RESEARCH ARTICLE

Protein kinase C is involved with upstream signaling of methyl farnesoate for photoperiod-dependent sex determination in the water flea *Daphnia pulex*

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ABSTRACT

Sex determination of *Daphnia pulex* is decided by environmental conditions. We established a suitable experimental system for this study using *D. pulex* WTN6 strain, in which the sex of the offspring can be controlled by photoperiod. Long-day conditions induced females and short-day conditions induced males. Using this system, we previously found that methy farnesoate (MF), which is a putative innate juvenile hormone molecule in daphnids, is necessary for male sex determination and that protein kinase C (PKC) is a candidate factor of male sex determiner. In this study, we demonstrated that a PKC inhibitor [bisindolylmaleimide IV (BIM)] application strongly suppressed male offspring induction in the short-day condition. Moreover, cotreatment of BIM with MF revealed that PKC signaling acts upstream of MF signaling for male sex determination. This is the first experimental evidence that PKC is involved in the male sex determination process associated with methyl farnesoate signaling in daphnid species.

KEY WORDS: *Daphnia pulex*, Environmental sex determination, Juvenile hormone, Methyl farnesoate, Photoperiod, Protein kinase C

INTRODUCTION

The micro-crustacean water flea, genus *Daphnia*, is a representative zooplankton in freshwater inland ecosystems. They have a unique sex determination system, which is tightly correlated with changeable habitat conditions. Under favorable environmental conditions, mothers exclusively produce female offspring via parthenogenesis, whereas under an unfavorable environment, such as low temperature and short-day length, they begin to produce parthenogenetic male offspring instead (Banta and Brown, 1929; Hobæk and Larsson, 1990; Kleiven et al., 1992).

Previous studies have demonstrated that juvenile hormone (JH) or its mimics could induce male offspring production in daphnid

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Received 16 September 2016; Accepted 5 December 2016

species even under female-producing conditions (Olmstead and LeBlanc, 2002; Tatarazako et al., 2003). Therefore, JH is considered to be a male sex determinant. Additionally, the JH-sensitive period for male offspring production occurs during late period of oocyte maturation, prior to embryo release into the brood pouch (Ignace et al., 2011; Kato et al., 2011). However, the regulatory mechanisms of a JH surge during this critical interval have not been elucidated yet due to the lack of a male-inducible system without JH treatment.

To fill this knowledge gap, we have recently established a femaleor male-inducible system using D. pulex WTN6 strain. The sex ratio of a clutch of this strain can be easily controlled by changing the photoperiod; a mother produces female progeny under long-day conditions (14 h light:10 h dark), whereas male progeny exclusively emerge under the short-day conditions (10 h light:14 h dark) (Toyota et al., 2015a). Based upon this induction system, we demonstrated that methyl farnesoate (MF) is likely an innate JH in daphnids (Toyota et al., 2015a), and furthermore, N-methyl-Daspartate (NMDA) receptors (a subtype of ionotropic glutamate receptors) act as upstream regulators of MF signaling during male offspring production (Toyota et al., 2015b). In addition to NMDA receptors, expression levels of serine/threonine kinase-coding genes were varied in both up- and downstream of MF signaling (Toyota et al., 2015b). We hypothesize that protein kinase C (PKC) might be a primary candidate factor of MF signaling since some previous works indicated that JH signaling is transduced via PKC activation (Yamamoto et al., 1988, 1997). However, whether PKC is involved in the MF pathway triggering male sex determination in daphnids is still largely unknown.

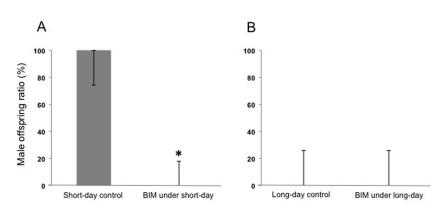
In this study, we conducted a pharmacological assay of PKC inhibitor treatment of the WTN6 strain, and demonstrated that this treatment strongly suppressed male offspring induction even under male-inducible (short-day) conditions. This is the first experimental evidence that PKC is involved in male sex determination associated with MF signaling, and provides a new picture of the signaling network underlying environmental sex determination in *Daphnia*.

