



# Quantitative assessment of pneumothorax by using Shannon entropy of lung ultrasound M-mode image and diaphragmatic excursion based on automated measurement

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**Background:** Lung ultrasound (LUS) and diaphragm ultrasound (DUS) are the appropriate modalities for conservative observation to those patients who are with stable pneumothorax, as well as for the timely detection of life-threatening pneumothorax at any location, due to they are portable, real-time, relatively cost effective, and most important, without radiation exposure. The absence of lung sliding on LUS M-mode images and the abnormality of diaphragmatic excursion (DE) on DUS M-mode images are the most common and novel diagnostic criteria for pneumothorax, respectively. However, visual inspection of M-mode images remains subjective and quantitative analysis of LUS and DUS M-mode images are required.

**Methods:** Shannon entropy of LUS M-mode image ( $\text{ShanEn}_{\text{LM}}$ ) and DE based on the automated measurement ( $\text{DE}_{\text{AM}}$ ) are adapted to the objective pneumothorax diagnoses and the severity quantifications in this study. Mild, moderate, and severe pneumothoraces were induced in 24 male New Zealand rabbits through insufflation of room air (5, 10 and 15, and 25 and 40 mL/kg, respectively) into their pleural cavities. *In vivo* intercostal LUS and subcostal DUS M-mode images were acquired using a point-of-care system for estimating  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$ .

**Results:**  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  as functions of air insufflation volumes exhibited U-shaped curves and were exponentially decreasing, respectively. Either  $\text{ShanEn}_{\text{LM}}$  or  $\text{DE}_{\text{AM}}$  had areas under the receiver operating characteristic curves [95% confidence interval (CI)] of 1.0000 (95% CI: 1.0000–1.0000), 0.9833 (95% CI: 0.9214–1.0000), and 0.9407 (95% CI: 0.8511–1.0000) for differentiating between normal and mild pneumothorax, mild and moderate pneumothoraces, and moderate and severe pneumothoraces, respectively.

**Conclusions:** Our findings imply that the combination of  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  give the promising potential for pneumothorax quantitative diagnosis.

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**Keywords:** Pneumothorax; lung ultrasound M-mode image (LUS M-mode image); Shannon entropy (ShanEn); diaphragm ultrasound (DUS); diaphragmatic excursion (DE)

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## Introduction

Pneumothorax is defined as the presence of excess air in the pleural cavity which caused by unknown reason, other lung diseases, trauma, or medical intervention (1-3). It may hinder lung inflation, deflation and gas exchange due to its high pleural pressure. Its clinical manifestations can express from asymptomatic to life threatening (1-3). For patients with stable pneumothorax, current clinical guidelines recommend the aggressive treatments (e.g., chest drainage) as first-line treatment; however, many studies have reported that conservative observation at the outpatient clinic until syndrome released by themselves has similar therapeutic outcomes as aggressive treatments while being more cost-effective and hospitalization exemption (3,4). In addition, timely detection and treatment can be life-saving for those tension pneumothorax or dyspnea patients who are irrespective of etiology, and even in the prehospital setting (2,3). Although pneumothorax diagnosis based on pleural manometry is intuitive, it has not received widespread clinical acceptance because of invasion nature and high interpatient variability (5-7). Computed tomography (CT) and chest X-ray (CXR) are the gold standard and primary methods, respectively, for confirming pneumothorax (1,2), with several drawbacks such as radiation exposure and incapable to perform serial images or for prehospital or bedside scanning.

Lung ultrasound (LUS) can overcome the limitations of CT and CXR for pneumothorax diagnosis. Because of the lack of radiation and low cost, LUS can be performed repeatedly without causing harm to the patients and clinicians, and budget overload. Moreover, its portability and real-time assessment capability enable clinicians to detect and monitor pneumothorax at any location—from the operating room to the bedside to outside the hospital. The LUS diagnostic criteria for pneumothorax include the presence of lung point and lung pulse, and the absence of lung sliding and A-line, with the absence of lung sliding being the most common criteria (2,8,9). Nevertheless, it is a great challenge to interpret lung sliding compared to identify it since the changes in lung sliding on traditional

LUS B-mode images may be subtle from normal to severe pneumothoraces. The M-mode image records the successive positions of a structure over time and captures a still image of this dynamic process, thereby improving visualization and providing more evidence of lung sliding (10,11). On LUS M-mode images, the seashore sign indicates lung sliding and excludes pneumothorax, whereas the stratosphere or barcode sign reflects the absence of lung sliding and indicates the presence of pneumothorax (11-13). However, visual inspection of LUS M-mode images depends on the operator's skill and experience and is subjective (14,15). To assist novice operators and alleviate the workloads of the experts, objective information obtained from the quantitative analysis of the LUS M-mode image is useful and helpful. Deep learning-based methods are powerful tools for detection of lung sliding and characterization of pneumothorax (16,17). However, requirement of large training data with labels, inconsistent system settings, scanning protocols, and annotations, and most importantly, unclear underlying mechanisms limited their use in clinical. Methods based on the occurrence probability of pixels with different brightness values are one of the most widely used for ultrasound quantification. They have been successfully applied for monitoring the progression of Duchenne muscular dystrophy (18) and liver ablation (19); detecting cataracts (20) and middle ear effusion (21); characterizing thrombi (22); and evaluating thrombolysis (23) and hepatic steatosis (24,25). Unfortunately, raw data (i.e., radiofrequency or envelope signals) are required for the methods used in these studies, and not every ultrasound system outputs the raw data of M-mode images. Thus, a quantitative method based on the occurrence probability of standard-output LUS M-mode images can enhance the applicability of this objective method for pneumothorax diagnosis.

