

Research Article

Metabolic Engineering of *Escherichia coli* for Poly(3-hydroxybutyrate) Production under Microaerobic Condition

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Received 2 January 2015; Revised 24 March 2015; Accepted 26 March 2015

Academic Editor: Yun-Peng Chao

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The alcohol dehydrogenase promoter P_{adhE} and mixed acid fermentation pathway deficient mutants of *Escherichia coli* were employed to produce poly(3-hydroxybutyrate) (P3HB) under microaerobic condition. The *E. coli* mutant with *ackA-pta*, *poxB*, *ldhA*, and *adhE* deletions accumulated 0.67 g/L P3HB, up to 78.84% of cell dry weight in tube cultivation. The deletion of pyruvate formate-lyase gene *pflB* drastically decreased P3HB production and P3HB content to 0.09 g/L and 24.44%, respectively. Overexpressing *pflB* via the plasmid in its knocked out mutant restored cell growth and P3HB accumulation, indicating the importance of the pyruvate formate-lyase in microaerobic carbon metabolism. The engineered *E. coli* BWapld (pWYC09) produced 5.00 g/L P3HB from 16.50 g/L glucose in 24 h batch fermentation, and P3HB production yield from glucose was 0.30 g/g, which reached up to 63% of maximal theoretical yield.

1. Introduction

Poly(3-hydroxybutyrate) (P3HB) is a polyester of 3-hydroxybutyrate that is synthesized by a variety of microorganisms as intracellular carbon and energy storages under unbalanced nutrients conditions [1]. The mechanical properties of P3HB are similar to those of petrol derived polymers such as polypropylene, which makes it a promising substitution of common plastics [2]. Moreover, P3HB can be obtained from renewable carbon sources and easily degraded into CO₂ and H₂O when discarded in natural environments. Thus, microbial production of P3HB via fermentation has attracted extensive attentions during the past few decades [3].

Although considerable efforts have been devoted to decreasing the production cost of P3HB to make it economically competitive, the present price of P3HB is still not feasible to replace traditional petro-based plastics. Generally, P3HB was mainly produced from renewable carbon sources through aerobic fermentation [4, 5]. Given that the feedstock

especially carbon source accounts up to a large portion of the production cost, low cost substrates such as crude glycerol and agricultural wastes have been explored for P3HB production [4, 6, 7]. On the other hand, aerobic fermentation process can be high demand of stir and air supply and contribute further energy cost besides feedstock. Microaerobic process could alleviate the energy requirement for fermentation by simplifying the bioreactor design and control strategies. Considering the potential advantages, P3HB production under microaerobic conditions has been investigated [8]. However, the P3HB content obtained under fed-batch microaerobic cultures was 51%, lower than that produced by aerobic fermentation [9]. Low polymer content decreases the purification efficiency and results in extra expenses in downstream process.

To achieve high P3HB content under microaerobic condition, the native promoter of P3HB biosynthesis operon *pha-CAB* from *Ralstonia eutropha* was changed to the anaerobic alcohol dehydrogenase promoter P_{adhE} , which was reported

TABLE 1: Strains, plasmids, and primers used in this study.

	Description	Reference
<i>E. coli</i> strains		
BW25113	<i>lacIq rrnBT14 ΔlacZWJ16 hsdR514 ΔaraBADAH33 ΔrhaBADLD78</i>	[11]
BWa	BW25113 <i>ΔackA-pta</i>	[11]
BWap	BW25113 <i>ΔackA-pta, ΔpoxB</i>	[11]
BWapl	BW25113 <i>ΔackA-pta, ΔpoxB, ΔldhA</i>	[11]
BWaplΔ	BW25113 <i>ΔackA-pta, ΔpoxB, ΔldhA, ΔadhE</i>	[11]
BWaplΔf	BW25113 <i>ΔackA-pta, ΔpoxB, ΔldhA, ΔadhE, ΔpflB</i>	[11]
Plasmids		
pWYC09	pBluescript II SK ⁻ derivatives containing <i>phaCAB</i> from <i>R. eutropha</i> with promoter P _{adhE} , Amp ^R	[10]
pBBR1MCS-2	Broad-host-range plasmid, Kan ^R	[10]
pMCS2pdc	Promoter P _{pdc} inserted into pBBR1MCS-2	This study
pMCS2pflB	<i>pflB</i> inserted into pMCS2pdc	This study
Primers		
pdcF	<u>5'-ATACTCGAGTTACGCTCATGATCGCGGCATGTC</u>	
pdcR	<u>5'-CCCCATATGTTACTCCATATATTCAAAAAC</u>	
plfF	<u>5'-GCTAGGCATATGTCCGAGCTTAATGAAAA</u>	
plfR	<u>5'-CCGAATTCTTACATAGATTGAGTGAAGGT</u>	