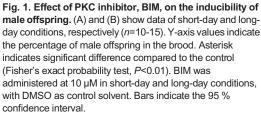
RESULTS AND DISCUSSION

To investigate the involvement of PKC in male offspring production under short-day conditions in *D. pulex*, we compared the sex ratio of offspring produced by females under PKC inhibition to control conditions for both long-day and short-day environments. Bisindolylmaleimide IV (BIM) was used as a PKC inhibitor (Denning et al., 1998). In the short-day control group (DMSOtreated), all mothers produced 100% male offspring, whereas in short-day BIM-treated groups, all mothers produced only femaleoffspring (Fig. 1A). In contrast, administration of BIM did not affect the proportion of female-producing mothers reared under the longday condition (Fig. 1B). These results strongly suggested that PKC is involved with the male-sex determining process in WTN6 strain.



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Next, we conducted the co-treatment of BIM with several concentrations of MF for dose-dependent male-inducible rates to clarify the hierarchical relationship between these signaling pathways. As shown in Fig. 2A, male offspring ratio showed an apparent increase in a dose-dependent manner to MF treatment. As a result of the co-treatment experiment, male induction rate did not change (Fig. 2B), indicating that PKC acts upstream of the MF signaling required for male sex determination.

Recent efforts have revealed that Methoprene-tolerant (MET) acts as a JH intracellular receptor in insects (Ashok et al., 1998; Jindra et al., 2013). In the presence of JH, MET binds to the partner protein SRC, which is also known as TAI in Drosophila melanogaster and FISC in Aedes aegypti, and forms the MET-SRC complex to activate its downstream factors (Charles et al., 2011; Li et al., 2011; Zhang et al., 2011). Moreover, it has been demonstrated that two daphnid species, D. pulex and D. magna, have an intracellular JH reception mechanism using MET-SRC complex that is similar to insects (Miyakawa et al., 2013). In addition to the intracellular JH signaling process, several lines of evidence have suggested that some actions of JH occur through membrane receptors via PKC signaling in male accessory glands in D. melanogaster (Yamamoto et al., 1988) and in ovarian follicles in Locusta migratoria (Zhou et al., 2002). Additionally, it has been reported using Drosophila cell line that PKC mediates phosphorylation of JH receptors to modulate binding of its receptors to JH-responsive elements (Kethidi et al., 2006). Furthermore, JH signaling via the PKC pathway may be widely present among crustaceans. For example, MF induces larval metamorphosis via PKC activation in the barnacle Balanus amphitrite (Yamamoto et al., 1997). Although PKC is considered as a crucial player in downstream mediation of JH signaling for nongenomic response, we reveal here that PKC also acts upstream of MF synthesis in daphnids.

Furthermore, our previous transcriptome analysis revealed that NMDA receptors act as an upstream factor of MF signaling (Toyota et al., 2015b); therefore, the next question addresses the regulatory relationship between the NMDA receptor and PKC involved with the MF synthesis in WTN6. Although these relationships have not been clarified yet in arthropods, including insects and crustaceans, a previous report has shown that PKC increases the channel-opening rate of NMDA receptors and induces a rapid delivery of functional NMDA receptors to the cell surface in *Xenopus* oocytes expressing NMDA receptors (Lan et al., 2001). This knowledge provides an important hint in the interpretation of our results and enables us to build the following hypothesis about male sex determination in *Daphnia* species: when a mother receives the short-day stimulus, (1) PKC is activated and recruits NMDA receptors, (2) NMDA

receptors mediate MF synthesis via expression of juvenile hormone *O*-methyltransferase (JHAMT) (Toyota et al., 2015a, b), and (3) MF binds to MET-SRC receptor complex to activate its downstream cascades for male sex determination (Miyakawa et al., 2013) (Fig. 3). To test this hypothesis and investigate whether this mechanism can be applied for other daphnid species, further experiments such as treatments of PKC activators will be necessary.