In information theory, Shannon entropy (ShanEn) is a metric used for measuring uncertainty, and it is proportional to the occurrence probability of message content. ShanEn is the highest when the message is totally unpredictable and decreases with content predictability (26).

Pixels with different brightness values result in different visual appearances of images, representing the uncertainty and occurrence probability of messages carried by images. Therefore, ShanEn based on occurrence probability can quantify the visual appearance of an image. In addition, ShanEn estimation can be made using any type of image—raw data, grayscale, or others—because ShanEn only considers the occurrence of pixel brightness, irrespective of the underlying origins (27-29). Accordingly, the ShanEn of LUS M-mode image (ShanEn<sub>LM</sub>) can help detect and quantify pneumothorax.

The diaphragm is a dome-shaped muscle and is the main source of pressure changes in the pleural cavity during respiration. It contracts and descends during inspiration, decreasing pleural pressure and thus allowing air to flow into the lungs, whereas it relaxes and ascends during expiration, increasing pleural pressure and thus causing air to flow out of the lungs (30,31). The position difference between diaphragmatic contraction and relaxation on diaphragm ultrasound (DUS) M-mode images is known as diaphragmatic excursion (DE) (32,33). DE may be abnormal in cases of diaphragmatic dysfunction, such as during spinal cord injury (34,35), muscular dystrophy (34,36), and diaphragmatic paralysis (34,37). Pneumothorax can also cause diaphragmatic dysfunction because high pleural pressure may push the diaphragm away from its normal position. Thus, similar to the use of ShanEn<sub>LM</sub>, DE based on automated measurement (DE<sub>AM</sub>) instead of manual measurement for pneumothorax quantification is worth exploring.

In this study, we implemented ShanEn<sub>LM</sub> and DE<sub>AM</sub> as the quantitative methods for the objective diagnosis of rabbit pneumothorax to validate our assumptions. We present this article in accordance with the STARD and ARRIVE reporting checklists (available at <https://qims.amegroups.com/article/view/10.21037/qims-23-636/rc>).

## Methods

### *Animal preparation*

The experimental protocols of this study were approved by the Institutional Animal Care and Use Committee of China Medical University (CMUIACUC-2020-381), in compliance with the Taiwan Council of Agriculture's Guideline for the Care and Use of Laboratory Animals. The animals, housing and experimental environments were not specific pathogen-free. Twenty-four male New Zealand

rabbits weighing 2.7–3.1 kg (age: fifteen weeks) were used according to Hill *et al.* (38). They were housed in air-conditioned cages with free access to laboratory chow and drinking water, and conventional cleaning. Randomization was not blinded in this study. Animals assigned to lab by animal house staff with no prior knowledge of experiment to be performed. After acclimatization for 1 week, the rabbits were all healthy according to regular eating and toileting, bright eyes, no significant discharge coming from nose, and normal coat without scurf, parasites, and fleas (39).

### *Experimental protocols*

The experiment was conducted in the operation room of China Medical University animal center after environmental sterilization by 75% ethanol. We used the pneumothorax induction methods by Hill *et al.* (38) and Summers *et al.* (40) for rabbits, with some modifications. Briefly, following an 8-h fast, each rabbit was anesthetized using 30 mg/kg pentobarbital sodium (Sigma-Aldrich, St. Louis, MO, USA). A sterile 16-gauge needle was inserted into the pleural cavity 1 cm below the right scapular tip and was connected to a 50-mL syringe and the homemade U-tube manometer through a three-way stopcock and flexible polyethylene tubes without respiratory monitoring and support by ventilator. Pneumothorax was induced only on the right side to minimize the interference from the beating heart during LUS scanning and to maximize the DUS acoustic window through the liver rather than through the spleen (31,41). Room air was insufflated into the pleural cavity at 5, 5, 5, 10, and 15 mL/kg sequentially to create excess air volumes of 5, 10, 15, 25, and 40 mL/kg, respectively. Immediately after each insufflation, LUS, DUS, and pleural pressure measurements with the homemade U-tube manometer were performed. Each insufflation only took a few seconds. However, the following scanning and measurements took 10 to 15 minutes.