All oligonucleotides were synthesized by AuGCT Biotechnology (Beijing, China). Restriction endonuclease digestion sites were underlined.

to be upregulated under oxygen limited condition. As a result, P3HB content was improved from 30% to 48% of cell dry weight [10]. Moreover, *E. coli* undertakes mixed acid fermentation pathway, producing lactate, succinate, acetate, formate, and ethanol under oxygen limited conditions, which decreases the carbon flux into P3HB accumulation. *E. coli* mutant with deletions of *ackA-pta*, *poxB*, *ldhA*, and *adhE* was constructed and doubled the cell dry weight and improved P3HB production by 3.5-folds compared to the control [11].

In this paper, we managed to further increase the microaerobic P3HB production by employing the P_{adhE} promoter to drive the expression of *phaCAB* operon in *E. coli* mutants with defected mixed acid fermentation pathway. Moreover, the pyruvate formate-lyase, which catalyzes the coenzyme A-dependent, nonoxidative cleavage of pyruvate to acetyl-CoA and formate under anaerobic or microaerobic conditions, was overexpressed to investigate its effect on cell growth and P3HB accumulation (Figure 1).

2. Materials and Methods

2.1. Bacterial Strains and Plasmids Construction. Bacterial strains, plasmids, and primers used in this study were listed in Table 1. *E. coli* BW25113 mutants with serial deletions of *ackA-pta*, *poxB*, *ldhA*, *adhE*, and *pflB* were constructed to eliminate the mixed acid fermentation pathway. Plasmid pWYC09 was constructed to express *phaCAB* operon from *R. eutropha* under the control of P_{adhE}. The pyruvate decarboxylase promoter of *Zymomonas mobilis* was amplified with primers pdcF/pdcR and then ligated into the *XhoI/NdeI* site of pBBR1MCS-2 to construct pMCS2pdc. *pflB* was amplified from *E. coli* BW25113 genome with primers plfF/plfR and inserted into the *NdeI/EcoRI* site of pMCS2pdc to generate pMCS2pflB.

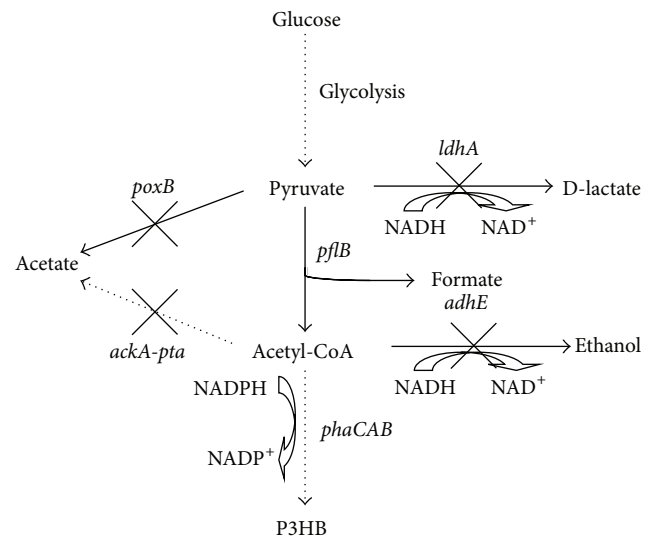


FIGURE 1: Strategies for improving P3HB production under microaerobic condition in *E. coli*.

