




Type 2 Diabetes (T2DM) and Parkinson's Disease (PD): a Mechanistic Approach

S. Sri Sabari^{1,2} · Kiruthika Balasubramani² · Mahalaxmi Iyer³ · Harysh Winstler Sureshbabu^{1,2} · Dhivya Venkatesan² · Abilash Valsala Gopalakrishnan⁴ · Arul Narayanaswamy⁵ · Nachimuthu Senthil Kumar⁶ · Balachandar Vellingiri^{1,2} 

Received: 30 January 2023 / Accepted: 19 April 2023

© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2023

Abstract

Growing evidence suggest that there is a connection between Parkinson's disease (PD) and insulin dysregulation in the brain, whilst the connection between PD and type 2 diabetes mellitus (T2DM) is still up for debate. Insulin is widely recognised to play a crucial role in neuronal survival and brain function; any changes in insulin metabolism and signalling in the central nervous system (CNS) can lead to the development of various brain disorders. There is accumulating evidence linking T2DM to PD and other neurodegenerative diseases. In fact, they have a lot in common patho-physiologically, including insulin dysregulation, oxidative stress resulting in mitochondrial dysfunction, microglial activation, and inflammation. As a result, initial research should focus on the role of insulin and its molecular mechanism in order to develop therapeutic outcomes. In this current review, we will look into the link between T2DM and PD, the function of insulin in the brain, and studies related to impact of insulin in causing T2DM and PD. Further, we have also highlighted the role of various insulin signalling pathway in both T2DM and PD. We have also suggested that T2DM-targeting pharmacological strategies as potential therapeutic approach for individuals with cognitive impairment, and we have demonstrated the effectiveness of T2DM-prescribed drugs through current PD treatment trials. In conclusion, this investigation would fill a research gap in T2DM-associated Parkinson's disease (PD) with a potential therapy option.

Keywords Parkinson's disease · Type 2 diabetes mellitus · Insulin · Pathophysiology · Therapeutics

Introduction

Parkinson's disease (PD) is a movement disorder depicted by dopaminergic (DA) neuron depletion in the basal ganglia region [1]. It is characterised by four cardinal symptoms of rigidity, postural imbalance, bradykinesia, and tremor [2]. The cause and severity of PD may be influenced by a variety of demographic and environmental factors [3, 4]. PD is multi-factorial, but ageing is considered to one of the prime causative agent amongst others [5, 6]; however, the pathophysiology of PD is still elusive [7]. People living with PD are more likely to develop several other diseases, including heart disease [8], cancer [9], gastrointestinal disorders [10], type 2 diabetes mellitus (T2DM) [11], vitamin D deficiency [12], peripheral neuropathy [13], and musculoskeletal diseases [14]. Amongst these associated diseases, T2DM has been the most commonly associated disease with PD. In an earlier study, it was stated that lysosomal disruption, mitochondrial dysfunction, aberrant protein build-up, and chronic inflammation are some of the common possible

✉ Balachandar Vellingiri
geneticbala@buc.edu.in

¹ Department of Zoology, School of Basic Sciences, Stem Cell and Regenerative Medicine/Translational Research, Central University of Punjab (CUPB), Bathinda 151401, Punjab, India

² Human Molecular Cytogenetics and Stem Cell Laboratory, Department of Human Genetics and Molecular Biology, Bharathiar University, Coimbatore 641 046, Tamil Nadu, India

³ Department of Biotechnology, Karpagam Academy of Higher Education (Deemed to Be University), Coimbatore 641021, Tamil Nadu, India

⁴ Department of Biomedical Sciences, School of Biosciences and Technology, Vellore Institute of Technology (VIT), Vellore 632 014, India

⁵ Department of Zoology, Bharathiar University, Coimbatore 641 046, Tamil Nadu, India

⁶ Department of Biotechnology, Mizoram University (A Central University), Aizawl 796004, Mizoram, India

signalling pathways that may cause protein misfolding and insulin resistance, in both PD and T2DM [15]. Interestingly, there is also an evidence for cross-seeding between islet amyloid polypeptide (IAPP) and α Syn [16]. Also, metabolically in T2DM, insufficient insulin production causes disruptions in glucose metabolism and chronic inflammation, whereas similar metabolic downregulation is seen in early PD conditions [17]. As a result, insulin is now widely recognised to play an important role in neuronal survival and brain function. Insulin action is vital for neuronal synaptic plasticity and aids learning and memory. Therefore, alterations in insulin metabolism and signalling in the central nervous system (CNS) can contribute to the development of many brain disorders [18]. The pathogenesis of these brain diseases suggested to be significantly influenced by both insulin resistance and reduced insulin action through many mechanisms [19]. Therefore, understanding the role of insulin and its signalling pathway is essential to investigate, in T2DM and PD [20]. Hence, in this review, we have discussed the link between T2DM and PD, by focusing the function of insulin in the brain, and its clinical and pre-clinical evidences on T2DM and neurodegeneration. We have also reported the efficiency of common therapeutic compounds on T2DM and PD, thereby concluding the need of research on the signalling pathways correlating T2DM and PD with its effectiveness on the development of therapeutic strategies for both.

Accompaniment Between T2DM and PD

The specific protein known as amylin misfolds may contribute to T2DM pathogenesis due to the loss of pancreatic beta cells, which in turn results in insulin insufficiency and hyperglycaemia and speed up of disease progression [21, 22]. Additionally, this amylin has been linked to PD pathophysiology since it interacts with alpha-synuclein (α Syn), which has been reported in an *in vitro* study [23]. This α Syn is also produced in pancreatic islets which plays a crucial role in glucose regulation [24]. Whereas in T2DM, IAPP aggregation takes place in the pancreatic cells [25]. As a result, it is important to maintain the balance of the protein levels between production, release, and clearance. Note that there may be an imbalance in the proportion of amylin or α Syn, which could encourage the formation of toxic aggregates in the brain and modulate pancreatic beta cells [26]. Ultimately, there is a possibility that these two proteins will interact indirectly or directly and result in T2DM and PD when their levels are altered [27]. Few reports have shown that oxidative stress, protein aggregation, neuroinflammation, insulin resistance, and mitochondrial dysfunction are involved in the aetiology of T2DM and PD. Almost all of the biomolecules in a cell can be damaged by oxidative stress, which can result in dysfunction and cell death. In T2DM and

PD, oxidative stress damages pancreatic beta-cells as well as dopaminergic cells [28]. Another mechanism is insulin resistance, which inhibits insulin signal transduction which contributes to T2DM pathogenesis, also an imbalance in insulin inside the brain could also promote PD progression by impairing mitochondrial function [29] through an increase in reactive oxygen species (ROS) generation [30, 31]. Another important mechanism is neuroinflammation, where the microglia plays a crucial role in PD [32], whereas its activation plays a key role in the dysregulation of energy homeostasis, promotes excessive glucose levels, and has an influence on the occurrence of T2DM [33]. In this review, we have incorporated and depicted an overview of studies with epidemiological evidences supporting the association between T2DM and PD in Table 1.

T2DM and Neurodegeneration: the Role of Insulin

The pancreatic beta-cells secrete insulin, a polypeptide hormone with a molecular weight of approximately 6000 Da [47]. The protein insulin is made up of two chains—an A chain, which has 21 amino acids, and a B chain, which has 30 amino acids—joined by sulphur atoms. Proinsulin, a prohormone molecule with 74 amino acids, is the precursor of insulin. Only a small amount of proinsulin is secreted normally since it is a relatively inert protein. The proinsulin molecule is split into two parts in the endoplasmic reticulum of beta cells, resulting in the A and B chains of insulin and an intervening, biologically inactive C peptide [48, 49]. Insulin enters the brain after being secreted by pancreatic b-cells, and this process is tightly controlled and may be altered by conditions like obesity, diabetes mellitus, fasting, and neurodegenerative diseases [50]. In fact, the brain has historically been assumed to be insensitive to insulin since it does not increase glucose absorption or metabolism in the brain. However, in the last two decades, studies in this domain have uncovered specific insulin effects in the brain, and there are growing evidences that reveals insulin to act in both central and peripheral brain regions and serves several functions [51, 52]. The development of the nervous system is found to be highly influenced by insulin action in the brain, which controls proliferation, specialisation, and nerve growth [18, 53]. Additionally, brain insulin functions as a neuroprotector by inhibiting the effects of apoptosis, β -amyloid toxicity, oxidative stress, and ischemia [31, 54, 55]. If the brain insulin metabolism is impaired, the protective effects of insulin may be reduced [56], which leads to cognitive impairment and depression as well as the lack of insulin action in the brain, and may increase the risk of neurodegenerative disorders [30, 57]. Insulin disturbances

Table 1 Concomitance between T2DM and PD

Study design	Study area	Objective	Sample size	Method	Result	Conclusion	References
Case-control	Italian population	Glucose metabolism abnormalities	110 patients	<ul style="list-style-type: none"> • Oral-glucose-tolerance-test 	<ul style="list-style-type: none"> • PD patients with dementia • ↑ disease duration • ↑ motor disability 	<ul style="list-style-type: none"> • Insulin resistance • ↑ PD with dementia • ↑ PD without dementia 	[34]
Inter-group comparative analysis	South Korea	Effect of DM on PD patients	671 patients	<ul style="list-style-type: none"> • Neuroimaging analyses • DAT • Assessment of longitudinal changes in LED 	<ul style="list-style-type: none"> • ↓ baseline DAT availability • ↓ working memory • ↓ frontal/executive function • ↑ longitudinal changes in LED 	<ul style="list-style-type: none"> • DM patients with PD • ↓ baseline striatal dopamine • ↓ cognitive function • ↓ brain structural alterations 	[35]
Retrospective, case-control	Italy	Clinical features of patients with idiopathic PD (IPD)	PD ($n=89$); control ($n=89$)	<ul style="list-style-type: none"> • UPDRS 	<ul style="list-style-type: none"> • Higher UPDRS motor score • Severe Hoehn and Yahr staging 	<ul style="list-style-type: none"> • The onset of DM before the onset of PD • ↑ extreme PD symptoms 	[36]
Meta-analysis	Italy	Relationship between pre-existing DM and PD	Nine studies/1,947 citations	<ul style="list-style-type: none"> • Data obtained from electronic databases 	<ul style="list-style-type: none"> • Onset of DM before the onset of PD • ↑ risk for future PD 	<ul style="list-style-type: none"> • DM is a risk factor for PD 	[37]
Cohort study	America	Association between T2DM and PD	21,841 participants	Questionnaire	<ul style="list-style-type: none"> • ↑ DM had an PD risk 	<ul style="list-style-type: none"> • No conclusive evidences, so more studies remain to be established 	[38]
Cross-sectional study	Spain	Association between PD and DM	79 PD and 4919 controls	NEDICES	<ul style="list-style-type: none"> • No association between prevalence of PD and DM 	<ul style="list-style-type: none"> • The risk of PD in DM might be limited to longer disease duration 	[39]
Retrospective, cohort study	Spain	Association between T2DM and subsequent PD	81,90,323 participants	Linked English national Hospital Episode Statistics and mortality data	<ul style="list-style-type: none"> • The relative increase was greater in those with complicated T2DM and when comparing younger individuals 	<ul style="list-style-type: none"> • ↑ rate of subsequent PD following T2DM 	[40]
Prospective	Northeast America	Obesity and DM are related to the risk of PD	656 PD	Medical history and anthropometric variables	<ul style="list-style-type: none"> • DM was not significantly associated with PD risk 	<ul style="list-style-type: none"> • No evidence between baseline DM and PD 	[41]
Prospective	Singapore	Impact of DM in PD patients	PD, DM ($n=12$); control ($n=65$)	MRI imaging and neuropsychological assessments	<ul style="list-style-type: none"> • ↓ routine cognitive screening tests (MMSE and MOCA) • ↑ atrophy in the cortical White matter 	<ul style="list-style-type: none"> • DM in PD • ↑ cognitive decline 	[42]
Retrospective, cohort study	Taiwan	Age-sex-specific incidence, relative risk of PD	DM ($n=603,416$); control ($n=472,188$)	Incidence rate and relative risk of PD evaluation	<ul style="list-style-type: none"> • DM elevated risk of PD patients especially in women and younger patients 	<ul style="list-style-type: none"> • A stronger link between DM and young-onset PD deserves further investigation 	[43]