A point-of-care system (LU700C, Leltek, Taipei) was employed for scanning of LUS and DUS. Before scanning, the upper bodies of the rabbits were shaved, and they were placed in the left lateral position. Transverse LUS scanning along the right midclavicular line was used to visualize the right lung surface. The M-mode interrogation line was adjusted to be perpendicular to the lung surface. We recorded 17-s M-mode images of the second to sixth intercostal spaces. Image depth, gain, dynamic range, central frequency, frame rate, pulse length, mechanical index, and thermal index were 3.2 cm, 50, 40, 3.6 MHz,

15 frames per second, 2.13 mm, 0.6, and 0.42, respectively. The synthetic transmit focusing of the plane waves (i.e., total focusing imaging without focus zone) was used without time-gain compensation. The unfavorable effects of motions on scans were minimized by manual operation of handheld transducer based on operator's skill and experience. The scan was abandoned if the quality was not good enough according to operator's judgments. Five qualified scans of each rabbit were saved. Transverse DUS scanning of the right subcostal area between the midsternal and anterior axillary lines was used to visualize the right diaphragmatic dome. The M-mode interrogation line was adjusted to be perpendicular to the diaphragmatic dome. Again, 17-s M-mode images were recorded. The scanning settings of DUS were the same as those of LUS, except for depth, gain, and dynamic range. To visualize the diaphragm at a deeper position, the image depth was increased to 4.7 cm. To simplify the automated measurement of DE by emphasizing the hyperechoic signal of the diaphragm through the elimination of the hypoechoic signals of its reverberations, other tissues, and noises, we set a lower gain and dynamic range: 30 and 20, respectively. One qualified scans of each rabbit were saved. The LUS and DUS M-mode grayscale images after beamforming and focusing by ultrasound system were all output in the .jpg format, with a resolution of 1,328×530. During pleural manometry, because of the oscillations of the fluid column caused by pressure swings during respiration, and open end of the homemade U-tube manometer, only average pleural pressure values relative to 1 atmosphere were obtained. Twenty measurements of each rabbit with each air insufflation volumes were taken. After LUS and DUS scanning, and pleural pressure measurements, all rabbits were immediately euthanized with an overdose of pentobarbital sodium. No rabbits died during the experiment, indicating the acceptable risk of our protocols without ventilator use.

### *ShanEn<sub>LM</sub> and DE<sub>AM</sub> estimations*

Programs for estimating ShanEn<sub>LM</sub> and DE<sub>AM</sub> were developed using MATLAB R2016a (MathWorks, Natick, MA, USA, RRID:SCR\_001622). For each LUS M-mode image, the rectangular region of interest (ROI) was manually selected. The ShanEn<sub>LM</sub>  $H$  can be estimated as follows (26):

$$H \equiv -\int_{y_{\min}}^{y_{\max}} \omega(y) \log_2 \omega(y) \quad [1]$$

where  $y_{\min}$  and  $y_{\max}$  are the minimal and maximal pixel

brightness of ROI, respectively, and  $\omega(y)$  represents the occurrence probability of pixel with each brightness. Although the optimal logarithm base of ShanEn remains unclear, we used a base of 2, following the first choice in previous studies (27-29).

For each DUS M-mode image, DE<sub>AM</sub> was estimated using the following steps (Figure 1):

- (I) A binary DUS M-mode image was binarized using Otsu's method.
- (II) The upper boundary between black and white regions was detected based on the first brightness transition between 0 and 1 from top to bottom.
- (III) The local extremum of the boundary was obtained. The minimum peak separation was set to 0 (the default value of MATLAB software). Therefore, the window size of local extremum calculation was the length of whole boundary.
- (IV) The extrema values above the third quartile were regarded as the positions of the relaxed diaphragms.
- (V) The line connecting the positions of the contracted diaphragms was acquired through the linear fitting of extrema values below the first quartile based on the assumption of the same position of the diaphragmatic contraction in each respiratory cycle.
- (VI) DE<sub>AM</sub> was defined as the average of the differences between the positions of relaxed diaphragms and their corresponding positions on the fitting line.

