

SCIENTIFIC REPORTS



OPEN

Rosa26-targeted sheep gene knock-in via CRISPR-Cas9 system

Mingming Wu¹, Caihong Wei², Zhengxing Lian¹, Ruizao Liu², Caiye Zhu², Huihua Wang², Jiaxue Cao², Yuelei Shen³, Fuping Zhao², Li Zhang², Zhu Mu⁴, Yayu Wang⁴, Xiaogang Wang⁴, Lixin Du² & Chudian Wang¹

Received: 28 September 2015

Accepted: 29 March 2016

Published: 11 April 2016

Recent advances in our ability to design DNA binding factors with specificity for desired sequences have resulted in a revolution in genetic engineering, enabling directed changes to the genome to be made relatively easily. Technologies that facilitate specific and precise genome editing, such as knock-in, are critical for determining the functions of genes and for understanding fundamental biological processes. The CRISPR/Cas9 system has recently emerged as a powerful tool for functional genomic studies in mammals. *Rosa26* gene can encode a non-essential nuclear RNA in almost all organizations, and become a hot point of exogenous gene insertion. Here, we describe efficient, precise CRISPR/Cas9-mediated Integration using a donor vector with tGFP sequence targeted in the sheep genomic *Rosa26* locus. We succeeded in integrating with high efficiency an exogenous tGFP (turboGFP) gene into targeted genes in frame. Due to its simplicity, design flexibility, and high efficiency, we propose that CRISPR/Cas9-mediated knock-in will become a standard method for the generation transgenic sheep.

Sheep is an important economic animals and production trait has become the main direction of sheep breeding. Transgenic animals with reporter gene expression in specific tissues or cell types are valuable experimental tools. Recently, programmable endonucleases, such as Zinc Finger Nucleases (ZFNs), Transcription activator-like effector nucleases (TALENs), and especially the CRISPR, the recently developed clustered regularly interspaced short palindromic repeats (CRISPR)/CRISPR-associated (Cas) 9 system, were shown to be a kind of revolutionary technologies for targeted genome editing¹. CRISPR loci contain an array of short palindromic repeats separated by proto spacer DNA snippets of about 20 bp that have been acquired from such invading agents. Compared to ZFNs and TALENs, CRISPR/Cas9-mediated genome engineering is easy to handle, highly specific, efficient, and multiplexable². Compared with protein-based genome editing tools with customizable DNA binding specificities, such as ZFNs and TALENs, the newer CRISPR/Cas9 platform is based on simple base-pairing between an engineered RNA and the targeted genomic site, which enables rapid design, ease of use, and low costs³. The Cas9 endonuclease is ushered to the specific site of interest by a single guide RNA (sgRNA), an engineered fusion molecule of the targeting CRISPR RNA (crRNA) and the trans-activating crRNA, to generate double-stranded DNA breaks (DSDBs) at the target site⁴. The CRISPR/Cas9 system has been demonstrated to be easy to handle, highly specific, efficient, and multiplexable, making it a suitable approach to the engineering of eukaryotic genomes⁵. This system has been successfully used to target genomic loci in several species, including mice, rats, zebrafish, and pigs^{6–10}.

However, so far, there have been no reports of the use of CRISPR/Cas9-mediated knock-in DNA cassettes in the sheep genome. *ROSA26* is a safe area, exogenous genes that decide a dot inside this site will not affect the expression of other genes. *Rosa26* is ubiquitously expressed in embryonic and adult tissues¹¹. *Rosa26* was first identified and targeted in mouse embryonic stem cells (ESCs) in the 1990s and then in human ESCs in 2007¹⁰. Efficient integration of transgenes at preselected chromosomal locations was achieved in mammalian cells by recombinase-mediated-cassette-exchange (RMCE), a novel procedure that makes use of the CRE recombinase together with *Lox* sites bearing different spacer regions. Here, the sheep *Rosa26* (*sRosa26*) locus was characterized and its locus tagged with a knock-in reporter gene using a CRISPR/Cas9-mediated method. Using this approach, transgenic sheep stably over-expressing a gene of interest were also created through RMCE¹².

