

Hydrogen potassium adenosine triphosphatase activity inhibition and downregulation of its expression by bioactive fraction DLBS2411 from *Cinnamomum burmannii* in gastric parietal cells

Raymond R Tjandrawinata^{1,2}
Florensia Nailufar²
Poppy F Arifin¹

¹Section of Molecular Pharmacology,
²Section of Animal Pharmacology,
Dexa Medica, Cikarang, Indonesia

Abstract: This study assessed the gastric acid antisecretory effect of DLBS2411 fractionated from *Cinnamomum burmannii*. Hydrogen potassium adenosine triphosphatase (H⁺/K⁺ ATPase) activity and its gene expression were observed, and the antioxidant activity of DLBS2411 was also investigated. Treatment of DLBS2411 decreased the level of H⁺/K⁺ ATPase messenger RNA expression on human embryonic kidney 293 cells and rat gastric parietal cells in a dose-dependent manner, in vitro and ex vivo. DLBS2411 also acted as a competitive inhibitor by showing inhibition in gastric H⁺/K⁺ ATPase activity at various pHs. In gastric ulcer animal models induced with indomethacin and ethanol, DLBS2411 showed a reduction in the number of petechiae, suggesting that the fraction also confers gastroprotective activity. Moreover, DLBS2411 was also found to have potent antioxidant activity. Taken together, DLBS2411 is a promising novel agent for the management of dyspepsia, a condition of hyperacidity and diseases in the stomach requiring gastroprotection.

Keywords: H⁺/K⁺ ATPase, proton-pump inhibitor, proton-pump downregulator, gastroprotective

Introduction

Gastrointestinal disorders such as dyspepsia and gastric and duodenal ulcers are a common diagnosis in the medical world. The diseases significantly affect millions of people worldwide and cause significant economic impact in terms of health care costs and work absenteeism, in addition to patients' decreased quality of life. Generally, these disorders are caused by a hypersecretion of gastric acid from parietal cells of gastric mucosa. This hyperacidic condition might be stimulated by caffeine, alcohol, such drugs as nonsteroidal anti-inflammatory drugs (NSAIDs), stress, and such bacteria as *Helicobacter pylori*.¹⁻³

The gastric enzyme hydrogen potassium adenosine triphosphatase (H⁺/K⁺ ATPase), also known as the proton pump, plays an important role in the acidification process in the stomach. This enzyme is found in parietal, kidney, and heart tissues.^{4,5} The inhibition of H⁺/K⁺ ATPase in gastric parietal cells reduces the overproduction of gastric acid.⁶ Therefore, a H⁺/K⁺ ATPase inhibitor (proton-pump inhibitor) could be used as a pharmacological target for drug development against the disturbances of gastric acid production.^{6,7} In addition to the disturbance of acid production, reactive oxygen species (ROS), especially the hydroxyl radical, may also play a major role in causing oxidative

Correspondence: Raymond R Tjandrawinata
Section of Molecular Pharmacology,
Research Innovation and Invention,
Dexa Laboratories of Biomolecular
Sciences, Dexa Medica, Jalan Industri
Selatan V – 7 Block PP, Jababeka
Industrial Estate II, Cikarang,
West Java 17550, Indonesia
Tel +62 21 8984 1901
Fax +62 21 8984 1905
Email raymond@dexa-medica.com

damage of mucosa.^{3,8} Excellent correlations exist between the production of ROS and tissue damage of gastric and intestinal mucosa.⁹

The current approach for gastrointestinal disorder therapy involves multiple mechanisms of pharmacological activities, including a gastric acid reducer, a gastroprotector, and an ROS scavenger. Currently, several pharmaceutical products, eg, histamine blockers and proton-pump inhibitors, are being used to manage gastric hyperacidity; however, these drugs also have limitations because of their side effects, such as headache, nausea, vomiting, and even nose inflammation.^{3,10–12} Development of a new drug, preferably that which comes from natural sources, with multiple pharmacological activities targeted toward alleviation of gastrointestinal disorders will therefore serve the unmet medical need.

We introduce a new natural bioactive fraction from *Cinnamomum burmannii* that can potentially be used as a pharmacological agent against gastric intestinal disorders, especially those related to hyperacidity. Our laboratory has in the past and is currently investigating the actions of natural products, some of which have subsequently been developed into agents to treat diseases related to internal medicine.^{13–15} We report here the activities of DLBS2411, a bioactive fraction of *C. burmannii*, as an H⁺/K⁺ ATPase downregulator and inhibitor as well as a gastroprotector.

Materials and methods

Chemicals

Minimum essential medium (MEM), Roswell Park Memorial Institute (RPMI) medium, fetal calf serum, and penicillin/streptomycin were purchased from Life Technologies (Carlsbad, CA, USA). Trizol reagent was also purchased from Life Technologies. Reverse-transcription reagents were purchased from Promega (Fitchburg, WI, USA). An EnzCheck[®] phosphate assay kit was bought from Life Technologies. Indomethacin was supplied by CSPC Ouyi Pharmaceutical (batch 2175080208; Shijiazhuang, People's Republic of China). Ethanol, ethidium bromide, 2,2-diphenyl-1-picrylhydrazyl (DPPH), ascorbic acid, trichloroacetic acid, potassium ferricyanide, and ferric chloride were obtained from Sigma-Aldrich (St Louis, MO, USA).