2.2. Culture Conditions. For tube cultivation, one percent seed culture was inoculated into a 250 mL sealed tube completely filled with Luria-Bertani (LB) medium supplemented with 10 g/L glucose and then maintained at 37°C for 48 h as static cultures. For 5.5 liter bioreactor cultivation, 50 mL seed culture was transferred to the bioreactor containing 3 L LB medium supplemented with 20 g/L glucose. The pH was maintained at 7.0 via automatic addition of 5 M sodium hydroxide solution. Batch cultures were performed without air supply; the agitation was set at 75 rpm to prevent biomass sedimentation and maintain heat transfer and substrate exchange.

TABLE 2: P3HB accumulation and by-product formation of *E. coli* BW25113 and its five mutants harboring P_{adhE} controlled *phaCAB* operon.

<i>E. coli</i>	CDW (g/L)	P3HB content (%)	Glucose consumed (g/L)	Succinate (g/L)	Lactate (g/L)	Formate (g/L)	Acetate (g/L)	Ethanol (g/L)
BW25113	1.24 ± 0.08	44.85 ± 1.46	7.39 ± 0.09	0.34 ± 0.04	1.21 ± 0.12	0.06 ± 0.01	1.59 ± 0.08	0.25 ± 0.04
BWa	0.73 ± 0.10	68.31 ± 4.86	5.31 ± 0.16	0.37 ± 0.06	1.47 ± 0.26	0.04 ± 0.01	0.35 ± 0.02	0.19 ± 0.02
BWap	0.57 ± 0.06	70.69 ± 5.35	4.89 ± 0.24	0.28 ± 0.06	1.41 ± 0.16	0.03 ± 0.01	0.26 ± 0.06	0.21 ± 0.04
BWapl	1.33 ± 0.08	73.58 ± 4.72	6.26 ± 0.18	0.58 ± 0.08	ND	0.06 ± 0.02	0.45 ± 0.04	0.53 ± 0.06
BWapl _d	0.85 ± 0.04	78.84 ± 3.27	5.14 ± 0.23	0.32 ± 0.04	ND	ND	0.31 ± 0.02	ND
BWapl _d f	0.36 ± 0.03	24.44 ± 3.95	2.23 ± 0.08	0.18 ± 0.02	ND	ND	0.20 ± 0.05	ND

The recombinants harboring pWYC09 were cultivated in 250 mL sealed tubes completely filled with LB medium supplemented with 10 g/L glucose at 37°C for 48 h. CDW: cell dry weight. ND: not detected. Data shown are the average and standard deviation of three parallel experiments.

2.3. Analytical Methods. Cells were harvested by centrifugation at 8,000 ×g and 4°C for 10 min. The cell pellets were washed twice with distilled water and then lyophilized for 12 h for cell dry weight (CDW) assay. Lyophilized cells were subjected to methanolysis at 100°C for 4 h in the presence of 3% (v/v) H₂SO₄ and then assayed with a gas chromatograph (GC) to measure the P3HB content. For glucose and by-products lactate, succinate, acetate, formate, and ethanol measurements, the supernatant of cell cultures was filtered through a 0.2 μm syringe filter and then analyzed by high-performance liquid chromatography (HPLC) equipped with an ion exchange column (Aminex HPX-87H, 300 × 7.8 mm) and a refractive index detector.