The most preferred integration site used for transgene insertion is the *Rosa26* locus in mice. As in mice, the human and rat *Rosa26* loci were identified and successfully targeted using traditional HR¹³. There have reported

¹College of animal science and technology, China Agricultural University, Beijing 100193, China. ²Institute of Animal Sciences, Chinese Academy of Agricultural Sciences, Beijing 100193, China. ³Beijing Biocytogen Co., Ltd, Beijing 100176, China. ⁴College of life science and technology, Jinan university, Guangzhou 510632, China. Correspondence and requests for materials should be addressed to X.W. (email: 421345163@qq.com) or L.D. (email: lxd@263.net) or C.W. (email: wangchudian@qq.com)

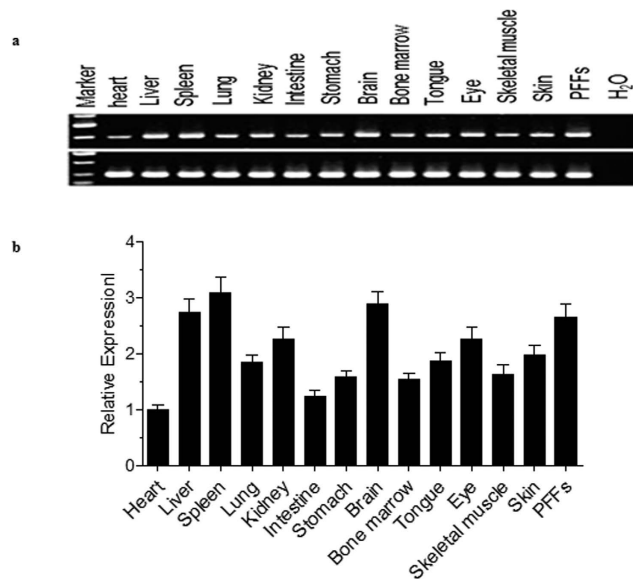


Figure 1. Characterization of *sRosa26* and highly efficient gene knock-in and replacement at the *sRosa26* locus. (a,b) *sRosa26* was expressed in a variety of organ tissues as determined by (a) RT-PCR and (b) quantitative RT-PCR. For RT-PCR, the designed primers annealed in *sRosa26* sequence and amplified a correctly spliced product of 485 bp. Ovis *GAPDH* served as a control (234 bp). For qPCR, primers were specific to the *sRosa26* sequence. The PCR product of the Ovis *ACTB* gene served as the reference control. Data are presented as the average expression levels of three individual RT/qPCR reactions.

that knock-in of a long DNA fragment via homology-independent DNA repair can be achieved in zebrafish using the CRISPR/Cas9 system. In this method, co-injection of a donor plasmid, short guide RNAs (sgRNAs) and Cas9 mRNA leads to concurrent digestion of the genomic DNA and the donor plasmid, resulting in the incorporation of the donor plasmid into the genome^{14,15}. In some cases, fluorescent proteins such as GFP have been used for the characterisation and quantification of homologous recombination events as well as for endogenous gene-tagging and determination of subcellular localisation of expressed proteins¹⁶.

Here, we have modified this method, and succeeded in generating knock-in transgenic sheep with reporter gene (tGFP) that mimics endogenous gene expression for the same targeted loci. The method is simple, and flexible in design. Furthermore, the efficiency of obtaining transgenic founders is 12.5%. We propose that CRISPR/Cas9 mediated knock-in will become a standard method for the generation of transgenic sheep lines.

Results

Confirmation of *Rosa26* locus. The ensemble ovis database was searched using the mouse *Rosa26* sequence as a template and a highly conserved region was located on the sheep chromosome 19 (Supplementary Table S1). Sequence alignments of porcine, murine, rat, and human *Rosa26* sequences showed high sequence conservation (>70%) among these species (Supplementary Fig. S1). According to the homologous sequence between them. In mice, rats, and humans, the *Rosa26* locus encodes non-coding RNAs that are ubiquitously expressed. RT-PCR and SYBR-green-based quantitative PCR assays demonstrated that this non-coding RNA was expressed in a wide variety of adult sheep tissues (Fig. 1a,b).

Designation of the sgRNA for *sRosa26* locus. Here, 5 guide RNAs (sgRNAs) were designed to target the sequence of the *sRosa26* locus (Fig. 2 and Table 1). The activity of the CRISPR/Cas9 was assessed using a UCA kit. The results of genotyping with the UCA kit showed that all 5 sgRNAs efficiently guided Cas9 for genome editing (Fig. 2a). This led to non-homologous end-joining mediated insertions and deletions (indels) in the *sRosa26* locus. The sgRNA with the highest activity was used to target the *sRosa26* locus.