Plant material and preparation of bioactive fraction DLBS2411

Bark of *C. burmannii* was purchased from Padang, West Sumatra, Indonesia. This plant has been identified by the Herbarium Bogoriense, Research Center of Biology, Indonesian Institute

of Sciences with certificate 1261/IPH.1.02/If.8/XII/2009. The extraction process was started by maceration of the bark in different solvents. Maceration of the material in hot water at temperatures of 60°C–90°C for 1–2 hours was seen to be giving the most biological activity. Miscella was collected during the filtration process and evaporated by vacuum using a rotary evaporator at a temperature of 60°C–80°C to obtain concentrate. The concentrate was further processed through liquid–liquid extraction using dichloromethane at a ratio of 1:2 to separate from organic components. Subsequently, the water phase was collected and then evaporated using the rotary evaporator at temperatures of 50°C–120°C to obtain the dry extract. This dry extract was referred to as bioactive fraction DLBS2411 and then subjected to further molecular and biochemical analysis.

Tissue culture

Gastric parietal cells were isolated from the stomach of Wistar strain rats by collagenase digestion on fundic mucosa followed by enrichment of cells, as described by Chew et al.¹⁶ The parietal cell preparation contained approximately 1×10^7 cells/well in six-well plates. Human embryonic kidney (HEK)293 cells were purchased from the American Type Culture Collection (Manassas, VA, USA). This cell line was also shown to express H⁺/K⁺ ATPase gene. HEK293 cells were cultured in MEM and gastric parietal cells in RPMI medium supplemented with 10% serum and 1% antibiotic penicillin/streptomycin in six-well plates. The media were incubated at 37°C, 5% CO₂ for 24 hours. Cell medium was refreshed every 2–3 days. A subconfluent monolayer of cells was used in all experiments. Prior to experimentation, the cell medium was changed to that without serum and incubated for 18–24 hours before treatment. HEK293 and gastric parietal cells were treated with DLBS2411 in various concentrations: 10 µg/mL, 25 µg/mL, and 50 µg/mL. Each cell in the medium grown without serum was treated with DLBS2411 for 24 hours.

RNA isolation and reverse-transcription polymerase chain reaction

Total RNA was extracted from cells using Trizol reagent according to the manufacturer's instructions. RNA was determined for concentration and purity using a spectrophotometer at 260 and 280 nm (Bio-Rad, Hercules, CA, USA). The integrity of the RNA was verified using gel electrophoresis to detect the 18S and 28S ribosomal bands.

Before reverse-transcription (RT) reaction, RNA was incubated at 65°C for 10 minutes. One microgram aliquot

of total RNA was reverse-transcribed with 20 U RNasin® (Promega, Fitchburg, WI, USA), 25 mM deoxyribonucleotide triphosphate mix, 0.5 ng Oligo dT, and 5 U avian myeloblastosis virus reverse transcriptase (RT). The reaction mixture was incubated at 30°C for 10 minutes, 45°C for 45 minutes, 99°C for 5 minutes, and 6°C for 5 minutes. Polymerase chain reaction (PCR) was performed using specific primers for H⁺/K⁺ ATPase (forward 5' GCT GCA GCT CCA TCC TTA TC 3'; reverse 5' AGG CGG GTA GTC CTT CTC AT 3'). PCR products were visualized by ethidium bromide staining after agarose gel electrophoresis, and the result was quantified using ChemiDoc™ (Bio-Rad).

In vitro H⁺/K⁺ ATPase activity assay

The effect of DLBS2411 as inhibitor was observed on H⁺/K⁺ ATPase enzyme activity wherein the assay was based on the assessment of the inorganic phosphate released from the hydrolysis of ATP. This assay was done using the Enzcheck phosphate assay kit (Life Technologies) according to the manual available from the kit. The enzyme assay was done in gastric parietal cells that had been isolated from Wistar rats and cultured with addition of 30 µg/mL DLBS2411, and the pH levels during the assay were varied: 7.4, 4, and 2. This enzyme-activity study was done with and without the addition of DLBS2411.

Free radical scavenging activity

The antioxidant activity of DLBS2411 on the basis of the scavenging activity of the stable DPPH free radical was determined using the method described by Brand-Williams et al.¹⁷ DPPH solution (0.1 mM) in methanol was prepared, and 1.0 mL of this solution was added to 3.0 mL of DLBS2411 solution at different concentrations (0–50 µg/mL). Thirty minutes later, the absorbance was measured at 517 nm. A blank was prepared without addition of DLBS2411. Ascorbic acid at various concentrations (0–50 µg/mL) was used as standard. The lower absorbance of the reaction mixture indicates higher free radical scavenging activity. The capability to scavenge the DPPH radical was calculated using the following equation:

$$\text{DPPH scavenged (\%)} = \frac{A_{\text{control}} - A_{\text{test}}}{A_{\text{control}}} \times 100$$

where A_{control} is the absorbance of the control reaction and A_{test} is the absorbance in the presence of the sample of the fractions. The antioxidant activity of DLBS2411 was expressed as DPPH scavenged (%).

