatic cancer. *J. Zhejiang Univ. Sci. B* **2016**, *17*, 352. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Huang, F.; Wan, J.; Hu, W.; Hao, S. Enhancement of Anti-Leukemia Immunity by Leukemia-Derived Exosomes Via Downregulation of TGF- β 1 Expression. *Cell. Physiol. Biochem.* **2017**, *44*, 240–254. [\[CrossRef\]](#)
120. Zhang, Y.; Luo, C.L.; He, B.C.; Zhang, J.M.; Cheng, G.; Wu, X.H. Exosomes derived from IL-12-anchored renal cancer cells increase induction of specific antitumor response in vitro: A novel vaccine for renal cell carcinoma. *Int. J. Oncol.* **2010**, *36*, 133–140.
121. Wen, S.W.; Sceneay, J.; Lima, L.G.; Wong, C.S.; Becker, M.; Krumeich, S.; Lobb, R.J.; Castillo, V.; Wong, K.N.; Ellis, S.; et al. The Biodistribution and Immune Suppressive Effects of Breast Cancer-Derived Exosomes Exosomes Regulate Immune Composition in Metastatic Organs. *Cancer Res.* **2016**, *76*, 6816–6827. [\[CrossRef\]](#)
122. Maybruck, B.T.; Pfannenstiel, L.W.; Diaz-Montero, M.; Gastman, B.R. Tumor-derived exosomes induce CD8⁺ T cell suppressors. *J. Immunother. Cancer* **2017**, *5*, 65. [\[CrossRef\]](#)
123. Bland, C.L.; Byrne-Hoffman, C.N.; Fernandez, A.; Rellick, S.L.; Deng, W.; Klinke, D.J. Exosomes derived from B16F0 melanoma cells alter the transcriptome of cytotoxic T cells that impacts mitochondrial respiration. *FEBS J.* **2018**, *285*, 1033–1050. [\[CrossRef\]](#)
124. Perez, C.R.; De Palma, M. Engineering dendritic cell vaccines to improve cancer immunotherapy. *Nat. Commun.* **2019**, *10*, 5408. [\[CrossRef\]](#) [\[PubMed\]](#)
125. Lindenbergh, M.F.S.; Wubolts, R.; Borg, E.G.F.; Van'T Veld, E.M.; Boes, M.; Stoorvogel, W. Dendritic cells release exosomes together with phagocytosed pathogen; potential implications for the role of exosomes in antigen presentation. *J. Extracell. Vesicles* **2020**, *9*, 1798606. [\[CrossRef\]](#)

126. Chaput, N.; Angevin, E.; Zitvogel, L.; Taïeb, J.; Scharztz, N.E.C.; André, F. Exosome-based immunotherapy. *Cancer Immunol. Immunother.* **2004**, *53*, 234–239. [[CrossRef](#)] [[PubMed](#)]
127. Viaud, S.; Théry, C.; Ploix, S.; Tursz, T.; Lapierre, V.; Lantz, O.; Zitvogel, L.; Chaput, N. Dendritic Cell-Derived Exosomes for Cancer Immunotherapy: What's Next? Dendritic Cell-Derived Exosomes Immunotherapy. *Cancer Res.* **2010**, *70*, 1281–1285. [[CrossRef](#)]
128. Hao, S.; Liu, Y.; Yuan, J.; Zhang, X.; He, T.; Wu, X.; Wei, Y.; Sun, D.; Xiang, J. Novel Exosome-Targeted CD4+ T Cell Vaccine Counteracting CD4+25+ Regulatory T Cell-Mediated Immune Suppression and Stimulating Efficient Central Memory CD8+ CTL Responses. *J. Immunol.* **2007**, *179*, 2731–2740. [[CrossRef](#)]
129. Amigorena, S. Cancer immunotherapy using dendritic cell-derived exosomes. *Med.-Buenos Aires* **2000**, *60*, 51–54.
130. Wang, L.; Xie, Y.; Ahmed, K.A.; Ahmed, S.; Sami, A.; Chibbar, R.; Xu, Q.; Kane, S.E.; Hao, S.; Mulligan, S.J.; et al. Exosomal pMHC-I complex targets T cell-based vaccine to directly stimulate CTL responses leading to antitumor immunity in transgenic FVBneun and HLA-A2/HER2 mice and eradicating trastuzumab-resistant tumor in athymic nude mice. *Breast Cancer Res. Treat.* **2013**, *140*, 273–284. [[CrossRef](#)]
131. Lu, Z.; Zuo, B.; Jing, R.; Gao, X.; Rao, Q.; Liu, Z.; Qi, H.; Guo, H.; Yin, H. Dendritic cell-derived exosomes elicit tumor regression in autochthonous hepatocellular carcinoma mouse models. *J. Hepatol.* **2017**, *67*, 739–748. [[CrossRef](#)]
132. Chen, Z.; You, L.; Wang, L.; Huang, X.; Liu, H.; Wei, J.Y.; Zhu, L.; Qian, W. Dual effect of DLBCL-derived EXOs in lymphoma to improve DC vaccine efficacy in vitro while favor tumorigenesis in vivo. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 190. [[CrossRef](#)]
133. Klinker, M.W.; Lizzio, V.; Reed, T.J.; Fox, D.A.; Lundy, S.K. Human B cell-derived lymphoblastoid cell lines constitutively produce Fas ligand and secrete MHCII+ FasL+ killer exosomes. *Front. Immunol.* **2014**, *5*, 144. [[CrossRef](#)]
134. Chen, W.; Wang, J.; Shao, C.; Liu, S.; Yu, Y.; Wang, Q.; Cao, X. Efficient induction of antitumor T cell immunity by exosomes derived from heat-shocked lymphoma cells. *Eur. J. Immunol.* **2006**, *36*, 1598–1607. [[CrossRef](#)]
135. Neelapu, S.S.; Tummala, S.; Kebriaei, P.; Wierda, W.; Gutierrez, C.; Locke, F.L.; Komanduri, K.V.; Lin, Y.; Jain, N.; Daver, N.; et al. Chimeric antigen receptor T-cell therapy—Assessment and management of toxicities. *Nat. Rev. Clin. Oncol.* **2017**, *15*, 47–62. [[CrossRef](#)]
136. Fu, W.; Lei, C.; Liu, S.; Cui, Y.; Wang, C.; Qian, K.; Li, T.; Shen, Y.; Fan, X.; Lin, F.; et al. CAR exosomes derived from effector CAR-T cells have potent antitumor effects and low toxicity. *Nat. Commun.* **2019**, *10*, 4355. [[CrossRef](#)]
137. Tang, X.-J.; Sun, X.-Y.; Huang, K.-M.; Zhang, L.; Yang, Z.-S.; Zou, D.-D.; Wang, B.; Warnock, G.L.; Dai, L.-J.; Luo, J. Therapeutic potential of CAR-T cell-derived exosomes: A cell-free modality for targeted cancer therapy. *Oncotarget* **2015**, *6*, 44179–44190. [[CrossRef](#)] [[PubMed](#)]