Preparation of the CRISPR/Cas plasmids for genome engineering. The targeting vector contains a 1.0 Kb left arm and a 1.0 Kb right arm for homologous recombination (Fig. 3a). It also contains a CAG promoter (2753 bp, Supplementary Fig. S2). To validate the targeting efficiency of these sgRNAs, expression vectors were constructed in which a sgRNA was driven by the CAG promoter. An expression cassette comprising a viral splice acceptor (SA), a CAG promoter gene, and an *tGFP* gene was inserted between the homologous arms (Fig. 3a). The tGFP (928 bp, Supplementary Table S2) expression vector was designed to determine whether the exogenous gene can be expressed at the *sRosa26* locus. The CAG promoter-driven tGFP vector was transfected into sheep fibroblasts. At 48 h post-transfection, a high level of tGFP expression was detected in sheep fibroblasts (Fig. 3b).

Cas9/sgRNA-Mediated Genome Targeting Efficiently Transmits into Ovaries of Sheep. The germline transmission of genetic mutations is the necessary condition to establish genetically modified animals¹⁷. *In vitro* synthesized Cas9 mRNA (40 ng/μl), sgRNA (20 ng/μl), and expression vector (20 ng/μl) were injected into

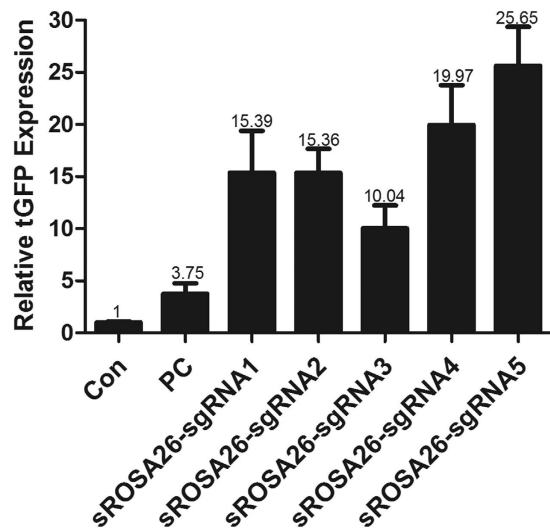


Figure 2. (a) Analysis of sgRNA activity. 5 guide RNAs (sgRNAs) were designed to target the sequence of the *sRosa26* locus. All 5 sgRNAs efficiently guided Cas9 for genome editing.

sgRNA		Sequence (5'-3')	Target sequence length
sROSA26-sgRNA1	target sequence	ggaggcgatgacgagatcgc ggg	20 + PAM
	sROSA26-sgRNA1-Up	cacc ggaggcgatgacgagatcgc	
	sROSA26-sgRNA1-Dn	aaac gcgatctctcatcgctcc	
sROSA26-sgRNA2	target sequence	ggggaggaggattcttct agg	20 + PAM
	sROSA26-sgRNA2-Up	cacc ggggaggaggattcttct	
	sROSA26-sgRNA2-Dn	aaac agaagaatcctcctcccc	
sROSA26-sgRNA3	target sequence	gacgagatcgcgggggagg agg	20 + PAM
	sROSA26-sgRNA3-Up	cacc ggacgagatcgcgggggagg	
	sROSA26-sgRNA3-Dn	aaac cctcccccgatctctcc	
sROSA26-sgRNA4	target sequence	gaggcgatgacgagatcgcg ggg	20 + PAM
	sROSA26-sgRNA4-Up	cacc ggaggcgatgacgagatcgcg	
	sROSA26-sgRNA4-Dn	aaac cgcatctctcatcgctcc	
sROSA26-sgRNA5	target sequence	gtcagtgctctctcgatta tgg	20 + PAM
	sROSA26-sgRNA5-Up	cacc ggtcagtgctctctcgatta	
	sROSA26-sgRNA5-Dn	aaac taatcggaggagactcgacc	

Table 1. 5 different sgRNA designs. 5 guide RNAs (sgRNAs) were designed to target the sequence of the *sRosa26* locus. The activity of the CRISPR/Cas9 was assessed using a UCA kit. All 5 sgRNAs efficiently guided Cas9 for genome editing.