Reducing-power ability

The reducing power of DLBS2411 was determined according to the method of Oyaizu.¹⁸ Various concentrations of DLBS2411 (0–50 µg/mL) in 1.0 mL of deionized water were mixed with 2.5 mL of 0.2 M phosphate buffer (pH 6.6) and 2.5 mL of 1% potassium ferricyanide. The mixture was incubated at 50°C for 20 minutes. Aliquots of trichloroacetic acid were added to 2.5 mL of the mixture and then centrifuged at 3,000 rpm for 10 minutes. Aliquots of the supernatant were mixed with 2.5 mL distilled water and 0.5 mL of a freshly prepared 0.1% ferric chloride solution. Ascorbic acid in various concentrations (0–50 µg/mL) was used as positive control. The absorbance was measured at 700 nm using a spectrophotometer. Increase in absorbance of the reaction mixture indicated an increase in reducing power.

Animal experiments

Experimental animals used were Wistar rats (*Rattus norvegicus*, Wistar strain), 2-month-old males with weight ranging from 200 to 210 g, obtained from D'Wistar (Bandung, Indonesia). They were housed individually and had access to food and water ad libitum. Experimental animals were maintained at a temperature of 22°C–23°C in regular light cycles of 12/12 hours' light/dark in accordance with the guidelines for the use and maintenance of experimental animals that have been published by the Association for Assessment and Accreditation of Laboratory Animal Care (AAALAC). Rats were acclimated for 1 week before treatment. The animal facility has received AAALAC accreditation. All experiments were reviewed for ethical treatment by the independent Institutional Animal Care and Use Committee members of Dexa Medica.

Gastric ulcer induction using indomethacin in rats

Rats were allocated into four groups, consisting of a negative control, a positive control, and two treatment groups. The negative-control group received normal saline, while the positive-control group received normal saline and indomethacin orally. Two treatment groups were treated with DLBS2411 at doses of 20 and 40 mg/kg body weight, respectively, for 7 consecutive days. At the end of the treatment period, the experimental animals were fasted overnight and surgeries to tie up the pylorus were performed following a fasting period. Prior to surgery, the animals were sedated using the anesthetizing drugs ketamine at a concentration of 75 mg/kg body weight and xylazine at a concentration of 10 mg/kg body weight. Their pylori were identified and then tied up using a piece of silk yarn with size of 3/0. Subsequently, test

animals received indomethacin 30 mg/kg body weight and water. After treatment for 6 hours, experimental animals were killed by decapitation. The gaster of the animals was then examined to check ulcer development. The final results are shown based on mean lesion formation. The data were analyzed statistically using Student's *t*-test.

Gastric ulcer induction using ethanol in rats

Rats were allocated into four groups, consisting of a negative control, a positive control, and two treatment groups. All groups received initial oral treatment with absolute ethanol and then received ethanol 15% for 1 week. After 1 week, the positive control group was treated with normal saline, while the two treatment groups were treated with DLBS2411 at doses of 20 and 40 mg/kg body weight. At the end of the treatment period, the experimental animals were fasted overnight. The animals were then killed by cervical decapitation, and their gaster was then examined for ulcer development.

Ex vivo H⁺/K⁺ ATPase assay

Enzyme assays were performed using gastric parietal cells isolated from the gaster of Wistar rats from the in vivo study. The rats were previously induced with ethanol and indomethacin. Enzyme-activity assays were carried out using the EnzCheck phosphate assay kit.

Results

Expression of H⁺/K⁺ ATPase on HEK293 and gastric parietal cells

The present in vitro study demonstrated that treatment with DLBS2411 decreased the level of H⁺/K⁺ ATPase messenger RNA (mRNA) expression of HEK293 and gastric parietal cells in a dose-dependent manner. The downregulation of H⁺/K⁺ ATPase mRNA expression of HEK293 cells was seen as high as 90% at a dose of 50 µg/mL (Figure 1A), a level which was greater than that seen in the gastric parietal cells (50%) (Figure 1B).

Inhibitory effect of DLBS2411 on H⁺/K⁺ ATPase gastric parietal cells in vitro

The effect of DLBS2411 on enzyme activity was investigated in various pH conditions. The result showed that DLBS2411 inhibited H⁺/K⁺ ATPase activity in a dose-dependent manner. DLBS2411 was shown to be most effective at pH 2 (Figure 2). Specifically, the half-maximal inhibitory concentration of DLBS2411 at pH 7.4 was 29.20 µg/mL, pH 4 was 18.70 µg/mL, and pH 2 was 9.20 µg/mL. In addition, the analysis of enzyme kinetics revealed that in the absence

