Supplementary tables

Supplementary table 1. Clinical characteristics of patients with LUAD from whom scRNA-seq and ST data were obtained.

No.	Gender	Age	Smoking	Tumor	Tumor	Radiological	Histological
			status	location	diameter	type	type
TD1	Female	57	Never	Left upper	17mm	SN	IAC
				lobe			
TD2	Male	47	Never	Right	20mm	SN	IAC
				lower lobe			
TD3	Male	37	Never	Left upper	10mm	pGGN	MIA
				lobe			
TD4	Female	45	Never	Left upper	12mm	pGGN	MIA
				lobe			
TD5	Female	56	Never	Left upper	16mm	SSN	AIS
				lobe			
TD6	Female	56	Never	Right	21mm	SSN	MIA
				upper lobe			
TD7	Female	40	Never	Left lower	8mm	SSN	AIS
				lobe			
TD8	Male	69	Ever	Left upper	7mm	pGGN	AIS
				lobe			
TD9	Female	62	Never	Right	15mm	SSN	IAC
				upper lobe			

Supplementary table 2. List of marker genes for major six types of cells.

Cell type	Epithelial cells	T/NK cells	Myeloid cells	B cells	Endothelial cells	Fibroblasts
Marker genes	CAPS	GZMK	LYZ	JCHAIN	CLDN5	DCN
	SCGB1A1	GZMB	CTSD	IGHM	RAMP2	LUM
	WFDC2	NKG7	CCL18	IGHG1	CAV1	COL1A2
	KRT8	GNLY	CD68	IGHA1	VWF	COL3A1
	KRT19	KLRD1	CD14	IGHG4	PECAM1	COL1A1
	SCGB3A1	KLRC1	HLA-DRA	IGHA2	CDH5	FN1
	SCGB3A2	TIGIT	HLA-DRB1	IGHG3	EMCN	COL6A2
	KRT18	LTB	TNF	IGHG2	CD34	COL6A3
	EPCAM	TNFRSF4	FCGR3A	MZB1	CD31	ACTA2
	MUC1	TNFRSF18	CD63	IGHD	THBD	COL6A1
	SFTPC	IL2RA	CD163	CD79A		COL5A2
	SFTPA1	GZMH	FCGR2A	MS4A1		COL4A2
	SFTPA2	CD3D	TPSB2			
	SFTPB	FOXP3	TPSAB1			
	PGC		CPA3			
	AGER		HPGDS			
	CAV1		MS4A2			
	CYP4B1		AIF1			

KRT7	IL1RN
ID1	CD86
SFTPD	THBD
CDH1	S100A9
	S100A8
	CXCL8
	FCGR3B
	IL1RN
	PTGS2
	CD1E
	FCGR2A
	THBD
	HLA-DPB1
	CDIC
	CD40

Supplementary table 3. Clinicopathological characteristics of patients with early LUAD verified by IF analysis.

Variable			Histological				
				type			
	n	AIS(14)	n	MIA(18)	n	IAC(17)	P value
Gender	-		<u></u>				0.573
Male	5		7		8		
Female	9		11		9		
Age							0.794
≥65	2		4		4		
< 65	12		14		13		
Smoking							0.100
status							
Never	12		14		9		
Ever	2		4		8		
Family							0.178
history of	?						
tumor							
Absent	13		13		11		
Present	1		5		6		
Tumor							0.092
diameter							
0-10mm	5		1		1		
10-20mm	6		13		11		
20-30mm	3		3		5		
Tumor							0.781
location							
RUL	5		6		6		
RML	1		1		0		

RLL	2	4	3	
LUL	4	5	8	
LLL	2	2	0	
Radiological				< 0.001
type				
pGGN	7	4	1	
SSN	6	14	8	
SN	1	0	8	
Surgical				0.005
approach				
Lobectomy	7	8	16	
Sub-lobectomy	7	10	1	

Supplementary table 4. Clinicopathological characteristics of patients with early LUAD verified by bulk RNA-seq.

Variable				Histological			
				type			
	n	AIS(20)	n	MIA(17)	n	IAC(23)	P value
Gender							0.377
Male	4		6		9		
Female	16		11		14		
Age							0.323
≥65	3		4		8		
< 65	17		13		15		
Smoking							0.490
status							
Never	17		13		16		
Ever	3		4		7		
Family							0.021
history of	f						
tumor							
Absent	14		13		23		
Present	6		4		0		
Tumor							< 0.001
diameter							
0-10mm	9		3		0		
10-20mm	8		13		12		
20-30mm	3		1		11		
Tumor							0.012
location							
RUL	8		5		12		
RML	0		3		4		
RLL	2		0		3		
LUL	4		8		4		
LLL	6		1		0		

Radiological				< 0.001
type				
pGGN	7	4	0	
SSN	13	13	12	
SN	0	0	11	
Surgical				< 0.001
approach				
Lobectomy	3	10	20	
Sub-lobectomy	17	7	3	

Supplementary table 5. The reproducibility of scRNA-seq data across nine patient donors through irreproducible discovery rate (IDR) analysis.

