



# **Review The Functional Roles and Applications of Immunoglobulins in Neurodegenerative Disease**

# Kyu-Young Sim, Kyeong Chan Im and Sung-Gyoo Park \*

School of Life Sciences, Gwangju Institute of Science and Technology (GIST), Gwangju 61005, Korea; rbdud0511@gist.ac.kr (K.-Y.S.); ikc412@gm.gist.ac.kr (K.C.I.)

\* Correspondence: sgpark@gist.ac.kr; Tel.: +82-62-715-2511; Fax: +82-62-715-2484

Received: 24 June 2020; Accepted: 24 July 2020; Published: 26 July 2020



**Abstract:** Natural autoantibodies, immunoglobulins (Igs) that target self-proteins, are common in the plasma of healthy individuals; some of the autoantibodies play pathogenic roles in systemic or tissue-specific autoimmune diseases, such as rheumatoid arthritis and systemic lupus erythematosus. Recently, the field of autoantibody-associated diseases has expanded to encompass neurodegenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD), with related studies examining the functions of Igs in the central nervous system (CNS). Recent evidence suggests that Igs have various effects in the CNS; these effects are associated with the prevention of neurodegenerative disease (AD and PD), focusing on the target antigens and effector cell types. In addition, we review the current knowledge about the roles of these antibodies as diagnostic markers and immunotherapies.

**Keywords:** immunoglobulin; Alzheimer's disease; Parkinson's disease; diagnostic marker; immunotherapy

## 1. Introduction

Antibodies bind to various foreign antigens (e.g., bacterial components and products, viruses, protozoa, and fungi) that enter the circulatory system of both humankind and animals. However, some antibodies bind to self-molecules such as cellular components (including nucleic acids, phospholipids, and proteins) in healthy individuals; these are referred to as natural antibodies or autoantibodies. The majority of natural autoantibodies are immunoglobulin (Ig) M class; as such, they are polyreactive and bind several unrelated antigens with different affinities, thereby contributing to homeostasis of the immune system. However, the adaptive immune responses are mediated primarily by high-affinity, somatically mutated IgG antibodies [1,2]. As with B-2 cells, B-1 cells (which are the main cell type that produces IgM isotype natural autoantibodies) have a mechanism for somatic hypermutation and class-switching. Therefore, the ability to bind to self-antigens can be a template for emergence of high-affinity IgG antibodies that recognize self-antigens [2–4]. Therefore, IgG autoantibodies are also present in the plasma of healthy individuals; these IgG autoantibodies have personal specific signatures that tend to be stable over time [5]. Newborns share a universal immune profile with respect to the IgM repertoire; by contrast, IgG autoantibody repertories are highly diverse and shared between the mother and newborn. This suggests that IgG signatures change according to personal immune "experience" [6]. Naturally occurring autoantibodies may provide clues regarding disruption of immune homeostasis and autoimmune diseases associated with recognition of autoantigens.

In some diseases, autoantibodies develop before clinical manifestations of autoimmune disease appear; examples include Sjögren's syndrome (SS), autoimmune hepatitis, multiple sclerosis, systemic lupus erythematosus (SLE), rheumatoid arthritis (RA), and primary biliary cirrhosis. The roles

of autoantibodies in the pathology of these diseases are different; indeed, these antibodies can have diverse effects against the same antigen depending on the target epitope [2,7–11]. Some autoantibodies in the sera of autoimmune disease patients exert various functions; for example, they act as pathogenic molecules that mimic hormone stimulation of receptors, block neural transmission by binding to receptors, affect signaling pathways, lyse cells, and induce inflammation at the site of autoantibody binding [8]. By contrast, some autoantibodies against autoantigens exposed during cell death increase phagocytosis of dead cells by forming a cell synapse between the phagocyte and the dead cell to induce engulfment [12]. Autoantibodies binding to the surfaces of necrotic cells from the serum of SLE patients increase phagocytic activity through complement component C4 [13]. Anti-dsDNA antibodies in sera from SLE patients participate in phagocytosis of the apoptotic cells by opsonizing the target cells [14]. Antiphospholipid antibodies also opsonize apoptotic cells by enhancing recognition of phagocytes [15]. In addition, apoptotic cell engulfment activates immunological signals that inhibit release of proinflammatory cytokines and induce an anti-inflammatory state in innate immune cells [12,16]. Therefore, autoantibodies can be used as biomarkers, thereby providing the opportunity to develop diagnostic tools and immunotherapies [2,7,8].

The concept of the brain as a site of immune privilege has been revised; we now know that immune cells provide immune surveillance within the central nerve system (CNS). Thus, research into the adaptive immune system is expanding into the CNS [17]. Mass cytometry of the mouse brain characterized the various resident and infiltrating immune populations in the brain compartment. The results indicate that small but significant numbers of immune cells, such as T cells, B cells, dendritic cells, and natural killer (NK) cells, migrate into the choroid plexus and meninges [18]. Lymphatic vessels in the brain have functional characteristics that include transportation of both fluid and immune cells; these vessels are connected to the deep cervical lymph nodes [19]. Therefore, many studies have examined the contributions of adaptive immune systems to neurodegenerative diseases such as Alzheimer's disease (AD) and Parkinson's disease (PD), which are related to neuroinflammation [20].

Recent clinical evidence shows that autoantibodies play roles in disease; therefore, the concept that neurodegenerative diseases may have an autoimmune etiology has been suggested [21–23]. Due to recent advances in technology related to antibody screening, many studies are attempting to identify autoantibodies associated with neurodegenerative diseases [24,25]. Some studies have identified autoantibodies that can either accelerate or prevent neurodegeneration, depending on the target antigens and cell types involved. Several reports show that Igs may be potential markers (on which we can base diagnostic techniques) and agents or targets for immunotherapy. In this review, we summarize the functional roles of Igs in the CNS, focusing on neurodegenerative diseases AD and PD, as well as their potential utility as diagnostic markers and immunotherapy agents or targets.

#### 2. Role of Igs in AD

#### 2.1. Alteration of Adaptive Immune Responses in AD

AD is a degenerative disease of the brain, accounting for an estimated 60–80% of dementia cases. AD causes difficulties with memory, language, and problem-solving due to damage or destruction of nerve cells. The major hallmarks of AD are accumulation of beta-amyloid (A $\beta$ ) around neurons and tau phosphorylation inside neurons [26], which are accompanied by neurinflammation in the CNS [20]. A pathogenic form of A $\beta$ , A $\beta_{1-42}$ , induces A $\beta$  amyloid fibril formation; these accumulated A $\beta$  amyloid fibrils in turn induce formation of senile plaques, resulting in neurotoxicity and induction of tau pathology. Tau is a microtubule-associated protein involved in microtubule polymerization and structural stabilization; however, the pathogenic form of tau forms aggregates and fibril seeds that damage the cell [27].

Recent research has focused on the contribution of the adaptive immune system to AD pathogenesis. One study shows that the number of clonally expanded  $CD8^+$  T effector memory  $CD45RA^+$  cells in

the blood of AD patients is increased, and that these cells patrol the cerebrospinal fluid (CFS) [28]. Lymphocyte profiling has revealed a significant decline in CD4<sup>+</sup> T cell populations in the CSF in brain regions in which A $\beta$  is deposited. By contrast, the A $\beta$  burden shows a positive correlation with increased numbers of memory B cells in the CSF; this is exacerbated in *APOE*  $\varepsilon$ 4 carriers [29]. Increased numbers of double-negative (IgD<sup>-</sup>CD27<sup>-</sup>) memory B cells and a reduction in the number of naïve B cells (IgD<sup>+</sup>CD27<sup>-</sup>) in the peripheral blood have been identified in AD [30], and the number of cells producing antibodies targeting A $\beta$ <sub>1-42</sub> is increased in AD [31]. These studies suggest a functional role for the adaptive immune system in AD pathogenesis, particularly with respect to B cell function [32].

#### 2.2. Evidence for an Association between Ig Responses and AD

Clinical evidence suggests that the Ig response is associated with AD pathogenesis. Ig labeling shows that Ig<sup>+</sup> neurons have neurodegenerative apoptotic characteristics that are not observed in Ig<sup>-</sup> neurons [33]; Ig binding to apoptotic debris may induce apoptotic cell clearance [12,14,15]. Additionally, C1q and C5b-9 were detected in these Ig<sup>-</sup> positive neurons [22]. In the brain of human AD patients immunized (or not) with AN-1792, IgG was detected in Aβ plaques, on plaques surrounding microglia, and within neurons adjacent to plaques; antibody deposition correlated with the C1q load [34]. Biochemical studies provide evidence for an interaction between IgG and tau protein, supporting a pathological role for Igs in AD brains [35]. These data support the autoimmune disease theory for AD; blood-brain barrier (BBB) dysfunction in AD allows autoantibodies access to targets in the brain, which leads to autoimmunity-induced neural cell death [22]. In addition, changes of autoantibody levels in blood and CSF have been identified. The blood of AD patients contains higher levels of  $A\beta$ -IgG immune complexes than that of controls. In addition, the levels of A $\beta$ -IgG complexes correlate negatively with performance in cognitive tests [36]. Antibodies specific for tau protein and heavy neurofilaments are increased in AD patients [37]. Moreover, elevated plasma levels of nicotinic acetylcholine receptor  $\alpha 7$  ( $\alpha 7$  nAChR)-specific autoantibodies are characteristic of early AD patients [38]. The neuronal  $\alpha$ 7 nAChRs are widely distributed in the CNS and are involved in PI3K/Akt signaling, which protects brain cells from apoptotic signals [39,40]. The  $\alpha$ 7 nAChRs interact directly with A $\beta$  peptide and stimulate the neuroprotective pathways that protect against A $\beta$ -toxicity [41]. Studies in an lipopolysaccharide injected mice model show that  $\alpha$ 7 nAChRs-specific antibodies aggravate neuroinflammation by stimulating proinflammatory cytokines and downregulating anti-pro-inflammatory miRNAs, which are responsible for limiting inflammatory signals in the CNS [42]. An autoantibody screening study identified autoantibodies in CSF that target glia-derived nexin, actin-interacting proteins, metalloproteinase inhibitor 2, quinone oxidoreductase, inositol trisphosphate receptor 1, and endoplasmic reticulum calcium ATPase 2 [43]. The levels of autoantibodies targeting ATP synthase, angiotensin II type 1, and 5-hydroxytryptamine also are altered in AD [44]. The concentration of IgM and IgG autoantibodies targeting alpha B-crystallin is increased in AD patients [45]. These data suggest that some types of Ig have functional roles related to AD pathogenesis.

#### 2.3. BBB Breakdown and Ig Infiltration of the CNS

The brain has a lymphatic drainage system for clearing waste, which includes abnormal proteins that contribute to neurodegeneration [46,47]; in addition, meningeal lymphatics vessels provide a direct pathway by which CSF components and immune cells drain to the cervical lymph nodes [48]. Therefore, neuronal antigens could be presented to immune cells in cervical lymph nodes. However, as antineuronal surface antibodies are rarely found in the CSF and blood of healthy participants, it seems that production of neuronal surface antibodies is induced by neuronal inflammatory conditions [49].