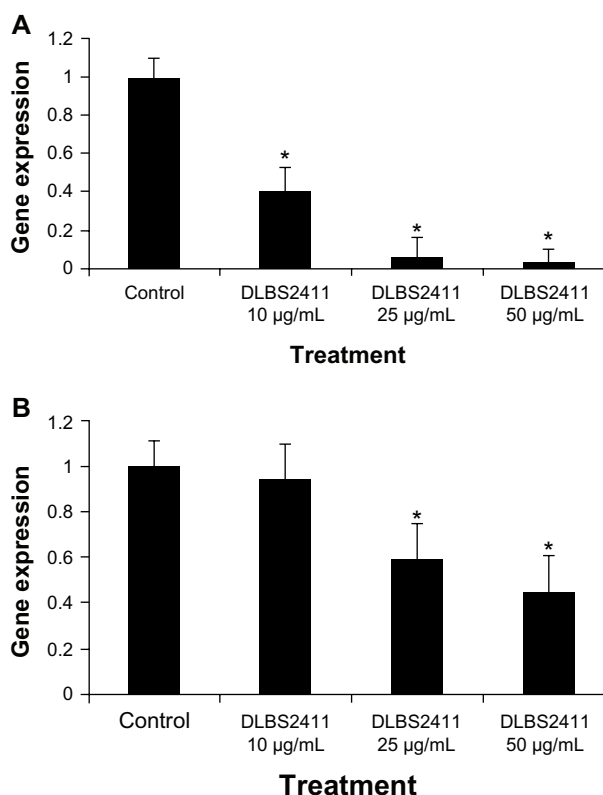


Figure 1 Expression of hydrogen potassium adenosine triphosphatase studied on (A) human embryonic kidney 293 cells and (B) gastric parietal cells after treatment with DLBS2411. The gene expressions were analyzed by real-time polymerase chain reaction.

Note: **P* < 0.05.

of DLBS2411, the Michaelis constant (K_m) and maximum velocity (V_{max}) were 0.3076 µmol/mL and 7.69 mmol Pi/hour, respectively. However, treatment with DLBS2411 yielded K_m and V_{max} values of 0.4002 µmol/mL and 6.67 mmol Pi/hour, respectively (Figure 3A and B). These data suggested that DLBS2411 served as a competitive inhibitor of the enzyme, having an inhibition constant (K_i) of 0.1153 µmol/mL.

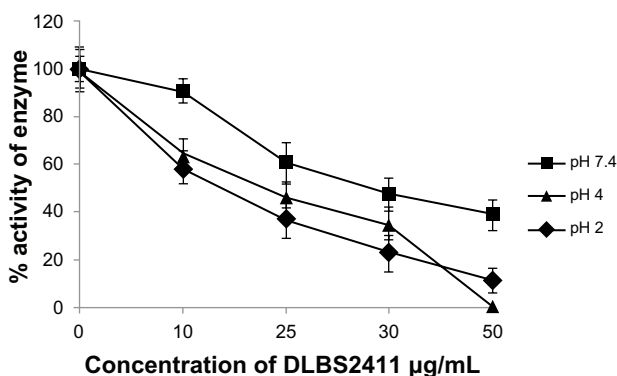


Figure 2 DLBS2411 effect on hydrogen potassium adenosine triphosphatase at pH 2, 4, and 7.4. Enzyme activity was studied using the Enzcheck phosphate assay kit.

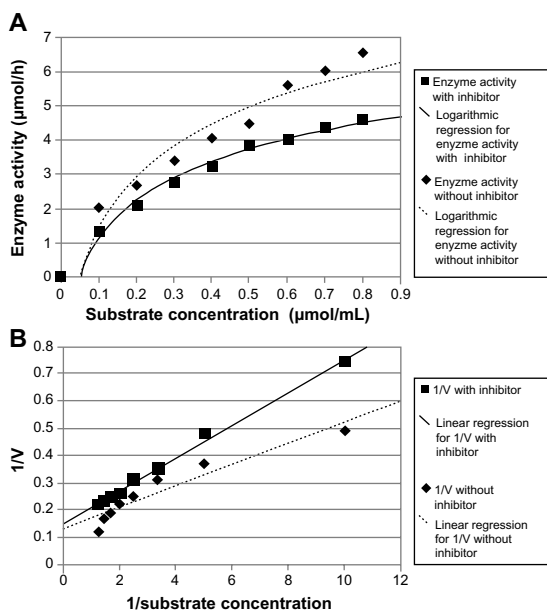


Figure 3 Enzyme kinetic assay of DLBS2411, which shows its properties as a competitive inhibitor. (A) Substrate-velocity curve and (B) Lineweaver-Burk plot of kinetic analysis for H⁺/K⁺ ATPase with DLBS2411 as inhibitor.

Inhibitory effect of DLBS2411 on ulceration in vivo

Indomethacin as a gastric ulcer inducer in rats

The activity of DLBS2411 in reducing gastric ulcers was also shown in the animals treated with indomethacin. Under

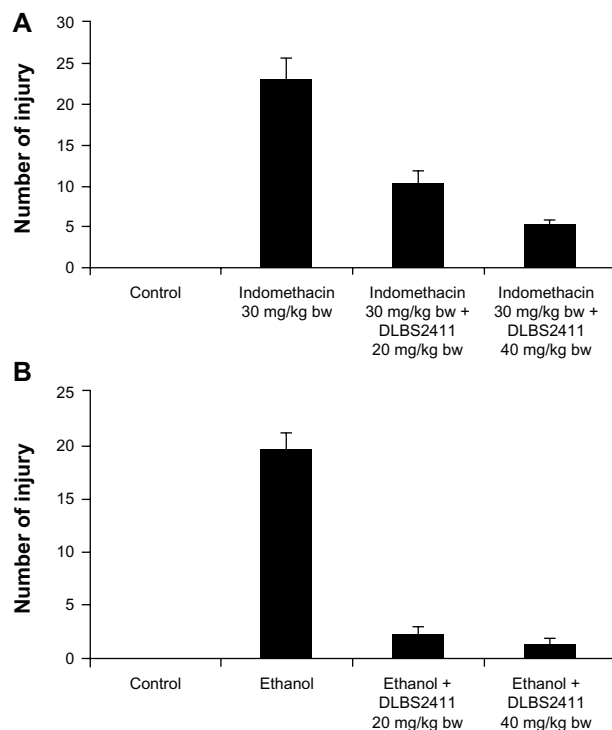


Figure 4 Number of petechiae on Wistar rats after administration of (A) absolute ethanol and 15% ethanol for 1 week and subsequently treated with DLBS2411, and (B) 30 mg/kg body weight of indomethacin after the rats received pylorus ligation treatment and DLBS2411 for 1 week.

