

Robust aptamer–polydopamine-functionalized M-PLGA–TPGS nanoparticles for targeted delivery of docetaxel and enhanced cervical cancer therapy

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Abstract: One limitation of current biodegradable polymeric nanoparticles (NPs) is the contradiction between functional modification and maintaining formerly excellent bioproperties with simple procedures. Here, we reported a robust aptamer–polydopamine-functionalized mannitol-functionalized poly(lactide-*co*-glycolide) (M-PLGA)–D- α -tocopheryl polyethylene glycol 1000 succinate (TPGS) nanoformulation (Apt-pD-NPs) for the delivery of docetaxel (DTX) with enhanced cervical cancer therapy effects. The novel DTX-loaded Apt-pD-NPs possess satisfactory advantages: 1) increased drug loading content and encapsulation efficiency induced by star-shaped copolymer M-PLGA–TPGS; 2) significant active targeting effect caused by conjugated AS1411 aptamers; and 3) excellent long-term compatibility by incorporation of TPGS. Therefore, with simple preparation procedures and excellent bioproperties, the new functionalized Apt-pD-NPs could maximally increase the local effective drug concentration on tumor sites, achieving enhanced treatment effectiveness and minimizing side effects. In a word, the robust DTX-loaded Apt-pD-NPs could be used as potential nanotherapeutics for cervical cancer treatment, and the aptamer–polydopamine modification strategy could be a promising method for active targeting of cancer therapy with simple procedures.

Keywords: dopamine, AS1411 aptamer, active targeting, polymeric NPs, enhanced cervical chemotherapy

Introduction

Cervical cancer is one of the most common malignant tumors and serious health problems, especially in developing countries. Both the morbidity and the mortality of cervical cancer have substantially increased in recent years. However, chemotherapy, which is one of the most used clinical treatments for cervical cancer nowadays, is nonspecific, and huge disadvantages such as serious side effects due to the lack of effective carriers still exist.^{1–3} As an efficient solution, polymeric nanoparticles (NPs) have been employed as highly promising carriers for anticancer drugs, functioning as sustained, controlled, and targeted drug delivery systems to improve the therapeutic effects for cervical cancer and to reduce the side effects on normal organs.^{4–8} In order to transport the NPs to tumor cells *in vivo*, a relatively long circulating half-life and sufficient target interaction are the two essential requirements that could be achieved through surface modification.⁹

Aptamers, which are single-stranded RNA or DNA ligands, are oligonucleotides or peptide molecules that bind a wide variety of biological targets on cancer cells with high affinity and specificity.^{10–12} Aptamers can be used for both basic research and clinical purposes as macromolecular drugs. Aptamers can also be combined with ribozymes

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to self-cleave in the presence of their target molecules. These compound molecules have additional research, industrial, and clinical applications. Owing to higher in vivo stability¹³ and good targeting moieties, aptamers are widely used in therapeutic applications.¹⁴ AS1411 is one of the frequently used DNA aptamers that can inhibit the viabilities of various cancer cells, such as prostate cancer cells (DU145), breast cancer cells (MCF-7),¹⁵ and cervical cancer cells (HeLa). Due to its superior performance on enhanced therapeutic effects and targeting efficacy, it can be used as an efficient targeting ligand for surface modification of NPs in cervical cancer treatment. However, for NPs prepared using prefunctionalized polymers, the synthesis of polymer–aptamer conjugates can be inefficient, lengthy with high cost, at a risk of altering the chemical properties, and compromising the ability of the polymer to encapsulate drugs.¹⁶

To successfully address these issues, we adopt a novel, simple, and versatile modification technique based on dopamine polymerization. Modification of the NP surface with hydrophilic, electrically neutral polymers such as polydopamine (pD) can achieve a crucial effect to incorporate functionalized ligands (eg, peptides, nucleic acids, and small molecules). Dopamine, which was a kind of traditional neurotransmitter inspired by the chemical constituents of adhesive proteins in mussels,¹⁷ has been reported to be used in modification of the surface of materials.¹⁸ Park et al¹⁹ also demonstrated that the dopamine polymerization method is a simple and versatile surface modification pathway, applicable to a variety of NP drug carriers irrespective of their chemical reactivity and the types of ligands (eg, small molecules, peptides, and polymers). In brief, dopamine catechol can be oxidized to quinone that reacts with other catechols or quinones to form polydopamine in weak alkaline conditions (~pH 8–pH 8.5), eventually forming a water-insoluble polymer film on the surface of NPs.²⁰ After that, ligands possessing nucleophilic functional groups (eg, amine and thiol) can be incorporated into the surface layer via Michael addition or Schiff base reactions.^{21–23}

In our previous research,²⁴ a star-shaped mannitol-functionalized poly(lactide-*co*-glycolide) (M-PLGA)-D- α -tocopheryl polyethylene glycol 1000 succinate (TPGS) copolymer was successfully synthesized and characterized, and the docetaxel (DTX)-loaded M-PLGA–TPGS NPs have been demonstrated to have many superiorities, for example, increased bioavailability of anticancer drugs (solubility, permeability, and stability), improved drug loading content (LC) and encapsulation efficiency (EE), accelerated drug release, and good in vitro antitumor efficiency. Hence, we continued our further research on M-PLGA–TPGS NPs for

cervical cancer therapy and inferred that aptamers could be conjugated to pD-modified M-PLGA–TPGS NPs. The new and improved functionalized NPs were characterized, and the antitumor effects were investigated both in vitro and in vivo.

Materials and methods

Materials

Mannitol (C₆H₁₄O₆), (TPGS (C₃₃O₅H₅₄ (CH₂CH₂O)₂₃)), coumarin-6 (C₆), D,L-lactide (3,6-dimethyl-1,4-dioxane-2,5-dione, C₆H₈O₄), 2-(3,4-dihydroxyphenyl) ethylamine (dopamine) hydrochloride, propidium iodide, and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) were purchased from Sigma-Aldrich Co. (St Louis, MO, USA). PLGA (LA:GA =75:25, Mn \approx 25,000) was purchased from Jinan Daigang Biomaterial Co. Ltd (Jinan, People's Republic of China). DTX was provided by Beijing InnoChem Science & Technology Co., Ltd (Beijing, People's Republic of China). Commercial Taxotere[®] (purity: 99.9%) was bought from Shanghai Jinhe Bio-Technology Co., Ltd (Shanghai, People's Republic of China). All other agents used were of the highest commercial grade available. Human cervix carcinoma cell line HeLa was purchased from American Type Culture Collection (Manassas, VA, USA). The Administrative Committee on Animal and Cell Research in Tsinghua University approved the protocols for cellular assays.

Synthesis of DTX-loaded Apt-pD-NPs

Preparation of DTX-loaded M-PLGA–TPGS NPs

A modified nanoprecipitation method with an acetone/water system was used to prepare the DTX-loaded M-PLGA–TPGS NPs (DTX-loaded NPs).^{25,26} Briefly, 20 mg DTX and 200 mg M-PLGA-*b*-TPGS copolymers powder were dissolved in 16 mL of acetone. The blend solution was titrated into 200 mL of 0.03% (w/v) TPGS aqueous solution under stirring. After stirring overnight (800 rpm) at room temperature to remove acetone, the DTX-loaded NPs were centrifuged at 20,000 rpm (4°C) for 20 minutes and washed three times in 20 mL of deionized water to remove unencapsulated drug and TPGS emulsifier. At last, we selected precalculated amount of the precipitated DTX-loaded NPs for pD coating, while the left DTX-loaded NPs were dispersed in deionized water and lyophilized for 2 days for further use.