Cell type	Cell subtype	Group	Marker gene	Irreproducible Discovery Rate
Epithelial	TM4SF1+ cancer cells (Epi-C0)	AIS	TM4SF1	4.48E-06
	TM4SF1+ cancer cells (Epi-C0)	MIA	TM4SF1	4.06E-05
	TM4SF1+ cancer cells (Epi-C0)	IAC	TM4SF1	0.00097041
	CRABP2+ cancer cells (Epi-C3)	AIS	CRABP2	0.266227814
	CRABP2+ cancer cells (Epi-C3)	MIA	CRABP2	0.090658529
	CRABP2+ cancer cells (Epi-C3)	IAC	CRABP2	0.019398353
	UBE2C+ cancer cells (Epi-C6)	IAC	UBE2C	0.003198881
T/NK	NK (T/NK-C4: GNLY+)	AIS	GNLY	7.77E-16
	NK (T/NK-C4: GNLY+)	MIA	GNLY	2.05E-13
	NK (T/NK-C4: GNLY+)	IAC	GNLY	8.10E-11
	Treg (T/NK-C6: FOXP3+)	AIS	FOXP3	0.000171547
	Treg (T/NK-C6: FOXP3+)	MIA	FOXP3	6.86E-08
	Treg (T/NK-C6: FOXP3+)	IAC	FOXP3	2.87E-10
Myeloid	Mast cells (Mye-C0:	AIS	TPSB2	0

	TPSB2+)				
	Mast cells				
	(Mye-C0:	MIA	TPSB2	4.97E-06	
	TPSB2+)				
	Mast cells				
	(Mye-C0:	IAC	TPSB2	5.62E-05	
	TPSB2+)				
	Monocytes				
	(Mye-C7:	AIS	FCN1	2.59E-05	
	FCN1+)				
	Monocytes				
	(Mye-C7:	MIA	FCN1	0.024205964	
	FCN1+)				
	Monocytes				
	(Mye-C7:	IAC	FCN1	0.365352387	
	FCN1+)				
D 11	MALT B cells	ATG	ICID	0.775010004	
B cells	(C4: IGHD+)	AIS	IGHD	0.775010024	
	MALT B cells	TA C	ICID	0.0702222	
	(C4: IGHD+)	IAC	IGHD	0.87832332	
	Secretory B cells	ATG	CZD CD	0.055046055	
	(C7: GZMB+)	AIS	GZMB	0.055846255	
	Secretory B cells	MA	CZD CD	0.042646200	
	(C7: GZMB+)	MIA	GZMB	0.042646208	
	Secretory B cells	TA C	CZD CD	0.000200401	
	(C7: GZMB+)	IAC	GZMB	0.000380491	
	Lymphatic				
T 1 4 1 1 11	endothelial cells	ATG	DDDM	1 12E 00	
Endothelial cells	(End-C4:	AIS	PDPN	1.13E-09	
	PDPN+)				
	Lymphatic				
	endothelial cells	3.674	2001	4.457.00	
	(End-C4:	MIA	PDPN	1.15E-08	
	PDPN+)				
	Lymphatic				
	endothelial cells				
	(End-C4:	IAC	PDPN	0	
	PDPN+)				
	Cancer-associated				
Fibroblast	fibroblast	AIS	FAP	0.000481336	
-	(Fib-C1)				
	Cancer-associated				
	fibroblast	MIA	FAP	2.74E-07	

Cancer-associa	ited		
fibroblast	IAC	FAP	0.001128006
(Fib-C1)			

Supplementary table 6. Ligands and receptors of TGF-β pathway.

Receptor type	Ligands
Type receptor	
ALK1/ACVRL1	TGF- β , BMP9 and BMP10
ALK2/ACVR1	BMPs and GDFs
ALK3/BMPR1A	BMPs
ALK4/ACVR1B	Activins,GDF8/myostatin and GDF11
ALK5/TGFBR1	TGF-βs, GDF/myostatin and GDF11
ALK6/BMPR1B	BMPs
ALK7/ACVR1C	BMP16/nodal
Type receptor	
TGFBR2/TBR	TGF-βs
BMPR2/BMPR	BMPs and GDFs
ACVR2/ActR A	Activins, BMPs and GDFs
ACVR2B/ActR B	Activins, BMPs and GDFs and BMP16/nodal
AMHR2/AMHR	MIF
Type receptor	
Betaglycan	TGF- β 1, TGF- β 2, TGF- β 3, Activin-A,BMP2,BMP4,BMP7 and GDF5
Endoglin	TGF-β1, TGF-β3, Activin-A, BMP2, BMP7 and BMP9

Supplementary table 7. Up-regulated genes in response to TGF-β pathway.