3. Results and Discussion

3.1. P_{adhE} Controlled P3HB Production in *E. coli* Mixed Acid Fermentation Mutants. The alcohol dehydrogenase promoter P_{adhE} has been demonstrated to improve the expression level of *phaCAB* operon under microaerobic condition [10]. Moreover, the disruption of mixed acid fermentation pathway was proved to be another effective strategy to increase P3HB accumulation [11]. To investigate the possibility of the synergistic effect, plasmid pWYC09 carrying P_{adhE} controlled *phaCAB* was transformed into the wild type *E. coli* and a series of mixed acid fermentation mutants BWa, BWap, BWapl, BWapl_d, and BWapl_df (Table 1). The recombinant strains were cultivated in LB medium supplemented with glucose to produce P3HB under microaerobic tubes. The mixed acid fermentation mutants showed higher P3HB content compared to that of wild type except for the *pflB* mutant, and P3HB accumulated up to 78.84% of cell dry weight in the quadruple Δ *ackA-pta*, Δ *poxB*, Δ *ldhA*, and Δ *adhE* deletion mutant BWapl_d, which produced the fewest amount of mixed acid (Table 2). Compared to the strains containing *phaCAB* operon with its native promoter, P_{adhE} driven operon led to much higher P3HB production [11].

When *pflB* was knocked out, both cell growth and P3HB accumulation decreased significantly (Table 2). It was reported that the expression of pyruvate formate-lyase was induced to generate acetyl-CoA and formate from pyruvate when *E. coli* was cultivated with limited oxygen. Therefore, the deletion of *pflB* may cause the disruption of glucose metabolism and led to much lower cell biomass and P3HB

accumulation. The profile of by-product formation was similar to the previous report [11]. Lactate production was totally eliminated by deleting *ldhA*, and only minute amount of acetate and succinate was detected in BWapl_d and BWapl_df (Table 2).

3.2. Effect of *pflB* Overexpression on Cell Growth, P3HB Accumulation, and By-Product Formation. Considering the importance of *pflB* in glucose metabolism under oxygen limited condition, plasmid pMCS2*pflB* was constructed to constitutively expressing *pflB* and cotransformed with pWYC09 into *E. coli* to study its influence on cell metabolism profile. The plasmid carrying *pflB* overexpression restored the cell growth and P3HB production completely in *pflB* deletion mutant BWapl_df. Cell dry weight was increased from 0.36 g/L to 0.70 g/L, and P3HB content was increased from 24.44% to 44.57%, which confirmed the crucial role of *pflB* during the microaerobic P3HB production (Table 3). However, *pflB* overexpression did not affect the cell growth or P3HB accumulation significantly in other mutants; only a slight improvement in terms of the CDW and P3HB content was observed in BWapl_d mutant. Compared to those of the control without *pflB* overexpression, BWapl_d harboring pWYC09 and pMCS2*pflB* grew up to 1.16 g/L CDW and accumulated 84.79% of P3HB, exhibiting 37% and 8% increase, respectively (Table 3). These results indicate that the expression of *pflB* from its native genomic copy could satisfy the requirement of *E. coli* cells for most of cases of experimental condition, and its expression was proved be of crucial importance to oxygen limited glucose metabolism. Furthermore, no large amount of formate production was observed in all *pflB* overexpressing strains, probably due to the expression of formate dehydrogenase, converting formate to CO₂ and H₂O when formate amount exceeds the tolerance of *E. coli* [12].

3.3. Growth and P3HB Production by *E. coli* BWapl_d Mutants in a 5.5-Liter Fermenter. The recombinant *E. coli* BWapl_d (pWYC09) and BWapl_d (pWYC09 + pMCS2*pflB*) were cultivated in oxygen limited 5.5 liter bioreactor to investigate cell growth and P3HB production profile. After 24 h cultivation, the recombinant BWapl_d (pWYC09) consumed 16.50 g/L glucose and produced 5.00 g/L P3HB; the P3HB content reached 73.58% of cell dry weight. The by-product formation

TABLE 3: Influence of *pflB* overexpression on cell growth, P3HB accumulation, and by-product formation.