Table 1 (continued)

Study design	Study area	Objective	Sample size	Method	Result	Conclusion	References
Cohort study meta-analysis	China	Associations between DM and the risk of PD	1,761,632 individuals from 7 population	Data obtained from electronic databases (PubMed, Embase, and Scopus)	↑DM associated with risk 38%	More large-scale prospective studies are warranted to further clarify this association and its mechanism	[44]
Case-control meta-analysis	China	Association between DM and the risk of PD	PD patients ($n=21,395$); control ($n=84,579$)	Data obtained from databases	↓DM association with future PD	DM individuals may have a decreased incidence of PD despite significant heterogeneity	[45]
Cohort study	USA	Relationship between DM and future risk of PD	288,662 with and without PD	Questionnaire	PD risk was higher amongst diabetic patients	↑ PD with DM ~40%	[46]

PD, Parkinson's disease; T2DM, type 2 DM mellitus; DM, DM mellitus; DAT, dopamine transporter; LED, levodopa-equivalent dose; UPDRS, Unified Parkinson's Disease Rating Scale; NEDICES, Neurological Disorders in Central Spain study; HR, hazard ratio; CI, confidence interval; MMSE, Mini-Mental State Examination; MOCA, Montreal Cognitive Assessment

associated with the diabetic phenotype (hyperglycemia, hyperinsulinemia, and hypercholesterolemia) have been linked to brain atrophy and PD pathological features [58]. It is still unclear whether insulin resistance is a cause or a result of PD; however, insulin action has been shown to play an important role in PD pathogenesis. Emerging data highlights the significance of insulin dysregulation that may lead to neuropathophysiological conditions, given the fact that the role and processes behind insulin action in the human CNS still raise many issues and are far from being fully understood (Fig. 1).

Insulin Signalling in Brain

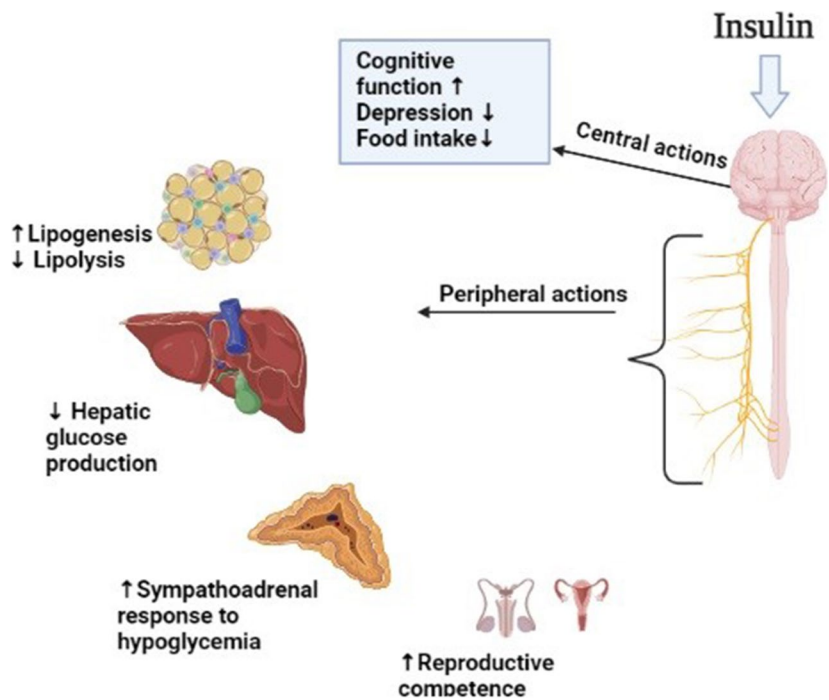
In the brain, insulin will bind to its receptors causing the phosphorylation of substrates, Shc, and insulin receptor substrate (IRS) where IRS is responsible for the PI3K (Phosphatidylinositol-3-kinase)-Akt (also known as PKB (Protein kinase B) cascade and Shc for Ras-MAPK (mitogen-activated protein kinase) that functions in food behaviour, learning, memory, and neuromodulation [59]. For the activation of PI3K-AKT, initially, insulin binds to the α -subunit of the insulin receptor, leading to the formation of dimers and autophosphorylation of β -subunits [60]. PI3K first transforms the minor phospholipid from phosphatidylinositol (3,4)-bisphosphate (PIP₂) to phosphatidylinositol (3,4,5)-trisphosphate (PIP₃), and this conversion is important for engaging PKB or AKT to the membrane of the cell [61]. AKT is responsible for the regulation of several other proteins in the insulin signalling cascade, including mTORC1 and FOXO1 [62]. It is also an active participant in several different signalling pathways of cell proliferation, cell growth, and metabolism [63]. Insulin binding also activates another series of cascades leading to MAPK pathway [62]. Small G-protein Ras is activated by the phosphorylation of guanosine triphosphate which further results in the MAPK-ERK cascade signalling [64]. MAPK is the pathway through which insulin can affect both learning and memory [65]. It is also responsible for the initiation of synaptic plasticity and cell differentiation through the transcription of various genes [66]. From these studies, it is evident that the insulin plays a major role in the brain and the deviation in their levels do have an impact on the metabolic and cognitive functions (Fig. 2).

Impact of Methylglyoxal in T2DM and PD

Methylglyoxal and T2DM

Certain conditions such as hyperglycaemia and glycation, a random event that typically takes place under biological circumstances, are generally aggravated in T2DM [67]. The key

Fig. 1 Function of insulin in the brain. Insulin controls peripheral activities in the brain by autonomic nervous system (ANS) and the hypothalamic-pituitary axis (HPA). Insulin specifically acts in the hippocampus and prefrontal cortex to enhance cognitive function and lessen depression symptoms. Insulin reduces food intake and helps people lose weight by acting in the hypothalamic nuclei. Insulin functions in the brain to reduce hepatic glucose synthesis, enhance lipogenesis, reduce lipolysis, and boost the sympathoadrenal response to hypoglycemia via the efferent ANS to target organs. The hypothalamic-pituitary-gonadal axis is the pathway via which insulin improves reproductive competence



factor thought to be responsible for diabetes complications is hyperglycaemia, which occurs when there is an overwhelming quantity of glucose in the blood [68]. Methylglyoxal (MGO) is the main hazardous oxoaldehyde that is formed during hyperglycaemia in diabetes mellitus. In reality, the healthy levels of MGO in human plasma are about 150 nM, whilst

T2DM individuals have been shown to have two to six times elevated levels [69, 70]. Prevailing thought is that methylglyoxal (MG), a reactive metabolite generated from glucose, and its breakdown by the glyoxalase pathway play a significant role in metabolic dysfunction, which connects hyperglycaemia to the onset of vascular problems in diabetes [71]. With a

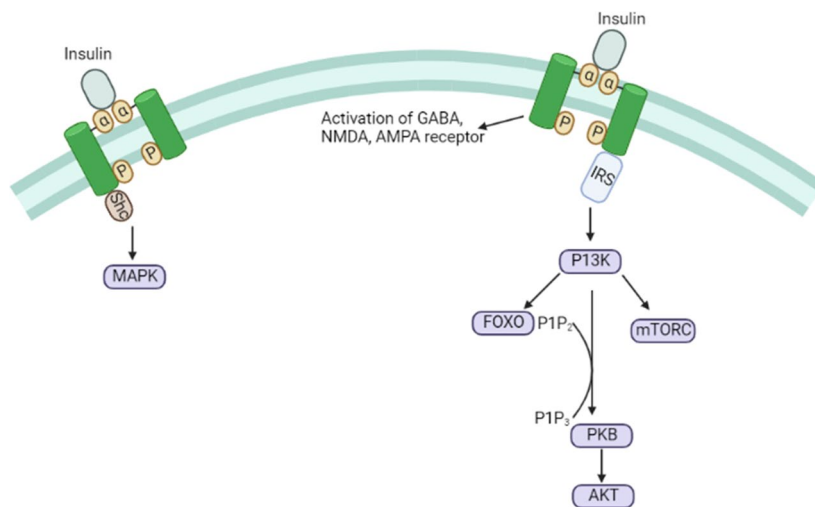


Fig. 2 Signalling pathways involved in insulin signals in the brain. When insulin ligand binds to its receptor, the receptor substrates IRS and Shc were phosphorylated. IRS activates PI3K-Akt pathway and insulin signalling cascade such as mTORC and FOXO1. PI3K first converts PIP₂ to PIP₃ that helps in engaging the Akt to the cell membrane. Shc activates MAPK pathway by activating the Ras protein through phosphorylation. Insulin binding also helps in the expression and recruit-

ment of other receptors like GABA, NMDA, and AMPA. IRS, insulin receptor substrate; mTORC, mammalian target of rapamycin complex; FOXO1, Forkhead box protein O1; MAPK, mitogen-activated protein kinase; PI3K, phosphatidylinositol-3-kinase; PIP₂, phosphatidylinositol (3,4)-bisphosphate; PIP₃, phosphatidylinositol (3,4,5)-trisphosphate; GABA, gamma-aminobutyric acid; NMDA, N-methyl-D-aspartate; AMPA, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid

molecular weight of 72 Da, MGO is a-oxoaldehyde metabolite that is primarily produced as a by-product of glycolysis after the spontaneous breakdown of the triose phosphate intermediates glyceraldehyde-3-phosphate (G3P) and dihydroxyacetone phosphate (DHAP) [72]. Other minor sources of MGO production include the breakdown of glycated proteins [73], the threonine catabolism [74], and the metabolic activity of ketone bodies, where MGO is produced from the further oxidation of hydroxyacetone, which is generated from acetone hydroxylation [75, 76]. Nucleotides, lipids, and proteins undergo spontaneous chemical change as a result of MGO activity. It alters DNA primarily by forming the imidazopurinone adduct 3-(2'-deoxyriboseyl)-6,7-dihydro-6,7-dihydroxy-6/7-methylimidazo-[2,3-b]purine-9(8H)-one (MGdG) by interacting with deoxyguanosine (dG) [77]. In addition, it can alter amino acids in proteins by forming advanced glycation end products (AGEs) by interacting with arginine, cysteine, and lysine residues in proteins [78]. The primary pathway for MGO detoxification in normoglycemic circumstances is the glyoxalase system, which also protects cells from MGO toxicity. During physiologic conditions, glyoxalases 1 and 2 (Glo-1 and Glo-2) are primarily responsible for the metabolism of MG, which results in the synthesis of D-lactate through a process that employs glutathione as a coenzyme. When reduced glutathione (GSH) is present, these enzymes react with MGO to create lactate [79]. In diabetic individuals, MGO plays a critical role in endothelial dysfunction that results in insulin resistance, hypertension, and nephropathy. In glycation processes, it is noticeably more sensitive than glucose. Additionally, AGEs are well-established, scientifically proven risk factors for the development of diabetes and associated consequences [78]. MG-modified proteins and MG-modified amino acids, also known as protein-bound AGEs and AGE-free adducts, correspondingly, are present in tissues and bodily fluids [80]. According to Stitt (2010), there were comparable increases in MG-H1 and CEL protein-bound and free adducts in plasma, suggesting that clinical translation may make MG-derived AGEs evaluated in plasma or serum biomarkers or risk indicators for vascular consequences in diabetes. These pieces of evidences show that AGEs and MGO produced from MG may be the best diagnostic biomarkers for the disease.