11	 <u> </u>	1 1 1
Genes		Gene names
		APC, ARID4B, BCAR3, BMP2, BMPR1A,
		BMPR2, CDH1, CDK9, CDKN1C, CTNNB1,
		ENG, FKBP1A, FNTA, FURIN, HDAC1, HIPK2,
		ID1, ID2, ID3, IFNGR2, JUNB, KLF10, LEFTY2,
		LTBP2, MAP3K7, NCOR2,
		NOG, PMEPA1, PPM1A, PPP1CA, PPP1R15A,
		RAB31, RHOA, SERPINE1, SKI, SKIL, SLC20A1,
		SMAD1, SMAD3, SMAD6, SMAD7, SMURF1,
		SMURF2, SPTBN1, TGFB1
		, TGFBR1, TGIF1, THBS1, TJP1, TRIM33,
		UBE2D3, WWTR1, XIAP

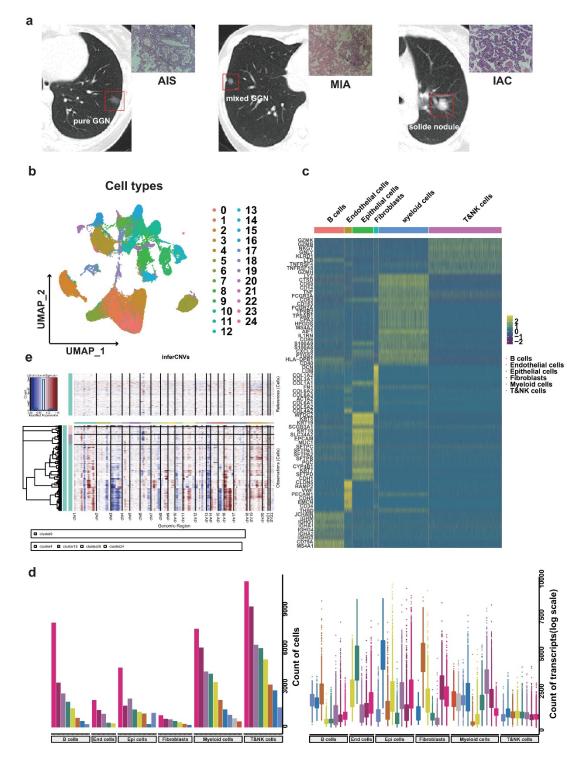
Supplementary table 8. The reproducibility of ST data across six patient donors through irreproducible discovery rate (IDR) analysis.

Cell type	Cell subtype	Group	Marker gene	Irreproducible Discovery Rate
Cancer cells	TM4SF1+ cancer cells (Epi-C0)	AIS	TM4SF1	0.001465066
	TM4SF1+ cancer	MIA	TM4SF1	0.984152457

	cells (Epi-C0)			
	TM4SF1+ cancer	IAC	TM4SF1	0.01611888
	cells (Epi-C0)			
	CRABP2+			
	cancer cells	AIS	CRABP2	0.874261321
	(Epi-C3)			
	CRABP2+			
	cancer cells	MIA	CRABP2	0.920942293
	(Epi-C3)			
	CRABP2+			
	cancer cells	IAC	CRABP2	0.478043675
	(Epi-C3)			
	UBE2C+ cancer	AIS	UBE2C	0.8686264 0.968448944
	cells (Epi-C6)			
	UBE2C+ cancer	MIA	UBE2C	
	cells (Epi-C6)	WIIA	OBE2C	
	UBE2C+ cancer	IAC	UBE2C	0.62342401
	cells (Epi-C6)	n ic	OBL2C	0.02342401
Lymph	T/NK	AIS	CD3D	0.780143153
	T/NK	MIA	CD3D	0.926281547
	T/NK	IAC	CD3D	0.409710723
	B cells	AIS	CD79A	0.3044021
	B cells	MIA	CD79A	0.882320403
	B cells	IAC	CD79A	0.495937474
Stromal	Endothelial cells	AIS	CLDN5	0.93870687
	Endothelial cells	MIA	CLDN5	0.79757546
	Endothelial cells	IAC	CLDN5	0.851591735
	Fibroblast	AIS	COL1A1	0.021762812
	Fibroblast	MIA	COL1A1	0.780095108
	Fibroblast	IAC	COL1A1	0.254921785
Epithelial cells	AT1	AIS	AGER	5.59E-08
	AT1	MIA	AGER	0.003351366
	AT1	IAC	AGER	0.001500678
	AT2	AIS	SFTPC	6.76E-08
	AT2	MIA	SFTPC	0.013318399
	AT2	IAC	SFTPC	2.37E-05

Supplementary figures

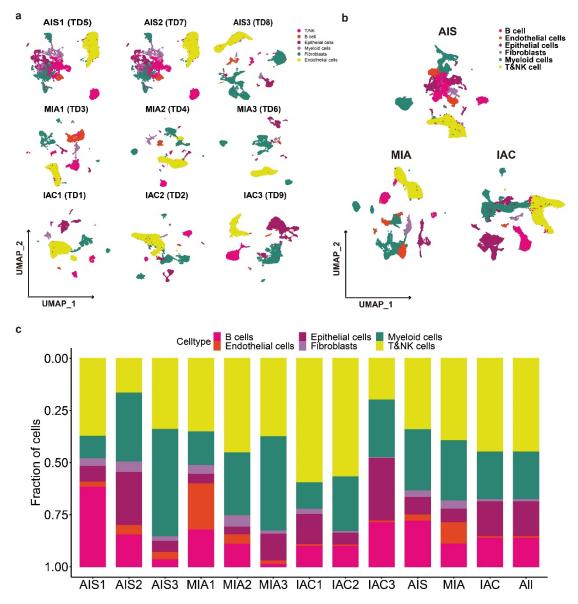
Supplementary Fig.1. Clinical characteristics and cell type identification of LUAD within three pathological stages, related with Fig. 1.