<i>E. coli</i>	CDW (g/L)	P3HB content (%)	Glucose consumed (g/L)	Succinate (g/L)	Lactate (g/L)	Formate (g/L)	Acetate (g/L)	Ethanol (g/L)
BW25113	0.85 ± 0.03	38.44 ± 5.91	6.95 ± 0.17	0.68 ± 0.03	0.97 ± 0.09	0.23 ± 0.02	1.68 ± 0.01	0.45 ± 0.06
BWa	0.58 ± 0.08	71.58 ± 6.93	5.31 ± 0.13	0.72 ± 0.06	1.29 ± 0.12	0.20 ± 0.01	0.64 ± 0.06	0.29 ± 0.04
BWap	0.64 ± 0.02	66.22 ± 4.67	4.89 ± 0.12	0.64 ± 0.01	1.16 ± 0.08	0.18 ± 0.02	0.24 ± 0.07	0.25 ± 0.03
BWapl	1.12 ± 0.07	76.47 ± 2.98	5.36 ± 0.26	0.84 ± 0.02	ND	0.11 ± 0.04	0.29 ± 0.04	0.38 ± 0.05
BWapl _d	1.16 ± 0.05	84.79 ± 2.37	6.24 ± 0.32	0.85 ± 0.05	ND	0.25 ± 0.06	0.35 ± 0.04	ND
BWapl _d _f	0.70 ± 0.06	44.57 ± 4.02	3.78 ± 0.26	0.73 ± 0.02	ND	0.15 ± 0.04	0.16 ± 0.05	ND

The recombinants harboring pWYC09 and pMCS2pflB were cultivated in 250 mL sealed tubes completely filled with LB medium supplemented with 10 g/L glucose at 37°C for 48 h. CDW: cell dry weight. ND: not detected. Data shown are the average and standard deviation of three parallel experiments.

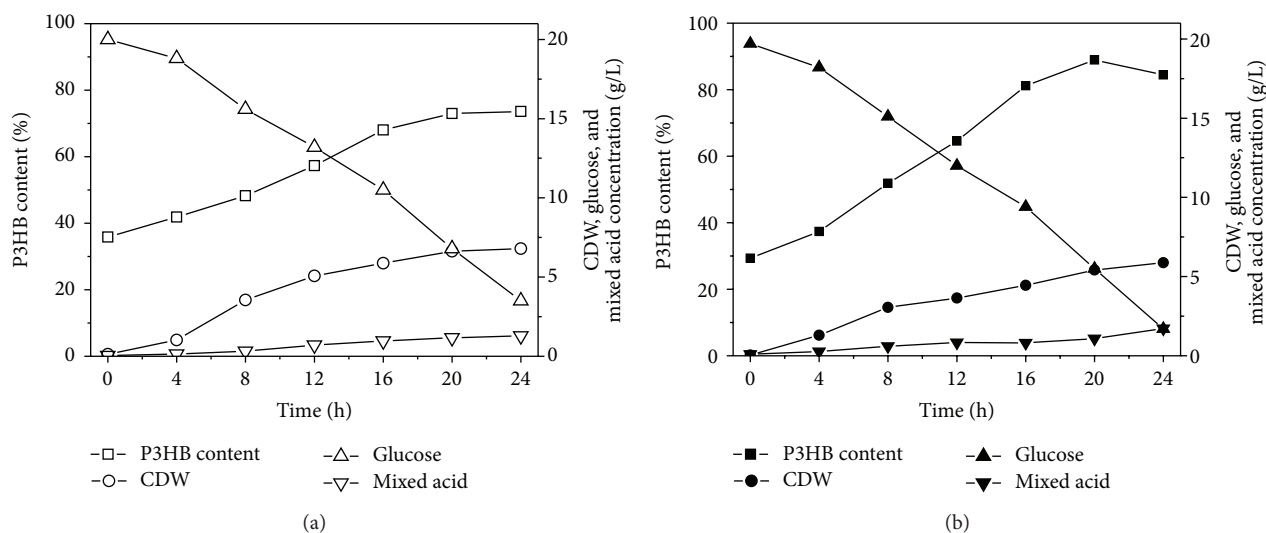


FIGURE 2: Cell dry weight (CDW), P3HB content (%), glucose consumption, and mixed acid production profile of *E. coli* BWapl_d harboring P_{adhE} controlled *phaCAB* operon without (a) or with (b) *pflB* overexpression in 5.5 liter bioreactor under microaerobic condition.