138. Seo, N.; Shirakura, Y.; Tahara, Y.; Momose, F.; Harada, N.; Ikeda, H.; Akiyoshi, K.; Shiku, H. Activated CD8+ T cell extracellular vesicles prevent tumour progression by targeting of lesional mesenchymal cells. *Nat. Commun.* **2018**, *9*, 435. [[CrossRef](#)] [[PubMed](#)]
139. Mittelbrunn, M.; Gutiérrez-Vázquez, C.; Villarroya-Beltrí, C.; González, S.; Sánchez-Cabo, F.; González, M.Á.; Bernad, A.; Sánchez-Madrid, F. Unidirectional transfer of microRNA-loaded exosomes from T cells to antigen-presenting cells. *Nat. Commun.* **2011**, *2*, 282. [[CrossRef](#)] [[PubMed](#)]
140. Tumne, A.; Prasad, V.S.; Chen, Y.; Stolz, D.B.; Saha, K.; Ratner, D.M.; Ding, M.; Watkins, S.C.; Gupta, P. Noncytotoxic suppression of human immunodeficiency virus type 1 transcription by exosomes secreted from CD8+ T cells. *J. Virol.* **2009**, *83*, 4354–4364. [[CrossRef](#)]
141. Azimi, M.; Ghabaee, M.; Moghadas, A.N.; Izad, M. Altered Expression of miR-326 in T Cell-derived Exosomes of Patients with Relapsing-remitting Multiple Sclerosis. *Iran. J. Allergy Asthma Immunol.* **2019**, *18*, 108–113. [[CrossRef](#)]
142. de Carvalho, J.V.; de Castro, R.O.; da Silva, E.Z.; Silveira, P.P.; da Silva-Januario, M.E.; Arruda, E.; Jamur, M.C.; Oliver, C.; Aguiar, R.S.; DaSilva, L.L.P. Nef neutralizes the ability of exosomes from CD4+ T cells to act as decoys during HIV-1 infection. *PLoS ONE* **2014**, *9*, e113691. [[CrossRef](#)]
143. Zakharova, L.; Svetlova, M.; Fomina, A.F. T cell exosomes induce cholesterol accumulation in human monocytes via phosphatidylserine receptor. *J. Cell. Physiol.* **2007**, *212*, 174–181. [[CrossRef](#)]
144. Fais, S. NK cell-released exosomes: Natural nanobullets against tumors. *Oncoimmunology* **2013**, *2*, e22337. [[CrossRef](#)]
145. Di Pace, A.L.; Tumino, N.; Besi, F.; Alicata, C.; Conti, L.A.; Munari, E.; Maggi, E.; Vacca, P.; Moretta, L. Characterization of human NK cell-derived exosomes: Role of DNAM1 receptor in exosome-mediated cytotoxicity against tumor. *Cancers* **2020**, *12*, 661. [[CrossRef](#)] [[PubMed](#)]
146. Wu, C.; Li, J.; Li, L.; Sun, J.; Fabbri, M.; Wayne, A.S.; Seeger, R.C.; Jong, A.Y. Extracellular vesicles derived from natural killer cells use multiple cytotoxic proteins and killing mechanisms to target cancer cells. *J. Extracell. Vesicles* **2019**, *8*, 1588538. [[CrossRef](#)]
147. Fu, Y.; Liu, S.; Zeng, S.; Shen, H. From bench to bed: The tumor immune microenvironment and current immunotherapeutic strategies for hepatocellular carcinoma. *J. Exp. Clin. Cancer Res.* **2019**, *38*, 396. [[CrossRef](#)] [[PubMed](#)]
148. Zöller, M.; Zhao, K.; Kutlu, N.N.; Bauer, N.; Provaznik, J.; Hackert, T.; Schnölzer, M. Immunoregulatory Effects of Myeloid-Derived Suppressor Cell Exosomes in Mouse Model of Autoimmune Alopecia Areata. *Front. Immunol.* **2018**, *9*, 1279. [[CrossRef](#)] [[PubMed](#)]

149. Geis-Asteggianti, L.; Belew, A.T.; Clements, V.K.; Edwards, N.J.; Ostrand-Rosenberg, S.; El-Sayed, N.M.; Fenselau, C. Differential Content of Proteins, mRNAs, and miRNAs Suggests that MDSC and Their Exosomes May Mediate Distinct Immune Suppressive Functions. *J. Proteome Res.* **2017**, *17*, 486–498. [[CrossRef](#)]
150. Deng, Z.; Rong, Y.; Teng, Y.; Zhuang, X.; Samykutty, A.; Mu, J.; Zhang, L.; Cao, P.; Yan, J.; Miller, D.; et al. Exosomes miR-126a released from MDSC induced by DOX treatment promotes lung metastasis. *Oncogene* **2016**, *36*, 639–651. [[CrossRef](#)]
151. Cheng, L.; Wang, Y.; Huang, L. Exosomes from M1-Polarized Macrophages Potentiate the Cancer Vaccine by Creating a Pro-inflammatory Microenvironment in the Lymph Node. *Mol. Ther.* **2017**, *25*, 1665–1675. [[CrossRef](#)]
152. Singhto, N.; Kanlaya, R.; Nilnumkhum, A.; Thongboonkerd, V. Roles of Macrophage Exosomes in Immune Response to Calcium Oxalate Monohydrate Crystals. *Front. Immunol.* **2018**, *9*, 316. [[CrossRef](#)]
153. Lan, J.; Sun, L.; Xu, F.; Liu, L.; Hu, F.; Song, D.; Hou, Z.; Wu, W.; Luo, X.; Wang, J.; et al. M2 Macrophage-Derived Exosomes Promote Cell Migration and Invasion in Colon Cancer. *Cancer Res.* **2019**, *79*, 146–158. [[CrossRef](#)]
154. Cianciaruso, C.; Beltraminelli, T.; Duval, F.; Nassiri, S.; Hamelin, R.; Mozes, A.; Gallart-Ayala, H.; Torres, G.C.; Torchia, B.; Ries, C.H.; et al. Molecular Profiling and Functional Analysis of Macrophage-Derived Tumor Extracellular Vesicles. *Cell Rep.* **2019**, *27*, 3062–3080.e11. [[CrossRef](#)]
155. Li, M.; Zhao, J.; Cao, M.; Liu, R.; Chen, G.; Li, S.; Xie, Y.; Xie, J.; Cheng, Y.; Huang, L.; et al. Mast cells-derived MiR-223 destroys intestinal barrier function by inhibition of CLDN8 expression in intestinal epithelial cells. *Biol. Res.* **2020**, *53*, 12. [[CrossRef](#)] [[PubMed](#)]
156. Xiao, H.; Lässer, C.; Shelke, G.V.; Wang, J.; Rådinger, M.; Lunavat, T.R.; Malmhäll, C.; Lin, L.H.; Li, J.; Li, L.; et al. Mast cell exosomes promote lung adenocarcinoma cell proliferation—Role of KIT-stem cell factor signaling. *Cell Commun. Signal.* **2014**, *12*, 64. [[CrossRef](#)] [[PubMed](#)]