the single-cell sheep embryos. Out of the 35 injected embryos, 30 healthy embryos were transferred into surrogate mothers and 8 pups were born (Fig. 4 and Table 2). Out of these 8 pups, 1 had the tGFP gene at the *sRosa26* locus and it was expressed in lamb ear tissues as indicated by PCR, Southern blot, RT-PCR and Western blot analysis (Fig. 4). And the *sRosa26* locus sequence were detected by PCR and sequencing (Supplementary Fig. S3). The PCR and sequencing results showed that the tGFP gene locus was the *sRosa26* target locus. These results suggest that the CRISPR/Cas9 system can induce mutations and knock-in DNA fragments in the *sRosa26* locus very efficiently.

Discussion

Instead of knocking-in DNA constructs in the exon of a gene of interest, we knocked in constructs in the upstream region of a gene with the CAG promoter construct (Fig. 3a). There are advantages and a potential disadvantage in this approach. An advantage is that expression levels of transgenes are increased¹⁸. In our experience generating transgenic sheep, the usage of the CAG promoter instead of promoters of endogenous gene promoters increased the expression level of the transgene for many genes including tGFP. Another advantage is that a relatively large genomic region can be a subject of integration. This allowed that many sgRNAs can be tested by the usage of the CAG promoter. Transmission of the precise genome modification to the germline is a key to establishing

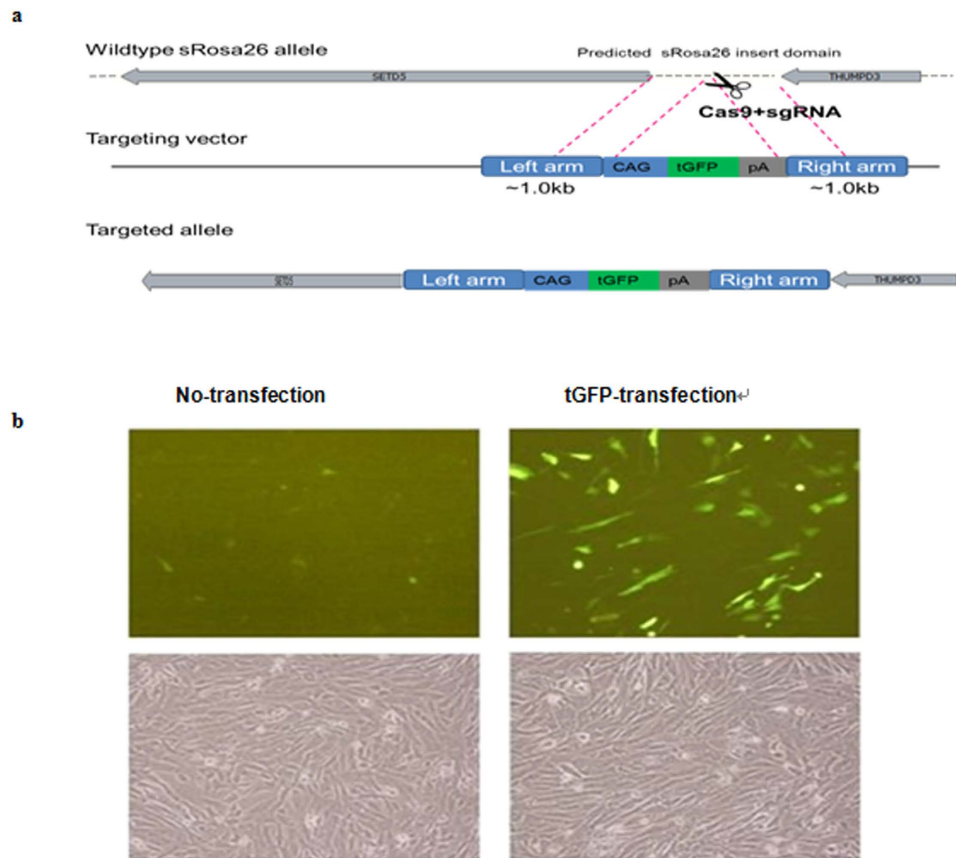


Figure 3. (a) Diagram of Cas9-mediated knock-in of LA-CAG-tGFP-polyA-RA into the *sRosa26* locus. LA, left arm (1.0 kb); RA, right arm (1.0 kb). (b) CAG promoter-driven tGFP expression in sheep fibroblasts (CAG promoter region is shown in supplementary information).

a targeted gene knock-in line. In mice, rats, and humans, the *Rosa26* locus encodes non-coding RNAs that are ubiquitously expressed¹³. So 5 guide RNAs (sgRNAs) were designed to target the sequence of the *sRosa26* locus.

