

OPEN

RAS mutations in AUS/FLUS Cytology: Does it Have an Additional Role in BRAF^{V600E} Mutation-Negative Nodules?

Jung Hyun Yoon, MD, Hyeong Ju Kwon, MD, PhD, Hye Sun Lee, MS, Eun-Kyung Kim, MD, PhD, Hee Jung Moon, MD, PhD, and Jin Young Kwak, MD, PhD

Abstract: The object of this study is to evaluate the additional role of RAS mutation in detecting thyroid malignancy among BRAF^{V600E} mutation-negative nodules diagnosed as atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS) on cytology.

From December 2009 to December 2011, 202 BRAF^{V600E} mutation-negative thyroid nodules diagnosed as AUS/FLUS cytology in 201 patients were included in this study. RAS mutation analysis was performed using residual material from ultrasonography-guided fine needle aspiration (US-FNA) cytology testing for K-RAS, N-RAS, and H-RAS codons 12/13 and 61 point mutations. The authors evaluated the association between RAS mutation status and cytopathologic characteristics.

Of the 202 BRAF^{V600E} mutation-negative thyroid nodules with AUS/FLUS cytology, 4 were considered insufficient for mutation analysis. Of the 198 thyroid nodules, 148 (74.7%) were confirmed as benign and 50 (25.3%) as malignant. Thirty-one (15.7%) of the 198 thyroid nodules were positive for any RAS mutation, 4 positive for K-RAS 12/13, 26 for N-RAS 61, and 1 positive for H-RAS 61. Seven (22.6%) of the RAS mutation positive nodules were malignant, 1 with K-RAS 12/13, 6 with N-RAS 61. Twenty-four (77.4%) of the 31 nodules positive for K-RAS 12/13 (N = 3), N-RAS 61 (N = 20), or H-RAS 61 (N = 1) mutations were proven benign. None of the 198 thyroid nodules were positive for K-RAS 61, N-RAS 12/13, or H-RAS 12/13 mutations.

N-RAS 61 mutation is the most common mutation detected among BRAF^{V600E} mutation-negative nodules with AUS/FLUS cytology. RAS mutation has limited value in predicting malignancy among BRAF^{V600E} mutation-negative thyroid nodules with AUS/FLUS cytology and further,

investigation is anticipated to evaluate the true role of RAS mutation in thyroid malignancy.

(*Medicine* 94(27):e1084)

Abbreviations: AUS/FLUS = atypia of undetermined significance/follicular lesion of undetermined significance, FNA = fine needle aspiration, US = ultrasonography.

INTRODUCTION

Although ultrasonography-guided fine needle aspiration (US-FNA) is considered the gold standard showing excellent diagnostic performances in the diagnosis of thyroid nodules,¹⁻⁴ 1 of the main flaws for this diagnostic method is “indeterminate” categories,⁵ including atypia of undetermined significance/follicular lesion of undetermined significance (AUS/FLUS), follicular neoplasm or suspicious for follicular neoplasm, and suspicious for malignancy categories according to the recently proposed Bethesda system for reporting thyroid cytopathology.⁶ Considering the broad range of malignancy rates among these categories, 5% to 75%, cytology results of these categories warrant further invasive investigations such as repeat US-FNA or diagnostic lobectomy,⁵⁻⁷ among which approximately 66% will eventually be proven benign.^{8,9} Atypia of undetermined significance/follicular lesion of undetermined significance in particular, is a heterogeneous cytologic category, including a broad disease spectrum, from patients with air-drying artifact, low cellularity, or obscuring elements to architectural or nuclear atypia.⁶ Malignancy risk for this category has a broad range, reported from 6% to 42.1%,¹⁰⁻¹⁵ with management guidelines varying among reports or institutions accordingly. At present, improving the accuracy of preoperative diagnosis for patients with this ambiguous cytologic category is critical in patient management.

With the rapid evolution in molecular genetics, molecular analysis has been reported and proposed as an effective adjunct to cytologic evaluation in the differential diagnosis of thyroid nodules, especially those with nondiagnostic or indeterminate cytology.¹⁶⁻¹⁹ BRAF^{V600E} mutation in particular, is present in 29% to 84% of papillary thyroid carcinoma (PTC),^{16,20-23} showing high specificity up to 100% and is potentially a useful marker in predicting PTC in clinical practice. Several recent studies have applied BRAF^{V600E} mutation analysis in AUS/FLUS nodules,^{11,24,25} and although this additional method was helpful in stratifying nodules among this cytologic category, approximately 39.7% to 70.8% of thyroid nodules negative for BRAF^{V600E} mutation proved to be malignant.^{11,24} RAS mutations are the second most common genetic alteration seen in thyroid cancers,^{26,27} and the most common genetic alteration seen in indeterminate thyroid nodules,²⁸ but how it helps in detecting malignancy among AUS/FLUS nodules,

Editor: Maohua Xie.