Some evidence suggests that Igs in the CNS have infiltrated from the blood. A study of CSF and blood from hip fracture patients showed that CSF and blood contain similar autoantibody repertoires [50]. However, the amounts of each antibody type in the CSF were far lower than those in blood [50,51]. The similarity of the autoantibody repertoires and the lower amounts of antibodies in

the CSF suggest that blood Igs infiltrate the CNS. Additionally, low levels of IgG were observed in the adult human cortex; in particular, diffuse patterns were observed around blood vessels, but extending into the brain parenchyma [52].

BBB breakdown may result in greater infiltration of the brain by Igs. BBB breakdown was identified in AD post-mortem human studies measuring perivascular accumulation of blood-derived fibrinogen, Ig, thrombin, albumin, and loss of BBB tight junctions. In addition, genetic risk factors for AD, such as *APOE*, *APP*, and *PSEN1*, induce BBB breakdown in animal models [53–56]. A study revealed that physical BBB breakdown induced by needle insertion can trigger selective IgG localization in the brain parenchyma at the site of injury; in addition, IgG antibodies from neuromyelitis optica spectrum disease patients induce astrocyte pathology in this region [57]. These studies suggest that BBB breakdown in AD results in deposition of Igs in the brain. In addition, failure of the lysosome system in AD could be another possible factor that causes antibody deposition in the brain [58,59].

#### 2.4. Protective Role of Natural Antibodies in AD

Many studies have identified functional roles for Igs in AD. However, the function of Igs depends on the target antigen and CNS cell type. Some reports suggest a protective role for IgG in AD. Studies identified an increase in IgG associated with microglia in the brains of AD transgenic mice [60,61]. One study used the immune-deficient AD mouse model, which lacks B, T and NK cells, to examine the functional role of Igs. They noted several changes in the immune-deficient AD mouse model, including increased levels of  $A\beta$  and several proinflammatory cytokines, and a shift in microglial phenotype. Delivery of normal mouse IgG to mice via either bone marrow transplantation or direct stereotactic injection led to a marked reduction in AD pathology. The Src/spleen tyrosine kinase (Syk)/phosphatidylinositol 3-kinase signal transduction pathway is involved in the IgG-induced uptake of A $\beta$  by microglia [60]. Another study showed that stress granules (SGs) containing Syk and phosphotyrosine are prevalent in the brains of AD patients. These SGs modulate Syk and cause microglial cell dysfunction with respect to phagocytosis of *Escherichia coli* or Aβ fibrils. Phagocytic activity was restored by treatment with non-specific rabbit IgG, suggesting a mechanism that explains the therapeutic efficacy of intravenous IgG [62]. Another research study examined how AD risk factors (APOE genotype, aging, and gender) affect IgG levels in various brain regions, especially in AD-susceptible regions, such as the hippocampus and cortex. IgG was detected mainly on microglia and some neurons, but not astrocytes [61]. These data show that Igs might prevent A $\beta$  pathology by increasing phagocytosis by microglial cells, leading to increased clearance of A $\beta$ . Another study showed that human IgG prevents A $\beta$ -mediated neurotoxicity by inhibiting A $\beta$  aggregation. The protective effects of IgG seem to be independent of IgG specificity; the Fab region is responsible for inhibiting Aß aggregation, and bioinformatics tools show that the Fab regions bind to A $\beta$  [63]. These studies show that non-specific IgG may also inhibit  $A\beta$  aggregation.

Antibodies specific for some epitopes are protective against AD. Natural antibodies specific for toxic A $\beta$  and amyloidogenic non-A $\beta$  species are present in blood and CSF samples from both AD patients and healthy controls. IgG targeting oligomeric A $\beta_{1-42}$  declines with age and advancing AD, and IgG isolated from the plasma of AD patients or healthy controls protects primary neurons from A $\beta$ -induced toxicity [51]. These results suggest that natural Igs, even in healthy individuals, protect against AD pathogenesis. Thus, administering these antibodies to the elderly may prevent AD (Table 1).

#### 2.5. Pathogenic Natural Antibodies in AD

By contrast, some research shows that Igs have pathogenic effects. The Fc gamma receptor (Fc $\gamma$ R) is a receptor for IgG that has both activating and inhibitory activities [75,76]. Multiple cell types within the CNS (e.g., neurons and astrocytes) express Fc $\gamma$ Rs, and various stimuli (e.g., immune complexes) can activate expression of Fc $\gamma$ Rs [67,76–78]. Fc $\gamma$ Rs participate in uptake of Igs and increase calcium concentrations and neurotransmitter release from motor neurons [79]. IgG immune complexes increase

phosphorylation of extracellular signal-regulated kinase and levels of intracellular calcium, and induce  $Fc\gamma R$ -mediated internalization of IgG in cortical and hippocampal cells [77]. In addition, IgG immune complexes activate sensory neurons directly [68]. These findings suggest that IgG immune complexes affect neuronal function via direct interactions with  $Fc\gamma R$  expressed by non-immune cells in the brain.

ia [60–6 euron [63] l cell	Effector Cell Microglia	Antibody Target	F ('		
euron [63]	Microglia		Function	Disease	
l cell [63]		Non-specific	Enhance the phagocytosis of $A\beta$	AD	
	Cortical Neuron Endothelial cell	Fab-mediated function	Inhibit Aß aggregation		
euron [51]	Primary Neuron	Oligomeric Aβ <sub>1-42</sub>	Neutralize A <sub>β</sub> -induced toxicity		
ia [64]	Microglia	Tau	Reduce tau toxicity		
n [65]	Neuron		Neutralize the intracellular α-syn aggregation	PD	
ia [66]	Microglia	α-syn	Remove the extracellular α-syn proteins		
			Prevent $\alpha$ -syn transfer between neuron and astrocyte		
		Pathogenic Role	I		
Cell Refere	Effector Cell	Antibody Target	Function	Disease	
	FcγR-expressing cells in the CNS	FcyR-mediated function	Induce the neuroinflammation as Immune complexes	AD	
70	Neuron Endothelial cell	Ceramide	Increase Aβ plaque formation		
ia [71]	Microglia	FcyR-mediated function	Release the proinflammatory cytokines		
[64]		Tau	Release the proinflammatory cytokines		
ia [72,73	Microglia	FcyR-mediated function	Induce neuronal damage via microglial activation	PD	PD
n [74]	Neuron	-	Promote the $\alpha$ -syn transfer		
י ז [	Neuron		,		

Table 1. The potential roles of autoantibodies in neuronal degenerative diseases.

AD: Alzheimer's disease; Aβ: beta-amyloid; PD: Parkinson's disease; α-syn: α-synuclein; FcγR: Fc gamma receptor.

Increased expression of FcyRs in human brain tissue from individuals with neurodegenerative diseases, such as AD and PD, along with ligation of specific FcyRs in the CNS by IgG immune complexes and alternate ligands, promotes neuroinflammation and increases neurodegeneration [67,69,78]. For example, neuronal cells exposed to synthetic A $\beta$  and the hippocampus of AD brains both show upregulation of FcyRIIb, which activates endoplasmic reticulum stress and caspase-12; knockout of Fc $\gamma$ RIIb induces resistance to synthetic A $\beta$ -induced cell death in vitro. Moreover, genetic depletion of FcyRIIb rescued memory impairment in an AD mouse model [80]. A study using the APOE-deficient mouse showed upregulation of Fc receptors, predominantly type IV, and increased amounts of serum and brain IgGs compared with controls. Interestingly,  $Fc\gamma R$  engagement induces A $\beta$  accumulation in neurons, but genetic deletion of the  $\gamma$ -chain of activating Fc receptors reduces learning and memory impairment. In vitro tests show that activation of Fc receptors increases expression of mitogen-activated protein kinases and  $\beta$ -site amyloid precursor protein cleaving enzymes, which cause hyperphosphorylation of tau, A $\beta$  accumulation, and synapse loss in primary neurons [81]. This suggests that immune complexes induce FcyR overexpression by neurons and that activation of neuronal FcyRs induces AD-like pathology. Some antibodies can have pathogenic effects in AD, depending on the target antigen. AD model mice show age-dependent increases in expression of antibodies targeting

ceramide, suggesting that autoimmune reactions against ceramide play a role in AD pathogenesis. In addition, induction of serum anticeramide IgG by ceramide administration increases the plaque burden in female mice compared with controls, suggesting that systemic anticeramide IgG and exosome levels correlate with increased plaque formation [70].

Activation of microglia also has functional effects on neurodegeneration. Activation of microglia increases phagocytosis, clearance, and degradation of A $\beta$ , which prevents formation of A $\beta$  plaques in the brain. However, prolonged activation of microglia leads to release of proinflammatory cytokines, which initiate a proinflammatory cascade [71]. Immune complexes play important roles in IgG-mediated phagocytosis by microglia, and these cells are also affected by cytokines in the CNS [82]. However, immune complexes also induce inflammatory reactions in the brain parenchyma and induce neuronal tissue damage indirectly through activation of Fc $\gamma$ R on microglia [69]. IgG in lupus serum induces M1 polarization of brain microglia. IgG antibodies bind to microglia through the Fc region; then, the B cell activating factor in lupus serum upregulates expression of Fc $\gamma$ R on the surface of microglia, thereby triggering Fc $\gamma$ R-mediated signaling pathways to induce inflammatory responses [83]. Another study revealed that tau antibodies (both with and without full effector function) in the presence of microglia protect neurons from tau toxicity, but effector tau antibodies induce release of proinflammatory cytokines by microglia [64].

IgG from AD and PD are more toxic to neurons than control IgG [72,84]. In addition, disease alters IgG Fc sugar moieties, known as *N*-glycans, which affect the affinity of the Fc domain [85,86]. Therefore, antibody function is complex; in particular, function can differ depending on environmental conditions, effector cell type, and target antigen (Table 1). Therefore, it is necessary to comprehensively consider various factors, including Fc $\gamma$ R-IgG ligation, when developing treatment modalities based on antibodies. Some studies show that anti-A $\beta$  and -tau antibodies with no effector function can inhibit AD pathogenesis, suggesting utilization of immunotherapies based on modified antibodies that lack adverse effects [64,87,88]. Figure 1 summarizes a model of antibody-mediated pathogenesis in neurodegenerative disease.

#### 2.6. Diagnostic Application of Antibodies for AD

As the number of people developing AD is expected to increase, we need to develop low-cost and patient-friendly diagnostic techniques based on blood biomarkers. Autoantibodies are one candidate blood biomarker; indeed, anti-A $\beta$  and -tau protein antibodies have been used as biomarkers [44]. Autoantibodies targeting A $\beta$  have been studied by many groups. A meta-analysis of 2901 individuals (1311 AD patients and 1590 healthy control subjects) revealed a significant increase in the amount of endogenous IgG autoantibodies targeting A $\beta$  in the blood; by contrast, anti-A $\beta$  IgM autoantibody levels were markedly lower in patients with AD than in control subjects, suggesting the possibility that alterations in autoantibody levels may be a biomarker [89]. With respect to anti-tau antibodies, several groups have measured concentrations in the blood. However, there were no statistically significant differences between healthy controls and AD patients [90]. This lack of difference was also shown in another study [91].