Prime coating with pD

The DTX-loaded and pD-coated NPs (DTX-loaded pD-NPs) were synthesized by incubating DTX-loaded NPs in 0.1 mg/mL dopamine hydrochloride/Tris buffer solution (pH 8.5, dopamine hydrochloride dissolved in a 10 mM

Tris buffer) under stirring at room temperature. After magnetic stirring for 6 hours, the suspensions turned dark when dopamine hydrochloride was added, indicating that dopamine was successfully polymerized. The obtained DTX-loaded pD-NPs were centrifuged and lyophilized for aptamer conjugation.

Conjugation of aptamers to pD-coated NPs

The AS1411 aptamers (5'-GGT GGT GGT GGT TGT GGT GGT GGT TTT TTT TTT-thiol-3'), whose sequence contains ten extra T bases at the 3-terminus, were connected to the surface of DTX-loaded pD-NPs via the Michael addition reaction.¹⁹ In brief, the DTX-loaded pD-NPs were resuspended in Tris buffer (pH 8.0) containing aptamers. The final concentration of DTX-loaded pD-NPs and aptamers was 1 mg/mL and 0.5 mg/mL, respectively. After magnetic stirring for 2 hours at room temperature, the resulting NPs (designated as DTX-loaded Apt-pD-NPs) were centrifuged, washed three times with deionized water, and lyophilized for 2 days.

All the fluorescent (C6- or IR-780-loaded) NPs were fabricated in a similar manner at each of the three steps described earlier. All the lyophilized NPs were redispersed in phosphate-buffered saline (PBS) before use.

Characterization of functionalized NPs

Particle size, zeta potential, and morphology

The size of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs was analyzed using Malvern Mastersizer 2000 (Zetasizer Nano ZS90; Malvern Instruments, Malvern, UK).²⁷ The NPs were resuspended in deionized water before experiments. These data were obtained by averaging the values obtained at three time points. The surface morphology of the NPs was measured by a field emission scanning electron microscopy (FESEM; JEOL JSM-6700F, Tokyo, Japan) and a transmission electron microscopy (TEM; Tecnai G² F30; FEI Company, Hillsboro, OR, USA).

Fourier transform infrared spectroscopy analysis

The Fourier transform infrared (FT-IR) spectroscopy analysis of drug-free NPs, drug-free pD-NPs, and drug-free Apt-pD-NPs was recorded by FT-IR spectrophotometer (Thermo Nicolet, Madison, WI, USA) using KBr.

X-ray photoelectron spectroscopy

The X-ray photoelectron spectroscopy (XPS) of the DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs was obtained by a Kratos Axis Ultra DLD spectrometer (Kratos Analytical Ltd, Manchester, UK) with monochromatic Al K α radiation ($h\nu = 1,486.58$ eV). Survey and high-resolution spectra were collected at a fixed analyzer

pass energy of 160 eV and 20 eV, respectively. Binding energy values were referenced to the Fermi edge, and charge correction was performed setting the C 1s peak at 284.80 eV.

Drug LC and EE

The solution, which was prepared using 5 mg NPs dissolved in 1 mL of dichloromethane under vigorous vortexing, was transferred to 5 mL of mobile phase consisting of acetonitrile and deionized water (50:50, v/v). The LC and EE of the modified NPs were determined by high-performance liquid chromatography (HPLC; LC 1200; Agilent Technologies, Santa Clara, CA, USA), and to obtain a clear solution for HPLC, a nitrogen stream was introduced for ~15 minutes to evaporate dichloromethane. A reverse-phase C18 column (150×4.6 mm, 5 μ m, C18; Agilent Technologies) was used at 35°C. The flow rate of mobile phase was set at 1.0 mL/min. The column effluent was detected at 227 nm using a UV/VIS detector. Measurements were carried out three times for each batch. The LC and EE of modified NPs were calculated by the following equations, respectively.

$$\text{LC (\%)} = \frac{\text{Weight of DTX in NPs}}{\text{Weight of NPs}} \times 100\% \quad (1)$$

$$\text{EE (\%)} = \frac{\text{Weight of DTX in NPs}}{\text{Weight of the feeding DTX}} \times 100\% \quad (2)$$

In vitro drug release profiles

Before the testing of in vitro DTX release profiles, 5 mg of the lyophilized NPs was put in a centrifuge tube and dispersed in 10 mL of PBS (pH 7.4, containing 0.1% w/v Tween 80). Tween 80 was added to avoid the binding of DTX to the tube wall and to increase the solubility of DTX in the buffer. The supernatant was then transferred into a dialysis membrane bag (molecular weight cut off [MWCO] = 3,500; Shanghai Sangon, People's Republic of China) that was immersed in 10 mL of fresh PBS release medium in a centrifuge tube. At designated time intervals, the tube was transferred into an orbital water bath and shaken at 200 rpm at 37°C. The cumulative release buffer outside the dialysis bag was plotted to HPLC analysis.

Endocytosis of fluorescent NPs

HeLa cells were cultured in a chambered cover glass system in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% fetal bovine serum, 100 U/mL penicillin, and 100 mg/mL streptomycin in 5% CO₂ at 37°C. The culture was kept in 95% air humidified atmosphere. The cells were incubated with 250 μ g/mL C6-loaded NPs (or pD-NPs or

Apt-pD-NPs) for 2 hours, washed with cold PBS three times, and fixed with cold methanol for 20 minutes. After that, the nuclei were counterstained with propidium iodide for 30 minutes and washed twice with PBS. In order to visualize the cells, the chambers were mounted onto the confocal laser scanning microscope (FV-1000, Olympus FluoView™, Tokyo, Japan) with the following channels: a blue channel excited at 340 nm and a green channel excited at 485 nm.

For quantitative analysis, HeLa cells were plated in 96-well black plates and incubated overnight with their initial density: 1×10^4 cells/well. The cells were then equilibrated with Hank's buffered salt solution for 1 hour at 37°C, and C6-loaded NPs (or pD-NPs or Apt-pD-NPs) were added at concentrations 100 µg/mL, 250 µg/mL, and 500 µg/mL. After incubation for 2 hours, the medium was removed and the wells were washed three times with 50 µL cold PBS. Finally, 50 µL of 0.5% Triton X-100 in 0.2 N sodium hydroxide was put into each sample well to lyse the cells.

Cell viability evaluation

To evaluate the in vitro antitumor cytotoxicity of the DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs, HeLa cells were seeded at a density of 5,000 cells/well in 96-well plates and incubated overnight. Afterward, the cells were incubated with drug-free NPs, drug-free pD-NPs, drug-free Apt-pD-NPs, commercial Taxotere, DTX-loaded NPs, DTX-loaded pD-NPs, or DTX-loaded Apt-pD-NPs suspension at 0.25 µg/mL, 2.5 µg/mL, 12.5 µg/mL, and 25 µg/mL equivalent DTX concentrations for 24 hours and 48 hours. The formulations were changed with MTT-contained DMEM (5 mg/mL), and the cells were incubated for an additional 4 hours at a determined time. Then, dimethyl sulfoxide was added to dissolve the formazan crystals (dark, 2 hours), and MTT was removed at 37°C. A microplate reader (Bio-Rad Model 680, Bio-Rad Laboratories Inc., Hercules, CA, USA) was used to measure the absorbance at 570 nm. Untreated cells served as 100% viability and cells incubated with MTT-free medium were taken as blank to calibrate the spectrophotometer to zero absorbance. The drug concentration, at which the growth of 50% cells was inhibited (defined as IC_{50}), was detected by curve fitting of the cell viability data in comparison with that of the control groups.