this condition, the animals developed petechiae as a result of the treatment. The number of petechiae in the positive group that received indomethacin only was 23 ± 2.65 , while the number receiving DLBS2411 with doses of 20 and 40 mg/kg body weight, prior to addition of indomethacin, was 10.33 ± 1.53 and 5.33 ± 0.58 , respectively (Figure 4B). The fact that the number of petechiae was significantly different when DLBS2411 was given prior to indomethacin compared to indomethacin alone suggested that DLBS2411 conferred its gastroprotection activity to the microenvironment of the gaster.

Ethanol induction of gastric ulcers in rats

Experimentation on gastric ulceration was performed on rats using ethanol treatment. The number of petechiae in experimental animals that received only absolute and 15% ethanol was 19.67 ± 1.53 , while animals that were treated with DLBS2411 at doses of 20 and 40 mg/kg body weight had a significantly lower number of petechiae of 2.33 ± 0.58 and 1.33 ± 0.58 , respectively (Figure 4A). As in the case of indomethacin treatment, these data suggested that DLBS2411 also conferred protection to the gaster, which led to reduced gastric injuries.

Inhibitory effect of DLBS2411 on H⁺/K⁺ ATPase gastric parietal cells ex vivo

H⁺/K⁺ ATPase enzyme-activity assays were conducted on gastric parietal cells isolated from the rats from the in vivo study. When the animals were administered indomethacin or ethanol, the enzyme activity increased (Figure 5A and B). However, simultaneous treatment of indomethacin or ethanol with DLBS2411 caused the enzyme activity to be decreased in a dose-dependent manner. It can be inferred from these data that DLBS2411 confers gastroprotection partly through inhibition of H⁺/K⁺ ATPase activity.

Free radical scavenging activity and reducing-power activities

The free radical scavenging activity of DLBS2411 was observed and compared to the activity of ascorbic acid (Figure 6). At a concentration of 10 µg/mL, DLBS2411 was seen to yield similar free radical scavenging activity to that of ascorbic acid. An experiment was conducted to investigate the reducing power of DLBS2411, compared to an active control – ascorbic acid. As shown in Figure 7, DLBS2411 was able to increase its reducing power in a dose-dependent manner. Taken together, the data suggested DLBS2411 might have an antioxidant activity, although the activity is lower than that of ascorbic acid.

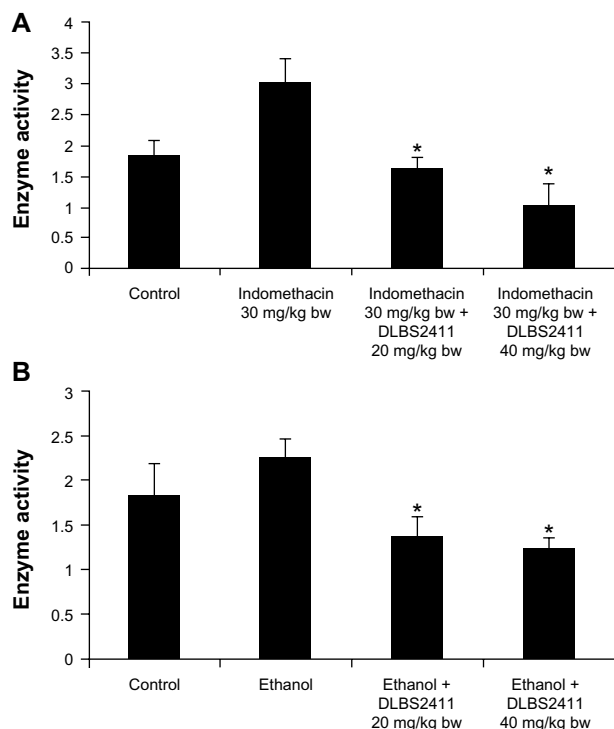


Figure 5 Effect of DLBS2411 on the reduction of hydrogen potassium adenosine triphosphatase enzyme activity on the gastric parietal cells isolated from Wistar rats which were previously treated with (A) 30 mg/kg body weight (bw) of indomethacin and (B) 15% ethanol.

Note: * $P < 0.05$.