(a) Radiological and pathological features of enrolled cases (left panel: AIS represented by TD8, middle panel: MIA represented by TD3, right panel: IAC represented by TD1); (b) UMAP plot exhibit 25 cell type clusters with high confidence from all nine patients by scRNA-seq; (c) Heat map exhibit marker genes of six dominant cell types from scRNA-seq, the details of marker genes were listed in Table 4.d; (d) Quantity display of number cell (by histogram, left panel) and transcripts (by box diagram, right panel) of each subcluster from all patients in

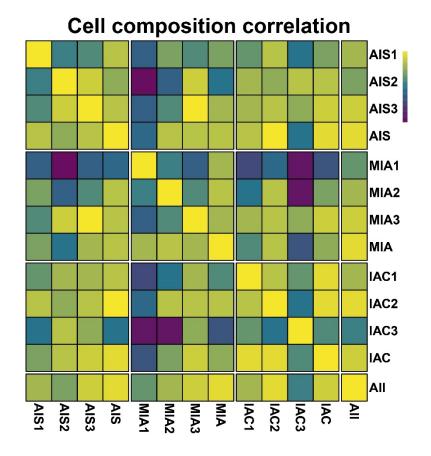
scRNA-seq data; (e) CNV atlas of cancer cell and normal epithelial cells in LUAD by scRNA-seq. Cluster 4 and Cluster 24 are identified as cancer cells, Cluster 19 and Cluster 20 are identified as normal epithelial cells.

Supplementary Fig.2. Duplicate results for cell type clustering.

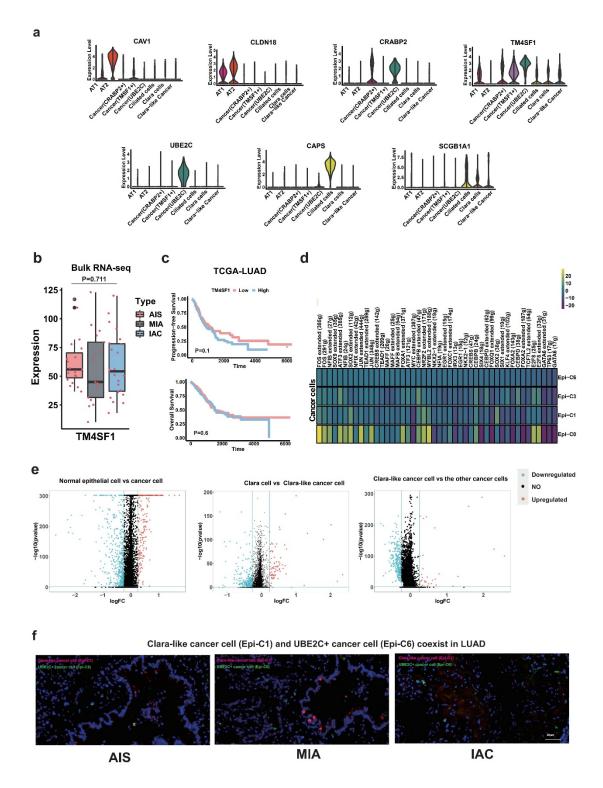


(a) Cell type clustering for each patient in all nine patients; (b) Cell type clustering in three stages of LUAD; (c) Distribution of major cell types in three stages and in each patient of LUAD.

Supplementary Fig.3. Correlation analysis of cellular components from scRNA-seq data.



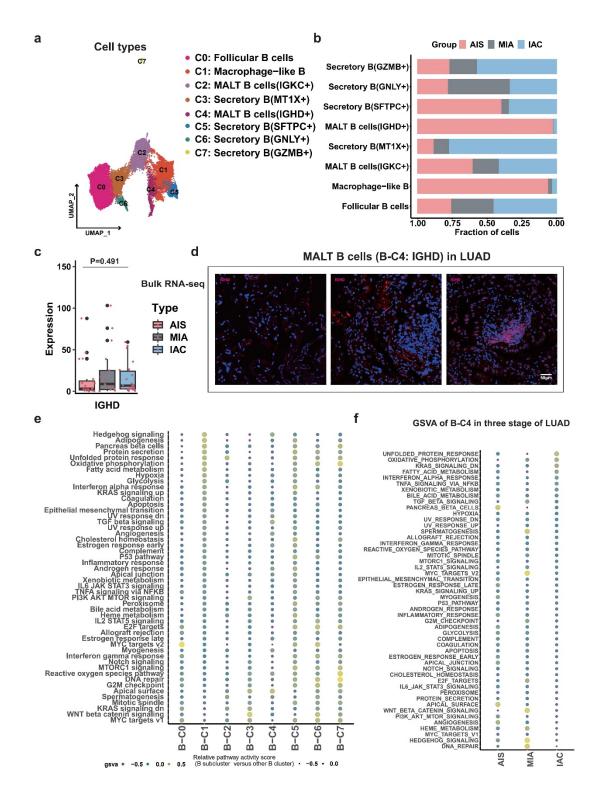
Supplementary Fig.4. Polyclonal origins of cancer cells, related with Fig. 2.