Various technological advances related to high-throughput antibody screening have meant that many studies are attempting to identify other antibodies that may be blood biomarkers for AD. Microarrays for screening of autoantibodies have identified a combination of autoantibodies in blood that can differentiate patients with mild cognitive impairment from age-matched controls with high accuracy. The target proteins are a variety of receptors, adapters, kinases, cytoskeletal components, potassium channel subunits, ribosomal and mitochondrial proteins, and other accessory proteins [25]. Furthermore, 44 autoantibodies from the panel enrolled in a previous study can distinguish prodromal AD pre-surgically and with high accuracy in individuals admitted to the hospital for hip fracture repair surgery [92]. In addition, various protein and peptide arrays have led to identification of autoantibodies specific to patients with AD [4,93]. These antibodies target proteins involved in synaptic activity, including neurotransmitters [94,95] and receptors [96], and autoantibodies specific for proteins

modulating inflammation [95] and the BBB [96]. The levels of autoantibodies targeting proteins involved in energy metabolism, such as aldolase and adenosine triphosphate synthase  $\beta$ , are higher in the serum of patients with AD than in controls [93]. This suggests that the antibody repertoire responsible for AD pathogenesis changes; such changes could be utilized as blood biomarkers for AD. However, actual application would require clear criteria for calculating clinical scores on which a diagnosis of AD would be based.



**Figure 1.** A model showing antibody-mediated pathogenesis of Alzheimer's disease (AD) and Parkinson's disease (PD). Changes in brain conditions in AD and PD (for example, cell death, inflammation, and aggregation of abnormal proteins) promote the outflow of neo-antigens through lymphatic vessels, causing changes in antibody repertoires. Damage to the blood–brain barrier (BBB) generated in AD and PD induces infiltration by antibodies. Antibodies activate FcR expression by brain cells, thereby promoting pathogenesis of AD and PD in the brain. A $\beta$ : beta-amyloid;  $\alpha$ -syn:  $\alpha$ -synuclein.

#### 2.7. Therapeutic Application of Antibodies in AD

Several immunotherapies have been developed using antibodies against A $\beta$ , the major marker of AD. Two immunotherapeutic strategies, active immunization and passive immunization, have been used as immune therapies [76]. Passive immunotherapy with antibodies against A $\beta$  is based on binding of transferred IgG to A $\beta$ , which may prevent or inhibit AD pathogenesis. However, the lack of clinical benefit has led to repeated failures of clinical trials. Phase 3 clinical trials (NCT01900665) revealed that solanezumab, a monoclonal antibody against A $\beta$ , showed no significant benefit with respect to preventing cognitive decline in patients with mild AD [97]; this failure suggests that earlier treatment may be critical for cognitive benefits. Phase III trials (NCT00667810 and NCT00676143) also confirmed that bapineuzumab (as a passive vaccination for mild to moderate AD) lacks efficacy [98]. A study of gantenerumab was halted early due to lack of a positive effect (NCT01224106) [99], but high doses of gantenerumab did result in robust A $\beta$  plaque removal (NCT01224106 and NCT02051608) [100]. An active vaccine against A $\beta$  was also used to stimulate antibody production. One of these, AN-1792, comprises synthetic full-length A $\beta$  peptide plus a QS-21 adjuvant. Long-term follow-up

patients immunized with AN1792 revealed sustained anti-AN1792 antibody titers and a marked reduction in functional decline and plaque numbers [101,102]. However, the vaccination caused meningoencephalitis in some cases [103]. Nevertheless, to date, researchers are developing and testing several possible interventions in clinical trials; these are based on anti-A $\beta$ , anti-tau antibodies, and agents that prevent neuroinflammation (Table 2) [76,104].

Drug	Antibody Type (Target)	FDA Status	NCT (Patients)
Solanezumab	Passive (Mid-domain of the Aβ peptide)	Phase 3	NCT02008357 (Older individuals who may be at risk for memory loss) NCT02760602 (Prodromal AD)
Bapineuzumab	Passive (N-terminal region of Aβ)	Discontinued	NA
Gantenerumab	Passive (Conformational epitope on Aβ fibrils)	Phase 3	NCT03443973 and NCT03444870 (Early AD)
Crenezumab	Passive (Multiple forms of aggregated Aβ)	Phase 2	NCT02670083 (Prodromal to mild AD) NCT01998841 (Autosomal-dominant AD) NCT01397578 (Mild to moderate AD)
Aducanumab	Passive (Conformational epitope on Aβ)	Phase 3	NCT02477800 and NCT02484547 (Early AD)
BAN2401	Passive (Large, soluble Aβ protofibrils)	Phase 3	NCT01767311 and NCT03887455 (Early AD)
AN-1792	Active (Synthetic full-length Aβ peptide with QS-21 adjuvant)	Discontinued	NA
Gosuranemab	Passive (N-terminal fragments of tau)	Phase 2	NCT03352557 (Early AD)
AADvac1	Active (Synthetic tau peptide withaluminum hydroxide adjuvant)	Phase 2	NCT01117818 (Early AD)
Prasinezumab	Passive (Against aggregated α-syn)	Phase 2	NCT02157714 and NCT03100149 (PD)
BIIB054	Passive $(\alpha$ -syn residues 1-10)	Phase 2	NCT03318523 and NCT02459886 (PD)
ABBV-0805	Passive (Oligomeric/protofibrillar α-syn)	Phase1	NCT04127695 (PD)
Activa Affitope (PD01A)	Active (Synthetic α-syn peptide with adjuvant)	Phase1	NCT02618941 (PD)

Table 2. Clinical trials of immunotherapies for neuronal degenerative diseases.

A $\beta$ : beta-amyloid; AD: Alzheimer's disease;  $\alpha$ -syn:  $\alpha$ -synuclein; PD: Parkinson's disease.

#### 3. Role of Igs in PD

#### 3.1. Changes to the Adaptive Immune System in PD

PD is a progressive neurodegenerative disease that affects 2–3% of the population aged  $\geq$ 65 years. PD results in impaired movement and cognitive problems, which become increasingly prevalent as the disorder progresses. The major hallmarks of PD are accumulation and aggregation of  $\alpha$ -synuclein ( $\alpha$ -syn) and loss of neurons in the substantia nigra (SN) [105]. Although neuroinflammation may not be the first step in PD, neuroinflammation is strongly implicated in disease pathogenesis [20]. Therefore, several studies have profiled the lymphocyte populations in PD to identify the role of the adaptive immune system. The data show that in human and mouse models of PD, there are fewer naïve T cells, CD4<sup>+</sup> Th (helper) cells, and Treg cells than in controls. Also, there is an increasing tendency toward clonally expanded CD45RA<sup>+</sup> cells, activated microglial cells, and activated T cells in the CSF of patients with PD [28,106,107]. Another study reports that the CD4<sup>+</sup> T cell population in PD has a Th1-biased profile [108]. However, there is a lack of evidence related to changes in the B cell population in PD [107].

#### 3.2. Evidence for an Association Between Ig Response and PD

PD is associated with altered Ig levels. One study identified an increased CSF/serum IgG ratio in cases of advanced PD [109]. Another showed that the subthalamic nucleus of the brain in PD patients showed widespread extravascular staining for IgG, indicating BBB impairment; however, IgG staining was restricted to the lumen of vessels in the brains of healthy controls [110]. Another study revealed that some pigmented dopamine neurons in the SN of PD patients have more IgG than those of healthy controls, and that the IgG co-localizes with  $\alpha$ -syn. IgG-positive neurons correlate negatively with the degree of cell loss and positively with the number of HLA-positive microglia. The most common subclass of neuronal IgG in the damaged SN is IgG1; in addition, the high-affinity activating IgG receptor, FcγRI, is expressed on nearby activated microglia [111]. These results suggest that Igs play functional roles related to activation of microglia, leading to loss of dopamine nigral neurons in PD.

The majority of studies of Igs have focused on changes in the amounts of  $\alpha$ -syn antibodies, but the results are inconsistent and contradictory [112,113]. However, a study that measured the affinity of these antibodies revealed that the amount of high-affinity  $\alpha$ -syn antibodies in the plasma of PD patients is lower than that in healthy controls, and that there are markedly fewer  $\alpha$ -syn-antibody immunocomplexes in the plasma of PD patients. Furthermore, cross binding of  $\alpha$ -syn antibodies to  $\beta$ - and  $\gamma$ -synuclein monomers suggests that antibodies target mainly C-terminal epitopes [114]. Another group showed that glycosylation of IgG was significantly different between patients with PD and controls. They suggest that the type of IgG glycosylations in PD decrease its capacity to inhibit Fc $\gamma$ -RIII $\alpha$  binding, so IgG may trigger antibody-dependent cell cytotoxicity and inflammation in individuals with PD [86].

#### 3.3. BBB Breakdown in PD

Similar to AD, studies report BBB breakdown in PD, which may allow antibody infiltration. One study used positron emission tomography to show BBB dysfunction in the midbrain of PD patients relative to controls [115]. Cerebral microbleeds are more common in PD patients with dementia than in those without, suggesting that the burden of cerebral microbleeds may contribute to further cognitive impairment in PD [116]. In addition, studies show markedly increased permeability of the BBB in the post-commissural putamen of PD patients [117], as well as microvascular changes in post-mortem SN samples [110]. There are high levels of  $\alpha\nu\beta3$  expression in the locus ceruleus and the SN pars compacta of PD brains, and the presence of  $\alpha\nu\beta3$  reactive vessels suggests newly created vessels that have not developed restrictive properties of the BBB [118]. This evidence suggests that the brain of PD patients is susceptible to infiltration by antibodies, as in AD.

#### 3.4. Protective Role of Natural Antibodies in PD

Igs play a bifunctional role (protecting or accelerating) in PD pathogenesis. In particular, antibodies against  $\alpha$ -syn protect against PD. One of the antibodies secreted by memory B cells in PD patients has high affinity for epitopes at the C terminus of  $\alpha$ -syn. An in vitro  $\alpha$ -syn seeding assay revealed that these antibodies neutralize the seeding of intracellular syn aggregates [65]. Immunization against  $\alpha$ -syn results in reduced  $\alpha$ -syn accumulation and synaptic loss in mouse models of Lewy body disease [66,119,120]. Antibodies against  $\alpha$ -syn induce clearance of extracellular  $\alpha$ -syn proteins by microglia through Fc $\gamma$ R binding. In addition, stereotaxic administration of antibodies into the brain prevent neuron-to-astroglia transmission of  $\alpha$ -syn, thereby inducing increased co-localization of  $\alpha$ -syn

and the antibody on microglia [66]. These findings suggest that antibodies against  $\alpha$ -syn play protective roles and can be utilized as potential therapeutic targets (Table 1).