Discussion

This study was carried out to elucidate gastroprotective and gastric acid antisecretory effects of DLBS2411, a bioactive fraction of *C. burmannii*. DLBS2411 was particularly investigated for its effect on the gene expression and activity of

the proton-pump H^+/K^+ ATPase enzyme. This enzyme has a significant role in the regulation of acid secretion.^{4,5}

In this study, it was shown that treatment of DLBS2411 downregulated H^+/K^+ ATPase mRNA expression (Figure 1A and B) as well as inhibited H^+/K^+ ATPase activity observed in the genes isolated from the HEK293 and gastric parietal cells (Figure 2). The results reflected a positive correlation between gene expression and protein translations. The reduction in H^+/K^+ ATPase enzyme activity might have been partly due to reduced expression of the gene. In addition, we also studied H^+/K^+ ATPase mRNA expression of the gastric parietal cells with epinephrine as inducer (data not shown). Epinephrine has been shown to stimulate gastric acid secretion in humans and animals.^{19,20} Under this condition, expression was also seen to decrease (data not shown). As we know that epinephrine binds through the beta-adrenergic receptor and uses cyclic adenosine monophosphate and protein kinase A as signal-transduction molecules, the data suggested that DLBS2411 may have used a cyclic adenosine monophosphate-dependent mechanism through which it regulated the expression of H^+/K^+ ATPase.

Several factors have been shown to affect enzyme activity. These may include temperature and pH as well as the presence of an inhibitor. It was found that DLBS2411 was able to exert its function in inhibiting H^+/K^+ ATPase enzyme activity at various gastric pH levels. The inhibition was most pronounced at the lowest gastric pH of 2 (Figure 2). Ulceration occurs when the gaster becomes hyperacidic, where the level of gastric pH is very low. The root cause of this condition is partly related to the activity of H^+/K^+ ATPase

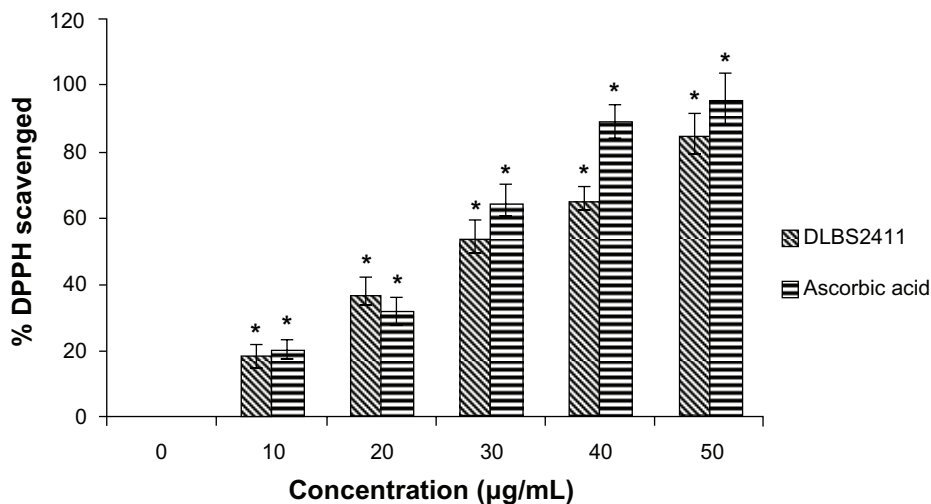


Figure 6 Free radical scavenging activity of DLBS2411 compared to ascorbic acid, suggesting its antioxidant property.

Note: * $P < 0.05$.

Abbreviation: DPPH, 2,2-diphenyl-1-picrylhydrazyl.

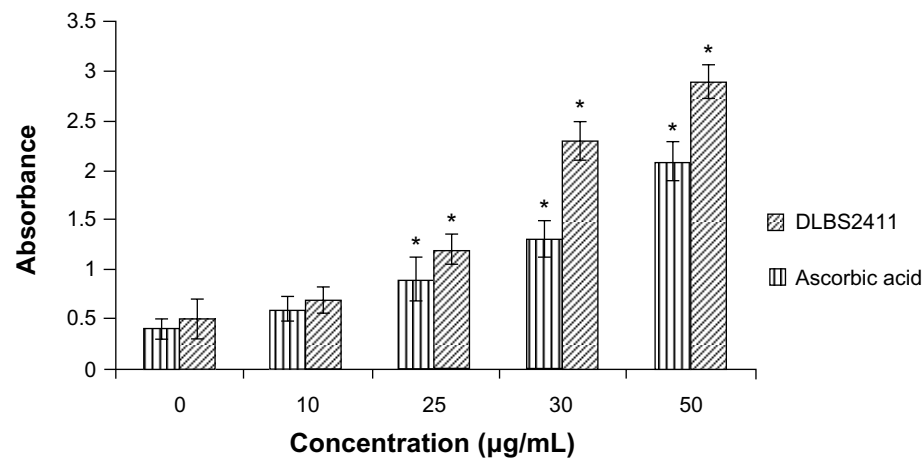


Figure 7 Reducing power of DLBS2411 in comparison with ascorbic acid, suggesting its antioxidant property.

Note: * $P < 0.05$.

in the acid-secretion process. The fact that DLBS2411 is able to work more effectively in low gastric pH suggests that DLBS2411 could serve as a potent pharmacological agent to suppress hyperacidity in the clinical setting and reduce gastric ulceration. The result of chemical identification using thin-layer chromatography (data not shown) showed that the major compound of DLBS2411 is a molecule currently being investigated, but belonging to the class of polyphenol. Reyes-Chilpa et al reported that phenolic compounds are important in the interaction with H⁺/K⁺ ATPase, as they are potent in blocking the activity of this enzyme.⁷ This suggested that as-yet-unidentified phenolic compounds may be present in DLBS2411 and may also contribute to the inhibition of H⁺/K⁺ ATPase.