Received: March 24, 2015; revised: June 4, 2015; accepted: June 5, 2015. Department of Radiology, Severance Hospital, Research Institute of Radiological Science (JHY, E-KK, HJM, JYK); Department of Pathology, Yonsei University, College of Medicine (HJK); Department of Pathology, Yonsei University, Wonju College of Medicine (HJK); and Biostatistics Collaboration Unit, Medical Research Center, Yonsei University, College of Medicine (HSL), Seoul, South Korea.

Reprints: Jin Young Kwak, MD, PhD, Department of Radiology, Research Institute of Radiological Science, Yonsei University, College of Medicine, 50 Yonsei-ro Seodaemun-gu, 120-752 Seoul, South Korea (e-mail: docjin@yuhs.ac), Hyeong Ju Kwon, MD, PhD, Department of Pathology, Yonsei University, College of Medicine, Department of Pathology, Yonsei University, Wonju College of Medicine, 20 Ilisan-ro, 220-701, Wonju, South Korea.

This study was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) by the Ministry of Education (2013R1A1A2058817). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

The authors have no funding and conflicts of interest to disclose.

Copyright © 2015 Wolters Kluwer Health, Inc. All rights reserved.

This is an open access article distributed under the Creative Commons Attribution-ShareAlike License 4.0, which allows others to remix, tweak, and build upon the work, even for commercial purposes, as long as the author is credited and the new creations are licensed under the identical terms.

ISSN: 0025-7974

DOI: 10.1097/MD.0000000000001084

especially in those negative for BRAF^{V600E} mutation, has not been clarified.

In this study, we evaluated the usefulness of *RAS* mutations when used as an adjunct for definitive diagnosis of thyroid nodules with AUS/FLUS cytology, which are negative for BRAF^{V600E} mutation.

METHODS

This retrospective study has been approved by the Institutional Review Board (IRB) of Severance Hospital, Seoul, South Korea. Neither patient approval nor informed consent was required for review of medical records, cytology specimen, or US images. Signed informed consent was obtained from all patients before US-FNA or surgical procedures.

Patients

The Bethesda system has been used in reporting thyroid cytopathology starting from December 2009 at our institution (a referral center), and since then, this reporting system has been consistently used. We performed a retrospective review of our institutional database for all thyroid US-FNAs performed on nodules measuring larger than 5 mm during the period from December 2009 to December 2011, and 13,456 US-FNAs had been performed during this period. Among them, approximately 725 (5.4%) nodules had been diagnosed as AUS/FLUS on US-FNA cytology. BRAF^{V600E} mutation analysis was performed in 300 (41.4%) nodules, of which 262 (87.3%) nodules had negative results. Thyroid nodules fulfilling the following inclusion criteria were included in this study: nodules confirmed with surgery or nodules that had been diagnosed as benign or malignancy on follow-up US-FNA. Finally, 202 nodules in 201 patients were included in this study. Mean age of the 201 patients was 58.5 ± 11.8 years (range, 18–76 years). Mean size of the 202 thyroid nodules were 19.0 ± 11.4 mm (range, 5–61 mm).

US and US-FNA Procedures

Real-time US and US-FNA procedures were performed by 1 of 14 board-certified radiologists (3 faculty, 11 fellows, 1–15 years of experience) using a 5- to 12 MHz linear array transducer (iU22; Philips Medical Systems, Bothell, WA). Fine needle aspiration was performed on thyroid nodules exhibiting suspicious US features or at the largest mass among multiple benign-appearing nodules.

Ultrasonography-guided fine needle aspiration was performed with a 23-gauge needle connected to a 2 mL disposable syringe, using a freehand technique. Aspiration was performed at least twice for each thyroid nodule, during which aspirated material was expelled, smeared on glass slides, and placed immediately in 95% ethanol for Papanicolaou staining. The syringe used for aspiration was rinsed in normal saline for cell block processing. One of 5 cytopathologists specializing in thyroid cytopathology was involved in slide interpretation. Cytopathologists were not on-site during procedures, and additional immunohistochemical staining was performed on the cytopathologist's request. Since December 2009, cytology reports of our institution were based on the Bethesda system for reporting thyroid cytopathology.⁶

BRAF^{V600E} Mutation Analysis

Additional BRAF^{V600E} mutation analysis was performed when the referring clinicians requested them, or when it seemed relevant to providing definitive diagnosis of thyroid nodules

exhibiting suspicious US features. Aspiration was performed once more for BRAF^{V600E} mutation analysis after cytology slide preparations, which were rinsed in 1 mL of normal saline.