Methylglyoxal and PD

Researches have suggested that PD and T2DM may also be related [36, 81]. Here, it is hypothesised that PD and T2DM share a same fundamental causative mechanism that entails elevated levels of MG. One of the key characteristics of PD is a rapid decline in cellular reduced glutathione (GSH) levels in the initial phases of the disease. This leads to a reduction in the activity of the glyoxalase system, the primary catabolic pathway of the most significant glycation agent, MG [82]. The AGE levels rise as a result of the carbonyl stress, which

eventually causes oxidative stress and further AGE formation. The loss of dopaminergic neurons and an increase in oxidative stress in the brain may both be caused by this harmful loop. According to Lee et al. [83], it was found that MGO could cause a-syn to oligomerize and also reduce the ability of glycated a-syn to bind to membrane in PD. Since PD is well recognised to be closely linked to the build-up and aggregation of α -synuclein. Additionally, beta-synuclein is highly glycation-susceptible, and beta-synuclein “knockout” animals had higher levels of MG, glycation stress, and glyoxalase activity [84]. Parkin affects energy metabolism, as well as a cell-based investigation has shown this [85]. Its over-expression was discovered to inhibit glucose uptake, reduce glycolytic rate, and promote oxygen consumption [86]. The generation of MG as a result of triosephosphate accumulation due to diminished NAD⁺ accessibility or impaired glyceraldehyde-3-phosphate dehydrogenase activity is one effect of excessive glycolysis, as mentioned above. It is intriguing to learn that PD is linked to mitochondrial malfunction, which may impede NADH from being oxidised back to NAD⁺ (through the malate or glycerol-phosphate shuttles) and enhance oxidative stress to glyceraldehyde-3-phosphate dehydrogenase in the cerebral cortex [87]. Therefore, these findings suggest that enhanced or inadequate glycolysis due to over production of MG leads to the PD phenotypes (Fig. 3).

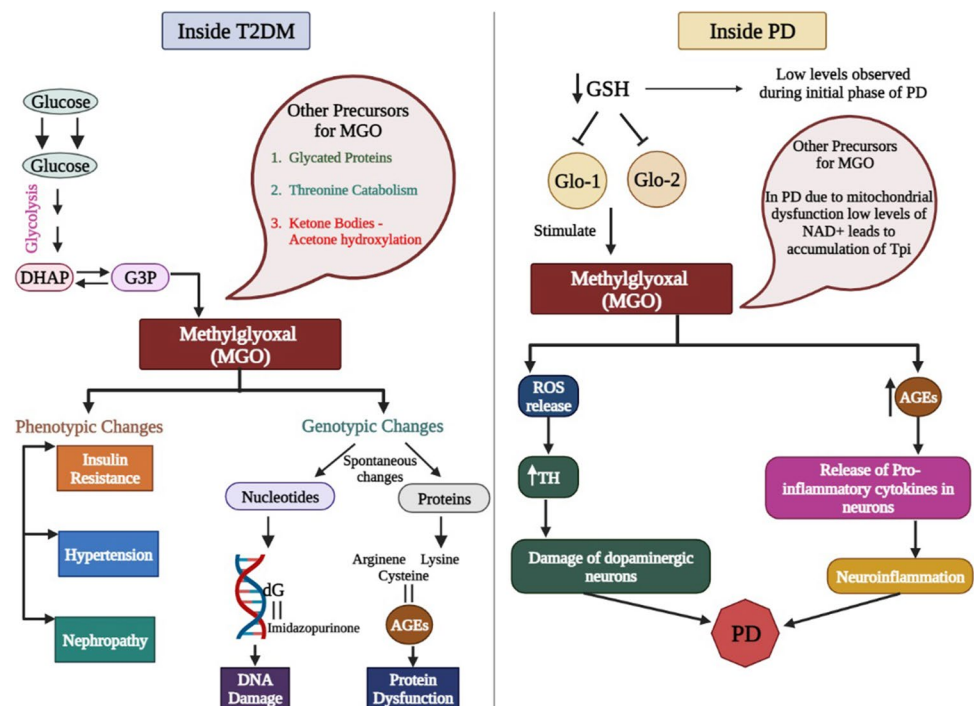
Common Problems Observed in Both T2DM and PD

An association between T2DM and PD was first reported by Sandyk in 1993, where it was noted that PD patients with co-existent T2DM had worse motor symptoms and reduced response to treatment [88]. Also, a high prevalence of impaired glucose tolerance has been reported amongst PD patients (50–80%); however, a more recent estimate suggests that overtly impaired glucose metabolism occurs in only around 20% [89]. So there has to be a commonality between the characteristic features of the both the disease conditions in a similar pattern which has been explained below elaborately.

Insulin Dysregulation in PD

A peptide hormone called insulin is released into the bloodstream by pancreatic beta cells in response to postprandial hyperglycemia. Insulin receptors are expressed in the basal ganglia [90] and in the substantia nigra [91], which are the areas of the brain that are mostly affected in PD. Since a long time ago, it has been discovered that the hypothalamus, olfactory bulb, and midbrain all contain detectable levels of insulin [92]. According to experimental data, adult neuronal cells coming from the hippocampus and olfactory bulb as well as pyramidal neurons in the cortex produce insulin [93]. It has

Fig. 3 Correlating the influence of methylglyoxal on T2DM and PD. This image depicts the possible influence of methylglyoxal on causing the T2DM and PD pathogenesis. In T2DM, glycolysis is the major factor for the release of MGO, whereas in PD it gets released due to low levels of GSH. Further in T2DM, it has a role in causing both phenotypic as well as genotypic changes. In PD, it impacts in two ways: (1) by increasing oxidative stress and (2) neuroinflammation in the cells



been discovered that there is a reduction in insulin signalling as a result of physiological changes brought on by ageing, particularly in PD patients. According to studies, patients with PD had much more insulin resistance and less insulin receptor mRNA in their substantia nigra pars compacta (SNpc) than age-matched controls [94, 95]. The loss of IGF-1 in the frontal cortex is substantially greater than that of insulin and IGF-1 signalling, and these changes are linked to higher levels of oxidative stress indicators and α Syn build-up in PD [96]. The suppression of IR-IRS-1-PI3K/AKT pathway in the hippocampus region of the brain, which in turn boosts the levels of phosphorylated IRS-1 at serine residues 636 and 616, would be the most likely molecular route by which insulin would be reduced in PD. Also, the phosphorylation of IRS-1 on serine residues, a crucial element of functional insulin signalling, prevents insulin/IGF-1 from binding to the IR and subsequent activation of downstream effectors. Further IR stimulates the aggregation of α Syn in PD [29]. Insulin resistance is linked to a more severe phenotype, a faster pace of disease progression, and a higher likelihood of cognitive deterioration in PD patients. These findings demonstrate that insulin dysregulation is a significant issue in both PD and T2DM. Figure 4a provides a brief explanation of the insulin dysregulation mechanisms between PD.

Oxidative Stress and Mitochondrial Dysfunction

An imbalance in the levels of ROS and difficulty of the body to outrage its toxic reactive intermediates leading to

cellular damages are known as oxidative stress. A hyperglycaemic state can lead to an increase in the levels of oxidative stress-induced DNA damage markers such as 8-hydroxy-2'-deoxyguanosine (8-OHdG) and 8-oxo-7, 8-dihydro-2'-deoxyguanosine; lipid-peroxidation products measured as thiobarbituric acid-reactive substances (TBARS); protein oxidation products such as nitrotyrosine and carbonyl levels and also lower the activity of antioxidant enzymes. Exposure of β -cell line and isolated pancreatic islet cells to oxidative stress has been shown to inhibit the promoter activity and mRNA expression of the insulin gene therefore, decreasing insulin gene expression [97]. Oxidative stress is also strongly suspected to be involved in chronic hyperglycaemia-induced insulin resistance [98]. Increased ROS production in T2DM patients is thought to activate many detrimental pathways including hexosamine pathways, advanced glycation end-products (AGEs) formation, and PKC β 1/2 [99]. Hyperglycemia condition can induce oxidative stress by several mechanisms such as glucose autooxidation, polyol pathway, AGE formation, and PKC β 1/2 kinase. Elevated free fatty acids, leptin, and other circulating factors in T2DM patients may also contribute to cause ROS overproduction. Similarly, even in PD brains, the loss of dopaminergic neurons occurs mainly due to ROS levels which result from dopamine metabolism, low glutathione (GSH), and high levels of iron and calcium in the SNpc [100]. Additionally, the brain contains high concentrations of

polyunsaturated fatty acids, which under oxidative stress conditions result in lipid peroxidation and the generation of toxic products [101]. Therefore, it is evident that excessive free radical generation, known to promote oxidative stress and cytokine production, appears to be the main

trigger factors for cellular dysfunction including impaired insulin signalling and mitochondrial dysfunction resulting into T2DM and PD. In Fig. 4b, we have depicted the possible mechanism of oxidative stress in T2DM and PD in a comparable manner.

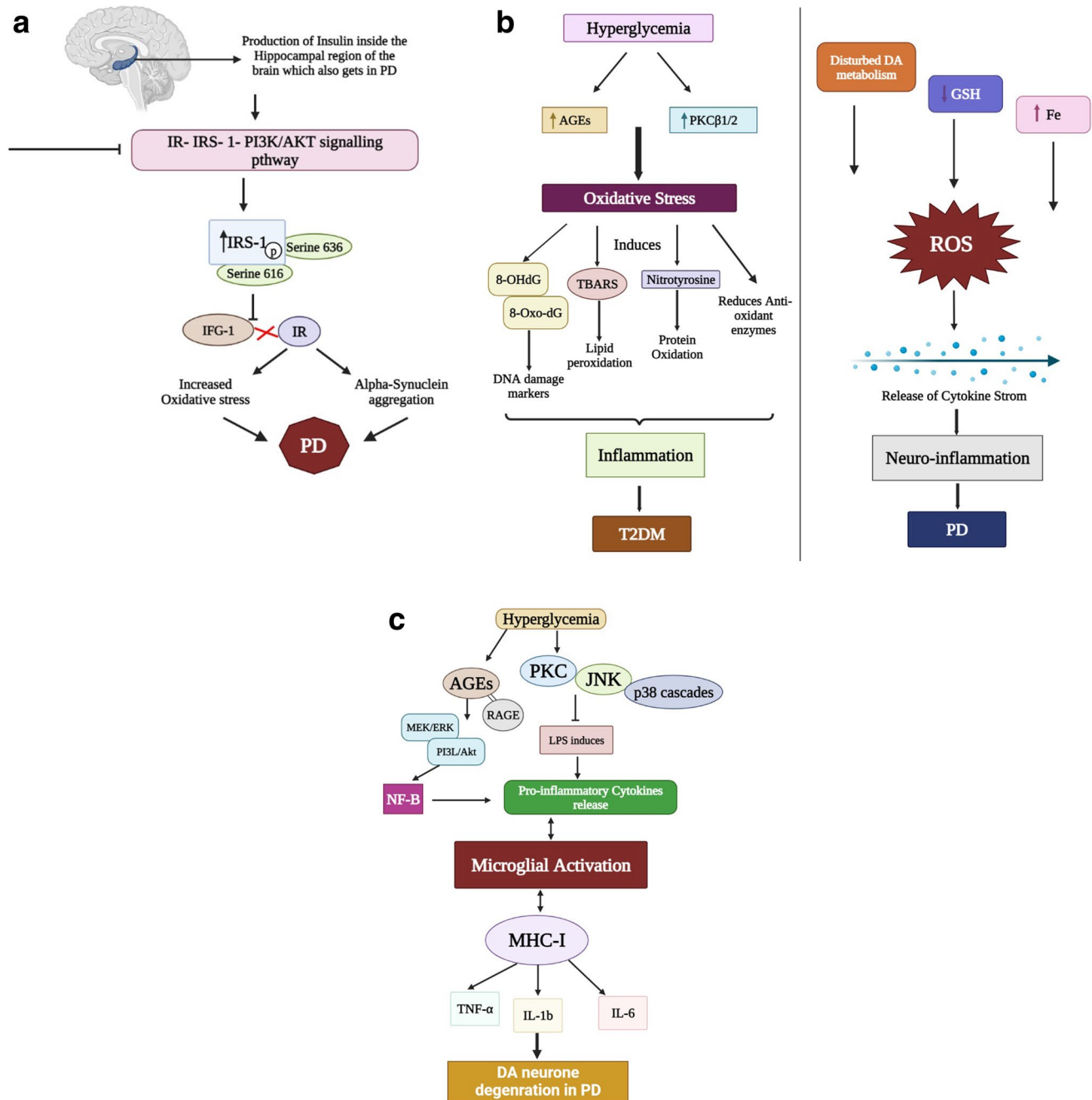


Fig. 4 (a) Dysregulation of Insulin in PD pathogenesis. In PD, the production of insulin is hindered which directly or indirectly inhibits the IR-IRS-1-PI3K/AKT signalling pathway of insulin production. This results into consecutive production of either oxidative stress or aggregation of α -syn which is mostly observed in PD. (b) Impact of oxidative stress on T2DM and PD. This image lets us to easily compare the similarities and differences amongst the impact

of oxidative stress on T2DM and PD. In both the cases, inflammation is the major role of disease pathogenesis. (c) Importance of microglia in T2DM and PD. Generally, the activation of microglia causes morphological as well as functional changes which results into disease conditions. This image enables us to understand the role of microglia activation in both T2DM and PD and how it triggers inflammation more easily