In order to investigate further the activity of DLBS2411 in inhibiting H⁺/K⁺ ATPase, an enzyme kinetic experiment was performed. It was found that the K_m of enzyme activity treated with DLBS2411 was higher than the K_m of control (Figure 4A and B). This indicates that DLBS2411 has a characteristic as a competitive inhibitor for the enzyme. The competitive inhibitor mimics the action of an enzyme substrate and binds to the active site of the enzyme. The substrate is thereby prevented from binding to the same active site. A competitive inhibitor diminishes the rate of catalysis by reducing the proportion of enzyme molecules bound to a substrate.²¹ K_m values used as a measure of affinity-enzyme substrate are also linked to a complex dissociation balance constant. In this regard, DLBS2411 competitively inhibits H⁺/K⁺ ATPase, an action which leads to the inhibition of enzyme activity in the acid-secretion process.

In addition to the in vitro studies, we also conducted two animal studies with indomethacin and ethanol as ulcer inducers. The ability of NSAIDs such as indomethacin to

cause ulceration and bleeding in the upper gastrointestinal tract has been shown in many studies, including one reference study.²² Indomethacin has the ability to suppress gastric prostaglandin synthesis. A study by Yoshikawa et al suggested that the enhanced gastric motility induced by indomethacin caused microcirculatory disturbances that led to an increase of microvascular permeability and cellular damage.²³ The gastric ulceration in the indomethacin-induced group was also induced by pylorus ligation treatment. This procedure caused the accumulation of gastric acid, which then led to the development of gastric ulcers. Both treatments – indomethacin and pylorus ligation – were chosen as the method to stimulate the development of gastric ulcers. In the other experiment, in vivo ulceration was also induced with ethanol. Previous data have suggested acute damage of gastric mucosa can develop at a higher rate in ethanol-treated experimental animals.²⁴ This gaster-damaging agent induced mucosal lesions that were associated with the production of oxygen free radicals and lipid peroxidation.^{25,26} In the case of gastric injury by these gaster-damaging treatments, the injuries can be shown to cause the development of petechiae in the stomach. DLBS2411 in doses of 20 and 40 mg/kg reduced the number of petechiae significantly in a dose-dependent manner in both treatment groups (Figure 4A and B). This finding correlated with the in vitro antioxidant activity of DLBS2411, where DLBS2411 also reduced the numbers of free radicals that might cause ulceration. These data also suggested that DLBS2411 not only confers activity in proton-pump inhibition but also in protecting mucosa and in minimizing gastric injury.

In order to confirm the mechanism of the DLBS2411 toward the H⁺/K⁺ ATPase enzyme, gastric parietal cells were isolated from the animals in the in vivo study. It was found

that H⁺/K⁺ ATPase enzyme activity was increased with the addition of indomethacin and 15% ethanol (Figure 5A and B), suggesting that those agents induced gastric wounds that could lead to the induction of ulceration. DLBS2411 was able to decrease the activity of H⁺/K⁺ ATPase. These data may also suggest the interaction of compounds contained in the DLBS2411 with H⁺/K⁺ ATPase, which then inhibited the acid-secreting enzyme.

In addition to its activity as enzyme inhibitor, DLBS2411 was also found to possess free radical scavenging activity and reducing-power capability. These data suggested that this agent is also acting as a potent antioxidant. DPPH free radical scavenging is a common method used to evaluate the antioxidant potential of an agent. The free radical chain reaction is widely accepted as a mechanism of lipid peroxidation. For measurements of the reducing power of DLBS2411, the concept of transformation of Fe³⁺–Fe²⁺ adopted from Oyaizu was applied.¹⁸ This study revealed the free radical scavenging activity and reducing power of DLBS2411, which were increased in a dose-dependent manner (Figures 6 and 7). This is in line with the finding that DLBS2411 works in inhibiting enzyme activity that further reduced ulceration. Ulceration often happens due to a hyperoxidation process in the gaster.^{3,8,9} With a highly effective reductive ability, it is expected that DLBS2411 could reduce the hyperoxidation process. This oxygen radical reductive activity indicated that DLBS2411 has the potential to be an antioxidant agent that contributes to the alleviation of gastric ulcer problems.

In conclusion, we have shown in this report some pharmacological data that collectively indicate that DLBS2411 possesses an efficacy to inhibit and downregulate gastric H⁺/K⁺ ATPase, as well as act as an antioxidant. This agent can potentially be used for treating gastric ulcers from various etiologies, such as alcohol, drug, and stress. These findings provide evidence for DLBS2411 to potentially be administered as a natural alternative for gastric acid regulation. Currently, a number of clinical trials are ongoing to prove this concept in subjects suffering from this condition in several hospitals in Asia.

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Disclosure

The authors report no conflicts of interest in this work.