Dual priming oligonucleotide-based multiplex polymerase chain reaction analysis (DPO-PCR) was used for BRAF^{V600E} mutation analysis from May 2008 to November 2011,²⁹ and real-time PCR was used from November 2011 to the current date, according to the methods reported in prior studies.^{29,30}

RAS Mutation Analysis

RAS mutation analysis was performed at *N-RAS*, *H-RAS*, and *K-RAS* codon 12/13, and 61. Atypical cells of interest were isolated from US-FNA cytology smear slides, and DNA was extracted with QIAamp DNA mini kit (Qiagen, Germany) for PCR. A specific primer for exon 2 for amplification of codon 12/13 mutations and a primer for exon 4 for amplification of codon 61 mutations of the *N-RAS*, *H-RAS*, and *K-RAS* genes has been prepared for PCR reaction. The PCR reaction medium consisted of 2 μL DNA, 1X Real HelixTM qPCR mixture (Nanohelix, Korea), and optimized concentrations (10 nM) of primers in a final volume of 20 μL. Polymerase chain reaction was performed under the following cycling conditions: pre-denaturation at 95°C for 10 minutes, 45 cycles at 95°C for 30 seconds, 60°C for 30 seconds, and 72°C for 30 seconds, followed by final extension at 72°C for 5 minutes.

For sequencing analysis, the PCR product was purified, and single-strand reaction was carried out with each sequencing primer sets and BigDye® Terminator v3.1 Cycle Sequencing Kit (ABI, CA). The DNA sequence was obtained with an ABI PRISM 3730xl DNA Analyzer (ABI), and analyzed with the Sequencing Analysis 5.1.1 software.

Data and Statistical Analysis

Histopathologic results from surgery and US-FNA were used as the standard reference. Nodules with benign cytology results on follow-up US-FNA, which had been followed with US examinations performed at an interval of more than 12 months, also showing stable or decreased size were considered benign. Nodules diagnosed as malignant on US-FNA but had not undergone surgery were considered malignant.

Independent 2-sample *t*-test was used in comparison of continuous variables. Chi-square test or Fisher exact test was used in comparison of categorical variables. All tests were 2-sided, and *P* values of less than 0.05 were considered to have statistical significance. Analyses were performed using SAS (version 9.2, SAS Inc., Cary, NC).

RESULTS

Of the 202 BRAF^{V600E} mutation-negative thyroid nodules with AUS/FLUS cytology, 4 were found to have insufficient amount of isolated nucleic acid for mutation analysis. The remaining 198 thyroid nodules had undergone *RAS* mutation analysis. Of the 198 thyroid nodules, 148 (74.7%) were confirmed as benign, and 50 (25.3%) as malignant. Mean age of the patients with malignant nodules did not show significant differences to patients with benign nodules, 45.5 ± 10.9 years to 49.1 ± 11.8 years (*P* = 0.06). Mean size of the malignant nodules were significantly smaller than benign ones, 12.2 ± 10.3 mm to 16.2 ± 11.3 mm (*P* = 0.03).

Sixty-two patients with 62 (31.3%) thyroid nodules had undergone surgery; 32 were confirmed as classical type (PTC),

12 as PTC-follicular variant (FV-PTC), 9 as adenomatous hyperplasia, 3 as follicular adenoma, 2 as lymphocytic thyroiditis, 1 as medullary carcinoma, 2 as Hürthle cell adenoma, and 1 as Hürthle cell carcinoma, minimally invasive. One hundred thirty-six (69.7%) nodules in 135 patients had undergone follow-up US-FNA for diagnosis, and among them, 113 had benign cytology and 3 had malignant results on follow-up US-FNA. All the nodules with nondiagnostic (N=6) and AUS/FLUS (N=14) cytology on follow-up US-FNA had shown benign cytology results on third US-FNA.