(a) Violin diagram reveal marker genes of four sub-clusters of normal epithelial cells and four sub-clusters of cancer cells. SCGB1A1 is common marker gene of Clara cell and Clara cell-like cancer cell; (b) Bulk RNA-seq validated that the expression level of the TM4SF1 gene increased in the process of cancer invasion; (c) Survival analysis of different TYM4SF1 expression levels in lung adenocarcinoma, from TCGA database; (d) Heat map of the SCENIC results showing the area under the curve (AUC) (enrichment score) for the regulation of TF expression for each kind of tumor

cell sub-cluster; (e) Volcano maps show comparisons of DEGs of Clara-like cancer cells and the other cancer cells from scRNA-seq data. The other normal epithelia cells (Epi-C2, Epi-C4, Epi-C5) vs the other cancer cells (Epi-C0, Epi-C3, Epi-C6); Clara cells (Epi-C7) vs Clara-like cancer cells (Epi-C1), Clara-like cancer cells (Epi-C1) vs the other cancer cells (Epi-C0, Epi-C3, Epi-C6). Blue represent downregulated DEGs, black represent no DEGs, red represent upregulated DEGs; (f) IF staining exhibit coexistence of Clara-like cancer (Epi-C1) and UBE2C (Epi-C6) with three pathological stages in LUAD. Red: Clara-like cancer (Epi-C1); Green: UBE2C (Epi-C6).

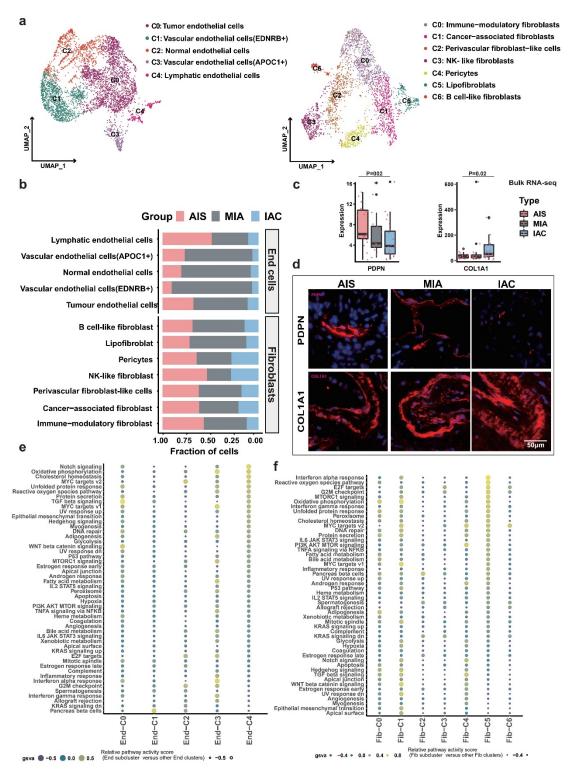
Supplementary Fig.5. Decrease of MALT B cells (B-C4: IGHD) drive invasion of LUAD, related with Fig. 4.



(a) U-MAP of scRNA-seq exhibit eight sub-clusters of B cells from 18,467 cells; (b) Proportions of eight sub-clusters of B cells distributing in three pathological stages of LUAD; (c) Bulk RNA-seq confirm that IGHD expression levels in three stages of LUAD; (d) IF validate that protein expression level of IGHD gradually decreased in the process of cancer invasion in LUAD; (e) Bubble diagram of GSVA exhibit the pathway activity score in each subtype of B cell; (f) Heatmap of GSVA results

showing distinct signaling pathways of MALT B cells (B-C4: IGHD) in three stages of lung adenocarcinoma

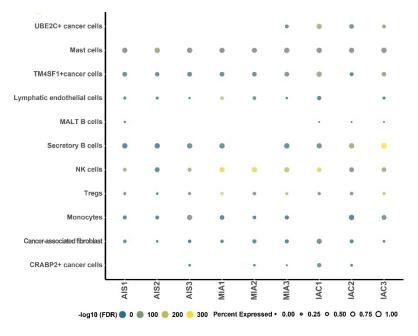
Supplementary Fig.6. Decrease of lymphatic endothelial cells (End-C4: PDPN) mediates the invasion of LUAD, related with Fig. 4.