Xenograft tumor model and in vivo antitumor effects

The Institute of Laboratory Animal Sciences, Chinese Academy of Medical Science, provided the female severe combined immunodeficient (SCID) mice aged 4–5 weeks old. The Administrative Committee on Animal Research in Tsinghua

University approved the protocols for animal assays and all experimental procedures were carried out according to the Animal Care guidelines. About 100 µL of HeLa cells in PBS were implanted subcutaneously into the backs of mice (15–20 g) at a dosage of 2×10^6 cells per mouse. After that, we observed the tumor growth in each mouse at frequent intervals. A Vernier caliper was used to survey the tumor size, and the tumor volume (V) was calculated as $4\pi/3 \times (\text{length}/2) \times (\text{width}/2)^2$. Ninety-five percent of the mice injected developed a tumor with an average volume of $\sim 80 \text{ mm}^3$ after 10 days. Treatments were started after the tumor size reached $\sim 80 \text{ mm}^3$. Two experienced researchers randomly divided the mice into five groups ($n=5$), which were intravenously injected with saline as a control, DTX-loaded NPs, DTX-loaded pD-NPs, DTX-loaded Apt-pD-NPs, and Taxotere, respectively, at a DTX dose of 10 mg/kg in 100 µL PBS on days 0, 4, 8, 12, 16, and 20. The body weights and tumor size of each mouse were recorded every other day. The mice were humanely killed after 24 days of treatment. The antitumor efficacy was investigated via tumor growth in this study.

The antitumor efficacy of the DTX-loaded Apt-pD-NPs was also investigated on another batch of SCID mice bearing HeLa tumor xenograft, by measuring their survival time after treatments. After choosing 20 mice with similar body status (ie, age, weight, and 80 mm³ of tumor size), the mice were randomly divided into five groups ($n=5$) and injected with Taxotere, DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs (DTX dose of 10 mg/kg), and saline via tail vein for every 4 days. The survival data were presented as Kaplan–Meier plots and analyzed with a log-rank test.

Statistical methodology

All tests were carried out at least three times in all experiments unless otherwise stated. The results are expressed as mean \pm SD, and the statistical significance of all the results was determined by the Student's t -test. The results were considered to be statistically significant if $P < 0.05$.

Results and discussion

Fabrication of DTX-loaded Apt-pD-NPs

The preparation of DTX-loaded Apt-pD-NPs is exhibited in Figure 1. During the polymerization of dopamine, a tight pD layer surrounded the surface of M-PLGA–TPGS NPs, which were immersed in the dopamine solution. As shown in Figure 1, there were two fatal processes of functionalized DTX-loaded Apt-pD-NPs: in a weak alkaline condition, dopamine was able to undergo oxidative polymerization in the presence of oxygen as an oxidant on the surface of M-PLGA–TPGS (Process I) and conjugation of aptamers to pD-coated NPs (DTX-loaded pD-NPs) with a Michael

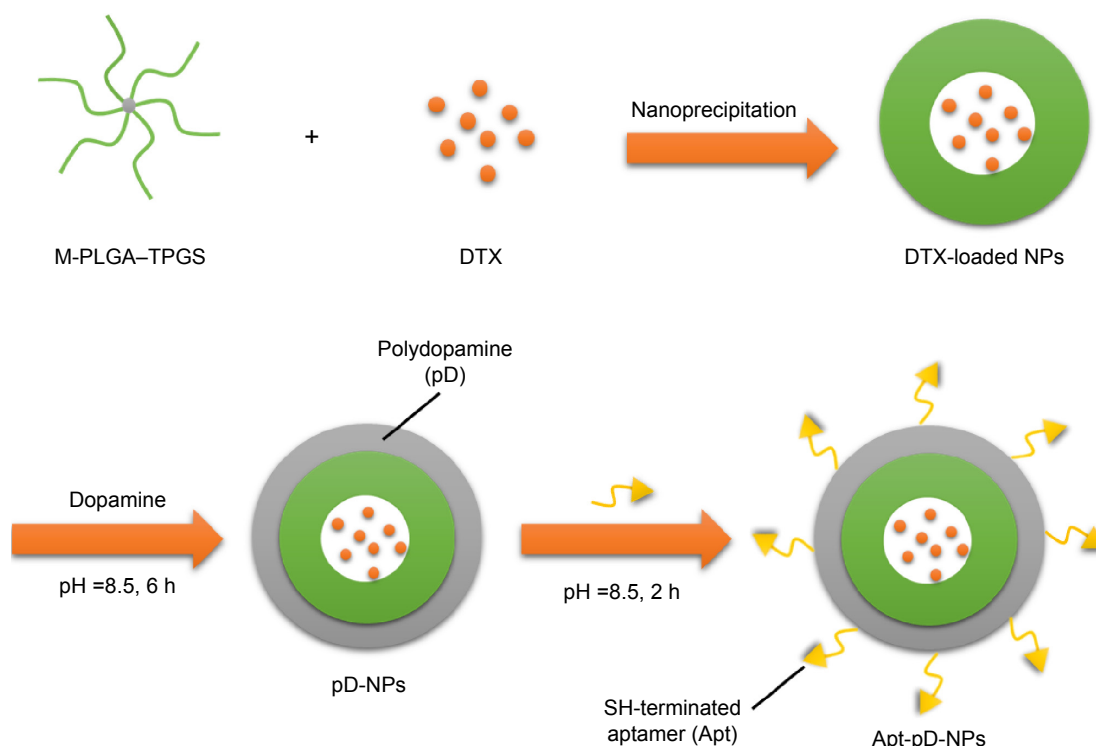


Figure 1 Schematic representation of the preparation procedure for targeted and DTX-loaded Apt-pD-NPs.

Notes: There were two fatal processes of functionalized DTX-loaded Apt-pD-NPs: oxidative polymerization of dopamine in the presence of oxygen as an oxidant on the surface of M-PLGA-TPGS (Process I) and conjugation of aptamers to pD-coated NPs (DTX-loaded pD-NPs) with a Michael addition reaction (Process II).

Abbreviations: DTX, docetaxel; Apt, aptamer; pD, polydopamine; NPs, nanoparticles; M-PLGA, mannitol-functionalized poly(lactide-co-glycolide); TPGS, D- α -tocopheryl polyethylene glycol 1000 succinate; h, hours.

addition reaction (Process II). It turned out that dopamine was successfully polymerized, while the suspensions turned dark with some particulate pD appearing during the washing steps. Afterward, a 26-mer SH-terminated DNA aptamer (AS1411) was conjugated with the pD-coated NPs, and it was deemed to be targeted therapy for cervical cancer.