Thirty-one (15.7%) of the 198 thyroid nodules were positive for RAS mutation, of which 7 (22.6%) were proven as malignant. Table 1 summarizes the RAS mutations to final pathology of the thyroid nodules included in this study. Of the 198 thyroid nodules, 4 were positive for K-RAS 12/13, 26 were positive for N-RAS 61, and 1 was positive for H-RAS 61. Of the 4 nodules positive for K-RAS 12/13, 1 (25.0%) was confirmed as papillary thyroid microcarcinoma (PTMC) (G12V). Of the 26 nodules positive for N-RAS 61, 6 (23.1%) were confirmed as malignant (2PTCs: Q61R, 1PTC: Q61K, 3 FV-PTC: Q61R). The single nodule positive for H-RAS 61 was diagnosed as benign on US-FNA cytology. Twenty-four (84.3%) of the nodules positive for K-RAS 12/13 (N=3), N-RAS 61 (N=20), or H-RAS 61 (N=1) mutations were proven benign. None of the 198 thyroid nodules were positive for K-RAS 61, N-RAS 12/13, or H-RAS 12/13 mutations.

Based on the number of RAS mutations present in this study, clinical features among thyroid nodules were analyzed for nodules with any RAS mutation (Table 2). Nodule size was significantly larger in nodules positive for N-RAS 61 mutation, 20.2 ± 14.5 mm to 14.4 ± 10.4 mm (P=0.01). Patient's age and sex did not show significant differences in thyroid nodules positive for RAS mutation (all P > 0.05).

DISCUSSION

Indeterminate cytology, including AUS/FLUS, has been considered problematic both to the patient and the clinician because a definite diagnosis has not been reached, but still, both the possibility of benignity or malignancy has been suggested with the ambiguous cytologic result. Atypia of undetermined significance/follicular lesion of undetermined significance cytology in particular, has a recommended malignancy rate of 15% to 30% by the Bethesda system,³¹ and although diagnostic lobectomy is recommended in patients with suspicious features for malignancy, the majority of patients with AUS/FLUS cytology is confirmed as benign for which surgery is rather unnecessary. In effort to improve the accuracy of cytological diagnosis, molecular testing has been recently introduced and BRAF, RAS, and RET/PTC mutations have been popularly used and evaluated as an adjunctive tool in detecting thyroid malignancy.^{32,33} BRAF^{V600E} mutation is the most common genetic alteration, which is highly specific for PTC,^{5,16,34,35} has been proven as a useful adjunct to US-FNA cytology in detecting malignancy among thyroid nodules with indeterminate cytology with high positive predictive value ranging from 87% to 100%.^{19,33} But, a considerable proportion of BRAF^{V600E} negative-nodules are proven as malignant, in particular, malignancy rate of BRAF^{V600E} negative-nodules with AUS/FLUS cytology has been reported to be 39.7% to 62.7%.^{11,36,37} Malignancy rate of BRAF^{V600E} mutation-negative nodules with AUS/FLUS cytology in this study was 25.3%, slightly lower than the prior reports, but showing that a

TABLE 1. Correlation of K-RAS, N-RAS, H-RAS to Final Pathology Results Among the 198 Thyroid Nodules With AUS/FLUS Cytology and Negative BRAF^{V600E} Mutation

	Benign (N = 148)	Malignant (N = 50)	Total
K-RAS 12/13			
Negative	145 (98.0%)	49 (98.0%)	198 (98.0%)
	Benign (129)	PTC (35)	
	AH (9)	FV-PTC (12)	
	LT (2)	HCC, MI (1)	
	FA (3)	Medullary (1)	
	HCA (2)		
Positive	3 (2.0%)	1 (2.0%)	4 (2.0%)
	Benign (3)	PTMC (1)	
	G12V:2	G12V:1	
	G12D:1		
K-RAS 61			
Negative	148 (100.0%)	50 (100.0%)	198 (100.0%)
Positive	0 (0.0%)	0 (0.0%)	0 (0.0%)
N-RAS 12/13			
Negative	148 (100.0%)	50 (100.0%)	198 (100.0%)
Positive	0 (0.0%)	0 (0.0%)	0 (0.0%)
N-RAS 61			
Negative	128 (86.5%)	44 (88.0%)	172 (87.1%)
	Benign (116)	PTC (33)	
	AH (6)	FV-PTC (9)	
	LT (2)	HCC, MI (1)	
	FA (2)	Medullary (1)	
	HCA (2)		
Positive	20 (13.5%)	6 (12.0%)	26 (12.9%)
	Benign (16)	PTC (3)	
	Q61R: 13	Q61R: 2,	
	Q61K: 3	Q61K: 1	
	AH (3)	FV-PTC (3)	
	Q61R: 3	Q61R: 3	
	FA (1)		
	Q61R: 1		
H-RAS 12/13			
Negative	148 (100.0%)	50 (100.0%)	198 (100.0%)
Positive	0 (0.0%)	0 (0.0%)	0 (0.0%)
H-RAS 61			
Negative	147 (99.3%)	50 (100.0%)	197 (99.5%)
	Benign (131)	PTC (36)	
	AH (9)	FV-PTC (12)	
	LT (2)	HCC, MI (1)	
	FA (3)	Medullary (1)	
	HCA (2)		
Positive	1 (0.7%)	0 (0.0%)	1 (0.5%)
	Benign (1)	-	
	Q61K:1		