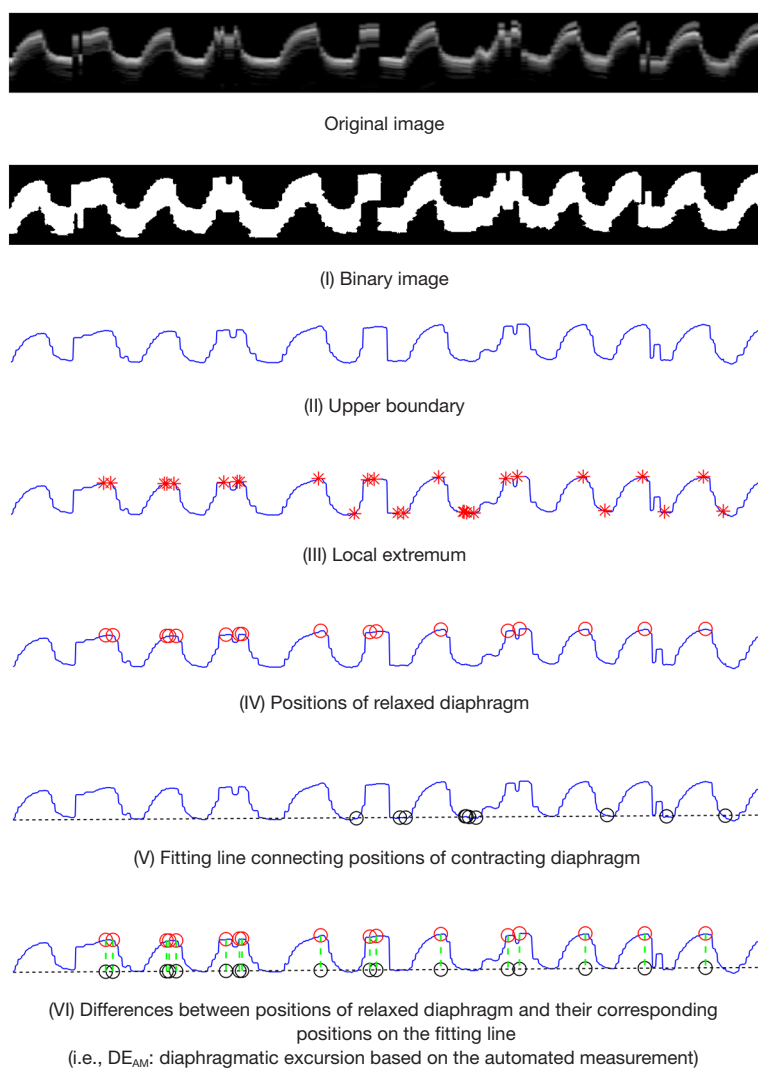
### *Statistical analysis*

All plots and statistical analyses were performed using SigmaPlot software (Version 12.0, Systat Software, Inc., Chicago, IL, USA, RRID:SCR\_003210). Because our lab lacked the dedicated pleural manometer as references, the accuracy of the homemade U-tube manometer was estimated using measurements themselves. We treated the average of 20 measurements and each measurement of pleural pressures as true and observation values of each rabbit with each air insufflation volume, respectively. The accuracy of each measurement using the homemade U-tube manometer  $A$  can be estimated based on relative errors  $E_r$ , as follows (42,43):

$$A = (1 - E_r) \times 100\% \quad [2]$$

$$E_r = \left| \frac{x_i - x_t}{x_t} \right| \quad [3]$$

where  $x_i$  and  $x_t$  are observation (i.e., each measurement) and true (i.e., the average of 20 measurements) values,



**Figure 1** Flowchart for estimating  $DE_{AM}$ .  $DE_{AM}$ , diaphragmatic excursion based on automated measurement.

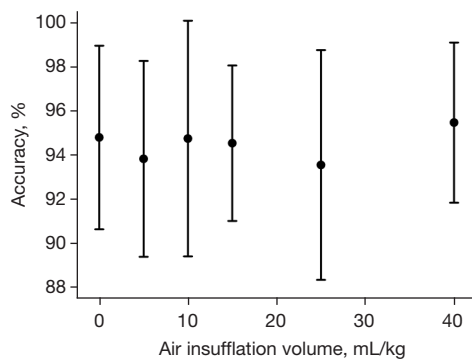
respectively. The accuracy of each rabbit with each air insufflation volumes was obtained from the average of 20 measurements. The accuracy of each air insufflation volume was obtained from the average of 24 rabbits and expressed as scatter plots with mean  $\pm$  standard deviation (SD). The pleural pressures of each rabbit with different air insufflation volumes were the average results of 20 measurements and illustrated using line and scatter plots with mean  $\pm$  SD.  $ShanEn_{LM}$  of each rabbit was the average of five LUS M-mode scans.  $ShanEn_{LM}$  and  $DE_{AM}$  as functions of air insufflation volumes were expressed as scatter plots with mean  $\pm$  SD and curve fitting. The distributions of  $ShanEn_{LM}$  and  $DE_{AM}$  with different air insufflation volumes were derived using scatter plots, respectively, and presented

as mean  $\pm$  SD. Because the severity of pneumothorax has not been defined with a consensus (44), pneumothorax with air insufflation volumes of 5, 10 and 15, and 25 and 40 were denoted as mild, moderate, and severe, respectively, according to data distribution. Receiver operating characteristic curves (ROCs) illustrated performances of  $ShanEn_{LM}$  and  $DE_{AM}$  for differentiating normal and mild pneumothorax, mild and moderate pneumothoraces, and moderate and severe pneumothoraces. The ROC curves were produced by plotting sensitivity [true positive/(true positive + false positive)] on the y-axis against 1-specificity [true negative/(false positive + true negative)] on the x-axis under varying thresholds. Areas under the ROCs (AUROCs) with 95% confidence interval (CI) and ROC

indices, including cut off value, positive predictive value [true positive/(true positive + false positive)], negative predictive value [true negative / (true negative + false negative)], and accuracy [(true positive + true negative) / (positive + negative)] were estimated.