Microglial Activation and Inflammation

Microglial activation is a process of morphological and functional change that can be triggered by inflammation and vascular injury. Microglial stimulation and the consequent production of inflammatory mediators, which ultimately cause injury to brain tissue, are the principal manifestations of neuroinflammation. Microglial activation and inflammatory responses are modulated by signalling molecules, such as MAPKs and triggering receptor expressed on myeloid cells 2 (TREM-2) [102, 103]. The three main MAPKs are extracellular signal-regulated kinases 1 and 2 (ERK1/2), p38 MAPK, and c-Jun N-terminal kinases (JNK), which control a variety of cellular processes and are crucial in controlling the expression of pro-inflammatory cytokines like tumour necrosis factor- α (TNF- α) and interleukin-1 α (IL-1 α) [104]. According to a recent study, chronic cerebral hypoperfusion CCH-induced stimulation of microglia encourages the production of pro-inflammatory substances, which further contribute to cognitive dysfunction and long-term potentiation (LTP) impairment [105]. Additionally, in humans and rats with T2DM, microglial activation and an inflammation are also involved [106, 107]. Inflammation can be exacerbated by mitochondrial ROS brought on by hyperglycemia by activating the PKC, JNK, and p38 cascades [108]. JNK inhibition lessens the lipopolysaccharide (LPS)-induced activity of inflammatory cytokines in microglia, as activation of p38 is crucial for cytokine production and release [109]. In an in vitro model using microglial cell line, AGEs in diabetes trigger the MEK/ERK, PI3K/Akt, and NF- κ B pathways via RAGE, which causes the activation of pro-inflammatory cytokines [110]. These studies suggest that the JNK cascade is the most important mechanism contributing to oxidative/inflammatory stress and microglial activation during acute glucose fluctuations by evaluating the efficacy of all signalling pathway inhibitors especially in T2DM. In an effort to pinpoint a putative mechanism that might contribute to neuroinflammation and possibly underlying the aetiology of PD, the intricacy of microglial stimulation signals came under great scrutiny. In particular, areas of the SN and striatum in PD patients were found to have MHC-II immunoreactive microglia [111, 112]. M1 pro-inflammatory microglial cell activation encourages the production of substances that damage DA neurons in the brain. Followed which nuclear factor- κ B is the primary initiator of inflammation and the first signal to do so. Other pro-inflammatory cytokines, such as tumour necrosis factor- α , interleukin 1b, and IL-6, are then released into the body [4]. Therefore, these results demonstrate that there is a tight relationship between microglia activation and inflammation in both T2DM and PD, which requires further study (Fig. 4c).

Drugs Available for T2DM

People with T2DM are probable to develop many health complications such as heart problems, retinal issues, and brain strokes. The finest kind of medication will depend on a number of factors, including the severity of diabetes, age, and whether with additional health issues. Here, we describe the drugs developed to treat T2DM, which has lack of scientific findings with convincing results (Table 2). As previously mentioned, T2DM and PD share several common characteristics, and it has been suggested that there is a list of drugs that have influenced the signalling pathways in both T2DM and PD, and are listed in detail in (Table 3).

Mechanism of Action of Metformin in T2DM

Since a decade, metformin has been prescribed as a common drug to treat T2DM [174]. Metformin acts as an inhibitor of mitochondrial complex I, which ultimately results in the decrease in ATP synthesis and an increase in AMP levels activating the AMPK pathway [175]. AMPK regulates glucose metabolic pathway and energy balance [176]. As an AMPK activator, metformin regulates all the major metabolic pathways for glucose uptake, carbohydrate metabolism, and lipid metabolism [176]; and it was thought to be a key therapeutic target for obesity, T2DM, ageing, and neurodegeneration [215]. It functions by lowering the insulin-mediated hepatic glucose synthesis levels and by improving insulin sensitivity in T2DM patients [216], it is neuroprotective in human neural stem cells against amyloid-beta-induced mitochondrial dysfunction mediated through the PI3K/Akt pathway [217], and long-term treatment with metformin seems to decrease the risk of cognitive decline in diabetic patients [218] and improve depressive and cognitive performance, changing the glucose metabolism in depressed patients [164]. The inhibition of insulin and insulin growth factor receptor signalling by metformin results in alterations in metabolic balance. Metformin's antidiabetic actions are primarily brought by the AMPK pathway [177] but it can also occur in an AMPK-independent mechanism [178]. Genes coding for the enzymes involved in glucogenesis are reduced by AMPK through phosphorylation of CREB binding protein (CBP) [179]. Activation of AMPK results in decreased adenosine monophosphate which inhibit fructose-1,6-bisphosphatase-1, a major enzyme in gluconeogenesis [180]. Similarly, metformin causes AMPK activation which involves in reduction of PD pathogenesis [181]. When AMPK deficiency is detected in *Drosophila*, the characteristics of PD was observed [182]. From the data, a possible mechanism of metformin for its neuroprotective role has also been revealed and it is reported to be a hotspot in linking PD and T2DM. Metformin functions similarly in PD too by inhibiting complex I which then activates the AMPK pathway. Complex I

Table 2 List of drugs used in T2DM

S. no	Plant/drug	Extract/compound	Mechanism of action	Organ function	Findings	Recommended/not recommended	Reference
1	<i>Moringa oleifera</i>	Moringa seeds	Lessening the expression of iNOS, IL-1 β , NO, TNF- α	Liver	↓ plasma glucose levels, HbA1c	Recommended	[113]
2	<i>Garcinia macrophylla</i>	5,7,4',5'',7'',3''',4''-heptahydroxyflavanone [3–6''] flavones	Antibacterial, antioxidant, and anti-T2DM	-	↓ glucose level	Recommended	[114]
3	Albendazole lansoprazole	-	Activation of AMPK and PPAR pathways and incretin-like effect	-	↑ glucose, HbA1c, triglyceride, LDL, leptin ↓ insulin and HOMA- β	Recommended	[115]
4	Remogliflozin etabonate (100 mg and 250 mg)	-	SGLT2 inhibition improves urinary glucose excretion thereby decreases blood glucose levels	-	↓ glucose levels	Recommended	[116]
5	Metformin hydrochloride	-	Lessens intestinal glucose absorption, declines hepatic glucose production, increases insulin sensitivity	Liver	↓ HbA1c levels, FBG, and weight loss	Recommended	[117]
6	Remogliflozin metformin teneligliptin	-	-	-	↓ glucose levels, weight	Recommended	[118]
7	<i>Rhizoma coptidis</i>	Berberine and polyphenols	Improves β cell dysfunction	Liver adipose tissues	↓ IL-6, TNF α ↑ islet cell viability	Recommended	[119]
8	Alkaline and flavone category compounds	MAPK1, STAT3, INSR, and 38 signalling pathways	-	-	Insulin resistance	Recommended	[120]
9	<i>Morus alba</i>	Chlorogenic acid, isoquercitrin, and quercitrin	-	-	Glucose absorption, insulin secretion, antioxidant, anti-inflammatory, antihyperglycemic, and antihyperlipidemic activities, declined obesity	Recommended	[121]
10	<i>Zingiber officinale</i>	-	-	Heart	↑ diastole function and blood pressure ↓ HbA1c, triglycerides	Recommended	[122]
11	Glargine/lixisenatide	-	GLP-1 RA, control postprandial glucose improved β -cell function	-	↓ HbA1c, weight gain ↓ glucose levels	Recommended	[123]
12	Gegen Qinlian decoction berberine	-	Proinflammatory cytokines Nfkb1, Stat1, and Ifng1	Pancreas	Enriched gut microflora ↓ glucose levels	-	[124]
13	Gegen Qinlian decoction (meta-analysis study)	-	-	Pancreas	↓ glucose levels ↑ islet function ↓ insulin resistance and sensitivity indexes	A need for clinical prognosis and islet function	[125]

Table 2 (continued)

S. no	Plant/drug	Extract/compound	Mechanism of action	Organ function	Findings	Recommended/not recommended	Reference
14	Huanglian Jiedu decoction	Decoction has Huanglian, Huangqin, Huangbo, and Zhizi mixture	Targets AKT1, IL-6, FOS, VEGFA, CASP3	Pancreas	↓ glucose levels, HbA1c	Recommended	[126]
15		-	Enhance GLUT4, INSR, MAPK1	-	Improve in fasting glucose level, lipid level, insulin sensitivity index	Recommended	[127]
16	<i>Nigella sativa</i> (meta-analysis study)	Thymol, thymohydroquinone, dithymoquinone, nigellone, alpha-hederin, flavonoids, and fatty acids	Antidiabetic, antioxidant, anticancer, hypolipidemic, and anti-inflammatory properties	-	↓ HbA1c, glucose level, insulin resistance	Recommended	[128]
17	Meformin with Chinese traditional medicine	-	-	-	Lessened the hyperglycemia, altered gut microflora, improved insulin resistance	Recommended	[129]
18	GLP-1RAs and oral anti-diabetic drugs (meta-analysis)	-	-	Heart	Improvement in left ventricular diastolic function	Recommended	[130]
19	GLP-1 RA and SGLT-2i (meta-analysis)	-	-	Pancreas	5 studies—GLP-1 RA showed decreasing effect in cardiovascular complications, glucose reduction	Recommended	[131]
20	Metformin and DDP-4 inhibitors	-	AMPK pathway activation, NFK-β, and mTOR suppression	-	Adverse outcome from COVID-19	Not recommended	[132]
21	Statin, fibrates, and its combination	-	-	-	↓ lipid levels	Recommended	[133]
22	Incretin, DDP-4 inhibitors, and GLP-1 RA	-	-	-	↓ HbA1c	Recommended	[134]
23	Anti-hyperglycaemic therapies (GLP-1 RA and SGLT2i)	-	-	-	No change in HbA1c levels	Recommended	[135]
24	Metformin (meta-analysis)	-	-	-	↓ HbA1c ↓ glucose levels	Recommended	[136]
25	Aspirin and statin	-	-	Heart	↓ cardiovascular complications in T2DM	Recommended	[133]
26	Tofogliflozin with DDP-4 inhibitors	-	-	Pancreas	↓ postprandial glucose	Recommended	[137]
27	<i>Momordica charantia</i>	-	-	Pancreas	↑ insulin secretion ↓ glucose uptake	Recommended	[138, 139]

Table 2 (continued)