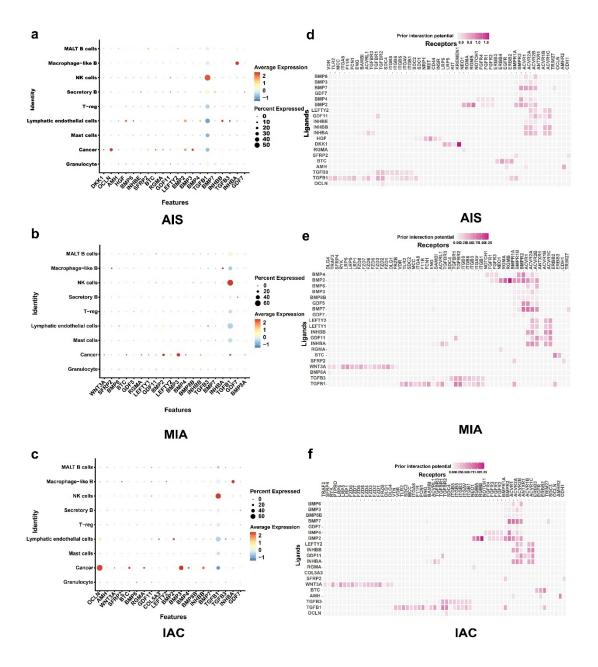
(a) UMAP of scRNA-seq exhibits sub-clusters of endothelial cells (left panel, 4,739

cells, 5 cell subtypes) and fibroblasts (right panel, 2,922 cells, 7 cell subtypes); (b) Proportions of five sub-clusters of endothelial cells (top) and seven sub-clusters of fibroblasts (bottom) distributed in three pathological stages of LUAD; (c) Bulk RNA-seq confirms that the expression level of PDPN gene (marker gene of lymphatic endothelium) gradually decreased from AIS to MIA and then to IAC (left panel), yet COL1A1 gene (marker gene of fibroblasts) show enrich in MIA; (d) IF validate that protein expression level of PDPN gene gradually decreased in the process of cancer invasion in LUAD, however COL1A1 gene show reverse trend; (e) Bubble diagram of GSVA exhibit the pathway activity score in each type of endothelial cell sub-clusters; (f) Bubble diagram of GSVA exhibit the pathway activity score in each type of fibroblasts sub-clusters.

Supplementary Fig.7. The reproducibility of gene expression for each major cell type through Wilcoxon on test for each individual patient.

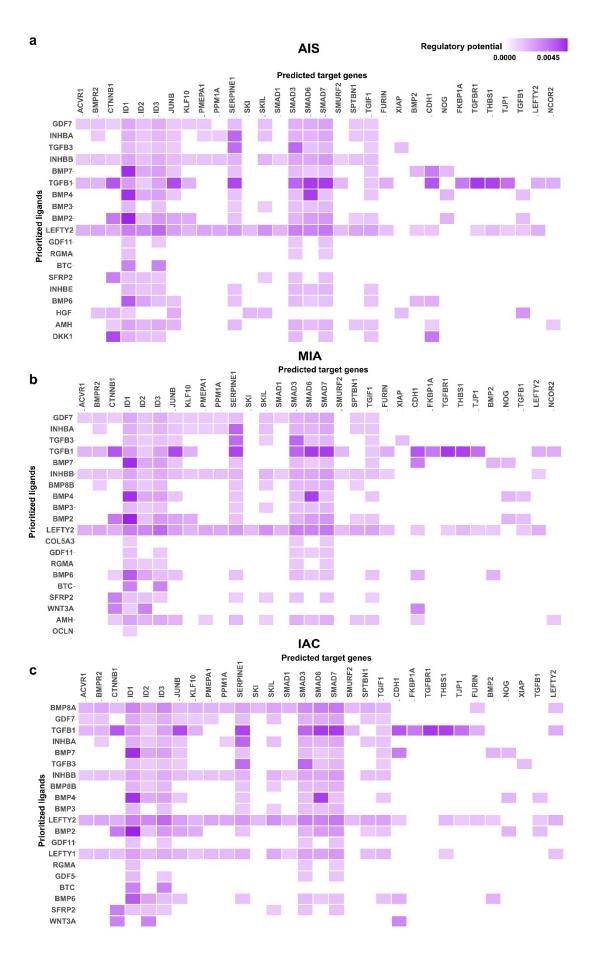


Supplementary Fig.8. Prediction of the interaction of receptors and ligands in cancer cells and TME cells.



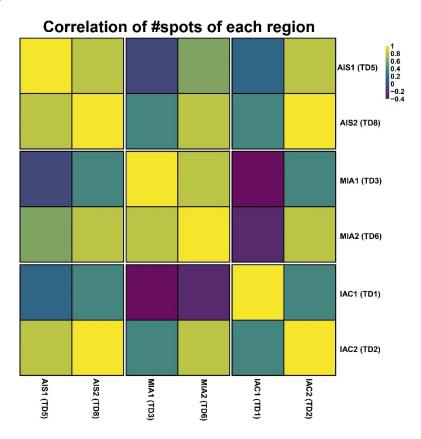
Top predicted ligand-receptor interactions presented between cancer cells and TME cells in AIS (a), MIA (b) and IAC (c). Top ligand-receptor interactions predicted by NicheNet in AIS (d), MIA (e) and IAC (f).

Supplementary Fig.9. Prediction of signaling pathways in cancer cells and TME cells.