In this study, we showed that the precise integration of exogenous DNA into the targeted genomic locus in sheep can be efficiently achieved using a donor vector containing short homology arms. We improved this method by introducing homology arms into a donor vector. In comparison to ZFNs and TALENs, the CRISPR/Cas9 system is suitable for this method because of the ease of donor-vector construction and of the multiple gRNA design^{18,19}. A conventional knock-in vector carries more than 800 bp of homology arms without cleavable sites^{20,21}. And the targeting vector contains a 1.0 Kb left arm and a 1.0 Kb right arm for homologous recombination.

The CRISPR/Cas9 system has been used to introduce defined genetic modification in cultured cell system^{22,23}, but whether this could be applied to *in vivo* system and what parameter(s) is influential for its efficiency was less explored²⁴. This is the first study to use oligonucleotides as templates to successfully knock DNA cassettes into the sheep genome via the CRISPR/Cas9 system. This is also the first recorded use of CRISPR/Cas9-mediated knock in of DNA sheep into the *sRosa26* locus as part of a research exercise.

Collectively, these data strongly suggest that the *sRosa26* locus is an ideal site for expression of exogenous genes. But the using of Cas9-mediated knock-ins has not yet been reported in sheep. Here, this simple and efficient CRISPR/Cas9 system was successfully extended to the modification of the sheep genome at the *sRosa26* locus.

Unlike other ways of editing genes, which require construction of double-stranded DNA templates, this CRISPR/Cas9 system allows rapid and seamless editing of the genome at precise locations. It may become a powerful tool for assessing the functions of genes, altering critical residues in proteins to create desirable gain-of-function or loss-of-function mutations, and generating mutations in highly conserved proteins in sheep to facilitate the study of corresponding human diseases and agricultural production. All the results given above indicate that the *sRosa26* locus is an excellent site for ubiquitous expression of exogenous genes.

Taken together, these results indicate that transgenic sheep stably overexpressing a gene of interest could be generated through CRISPR/Cas9 system at the *sRosa26* locus without any need for more sophisticated technology.

Methods

Ethics statement. All animals were housed and handled according to China Agricultural University Institutional Animal Care and Use Committee guidelines and all animal work was approved by the appropriate committee (IACUC 0000000 and 0000000A-1). All experiments were performed according to institutional guidelines.