was maintained at low level during the process, and finally 0.23 g/L succinate, 0.86 g/L acetate, 0.17 g/L formate, and 0.03 g/L ethanol were detected (Figure 2(a)). When *pflB* was overexpressed, the final CDW and P3HB content were 5.88 g/L and 84.43%, respectively, accompanied with a total amount of 1.72 g/L mixed acid (Figure 2(b)). Previously, the wild type strain BW25113 harboring the same plasmid produced 6.72 g/L CDW with 44.53% P3HB under the same condition. In this study, with the disruption of mixed acid fermentation pathways, the recombinant accumulated P3HB up to 73.58% of cell dry weight, and overexpressing *pflB* helped to further improve P3HB content to 84.43%, which will facilitate downstream biopolymer purification processes.

In terms of byproduct formation and P3HB production yield, the wild type strain BW25113 produced 13.60 g/L mixed acids, and *pta* mutant JW2294 generated 16.60 g/L mixed acids (Table 4). In this study, the mixed acid fermentation pathway mutant BWapl_d only produced 1.29 g/L mixed acids. As a result, the P3HB production yield from glucose was significantly increased. The wild type obtained 0.10 g P3HB/g glucose, and the *pta* mutant showed a 70% increase to 0.17 g/g. BWapl_d mutant reached up to 0.30 g/g, which was 2 times higher than that obtained from the wild type (Table 4).

The maximal theoretical yield of P3HB from glucose is 0.48 g P3HB/g glucose. Previously, the *arcA* mutant showed 46% of maximal yield under optimized microaerobic fermentation condition [9]. The recombinant constructed in this study reached up to 63% of maximal yield. Hence, the engineered *E. coli* constructed here showed superior ability for microaerobic P3HB production compared to previous strains. Further optimization of different degrees of oxygen limitation in bioreactor batch cultivations should be performed to obtain much higher P3HB production titer and productivity. Since the overall redox balance is important to P3HB biosynthesis and the metabolic response [13], P3HB production with other substrates with different oxidation states of the carbon atoms such as gluconate and glycerol could be another choice for improvements under microaerobic condition.

4. Conclusion

The alcohol dehydrogenase promoter and mixed acid fermentation mutants were combined to improve P3HB production under microaerobic condition. The deletion of *pflB* gene significantly decreased cell biomass and P3HB production

TABLE 4: Summary of engineered *E. coli* for improving P3HB production under microaerobic condition.

<i>E. coli</i>	Description	Cell dry weight (g/L)	P3HB content (%)	P3HB concentration (g/L)	Mixed acid (g/L)	Yield (g P3HB/g glucose)
BW25113	Wild type	6.72	44.53	3.06	13.60	0.10
JW2294	<i>pta</i> knockout	7.75	64.31	4.98	16.60	0.17
BWapld	Mixed acid fermentation deficient	6.79	73.58	5.00	1.29	0.30
BWapld (pMCS2pflB)	Mixed acid fermentation deficient and <i>pflB</i> overexpression	5.88	84.43	4.96	1.72	0.27

All *E. coli* strains carried plasmid pWYC09, which harbored *phaCAB* operon under the control of P_{adhE}. Mixed acid referred the total amount of succinate, formate, acetate, and ethanol.

and overexpressing *pflB* from the plasmid restored the cell growth and P3HB accumulation, indicating the importance of pyruvate formate-lyase for microaerobic metabolism. The engineered strain BWapld harboring pWYC09 accumulated 5.00 g/L P3HB, up to 73.58% of cell dry weight in 5.5 liter bioreactor fermentation. The P3HB production yield from glucose was 0.30 g/g, up to 63% of maximal theoretical yield. The optimization of *phaCAB* expression in mixed acid fermentation mutants was proved to be effective for improving microaerobically P3HB production.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This research was sponsored by Grants from National Natural Science Foundation of China (31100025 and 31260015), National Basic Research Program of China (2013CB733600 and 2012CB725200), Beijing Higher Education Young Elite Teacher Project (YETP0523), and Key Laboratory for Agrobiotechnology of China Agricultural University (2014SKLAB06-10).