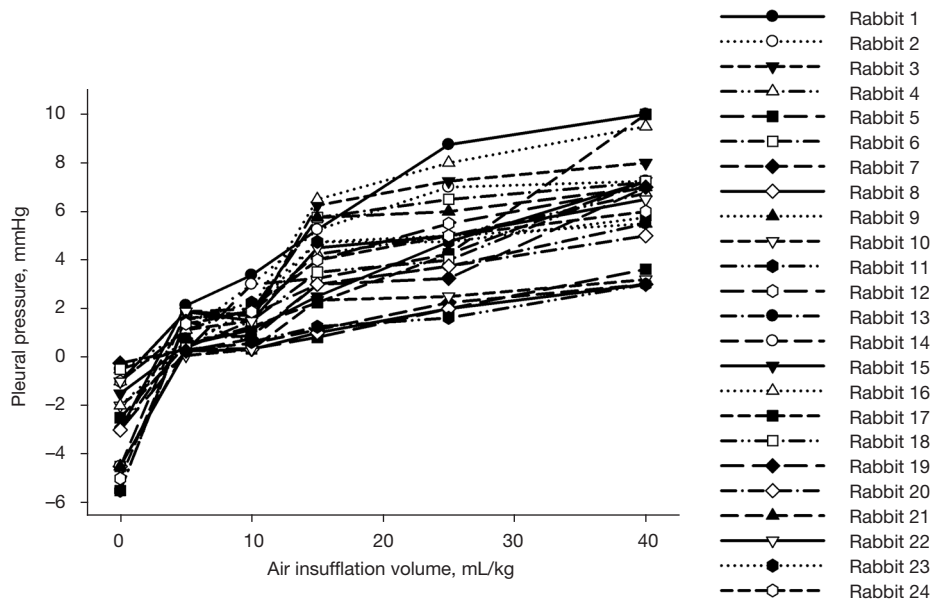
**Results**

Figure 2 shows the accuracies with different air insufflation volumes. They had similar performances. Figure 3 depicts each rabbit’s pleural pressure with different air insufflation

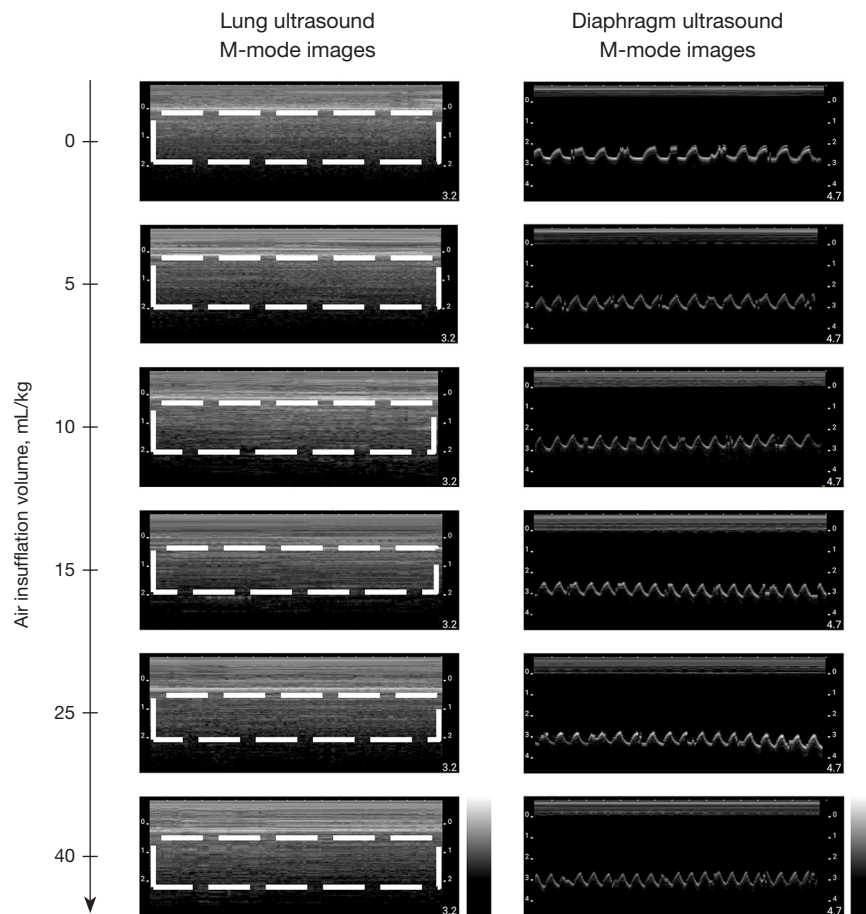


**Figure 2** Accuracies with different air insufflation volumes. They had similar performances.

volumes. The all increased with air insufflation volumes and exhibited high interindividual variability. Figure 4 presents the representative LUS and DUS M-mode images of different air insufflation volumes. The changes of them between air insufflation volumes may be subtle and undetectable by the naked eye. In LUS M-mode images, as the air insufflation volumes increased from 0 to 15 mL/kg, the visual appearances of the subpleural region changed from the granular pattern to parallel lines, being almost restored to the granular pattern at volumes >25 mL/kg. In DUS M-mode images, DE decreased significantly as the air insufflation volumes increased from 0 to 10 mL/kg and remained stable thereafter. ShanEn<sub>LM</sub> and DE<sub>AM</sub> results supported the qualitative findings in Figure 4. Figure 5A,5B illustrate the U-shaped and exponentially decreasing fitting curves of ShanEn<sub>LM</sub> and DE<sub>AM</sub> as functions of air insufflation volumes, respectively. The determination coefficients ( $r^2$ ) between them and air insufflation volumes were 0.6205 and 0.8506, respectively. Figure 5C illustrates the distributions of ShanEn<sub>LM</sub> and DE<sub>AM</sub> with different air insufflation volumes based on scans of the same 24 rabbits induced by sequential insufflation of room air. ShanEn<sub>LM</sub> could differentiate between mild (i.e., air insufflation volume of 5) and moderate (i.e., air insufflation volumes of 10 and 15) pneumothoraces, and moderate and severe (i.e., air insufflation volumes of 25 and 40) pneumothoraces. In contrast, DE<sub>AM</sub> could



**Figure 3** Pleural pressure of each rabbit with different air insufflation volumes. They all increased with air insufflation volumes and showed high interindividual variability.