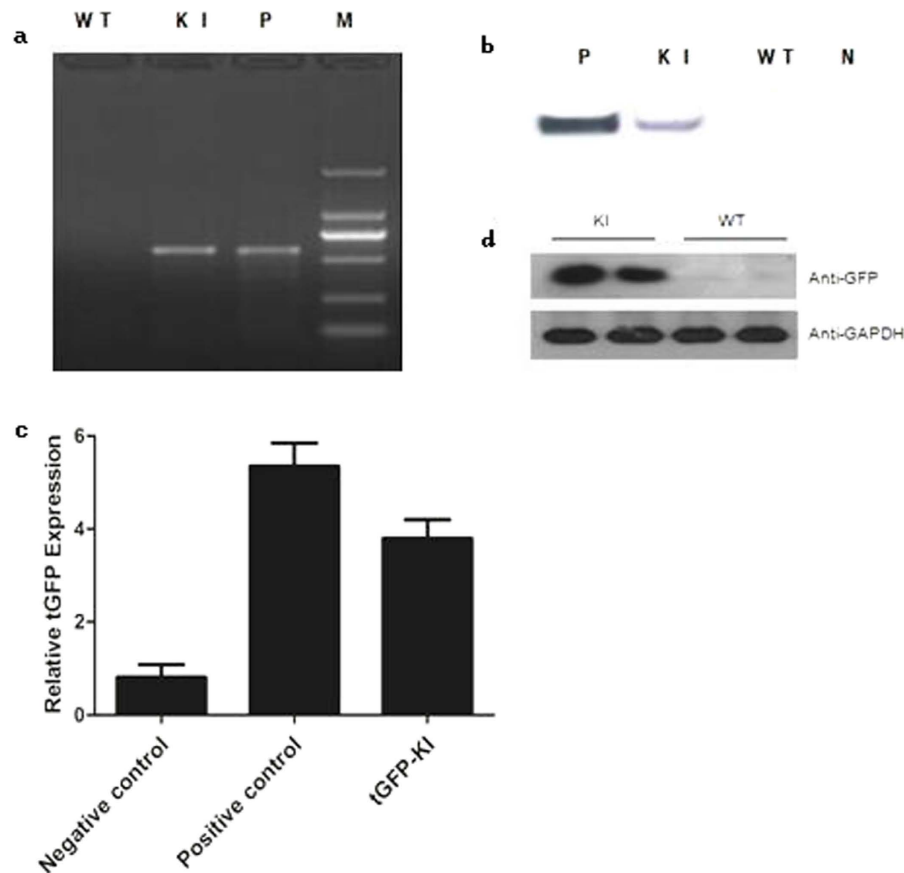


Figure 4. (a) PCR analysis of tGFP in tissues knock-in lambs. M = marker, N = negative control, P = positive control. The size of target band is about 650 bp. (b) Southern blot analysis of tGFP-KI mutant sheep. Both the negative control and tGFP-KI sheep can be detected the target sequence. (c) RT-PCR analysis of tGFP expression in tissues knock-in lambs. All the lambs were 1 month old. tGFP gene expression quantity is 4.5 times that of the negative control. (d) Western blot analysis of tGFP-KI mutant sheep. Total protein from tGFP-KI sheep was subjected to SDS-PAGE on a 12% acrylamide gel. GAPDH was used as a loading control. The tGFP gene expression can't be detected in controls.

Gene	Number of embryos injected	Number of embryos transferred	Number of new borns	Number of mutations
sRosa26	35	30	8	1
Ration	/	85.7%	26.7%	12.5%

Table 2. Numbers of injected and transferred embryos, newborns, and mutants generated during the establishment of knock-in sheep using gRNA/Cas9 system. Cas9 mRNA and sgRNA were mixed and injected into sheep zygotes. The injected eggs were transferred into pseudopregnant females. The mutations were identified by sequencing PCR amplified 0.5 kbp genomic fragment containing target in the center. GMO; gene modified organism.

Animals. All animals were handled according to the Guidelines for the Care and Use of Laboratory Animals established by the Beijing Association for Laboratory Animal Science. Animal experiments were approved by the Animal Ethics Committee of College of Biological Sciences of China Agricultural University.

Cell culture. A sheep fetal fibroblast (SFB) cell line (China Agricultural University) was cultured in the medium containing 10% fetal bovine (Gibco, USA) serum in 5% CO₂ at 37 °C. The cells were seeded in 6-well plates (Thermo Scientific, USA). After 24 h, cells were co-transfected with a mixture of plasmid, pCDNA3.1(+)-Cas9, pMD-19T-U6-sgRNA, and tGFP expression vector with the mass ratio of 2:1:1 (2500 ng in total per well), following the instruction of Lipofectamine3000 (Life Technologies, USA). Cells were harvested 48 h after transfection.

Preparation of Cas9 mRNA and sgRNA. Cas9 and sgRNA coding regions containing the T7 promoter were PCR-amplified using DNA polymerase from each plasmid constructed above. The T7-Cas9 PCR products were gel purified and used as the template for *in vitro* transcription (IVT) using an mMESSEGE mMACHINE T7 ULTRA Transcription Kit (Life Technologies, USA). The T7-sgRNA PCR product was gel-purified and used as the template for IVT using MEGA shortscript T7 Kit (Life Technologies). The sgRNA was purified by ethanol precipitation, and all products were re-dissolved in RNase-free water and stored at -80°C . 5 guide RNAs (sgRNAs) were designed to target the sequence of the *sRosa26* locus (Fig. 2 and Table 1). The activity of the CRISPR/Cas9 was assessed using a UCA kit (Beijing Biocytogen Co., Ltd, China).