Characteristics of DTX-loaded Apt-pD-NPs

The FT-IR spectroscopy of the drug-free NPs, pD-NPs, and Apt-pD-NPs is exhibited in Figure 2. As for drug-free NPs, the absorption band at $1,750\text{ cm}^{-1}$ was assigned to the carbonyl band of TPGS and PLGA.²⁸ The absorption band at $1,600\text{ cm}^{-1}$ with aromatic rings stretching vibration of pD supported the fact that the pD was coated on the surface of NPs.²⁹ In addition, the $1,750\text{ cm}^{-1}$ band significantly decreased and nearly disappeared. It turned out that both the pD-coating and the aptamers for Apt-pD-NPs were conjugated. XPS was studied to further demonstrate the successful modification of pD and conjugation of aptamers. As displayed in Figure 3, both pD-NPs and Apt-pD-NPs showed two nitrogen peaks (N 1s) at $\sim 398.8\text{ eV}$ (nonprotonated amine groups) and $\sim 402.1\text{ eV}$ (protonated amine groups) in XPS spectra, but bare NPs did not, verifying the presence of pD films

on the NP surface. In the case of carbon peaks (C 1s), the increased intensity (pD-NPs or Apt-pD-NPs vs bare NPs) at $\sim 286.2\text{ eV}$ and $\sim 284.7\text{ eV}$ was contributed to the C–O groups and C–C groups from pD films on the NP surface. Taken together, the FT-IR spectroscopy and XPS studies indicated

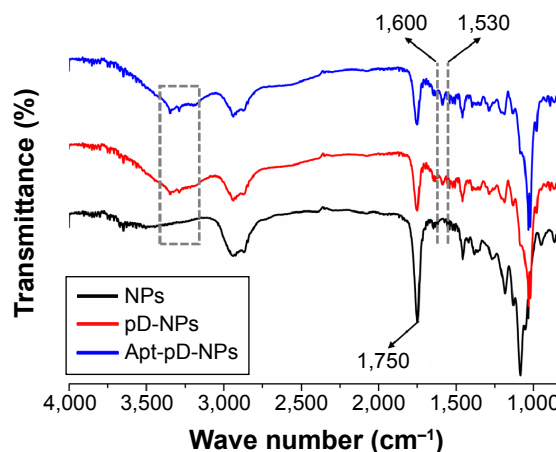


Figure 2 Fourier transform infrared spectra of drug-free NPs, drug-free pD-NPs, and drug-free Apt-pD-NPs.

Notes: The absorption band at $1,750\text{ cm}^{-1}$ is assigned to the carbonyl band of TPGS and PLGA. The absorption band at $1,600\text{ cm}^{-1}$ with aromatic rings stretching vibration of pD supported the fact that the pD was coated on the surface of NPs.

Abbreviations: NPs, nanoparticles; pD, polydopamine; Apt, aptamer; PLGA, poly(lactide-co-glycolide); TPGS, D- α -tocopheryl polyethylene glycol 1000 succinate.

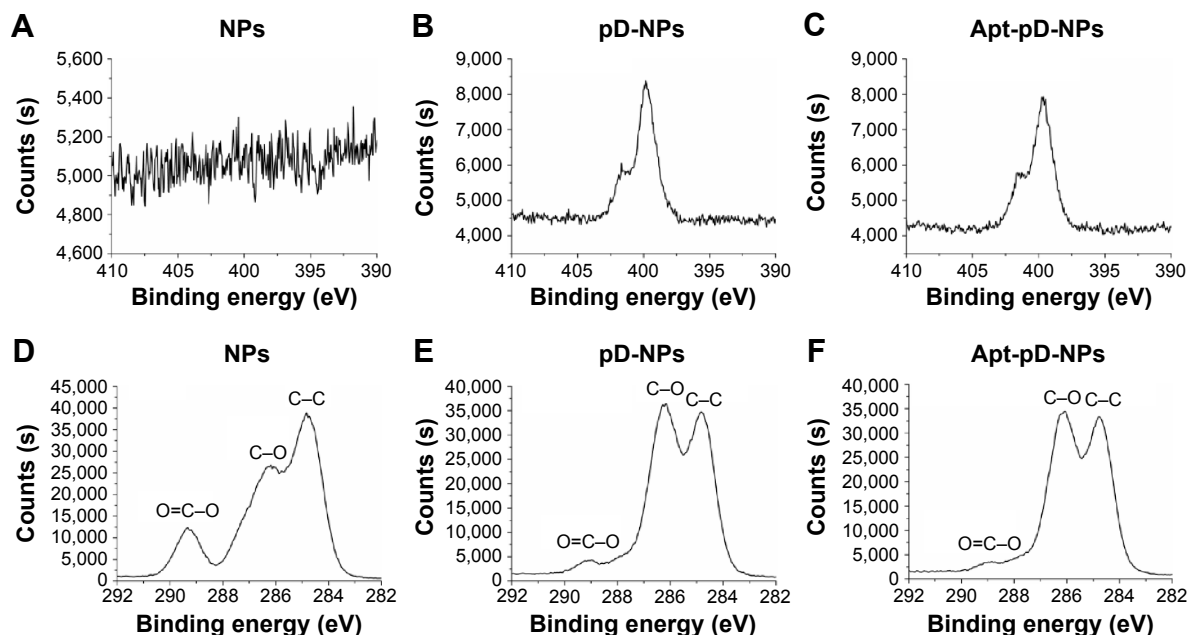


Figure 3 XPS spectra of NPs, pD-NPs, and Apt-pD-NPs.

Notes: (A–C) Narrow scan for N 1s peaks. (D–F) Narrow scan for C 1s peaks. Both pD-NPs and Apt-pD-NPs showed two nitrogen peaks (N 1s) at ~398.8 eV (nonprotonated amine groups) and ~402.1 eV (protonated amine groups) in XPS spectra, but bare NPs did not, verifying the presence of pD films on NP surface.

Abbreviations: XPS, X-ray photoelectron spectroscopy; NPs, nanoparticles; pD, polydopamine; Apt, aptamer; s, seconds.

the successful incorporation of pD film and conjugation of AS1411 aptamers on the surface of NPs.

Furthermore, the size and surface properties of NPs have a vital position in drug release, cellular uptake, and in vivo pharmacokinetics.³⁰ The FESEM and TEM images of

DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs are illustrated in Figure 4A–F. Thin spherical films with a single surface could be obviously visualized on the surface of both DTX-loaded pD-NPs and DTX-loaded Apt-pD-NPs, which indicates that pD films were successfully

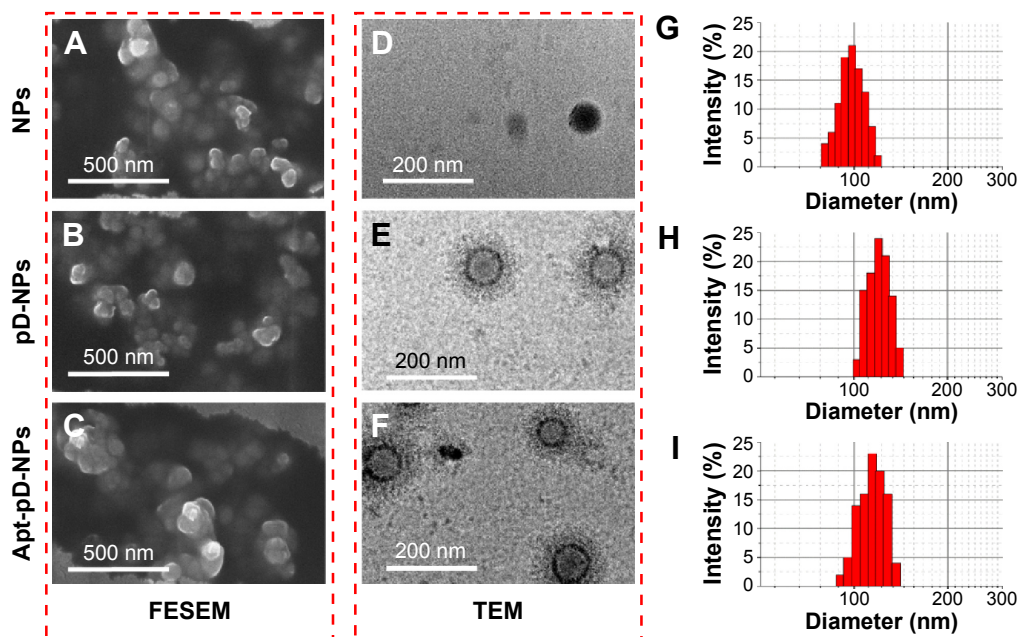


Figure 4 Surface morphology and particle size of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs.