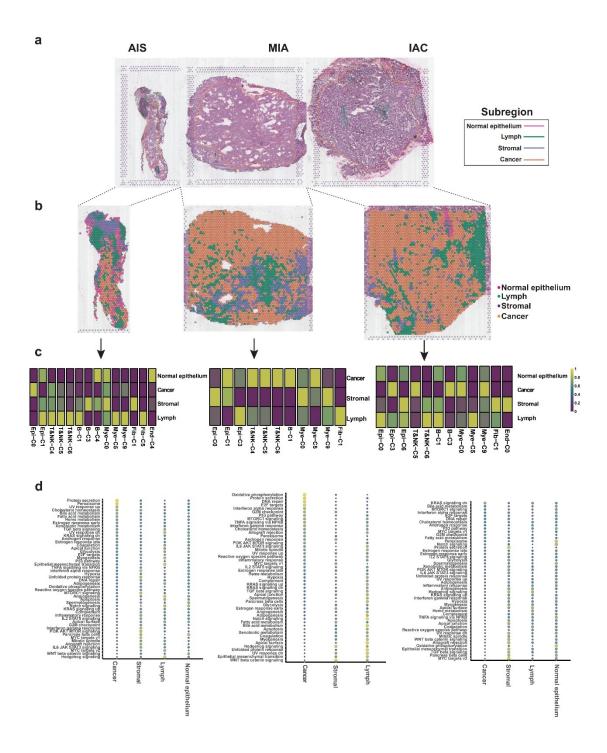


(a) The predicted target genes in AIS are predicted by NicheNet; (b) The predicted target genes in MIA are predicted by NicheNet; (c) The predicted target genes in IAC predicted by NicheNet.

Supplementary Fig.10. Correlation analysis of the number of spots in each region from ST data.



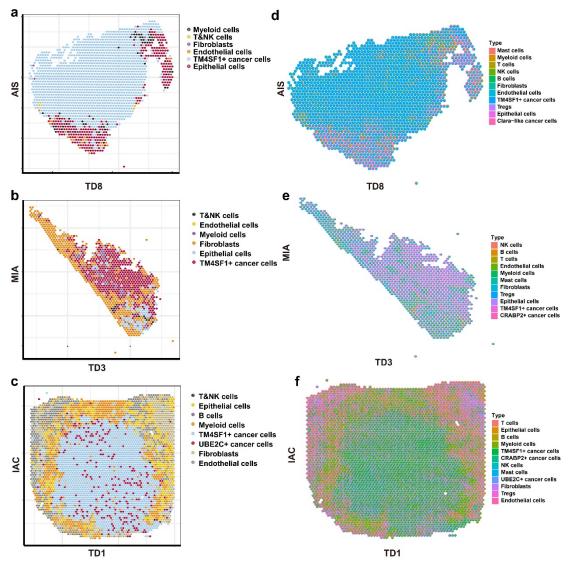
Supplementary Fig.11. Analysis of cellular spatial distribution and integration of single cell data in three pathological stages of LUAD, related with Fig. 6.



(a) HE staining exhibits the histological regions of AIS (left panel, TD5), MIA (middle panel, TD6) and IAC (right panel, TD2). Pink label: normal epithelium, green label: lymph, purple label: stromal, yellow label: cancer; (b) The spatial subregions of three pathological stages of LUAD annotated by ST; (c) Hyper-geometric maps of all certain cell types identified by scRNA-seq and regions defined by ST, yellow indicates high overlap enrichment; purple indicates low overlap enrichment; (d) Bubble diagram of GSVA exhibit the pathway activity score in each type of spatial

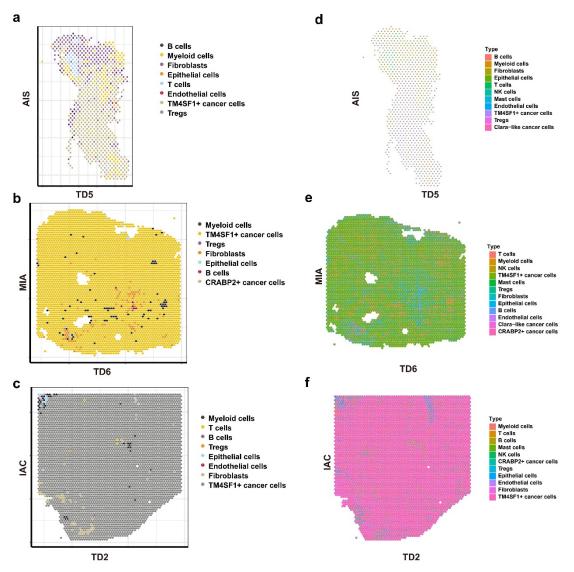
region.

Supplementary Fig.12. Cell types of each spatial spot for three stages of LUAD (TD8, TD3, and TD1) revealed by RCTD.