considerable amount of thyroid cancers are missed by BRAF^{V600E} mutation analysis alone. In addition to BRAF^{V600E} mutation, RAS mutation, the second most common genetic alteration in thyroid cancer,^{38,39}

TABLE 2. Comparison of Clinical Features Among Thyroid Nodule According to N-RAS 61 Mutation

Clinical Features	K-RAS 12/13		N-RAS 61		H-RAS 61		Any RAS		P
	Negative (N = 194)	Positive (N = 4)	Negative (N = 172)	Positive (N = 26)	Negative (N = 197)	Positive (N = 1)	Negative (N = 167)	Positive (N = 31)	
Age (years)	48.3 ± 11.7	45.3 ± 13.0	48.5 ± 11.9	46.8 ± 10.5	48.3 ± 11.6	30.0	48.6 ± 11.8	46.0 ± 10.9	0.26
Sex									0.35
Men	39 (20.1%)	0 (0.0%)	31 (18.0%)	8 (30.8%)	158 (80.2%)	1 (100.0%)	31 (18.6%)	8 (25.8%)	
Women	155 (79.9%)	4 (100.0%)	141 (82.0%)	18 (69.2%)	39 (19.4%)	0 (0.0%)	136 (81.4%)	23 (74.2%)	
Nodule size (mm)	15.3 ± 11.2	10.0 ± 4.8	14.4 ± 10.4	20.2 ± 14.5	15.2 ± 11.2	15.0	14.5 ± 10.5	18.9 ± 13.8	0.06

has been reported to be helpful in detecting follicular-patterned thyroid tumors, especially FV-PTC among thyroid nodules of indeterminate cytology.^{28,38–40} Results of a recent study show that approximately 40.4% (23 of 57) BRAF^{V600E} mutation-negative nodules with AUS/FLUS cytology had RAS 61 mutations,²⁸ among which 52.2% were surgically confirmed as FV-PTC. The presence of RAS mutations suggests a possibility of a broad spectrum of tumors from benign follicular adenoma to follicular thyroid carcinoma, FV-PTC, anaplastic carcinoma and poorly differentiated thyroid carcinoma,^{28,38,39,41,42} conditions that are well known to be difficult to diagnose with cytology alone since they are often diagnosed as indeterminate with FNA, also with the majority negative for BRAF^{V600E} mutation. Most of the studies evaluating the efficacy of RAS mutation focus on thyroid nodules with indeterminate cytology, including AUS/FLUS, follicular neoplasm or suspicious for follicular neoplasm, and/or suspicious for malignancy categories,^{28,38–40} and to our knowledge, there are no studies published evaluating the usefulness of RAS mutation among AUS/FLUS cytology category alone in detecting thyroid cancers.

In this study, 15.7% (31 of 198) of the BRAF^{V600E} mutation-negative thyroid nodules with AUS/FLUS cytology were positive for RAS mutation, of which 22.6% were proven malignant. Also, 42.9% (3 out of 7 malignant nodules) were confirmed as FV-PTC on surgery, the remaining 57.1% diagnosed as PTC. Both the proportion of the presence of RAS mutation and the positive predictive value of RAS mutation was low, even when considering that this study had exclusively included nodules with AUS/FLUS cytology. Similar to the results of our study, in another study comparing the diagnostic utility of BRAF^{V600E} mutation to RAS and RET/PTC rearrangements, the presence of RAS mutation in AUS/FLUS cytology was 7.7%, whereas cancer risk was 0%.⁵ Unlike BRAF^{V600E} mutation, in which the presence of mutation almost always predicts malignancy, the predictive value of RAS mutation is not as confirmative, since RAS mutations is also detected in benign follicular adenomas, which obscures the performance of this specific mutation, which must be considered when applying it as an adjunctive diagnostic tool. As the prior study concluded, based on the results of our study, we feel that the presence of RAS mutation do not enhance the detection of thyroid cancers BRAF^{V600E} mutation-negative thyroid nodules with AUS/FLUS cytology, even those of follicular patterned growth.