In this study, we demonstrated that PKC is involved with the male sex determination process in *D. pulex*. Moreover, co-treatment of PKC inhibitor with MF revealed that PKC acts upstream of MF signaling. This is the first evidence that PKC might be involved in MF synthesis. Although more detailed investigations examining the role of PKC in male sex determination process in daphnids will be required, our current findings provide important clues and a new hypothesis of the signaling network underlying photoperioddependent sex determination process in *D. pulex*.

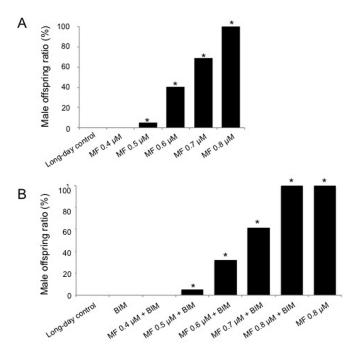


Fig. 2. Dose-dependent effects of methyl farnesoate (MF) with BIM on the inducibility of male offspring under long-day condition. (A) and (B) show data of MF and MF with BIM, respectively (n=16 and 15). Y-axis values indicate the percentage of male offspring in the brood. Asterisks indicate significant differences compared to respective controls (Fisher's exact probability test, P<0.01). Concentration of BIM was 10 µM.

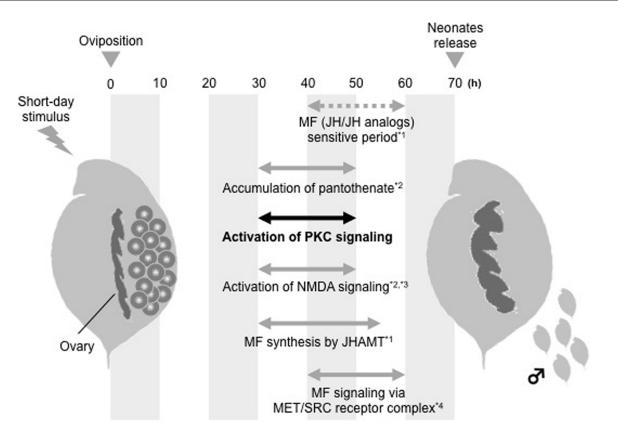


Fig. 3. Schematic illustration of a putative male sex-determining process triggered by short-day stimulus in *D. pulex* WTN6 strain. MF (JH/JH analog)sensitive period corresponds to an oocyte maturation period (40-60 h after oviposition) inside of mother's ovary. *1: Toyota et al. (2015a); *2: Toyota et al. (2015b); *3: Toyota et al. (2016); *4: Miyakawa et al. (2013).

MATERIALS AND METHODS

Daphnia pulex strain and its female- or male-producing conditions

The WTN6 strain of *D. pulex*, obtained from the Center for Genomics and Bioinformatics (Indiana University, IN, USA), was maintained at the National Institute for Basic Biology (Aichi, Japan) for more than 2 years. This strain was reared in dechlorinated fresh water, which was aerated and filtered through activated carbon for 2 weeks prior to use. A 0.04-ml suspension of 4.3×10^8 cells ml⁻¹ of *Chlorella vulgaris* was added daily to each culture (40 individuals/2 1). In this strain, the production of female and male offspring can be induced by rearing under 14 h light:10 h dark (long day) or 10 h light:14 h dark (short day) conditions, respectively (Toyota et al., 2015a). We determined the sex of offspring based on the length of the first antenna (Tatarazako et al., 2003).