S. no	Plant/drug	Extract/compound	Mechanism of action	Organ function	Findings	Recommended/not recommended	Reference
28	Metformin	-	-	Pancreas	↑ epigenetic markers	Recommended for diagnosis	[140]
29	Chenpi	5-OH PMFs	Antidigestive and anti-inflammatory AMPK activation		↓ hepatic steatosis ↓ glucose levels	Recommended	[141]
30	Aloe vera	-	-	-	↓ glucose levels	Recommended	[142]
31	Berberine	-	-	Pancreas	↓ HbA1c levels ↓ cholesterol and triglyceride	Recommended	[143]
32	<i>Pueraria lobata</i> (meta-analysis)	-	-	Pancreas	↑ insulin secretion Anti-diabetic activity	Recommended	[144]
33	SGLT-2i and GLP-1 RA	-	-	Heart	↓ cardiorenal problems, stroke	Recommended	[145]
34		-	-	Heart kidney	↓ renal risk No significance in cardiovascular condition	Recommended	[146, 147]
35	Sulfonylureas, DPP-4 inhibitors, meglitinides	-	-	Heart	↑ myocardium infection	Recommended	[148]
36	Acupuncture with Chinese herbal medicine (meta-analysis)	-	-		↓ glucose levels ↑ insulin sensitivity and resistance	Recommended	[149]
37	Ipragliflozin (meta-analysis)	-	-	-	↑ genital infection No significance in glucose level	Not recommended	[150]
38	Ginseng supplementation	-	-	-	↑ fasting glucose, postprandial insulin, and HOMA-IR	Recommended	[151]
39	Shenqi Jiangtang Granules Baihu Jia Renshen Decoction Tianqi Jiangtang Capsule (meta-analysis)	-	-	Pancreas	Lipid levels were altered ↓ hypoglycemia levels	Recommended	[152–154]
40	Chinese tradition medicine vs. Western medicine (meta-analysis)	-	-	Pancreas	Traditional medicine showed enhanced insulin sensitivity, decreasing body weight, protects from β cells	Recommended	[155]

Table 2 (continued)

S. no	Plant/drug	Extract/compound	Mechanism of action	Organ function	Findings	Recommended/not recommended	Reference
41	Cotadutide (meta-analysis)	-	-	Pancreas	↓ glucose levels, HbA1c and body weight	Recommended	[156]
42	SGD	-	Influences PI3K-Akt, AMPK, and PPAR	Pancreas	↓ glucose levels and lipid levels ↓	Recommended	[157]
43	Sancai powder	-	-	Pancreas	↓ glucose levels, HbA1c, lipid levels	Recommended	[117]
44	Zhimu-Huangbai herb pair	-	-	Pancreas	↓ glucose levels	Recommended	[158]
45	Naointong capsule	-	Arachidonic acid metabolism, fatty acid β-oxidation, and glycerophospholipid metabolism	Pancreas	↓ hyperlipidemia, hyperglycemia, myocardial infarction, insulin resistance	Recommended	[159, p. 20]
46	Pioglitazone and canagliflozin	-	-	-	↓ glucose levels, lipid levels, HbA1c	Recommended	[160]
47	Canagliflozin	-	-	Kidney	↑ renal oxygen in T2DM	Recommended	[161]
48	Pine bark extract	-	-	Pancreas	↓ HbA1c, VCAM-1, cholesterol levels, UACR	Recommended	[162]
49	Jinlida granules	-	-	Pancreas	↓ HbA1c and glucose levels	Recommended	[163]

iNOS, inducible nitric oxide synthase; *IL-1β*, interleukin-1 beta; *NO*, nitric oxide; *TNF-α*, tumour necrosis factor alpha; *HbA1c*, glycated haemoglobin; *T2DM*, type 2 diabetes mellitus; *LDL*, low density cholesterol; *HOMA-β*, homeostasis model assessment of β-cell function; *AMPK*, adenosine 5′-monophosphate activated protein kinase; *PPAR*, peroxisome proliferator-activated receptor; *SGLT-2*, sodium glucose co-transporter-2; *SGLT-2i*, sodium-glucose cotransporter-2 inhibitors; *FBG*, fasting blood glucose; *GLP-1 RA*, glucagon-like peptide 1 receptor agonists; *DDP-4*, dipeptidyl peptidase-4 inhibitors; *NFK-β*, nuclear factor kappa B; *mTOR*, mammalian target of rapamycin; *COVID-19*, coronavirus disease-19; *5-OH PMFs*, 5-demethylated poly methoxy flavones; *HOMA-IR*, homeostatic model assessment of insulin resistance; *SGD*, Shengmai-Yin and Ganmai dazao decoction; *VCAM-1*, vascular cell adhesion molecule 1; *UACR*, urinary albumin-to-creatinine ratio; *PPAR*, peroxisome proliferator-activated receptors; *PI3K-Akt*, phosphatidylinositol 3-kinase-protein kinase B; *GLUT4*, glucose transporter type 4; *INSR*, insulin receptor; *MAPK1*, mitogen-activated protein kinase 1; *IL-6*, interleukin-6; *FOS*, Fos proto-oncogene; *AP-1*, transcription factor subunit; *VEGFA*, vascular endothelial growth factor A; *CASP3*, caspase 3; *Stat1*, signal transducer and activator of transcription 1; *Irfgr1*, interferon gamma receptor 1; *STAT3*, signal transducer and activator of transcription 3

Table 3 Therapeutics targeting signalling pathways common in both T2DM and PD

Therapeutic drug	Mechanism	Model	Dosage	Methods	Result	Conclusion	References
Metformin	Amyloid aggregation	Healthy human blood	200 mmol/L	<ul style="list-style-type: none"> • Spectrophotometry • In vitro assays 	<ul style="list-style-type: none"> • ↑anti-AChE • ↑anti-Aβ aggregation 	Metformin may be regarded as an effective adjuvant to donepezil	[233]
	Oxidative stress and mitochondrial dysfunction	ASCs	25 μmol/L	<ul style="list-style-type: none"> • Western blot • Immunoblot • SDS-PAGE 	<ul style="list-style-type: none"> • ↓oxidative stress • ↓mitochondrial dysfunction • ↑adipogenicity • ↑insulin sensitivity 	Metformin may improve impaired adipogenesis and insulin sensitivity	[234]
	Microglial activation and chronic inflammation	BV-2 and bEnd3 cells	50 mg/kg	<ul style="list-style-type: none"> • Western blot • Immunohistochemistry • PCR 	<ul style="list-style-type: none"> • ↓p38 MAPK phosphorylation • ↓pro-inflammatory cytokines • ↓TLR-4 mRNA expression • ↓IL-6 • ↓TNF-α mRNA expression 	Metformin suppresses microglial activation by increasing AMPK phosphorylation	[235]
GLP-1R agonists/ GIP receptor	Amyloid aggregation	Clonal mouse insulin-secreting cell line (min6)	25 mM	<ul style="list-style-type: none"> • RT-PCR • ELISA • Western blotting 	<ul style="list-style-type: none"> • ↓hAPP-induced LC3II/I ratio • ↓hAPP-induced cleaved caspase-3 • ↓p62 expression 	Exendin-4's has the potential in the treatment of diabetic β-cell failure	[236]
	Microglial activation and chronic inflammation	Mice	5.0 mg/ml	<ul style="list-style-type: none"> • Immunofluorescence • Western blotting • Immunofluorescence staining 	<ul style="list-style-type: none"> • ↓Iba-1 • ↓IL-1β • ↓TNF-α release • ↓TNC microglial number • ↑allodynia • ↓CGRP and c-fos in the TNC 	GLP-1R agonist liraglutide might represent a new therapeutic approach for treating chronic migraine	[237]
	Impaired synaptic plasticity	Mice	–	<ul style="list-style-type: none"> • Open field assessment • Object recognition task • Water maze task 	<ul style="list-style-type: none"> • Glp1r^{-/-} mice • ↓motor activity • Normal exploratory activity and anxiety levels 	GLP-1R is crucial for some forms of memory formation and synaptic plasticity	[238, p. 20]
Insulin	Microglial activation and chronic inflammation	Mice	25 nmol/kg	Rotarod and grip strength assessment	<ul style="list-style-type: none"> • ↓increased levels of activated microglia and astrocytes • ●●●↑ GDNF 	GLP-1/GIP dual agonist has the potential to exhibit superior neuroprotective effects	[239]
	Impaired synaptic plasticity	Mice	10 nmol/kg	<ul style="list-style-type: none"> • Western blotting • Water maze • Probe test 	<ul style="list-style-type: none"> • ↓memory loss • ↑synaptic plasticity • ↓amyloid plaques • ↓pro-inflammatory cytokine levels 	GLP-1/GIP dual receptor agonist was more effective at reversing memory loss	[240]
	Oxidative stress	Cultured cortical neurons	10 M	<ul style="list-style-type: none"> • MTT assay • SDS-PAGE • UV-spectrophotometry 	<ul style="list-style-type: none"> • ↓oxidative stress • ↓necrotic and apoptotic cell death • ↓ascorbate/Fe2+-mediated lipid and protein oxidation 	Insulin may help reduce the damage caused by oxidative stress that develops in several neurodegenerative diseases	[31]

Table 3 (continued)

Therapeutic drug	Mechanism	Model	Dosage	Methods	Result	Conclusion	References
	Impaired synaptic plasticity	Mice	20 µl	<ul style="list-style-type: none"> Immunohistochemistry and image analysis Western blotting 	<ul style="list-style-type: none"> ↓rapamycin complex 1 (mTORC1) 	Intranasal insulin prevents general anaesthesia-induced apoptosis of hippocampal cells and deficits in synaptic plasticity and memory	[241]
	Microglial activation and chronic inflammation	Microglial cell line	125 ng/ml	<ul style="list-style-type: none"> Western blot Immunocytochemistry Phagocytosis assay 	<ul style="list-style-type: none"> ↑ phagocytic activity ↓NO ↓TNF production ↓ROS 	Insulin has beneficial effects on CNS injury or neurodegenerative conditions	[242]

PI3K, phosphatidylinositol 3-kinase; *Akt*, protein kinase B; *GSK-3 beta*, glycogen synthase kinase-3 beta; *FOXO1*, Forkhead box protein O1; *NF-κB*, nuclear factor kappa-light-chain-enhancer of activated B cells; *mTOR*, mammalian target of rapamycin; *DA4-JC*, dual GLP-1/GIP receptor agonist; *ASCs*, adipose-derived stromal cells; *HIAPP*, human islet amyloid polypeptide; *TNC*, trigeminal nucleus caudalis; *CM*, chronic migraine; *AMPK*, 5' AMP-activated protein kinase; *NO*, nitric oxide; *TNF*, tumour necrosis factor; *ROS*, reactive oxygen species; *iNOS*, inducible NO synthase; *ACHE*, human acetylcholinesterase; *Aβ*, amyloid beta; *bEnd3*, endothelial cells; *TLR-4*, Toll-like receptor-4; *TNF-α*, tumour necrosis factor-α; *GFNF*, glial derived neurotrophic factor

is one of the primary sources of ROS where metformin reduces the ROS generation through reverse electron transfer but will not boost their production [183]. Metformin therapy drastically reduced the amount of cytochrome C released from mitochondria into the cytosol, especially in the aged brain [35]. In the 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP)-treated PD mice model, metformin exhibits neuroprotective benefits where it enhances mobility and muscular activity [184]. Another way through which ROS is reduced is by NAD(P)H inhibition and activation of AMPK [185]. Experimentally gathered data shows that ROS is the major factor in the death of dopaminergic neurons in PD patients [186]. A drop in ROS levels will result in a significant attenuation of DNA damage [187]. The incidence of cognitive decline in people with T2DM was considerably lowered by metformin therapy [188]. Metformin reduces neuroinflammation by an increase in the antiapoptotic protein Bcl-2 [189]. In addition, metformin mediated activation of AMPK also increases serine/threonine *PP2A* (protein phosphatase 2A) that dephosphorylates and reduce αSyn levels [190]. As a result, there are numerous molecular processes explaining metformin's multiple beneficial effects, but there are limited studies conducted on therapeutic potential in PD; hence, more studies need to be elucidated in PD pathogenesis. Hence, from the studies, it is probable that AMPK targeted therapy might lessen the neurodegeneration, in PD where more research is required on therapeutics effects of metformin to confirm its neuroprotective role. The probable mechanism is depicted in Fig. 5.