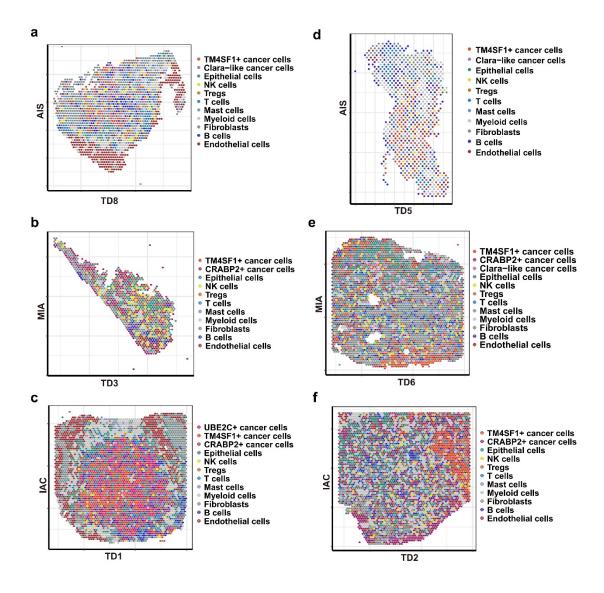


Major cell types in each spatial spot for AIS (TD8, a), MIA (TD 3, b), and IAC (TD 1, c); Pie chart of cell types in each spatial spot for AIS (d), MIA (e), and IAC (f).

Supplementary Fig.13. Cell types of each spatial spot for three stages of LUAD (TD5, TD6, and TD2) revealed by RCTD.

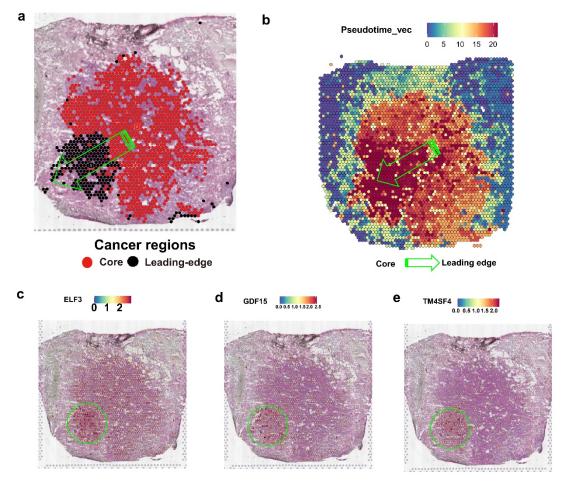


Major cell types in each spatial spot for AIS (TD5, a), MIA (TD 6, b), and IAC (TD 2, c); Pie chart of cell types in each spatial spot for AIS (d), MIA (e), and IAC (f). Supplementary Fig.14. Prediction of major cell types in LUAD based on each spatial spot by using Cell-ID.



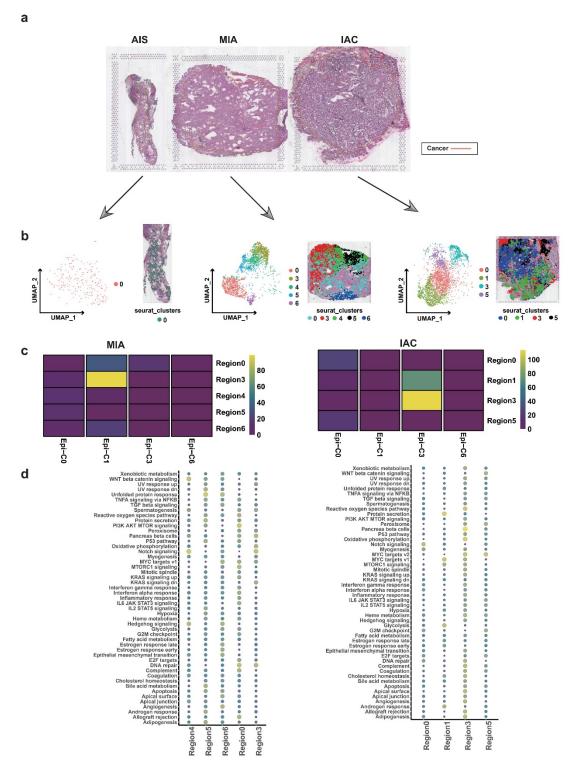
Cell types in the spatial spot of AIS (a, d), MIA (b, e) and IAC (c, f).

Supplementary Fig.15. Spatial differentiation trajectories of LUAD.



(a) ST analysis is used to sub-regionalize the cancer region; (b) The pseudo-time analysis of different regions (edge and core) of the tumor using SPAPA; Key marker gene of the leading edge were ELF3(c), GDF15 (d), and TM4SF4 (e).

Supplementary Fig.16. Spatial transcriptome and re-clustering of cancer cells, related with Fig. 7.



(a) HE staining exhibit the histological cancer subregions of AIS (left panel, TD5), MIA (middle panel, TD6) and IAC (right panel, TD2); (b) UMAP plot exhibit varieties of cancer subregions in HE-stained cancer region by ST; (c) Heat maps showing four cancer subclusters (Epi-C0, Epi-C1, Epi-C3, Epi-C6) obtained upon hyper-geometric distribution analysis of all specific cell types identified by scRNA-seq analysis and regions defined by ST analysis (left panel: MIA represented by TD6, right panel: IAC represented by TD2); (d) Bubble diagram of GSVA exhibit

the pathway activity score in each type of spatial subregion (left panel: MIA represented by TD6, right panel: IAC represented by TD2).