Chemicals and treatment procedure

We used bisindolylmaleimide IV (BIM; \geq 98%; Sigma-Aldrich, St. Louis, MO, USA) as an inhibitor of PKC (Davis et al., 1992), and methyl farnesoate (MF; Echelon Bioscience, Salt Lake City, UT, USA) as a JH. Dimethylsulfoxide (DMSO; Nacalai Tesque, Kyoto, Japan) and dimethylformamide (DMF; analytical grade, Wako, Osaka, Japan) were used as solvent controls for BIM and MF, respectively, at concentrations below 0.01% (v/v). BIM dissolved in DMSO and MF dissolved in DMF were stored as 15 mM and 4 mM stock solutions, respectively, and kept at -20°C until use. All experiments (experiments 1 and 2) were conducted in 5 ml media in a 5 ml sampling tube (INA OPTICA, Osaka, Japan) containing one adult female (one-month-old or older) 30 h after ovulation, which is just before the MF-sensitive period. A total of 10-16 individuals were used for each treatment as replicates. In experiment 1, females were transferred to one of two ambient conditions: short-day or long-day, with a subset of females in each also exposed to BIM (10 µM). In experiment 2,

all females were subjected to the long-day conditions. Animals were exposed to several concentrations of MF (0.4, 0.5, 0.6, 0.7 and 0.8 μ M) either with or without a dose of BIM (10 μ M). We tried the BIM exposure experiments using 15, 20, and 25 μ M, however, all test animals died within 48 h after exposure. We checked sexes of all offspring produced and difference of sex ratio between treatments were statistically analyzed by Fisher's exact probability test with Holm's correction using R 2.15.3 (R Development Core Team, 2013).

Acknowledgements

We are grateful to Dr Margaret Beaton (Biology Department, Mount Allison University, Canada) and Dr Ryohei Yatsu (Department of Developmental Genetics, Wakayama Medical University, Japan) for their critical reading of the manuscript.

Competing interests

The authors declare no competing or financial interests.

Author contributions

K.T., T.S., N.T. and T.I. participated in the experimental design. K.T. prepared all daphnid samples and conducted all experiments. K.T. and T.I. wrote the first draft of the manuscript. All authors contributed to the final version of the manuscript.

Funding

This work was supported by the Japan Society for the Promotion of Science [Research Fellowship for Young Scientists 12J05579 to K.T.; KAKENHI Grant 15H06839 to K.T. and 15H04396 to T.I.].

References

- Ashok, M., Turner, C. and Wilson, T. G. (1998). Insect juvenile hormone resistance gene homology with the bHLH-PAS family of transcriptional regulators. *Proc. Natl. Acad. Sci. USA* 95, 2761-2766.
- Banta, A. M. and Brown, L. A. (1929). Control of sex in Cladocera. II. The unstable nature of the excretory products involved in male production. *Physiol. Zool.* 2, 93-98.

- Charles, J.-P., Iwema, T., Epa, V. C., Takaki, K., Rynes, J. and Jindra, M. (2011). Ligand binding properties of a juvenile hormone receptor, Methoprene-tolerant. *Proc. Natl. Acad. Sci. USA* **108**, 21128-21133.
- Davis, P. D., Hill, C. H., Lawton, G., Nixon, J. S., Wilkinson, S. E., Hurst, S. A., Keech, E. and Turner, S. E. (1992). Inhibitors of protein kinase C. 1,2,3-Bisarylmaleimides. J. Med. Chem. 35, 177-184.
- Denning, M. F., Wang, Y., Nickoloff, B. J. and Wrone-Smith, T. (1998). Protein kinase Cδ is activated by caspase-dependent proteolysis during ultraviolet radiation-induced apoptosis of human keratinocytes. J. Bio. Chem. 273, 29995-30002.
- Hobæk, A. and Larsson, P. (1990). Sex determination in *Daphnia magna*. Ecology **71**, 2255-2268.
- Ignace, D. D., Dodson, S. I. and Kashian, D. R. (2011). Identification of the critical timing of sex determination in *Daphnia magna* (Crustacea, Branchipoda) for use in toxicological studies. *Hydrobiologia* **668**, 117-123.
- Jindra, M., Palli, S. R. and Riddiford, L. M. (2013). The juvenile hormone signaling pathway in insect development. Annu. Rev. Entomol. 58, 181-204.
- Kato, Y., Kobayashi, K., Watanabe, H. and Iguchi, T. (2011). Environmental sex determination in the branchiopod crustacean *Daphnia magna*: Deep conservation of a *Doublesex* gene in the sex-determining pathway. *PLoS Genet.* 7, e1001345.
- Kethidi, D. R., Li, Y. and Palli, S. R. (2006). Protein kinase C mediated phosphorylation blocks juvenile hormone action. *Mol. Cell Endocrinol.* 247, 127-134.
- Kleiven, O. T., Larsson, P. and Hobaek, A. (1992). Sexual reproduction in Daphnia magna requires three stimuli. Oikos 65, 197-206.
- Lan, J., Skeberdis, V. A., Jover, T., Grooms, S. Y., Lin, Y., Araneda, R. C., Zheng, X., Bennett, M. V. L. and Zukin, R. S. (2001). Protein kinase C modulates NMDA receptor trafficking and gating. *Nat. Neurosci.* 4, 382-390.
- Li, M., Mead, E. A. and Zhu, J. (2011). Heterodimer of two bHLH-PAS proteins mediates juvenile hormone-induced gene expression. *Proc. Natl. Acad. Sci. USA* 108, 638-643.
- Miyakawa, H., Toyota, K., Hirakawa, I., Ogino, Y., Miyagawa, S., Oda, S., Tatarazako, N., Miura, T., Colbourne, J. K. and Iguchi, T. (2013). A mutation in the receptor Methoprene-tolerant alters juvenile hormone response in insects and crustaceans. Nat. Commun. 4, 1856.