157. Li, F.; Wang, Y.; Lin, L.; Wang, J.; Xiao, H.; Li, J.; Peng, X.; Dai, H.; Li, L. Mast Cell-Derived Exosomes Promote Th2 Cell Differentiation via OX40L-OX40 Ligation. *J. Immunol. Res.* **2016**, *2016*, 3623898. [[CrossRef](#)]
158. Genschmer, K.R.; Russell, D.W.; Lal, C.; Szul, T.; Bratcher, P.E.; Noerager, B.D.; Roda, M.A.; Xu, X.; Rezonzew, G.; Viera, L.; et al. Activated PMN Exosomes: Pathogenic Entities Causing Matrix Destruction and Disease in the Lung. *Cell* **2019**, *176*, 113–126.e15. [[CrossRef](#)]
159. Li, L.; Zuo, X.; Xiao, Y.; Liu, D.; Luo, H.; Zhu, H. Neutrophil-derived exosome from systemic sclerosis inhibits the proliferation and migration of endothelial cells. *Biochem. Biophys. Res. Commun.* **2020**, *526*, 334–340. [[CrossRef](#)]
160. Vargas, A.; Roux-Dalvai, F.; Droit, A.; Lavoie, J.-P. Neutrophil-Derived Exosomes: A New Mechanism Contributing to Airway Smooth Muscle Remodeling. *Am. J. Respir. Cell Mol. Biol.* **2016**, *55*, 450–461. [[CrossRef](#)]
161. Martinon, F.; Burns, K.; Tschopp, J. The inflammasome: A molecular platform triggering activation of inflammatory caspases and processing of proIL- β . *Mol. Cell* **2002**, *10*, 417–426. [[CrossRef](#)]
162. Broz, P.; Pelegrin, P.; Shao, F. The gasdermins, a protein family executing cell death and inflammation. *Nat. Rev. Immunol.* **2020**, *20*, 143–157. [[CrossRef](#)]
163. Ariffin, J.K.; Sweet, M.J. Differences in the repertoire, regulation and function of Toll-like Receptors and inflammasome-forming Nod-like Receptors between human and mouse. *Curr. Opin. Microbiol.* **2013**, *16*, 303–310. [[CrossRef](#)]
164. Sauter, K.A.; Wood, L.J.; Wong, J.; Iordanov, M.; Magun, B.E. Doxorubicin and daunorubicin induce processing and release of interleukin-1 β through activation of the NLRP3 inflammasome: Progress at a snail's pace. *Cancer Biol. Ther.* **2011**, *11*, 1008–1016. [[CrossRef](#)] [[PubMed](#)]
165. Bürckstümmer, T.; Baumann, C.; Blüml, S.; Dixit, E.; Dürnberger, G.; Jahn, H.; Planyavsky, M.; Bilban, M.; Colinge, J.; Bennett, K.L.; et al. An orthogonal proteomic-genomic screen identifies AIM2 as a cytoplasmic DNA sensor for the inflammasome. *Nat. Immunol.* **2009**, *10*, 266–272. [[CrossRef](#)]
166. Fernandes-Alnemri, T.; Yu, J.-W.; Datta, P.; Wu, J.; Alnemri, E.S. AIM2 activates the inflammasome and cell death in response to cytoplasmic DNA. *Nature* **2009**, *458*, 509–513. [[CrossRef](#)]
167. Lightfield, K.L.; Persson, J.; Brubaker, S.W.; E Witte, C.; von Moltke, J.; A Dunipace, E.; Henry, T.; Sun, Y.-H.; Cado, D.; Dietrich, W.F.; et al. Critical function for Naip5 in inflammasome activation by a conserved carboxy-terminal domain of flagellin. *Nat. Immunol.* **2008**, *9*, 1171–1178. [[CrossRef](#)]
168. Zhao, Y.; Yang, J.; Shi, J.; Gong, Y.-N.; Lu, Q.; Xu, H.; Liu, L.; Shao, F. The NLRC4 inflammasome receptors for bacterial flagellin and type III secretion apparatus. *Nature* **2011**, *477*, 596–600. [[CrossRef](#)]
169. Xu, H.; Yang, J.; Gao, W.; Li, L.; Li, P.; Zhang, L.; Gong, Y.-N.; Peng, X.; Xi, J.J.; Chen, S.; et al. Innate immune sensing of bacterial modifications of Rho GTPases by the Pyrin inflammasome. *Nature* **2014**, *513*, 237–241. [[CrossRef](#)] [[PubMed](#)]
170. Tentherey, J.L.; Haloupek, N.; López-Blanco, J.R.; Grob, P.; Adamson, E.; Hartenian, E.; Lind, N.A.; Bourgeois, N.M.; Chacón, P.; Nogales, E.; et al. The structural basis of flagellin detection by NAIP5: A strategy to limit pathogen immune evasion. *Science* **2017**, *358*, 888–893. [[CrossRef](#)]
171. Swanson, K.V.; Deng, M.; Ting, J.P.-Y. The NLRP3 inflammasome: Molecular activation and regulation to therapeutics. *Nat. Rev. Immunol.* **2019**, *19*, 477–489. [[CrossRef](#)]

172. Baldwin, A.G.; Brough, D.; Freeman, S. Inhibiting the Inflammasome: A Chemical Perspective. *J. Med. Chem.* **2016**, *59*, 1691–1710. [\[CrossRef\]](#)
173. Shadab, A.; Mahjoor, M.; Abbasi-Kolli, M.; Afkhami, H.; Moeinian, P.; Safdarian, A.-R. Divergent functions of NLRP3 inflammasomes in cancer: A review. *Cell Commun. Signal.* **2023**, *21*, 232. [\[CrossRef\]](#)
174. Fleischmann, R.M.; Schechtman, J.; Bennett, R.; Handel, M.L.; Burmester, G.R.; Tesser, J.; Modafferi, D.; Poulakos, J.; Sun, G. Anakinra, a recombinant human interleukin-1 receptor antagonist (r-metHuIL-1ra), in patients with rheumatoid arthritis: A large, international, multicenter, placebo-controlled trial. *Arthritis Rheum. Off. J. Am. Coll. Rheumatol.* **2003**, *48*, 927–934. [\[CrossRef\]](#) [\[PubMed\]](#)
175. He, A.; Shao, J.; Zhang, Y.; Lu, H.; Wu, Z.; Xu, Y. CD200Fc reduces LPS-induced IL-1 β activation in human cervical cancer cells by modulating TLR4-NF- κ B and NLRP3 inflammasome pathway. *Oncotarget* **2017**, *8*, 33214. [\[CrossRef\]](#)
176. Li, S.; Liang, X.; Ma, L.; Shen, L.; Li, T.; Zheng, L.; Sun, A.; Shang, W.; Chen, C.; Zhao, W.; et al. MiR-22 sustains NLRP3 expression and attenuates H. pylori-induced gastric carcinogenesis. *Oncogene* **2017**, *37*, 884–896. [\[CrossRef\]](#)
177. Tang, Z.; Ji, L.; Han, M.; Xie, J.; Zhong, F.; Zhang, X.; Su, Q.; Yang, Z.; Liu, Z.; Gao, H.; et al. Pyroptosis is involved in the inhibitory effect of FL118 on growth and metastasis in colorectal cancer. *Life Sci.* **2020**, *257*, 118065. [\[CrossRef\]](#)