#### 3.5. Pathogenic Role of Natural Antibodies in PD

Igs also play a role in disease pathology, principally through FcγR activation. Activation of FcyR expression by neurons induces PD pathogenesis as follows:  $\alpha$ -syn fibrils bind to FcyRIIB of neurons; Fc $\gamma$ RIIB then mediates cell-to-cell transmission of  $\alpha$ -syn via activation of Src homology region 2 domain containing phosphatase-1/2. Blocking this signaling pathway attenuates formation of Lewy-body-like inclusion bodies [74]. FcyRs are expressed on the plasma membrane of microglia; these receptors bind IgG and activate microglia, resulting in damage to dopaminergic neurons in the SN. Virus-induced overexpression of  $\alpha$ -syn in wild-type animals induces activation of NF- $\kappa$ B p65 and other proinflammatory molecules, which activate microglia and result in loss of DA neurons from the SN. However,  $Fc\gamma R^{-/-}$  mice exhibit no neuritic changes in the absence of  $\alpha$ -syn-induced neurodegeneration; these mice show attenuated microglial activation [73]. In vitro experiments show that IgG in PD activates microglia via the FcγR to induce dopaminergic cell injury. Stereotaxic injection of PD IgG into the SN of mice increases the number of microglial cells, accompanied by a 40% loss in tyrosine hydroxylase (TH)-positive neuronal cells in the SN. However, there is no significant increase in the number of microglia and no loss of TH-positive cells in  $Fc\gamma R^{-/-}$  mice [121]. In PD patients, antimyelin-associated glycoprotein (MAG)-IgM autoantibody levels are significantly higher than those in healthy control or patients with atypical parkinsonism. Furthermore, studies report increasing levels of antimyelin basic protein and antiproteolipid protein IgM autoantibodies in PD [122,123]. The MAG–IgM autoantibodies induce neuropathy, which is a common symptom of PD [124–126]. IgG from PD and recombinant human C5a act synergistically to induce selective dopaminergic neurodegeneration in rat mesencephalic neuron-glia cultures; however, IgG from disease controls and normal controls does not show dopaminergic neurotoxicity. The results from microglia-supplemented neuronal cultures indicate that microglia play pivotal roles in neurotoxicity. PD IgG and C5a act synergistically to activate microglia, which produce superoxide and nitric oxide, both of which are neurotoxic. Finally, F(ab')<sub>2</sub> fragments of PD IgG play important roles in neurotoxicity [72]. Taken together, these results suggest that FcyR-IgG interactions, especially IgG from PD patients, activate microglia, resulting in loss of neurons and activated synuclein transmission (Table 1 and Figure 1).

#### 3.6. Diagnostic Application of Antibodies in PD

Identifying blood biomarkers for PD is the goal of much research, however an effective biomarker remains elusive. However, antibodies in the blood of PD patients are promising candidate PD biomarkers. In particular,  $\alpha$ -syn IgG is a strong candidate. A study of four IgG subclasses (IgG1, IgG2, IgG3, and IgG4) found higher levels of anti- $\alpha$ -syn IgG2 and lower levels of anti- $\alpha$ -syn IgG4 in PD than in controls. Anti- $\alpha$ -syn IgM levels were also lower in PD. Increased or decreased levels of IgG subclasses are expected to be one biomarker of PD [112]. A recent study reports that the repertoire of high-affinity or -avidity IgG autoantibodies targeting a-syn is significantly reduced in PD [127]. The pathological changes associated with PD might contribute to the levels of naturally occurring  $\alpha$ -syn autoantibodies in both serum and CSF. Both of these are potential biomarkers for PD patients [128,129]. However, studies of  $\alpha$ -syn antibodies report inconsistent measurements of antibody levels. Several studies demonstrated that serum  $\alpha$ -syn antibody titers in patients are different depending on age, disease duration, severity, and genetic inheritance. The distinct pattern of pathology and symptom progression associated with some isolated naturally occurring human anti- $\alpha$ -syn antibodies might be a potent diagnostic marker for PD [65,112,113]. However, a systematic review and meta-analysis highlight many caveats to this conclusion based on the limitations of the assays used, the clinical heterogeneity of the study cohorts, the lack of longitudinal data, and poor matching of controls to patients; thus, the overall quality of the evidence is poor. Hence, the value of  $\alpha$ -syn

autoantibodies as diagnostic or prognostic biomarkers remains uncertain [113]. Another study tried to identify changes in a panel of autoantibodies in the blood of PD. They selected a panel of blood-borne autoantibodies as biomarkers, which distinguished early-stage PD subjects from controls with an overall accuracy of 87.9%; it also distinguished PD from neurological diseases. These autoantibodies target microtubule-affinity-regulating kinase 1, pseudouridine synthase-like 1, interleukin 20, and C-C motif chemokine ligand 19, among others [24].

## 3.7. Therapeutic Application of Antibodies in PD

Since  $\alpha$ -syn is a major marker for PD pathogenesis, use of  $\alpha$ -syn antibodies has been put forward as a potential immunotherapy. Anti- $\alpha$ -syn antibodies may stimulate microglial cells to scavenge extracellular  $\alpha$ -syn and prevent its transfer from one neuron to another. Current immunotherapies for PD target microglia activation, which regulates lysosome function for effective clearance from the brain [130]. A passive vaccination for PD, prasinezumab (PRX002; a humanized IgG1 monoclonal antibody), has been developed. Clinical phase 1 (NCT02157714) trials show that the antibody can regulate serum  $\alpha$ -syn levels in a dose-dependent manner [131]; thus, passive vaccination for PD is being examined in a clinical phase 2 trial (NCT03100149). Active vaccination with a synthetic  $\alpha$ -syn-mimicking peptide called Affitope (PD01A) resulted in a significant increase in PD01-specific antibody titers; these antibodies bind to both oligometric and fibrillar  $\alpha$ -syn. At week 26, there was a trend toward a reduction in the levels of oligometric  $\alpha$ -syn in plasma and CSF after treatment with PD01A. Therefore, PD01A is being prepared for a phase 2 clinical trial. A second vaccination with PD03A resulted in a clear dose-dependent immune response against the  $\alpha$ -syn-targeted epitope over time, with antibody reactivation upon booster immunization (NCT02267434); however, a parallel phase 1 study did not find any immune responses (NCT02270489) [132,133]. Multiple immunotherapeutic agents for PD are currently in clinical trials (Table 2) [134].

#### 4. Conclusions

The increased or decreased risk of AD and PD due to autoimmune mechanisms suggests that autoantibodies produced in those with autoimmune diseases can affect neurodegeneration. A previous study showed that primary SS patients have a 2.69-fold increased risk of developing AD; the risk is even higher in older people [135]. Another study found that 18 out of 25 autoimmune diseases were significantly associated with a risk of dementia [136]. Patients with autoimmune rheumatic diseases such as RA and SS have a significantly higher risk of PD [137]. By contrast, patients of SLE have a lower risk of PD [138]. These results suggest that autoimmune disease, possibly through autoantibodies, can either induce or prevent neurodegenerative conditions. However, further research is needed to identify the specific autoantibodies that are associated with neurodegeneration and to identify their functions.

In recent decades, extensive efforts by researchers, together with advances in molecular and biochemical techniques, have made it possible to elucidate the lymphatic drainage system in the brain and examine infiltrating adaptive immune components in the CNS. In particular, the appreciation of the relationship between Igs and neurodegenerative disease such as AD and PD and the potential role of Igs in disease pathology have grown markedly. Our summary indicates that the functional roles of Igs depend on effector cells and target antigens. Igs or immune complexes can be neurotoxic by inducing proinflammatory responses through microglia activation in the CNS via activation of Fc $\gamma$ R. By contrast, they may exert protective effects through triggering immune-complex-mediated phagocytosis by microglia via Fc $\gamma$ R activation. Interestingly, IgG from PD and AD patients are more neurotoxic than those from controls, suggesting that regulation of Ig-mediated Fc $\gamma$ R signaling is a complicated process controlled by additional factors. Recently, studies have shown that Ig glycosylation affects the affinity of antibodies for Fc $\gamma$ R. In addition, microarray studies identified promising autoantibodies in the blood of AD and PD patients. Data suggest that changes in both Fab and Fc regions of Igs have important roles with respect to interactions with CNS cells in neuronal

disease. Although many attempts have been made to develop vaccines based on antibodies specific for A $\beta$  and  $\alpha$ -syn, most have failed. A more comprehensive understanding of the function and roles of Fc and Fab fragments may enable us to develop advanced vaccines for the treatment of CNS disease.

Author Contributions: K.Y.S. and S.G.P. wrote the manuscript with support from K.C.I. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research was supported by grants from the Research of Korea Centers for Disease Control and Prevention (2020-ER6902-00) and the National Research Foundation of Korea (NRF-2016M3C7A1905475 and 2017R1E1A1A01074299).

Conflicts of Interest: The authors declare no conflict of interest.

#### Abbreviations

Immunoglobulin			
Sjögren's syndrome			
Systemic lupus erythematosus			
Rheumatoid arthritis			
Alzheimer's disease			
Parkinson's disease			
Central nervous system			
Cerebrospinal fluid			
Natural killer			
Blood-brain barrier			
Nicotinic acetylcholine receptor $\alpha 7$			
Spleen tyrosine kinase			
Fc gamma receptor			
Stress granules			
Beta-amyloid			
Substantia nigra			
α-Synuclein			
Tyrosine hydroxylase			
Myelin-associated glycoprotein			

## References

- 1. Casali, P.; Schettino, E.W. Structure and function of natural antibodies. *Curr. Top. Microbiol. Immunol.* **1996**, 210, 167–179. [PubMed]
- 2. Elkon, K.; Casali, P. Nature and functions of autoantibodies. *Nat. Clin. Pract. Rheumatol.* **2008**, *4*, 491–498. [CrossRef] [PubMed]
- 3. Rodriguez-Zhurbenko, N.; Quach, T.D.; Hopkins, T.J.; Rothstein, T.L.; Hernandez, A.M. Human B-1 Cells and B-1 Cell Antibodies Change With Advancing Age. *Front. Immunol.* **2019**, *10*, 483. [CrossRef] [PubMed]
- 4. Sim, K.Y.; Park, S.H.; Choi, K.Y.; Park, J.E.; Lee, J.S.; Kim, B.C.; Gwak, J.; Song, W.K.; Lee, K.H.; Park, S.G. High-throughput epitope profiling of antibodies in the plasma of Alzheimer's disease patients using random peptide microarrays. *Sci. Rep.* **2019**, *9*, 4587. [CrossRef]
- 5. Neiman, M.; Hellstrom, C.; Just, D.; Mattsson, C.; Fagerberg, L.; Schuppe-Koistinen, I.; Gummesson, A.; Bergstrom, G.; Kallioniemi, O.; Achour, A.; et al. Individual and stable autoantibody repertoires in healthy individuals. *Autoimmunity* **2019**, *52*, 1–11. [CrossRef] [PubMed]
- Madi, A.; Hecht, I.; Bransburg-Zabary, S.; Merbl, Y.; Pick, A.; Zucker-Toledano, M.; Quintana, F.J.; Tauber, A.I.; Cohen, I.R.; Ben-Jacob, E. Organization of the autoantibody repertoire in healthy newborns and adults revealed by system level informatics of antigen microarray data. *Proc. Natl. Acad. Sci. USA* 2009, 106, 14484–14489. [CrossRef] [PubMed]
- Ma, W.T.; Chang, C.; Gershwin, M.E.; Lian, Z.X. Development of autoantibodies precedes clinical manifestations of autoimmune diseases: A comprehensive review. *J. Autoimmun.* 2017, *83*, 95–112. [CrossRef]