Microinjection of zygotes and embryo transfer. The sheep zygotes were obtained by super-ovulation of females and artificial insemination. The zygotes were flushed from the oviduct using sterile filtered embryo flushing solution. After that, 2–5 μl TE solution containing 20 ng/ μl of sgRNA, 40 ng/ μl of Cas9 mRNA and expression vector (20 ng/ μl) were injected into the cytoplasm of pronucleus embryos using injection needles. Injections were performed using an Eppendorf TransferMan NK2 micromanipulator.

Detection and Analysis of Indel Mutation in Lambs. Ear and muscle in hind leg were collected and digested in lysis buffer (10 μM Tris-HCl, 0.4 M EDTA, 1% SDS, and 100 $\mu\text{g}/\text{ml}$ Proteinase K). And incubated at 50°C for 1 hr, followed by extraction in 400 μl of phenolchloroform. The mixture was then centrifuged at $21,000 \times g$ for 20 min at 4°C . The supernatant was transferred to a new tube. An equal volume of isopropanol was added, and the tube was vortexed thoroughly. The mixture was then kept at -20°C for at least 1 hr, followed by centrifugation at $21,000 \times g$ for 20 min at 4°C . The supernatant was removed, and the DNA pellet was washed with 500 μl of 75% ethanol, followed by centrifugation at $21,000 \times g$ for 5 min at 4°C . Finally, the pellet was dried for 10 min and resuspended in 30 μl of DNase-free water. For each samples, at least 100 clones were picked up randomly and sequenced.

Southern Blot detection. To confirm transgene insertion in the sheep genome, Southern Blot was performed using DIG High Prime DNA Labeling and Detection Starter Kit II (Roche, 11585614910). DNA was isolate from the transgenic lambs and a wild type hu sheep tissue, Plasmid *sRosa26*-Gibson-HR1 was used as a positive control. The probe hybridizes to a 0.92 kb fragment depicted in Fig. 4b, indicating site-specific gene insertion.

Western blotting analysis. For Western blot analysis, total proteins were isolated from the samples by homogenization in lysis buffer (50 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1% Triton X-100, 0.25% sodium deoxycholate, and complete protease inhibitor cocktail (Roche)). The concentration of proteins was measured by Bradford reagent (Sigma), separated on 10% SDS-PAGE gels and transferred to Immobilon-P membranes (Millipore). After blocking in 5% low-fat milk in PBST (0.1% Tween 20 in PBS) for 1 h, the membranes were incubated with tGFP antibody (1:500, SantaCruz Biotechnology), Fst antibody (1:500, Santa Cruz Biotechnology) or mouse GAPDH antibody (1:2000, Santa Cruz Biotechnology) overnight at 4°C . After washing in PBST, the membranes were incubated in goat anti-rabbit antibody conjugated with horseradish peroxidase (1:5000) for 1 h, followed by three washes in PBST. The signals were detected by ECL Chemiluminescent kit (Amersham Pharmacia Biotech, Arlington Heights).

Off-target Analyses. Potential targets of tGFP gene and CAG promoter were detected using PCR amplifications with the program: 95°C for 5 min; 95°C for 1 min; 58°C for 1 min, 72°C for 1 min, 30 cycles, 72°C for 10 min, store at 4°C and the expression of tGFP gene were detected using RT-PCR, The target *Rosa26* sequence were detected using PCR and Sanger sequencing (Invitrogen).