**Figure 4** Representative M-mode images of LUS and DUS with different air insufflation volumes. The subtle changes of subpleural region visual appearances of LUS M-mode images were from granular pattern to parallel lines and nearly restored to the granular pattern with air insufflation volumes from 0 to 15 mL/kg, and above 25 mL/kg, respectively. Diaphragmatic excursion of DUS M-mode images decreased significantly and remained stable with air insufflation volumes from 0 to 10 mL/kg, and above 10 mL/kg, respectively. The white dashed rectangles were regions of interest.  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  as functions of air insufflation volumes were expressed as scatter plots with mean  $\pm$  SD and curve fitting. The distributions of  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  with different air insufflation volumes were derived using scatter plots, respectively. The U-shaped and exponentially decreasing fitting curves of  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  as functions of air insufflation volumes, respectively. LUS, lung ultrasound; DUS, diaphragm ultrasound;  $\text{ShanEn}_{\text{LM}}$ , Shannon entropy of lung ultrasound M-mode image;  $\text{DE}_{\text{AM}}$ , diaphragmatic excursion based on automated measurement; SD, standard deviation.

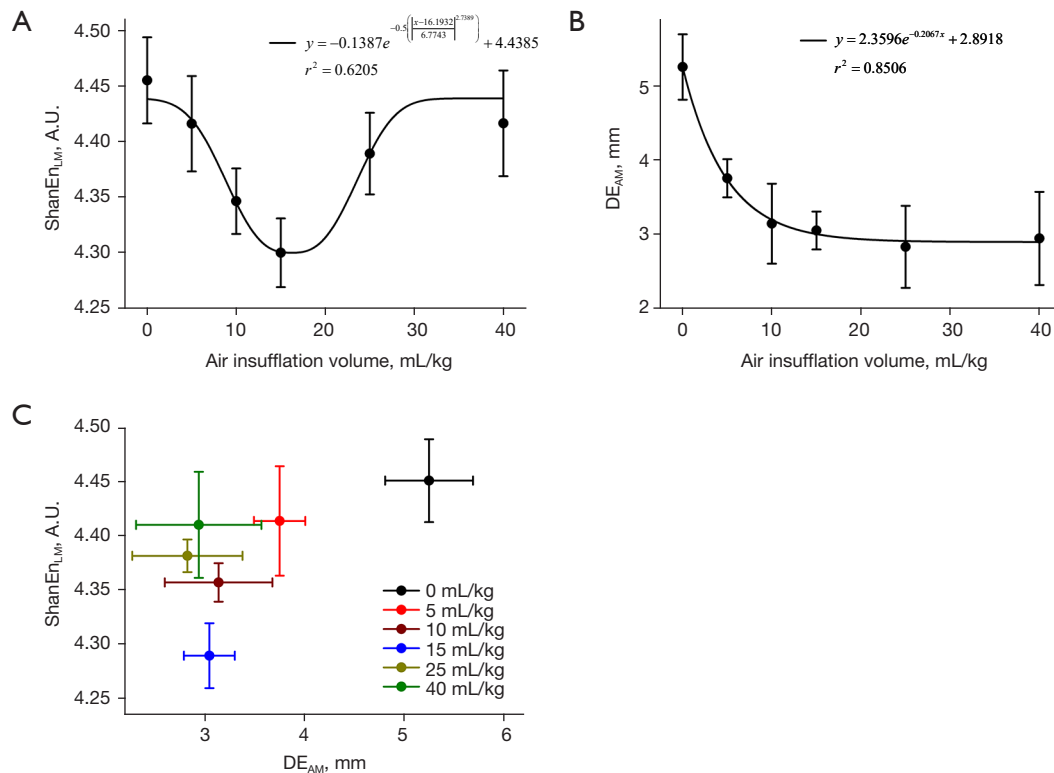
differentiate between normal and mild pneumothorax, and mild and moderate pneumothoraces. *Table 1* summarizes the AUROCs and ROC indices of  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  for differentiating between normal and mild pneumothorax, mild and moderate pneumothoraces, and moderate and severe pneumothoraces. Either  $\text{ShanEn}_{\text{LM}}$  or  $\text{DE}_{\text{AM}}$  had AUROCs of 1.0000 (95% CI: 1.0000–1.0000), 0.9833 (95% CI: 0.9214–1.0000), and 0.9407 (95% CI: 0.8511–1.0000) for differentiating between normal and mild pneumothorax, mild and moderate pneumothoraces, and moderate and

severe pneumothoraces, respectively, indicating the effectiveness of combination of  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$  for pneumothorax quantification.