Notes: (A–C) FESEM images, (D–F) TEM images, and (G–I) DLS size distribution. Thin spherical films with a single surface could be obviously visualized on the surface of both DTX-loaded pD-NPs and DTX-loaded Apt-pD-NPs could further confirm the existence of pD film.

Abbreviations: DTX, docetaxel; FESEM, field emission scanning electron microscopy; TEM, transmission electron microscopy; DLS, dynamic light scattering; NPs, nanoparticles; pD, polydopamine; Apt, aptamer.

Table 1 Characterization of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs

Samples (n=3)	Size (nm)	PDI	ZP (mV)	LC (%)	EE (%)
NPs	90.8±4.2	0.163	-12.9±4.3	9.99±0.47	98.54±1.63
pD-NPs	129.8±6.7	0.137	-14.1±4.8	9.83±0.51	98.13±1.57
Apt-pD-NPs	130.1±2.9	0.168	-14.9±5.1	9.47±0.42	98.06±1.46

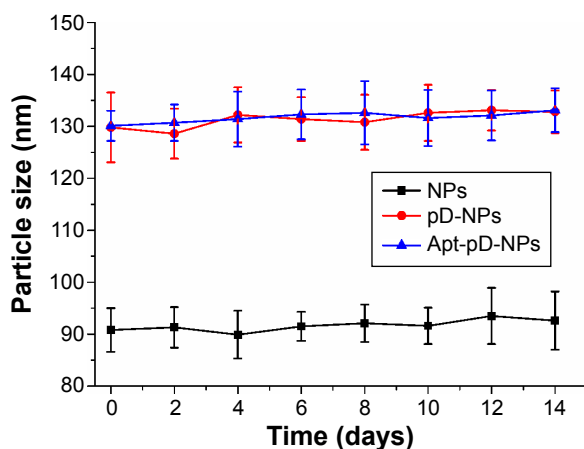
Note: Data presented as mean ± standard deviation.

Abbreviations: DTX, docetaxel; NPs, nanoparticles; pD, polydopamine; Apt, aptamer; PDI, polydispersity index; ZP, zeta potential; LC, loading content; EE, encapsulation efficiency.

deposited on the surface of NPs via an oxidative polymerization reaction. The attenuated average particle size (Table 1), which may be induced by a tendency to shrink and collapse while the NPs are in the dry state,²⁴ was ~80 nm in TEM images for DTX-loaded NPs and ~120 nm for DTX-loaded pD-NPs or DTX-loaded Apt-pD-NPs.

As shown in Table 1, the increased size of DTX-loaded pD-NPs or DTX-loaded Apt-pD-NPs, which reached nearly 40 nm, may be attributed to the thin pD films (ie, ~20 nm, half of the total increased size) formed around the DTX-loaded NPs and demonstrated the successful coating of pD through the oxidative polymerization reaction. The size distribution of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs is illustrated in Figure 4G–I. The mean hydrodynamic size of DTX-loaded NPs was ~125–135 nm in diameter, which could easily lead to the accumulation of tumor vasculature under the influence of the enhanced permeability and retention effect.³¹ Because of the good redispersion stability of the DTX-loaded pD-NPs shown in Figure 5, the average size of the NPs hardly changed in the 14 days experiment.

Zeta potential is a vital factor for the stability of the NPs in cellular uptake, drug release, in vivo pharmacokinetics, and biodistribution.³² As shown in Table 1, the zeta potential

**Figure 5** Stability of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs in PBS, respectively, by monitoring particle size over a span of 2 weeks (n=3).

Abbreviations: DTX, docetaxel; NPs, nanoparticles; pD, polydopamine; Apt, aptamer; PBS, phosphate-buffered saline.

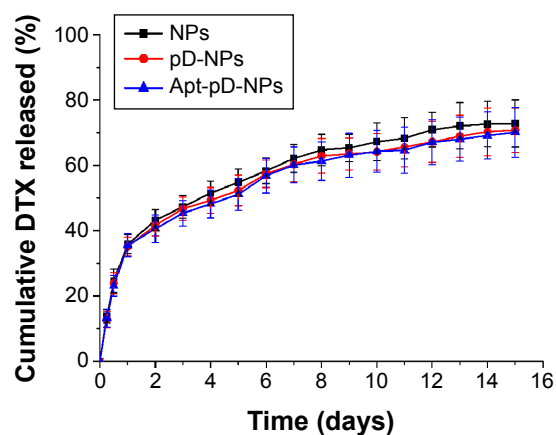
of the DTX-loaded pD-NPs and DTX-loaded Apt-pD-NPs was observed to be -14.1 mV and -14.9 mV, respectively, which is slightly higher than that of the DTX-loaded NPs (~-12.9 mV), which may be caused by the slightly negative charge of NPs. As can be seen from Table 1, the LC and EE of Apt-pD-DTX/NPs (9.47%±0.42%, 98.06%±1.46%) were a little bit lower than DTX/NPs (9.99%±0.47%, 98.54%±1.63%) and pD-DTX/NPs (9.83%±0.51%, 98.13%±1.57%), which may be caused by the loss of a few loaded drugs when aptamers were conjugated to the NPs.

In vitro drug release kinetics

In vitro DTX release from DTX-loaded NPs, DTX-loaded pD-NPs, or DTX-loaded Apt-pD-NPs was surveyed in the release medium (pH 7.4, PBS containing 0.1% w/v Tween 80) at 37°C (Figure 6). A fast release of DTX-loaded NPs, DTX-loaded pD-NPs, or DTX-loaded Apt-pD-NPs was exhibited in the initial 48 hours, that is, ~45% of the encapsulated drugs were released in all NPs. After 14 days, 72.57%, 70.48%, and 69.87% of drugs were released from DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs, respectively. The release rate from DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs was quite similar, indicating that the pD-coating and AS1411 conjugation did not have much influence on the drug release properties of the DTX-loaded NPs. Thus, the DTX-loaded Apt-pD-NPs can be a promising drug delivery vector in nanomedicine research.

Cellular uptake of fluorescent NPs and in vitro cellular targeting

The cellular uptake profile of the NPs was investigated because of the important roles that the cellular internalization

**Figure 6** In vitro drug release profile of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs.

Notes: The pD-coating and AS1411 conjugation seemed to not have much influence on the drug release properties of the DTX-loaded NPs.

Abbreviations: DTX, docetaxel; NPs, nanoparticles; pD, polydopamine; Apt, aptamer.

and sustained retention play on the therapeutic effects of the NPs.³³ Figure 7A shows the confocal laser scanning microscope images of the HeLa cells after 2-hours incubation with the suspension of 250 $\mu\text{g/mL}$ C6-loaded NPs in DMEM. As shown in the figure, the fluorescence intensity, which represents the cellular uptake efficiency, was nearly equivalent between the cells incubated with C6-loaded NPs and C6-loaded pD-NPs. Furthermore, the existence of aptamers on the surface of NPs may lead to high-efficiency in vitro cellular targeting, and the fluorescence intensity increased significantly after 2 hours incubation with the C6-loaded Apt-pD-NPs. To verify the significant role of aptamers in the cellular uptake of the C6-loaded Apt-pD-NPs, we performed a receptor competition assay by selecting aptamers as the competitive reagent. When the C6-loaded Apt-pD-NPs and aptamers were added to wells at the same time, the fluorescence was significantly quenched. Furthermore, with LNCaP cells (AS1411 nonspecific cancer cells) used as control, the fluorescence observed in LNCaP cells was also significantly decreased compared with HeLa cells

incubated with the C6-loaded Apt-pD-NPs. These tests certified the tumor targeting specificity of the C6-loaded Apt-pD-NPs.