Glucagon-Like Peptide-1 (GLP-1) Receptor Agonist

The GLP-1 receptor agonists are primarily known to be a group of anti-diabetic medications, and this GLP-1 has an intestinal peptide known as incretin that promotes glucose-dependent insulin secretion whilst inhibiting glucagon release to maintain blood sugar levels [191] and also GLP-1 exhibits potent and strictly glucose-dependent insulinotropic activity through unique GLP-1 receptors in the plasma membrane of pancreatic cells [192]. This inhibits glucagon secretion, which may be as clinically relevant as GLP-1's insulinotropic action [193]. Furthermore, GLP-1 decreases food intake (most likely by activating GLP-1 receptors in the central nervous system and slows stomach voidance, resulting in lower postprandial glucose excursions and food intake) [192]. Few studies have also discovered that as one progresses from normal glucose tolerance to insulin resistance, plasma GLP-1 concentrations decreases [194, 195]. Also, GLP-1 is expressed in all regions of the brain, including the frontal cortex, hypothalamus, thalamus, hippocampus, cerebellum, and substantia nigra and has a key role in neuronal metabolism and neuroprotective effect in the brain through crossing the blood brain barrier (BBB) [196]. Additionally, GLP-1 exhibits regulatory effects that includes stimulation of cell

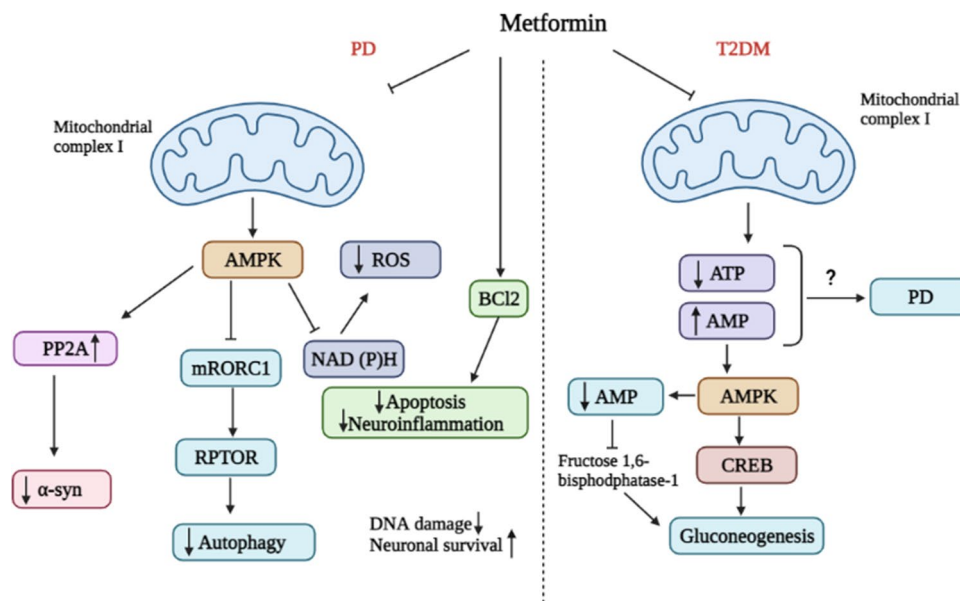


Fig. 5 Metformin: anti-diabetic and neuroprotective role. Metformin has a neuroprotective role through the AMPK pathway which is also their anti-diabetic pathway. AMPK is activated through inhibition of mitochondrial complex I and it drastically reduces the ROS levels causing neuronal survival. Metformin enhances autophagy by inhibiting mTORC1 which was phosphorylated by AMPK, anti-

apoptosis by increasing anti-apoptotic protein Bcl-2, reduces DNA damage, and neuroinflammation hence accounting for neuronal protection. Metformin reduces the α Syn aggregates by increasing PP2A. AMPK, AMP-activated protein kinase; ROS, reactive oxygen species; mTORC1, mammalian target of rapamycin complex 1; Bcl-2, B-cell lymphoma 2; PP2A, protein phosphatase 2

neogenesis, growth, and differentiation; prevention of cell apoptosis; and boosting cell survival [197]. There are several GLP-1 analogues, particularly exenatide, liraglutide, and lixisenatide, which regulates cellular pathways that protect the neurons, maintain mitochondrial function, prevent apoptosis, and respond to oxidative stress in addition to their effects on glucose homeostasis [198, 199]. GLP-1 receptor agonists exhibits neuroprotective and neurorestorative characteristics in various PD experimental models. Exenatide-treated animal models can halt 6-hydroxydopamine (6-OHDA), MPTP, and LPS-induced dopaminergic degeneration and restore DA imbalance in PD, resulting in significant improvements in behaviour and motor function [200–202]. Longer-acting GLP-1 analogues (liraglutide and lixisenatide) have also shown protective benefits and enhanced motor function in MPTP induced PD mouse model [198]. Taken together, GLP-1 receptor agonists are already a proven fundamentally anti-diabetic medication, and also there is strong evidence that GLP-1 activity has a positive impact on the treatment of PD pathogenesis, as shown in numerous PD models.

Dipeptidyl Peptidase-4 Inhibitors (DPP-4i)

The anti-diabetic medication DPP-4i (gliptins) inhibits DPP-4, a proteolytic enzyme that has shown neuroprotective effects in the brain [203], and in T2DM, the

inactivation of GLP-1 in the peripheral circulation is prevented by DPP-4 inhibitors by reducing DPP-4 activity in peripheral plasma thereby prevents T2DM condition and improves glucose metabolism [73]. DPP-4 inhibitors on elderly people with T2DM and mild cognitive impairment enhances glucose control, prevents deterioration of cognitive abilities, and also decreases β -amyloid toxicity and tau hyperphosphorylation in human neural cells [204]. DPP-4 inhibitors might improve synaptic function, lessen the loss of dopaminergic cells, and/or assist the regenerative process in PD by either increasing endogenous levels of GLP-1 and GIP both systemically and in the brain, or by acting directly on them [205]. Data from DPP-4 inhibitors in PD experimental models are inconsistent. In a rotarod test, rats were given saxagliptin before the induction of rotenone, which showed increased striatal DA production and decreased dopaminergic neuronal loss, resulting in improved motor performance and coordination [206]. However, rats given supramaximal doses of sitagliptin (a DPP-4 inhibitor with a significantly longer half-life than saxagliptin) were not protected against MPTP-induced striatal dopaminergic degeneration [207]. Thus, DPP-4 may be useful for T2DM with PD patients in terms of their initial dopamine degradation and in long-term effects it may also play a potential role in non-diabetic PD patients.

Insulin

Insulin is found to be beneficial in treating both T2DM and PD conditions. In general, insulin interacts with the brain in several ways, likely through the complex insulin/IR signalling pathway, including effects on cognition, memory, learning, and synaptic plasticity [208]; and importantly exogenous insulin can be administered nasally to prevent effects on peripheral glucose levels, and it may have benefits for PD patients [57]. At clinically significant levels, insulin enters the brain by passing the BBB, preventing neuronal death by activating protein kinase B, and regulating tau phosphorylation, beta-amyloid precursor protein metabolism, and beta-amyloid clearance in vivo [209] [210]. Intranasal insulin administration increases mood and memory whilst preserving the volume of brain regions damaged by neuropathology, and it also improves cerebral glucose metabolism which is mediated through PI3K/Akt pathway [211]. Insulin reduced α -synuclein levels in MPP-positive C6 glial cells [212]. Insulin contributes in directing the autophagic process to protect the neurons by enhancing the autophagy of harmful proteins, by suppressing mTORC1 activity, activating Akt survival molecule, and increasing mTORC2 activity [213]. Injection of intranasal insulin was conducted in PD patients which revealed that the insulin-treated patients showed improvements in verbal fluency and motor severity compared to baseline scores at the end of the exposure period, and, more importantly, there were no reports of changes in serum glucose levels or hypoglycaemia [214]. Therefore, as we know that insulin is primarily known to regulate blood sugar levels, and with the evidence provided above, it is clear that insulin, through interactions with the complex insulin/IR signalling network, can help to treat multiple processes associated with the onset of PD.

Sulfonylureas

Sulfonylurea is primarily an antidiabetic drug and there are generations of drugs under the sulfonylurea class. The sulfonylurea glimepiride is linked to altered cell signalling and synaptic membranes and provides several protective advantages for neurons and is used to treat T2DM [220]. Gliclazide is a second-generation sulfonylurea hypoglycaemic medication. It reduces the free radical synthesis or increases their scavenging by lowering oxidative stress, protecting against DNA damage, and boosting antioxidant status, and may also help to regulate the physiological diseases that underlie both T2DM and ageing [221]. Glibenclamide is another generation of sulfonylurea, it is a KATP channel inhibitor where it decreases the amyloid-induced behavioural impairments and anxiety in a rat model, and it may be used as a therapeutic target for neurodegenerative diseases which is mediated through PI3K/Akt pathway [222]. In a rodent

PD model, glibenclamide protected dopaminergic neurons by inhibiting inflammasome activation, microglial polarisation, and oxidative stress [223]. In both BV2 microglial cells and mouse PD models, glibenclamide was able to limit microglial activation and reduce excessive neuroinflammation. Furthermore, glibenclamide's anti-neuroinflammation actions may be caused through mechanisms that regulate the c-Jun N-terminal kinase (JNK) and NF- κ B signalling pathways [224]. Together, these data suggest a possible role of sulfonylureas in treating both PD and T2DM.

Thiazolidinediones

Thiazolidinediones (TZDs), such as pioglitazone and rosiglitazone, are useful to improve insulin sensitivity in muscle, adipose, and hepatic tissue and reduce systemic insulin resistance. This includes the transfer of extra fatty acids to peripheral fat, and this improves insulin sensitivity by lowering the availability of fatty acids in the liver, muscles, and circulation to treat the T2DM condition [225]. Pioglitazone, the only commercially available TZD, can enter the brain, decrease glial activation, lessen neurodegenerative-related diseases [226], lessen spatial learning impairment, and prevent tau hyperphosphorylation by improving Akt signalling [227]. Various toxin induced PD models, including MPTP [228], LPS [229], 6-OHDA [230], and rotenone [231] showed that the pioglitazone and rosiglitazone have neuroprotective effects that improve behavioural and motor responses. The suppression of pro-inflammatory pathways, as well as the modification of mitochondrial activity and oxidative stress responses, is suggested to be the causes of these effects [232]. Considered together, there is evidence that TZD action has a beneficial impact on the treatment of both T2DM and the aetiology of PD.

Traditional Indian Medicinal Plants to Treat T2DM and PD

Traditionally, moringa seeds containing isothiocyanates reported to delay T2DM rat models [113]. Similarly, biflavonoid compound isolated from *Garcinia macrophylla* plant suggested to decline blood sugar level in diabetic rats [114]. Anti-diabetic activity by benzimidazole derivatives, alben-dazole, and lansoprazole in rats showed effective treatment strategy in curing T2DM [115]. In T2DM patients, remogliflozin etabonate reported to be an effective therapeutic option in lowering glucose levels [116]. In a comparative study using metformin sustainable-release tablet and original metformin tablet, metformin sustainable-release had significantly showed the therapeutic efficiency in T2DM than original tablet [164]. In participants with insulin glargine along with oral hypoglycemic therapy received