## Discussion

### Study significance

This study explored the feasibility of quantifying pneumothorax by using  $\text{ShanEn}_{\text{LM}}$  and  $\text{DE}_{\text{AM}}$ . Pneumothorax was induced in rabbits by air insufflation through a needle



**Figure 5** ShanEn<sub>LM</sub> and DE<sub>AM</sub> with different air insufflation volumes, respectively. (A) ShanEn<sub>LM</sub> as a function of air insufflation volumes. The correlation between them was U-shaped according to curve fitting. The determination coefficient ( $r^2$ ) between them was 0.6205. (B) DE<sub>AM</sub> as a function of air insufflation volumes. The correlation between them decreased exponentially according to curve fitting. The determination coefficient ( $r^2$ ) between them was 0.8506. (C) The distributions of ShanEn<sub>LM</sub> and DE<sub>AM</sub> with different air insufflation volumes. ShanEn<sub>LM</sub> could differentiate between mild (i.e., air insufflation volume of 5) and moderate (i.e., air insufflation volumes of 10 and 15) pneumothoraces, and moderate and severe (i.e., air insufflation volumes of 25 and 40) pneumothoraces. In contrast, DE<sub>AM</sub> could differentiate between normal and mild pneumothorax, and mild and moderate pneumothoraces. ShanEn<sub>LM</sub>, Shannon entropy of lung ultrasound M-mode image; DE<sub>AM</sub>, diaphragmatic excursion based on automated measurement.

inserted into the pleural cavity. The pleural pressure measured using the homemade U-tube manometer was planned to be the reference for indicating pneumothorax severity. Although increasing pleural pressure indicates the successful induction of pneumothorax, high interindividual variability renders it a qualitative description rather than a quantitative reference as the same as it measured in clinical (5-7). Therefore, we replaced pleural pressure with air insufflation volumes because of the positive correlation between them of each rabbit. Our results revealed that ShanEn<sub>LM</sub> and DE<sub>AM</sub> exhibited U-shaped curves and exponentially decreasing correlations, respectively, with air insufflation volumes. Furthermore, the AUROCs of either ShanEn<sub>LM</sub> or DE<sub>AM</sub> for differentiating between normal and mild pneumothorax (i.e., air insufflation volume of 5),

mild and moderate (i.e., air insufflation volumes of 10 and 15) pneumothoraces, and moderate and severe (i.e., air insufflation volumes of 25 and 40) pneumothoraces were 1.0000 (95% CI: 1.0000–1.0000), 0.9833 (95% CI: 0.9214–1.0000), and 0.9407 (95% CI: 0.8511–1.0000), respectively. The U-shaped curve between ShanEn<sub>LM</sub> and air insufflation volumes had better ability to differentiate between moderate and severe pneumothoraces. In contrast, the exponentially decreasing correlations with air insufflation volumes of DE<sub>AM</sub> had better ability to differentiate between normal and mild pneumothorax, and mild and moderate pneumothoraces. These findings demonstrate that ShanEn<sub>LM</sub> and DE<sub>AM</sub> provide objective information of pneumothorax, in particularly to combine both methods.



**Table 1** Receiver operating characteristic curve indices of ShanEn<sub>LM</sub> and DE<sub>AM</sub> for differentiating between mild, moderate, and severe pneumothoraces

Parameters/indices	Cut off value	AUROC (95% CI)	Sensitivity (%)	Specificity (%)	PPV (%)	NPV (%)	Accuracy (%)
<b>ShanEn<sub>LM</sub></b>							
Normal and mild	4.4220	0.7292 (0.4840–0.9735)	75.0000	75.0000	66.6667	81.8182	75.0000
Mild and moderate	4.3514	0.9750 (0.9135–1.0000)	86.6667	100.0000	100.0000	80.0000	91.3043
Moderate and severe	4.3514	0.9407 (0.8511–1.0000)	86.6667	94.4444	92.8571	89.4737	90.9090
<b>DE<sub>AM</sub></b>							
Normal and mild	4.2183	1.0000 (1.000–1.0000)	100.0000	100.0000	100.0000	100.0000	100.0000
Mild and moderate	3.4618	0.9833 (0.9214–1.0000)	90.0000	100.0000	100.0000	85.7143	93.7500
Moderate and severe	2.8611	0.6000 (0.3278–0.8722)	66.6667	70.0000	66.6667	70.0000	68.4211

ShanEn<sub>LM</sub>, Shannon entropy of lung ultrasound M-mode image; DE<sub>AM</sub>, diaphragmatic excursion based on automated measurement; AUROC, area under the receiver operating characteristics curve; CI, confidence interval; Sensitivity, [true positive/(true positive + false negative)]; Specificity, [true negative/(false positive + true negative)]; PPV, positive predictive value [true positive/(true positive + false positive)]; NPV, negative predictive value [true negative/(true negative + false negative)]; Accuracy, [(true positive + true negative)/(positive + negative)].

### Effects of pneumothorax on ShanEn<sub>LM</sub>

Lung sliding refers to the cyclic to-and-fro movements of the pleural line (i.e., relative motions between the parietal and visceral pleurae) caused by lung inflation and deflation (8-11). Above the pleural line, the soft tissues of the chest wall did not affect by the lung movement and are almost static; they thus generate parallel lines on LUS M-mode images because of reverberations, which appear like waves approaching the beach. Moving pleural line, superficial lung structures and their reverberations change positions over time and create a granular pattern on LUS M-mode images, appearing like sand on beach. These two appearances together constitute the seashore sign of LUS M-mode images, indicating the presence of lung sliding and the absence of pneumothorax (2,12,13). High pleural pressure of pneumothorax separates the two pleurae. The air beneath the parietal pleura almost completely reflects the ultrasound waves, thus preventing the visualization of visceral pleura and superficial lung structures. Consequently, relative motions between two pleurae and resulting lung sliding are lost; the sand-like appearance is replaced by parallel lines, identical to that observed for tissues above the pleural line. This appearance is termed as the stratosphere or barcode sign; this indicates the absence of lung sliding and the presence of pneumothorax (2,12,13). Because the appearances above the pleural line on LUS M-mode images are similar between normal lungs and lungs with pneumothorax, this study paid more attention to the changes in subpleural appearances.