References

- Miller TA. Mechanisms of stress-related mucosal damage. *Am J Med.* 1987;83:8–14.
- Konturek PC, Brzozowski T, Konturek SJ, et al. Apoptosis in gastric mucosa with stress-induced gastric ulcers. *J Physiol Pharmacol.* 1999;50:211–225.
- Bandyopadhyay U, Biswas K, Chatterjee R, et al. Gastroprotective effect of Neem (*Azadirachta indica*) bark extract: possible involvement of H(+)-K(+)-ATPase inhibition and scavenging of hydroxyl radical. *Life Sci.* 2002;71:2845–2865.
- Sachs G, Shin JM, Briving C, Wallmark B, Hersey S. The pharmacology of the gastric acid pump: the H⁺, K⁺ ATPase. *Annu Rev Pharmacol Toxicol.* 1995;35:277–305.
- DuBose TD Jr, Codina J, Burges A, Pressley TA. Regulation of H(+)-K(+)-ATPase expression in kidney. *Am J Physiol.* 1995;269:F500–F507.
- Ono K, Sawada T, Murata Y, et al. Pentagoloylglucose an antisecretory component of Paeoniae radix inhibits gastric H⁺/K⁺ ATPase. *Clin Chim Acta.* 2000;290:159–167.
- Reyes-Chilpa R, Baggio CH, Alavez-Solano D, et al. Inhibition of gastric H⁺, K⁺-ATPase activity by flavonoids, coumarins and xanthenes isolated from Mexican medicinal plants. *J Ethnopharmacol.* 2006;105: 167–172.
- Maity P, Biswas K, Chattopadhyay I, Banerjee RK, Bandyopadhyay U. The use of neem for controlling gastric hyperacidity and ulcer. *Phytother Res.* 2009;23:747–755.
- Bagchi D, Carryl OR, Tran MX, et al. Stress, diet and alcohol-induced oxidative gastrointestinal mucosal injury in rats and protection by bismuth subsalicylate. *J Appl Toxicol.* 1998;18:3–13.
- Fort FL, Miyajima H, Ando T, et al. Mechanism for species-specific induction of Leydig cell tumors in rats by lansoprazole. *Fundam Appl Toxicol.* 1995;26:191–202.
- Powers RE, Lawton GP, Modlin IM. Genotoxicity, carcinogenicity and acid-suppressing medications. *Pharmacol Ther.* 1995;65:303–317.
- Martelli A, Mattioli F, Mereto E, et al. Evaluation of omeprazole genotoxicity in a battery of in vitro and in vivo assays. *Toxicology.* 1998;30:29–41.
- Trisina J, Sunardi F, Suhartono MT, Tjandrawinata RR. DLBS1033, a protein extract from *Lumbricus rubellus*, possesses antithrombotic and thrombolytic activities. *J Biomed Biotech.* 2011;2011:519652.
- Tandrasasmita OM, Wulan DD, Nailufar F, Sinambela J, Tjandrawinata RR. Glucose-lowering effect of DLBS3233 is mediated through phosphorylation of tyrosine and upregulation of PPARγ and GLUT4 expression. *Int J Gen Med.* 2011;4:345–357.
- Tjandrawinata RR, Arifin PF, Tandrasasmita OM. DLBS1425, a *Phaleria macrocarpa* (Scheff.) Boerl. extract confers anti proliferative and proapoptosis effects via eicosanoid pathway. *J Exp Ther Oncol.* 2010;8:187–201.
- Chew CS, Nakamura K, Ljungström M. Calcium signaling mechanisms in the gastric parietal cell. *Yale J Biol Med.* 1992;65:561–623.
- Brand-Williams W, Cuvelier ME, Berset C. Use of a free radical method to evaluate antioxidant activity. *Food Sci Technol.* 1995;28:25–30.
- Oyaizu M. Studies on products of browning reaction: antioxidant activities of products of browning reaction prepared from glucosamine. *Jpn J Nutr.* 1986;44:307–315.
- Christensen KC, Stadil F. Effect of epinephrine and norepinephrine on gastrin release and gastric secretion of acid in man. *Scand J Gastroenterol Suppl.* 1976;37:87–92.
- Marolf CJ, Schultz BD, Clemens ET. Epinephrine effects on gastrin and gastric secretions in normal and stress-susceptible pigs and in dogs. *Comp Biochem Physiol C.* 1993;106:367–370.
- Berg JM, Tymoczko JL, Stryer L. *Biochemistry.* 5th ed. New York: WH Freeman; 2002.
- Wallace JL. How do NSAIDs cause ulcer disease? *Baillieres Best Pract Res Clin Gastroenterol.* 2000;14:147–159.
- Yoshikawa T, Naito Y, Kishi A, et al. Role of active oxygen, lipid peroxidation, and antioxidants in the pathogenesis of gastric mucosal injury induced by indomethacin in rats. *Gut.* 1993;34:732–737.

24. Bienia A, Sodolski W, Luchowska E. The effect of chronic alcohol abuse on gastric and duodenal mucosa. *Ann Univ Mariae Curie Sklodowska Med.* 2002;57:570–582.
25. Patil PB, Mahadik VJ, Patil SB, Naikwade NS. Evaluation of antiulcer activity of aqueous and ethanolic extract of *Oxalis corniculata* leaf in experimental rats. *Int J Pharm Res Dev.* 2011;3:98–104.
26. Chen SH, Liang YC, Chao JC, et al. Protective effects of *Ginkgo biloba* extract on the ethanol-induced gastric ulcer rats. *World J Gastroenterol.* 2005;11:3746–3750.

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