remogliflozin which showed an alternative approach with drastic reduction of T2DM [118]. By using network pharmacology and docking analysis, mechanistic insights of *Rhizoma coptidis* were explored against T2DM which suggested to show anti-inflammatory effects thus proving to be competent in antidiabetic research [119, 120]. In a systemic review, *Morus alba* reported to be a potential therapeutic target for T2DM though it lacks more studies on synergistic effects [121]. Apart from drugs, exercise was recommended for glucose management and T2DM control [165]. In a clinical trial using *Zingiber officinale* as an add-on therapy in T2DM showed improved blood pressure, proper function of diastole, and maintenance of lipid profile [122]. In a 26-week treatment with insulin glargine/lixisenatide in Japanese T2DM patients inadequately controlled over oral antidiabetic drugs [123]. In a rat model, Gegen Qinlian Decoction and berberine showed the decline of blood glucose levels and alteration of gut microbiota [124]. Likewise, Ren et al. (2021) evaluated the effect of Gegen Qinlian Decoction on clinical prognosis and islet function in T2DM. The berberine effects in reducing glucose levels and maintenance of intestinal flora has revealed to be beneficial in T2DM [166]. Similarly, *Huanglian Jiedu* Decoction and its active compounds showed effective against T2DM [126, 127]. Studies have reviewed on beneficial aspects of curcumin and cinnamon on T2DM [167, 168]. Similarly, *Nigella sativa* and *Taraxacum officinale* revealed to show anti-diabetic properties on T2DM [128, 169]. In an open label clinical trial, metformin with Chinese traditional medicines alleviated T2DM along with alterations in gut microbiome [129]. Also, the Chinese traditional medicines effect on gut microflora reduced T2DM [124]. In a meta-analysis study, oral antidiabetic drugs and glucagon-like peptide-1 receptor agonists were assessed in heart failure occurring in T2DM, where the drug liraglutide showed a promising effect in preventing heart failure [130]. In a review, a placebo-controlled outcomes on lessening the glucose levels by glucagon-like peptide 1 receptor agonists, sodium-glucose cotransporter-2 inhibitors, and dipeptidyl peptidase-4 inhibitors were deliberated by Elbelt. (2018) and González-González et al. [132]. Recently, T2DM patients confirmed with COVID-19 infection showed that the usage of metformin and dipeptidyl peptidase-4 inhibitors had minimal contrary outcomes when compared to non-users. In T2DM patients receiving statin, fibrates and its combination showed decrease in lipid levels which shows the necessity in treating diabetes [170]. Therapeutic approaches using incretin and sodium-glucose cotransporter-2 inhibitors have made beneficial effect in renal function in T2DM [134]. Anti-hyperglycaemic drugs showed no significant change in haemoglobin A levels in glycaemic patients [135, p. 202]. In a meta-analysis study, the efficiency of metformin decreased the plasma glucose levels in T2DM patients [136]. The use of statins in T2DM

patients has increased drastically since it has reduced the cardiovascular complications and mortality [133]. The add-on therapy using tofogliflozin to dipeptidyl peptidase-4 inhibitors has shown a reduced glucose level in T2DM [137]. In a review, *Momordica charantia* (bitter melon) has been recommended as an anti-diabetic drug and more studies on its preparation, dosage, and duration were suggested to prove its efficiency in T2DM [138]. *Momordica charantia* has shown drastic reduction in glycaemic levels in T2DM patients [139]. Tolerance and intolerance of metformin was investigated in T2DM patients by measuring the epigenetic markers in blood which showed the effectiveness of epigenetics as a diagnostic tool in diabetes [170]. The extract of chenpi, 5-demethylated polymethoxyflavones (5-OH PMFs) reduced the T2DM as well as obesity with an enhancement in lipid profile [141]. In a meta-analysis study, aloe vera was reported to show an effective therapy in glycaemic control in T2DM and in pre-diabetic condition [142]. In diabetic rats, the effects of berberine were investigated to show a reduced lipid and glucose levels with an activation of adenosine 5'-monophosphate-activated protein kinase pathway [143]. In a meta-analysis, the root of *Pueraria lobata* has evidently showed that T2DM can be treated by increasing insulin levels and reducing glucose absorption [144]. By the use of sodium-glucose cotransporter-2 inhibitors in Japanese T2DM patients, the risk of heart failure and kidney related outcomes was reduced [145]. A comparison study was carried out using sodium-glucose cotransporter-2 inhibitors and glucagon-like peptide-1 receptor agonists in T2DM patients where the sodium-glucose cotransporter-2 inhibitors showed improvement on renal function with or without albuminuria [146]. Similar comparison study showed controlled effects on cardiorenal function in T2DM [147]. In a monotherapy study, metformin was compared with sulfonylureas and meglitinides showed increased risk to cardiovascular complications in T2DM [148]. In a meta-analysis study, acupuncture combined with Chinese herbal treatment showed positive therapeutic openings in T2DM [149]. The treatment using ipragliflozin has shown adverse effects causing genital infections in T2DM patients [150]. In a clinical trial, ginseng showed an efficient action on glucose levels with contradiction in lipid metabolites in T2DM [151]. The Chinese medicines, Shenqi Jiangtang Granules, Baihu Jia Renshen Decoction, and Tianqi Jiangtang Capsule reported as a safe therapy for T2DM [152–154]. In a Bayesian network meta-analysis study, the effective role of Chinese traditional medicine against T2DM was deliberated comparing with western medicines [152]. The cotadutide drug has been a potent drug in treating T2DM which has been effective in decreasing glucose levels, haemoglobin A, and body weight [156]. The Chinese medicine Astragali Radix-Coptis Rhizoma and *Allium sativum* L. was used to treat T2DM where its mechanistic action using network pharmacology was explained in

Table 4 Ongoing clinical trials on anti-diabetic drugs for PD

S. no	Study	Drug	Status	Organisation
1	Exenatide once weekly over 2 years as a potential disease modifying treatment for Parkinson's disease (exenatide-PD3)	Exenatide	Active, not recruiting	University College, London
2	Effects of exenatide on motor function and the brain	Exenatide	Active, not recruiting	University of Florida
3	A clinical study of NLY01 in patients with early Parkinson's disease	NLY01	Active, not recruiting	Neuraly, Inc
4	SR-exenatide (PT320) to evaluate efficacy and safety in patients with early Parkinson's disease	PT320	Active, not recruiting	Peptron, Inc
5	Safety and efficacy of liraglutide in Parkinson's disease	Liraglutide	Active, not recruiting	Cedars-Sinai Medical Centre
6	GLP1R in Parkinson's disease (GIPD)	Semaglutide	Not yet recruiting	Oslo University Hospital
7	Study to evaluate the effect of lixisenatide in patients with Parkinson's disease (LixiPark)	Lixisenatide	Active, not recruiting	University Hospital, Toulouse

a study [171, 172]. The combined effect of Shengmai-Yin and Ganmaidazao decoction (SGD) in T2DM showed declined insulin resistance [157]. In a review, the therapeutic effect of *Cyclocarya paliurus* was explained with mechanistic outcome in treating T2DM [173]. In a randomised clinical study, the Sancai powder showed effective results in T2DM patients in 12 weeks when compared with metformin [141]. In T2DM rats, Zhimu-Huangbai herb pair metabolites and ingredients reported with clinical efficacy in T2DM [158]. In T2DM rats treated with Naoxintong Capsule showed anti-diabetic activity with effective mechanistic strategy [159]. The blend of pioglitazone and canagliflozin treatment showed an improvement from T2DM and metabolites levels [160]. Canagliflozin improved the renal function in T2DM diagnosed patients [161]. The supplementation of pine bark extract was given to T2DM and has control on glycaemic levels and obesity [162]. The effect of Jinlida granules on glycemic levels was reported with a need in improvement in T2DM treatment [163]. From the studies discussed, it is well-known that ground-breaking discoveries have been coming up in treating T2DM. However, the mechanistic insights on previous drugs should be investigated for deliberating clinical assistance to the patients. Previously, it is known that many reviews, systemic reviews, and meta-analysis have been published on T2DM drugs; here, we have deliberated the commonly used western and traditional drugs involved in T2DM with recent outputs in Table 2.

Ongoing Clinical Trials About the Use of T2DM Drugs for PD

The relationship between PD and T2DM is supported by both epidemiological and experimental data. Both diseases are becoming more common worldwide and are strongly linked to ageing. At present, many clinical trials have been applied to repurpose prevailing drugs as well as to address the need for the development of vaccines and drugs against

T2DM associated PD. Currently, the established T2DM drugs undergoing clinical trials has the probability to treat PD (Table 4). Additional research is needed to explore the efficiency of T2DM drugs on PD cure.

Future Perspective and Conclusion

It is now understood that insulin can have a significant impact on how the brain functions. The regulation of neuropathological characteristics of neurodegenerative diseases, such as PD, can be influenced by changes in insulin metabolism and signalling. Numerous in vivo and in vitro studies indicate the connection between T2DM and neurodegeneration and raise the possibility of therapeutic use of several anti-diabetics in the prevention or treatment of conditions like PD. The pathophysiological connections between T2DM and neurodegenerative diseases, as well as how anti-diabetics affect neurodegeneration, are not fully understood, and further studies are necessary to understand these connections and their therapeutic implications.

Acknowledgements The authors would like to thank the Indian Council of Medical Research (ICMR), Department of Health Research—Grant-In-Aid (DHR-GIA) (grant number: GIA/2019/000276/PRCGIA), Department of Zoology, Central University of Punjab, Bathinda—151401, Punjab, Government of India and Bharathiar University, Coimbatore, to carry out this review work.

Author Contribution Balachandar Vellingiri, Mahalaxmi Iyer, and S Sri Sabari designed the review. S Sri Sabari, Dhivya Venkatesan, Kiruthika Balasubramani, Mahalaxmi Iyer, Abilash Valsala Gopalakrishnan, Arul N, Senthil Kumar Nachimuthu, and Balachandar Vellingiri wrote the manuscript. Balachandar Vellingiri conceived and supervised the review.

Funding This work was supported by the Indian Council of Medical Research (ICMR), Department of Health Research—Grant-In-Aid (DHR-GIA) (grant number: GIA/2019/000276/PRCGIA).

Data Availability Data sharing is not applicable to this article as no new data were created or analysed in this study.

Declarations

Ethics Approval Not applicable.

Consent to Participate Not applicable.

Consent for Publication All authors provide consent for publication.

Conflicts of Interest The authors declare no competing interests.