RAS mutation consists of 3 highly homologous human RAS genes, N-RAS, K-RAS, and H-RAS carrying mutations in codons 12, 13, and 61 have been described to be related to thyroid tumorigenesis.^{38,39} The presence of different subtypes of RAS mutation has been reported to be related to different clinicopathological outcomes, K-RAS 12/13 mutation showing significantly lower carcinoma outcome compared with N-RAS 61 or H-RAS 61 mutations.²⁶ Our study showed similar results in that K-RAS 12/13 mutation has a low malignancy rate, 75% (3 of 4) of nodules with K-RAS 12/13 mutation were proven benign. N-RAS 61 mutation was the most common mutation (83.9%, 26 of 31), as with the results of previous reports,^{26,43} of which approximately 50% of nodules with N-RAS 61 mutation was proven as FV-PTC, consistent to the prior report in that FV-PTC is the cancer subtype mostly related to RAS 61 mutation.^{26,28,43} Only 1 benign thyroid nodule had positive results for H-RAS 61 mutation, which is in contrast with the prior report showing relatively high cancer rates in nodules with H-RAS 61 mutation.²⁸ In addition, malignancy rate of N-RAS 61 mutation was 23.1% (6 of 26), lower than the 25.0% (1 of 4) for K-RAS 12/13, results which are in contrast to the report in that K-RAS mutation withholds lower malignancy rate

than *N-RAS* mutation.²⁶ The small number of nodules positive for mutation may have affected the results, and as little has been proven in the true characteristics of *RAS* mutation subclassification, further studies including a large number of patients are anticipated to evaluate the true utility of *RAS* mutation when applied to lesion characterization or for risk stratification for patient management.

There are several limitations to this study. First, this study is of a retrospective design in which selection bias may have occurred during patient inclusion. Second, only 31.2% of the nodules included in this study had been confirmed with surgery, whereas the remaining 68.8% had been diagnosed based on cytology results. Third, subcategorization of AUS/FLUS cytology has not been applied. Prevalence of *RAS* mutation and predictive value for thyroid malignancy may differ among subcategories of AUS/FLUS which has not been considered in this study. Fourth, 1 of 5 cytopathologists was involved in cytology interpretation, in which interobserver variability may have existed. Fifth, direct sequencing has been used for mutation analysis in this study, a method which has been known to exclude mutations present in minor fractions of tumor cells,⁴⁴ which may have affected our results.

In conclusion, *N-RAS* 61 mutation is the most common mutation detected among BRAF^{V600E} mutation-negative nodules with AUS/FLUS cytology. *RAS* mutation has limited value in predicting malignancy among BRAF^{V600E} mutation-negative thyroid nodules with AUS/FLUS cytology and further investigation is anticipated to evaluate the true role of *RAS* mutation in thyroid malignancy.