As displayed in Figure 7B, HeLa cells were incubated with the C6-loaded NPs, the C6-loaded pD-NPs, or the C6-loaded Apt-pD-NPs for 2 hours at the concentration of 100 $\mu\text{g/mL}$, 250 $\mu\text{g/mL}$, and 500 $\mu\text{g/mL}$, respectively, to quantitatively investigate the cellular uptake efficacy. The cellular uptake efficiency of C6-loaded NPs, which was similar to the C6-loaded pD-NPs at all NPs concentration, was only 87.83%, 78.19%, and 72.12% of the C6-loaded Apt-pD-NPs at the concentration of 100 $\mu\text{g/mL}$, 250 $\mu\text{g/mL}$, and 500 $\mu\text{g/mL}$, respectively. Therefore, we consider that the high uptake efficiency can be attributed to the targeting ligands (ie, AS1411 aptamers) on the surface of the NPs. The specific interactions caused by AS1411 aptamers reinforce that the endocytosis of NPs was resulted in the fluorescence intensity of cells treated with the C6-loaded Apt-pD-NPs. The endocytosis of the C6-loaded NPs, the C6-loaded pD-NPs, and the C6-loaded Apt-pD-NPs was

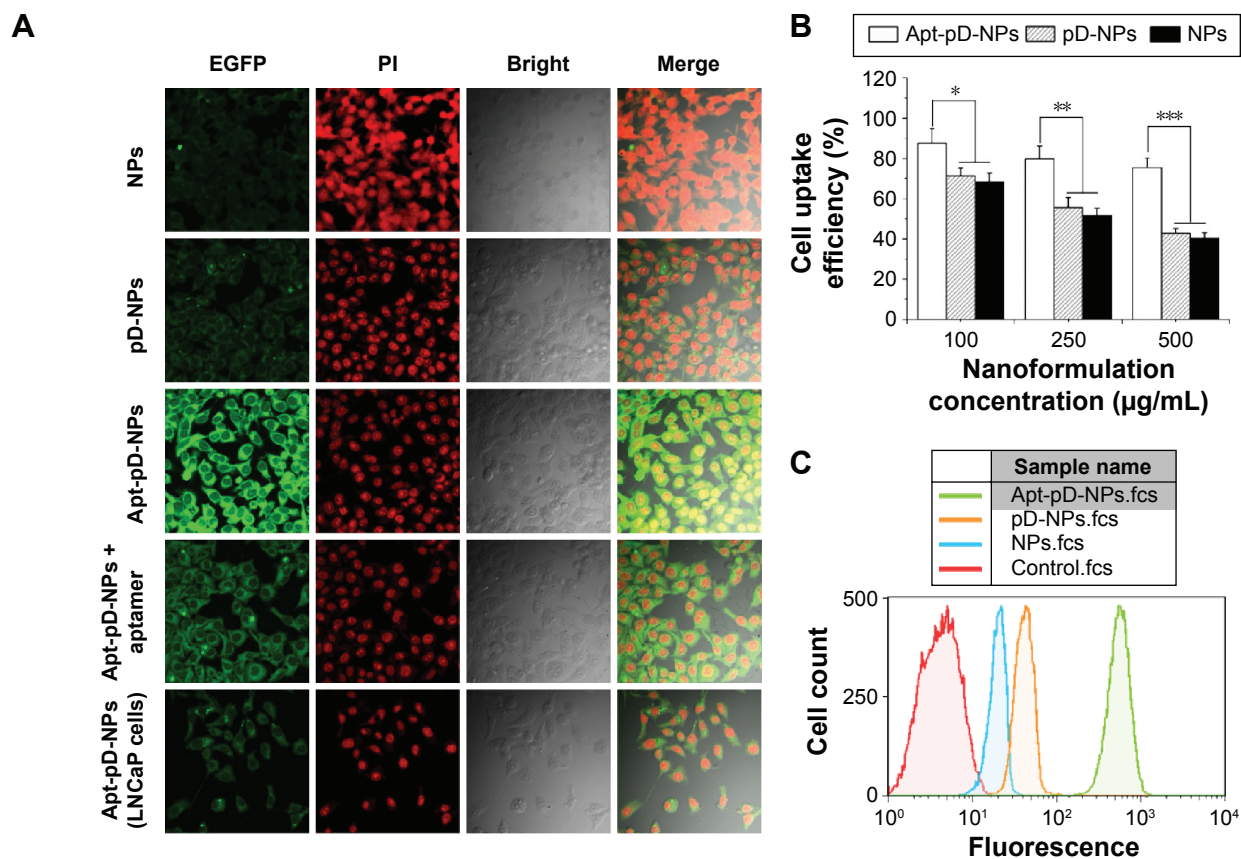


Figure 7 Endocytosis of C6-loaded NPs, C6-loaded pD-NPs, and C6-loaded Apt-pD-NPs.

Notes: (A) CLSM images of HeLa and LNCaP cells after 2 hours incubation at 37°C. (B) Cellular uptake efficiency of all kinds of NPs in HeLa cells after 2 hours incubation at 37°C (Student's *t*-test, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$). (C) FCM histograms for all kinds of NPs in HeLa cells after 1 hour incubation at 37°C.

Abbreviations: C6, coumarin-6; NPs, nanoparticles; pD, polydopamine; Apt, aptamer; CLSM, confocal laser scanning microscope; FCM, flow cytometry method; EGFP, enhanced green fluorescent protein; PI, propidium iodide.

further confirmed by flow cytometry method assay with similar results shown in Figure 7C.

Effects of NPs on cell viability

MTT assay was used to evaluate the cytotoxicity (in vitro cell viability) of DTX-loaded NPs, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs. Taxotere, a kind of DTX formulation that is clinically available, was chosen as the reference.

After excluding any contamination, HeLa cells were treated at the concentration of 0.25 $\mu\text{g/mL}$, 2.5 $\mu\text{g/mL}$, 12.5 $\mu\text{g/mL}$, and 25 $\mu\text{g/mL}$ equivalent DTX concentrations for 24 hours, 48 hours, and 72 hours, respectively. We can conclude from Figure 8 that 1) the cellular viability decreased with the increase of incubation time for both Taxotere and nanoformulations, exhibiting a dose-dependent and time-dependent effect, especially for the DTX-loaded Apt-pD-NPs;