References

- [1] G.-Q. Chen and M. K. Patel, "Plastics derived from biological sources: present and future. A technical and environmental review," *Chemical Reviews*, vol. 112, no. 4, pp. 2082–2099, 2012.
- [2] S. Philip, T. Keshavarz, and I. Roy, "Polyhydroxyalkanoates: biodegradable polymers with a range of applications," *Journal of Chemical Technology and Biotechnology*, vol. 82, no. 3, pp. 233–247, 2007.
- [3] G.-Q. Chen, "A microbial polyhydroxyalkanoates (PHA) based bio- and materials industry," *Chemical Society Reviews*, vol. 38, no. 8, pp. 2434–2446, 2009.
- [4] S. Obruca, P. Benesova, J. Oborna, and I. Marova, "Application of protease-hydrolyzed whey as a complex nitrogen source to increase poly(3-hydroxybutyrate) production from oils by *Cupriavidus necator*," *Biotechnology Letters*, vol. 36, no. 4, pp. 775–781, 2014.
- [5] J. G. da Cruz Pradella, J. L. Ienczak, C. R. Delgado, and M. K. Taciro, "Carbon source pulsed feeding to attain high yield and high productivity in poly(3-hydroxybutyrate) (PHB) production from soybean oil using *Cupriavidus necator*," *Biotechnology Letters*, vol. 34, no. 6, pp. 1003–1007, 2012.
- [6] M. T. Cesário, R. S. Raposo, M. C. M. D. de Almeida, F. van Keulen, B. S. Ferreira, and M. M. R. da Fonseca, "Enhanced bioproduction of poly-3-hydroxybutyrate from wheat straw lignocellulosic hydrolysates," *New Biotechnology*, vol. 31, no. 1, pp. 104–113, 2014.
- [7] D. Pleissner, W. C. Lam, W. Han et al., "Fermentative polyhydroxybutyrate production from a novel feedstock derived from bakery waste," *BioMed Research International*, vol. 2014, Article ID 819474, 8 pages, 2014.
- [8] P. I. Nickel, M. J. Pettinari, M. A. Galvagno, and B. S. Méndez, "Poly(3-hydroxybutyrate) synthesis by recombinant *Escherichia coli arcA* mutants in microaerobiosis," *Applied and Environmental Microbiology*, vol. 72, no. 4, pp. 2614–2620, 2006.
- [9] P. I. Nickel, M. J. Pettinari, M. A. Galvagno, and B. S. Méndez, "Poly(3-hydroxybutyrate) synthesis from glycerol by a recombinant *Escherichia coli arcA* mutant in fed-batch microaerobic cultures," *Applied Microbiology and Biotechnology*, vol. 77, no. 6, pp. 1337–1343, 2008.
- [10] X.-X. Wei, Z.-Y. Shi, M.-Q. Yuan, and G.-Q. Chen, "Effect of anaerobic promoters on the microaerobic production of polyhydroxybutyrate (PHB) in recombinant *Escherichia coli*," *Applied Microbiology and Biotechnology*, vol. 82, no. 4, pp. 703–712, 2009.
- [11] J. Jian, S.-Q. Zhang, Z.-Y. Shi, W. Wang, G.-Q. Chen, and Q. Wu, "Production of polyhydroxyalkanoates by *Escherichia coli* mutants with defected mixed acid fermentation pathways," *Applied Microbiology and Biotechnology*, vol. 87, no. 6, pp. 2247–2256, 2010.
- [12] L. Beyer, C. Doberenz, D. Falke, D. Hunger, B. Suppmann, and R. G. Sawers, "Coordination of FocA and pyruvate formate-lyase synthesis in *Escherichia coli* demonstrates preferential translocation of formate over other mixed-acid fermentation products," *Journal of Bacteriology*, vol. 195, no. 7, pp. 1428–1435, 2013.
- [13] K. Sekar and K. E. Tyo, "Regulatory effects on central carbon metabolism from poly-3-hydroxybutyrate synthesis," *Metabolic Engineering*, vol. 28, pp. 180–189, 2015.