REFERENCES

1. Yoon JH, Kwak JY, Kim EK, et al. How to approach thyroid nodules with indeterminate cytology. *Ann Surg Oncol*. 2010;17:2147–2155.
2. Yoon JH, Moon HJ, Kim EK, et al. Inadequate cytology in thyroid nodules: should we repeat aspiration or follow-up? *Ann Surg Oncol*. 2011;18:1282–1289.
3. Miller B, Burkey S, Lindberg G, et al. Prevalence of malignancy within cytologically indeterminate thyroid nodules. *Am J Surg*. 2004;188:459–462.
4. Ferraz C, Eszlinger M, Paschke R. Current state and future perspective of molecular diagnosis of fine-needle aspiration biopsy of thyroid nodules. *J Clin Endocrinol Metab*. 2011;96:2016–2026.
5. Rossi M, Buratto M, Tagliati F, et al. Relevance of BRAF mutation testing versus *RAS* point mutations and RET/PTC rearrangements evaluation in the diagnosis of thyroid cancer. *Thyroid*. 2015;25:221–228.
6. Bossuyt PM, Reitsma JB, Bruns DE, et al. Toward complete and accurate reporting of studies of diagnostic accuracy. The STARD initiative. *Am J Clin Pathol*. 2003;119:18–22.
7. Chudova D, Wilde JJ, Wang ET, et al. Molecular classification of thyroid nodules using high-dimensionality genomic data. *J Clin Endocrinol Metab*. 2010;95:5296–5304.
8. Kim SW, Lee JI, Kim JW, et al. BRAFV600E mutation analysis in fine-needle aspiration cytology specimens for evaluation of thyroid nodule: a large series in a BRAFV600E-prevalent population. *J Clin Endocrinol Metab*. 2010;95:3693–3700.
9. Song JY, Chu YC, Kim L, et al. Reclassifying formerly indeterminate thyroid FNAs using the Bethesda system reduces the number of inconclusive cases. *Acta Cytol*. 2012;56:122–129.
10. Bongiovanni M, Krane JF, Cibas ES, et al. The atypical thyroid fine-needle aspiration: past, present, and future. *Cancer Cytopathol*. 2012;120:73–86.
11. Hyeon J, Ahn S, Shin JH, et al. The prediction of malignant risk in the category “atypia of undetermined significance/follicular lesion of undetermined significance” of the Bethesda system for reporting thyroid cytopathology using subcategorization and BRAF mutation results. *Cancer Cytopathology*. 2014;122:368–376.
12. Lee KH, Shin JH, Oh YL, et al. Atypia of undetermined significance in thyroid fine-needle aspiration cytology: prediction of malignancy by US and comparison of methods for further management. *Ann Surg Oncol*. 2014;21:2326–2331.
13. Nayar R, Ivanovic M. The indeterminate thyroid fine-needle aspiration: experience from an academic center using terminology similar to that proposed in the 2007 National Cancer Institute Thyroid Fine Needle Aspiration State of the Science Conference. *Cancer*. 2009;117:195–202.
14. Yang J, Schnadig V, Logrono R, et al. Fine-needle aspiration of thyroid nodules: a study of 4703 patients with histologic and clinical correlations. *Cancer*. 2007;111:306–315.
15. Yassa L, Cibas ES, Benson CB, et al. Long-term assessment of a multidisciplinary approach to thyroid nodule diagnostic evaluation. *Cancer*. 2007;111:508–516.
16. Xing M, Tufano RP, Tufano AP, et al. Detection of BRAF mutation on fine needle aspiration biopsy specimens: a new diagnostic tool for papillary thyroid cancer. *J Clin Endocrinol Metab*. 2004;89:2867–2872.
17. Kang G, Cho EY, Shin JH, et al. Role of BRAFV600E mutation analysis and second cytologic review of fine-needle aspiration for evaluating thyroid nodule. *Cancer Cytopathol*. 2012;120:44–51.
18. Lee ST, Kim SW, Ki CS, et al. Clinical implication of highly sensitive detection of the BRAF V600E mutation in fine-needle aspirations of thyroid nodules: a comparative analysis of three molecular assays in 4585 consecutive cases in a BRAF V600E mutation-prevalent area. *J Clin Endocrinol Metab*. 2012;97:2299–2306.
19. Nikiforov YE, Ohori NP, Hodak SP, et al. Impact of mutational testing on the diagnosis and management of patients with cytologically indeterminate thyroid nodules: a prospective analysis of 1056 FNA samples. *J Clin Endocrinol Metab*. 2011;96:3390–3397.
20. Cohen Y, Rosenbaum E, Clark DP, et al. Mutational analysis of BRAF in fine needle aspiration biopsies of the thyroid: a potential application for the preoperative assessment of thyroid nodules. *Clin Cancer Res*. 2004;10:2761–2765.
21. Johnson SJ, Hardy SA, Roberts C, et al. Pilot of BRAF mutation analysis in indeterminate, suspicious and malignant thyroid FNA cytology. *Cytopathology*. 2014;25:146–154.
22. Kimura ET, Nikiforova MN, Zhu Z, et al. High prevalence of BRAF mutations in thyroid cancer: genetic evidence for constitutive activation of the RET/PTC-RAS-BRAF signaling pathway in papillary thyroid carcinoma. *Cancer Res*. 2003;63:1454–1457.
23. Kleiman DA, Sporn MJ, Beninato T, et al. Preoperative BRAF (V600E) mutation screening is unlikely to alter initial surgical treatment of patients with indeterminate thyroid nodules: a prospective case series of 960 patients. *Cancer*. 2013;119:1495–1502.
24. Guerra A, Fugazzola L, Marotta V, et al. A high percentage of BRAFV600E alleles in papillary thyroid carcinoma predicts a poorer outcome. *J Clin Endocrinol Metab*. 2012;97:2333–2340.
25. Park HJ, Moon JH, Yom CK, et al. Thyroid “atypia of undetermined significance” with nuclear atypia has high rates of malignancy and BRAF mutation. *Cancer Cytopathol*. 2014;122: 512–520.
26. Radkay LA, Chiosea SI, Seethala RR, et al. Thyroid nodules with KRAS mutations are different from nodules with NRAS and HRAS mutations with regard to cytopathologic and histopathologic outcome characteristics. *Cancer Cytopathol*. 2014;122:873–882.