178. Xu, L.; Bi, Y.; Xu, Y.; Zhang, Z.; Xu, W.; Zhang, S.; Chen, J. Oridonin inhibits the migration and epithelial-to-mesenchymal transition of small cell lung cancer cells by suppressing FAK-ERK1/2 signalling pathway. *J. Cell. Mol. Med.* **2020**, *24*, 4480–4493. [\[CrossRef\]](#)
179. Yaw, A.C.K.; Chan, E.W.L.; Yap, J.K.Y.; Mai, C.W. The effects of NLRP3 inflammasome inhibition by MCC950 on LPS-induced pancreatic adenocarcinoma inflammation. *J. Cancer Res. Clin. Oncol.* **2020**, *146*, 2219–2229. [\[CrossRef\]](#)
180. Tengesdal, I.W.; Dinarello, A.; Powers, N.E.; Burchill, M.A.; Joosten, L.A.; Marchetti, C.; Dinarello, C.A. Tumor NLRP3-derived IL-1 β drives the IL-6/STAT3 axis resulting in sustained MDSC-mediated immunosuppression. *Front. Immunol.* **2021**, *12*, 661323. [\[CrossRef\]](#)
181. Tengesdal, I.W.; Menon, D.R.; Osborne, D.G.; Neff, C.P.; Powers, N.E.; Gamboni, F.; Mauro, A.G.; D'alessandro, A.; Stefanoni, D.; Henen, M.A.; et al. Targeting tumor-derived NLRP3 reduces melanoma progression by limiting MDSCs expansion. *Proc. Natl. Acad. Sci. USA* **2021**, *118*, e2000915118. [\[CrossRef\]](#)
182. Saito, H.; Fushida, S.; Harada, S.; Miyashita, T.; Oyama, K.; Yamaguchi, T.; Tsukada, T.; Kinoshita, J.; Tajima, H.; Ninomiya, I.; et al. Importance of human peritoneal mesothelial cells in the progression, fibrosis, and control of gastric cancer: Inhibition of growth and fibrosis by tranilast. *Gastric Cancer* **2017**, *21*, 55–67. [\[CrossRef\]](#) [\[PubMed\]](#)
183. Yang, J.; Ren, X.; Zhang, L.; Li, Y.; Cheng, B.; Xia, J. Oridonin inhibits oral cancer growth and PI3K/Akt signaling pathway. *Biomed. Pharmacother.* **2018**, *100*, 226–232. [\[CrossRef\]](#) [\[PubMed\]](#)
184. Takahashi, K.; Menju, T.; Nishikawa, S.; Miyata, R.; Tanaka, S.; Yutaka, Y.; Yamada, Y.; Nakajima, D.; Hamaji, M.; Ohsumi, A.; et al. Tranilast Inhibits TGF- β 1-induced Epithelial-mesenchymal Transition and Invasion/Metastasis via the Suppression of Smad4 in Human Lung Cancer Cell Lines. *Anticancer Res.* **2020**, *40*, 3287–3296. [\[CrossRef\]](#)
185. Zheng, Q.; Yao, D.; Cai, Y.; Zhou, T. NLRP3 augmented resistance to gemcitabine in triple-negative breast cancer cells via EMT/IL-1 β /Wnt/ β -catenin signaling pathway. *Biosci. Rep.* **2020**, *40*, 1–9. [\[CrossRef\]](#) [\[PubMed\]](#)
186. Shiravand, Y.; Khodadadi, F.; Kashani, S.M.A.; Hosseini-Fard, S.R.; Hosseini, S.; Sadeghirad, H.; Ladwa, R.; O'byrne, K.; Kulasinghe, A. Immune Checkpoint Inhibitors in Cancer Therapy. *Curr. Oncol.* **2022**, *29*, 3044–3060. [\[CrossRef\]](#)
187. Jiao, Z.; Zhang, J. Interplay between inflammasomes and PD-1/PD-L1 and their implications in cancer immunotherapy. *Carcinogenesis* **2023**, *44*, 795–808. [\[CrossRef\]](#)
188. Honda, H.; Nagai, Y.; Matsunaga, T.; Okamoto, N.; Watanabe, Y.; Tsuneyama, K.; Hayashi, H.; Fujii, I.; Ikutani, M.; Hirai, Y.; et al. Isoliquiritigenin is a potent inhibitor of NLRP3 inflammasome activation and diet-induced adipose tissue inflammation. *J. Leukoc. Biol.* **2014**, *96*, 1087–1100. [\[CrossRef\]](#)
189. Ahn, H.; Kang, S.G.; Yoon, S.; Ko, H.J.; Kim, P.H.; Hong, E.J.; An, B.S.; Lee, E.; Lee, G.S. Methylene blue inhibits NLRP3, NLRC4, AIM2, and non-canonical inflammasome activation. *Sci. Rep.* **2017**, *7*, 12409. [\[CrossRef\]](#)
190. Guo, W.; Sun, Y.; Liu, W.; Wu, X.; Guo, L.; Cai, P.; Wu, X.; Wu, X.; Shen, Y.; Shu, Y.; et al. Small molecule-driven mitophagy-mediated NLRP3 inflammasome inhibition is responsible for the prevention of colitis-associated cancer. *Autophagy* **2014**, *10*, 972–985. [\[CrossRef\]](#) [\[PubMed\]](#)
191. Zhang, Z.; Li, X.; Wang, Y.; Wei, Y.; Wei, X. Involvement of inflammasomes in tumor microenvironment and tumor therapies. *J. Hematol. Oncol.* **2023**, *16*, 24. [\[CrossRef\]](#)
192. Teng, J.-F.; Mei, Q.B.; Zhou, X.G.; Tang, Y.; Xiong, R.; Qiu, W.Q.; Pan, R.; Law, B.Y.-K.; Wong, V.K.-W.; Yu, C.-L.; et al. Polyphyllin VI induces caspase-1-mediated pyroptosis via the induction of ROS/NF- κ B/NLRP3/GSDMD signal axis in non-small cell lung cancer. *Cancers* **2020**, *12*, 193. [\[CrossRef\]](#)
193. Hu, B.; Elinav, E.; Huber, S.; Booth, C.J.; Strowig, T.; Jin, C.; Eisenbarth, S.C.; Flavell, R.A. Inflammation-induced tumorigenesis in the colon is regulated by caspase-1 and NLRC4. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 21635–21640. [\[CrossRef\]](#)

194. Ikuta, T.; Kobayashi, Y.; Kitazawa, M.; Shiizaki, K.; Itano, N.; Noda, T.; Pettersson, S.; Poellinger, L.; Fujii-Kuriyama, Y.; Taniguchi, S.; et al. ASC-associated inflammation promotes cecal tumorigenesis in aryl hydrocarbon receptor-deficient mice. *Carcinogenesis* **2013**, *34*, 1620–1627. [[CrossRef](#)] [[PubMed](#)]
195. Gasparoto, T.H.; de Oliveira, C.E.; de Freitas, L.T.; Pinheiro, C.R.; Hori, J.I.; Garlet, G.P.; Cavassani, K.A.; Schillaci, R.; da Silva, J.S.; Zamboni, D.S.; et al. Inflammasome Activation Is Critical to the Protective Immune Response during Chemically Induced Squamous Cell Carcinoma. *PLoS ONE* **2014**, *9*, e107170. [[CrossRef](#)]