For ShanEn<sub>LM</sub> estimation, the granular pattern has many pixels with different brightness values and corresponding high uncertainty. However, parallel lines only have white (i.e., lines themselves) and black (i.e., background) pixels. Thus, their uncertainty is smaller than that of the granular pattern. That ShanEn<sub>LM</sub> decreased with increasing pneumothorax severity is reasonable; however, the opposite was observed from moderate to severe pneumothorax. This finding can be explained as follows. First, the high pleural pressure in severe pneumothorax triggers intercostal muscle retraction for maintaining normal respiratory function. In the end, it further causes chest partial motion along the pleural line, which may be mistaken for normal lung sliding on LUS M-mode images (40). Second, our experimental setup may be another reason. We insufflated air into the pleural cavity from the posterior zone. The high air volume of severe pneumothorax may push the lung toward the chest wall and disrupt the air layer that separates the two pleurae during mild and moderate pneumothoraces, leading to restoration of the visualization of the visceral pleura—and lung sliding—in severe pneumothorax. Consequently, the corresponding uncertainty and ShanEn<sub>LM</sub> of severe pneumothorax may increase.

### Effects of pneumothorax on DE<sub>AM</sub>

The decreasing DE<sub>AM</sub> with pneumothorax severity in our study was similar to that with pleural effusion in previous studies (45,46). External pressure on the diaphragm, such as that exerted during pneumothorax and pleural effusion, may

push it caudally and reduce its length. Due to a contracted diaphragm is harder and more resistant to pressure than a relaxed diaphragm, the downward displacement of the contracted diaphragm becomes smaller than that of the relaxed diaphragm as the pleural cavity pressure increases (from normal to pneumothorax). Additionally, the shortened diaphragm is no longer the optimal length for force generation and has impaired contraction capacity against pressure according to the force-length relationship of muscle (47-49). Thus, the smaller position differences between the contracted and relaxed diaphragms during respiration (i.e., decreased  $DE_{AM}$ ) are observed.

### *Limitations, and future challenges and studies*

First, despite the promising performance of the combination of  $ShanEn_{LM}$  and  $DE_{AM}$  for differentiating pneumothorax severity, the correlation between  $ShanEn_{LM}$  and air insufflation volumes was U-shaped instead of a monotonic decrease as per the theoretical hypothesis. Future studies should include a technique unaffected by intercostal muscle retraction in severe pneumothorax and our method of air insufflation to validate our results. Second, DUS M-mode images has parameters other than DE, such as cycle and velocity of diaphragmatic contraction (30,34), which we did not evaluate. Future studies should explore their quantifications and performances in pneumothorax diagnosis. Third, although we used animal model rather than clinical data for concept validation of our proposed method in this study, the employment of commercial ultrasound system, widely available image types to  $ShanEn_{LM}$  and  $DE_{AM}$ , the anatomical, physiological, and biochemical similarities of lung between rabbit and human (50,51) make our proposed method easily translate to human *in vivo* applications. Recruitment of patients with pneumothorax is the next step of our group. Fourth, although the absence of lung sliding is the most common diagnostic criterion of pneumothorax (2,8,9), the presence of some other lung conditions (e.g., chronic obstructive pulmonary disease and large parenchymal tumor) may also have an absent lung sliding and confuse clinicians (1-3,52). Differentiating pneumothorax from these conditions based on our proposed method may be challenging, fortunately, using the bedside LUS in emergency (BLUE) protocol (53-55), the detection of other parameters, such as the B-line artifact, consolidation, and lung pulse, can help overcome this limitation. The integration of the BLUE protocol,  $ShanEn_{LM}$ , and  $DE_{AM}$  is the future direction of our group.

### **Conclusions**

In this study,  $ShanEn$  of LUS M-mode images (i.e.,  $ShanEn_{LM}$ ) and DE based on the automated measurement of DUS M-mode images (i.e.,  $DE_{AM}$ ) was applied as new quantitative methods for the objective diagnosis of pneumothorax in a rabbit model. Our study provided two key findings: (I)  $ShanEn_{LM}$  and  $DE_{AM}$  had U-shaped and exponentially decreasing correlations with air insufflation volumes, respectively, and (II) either  $ShanEn_{LM}$  or  $DE_{AM}$  had AUROCs of 1.0000 (95% CI: 1.0000–1.0000), 0.9833 (95% CI: 0.9214–1.0000), and 0.9407 (95% CI: 0.8511–1.0000) for differentiating between normal and mild pneumothorax (i.e., air insufflation volume of 5), mild and moderate (i.e., air insufflation volumes of 10 and 15) pneumothoraces, and moderate and severe (i.e., air insufflation volumes of 25 and 40) pneumothoraces, respectively. The combination of  $ShanEn_{LM}$  and  $DE_{AM}$  has great potential for diagnosing and quantifying pneumothorax.

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### **Footnote**

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*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. This study was approved by the Institutional Animal Care

and Use Committee of China Medical University (CMUIACUC-2020-381), in compliance with the Taiwan Council of Agriculture's Guideline for the Care and Use of Laboratory Animals.

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