- Ludwig, R.J.; Vanhoorelbeke, K.; Leypoldt, F.; Kaya, Z.; Bieber, K.; McLachlan, S.M.; Komorowski, L.; Luo, J.; Cabral-Marques, O.; Hammers, C.M.; et al. Mechanisms of Autoantibody-Induced Pathology. *Front. Immunol.* 2017, *8*, 603. [CrossRef]
- 9. Eriksson, C.; Kokkonen, H.; Johansson, M.; Hallmans, G.; Wadell, G.; Rantapaa-Dahlqvist, S. Autoantibodies predate the onset of systemic lupus erythematosus in northern Sweden. *Arthritis Res. Ther.* **2011**, *13*, R30. [CrossRef]
- Shmerling, R.H. Autoantibodies in systemic lupus erythematosus—There before you know it. N. Engl. J. Med. 2003, 349, 1499–1500. [CrossRef]
- Lichtbroun, A.S. Positive Findings on an Early Autoantibody Panel in Sjogren's Syndrome Often Predate Positive Findings on Classic Tests: Comment on the Article by Theander et al. *Arthritis Rheumatol.* 2016, 68, 2054–2055. [CrossRef] [PubMed]
- 12. Gronwall, C.; Silverman, G.J. Natural IgM: Beneficial autoantibodies for the control of inflammatory and autoimmune disease. *J. Clin. Immunol.* **2014**, *34*, S12–S21. [CrossRef]
- Grossmayer, G.E.; Munoz, L.E.; Weber, C.K.; Franz, S.; Voll, R.E.; Kern, P.M.; Kalden, J.R.; Schett, G.; Herrmann, M.; Gaipl, U.S. IgG autoantibodies bound to surfaces of necrotic cells and complement C4 comprise the phagocytosis promoting activity for necrotic cells of systemic lupus erythaematosus sera. *Ann. Rheum. Dis.* 2008, *67*, 1626–1632. [CrossRef] [PubMed]
- Sarmiento, L.F.; Munoz, L.E.; Chirinos, P.; Bianco, N.E.; Zabaleta-Lanz, M.E. Opsonization by anti-dsDNA antibodies of apoptotic cells in systemic lupus erythematosus. *Autoimmunity* 2007, 40, 337–339. [CrossRef] [PubMed]
- 15. Manfredi, A.A.; Rovere, P.; Galati, G.; Heltai, S.; Bozzolo, E.; Soldini, L.; Davoust, J.; Balestrieri, G.; Tincani, A.; Sabbadini, M.G. Apoptotic cell clearance in systemic lupus erythematosus. I. Opsonization by antiphospholipid antibodies. *Arthritis Rheum.* **1998**, *41*, 205–214. [CrossRef]
- 16. Maderna, P.; Godson, C. Phagocytosis of apoptotic cells and the resolution of inflammation. *Biochim. Biophys. Acta* **2003**, *1639*, 141–151. [CrossRef] [PubMed]
- Louveau, A.; Harris, T.H.; Kipnis, J. Revisiting the Mechanisms of CNS Immune Privilege. *Trends Immunol.* 2015, 36, 569–577. [CrossRef]
- Korin, B.; Ben-Shaanan, T.L.; Schiller, M.; Dubovik, T.; Azulay-Debby, H.; Boshnak, N.T.; Koren, T.; Rolls, A. High-dimensional, single-cell characterization of the brain's immune compartment. *Nat. Neurosci.* 2017, 20, 1300–1309. [CrossRef]
- Louveau, A.; Smirnov, I.; Keyes, T.J.; Eccles, J.D.; Rouhani, S.J.; Peske, J.D.; Derecki, N.C.; Castle, D.; Mandell, J.W.; Lee, K.S.; et al. Corrigendum: Structural and functional features of central nervous system lymphatic vessels. *Nature* 2016, 533, 278. [CrossRef]
- 20. Ransohoff, R.M. How neuroinflammation contributes to neurodegeneration. *Science* **2016**, *353*, 777–783. [CrossRef]
- 21. Benkler, M.; Agmon-Levin, N.; Hassin-Baer, S.; Cohen, O.S.; Ortega-Hernandez, O.D.; Levy, A.; Moscavitch, S.D.; Szyper-Kravitz, M.; Damianovich, M.; Blank, M.; et al. Immunology, autoimmunity, and autoantibodies in Parkinson's disease. *Clin. Rev. Allergy Immunol.* **2012**, *42*, 164–171. [CrossRef] [PubMed]
- 22. D'Andrea, M.R. Add Alzheimer's disease to the list of autoimmune diseases. *Med. Hypotheses* **2005**, *64*, 458–463. [CrossRef] [PubMed]
- 23. Jiang, T.; Li, G.; Xu, J.; Gao, S.; Chen, X. The Challenge of the Pathogenesis of Parkinson's Disease: Is Autoimmunity the Culprit? *Front. Immunol.* **2018**, *9*, 2047. [CrossRef] [PubMed]
- 24. DeMarshall, C.A.; Han, M.; Nagele, E.P.; Sarkar, A.; Acharya, N.K.; Godsey, G.; Goldwaser, E.L.; Kosciuk, M.; Thayasivam, U.; Belinka, B.; et al. Potential utility of autoantibodies as blood-based biomarkers for early detection and diagnosis of Parkinson's disease. *Immunol. Lett.* **2015**, *168*, 80–88. [CrossRef] [PubMed]
- DeMarshall, C.A.; Nagele, E.P.; Sarkar, A.; Acharya, N.K.; Godsey, G.; Goldwaser, E.L.; Kosciuk, M.; Thayasivam, U.; Han, M.; Belinka, B.; et al. Detection of Alzheimer's disease at mild cognitive impairment and disease progression using autoantibodies as blood-based biomarkers. *Alzheimers Dement. (Amsterdam)* 2016, 3, 51–62. [CrossRef]
- 26. Alzheimer's Association. 2020 Alzheimer's disease facts and figures. *Alzheimers Dement.* 2020, *16*, 391. [CrossRef]
- 27. Kametani, F.; Hasegawa, M. Reconsideration of Amyloid Hypothesis and Tau Hypothesis in Alzheimer's Disease. *Front. Neurosci.* **2018**, *12*, 25. [CrossRef]

- 28. Gate, D.; Saligrama, N.; Leventhal, O.; Yang, A.C.; Unger, M.S.; Middeldorp, J.; Chen, K.; Lehallier, B.; Channappa, D.; De Los Santos, M.B.; et al. Clonally expanded CD8 T cells patrol the cerebrospinal fluid in Alzheimer's disease. *Nature* **2020**, *577*, 399–404. [CrossRef]
- 29. Stowe, A.M.; Ireland, S.J.; Ortega, S.B.; Chen, D.; Huebinger, R.M.; Tarumi, T.; Harris, T.S.; Cullum, C.M.; Rosenberg, R.; Monson, N.L.; et al. Adaptive lymphocyte profiles correlate to brain Abeta burden in patients with mild cognitive impairment. *J. Neuroinflamm.* **2017**, *14*, 149. [CrossRef]
- Bulati, M.; Buffa, S.; Martorana, A.; Gervasi, F.; Camarda, C.; Azzarello, D.M.; Monastero, R.; Caruso, C.; Colonna-Romano, G. Double negative (IgG+IgD-CD27-) B cells are increased in a cohort of moderate-severe Alzheimer's disease patients and show a pro-inflammatory trafficking receptor phenotype. *J. Alzheimers Dis.* 2015, 44, 1241–1251. [CrossRef]
- 31. Sollvander, S.; Ekholm-Pettersson, F.; Brundin, R.M.; Westman, G.; Kilander, L.; Paulie, S.; Lannfelt, L.; Sehlin, D. Increased Number of Plasma B Cells Producing Autoantibodies Against Abeta42 Protofibrils in Alzheimer's Disease. *J. Alzheimers Dis.* **2015**, *48*, 63–72. [CrossRef] [PubMed]
- 32. Sabatino, J.J., Jr.; Probstel, A.K.; Zamvil, S.S. B cells in autoimmune and neurodegenerative central nervous system diseases. *Nat. Rev. Neurosci.* 2019, 20, 728–745. [CrossRef] [PubMed]
- D'Andrea, M.R. Evidence linking neuronal cell death to autoimmunity in Alzheimer's disease. *Brain Res.* 2003, 982, 19–30. [CrossRef]
- 34. Zotova, E.; Bharambe, V.; Cheaveau, M.; Morgan, W.; Holmes, C.; Harris, S.; Neal, J.W.; Love, S.; Nicoll, J.A.; Boche, D. Inflammatory components in human Alzheimer's disease and after active amyloid-beta42 immunization. *Brain* **2013**, *136*, 2677–2696. [CrossRef]
- 35. Bouras, C.; Riederer, B.M.; Kovari, E.; Hof, P.R.; Giannakopoulos, P. Humoral immunity in brain aging and Alzheimer's disease. *Brain Res. Brain Res. Rev.* **2005**, *48*, 477–487. [CrossRef]
- 36. Maftei, M.; Thurm, F.; Schnack, C.; Tumani, H.; Otto, M.; Elbert, T.; Kolassa, I.T.; Przybylski, M.; Manea, M.; von Arnim, C.A. Increased levels of antigen-bound beta-amyloid autoantibodies in serum and cerebrospinal fluid of Alzheimer's disease patients. *PLoS ONE* **2013**, *8*, e68996. [CrossRef]
- 37. Bartos, A.; Fialova, L.; Svarcova, J.; Ripova, D. Patients with Alzheimer disease have elevated intrathecal synthesis of antibodies against tau protein and heavy neurofilament. *J. Neuroimmunol.* **2012**, 252, 100–105. [CrossRef]
- Koval, L.; Lykhmus, O.; Kalashnyk, O.; Bachinskaya, N.; Kravtsova, G.; Soldatkina, M.; Zouridakis, M.; Stergiou, C.; Tzartos, S.; Tsetlin, V.; et al. The presence and origin of autoantibodies against α4 and α7 nicotinic acetylcholine receptors in the human blood: Possible relevance to Alzheimer's pathology. *J. Alzheimers Dis.* 2011, 25, 747–761. [CrossRef]
- 39. Báez-Pagán, C.A.; Delgado-Vélez, M.; Lasalde-Dominicci, J.A. Activation of the Macrophage α7 Nicotinic Acetylcholine Receptor and Control of Inflammation. *J. Neuroimmune Pharmacol.* **2015**, *10*, 468–476. [CrossRef]
- Parada, E.; Egea, J.; Romero, A.; del Barrio, L.; García, A.G.; López, M.G. Poststress treatment with PNU282987 can rescue SH-SY5Y cells undergoing apoptosis via α7 nicotinic receptors linked to a Jak2/Akt/HO-1 signaling pathway. *Free Radic. Biol. Med.* 2010, *49*, 1815–1821. [CrossRef]
- 41. Parri, H.R.; Dineley, K.T. Nicotinic Acetylcholine Receptor Interaction with Ab-Amyloid: Molecular, Cellular, and Physiological Consequences. *Curr. Alzheimer Res.* **2010**, *7*, 27–39. [CrossRef] [PubMed]
- Lykhmus, O.; Mishra, N.; Koval, L.; Kalashnyk, O.; Gergalova, G.; Uspenska, K.; Komisarenko, S.; Soreq, H.; Skok, M. Molecular Mechanisms Regulating LPS-Induced Inflammation in the Brain. *Front. Mol. Neurosci.* 2016, 9, 19. [CrossRef] [PubMed]
- Lim, B.; Tsolaki, M.; Batruch, I.; Anastasiou, A.; Frontistis, A.; Prassas, I.; Diamandis, E.P. Putative autoantibodies in the cerebrospinal fluid of Alzheimer's disease patients. *F1000Research* 2019, *8*, 1900. [CrossRef] [PubMed]
- 44. Wu, J.; Li, L. Autoantibodies in Alzheimer's disease: Potential biomarkers, pathogenic roles, and therapeutic implications. *J. Biomed. Res.* **2016**, *30*, 361–372.
- Papuc, E.; Krupski, W.; Kurys-Denis, E.; Rejdak, K. Antibodies against small heat-shock proteins in Alzheimer's disease as a part of natural human immune repertoire or activation of humoral response? J. Neural. Transm. 2016, 123, 455–461. [CrossRef] [PubMed]
- 46. Louveau, A.; Da Mesquita, S.; Kipnis, J. Lymphatics in Neurological Disorders: A Neuro-Lympho-Vascular Component of Multiple Sclerosis and Alzheimer's Disease? *Neuron* **2016**, *91*, 957–973. [CrossRef]