196. Feng, X.; Luo, Q.; Zhang, H.; Wang, H.; Chen, W.; Meng, G.; Chen, F. The role of NLRP3 inflammasome in 5-fluorouracil resistance of oral squamous cell carcinoma. *J. Exp. Clin. Cancer Res.* **2017**, *36*, 81. [[CrossRef](#)] [[PubMed](#)]
197. Wei, Q.; Guo, P.; Mu, K.; Zhang, Y.; Zhao, W.; Huai, W.; Qiu, Y.; Li, T.; Ma, X.; Liu, Y.; et al. Estrogen suppresses hepatocellular carcinoma cells through ER β -mediated upregulation of the NLRP3 inflammasome. *Mod. Pathol.* **2015**, *95*, 804–816. [[CrossRef](#)]
198. Wei, Q.; Zhu, R.; Zhu, J.; Zhao, R.; Li, M. E2-Induced Activation of the NLRP3 Inflammasome Triggers Pyroptosis and Inhibits Autophagy in HCC Cells. *Oncol. Res. Featur. Preclin. Clin. Cancer Ther.* **2019**, *27*, 827–834. [[CrossRef](#)]
199. Lu, F.; Zhao, Y.; Pang, Y.; Ji, M.; Sun, Y.; Wang, H.; Zou, J.; Wang, Y.; Li, G.; Sun, T.; et al. NLRP3 inflammasome upregulates PD-L1 expression and contributes to immune suppression in lymphoma. *Cancer Lett.* **2021**, *497*, 178–189. [[CrossRef](#)]
200. Zhong, F.L.; Mamaï, O.; Sborgi, L.; Boussofara, L.; Hopkins, R.; Robinson, K.; Szeverényi, I.; Takeichi, T.; Balaji, R.; Lau, A.; et al. Germline NLRP1 Mutations Cause Skin Inflammatory and Cancer Susceptibility Syndromes via Inflammasome Activation. *Cell* **2016**, *167*, 187–202.e17. [[CrossRef](#)] [[PubMed](#)]
201. Verma, D.; Bivik, C.; Farahani, E.; Synnerstad, I.; Fredrikson, M.; Enerbäck, C.; Rosdahl, I.; Söderkvist, P. Inflammasome polymorphisms confer susceptibility to sporadic malignant melanoma. *Pigment. Cell Melanoma Res.* **2012**, *25*, 506–513. [[CrossRef](#)]
202. Castaño-Rodríguez, N.; Kaakoush, N.O.; Goh, K.-L.; Fock, K.M.; Mitchell, H.M. The NOD-Like Receptor Signalling Pathway in *Helicobacter pylori* Infection and Related Gastric Cancer: A Case-Control Study and Gene Expression Analyses. *PLoS ONE* **2014**, *9*, e98899. [[CrossRef](#)]
203. Miskiewicz, A.; Szparecki, G.; Durlik, M.; Rydzewska, G.; Ziobrowski, I.; Górska, R. The Q705K and F359L Single-Nucleotide Polymorphisms of NOD-Like Receptor Signaling Pathway: Association with Chronic Pancreatitis, Pancreatic Cancer, and Periodontitis. *Arch. Immunol. Ther. Exp.* **2015**, *63*, 485–494. [[CrossRef](#)]
204. Deswaerte, V.; Nguyen, P.; West, A.; Browning, A.F.; Yu, L.; Ruwanpura, S.M.; Balic, J.; Livis, T.; Girard, C.; Preaudet, A.; et al. Inflammasome Adaptor ASC Suppresses Apoptosis of Gastric Cancer Cells by an IL18-Mediated Inflammation-Independent Mechanism. *Cancer Res.* **2018**, *78*, 1293–1307. [[CrossRef](#)] [[PubMed](#)]
205. Tu, S.; Bhagat, G.; Cui, G.; Takaishi, S.; Kurt-Jones, E.A.; Rickman, B.; Betz, K.S.; Penz-Oesterreicher, M.; Bjorkdahl, O.; Fox, J.G.; et al. Overexpression of Interleukin-1 β Induces Gastric Inflammation and Cancer and Mobilizes Myeloid-Derived Suppressor Cells in Mice. *Cancer Cell* **2008**, *14*, 408–419. [[CrossRef](#)]
206. Wei, Q.; Mu, K.; Li, T.; Zhang, Y.; Yang, Z.; Jia, X.; Zhao, W.; Huai, W.; Guo, P.; Han, L. Deregulation of the NLRP3 inflammasome in hepatic parenchymal cells during liver cancer progression. *Mod. Pathol.* **2014**, *94*, 52–62. [[CrossRef](#)] [[PubMed](#)]
207. Allen, I.C.; TeKippe, E.M.; Woodford, R.-M.T.; Uronis, J.M.; Holl, E.K.; Rogers, A.B.; Herfarth, H.H.; Jobin, C.; Ting, J.P.-Y. The NLRP3 inflammasome functions as a negative regulator of tumorigenesis during colitis-associated cancer. *J. Exp. Med.* **2010**, *207*, 1045–1056. [[CrossRef](#)]
208. Zaki, M.H.; Vogel, P.; Body-Malapel, M.; Lamkanfi, M.; Kanneganti, T.-D. IL-18 Production Downstream of the Nlrp3 Inflammasome Confers Protection against Colorectal Tumor Formation. *J. Immunol.* **2010**, *185*, 4912–4920. [[CrossRef](#)]
209. Sharma, D.; Malik, A.; Guy, C.S.; Karki, R.; Vogel, P.; Kanneganti, T.-D. Pyrin Inflammasome Regulates Tight Junction Integrity to Restrict Colitis and Tumorigenesis. *Gastroenterology* **2018**, *154*, 948–964.e8. [[CrossRef](#)]
210. Liu, W.; Luo, Y.; Dunn, J.H.; Norris, D.A.; Dinarello, C.A.; Fujita, M. Dual Role of Apoptosis-Associated Speck-Like Protein Containing a CARD (ASC) in Tumorigenesis of Human Melanoma. *J. Investig. Dermatol.* **2013**, *133*, 518–527. [[CrossRef](#)] [[PubMed](#)]
211. Drexler, S.K.; Bonsignore, L.; Masin, M.; Tardivel, A.; Jackstadt, R.; Hermeking, H.; Schneider, P.; Gross, O.; Tschopp, J.; Yazdi, A.S. Tissue-specific opposing functions of the inflammasome adaptor ASC in the regulation of epithelial skin carcinogenesis. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 18384–18389. [[CrossRef](#)]
212. Gao, J.; Qiu, X.; Xi, G.; Liu, H.; Zhang, F.; Lv, T.; Song, Y. Downregulation of GSDMD attenuates tumor proliferation via the intrinsic mitochondrial apoptotic pathway and inhibition of EGFR/Akt signaling and predicts a good prognosis in non-small cell lung cancer. *Oncol. Rep.* **2018**, *40*, 1971–1984. [[CrossRef](#)]
213. Wang, W.J.; Chen, D.; Jiang, M.Z.; Xu, B.; Li, X.W.; Chu, Y.; Zhang, Y.J.; Mao, R.; Liang, J.; Fan, D.M. Downregulation of gasdermin D promotes gastric cancer proliferation by regulating cell cycle-related proteins. *J. Dig. Dis.* **2018**, *19*, 74–83. [[CrossRef](#)]