References

- Kim, H. & Kim, J. S. A guide to genome engineering with programmable nucleases. *Nat. Rev. Genet.* **15**, 321–334 (2014).
- Mali, P., Esvelt, K. M. & Church, G. M. Cas9 as a versatile tool for engineering biology. *Nat. Methods* **10**, 957–963 (2013).
- Pennisi, E. The CRISPR craze. *Science* **341**, 833–836 (2013).
- Zhao, P. *et al.* Oligonucleotide-based targeted gene editing in *C. elegans* via the CRISPR/Cas9 system. *Cell Res* **24**, 247–250 (2014).
- Mali, P., Esvelt, K. M. & Church, G. M. Cas9 as a versatile tool for engineering biology. *Nat. Methods* **10**, 957–963 (2013).
- Li, D. *et al.* Heritable gene targeting in the mouse and rat using a CRISPR-Cas system. *Nat. Biotechnol.* **31**, 681–683 (2013).
- Ma, Y. *et al.* Generating rats with conditional alleles using CRISPR/Cas9. *Cell Res.* **24**, 122–125 (2014).
- Shen, B. *et al.* Generation of gene-modified mice via Cas9/RNA-mediated gene targeting. *Cell Res.* **23**, 720–723 (2013).
- Hwang, W. Y. *et al.* Efficient genome editing in zebrafish using a CRISPR-Cas system. *Nat. Biotechnol.* **31**, 227–229 (2013).
- Li, X. P. *et al.* *Rosa26*-targeted swine models for stable gene over-expression and Cre-mediated lineage tracing. *Cell Res.* **24**, 501–504 (2014).
- Friedrich, G. & Soriano, P. Promoter traps in embryonic stem cells: a genetic screen to identify and mutate developmental genes in mice. *Genes Dev.* **5**, 1513–1523 (1991).
- Bouhassira, E. E., Westerman, K. & LeBoulch, P. Transcriptional behavior of LCR enhancer elements integrated at the same chromosomal locus by recombinase-mediated cassette exchange. *Blood* **90**, 3332–3344 (1997).
- Kong, Q. R. *et al.* *Rosa26* locus supports tissue-specific promoter driving transgene expression specifically in pig. *PLoS One.* **9**, e107945 (2014).
- Auer, T. O. & Del Bene, F. CRISPR/Cas9 and TALEN-mediated knock-in approaches in zebrafish. *Methods* **69**, 142–150 (2014).
- Auer, T. O., Duroure, K., De Cian, A., Concorde, J. P. & Del Bene, F. Highly efficient CRISPR/Cas9-mediated knock-in in zebrafish by homology-independent DNA repair. *Genome Res* **24**, 142–153 (2014).
- Li, K., Wang, G., Andersen, T., Zhou, P. & Pu, W. T. Optimization of genome engineering approaches with the CRISPR/Cas9 system. *PLoS One* **9**, e105779, doi: 10.1371/journal.pone.0105779 (2014).
- Spitz, F. & Furlong, E. E. Transcription factors: from enhancer binding to developmental control. *Nat. Rev. Genet.* **13**, 613–626 (2012).
- Ota, S., Hisano, Y., Ikawa, Y. & Kawahara, A. Multiple genome modifications by the CRISPR/Cas9 system in zebrafish. *Genes Cells* **19**, 555–564 (2014).

19. Wang, H. *et al.* One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell* **153**, 910–918 (2013).
20. Zu, Y. *et al.* TALEN-mediated precise genome modification by homologous recombination in zebrafish. *Nat Methods* **10**, 329–331 (2013).
21. Shin, J., Chen, J. & Solnica-Krezel, L. Efficient homologous recombination-mediated genome engineering in zebrafish using TALE nucleases. *Development* (2014).
22. Ran, F. A. *et al.* Double Nicking by RNA-Guided CRISPR Cas9 for Enhanced Genome Editing Specificity. *Cell* **154**, 1380–9 (2013).
23. Wang, H. *et al.* One-Step Generation of Mice Carrying Mutations in Multiple Genes by CRISPR/Cas-Mediated Genome Engineering. *Cell* **153**, 910–918 (2013).
24. Masafumi, I., Mami, M. & Maki, I. Rapid generation of mouse models with defined point mutations by the CRISPR/Cas9 system. *Sci. Rep.* doi: 10.1038/srep 05396 (2014).

Acknowledgements

We thank Wang Yuqin, Xu Haitao, Deng Shoulong and Liu Chousheng for support. This work was supported by the Henan University of Science and Technology and The national animal husbandry station resources protection center. This work was funded in part by grants of the National Key Technology R&D Program of China (2011BAD28B05-2), and the National Meat Sheep System Specialist Program.

Author Contributions

W.C.H. and W.X.G. participated in study design, Z.L. and Z.C.Y. performed the experiments, L.R.Z. and Z.F.P. analyzed the data, W.M.M. wrote the manuscript. S.Y.L., W.H.H. and C.J.X. conducted the animal experiments. L.Z.X., M.Z. and W.Y.Y. performed the pathological analyses. D.L.X. and W.C.D. oversaw the study. All authors discussed the data. All authors reviewed the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Wu, M. *et al.* *Rosa26*-targeted sheep gene knock-in via CRISPR-Cas9 system. *Sci. Rep.* **6**, 24360; doi: 10.1038/srep24360 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>