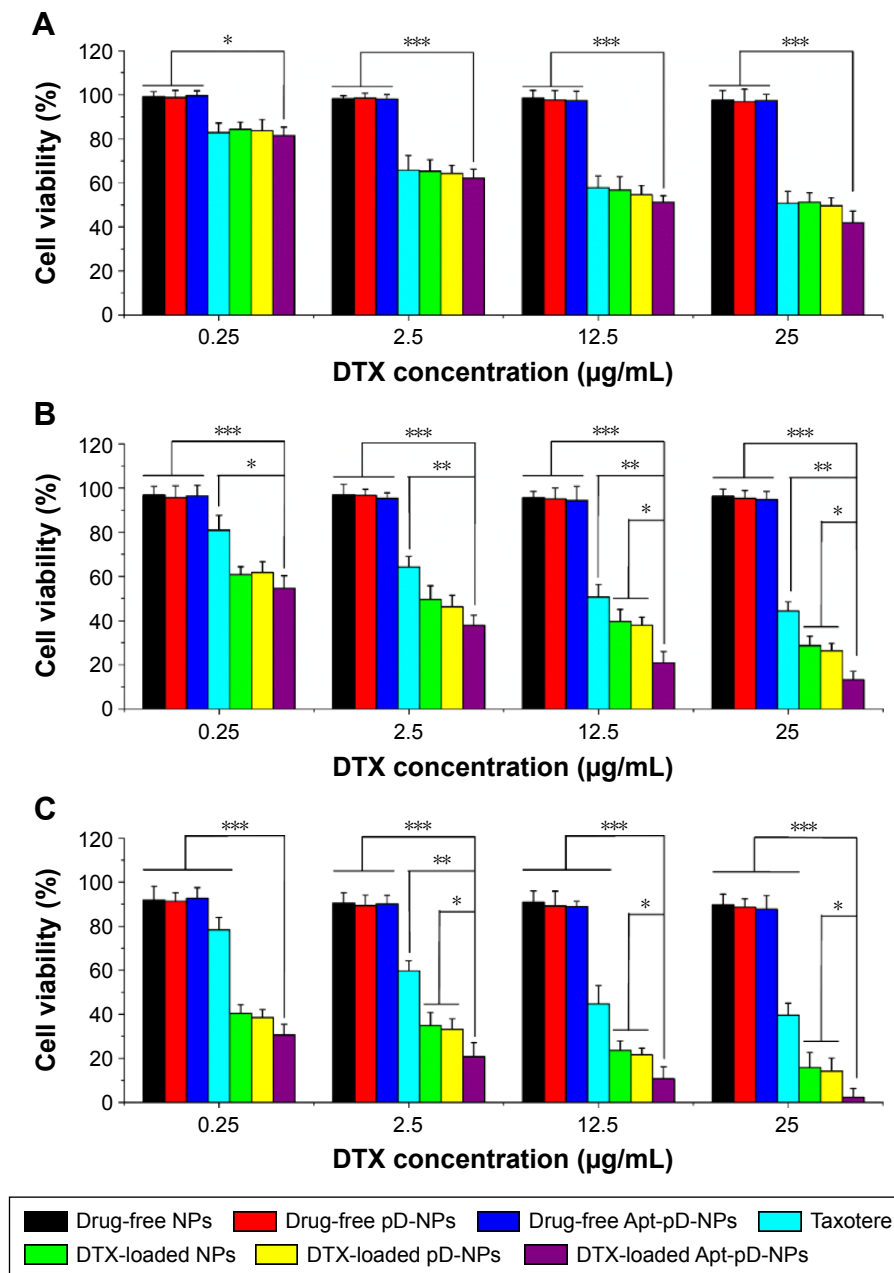


Figure 8 Cytotoxicity of all kinds of NPs detected by MTT assays (n=5).

Notes: Viability of HeLa cells cultured with DTX-loaded Apt-pD-NPs in comparison with that of Taxotere[®] at the same DTX dose and that of DTX-loaded NPs and pD-NPs with the same amount of NPs for (A) 24 hours, (B) 48 hours, and (C) 72 hours (Student's *t*-test, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$). Drug-free NPs, pD-NPs, and Apt-pD-NPs were taken as control.

Abbreviations: NPs, nanoparticles; MTT, 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; DTX, docetaxel; Apt, aptamer; pD, polydopamine.

2) none of the drug-free NPs exhibited obvious cytotoxicity to HeLa cells at various concentrations from 0.25 µg/mL to 25 µg/mL, illustrating that both the copolymer and the modification of NPs (pD coating and conjugation of aptamers) were biocompatible and nontoxic to cells and tissues; 3) the DTX-loaded Apt-pD-NPs exhibited better in vitro antitumor efficacy than Taxotere, the DTX-loaded NPs, and the DTX-loaded pD-NPs on HeLa cells. For example, the HeLa cellular viability (72 hours, 12.5 µg/mL) was 41.93% for Taxotere, 21.62% for the DTX-loaded NPs, 19.26% for the DTX-loaded pD-NPs, and 11.61% for the DTX-loaded Apt-pD-NPs. 4) The DTX-loaded NPs and the DTX-loaded pD-NPs showed similar cytotoxicity at various concentrations from 0.25 µg/mL to 25 µg/mL, further indicating that the pD film on the surface of the NPs did not influence the cell viability, being biocompatible and nontoxic.

In comparison with clinically available Taxotere, the DTX-loaded NPs, and the DTX-loaded pD-NPs, the cytotoxicity of HeLa cells after 72 hours of incubation with the DTX-loaded Apt-pD-NPs was 36.79%, 10.78%, and 6.75% larger at the DTX concentration of 25 µg/mL, indicating that the aptamers may have an outstanding targeting effect on the NP surface. Hence, targeted DTX-loaded Apt-pD-NPs could be used as a promising drug delivery system with their extremely high-efficiency in vitro therapeutic effects and in vivo treatment.

IC₅₀ value, which is defined as the drug inhibitory concentration causing 50% tumor cell mortality in a designated period, could be used to quantitatively evaluate the in vitro therapeutic effects of a pharmaceutical formulation.³⁴ Table 2 shows the IC₅₀ values of HeLa cells after 24 hours, 48 hours, and 72 hours incubation with clinically available Taxotere, the DTX-loaded NPs, the DTX-loaded pD-NPs, and the DTX-loaded Apt-pD-NPs (calculated from Figure 8). As revealed in Table 2, the IC₅₀ values of the DTX-loaded NPs and the DTX-loaded pD-NPs for HeLa cells were

19.89 µg/mL and 19.87 µg/mL after 24 hours incubation, respectively, 3.01 µg/mL and 3.07 µg/mL after 48 hours incubation, respectively, and 0.007 µg/mL and 0.006 µg/mL after 72 hours incubation, respectively, which was also consistent with the results obtained from cellular uptake experiments and indicated no obvious effect would be brought after pD coating. In the meantime, the IC₅₀ value of the DTX-loaded Apt-pD-NPs for HeLa cells was 14.31 µg/mL, only a little smaller than Taxotere (19.86 µg/mL) after 24 hours incubation. However, that of DTX-loaded Apt-pD-NPs was 25.60-fold smaller than that of Taxotere (0.47 µg/mL vs 12.03 µg/mL) after 48 hours incubation. The advantages in tumor inhibition of clinical Taxotere < the DTX-loaded NPs (or the DTX-loaded pD-NPs) < the DTX-loaded Apt-pD-NPs could be concluded, demonstrating significant in vitro therapeutic effects and great possibilities for in vivo treatment of the targeted DTX-loaded Apt-pD-NPs.