214. Bent, R.; Moll, L.; Grabbe, S.; Bros, M. Interleukin-1 beta—A friend or foe in malignancies? *Int. J. Mol. Sci.* **2018**, *19*, 2155. [[CrossRef](#)]
215. Yang, G.; Kang, H.C.; Cho, Y.-Y.; Lee, H.S.; Lee, J.Y. Inhibition of NLRP3 inflammasome in tumor microenvironment leads to suppression of metastatic potential of cancer cells. *Sci. Rep.* **2019**, *9*, 12277. [[CrossRef](#)]

216. Horio, D.; Minami, T.; Kitai, H.; Ishigaki, H.; Higashiguchi, Y.; Kondo, N.; Hirota, S.; Kitajima, K.; Nakajima, Y.; Koda, Y.; et al. Tumor-associated macrophage-derived inflammatory cytokine enhances malignant potential of malignant pleural mesothelioma. *Cancer Sci.* **2020**, *111*, 2895–2906. [[CrossRef](#)] [[PubMed](#)]
217. Jin, H.; Ko, Y.S.; Kim, H.J. P2Y2R-mediated inflammasome activation is involved in tumor progression in breast cancer cells and in radiotherapy-resistant breast cancer. *Int. J. Oncol.* **2018**, *53*, 1953–1966. [[CrossRef](#)] [[PubMed](#)]
218. Yang, D.; Cao, X.; Wang, F.; Jiang, H.; Feng, D.; Guo, H.; Du, L.; Jin, Y.; Chen, Y.; Yin, X.; et al. LFG-500, a novel synthetic flavonoid, suppresses epithelial–mesenchymal transition in human lung adenocarcinoma cells by inhibiting NLRP3 in inflammatory microenvironment. *Cancer Lett.* **2017**, *400*, 137–148. [[CrossRef](#)]
219. Tulotta, C.; Lefley, D.V.; Freeman, K.; Gregory, W.M.; Hanby, A.M.; Heath, P.R.; Nutter, F.; Wilkinson, J.M.; Spicer-Hadlington, A.R.; Liu, X.; et al. Endogenous Production of IL1B by Breast Cancer Cells Drives Metastasis and Colonization of the Bone Microenvironment. *Clin. Cancer Res.* **2019**, *25*, 2769–2782. [[CrossRef](#)]
220. Zhang, Y.; Yang, H.; Sun, M.; He, T.; Liu, Y.; Yang, X.; Shi, X.; Liu, X. Alpinumisoflavone suppresses hepatocellular carcinoma cell growth and metastasis via NLRP3 inflammasome-mediated pyroptosis. *Pharmacol. Rep.* **2020**, *72*, 1370–1382. [[CrossRef](#)]
221. Dupaul-Chicoine, J.; Arabzadeh, A.; Dagenais, M.; Douglas, T.; Champagne, C.; Morizot, A.; Rodrigue-Gervais, I.G.; Breton, V.; Colpitts, S.L.; Beauchemin, N.; et al. The Nlrp3 Inflammasome Suppresses Colorectal Cancer Metastatic Growth in the Liver by Promoting Natural Killer Cell Tumoricidal Activity. *Immunity* **2015**, *43*, 751–763. [[CrossRef](#)]
222. Deng, Q.; Geng, Y.; Zhao, L.; Li, R.; Zhang, Z.; Li, K.; Liang, R.; Shao, X.; Huang, M.; Zuo, D.; et al. NLRP3 inflammasomes in macrophages drive colorectal cancer metastasis to the liver. *Cancer Lett.* **2019**, *442*, 21–30. [[CrossRef](#)]
223. Reeves, E.; James, E. Antigen processing and immune regulation in the response to tumours. *Immunology* **2017**, *150*, 16–24. [[CrossRef](#)]
224. Cervantes-Villagrana, R.D.; Albores-García, D.; Cervantes-Villagrana, A.R.; García-Acevez, S.J. Tumor-induced neurogenesis and immune evasion as targets of innovative anti-cancer therapies. *Signal Transduct. Target. Ther.* **2020**, *5*, 99. [[CrossRef](#)] [[PubMed](#)]
225. Chai, D.; Shan, H.; Wang, G.; Li, H.; Fang, L.; Song, J.; Zhang, Q.; Bai, J.; Zheng, J. AIM2 is a potential therapeutic target in human renal carcinoma and suppresses its invasion and metastasis via enhancing autophagy induction. *Exp. Cell Res.* **2018**, *370*, 561–570. [[CrossRef](#)] [[PubMed](#)]
226. Lasithiotaki, I.; Tsitoura, E.; Samara, K.D.; Trachalaki, A.; Charalambous, I.; Tzanakis, N.; Antoniou, K.M. NLRP3/Caspase-1 inflammasome activation is decreased in alveolar macrophages in patients with lung cancer. *PLoS ONE* **2018**, *13*, e0205242. [[CrossRef](#)]
227. Theivanthiran, B.; Evans, K.S.; DeVito, N.C.; Plebanek, M.; Sturdivant, M.; Wachsmuth, L.P.; Salama, A.K.S.; Kang, Y.; Hsu, D.; Balko, J.M.; et al. A tumor-intrinsic PD-L1/NLRP3 inflammasome signaling pathway drives resistance to anti-PD-1 immunotherapy. *J. Clin. Investig.* **2020**, *130*, 2570–2586. [[CrossRef](#)]
228. Chai, D.; Zhang, Z.; Shi, S.Y.; Qiu, D.; Zhang, C.; Wang, G.; Fang, L.; Li, H.; Tian, H.; Li, H.; et al. Absent in melanoma 2-mediating M1 macrophages facilitate tumor rejection in renal carcinoma. *Transl. Oncol.* **2021**, *14*, 101018. [[CrossRef](#)]
229. Nakamura, K.; Kassem, S.; Cleynen, A.; Chrétien, M.-L.; Guillerey, C.; Putz, E.M.; Bald, T.; Förster, I.; Vuckovic, S.; Hill, G.R.; et al. Dysregulated IL-18 Is a Key Driver of Immunosuppression and a Possible Therapeutic Target in the Multiple Myeloma Microenvironment. *Cancer Cell* **2018**, *33*, 634–648.e5. [[CrossRef](#)]
230. Ghiringhelli, F.; Apetoh, L.; Tesniere, A.; Aymeric, L.; Ma, Y.; Ortiz, C.; Vermaelen, K.; Panaretakis, T.; Mignot, G.; Ullrich, E.; et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1 β -dependent adaptive immunity against tumors. *Nat. Med.* **2009**, *15*, 1170–1178. [[CrossRef](#)] [[PubMed](#)]
231. Hu, B.; Ren, J.; Luo, Y.; Keith, B.; Young, R.M.; Scholler, J.; Zhao, Y.; June, C.H. Augmentation of Antitumor Immunity by Human and Mouse CAR T Cells Secreting IL-18. *Cell Rep.* **2017**, *20*, 3025–3033. [[CrossRef](#)]