- Olmstead, A. W. and Leblanc, G. A. (2002). Juvenoid hormone methyl farnesoate is a sex determinant in the crustacean *Daphnia manga. J. Exp. Zool.* 293, 736-739.
- R Development Core Team (2011). R: A Language and Environment for Statistical Computing. Vienna, Austria: The R Foundation for Statistical Computing.
- Tatarazako, N., Oda, S., Watanabe, H., Morita, M. and Iguchi, T. (2003). Juvenile hormone agonists affect the occurrence of male *Daphnia*. *Chemosphere* 53, 827-833.
- Toyota, K., Miyakawa, H., Hiruta, C., Furuta, K., Ogino, Y., Shinoda, T., Tatarazako, N., Miyagawa, S., Shaw, J. R. and Iguchi, T. (2015a). Methyl farnesoate synthesis is necessary for the environmental sex determination in the water flea Daphnia pulex. J. Insect Physiol. 80, 22-30.
- Toyota, K., Miyakawa, H., Yamaguchi, K., Shigenobu, S., Ogino, Y., Tatarazako, N., Miyagawa, S. and Iguchi, T. (2015b). NMDA receptor activation upstream of methyl farnesoate signaling for short day-induced male offspring production in the water flea, *Daphnia pulex*. *BMC Genomics* 16, 186.
- Toyota, K., Gavin, A., Miyagawa, S., Viant, M. R. and Iguchi, T. (2016). Metabolomics reveals an involvement of pantothenate for male production responding to the short-day stimulus in the water flea, *Daphnia pulex. Sci. Rep.* 6, 25125.
- Yamamoto, H., Okino, T., Yoshimura, E., Tachibana, A., Shimizu, K. and Fusetani, N. (1997). Methyl farnesoate induces larval metamorphosis of the barnacle, *Balanus amphitrite* via protein kinase C activation. *J. Exp. Zool.* 278, 349-355.
- Yamamoto, K., Chadarevian, A. and Pellegrini, M. (1988). Juvenile hormone action mediated in male accessory glands of *Drosophila* by calcium and kinase C. *Science* 239, 916-919.
- Zhang, Z., Xu, J., Sheng, Z., Sui, Y. and Palli, S. R. (2011). Steroid receptor co-activator is required for juvenile hormone signal transduction through a bHLH-PAS transcription factor, methoprene tolerant. J. Biol. Chem. 286, 8437-8447.
- Zhou, S., Zhang, J., Hirai, M., Chinzei, Y., Kayser, H., Wyatt, G. R. and Walker, V. K. (2002). A locust DNA-binding protein involved in gene regulation by juvenile hormone. *Mol. Cell Endocrinol.* **190**, 177-185.