- 47. Ahn, J.H.; Cho, H.; Kim, J.H.; Kim, S.H.; Ham, J.S.; Park, I.; Suh, S.H.; Hong, S.P.; Song, J.H.; Hong, Y.K.; et al. Meningeal lymphatic vessels at the skull base drain cerebrospinal fluid. *Nature* **2019**, 572, 62–66. [CrossRef]
- 48. Sampson, J.H.; Gunn, M.D.; Fecci, P.E.; Ashley, D.M. Brain immunology and immunotherapy in brain tumours. *Nat. Rev. Cancer* 2020, *20*, 12–25. [CrossRef]
- 49. Lang, K.; Pruss, H. Frequencies of neuronal autoantibodies in healthy controls: Estimation of disease specificity. *Neurol. Neuroinflamm.* **2017**, *4*, e386. [CrossRef]
- 50. Kheirkhah, R.; D, C.; Sieber, F.; Oh, E.; Nagele, R.G. The origin and nature of the complex autoantibody profile in cerebrospinal fluid. *Brain Behav. Immun. Health* **2020**, *2*, 100032. [CrossRef]
- Britschgi, M.; Olin, C.E.; Johns, H.T.; Takeda-Uchimura, Y.; LeMieux, M.C.; Rufibach, K.; Rajadas, J.; Zhang, H.; Tomooka, B.; Robinson, W.H.; et al. Neuroprotective natural antibodies to assemblies of amyloidogenic peptides decrease with normal aging and advancing Alzheimer's disease. *Proc. Natl. Acad. Sci. USA* 2009, 106, 12145–12150. [CrossRef] [PubMed]
- 52. Glass, L.J.; Sinclair, D.; Boerrigter, D.; Naude, K.; Fung, S.J.; Brown, D.; Catts, V.S.; Tooney, P.; O'Donnell, M.; Lenroot, R.; et al. Brain antibodies in the cortex and blood of people with schizophrenia and controls. *Transl. Psychiatry* **2017**, *7*, e1192. [CrossRef] [PubMed]
- 53. Montagne, A.; Zhao, Z.; Zlokovic, B.V. Alzheimer's disease: A matter of blood-brain barrier dysfunction? *J. Exp. Med.* **2017**, *214*, 3151–3169. [CrossRef] [PubMed]
- 54. Nelson, A.R.; Sweeney, M.D.; Sagare, A.P.; Zlokovic, B.V. Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta* **2016**, *1862*, 887–900. [CrossRef]
- 55. Fullerton, S.M.; Shirman, G.A.; Strittmatter, W.J.; Matthew, W.D. Impairment of the blood-nerve and blood-brain barriers in apolipoprotein e knockout mice. *Exp. Neurol.* **2001**, *169*, 13–22. [CrossRef]
- 56. Ryu, J.K.; McLarnon, J.G. A leaky blood-brain barrier, fibrinogen infiltration and microglial reactivity in inflamed Alzheimer's disease brain. *J. Cell. Mol. Med.* **2009**, *13*, 2911–2925. [CrossRef]
- 57. Morch, M.T.; Sorensen, S.F.; Khorooshi, R.; Asgari, N.; Owens, T. Selective localization of IgG from cerebrospinal fluid to brain parenchyma. *J. Neuroinflamm.* **2018**, *15*, 110. [CrossRef]
- Villasenor, R.; Ozmen, L.; Messaddeq, N.; Gruninger, F.; Loetscher, H.; Keller, A.; Betsholtz, C.; Freskgard, P.O.; Collin, L. Trafficking of Endogenous Immunoglobulins by Endothelial Cells at the Blood-Brain Barrier. *Sci. Rep.* 2016, *6*, 25658. [CrossRef]
- 59. McBrayer, M.; Nixon, R.A. Lysosome and calcium dysregulation in Alzheimer's disease: Partners in crime. *Biochem. Soc. Trans.* 2013, *41*, 1495–1502. [CrossRef]
- Marsh, S.E.; Abud, E.M.; Lakatos, A.; Karimzadeh, A.; Yeung, S.T.; Davtyan, H.; Fote, G.M.; Lau, L.; Weinger, J.G.; Lane, T.E.; et al. The adaptive immune system restrains Alzheimer's disease pathogenesis by modulating microglial function. *Proc. Natl. Acad. Sci. USA* 2016, *113*, E1316–E1325. [CrossRef]
- 61. Zhang, L.; Xu, J.; Gao, J.; Chen, P.; Yin, M.; Zhao, W. Decreased immunoglobulin G in brain regions of elder female APOE4-TR mice accompany with Abeta accumulation. *Immun. Ageing* **2019**, *16*, 2. [CrossRef] [PubMed]
- Ghosh, S.; Geahlen, R.L. Stress Granules Modulate SYK to Cause Microglial Cell Dysfunction in Alzheimer's Disease. *EBioMedicine* 2015, 2, 1785–1798. [CrossRef] [PubMed]
- Valls-Comamala, V.; Guivernau, B.; Bonet, J.; Puig, M.; Peralvarez-Marin, A.; Palomer, E.; Fernandez-Busquets, X.; Altafaj, X.; Tajes, M.; Puig-Pijoan, A.; et al. The antigen-binding fragment of human gamma immunoglobulin prevents amyloid beta-peptide folding into beta-sheet to form oligomers. *Oncotarget* 2017, *8*, 41154–41165. [CrossRef] [PubMed]
- Lee, S.H.; Le Pichon, C.E.; Adolfsson, O.; Gafner, V.; Pihlgren, M.; Lin, H.; Solanoy, H.; Brendza, R.; Ngu, H.; Foreman, O.; et al. Antibody-Mediated Targeting of Tau In Vivo Does Not Require Effector Function and Microglial Engagement. *Cell Rep.* 2016, *16*, 1690–1700. [CrossRef]
- 65. Li, X.; Koudstaal, W.; Fletcher, L.; Costa, M.; van Winsen, M.; Siregar, B.; Inganas, H.; Kim, J.; Keogh, E.; Macedo, J.; et al. Naturally occurring antibodies isolated from PD patients inhibit synuclein seeding in vitro and recognize Lewy pathology. *Acta Neuropathol.* **2019**, *137*, 825–836. [CrossRef]
- Bae, E.J.; Lee, H.J.; Rockenstein, E.; Ho, D.H.; Park, E.B.; Yang, N.Y.; Desplats, P.; Masliah, E.; Lee, S.J. Antibody-aided clearance of extracellular alpha-synuclein prevents cell-to-cell aggregate transmission. *J. Neurosci.* 2012, 32, 13454–13469. [CrossRef]
- 67. Fuller, J.P.; Stavenhagen, J.B.; Teeling, J.L. New roles for Fc receptors in neurodegeneration-the impact on Immunotherapy for Alzheimer's Disease. *Front. Neurosci.* **2014**, *8*, 235. [CrossRef]