In vivo antitumor effects of NPs

On the basis of the earlier experiments in vitro cytotoxicity against HeLa cells, the DTX-loaded Apt-pD-NPs could be used as a promising drug delivery vector in cancer therapeutic research. Compared with saline (control group), both clinically available Taxotere and therapeutic NPs inhibited the tumor growth (shown in Figure 9A). The DTX-loaded Apt-pD-NPs had a better effect than the nontargeted DTX-loaded pD-NPs and clinical Taxotere, and these results (Figure 9A and 9B) were consistent with previous cellular uptake and cytotoxicity results. The tumor status of health and the morphology of tumors in each group on day 24 are shown in Figure 9C, and the tumor site was indicated by a dotted cycle. As shown in the figure, the size of the tumors could be directly observed and a similar conclusion could be achieved through this figure. Furthermore, only the mice treated in Taxotere group seemed to be emaciated with the protrusion of the vertebral column. No physical deterioration or unhealthy status was found in therapeutic NPs groups, indicating that less side effects would be caused by the therapeutic NPs. The survival data were studied using another batch of nude mice, and were presented as Kaplan–Meier plots and analyzed with a log-rank test. Compared with nontargeted DTX-loaded pD-NPs, DTX-loaded NPs, Taxotere, and saline, the survival time of targeted DTX-loaded Apt-pD-NPs was increased by 11 days, 10 days, 15 days, and 20 days, and the lifetime was extended by 24.44%, 21.74%, 36.59%, and 55.56%, respectively, as shown in Figure 9B, which turned out that targeted DTX-loaded Apt-pD-NPs worked significantly better than clinically used Taxotere

Table 2 IC₅₀ values of Taxotere®, DTX-loaded NPs, DTX-loaded pD-NPs and DTX-loaded Apt-pD-NPs on HeLa cells after 24 h, 48 h, and 72 h incubation

Incubation time (h)	IC ₅₀ (µg/mL)			
	Taxotere	NPs	pD-NPs	Apt-pD-NPs
24	19.86±1.62	19.89±1.51	19.87±1.64	14.31±1.27
48	12.03±0.89	3.01±0.63	3.07±0.59	0.47±0.09
72	7.84±0.76	0.007±0.002	0.006±0.004	0.0003±0.0001

Notes: IC₅₀, the drug concentration at which the growth of 50% cells was inhibited. Data presented as mean ± standard deviation.

Abbreviations: DTX, docetaxel; NPs, nanoparticles; pD, polydopamine; Apt, aptamer; h, hours.

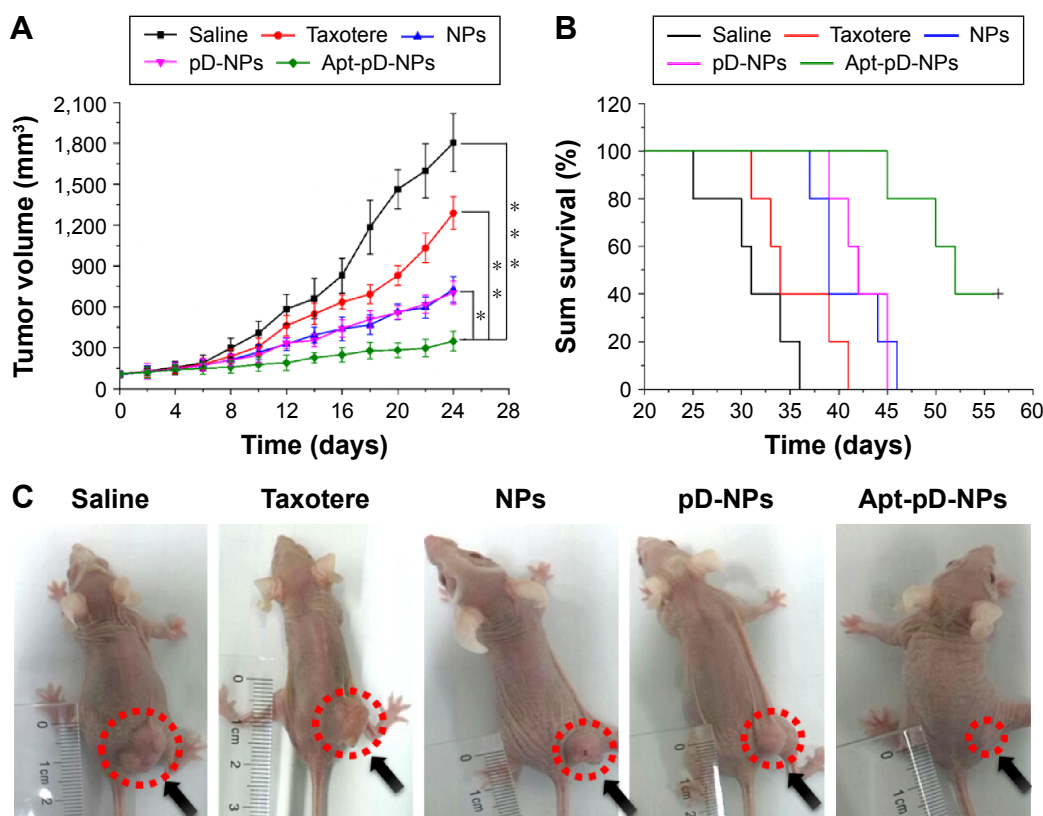


Figure 9 Antitumor efficacy of Taxotere®, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs on the SCID nude mice bearing HeLa xenograft (n=5).

Notes: (A) Tumor growth curve after intravenous injection with saline, Taxotere, DTX-loaded pD-NPs, and DTX-loaded Apt-pD-NPs (Student's t-test, * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$). (B) Kaplan-Meier survival log-rank analysis of the SCID nude mice bearing HeLa xenograft. The survival time of the animals that received DTX-loaded Apt-pD-NPs was significantly longer than that of those that received DTX-loaded NPs, DTX-loaded pD-NPs, Taxotere, and saline. (C) The status of health and the morphology of tumors in each group on day 24. The tumor site is indicated by a dotted circle.

Abbreviations: DTX, docetaxel; pD, polydopamine; NPs, nanoparticles; Apt, aptamer; SCID, severe combined immunodeficient.

and nontargeted DTX-loaded pD-NPs and DTX-loaded NPs, suggesting that DTX-loaded Apt-pD-NPs could be used as a promising and high-efficiency drug delivery system for cervical cancer therapy. In addition, the *in vivo* antitumor effect of DTX-loaded Apt-pD-NPs reported here was better than our previous studies using DTX-loaded M-PLGA-TPGS NPs,²⁴ DTX-loaded CA-(PCL-*ran*-PLA)-*b*-PEG_{1k} NPs,³⁵ or DTX-loaded H₄₀-PLA-*b*-TPGS³⁶ in tumor xenograft model.

Conclusion

In this work, we have successfully synthesized a kind of novel and robust DTX-loaded Apt-pD-NPs with star-shaped M-PLGA-TPGS copolymers for the first time, which was used as a promising and high-efficiency drug carrier for cervical cancer therapy. The M-PLGA-TPGS NPs were simply functionalized by aptamers based on the facile dopamine polymerization method. The DTX-loaded Apt-pD-NPs were characterized by DLS size and zeta potential, FT-IR, XPS, TEM, SEM, and *in vitro* drug release profile successfully. Through the endocytosis of fluorescent NPs and

in vitro cellular targeting assay, it was demonstrated that the C6-loaded Apt-pD-NPs can specifically target to HeLa cells compared with C6-loaded NPs and C6-loaded pD-NPs. The *in vitro* cytotoxicity assay showed that the DTX-loaded Apt-pD-NPs significantly inhibit cell proliferation in comparison with DTX-loaded NPs and DTX-loaded pD-NPs. The *in vivo* antitumor assays showed enhanced therapeutic effects of the DTX-loaded Apt-pD-NPs as well as the improved survival rate and reduced side effects. All data obtained from the *in vivo* assays were consistent with the *in vitro* experiments, suggesting that the robust DTX-loaded Apt-pD-NPs could be used as potential nanotherapeutics for cervical cancer treatment and the aptamer-polydopamine modification strategy could be a promising method for active targeting of cancer therapy with simple procedures.

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Disclosure

The authors report no conflicts of interest in this work.

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