27. Park SJ, Sun JY, Hong K, et al. Application of BRAF, NRAS, KRAS mutations as markers for the detection of papillary thyroid cancer from FNAB specimens by pyrosequencing analysis. *Clin Chem Lab Med*. 2013;51:1673–1680.
28. An JH, Song KH, Kim SK, et al. RAS mutations in indeterminate thyroid nodules are predictive of the follicular variant of papillary thyroid carcinoma. *Clin Endocrinol*. 2014;82:760–766.
29. Kwak JY, Kim EK, Kim JK, et al. Dual priming oligonucleotide-based multiplex PCR analysis for detection of BRAFV600E mutation in FNAB samples of thyroid nodules in BRAFV600E mutation-prevalent area. *Head Neck*. 2010;32:490–498.
30. Kwak JY, Han KH, Yoon JH, et al. BRAFV600E mutation testing in fine needle aspirates of thyroid nodules: potential value of real-time PCR. *Ann Clin Lab Sci*. 2012;42:258–265.
31. Cibas ES, Ali SZ. The Bethesda system for reporting thyroid cytopathology. *Am J Clin Pathol*. 2009;132:658–665.
32. Nikiforov YE, Nikiforova MN. Molecular genetics and diagnosis of thyroid cancer. *Nature reviews. Endocrinology*. 2011;7:569–580.
33. Nikiforov YE, Steward DL, Robinson-Smith TM, et al. Molecular testing for mutations in improving the fine-needle aspiration diagnosis of thyroid nodules. *J Clin Endocrinol Metab*. 2009;94:2092–2098.
34. Yoon JH, Kim EK, Moon HJ, et al. Is follow-up BRAF (V600E) mutation analysis helpful in the differential diagnosis of thyroid nodules with negative results on initial analysis? *PloS One*. 2013;8:e58592.
35. Xing M, Clark D, Guan H, et al. BRAF mutation testing of thyroid fine-needle aspiration biopsy specimens for preoperative risk stratification in papillary thyroid cancer. *J Clin Oncol*. 2009;27:2977–2982.
36. Jeong SH, Hong HS, Lee EH, et al. Outcome of thyroid nodules characterized as atypia of undetermined significance or follicular lesion of undetermined significance and correlation with Ultrasound features and BRAF (V600E) mutation analysis. *AJR Am J Roentgenol*. 2013;201:W854–860.
37. Lee EJ, Song KH, Kim DL, et al. The BRAF (V600E) mutation is associated with malignant ultrasonographic features in thyroid nodules. *Clin Endocrinol*. 2011;75:844–850.
38. Howell GM, Hodak SP, Yip L. RAS mutations in thyroid cancer. *Oncologist*. 2013;18:926–932.
39. Gupta N, Dasyam AK, Carty SE, et al. RAS mutations in thyroid FNA specimens are highly predictive of predominantly low-risk follicular-pattern cancers. *J Clin Endocrinol Metab*. 2013;98:E914–922.
40. Lee SR, Jung CK, Kim TE, et al. Molecular genotyping of follicular variant of papillary thyroid carcinoma correlates with diagnostic category of fine-needle aspiration cytology: values of RAS mutation testing. *Thyroid*. 2013;23:1416–1422.
41. Jang EK, Song DE, Sim SY, et al. NRAS codon 61 mutation is associated with distant metastasis in patients with follicular thyroid carcinoma. *Thyroid*. 2014;24:1275–1281.
42. Garcia-Rostan G, Zhao H, Camp RL, et al. ras mutations are associated with aggressive tumor phenotypes and poor prognosis in thyroid cancer. *J Clin Oncol*. 2003;21:3226–3235.
43. Fukahori M, Yoshida A, Hayashi H, et al. The associations between RAS mutations and clinical characteristics in follicular thyroid tumors: new insights from a single center and a large patient cohort. *Thyroid*. 2012;22:683–689.
44. Klebig F, Fischer C, Petri S, et al. Limitations in molecular detection of lymph node micrometastasis from colorectal cancer. *Diagn Mol Pathol*. 2007;16:91–95.