232. Li, X.-Y.; Moesta, A.K.; Xiao, C.; Nakamura, K.; Casey, M.; Zhang, H.; Madore, J.; Lepletier, A.; Aguilera, A.R.; Sundarajan, A.; et al. Targeting CD39 in Cancer Reveals an Extracellular ATP- and Inflammasome-Driven Tumor Immunity. *Cancer Discov.* **2019**, *9*, 1754–1773. [[CrossRef](#)]
233. Li, Y.; Cao, F.; Li, M.; Li, P.; Yu, Y.; Xiang, L.; Xu, T.; Lei, J.; Tai, Y.Y.; Zhu, J.; et al. Hydroxychloroquine induced lung cancer suppression by enhancing chemo-sensitization and promoting the transition of M2-TAMs to M1-like macrophages. *J. Exp. Clin. Cancer Res.* **2018**, *37*, 259. [[CrossRef](#)]
234. Zhou, M.; He, X.; Mei, C.; Ou, C. Exosome derived from tumor-associated macrophages: Biogenesis, functions, and therapeutic implications in human cancers. *Biomark. Res.* **2023**, *11*, 100. [[CrossRef](#)]
235. Yan, W.; Jiang, S. Immune Cell-Derived Exosomes in the Cancer-Immunity Cycle. *Trends Cancer* **2020**, *6*, 506–517. [[CrossRef](#)]
236. Chuang, H.-Y.; Su, Y.-K.; Liu, H.-W.; Chen, C.-H.; Chiu, S.-C.; Cho, D.-Y.; Lin, S.-Z.; Chen, Y.-S.; Lin, C.-M. Preclinical Evidence of STAT3 Inhibitor Pacritinib Overcoming Temozolomide Resistance via Downregulating miR-21-Enriched Exosomes from M2 Glioblastoma-Associated Macrophages. *J. Clin. Med.* **2019**, *8*, 959. [[CrossRef](#)] [[PubMed](#)]
237. Seo, N.; Akiyoshi, K.; Shiku, H. Exosome-mediated regulation of tumor immunology. *Cancer Sci.* **2018**, *109*, 2998–3004. [[CrossRef](#)] [[PubMed](#)]

238. Han, C.; Zhang, C.; Wang, H.; Zhao, L. Exosome-mediated communication between tumor cells and tumor-associated macrophages: Implications for tumor microenvironment. *OncoImmunology* **2021**, *10*, 1887552. [\[CrossRef\]](#)
239. Hazrati, A.; Soudi, S.; Malekpour, K.; Mahmoudi, M.; Rahimi, A.; Hashemi, S.M.; Varma, R.S. Immune cells-derived exosomes function as a double-edged sword: Role in disease progression and their therapeutic applications. *Biomark. Res.* **2022**, *10*, 30. [\[CrossRef\]](#)
240. Kim, S.H.; Kim, G.; Han, D.H.; Lee, M.; Kim, I.; Kim, B.; Kim, K.H.; Song, Y.-M.; Yoo, J.E.; Wang, H.J.; et al. Ezetimibe ameliorates steatohepatitis via AMP activated protein kinase-TFEB-mediated activation of autophagy and NLRP3 inflammasome inhibition. *Autophagy* **2017**, *13*, 1767–1781. [\[CrossRef\]](#)
241. Bretz, N.P.; Ridinger, J.; Rupp, A.-K.; Rimbach, K.; Keller, S.; Rupp, C.; Marmé, F.; Umansky, L.; Umansky, V.; Eigenbrod, T.; et al. Body Fluid Exosomes Promote Secretion of Inflammatory Cytokines in Monocytic Cells via Toll-like Receptor Signaling. *J. Biol. Chem.* **2013**, *288*, 36691–36702. [\[CrossRef\]](#)
242. Atay, S.; Gercel-Taylor, C.; Taylor, D.D. Human Trophoblast-Derived Exosomal Fibronectin Induces Pro-Inflammatory IL-1 β Production by Macrophages. *Am. J. Reprod. Immunol.* **2011**, *66*, 259–269. [\[CrossRef\]](#)
243. Bardi, G.T.; Smith, M.A.; Hood, J.L. Melanoma exosomes promote mixed M1 and M2 macrophage polarization. *Cytokine* **2018**, *105*, 63–72. [\[CrossRef\]](#)
244. Li, X.; Lei, Y.; Wu, M.; Li, N. Regulation of Macrophage Activation and Polarization by HCC-Derived Exosomal lncRNA TUC339. *Int. J. Mol. Sci.* **2018**, *19*, 2958. [\[CrossRef\]](#) [\[PubMed\]](#)
245. Chen, P.; Huang, Y.; Bong, R.; Ding, Y.; Song, N.; Wang, X.; Song, X.; Luo, Y. Tumor-Associated Macrophages Promote Angiogenesis and Melanoma Growth via Adrenomedullin in a Paracrine and Autocrine Manner. *Clin. Cancer Res.* **2011**, *17*, 7230–7239. [\[CrossRef\]](#)
246. Oh, K.; Lee, O.-Y.; Park, Y.; Seo, M.W.; Lee, D.-S. IL-1 β induces IL-6 production and increases invasiveness and estrogen-independent growth in a TG2-dependent manner in human breast cancer cells. *BMC Cancer* **2016**, *16*, 724. [\[CrossRef\]](#)
247. Wu, T.; Hong, Y.; Jia, L.; Wu, J.; Xia, J.; Wang, J.; Hu, Q.; Cheng, B. Modulation of IL-1 β reprogrammes the tumor microenvironment to interrupt oral carcinogenesis. *Sci. Rep.* **2016**, *6*, 20208. [\[CrossRef\]](#)
248. Dmitrieva-Posocco, O.; Dzutsev, A.; Posocco, D.F.; Hou, V.; Yuan, W.; Thovarai, V.; Mufazalov, I.A.; Gunzer, M.; Shilovskiy, I.P.; Khaitov, M.R.; et al. Cell-Type-Specific Responses to Interleukin-1 Control Microbial Invasion and Tumor-Elicited Inflammation in Colorectal Cancer. *Immunity* **2019**, *50*, 166–180.e7. [\[CrossRef\]](#)
249. Hu, P.; Yan, T.; Lv, S.; Ye, M.; Wu, M.; Fang, H.; Xiao, B. Exosomal HMGB3 released by glioma cells confers the activation of NLRP3 inflammasome and pyroptosis in tumor-associated macrophages. *Tissue Cell* **2024**, *88*, 102406. [\[CrossRef\]](#)
250. Liang, M.; Chen, X.; Wang, L.; Qin, L.; Wang, H.; Sun, Z.; Zhao, W.; Geng, B. Cancer-derived exosomal TRIM59 regulates macrophage NLRP3 inflammasome activation to promote lung cancer progression. *J. Exp. Clin. Cancer Res.* **2020**, *39*, 176. [\[CrossRef\]](#)
251. Shang, S.; Ji, X.; Zhang, L.; Chen, J.; Li, C.; Shi, R.; Xiang, W.; Kang, X.; Zhang, D.; Yang, F.; et al. Macrophage ABHD5 suppresses NF κ B-dependent matrix metalloproteinase expression and cancer metastasis. *Cancer Res.* **2019**, *79*, 5513–5526. [\[CrossRef\]](#)