- 68. Andoh, T.; Kuraishi, Y. Direct action of immunoglobulin G on primary sensory neurons through Fc gamma receptor I. *FASEB J.* 2004, *18*, 182–184. [CrossRef]
- 69. Teeling, J.L.; Carare, R.O.; Glennie, M.J.; Perry, V.H. Intracerebral immune complex formation induces inflammation in the brain that depends on Fc receptor interaction. *Acta Neuropathol.* **2012**, *124*, 479–490. [CrossRef]
- 70. Dinkins, M.B.; Dasgupta, S.; Wang, G.; Zhu, G.; He, Q.; Kong, J.N.; Bieberich, E. The 5XFAD Mouse Model of Alzheimer's Disease Exhibits an Age-Dependent Increase in Anti-Ceramide IgG and Exogenous Administration of Ceramide Further Increases Anti-Ceramide Titers and Amyloid Plaque Burden. J. Alzheimers Dis. 2015, 46, 55–61. [CrossRef]
- 71. Wang, W.Y.; Tan, M.S.; Yu, J.T.; Tan, L. Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease. *Ann. Transl. Med.* **2015**, *3*, 136. [PubMed]
- 72. Wang, X.J.; Yan, Z.Q.; Lu, G.Q.; Stuart, S.; Chen, S.D. Parkinson disease IgG and C5a-induced synergistic dopaminergic neurotoxicity: Role of microglia. *Neurochem. Int.* **2007**, *50*, 39–50. [CrossRef] [PubMed]
- 73. Cao, S.; Theodore, S.; Standaert, D.G. Fcgamma receptors are required for NF-kappaB signaling, microglial activation and dopaminergic neurodegeneration in an AAV-synuclein mouse model of Parkinson's disease. *Mol. Neurodegener.* **2010**, *5*, 42. [CrossRef]
- 74. Choi, Y.R.; Cha, S.-H.; Kang, S.-J.; Kim, J.-B.; Jou, I.; Park, S.M. Prion-like Propagation of α-Synuclein Is Regulated by the FcγRIIB-SHP-1/2 Signaling Pathway in Neurons. *Cell Rep.* **2018**, 22, 136–148. [CrossRef] [PubMed]
- 75. Bruhns, P.; Jonsson, F. Mouse and human FcR effector functions. *Immunol. Rev.* **2015**, *268*, 25–51. [CrossRef] [PubMed]
- Katsinelos, T.; Tuck, B.J.; Mukadam, A.S.; McEwan, W.A. The Role of Antibodies and Their Receptors in Protection Against Ordered Protein Assembly in Neurodegeneration. *Front. Immunol.* 2019, 10, 1139. [CrossRef]
- Stamou, M.; Lein, P.J. Commentary: Fc Gamma Receptors are Expressed in the Developing Rat Brain and Activate Downstream Signaling Molecules upon Cross-Linking with Immune Complex. *J. Neurol. Neuromed.* 2019, 4, 26–29. [CrossRef]
- 78. Murinello, S.; Mullins, R.F.; Lotery, A.J.; Perry, V.H.; Teeling, J.L. Fcgamma receptor upregulation is associated with immune complex inflammation in the mouse retina and early age-related macular degeneration. *Investig. Ophthalmol. Vis. Sci.* **2014**, *55*, 247–258. [CrossRef]
- Mohamed, H.A.; Mosier, D.R.; Zou, L.L.; Siklos, L.; Alexianu, M.E.; Engelhardt, J.I.; Beers, D.R.; Le, W.D.; Appel, S.H. Immunoglobulin Fc gamma receptor promotes immunoglobulin uptake, immunoglobulin-mediated calcium increase, and neurotransmitter release in motor neurons. *J. Neurosci. Res.* 2002, 69, 110–116. [CrossRef]
- 80. Kam, T.I.; Song, S.; Gwon, Y.; Park, H.; Yan, J.J.; Im, I.; Choi, J.W.; Choi, T.Y.; Kim, J.; Song, D.K.; et al. FcgammaRIIb mediates amyloid-beta neurotoxicity and memory impairment in Alzheimer's disease. *J. Clin. Investig.* **2013**, *123*, 2791–2802. [CrossRef]
- Fernandez-Vizarra, P.; Lopez-Franco, O.; Mallavia, B.; Higuera-Matas, A.; Lopez-Parra, V.; Ortiz-Munoz, G.; Ambrosio, E.; Egido, J.; Almeida, O.F.; Gomez-Guerrero, C. Immunoglobulin G Fc receptor deficiency prevents Alzheimer-like pathology and cognitive impairment in mice. *Brain* 2012, 135, 2826–2837. [CrossRef] [PubMed]
- 82. Quan, Y.; Moller, T.; Weinstein, J.R. Regulation of Fcgamma receptors and immunoglobulin G-mediated phagocytosis in mouse microglia. *Neurosci. Lett.* **2009**, *464*, 29–33. [CrossRef]
- Yang, C.; Hou, X.; Feng, Q.; Li, Y.; Wang, X.; Qin, L.; Yang, P. Lupus serum IgG induces microglia activation through Fc fragment dependent way and modulated by B-cell activating factor. *J. Transl. Med.* 2019, 17, 426. [CrossRef] [PubMed]
- Engelhardt, J.I.; Le, W.D.; Siklos, L.; Obal, I.; Boda, K.; Appel, S.H. Stereotaxic injection of IgG from patients with Alzheimer disease initiates injury of cholinergic neurons of the basal forebrain. *Arch. Neurol.* 2000, 57, 681–686. [CrossRef] [PubMed]
- Russell, A.; Adua, E.; Ugrina, I.; Laws, S.; Wang, W. Unravelling Immunoglobulin G Fc N-Glycosylation: A Dynamic Marker Potentiating Predictive, Preventive and Personalised Medicine. *Int. J. Mol. Sci.* 2018, 19, 390. [CrossRef]

- Russell, A.C.; Šimurina, M.; Garcia, M.T.; Novokmet, M.; Wang, Y.; Rudan, I.; Campbell, H.; Lauc, G.; Thomas, M.G.; Wang, W. The N-glycosylation of immunoglobulin G as a novel biomarker of Parkinson's disease. *Glycobiology* 2017, 27, 501–510. [CrossRef]
- 87. Bacskai, B.J.; Kajdasz, S.T.; McLellan, M.E.; Games, D.; Seubert, P.; Schenk, D.; Hyman, B.T. Non-Fc-mediated mechanisms are involved in clearance of amyloid-beta in vivo by immunotherapy. *J. Neurosci.* **2002**, *22*, 7873–7878. [CrossRef]
- 88. Tamura, Y.; Hamajima, K.; Matsui, K.; Yanoma, S.; Narita, M.; Tajima, N.; Xin, K.Q.; Klinman, D.; Okuda, K. The F(ab)'2 fragment of an Abeta-specific monoclonal antibody reduces Abeta deposits in the brain. *Neurobiol. Dis.* 2005, 20, 541–549. [CrossRef]
- 89. Li, X.W.; Li, X.X.; Liu, Q.S.; Cheng, Y. Blood and Cerebrospinal Fluid Autoantibody to Abeta Levels in Patients with Alzheimer's Disease: A Meta-Analysis Study. *J. Mol. Neurosci.* **2020**, *70*, 1208–1215. [CrossRef]
- Hromadkova, L.; Ovsepian, S.V. Tau-Reactive Endogenous Antibodies: Origin, Functionality, and Implications for the Pathophysiology of Alzheimer's Disease. J. Immunol. Res. 2019, 2019, 7406810. [CrossRef]
- Yu, Z.Y.; Li, W.W.; Yang, H.M.; Manucat-Tan, N.B.; Wang, J.; Wang, Y.R.; Sun, B.L.; Hu, Z.C.; Zhang, L.L.; Tan, L.; et al. Naturally Occurring Antibodies to Tau Exists in Human Blood and Are Not Changed in Alzheimer's Disease. *Neurotox. Res.* 2020, *37*, 1029–1035. [CrossRef] [PubMed]
- DeMarshall, C.; Oh, E.; Kheirkhah, R.; Sieber, F.; Zetterberg, H.; Blennow, K.; Nagele, R.G. Detection of early-stage Alzheimer's pathology using blood-based autoantibody biomarkers in elderly hip fracture repair patients. *PLoS ONE* 2019, 14, e0225178. [CrossRef] [PubMed]
- 93. Counts, S.E.; Ikonomovic, M.D.; Mercado, N.; Vega, I.E.; Mufson, E.J. Biomarkers for the Early Detection and Progression of Alzheimer's Disease. *Neurotherapeutics* **2017**, *14*, 35–53. [CrossRef] [PubMed]
- 94. Davydova, T.V.; Voskresenskaya, N.I.; Fomina, V.G.; Vetrile, L.A.; Doronina, O.A. Induction of autoantibodies to glutamate in patients with Alzheimer's disease. *Bull. Exp. Biol. Med.* **2007**, *143*, 182–183. [CrossRef]
- 95. Gruden, M.A.; Davidova, T.B.; Malisauskas, M.; Sewell, R.D.; Voskresenskaya, N.I.; Wilhelm, K.; Elistratova, E.I.; Sherstnev, V.V.; Morozova-Roche, L.A. Differential neuroimmune markers to the onset of Alzheimer's disease neurodegeneration and dementia: Autoantibodies to Abeta((25-35)) oligomers, S100b and neurotransmitters. *J. Neuroimmunol.* **2007**, *186*, 181–192. [CrossRef]
- 96. Hempel, P.; Heinig, B.; Jerosch, C.; Decius, I.; Karczewski, P.; Kassner, U.; Kunze, R.; Steinhagen-Thiessen, E.; Bimmler, M. Immunoadsorption of Agonistic Autoantibodies Against alpha1-Adrenergic Receptors in Patients With Mild to Moderate Dementia. *Ther. Apher. Dial.* 2016, 20, 523–529. [CrossRef]
- 97. Honig, L.S.; Vellas, B.; Woodward, M.; Boada, M.; Bullock, R.; Borrie, M.; Hager, K.; Andreasen, N.; Scarpini, E.; Liu-Seifert, H.; et al. Trial of Solanezumab for Mild Dementia Due to Alzheimer's Disease. *N. Engl. J. Med.* 2018, *378*, 321–330. [CrossRef]
- 98. Vandenberghe, R.; Rinne, J.O.; Boada, M.; Katayama, S.; Scheltens, P.; Vellas, B.; Tuchman, M.; Gass, A.; Fiebach, J.B.; Hill, D.; et al. Bapineuzumab for mild to moderate Alzheimer's disease in two global, randomized, phase 3 trials. *Alzheimers Res. Ther.* **2016**, *8*, 18. [CrossRef]
- 99. Ostrowitzki, S.; Lasser, R.A.; Dorflinger, E.; Scheltens, P.; Barkhof, F.; Nikolcheva, T.; Ashford, E.; Retout, S.; Hofmann, C.; Delmar, P.; et al. A phase III randomized trial of gantenerumab in prodromal Alzheimer's disease. *Alzheimers Res. Ther.* **2017**, *9*, 95. [CrossRef]
- 100. Klein, G.; Delmar, P.; Voyle, N.; Rehal, S.; Hofmann, C.; Abi-Saab, D.; Andjelkovic, M.; Ristic, S.; Wang, G.; Bateman, R.; et al. Gantenerumab reduces amyloid-beta plaques in patients with prodromal to moderate Alzheimer's disease: A PET substudy interim analysis. *Alzheimers Res. Ther.* 2019, *11*, 101. [CrossRef]
- 101. Vellas, B.; Black, R.; Thal, L.J.; Fox, N.C.; Daniels, M.; McLennan, G.; Tompkins, C.; Leibman, C.; Pomfret, M.; Grundman, M.; et al. Long-term follow-up of patients immunized with AN1792: Reduced functional decline in antibody responders. *Curr. Alzheimer Res.* 2009, *6*, 144–151. [CrossRef] [PubMed]
- 102. Nicoll, J.A.R.; Buckland, G.R.; Harrison, C.H.; Page, A.; Harris, S.; Love, S.; Neal, J.W.; Holmes, C.; Boche, D. Persistent neuropathological effects 14 years following amyloid-beta immunization in Alzheimer's disease. *Brain* 2019, 142, 2113–2126. [CrossRef] [PubMed]
- 103. Chantran, Y.; Capron, J.; Alamowitch, S.; Aucouturier, P. Anti-Abeta Antibodies and Cerebral Amyloid Angiopathy Complications. *Front. Immunol.* **2019**, *10*, 1534. [CrossRef] [PubMed]
- Huang, L.K.; Chao, S.P.; Hu, C.J. Clinical trials of new drugs for Alzheimer disease. J. Biomed. Sci. 2020, 27, 18. [CrossRef] [PubMed]

- 105. Poewe, W.; Seppi, K.; Tanner, C.M.; Halliday, G.M.; Brundin, P.; Volkmann, J.; Schrag, A.-E.; Lang, A.E. Parkinson disease. *Nat. Rev. Dis. Primers* **2017**, *3*, 17013. [CrossRef]
- 106. Garretti, F.; Agalliu, D.; Lindestam Arlehamn, C.S.; Sette, A.; Sulzer, D. Autoimmunity in Parkinson's Disease: The Role of α-Synuclein-Specific T Cells. *Front. Immunol.* **2019**, *10*, 303. [CrossRef]
- 107. Schröder, J.B.; Pawlowski, M.; Meyer Zu Hörste, G.; Gross, C.C.; Wiendl, H.; Meuth, S.G.; Ruck, T.; Warnecke, T. Immune Cell Activation in the Cerebrospinal Fluid of Patients With Parkinson's Disease. *Front. Neurol.* 2018, 9, 1081. [CrossRef]
- 108. Kustrimovic, N.; Comi, C.; Magistrelli, L.; Rasini, E.; Legnaro, M.; Bombelli, R.; Aleksic, I.; Blandini, F.; Minafra, B.; Riboldazzi, G.; et al. Parkinson's disease patients have a complex phenotypic and functional Th1 bias: Cross-sectional studies of CD4+ Th1/Th2/T17 and Treg in drug-naïve and drug-treated patients. J. Neuroinflamm. 2018, 15, 205. [CrossRef]
- Pisani, V.; Stefani, A.; Pierantozzi, M.; Natoli, S.; Stanzione, P.; Franciotta, D.; Pisani, A. Increased blood-cerebrospinal fluid transfer of albumin in advanced Parkinson's disease. *J. Neuroinflamm.* 2012, 9, 188. [CrossRef]
- 110. Pienaar, I.S.; Lee, C.H.; Elson, J.L.; McGuinness, L.; Gentleman, S.M.; Kalaria, R.N.; Dexter, D.T. Deep-brain stimulation associates with improved microvascular integrity in the subthalamic nucleus in Parkinson's disease. *Neurobiol. Dis.* 2015, 74, 392–405. [CrossRef]
- 111. Orr, C.F.; Rowe, D.B.; Mizuno, Y.; Mori, H.; Halliday, G.M. A possible role for humoral immunity in the pathogenesis of Parkinson's disease. *Brain* **2005**, *128*, 2665–2674. [CrossRef] [PubMed]
- 112. Folke, J.; Rydbirk, R.; Løkkegaard, A.; Salvesen, L.; Hejl, A.-M.; Starhof, C.; Bech, S.; Winge, K.; Christensen, S.; Pedersen, L.Ø.; et al. Distinct Autoimmune Anti-α-Synuclein Antibody Patterns in Multiple System Atrophy and Parkinson's Disease. *Front. Immunol.* **2019**, *10*, 2253. [CrossRef] [PubMed]
- Scott, K.M.; Kouli, A.; Yeoh, S.L.; Clatworthy, M.R.; Williams-Gray, C.H. A Systematic Review and Meta-Analysis of Alpha Synuclein Auto-Antibodies in Parkinson's Disease. *Front. Neurol.* 2018, *9*, 815.
  [CrossRef] [PubMed]
- 114. Brudek, T.; Winge, K.; Folke, J.; Christensen, S.; Fog, K.; Pakkenberg, B.; Pedersen, L.O. Autoimmune antibody decline in Parkinson's disease and Multiple System Atrophy; A step towards immunotherapeutic strategies. *Mol. Neurodegener.* **2017**, *12*, 44. [CrossRef]
- 115. Kortekaas, R.; Leenders, K.L.; van Oostrom, J.C.; Vaalburg, W.; Bart, J.; Willemsen, A.T.; Hendrikse, N.H. Blood-brain barrier dysfunction in parkinsonian midbrain in vivo. *Ann. Neurol.* 2005, 57, 176–179. [CrossRef] [PubMed]
- 116. Ham, J.H.; Yi, H.; Sunwoo, M.K.; Hong, J.Y.; Sohn, Y.H.; Lee, P.H. Cerebral microbleeds in patients with Parkinson's disease. *J. Neurol.* **2014**, *261*, 1628–1635. [CrossRef] [PubMed]
- 117. Gray, M.T.; Woulfe, J.M. Striatal blood-brain barrier permeability in Parkinson's disease. J. Cereb. Blood Flow Metab. 2015, 35, 747–750. [CrossRef]
- Desai Bradaric, B.; Patel, A.; Schneider, J.A.; Carvey, P.M.; Hendey, B. Evidence for angiogenesis in Parkinson's disease, incidental Lewy body disease, and progressive supranuclear palsy. *J. Neural. Transm.* 2012, 119, 59–71. [CrossRef]
- 119. Masliah, E.; Rockenstein, E.; Mante, M.; Crews, L.; Spencer, B.; Adame, A.; Patrick, C.; Trejo, M.; Ubhi, K.; Rohn, T.T.; et al. Passive immunization reduces behavioral and neuropathological deficits in an alpha-synuclein transgenic model of Lewy body disease. *PLoS ONE* **2011**, *6*, e19338. [CrossRef]
- 120. Spencer, B.; Valera, E.; Rockenstein, E.; Overk, C.; Mante, M.; Adame, A.; Zago, W.; Seubert, P.; Barbour, R.; Schenk, D.; et al. Anti-alpha-synuclein immunotherapy reduces alpha-synuclein propagation in the axon and degeneration in a combined viral vector and transgenic model of synucleinopathy. *Acta Neuropathol. Commun.* 2017, 5, 7. [CrossRef]
- 121. He, Y.; Le, W.D.; Appel, S.H. Role of Fcgamma receptors in nigral cell injury induced by Parkinson disease immunoglobulin injection into mouse substantia nigra. *Exp. Neurol.* **2002**, *176*, 322–327. [CrossRef] [PubMed]
- 122. Papuć, E.; Rejdak, K. Anti-MAG autoantibodies are increased in Parkinson's disease but not in atypical parkinsonism. *J. Neural. Transm.* 2017, 124, 209–216. [CrossRef] [PubMed]
- 123. Papuć, E.; Kurzepa, J.; Kurys-Denis, E.; Grabarska, A.; Krupski, W.; Rejdak, K. Humoral response against glial derived antigens in Parkinson's disease. *Neurosci. Lett.* **2014**, *566*, 77–81. [CrossRef] [PubMed]

- 124. Magy, L.; Kabore, R.; Mathis, S.; Lebeau, P.; Ghorab, K.; Caudie, C.; Vallat, J.M. Heterogeneity of Polyneuropathy Associated with Anti-MAG Antibodies. J. Immunol. Res. 2015, 2015, 450391. [CrossRef] [PubMed]
- 125. Garces-Sanchez, M.; Dyck, P.J.; Kyle, R.A.; Zeldenrust, S.; Wu, Y.; Ladha, S.S.; Klein, C.J. Antibodies to myelin-associated glycoprotein (anti-Mag) in IgM amyloidosis may influence expression of neuropathy in rare patients. *Muscle Nerve* **2008**, *37*, 490–495. [CrossRef] [PubMed]
- 126. Grambalova, Z.; Kaiserova, M.; Vastik, M.; Mensikova, K.; Otruba, P.; Zapletalova, J.; Dufek, J.; Kanovsky, P. Peripheral neuropathy in Parkinson's disease. *Neuro. Endocrinol. Lett.* **2015**, *36*, 363–367. [PubMed]
- 127. Feng, N.; Simanski, S.; Islam, K.; Hynan, L.S.; Kodadek, T.; German, D.C. Antibody biomarker for de novo Parkinson disease: Attempted validation. *NPJ Parkinsons Dis.* **2018**, *4*, 28. [CrossRef]
- 128. Akhtar, R.S.; Licata, J.P.; Luk, K.C.; Shaw, L.M.; Trojanowski, J.Q.; Lee, V.M. Measurements of auto-antibodies to α-synuclein in the serum and cerebral spinal fluids of patients with Parkinson's disease. *J. Neurochem.* 2018, 145, 489–503. [CrossRef]
- 129. Huang, Y.R.; Xie, X.X.; Ji, M.; Yu, X.L.; Zhu, J.; Zhang, L.X.; Liu, X.G.; Wei, C.; Li, G.; Liu, R.T. Naturally occurring autoantibodies against α-synuclein rescues memory and motor deficits and attenuates α-synuclein pathology in mouse model of Parkinson's disease. *Neurobiol. Dis.* **2019**, *124*, 202–217. [CrossRef]
- 130. George, S.; Brundin, P. Immunotherapy in Parkinson's Disease: Micromanaging Alpha-Synuclein Aggregation. J. Parkinsons Dis. 2015, 5, 413–424. [CrossRef]
- 131. Schenk, D.B.; Koller, M.; Ness, D.K.; Griffith, S.G.; Grundman, M.; Zago, W.; Soto, J.; Atiee, G.; Ostrowitzki, S.; Kinney, G.G. First-in-human assessment of PRX002, an anti-alpha-synuclein monoclonal antibody, in healthy volunteers. *Mov. Disord.* 2017, 32, 211–218. [CrossRef]
- McFarthing, K.; Simuni, T. Clinical Trial Highlights: Targetting Alpha-Synuclein. J. Parkinsons. Dis. 2019, 9, 5–16. [CrossRef] [PubMed]
- 133. Shin, J.; Kim, H.J.; Jeon, B. Immunotherapy Targeting Neurodegenerative Proteinopathies: Alpha-Synucleinopathies and Tauopathies. *J. Mov. Disord.* **2020**, *13*, 11–19. [CrossRef] [PubMed]
- Chatterjee, D.; Kordower, J.H. Immunotherapy in Parkinson's disease: Current status and future directions. *Neurobiol. Dis.* 2019, 132, 104587. [CrossRef] [PubMed]
- 135. Liliang, P.C.; Liang, C.L.; Lu, K.; Yang, S.N.; Hsieh, M.T.; Tai, Y.C.; Wang, K.W. Population-based study suggests an increased risk of Alzheimer's disease in Sjogren's syndrome. *Clin. Rheumatol.* 2018, 37, 935–941. [CrossRef]
- Wotton, C.J.; Goldacre, M.J. Associations between specific autoimmune diseases and subsequent dementia: Retrospective record-linkage cohort study, UK. J. Epidemiol. Commun. Health 2017, 71, 576–583. [CrossRef]
- Chang, C.C.; Lin, T.M.; Chang, Y.S.; Chen, W.S.; Sheu, J.J.; Chen, Y.H.; Chen, J.H. Autoimmune rheumatic diseases and the risk of Parkinson disease: A nationwide population-based cohort study in Taiwan. *Ann. Med.* 2018, 50, 83–90. [CrossRef]
- Liu, F.C.; Huang, W.Y.; Lin, T.Y.; Shen, C.H.; Chou, Y.C.; Lin, C.L.; Lin, K.T.; Kao, C.H. Inverse Association of Parkinson Disease With Systemic Lupus Erythematosus: A Nationwide Population-based Study. *Medicine* 2015, 94, e2097. [CrossRef]



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).