252. Rao, X.; Zhou, X.; Wang, G.; Jie, X.; Xing, B.; Xu, Y.; Chen, Y.; Li, J.; Zhu, K.; Wu, Z.; et al. NLRP6 is required for cancer-derived exosome-modified macrophage M2 polarization and promotes metastasis in small cell lung cancer. *Cell Death Dis.* **2022**, *13*, 891. [\[CrossRef\]](#)
253. Gutzeit, C.; Nagy, N.; Gentile, M.; Lyberg, K.; Gumz, J.; Vallhov, H.; Puga, I.; Klein, E.; Gabrielsson, S.; Cerutti, A.; et al. Exosomes derived from Burkitt's lymphoma cell lines induce proliferation, differentiation, and class-switch recombination in B cells. *J. Immunol.* **2014**, *192*, 5852–5862. [\[CrossRef\]](#)
254. Zhang, Z.; Yu, X.; Zhou, Z.; Li, B.; Peng, J.; Wu, X.; Luo, X.; Yang, L. LMP1-positive extracellular vesicles promote radioresistance in nasopharyngeal carcinoma cells through P38 MAPK signaling. *Cancer Med.* **2019**, *8*, 6082–6094. [\[CrossRef\]](#) [\[PubMed\]](#)
255. Dargani, Z.T.; Singla, D.K. Embryonic stem cell-derived exosomes inhibit doxorubicin-induced TLR4-NLRP3-mediated cell death-pyroptosis. *Am. J. Physiol. Circ. Physiol.* **2019**, *317*, H460–H471. [\[CrossRef\]](#)
256. Jeppesen, D.K.; Fenix, A.M.; Franklin, J.L.; Higginbotham, J.N.; Zhang, Q.; Zimmerman, L.J.; Liebler, D.C.; Ping, J.; Liu, Q.; Evans, R.; et al. Reassessment of Exosome Composition. *Cell* **2019**, *177*, 428–445.e18. [\[CrossRef\]](#) [\[PubMed\]](#)
257. Lee, H.; Groot, M.; Pinilla-Vera, M.; Fredenburgh, L.E.; Jin, Y. Identification of miRNA-rich vesicles in bronchoalveolar lavage fluid: Insights into the function and heterogeneity of extracellular vesicles. *J. Control. Release* **2019**, *294*, 43–52. [\[CrossRef\]](#) [\[PubMed\]](#)
258. Showalter, M.R.; Wancewicz, B.; Fiehn, O.; Archard, J.A.; Clayton, S.; Wagner, J.; Deng, P.; Halmai, J.; Fink, K.D.; Bauer, G.; et al. Primed mesenchymal stem cells package exosomes with metabolites associated with immunomodulation. *Biochem. Biophys. Res. Commun.* **2019**, *512*, 729–735. [\[CrossRef\]](#)
259. Cha, D.J.; Franklin, J.L.; Dou, Y.; Liu, Q.; Higginbotham, J.N.; Beckler, M.D.; Weaver, A.M.; Vickers, K.; Prasad, N.; Levy, S.; et al. KRAS-dependent sorting of miRNA to exosomes. *eLife* **2015**, *4*, e07197. [\[CrossRef\]](#) [\[PubMed\]](#)

260. Zhang, Y.; Liu, Y.; Liu, H.; Tang, W.H. Exosomes: Biogenesis, biologic function and clinical potential. *Cell Biosci.* **2019**, *9*, 19. [\[CrossRef\]](#)
261. Hurwitz, S.N.; Nkosi, D.; Conlon, M.M.; York, S.B.; Liu, X.; Tremblay, D.C.; Meckes, D.G., Jr. CD63 Regulates Epstein-Barr Virus LMP1 Exosomal Packaging, Enhancement of Vesicle Production, and Noncanonical NF- κ B Signaling. *J. Virol.* **2017**, *91*, 10–1128. [\[CrossRef\]](#)
262. Wozniak, A.L.; Adams, A.; King, K.E.; Dunn, W.; Christenson, L.K.; Hung, W.-T.; Weinman, S.A. The RNA binding protein FMR1 controls selective exosomal miRNA cargo loading during inflammation. *J. Cell Biol.* **2020**, *219*, e201912074. [\[CrossRef\]](#)
263. Savina, A.; Furlán, M.; Vidal, M.; Colombo, M.I. Exosome release is regulated by a calcium-dependent mechanism in K562 cells. *J. Biol. Chem.* **2003**, *278*, 20083–20090. [\[CrossRef\]](#)
264. King, H.W.; Michael, M.Z.; Gleadle, J.M. Hypoxic enhancement of exosome release by breast cancer cells. *BMC Cancer* **2012**, *12*, 421. [\[CrossRef\]](#) [\[PubMed\]](#)
265. Szul, T.; Bratcher, P.E.; Fraser, K.B.; Kong, M.; Tirouvanziam, R.; Ingersoll, S.; Sztul, E.; Rangarajan, S.; Blalock, J.E.; Xu, X.; et al. Toll-Like Receptor 4 Engagement Mediates Prolyl Endopeptidase Release from Airway Epithelia via Exosomes. *Am. J. Respir. Cell Mol. Biol.* **2016**, *54*, 359–369. [\[CrossRef\]](#)
266. Parolini, I.; Federici, C.; Raggi, C.; Lugini, L.; Palleschi, S.; De Mito, A.; Coscia, C.; Iessi, E.; Logozzi, M.; Molinari, A.; et al. Microenvironmental pH Is a Key Factor for Exosome Traffic in Tumor Cells. *J. Biol. Chem.* **2009**, *284*, 34211–34222. [\[CrossRef\]](#) [\[PubMed\]](#)
267. Välimäki, E.; Cypriak, W.; Virkanen, J.; Nurmi, K.; Turunen, P.M.; Eklund, K.K.; Åkerman, K.E.; Nyman, T.A.; Matikainen, S. Calpain Activity Is Essential for ATP-Driven Unconventional Vesicle-Mediated Protein Secretion and Inflammasome Activation in Human Macrophages. *J. Immunol.* **2016**, *197*, 3315–3325. [\[CrossRef\]](#)
268. Tzng, E.; Bayardo, N.; Yang, P.C. Current challenges surrounding exosome treatments. *Extracell. Vesicle* **2023**, *2*, 100023. [\[CrossRef\]](#)
269. Ju, M.; Bi, J.; Wei, Q.; Jiang, L.; Guan, Q.; Zhang, M.; Song, X.; Chen, T.; Fan, J.; Li, X.; et al. Pan-cancer analysis of NLRP3 inflammasome with potential implications in prognosis and immunotherapy in human cancer. *Briefings Bioinform.* **2020**, *22*, bbaa345. [\[CrossRef\]](#)
270. Zheng, T.; Wang, X.; Yue, P.; Han, T.; Hu, Y.; Wang, B.; Zhao, B.; Zhang, X.; Yan, X. Prognostic Inflammasome-Related Signature Construction in Kidney Renal Clear Cell Carcinoma Based on a Pan-Cancer Landscape. *Evidence-Based Complement. Altern. Med.* **2020**, *2020*, 3259795. [\[CrossRef\]](#)

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.