References

- Venkatesan D et al (2022) Genotypic-phenotypic analysis, metabolic profiling and clinical correlations in Parkinson's disease patients from Tamil Nadu population, India. *J Mol Neurosci* 72(8):1724–1737. <https://doi.org/10.1007/s12031-022-02028-4>
- Venkatesan D, Iyer M, Krishnan P, Wilson RS, Vellingiri B (2021) A late-onset Parkinson's disease in tribes in India – a case report. *Brain Disord* 3:100015. <https://doi.org/10.1016/j.dscb.2021.100015>
- Venkatesan D, Iyer M, Wilson R, Lakshmiopathy G, Vellingiri B (2021) The association between multiple risk factors, clinical correlations and molecular insights in Parkinson's disease patients from Tamil Nadu population, India. *Neurosci Lett* 755:135903. <https://doi.org/10.1016/j.neulet.2021.135903>
- M. Iyer et al., "Role of RhoA-ROCK signaling in Parkinson's disease," *European Journal of Pharmacology*, vol. 894, p. 173815, Mar. 2021, <https://doi.org/10.1016/j.ejphar.2020.173815>.
- B. Vellingiri et al (2022) "Influence of heavy metals in Parkinson's disease: an overview," *J Neuro* <https://doi.org/10.1007/s00415-022-11282-w>
- Vellingiri B et al (2022) Role of heavy metals (copper (Cu), arsenic (As), cadmium (Cd), iron (Fe) and lithium (Li)) induced neurotoxicity. *Chemosphere* 301:134625. <https://doi.org/10.1016/j.chemosphere.2022.134625>
- Vellingiri B et al (2022) Neurotoxicity of pesticides – a link to neurodegeneration. *Ecotoxicol Environ Saf* 243:113972. <https://doi.org/10.1016/j.ecoenv.2022.113972>
- Potashkin J, Huang X, Becker C, Chen H, Foltynie T, Marras C (2020) Understanding the links between cardiovascular disease and Parkinson's disease. *Mov Disord* 35(1):55–74. <https://doi.org/10.1002/mds.27836>
- Ejma M et al (2020) The links between Parkinson's disease and cancer. *Biomed* 8(10):416. <https://doi.org/10.3390/biomedicines8100416>
- Fasano A, Visanji NP, Liu LWC, Lang AE, Pfeiffer RF (2015) Gastrointestinal dysfunction in Parkinson's disease. *Lancet Neurol* 14(6):625–639. [https://doi.org/10.1016/S1474-4422\(15\)00007-1](https://doi.org/10.1016/S1474-4422(15)00007-1)
- Hu G, Jousilahti P, Bidet S, Antikainen R, Tuomilehto J (2007) Type 2 diabetes and the risk of Parkinson's disease. *Diabetes Care* 30(4):842–847. <https://doi.org/10.2337/dc06-2011>
- Zhou Z, Zhou R, Zhang Z, Li K (2019) The association between vitamin D status, vitamin D supplementation, sunlight exposure, and Parkinson's disease: a systematic review and meta-analysis. *Med Sci Monit* 25:666–674. <https://doi.org/10.12659/MSM.912840>
- Müller T et al (2013) Peripheral neuropathy in Parkinson's disease: levodopa exposure and implications for duodenal delivery. *Parkinsonism Relat Disord* 19(5):501–507. <https://doi.org/10.1016/j.parkreldis.2013.02.006>
- Kim YE, Lee W, Yun JY, Yang HJ, Kim H-J, Jeon BS (2013) Musculoskeletal problems in Parkinson's disease: neglected issues. *Parkinsonism Relat Disord* 19(7):666–669. <https://doi.org/10.1016/j.parkreldis.2013.03.002>
- Guariguata L, Whiting DR, Hambleton I, Beagley J, Linnenkamp U, Shaw JE (2014) Global estimates of diabetes prevalence for 2013 and projections for 2035. *Diabetes Res Clin Pract* 103(2):137–149. <https://doi.org/10.1016/j.diabres.2013.11.002>
- Pagano G et al (2018) Diabetes mellitus and Parkinson disease. *Neurol* 90(19):e1654–e1662. <https://doi.org/10.1212/WNL.0000000000005475>
- Adeghate E, Schattner P, Dunn E (2006) An update on the etiology and epidemiology of diabetes mellitus. *Ann N Y Acad Sci* 1084(1):1–29. <https://doi.org/10.1196/annals.1372.029>
- Chiu S-L, Cline HT (2010) Insulin receptor signaling in the development of neuronal structure and function. *Neural Dev* 5(1):7. <https://doi.org/10.1186/1749-8104-5-7>
- Biessels GJ, Strachan MWJ, Visseren FLJ, Kappelle LJ, Whitmer RA (2014) Dementia and cognitive decline in type 2 diabetes and prediabetic stages: towards targeted interventions. *Lancet Diabetes Endocrinol* 2(3):246–255. [https://doi.org/10.1016/S2213-8587\(13\)70088-3](https://doi.org/10.1016/S2213-8587(13)70088-3)
- White MF (2003) Insulin signaling in health and disease. *Sci* 302(5651):1710–1711. <https://doi.org/10.1126/science.1092952>
- Obasse I, Taylor M, Fullwood NJ, Allsop D (2017) Development of proteolytically stable N-methylated peptide inhibitors of aggregation of the amylin peptide implicated in type 2 diabetes. *Interface Focus* 7(6):20160127. <https://doi.org/10.1098/rsfs.2016.0127>
- Vijay A, Ranganathan P, Vellingiri B (2019) A survey to validate the traditional Siddha perception of diabetes mellitus. *J Public Health* 27(5):581–590. <https://doi.org/10.1007/s10389-018-0980-y>
- Poewe W et al (2017) Parkinson disease. *Nat Rev Dis Primers* 3(1):17013. <https://doi.org/10.1038/nrdp.2017.13>
- Rodriguez-Araujo G et al (2015) Low alpha-synuclein levels in the blood are associated with insulin resistance. *Sci Rep* 5(1):12081. <https://doi.org/10.1038/srep12081>
- Jaikaran ETAS, Nilsson MR, Clark A (2004) Pancreatic beta-cell granule peptides form heteromolecular complexes which inhibit islet amyloid polypeptide fibril formation. *Biochem J* 377(3):709–716. <https://doi.org/10.1042/bj20030852>
- Oueslati A, Ximerakis M, Vekrellis K (2014) Protein transmission, seeding and degradation: key steps for α -synuclein prion-like propagation. *Exp Neurol* 23(4):324–336. <https://doi.org/10.5607/en.2014.23.4.324>
- Horvath I, Wittung-Stafshede P (2016) Cross-talk between amyloidogenic proteins in type-2 diabetes and Parkinson's disease. *Proc Natl Acad Sci USA* 113(44):12473–12477. <https://doi.org/10.1073/pnas.1610371113>
- Yan MH, Wang X, Zhu X (2013) Mitochondrial defects and oxidative stress in Alzheimer disease and Parkinson disease. *Free Radical Biol Med* 62:90–101. <https://doi.org/10.1016/j.freeradbiomed.2012.11.014>
- Hong C-T et al (2020) Insulin resistance promotes Parkinson's disease through aberrant expression of α -synuclein, mitochondrial dysfunction, and deregulation of the polo-like kinase 2 signaling. *Cells* 9(3):740. <https://doi.org/10.3390/cells9030740>
- Kleinriders A et al (2015) Insulin resistance in brain alters dopamine turnover and causes behavioral disorders. *Proc Natl Acad Sci USA* 112(11):3463–3468. <https://doi.org/10.1073/pnas.1500877112>
- Duarte AI, Santos MS, Seica R, de Oliveira CR (2003) Insulin affects synaptosomal GABA and glutamate transport under oxidative stress conditions. *Brain Res* 977(1):23–30. [https://doi.org/10.1016/S0006-8993\(03\)02679-9](https://doi.org/10.1016/S0006-8993(03)02679-9)

32. Joers V, Tansey MG, Mulas G, Carta AR (2017) Microglial phenotypes in Parkinson's disease and animal models of the disease. *Prog Neurobiol* 155:57–75. <https://doi.org/10.1016/j.pneurobio.2016.04.006>
33. Maldonado-Ruiz R, Montalvo-Martínez L, Fuentes-Mera L, Camacho A (2017) Microglia activation due to obesity programs metabolic failure leading to type two diabetes. *Nutr & Diabetes* 7(3):e254–e254. <https://doi.org/10.1038/nutd.2017.10>
34. Bosco D et al (2012) Dementia is associated with insulin resistance in patients with Parkinson's disease. *J Neurol Sci* 315(1–2):39–43. <https://doi.org/10.1016/j.jns.2011.12.008>
35. Chung SJ et al (2019) Detrimental effect of type 2 diabetes mellitus in a large case series of Parkinson's disease. *Parkinsonism Relat Disord* 64:54–59. <https://doi.org/10.1016/j.parkreldis.2018.08.023>
36. Cereda E, Barichella M, Cassani E, Caccialanza R, Pezzoli G (2012) Clinical features of Parkinson disease when onset of diabetes came first: a case-control study. *Neurol* 78(19):1507–1511. <https://doi.org/10.1212/WNL.0b013e3182553cc9>
37. Cereda E et al (2011) Diabetes and risk of Parkinson's disease. *Diabetes Care* 34(12):2614–2623. <https://doi.org/10.2337/dc11-1584>
38. Driver JA, Smith A, Buring JE, Gaziano JM, Kurth T, Logroschino G (2008) Prospective cohort study of type 2 diabetes and the risk of Parkinson's disease. *Diabetes Care* 31(10):2003–2005. <https://doi.org/10.2337/dc08-0688>
39. De Pablo-Fernandez E, Sierra-Hidalgo F, Benito-León J, Bermejo-Pareja F (2017) Association between Parkinson's disease and diabetes: data from NEDICES study. *Acta Neurol Scand* 136(6):732–736. <https://doi.org/10.1111/ane.12793>
40. De Pablo-Fernandez E, Goldacre R, Pakpoor J, Noyce AJ, Warner TT (2018) Association between diabetes and subsequent Parkinson disease: a record-linkage cohort study. *Neurol* 91(2):e139–e142. <https://doi.org/10.1212/WNL.00000000000005771>
41. Palacios N et al (2011) Obesity, diabetes, and risk of Parkinson's disease: obesity, diabetes, and risk of Parkinson's disease. *Mov Disord* 26(12):2253–2259. <https://doi.org/10.1002/mds.23855>
42. Ong M et al (2017) Influence of diabetes mellitus on longitudinal atrophy and cognition in Parkinson's disease. *J Neurol Sci* 377:122–126. <https://doi.org/10.1016/j.jns.2017.04.010>
43. Sun Y, Chang Y-H, Chen H-F, Su Y-H, Su H-F, Li C-Y (2012) Risk of Parkinson disease onset in patients with diabetes. *Diabetes Care* 35(5):1047–1049. <https://doi.org/10.2337/dc11-1511>
44. Yue X, Li H, Yan H, Zhang P, Chang L, Li T (2016) Risk of Parkinson disease in diabetes mellitus: an updated meta-analysis of population-based cohort studies. *Medicine* 95(18):e3549. <https://doi.org/10.1097/MD.00000000000003549>
45. Lu L, Fu D, Li H, Liu A, Li J, Zheng G (2014) Diabetes and risk of Parkinson's disease: an updated meta-analysis of case-control studies. *PLoS ONE* 9(1):e85781. <https://doi.org/10.1371/journal.pone.0085781>
46. Xu Q et al (2011) Diabetes and risk of Parkinson's disease. *Diabetes Care* 34(4):910–915. <https://doi.org/10.2337/dc10-1922>
47. Marques RG, Fontaine MJ, Rogers J (2004) C-peptide: much more than a byproduct of insulin biosynthesis. *Pancreas* 29(3):231–238. <https://doi.org/10.1097/00006676-200410000-00009>
48. Jiménez JL, Nettleton EJ, Bouchard M, Robinson CV, Dobson CM, Saibil HR (2002) The protofilament structure of insulin amyloid fibrils. *Proc Natl Acad Sci USA* 99(14):9196–9201. <https://doi.org/10.1073/pnas.142459399>
49. Joshi SR, Parikh RM, Das AK (2007) Insulin—history, biochemistry, physiology and pharmacology. *J Assoc Physicians India* 55(Suppl):19–25
50. Banks WA, Kastin AJ (1998) Differential permeability of the blood–brain barrier to two pancreatic peptides: insulin and amylin. *Peptides* 19(5):883–889. [https://doi.org/10.1016/S0196-9781\(98\)00018-7](https://doi.org/10.1016/S0196-9781(98)00018-7)
51. Agrawal R, Reno CM, Sharma S, Christensen C, Huang Y, Fisher SJ (2021) Insulin action in the brain regulates both central and peripheral functions. *Am J Physiol-Endocrinol Metab* 321(1):E156–E163. <https://doi.org/10.1152/ajpendo.00642.2020>
52. Gray SM, Barrett EJ (2018) Insulin transport into the brain. *Am J Physiol Cell Physiol* 315(2):C125–C136. <https://doi.org/10.1152/ajpcell.00240.2017>
53. Apostolatos A et al (2012) Insulin promotes neuronal survival via the alternatively spliced protein kinase CδII isoform. *J Biol Chem* 287(12):9299–9310. <https://doi.org/10.1074/jbc.M111.313080>
54. Reno CM et al (2013) Antecedent glycemic control reduces severe hypoglycemia-induced neuronal damage in diabetic rats. *Am J Physiol-Endocrinol Metab* 304(12):E1331–E1337. <https://doi.org/10.1152/ajpendo.00084.2013>
55. Rensink AAM et al (2004) Insulin inhibits amyloid β-induced cell death in cultured human brain pericytes. *Neurobiol Aging* 25(1):93–103. [https://doi.org/10.1016/S0197-4580\(03\)00039-3](https://doi.org/10.1016/S0197-4580(03)00039-3)
56. Soto M, Cai W, Konishi M, Kahn CR (2019) Insulin signaling in the hippocampus and amygdala regulates metabolism and neurobehavior. *Proc Natl Acad Sci USA* 116(13):6379–6384. <https://doi.org/10.1073/pnas.1817391116>
57. Hölscher C (2019) Insulin signaling impairment in the brain as a risk factor in Alzheimer's disease. *Front Aging Neurosci* 11:88. <https://doi.org/10.3389/fnagi.2019.00088>
58. Kleinridders A, Ferris HA, Cai W, Kahn CR (2014) Insulin action in brain regulates systemic metabolism and brain function. *Diabetes* 63(7):2232–2243. <https://doi.org/10.2337/db14-0568>
59. Stockhorst U, Defries D, Steingrueber H, Scherbaum W (2004) Insulin and the CNS: effects on food intake, memory, and endocrine parameters and the role of intranasal insulin administration in humans. *Physiol Behav* 83(1):47–54. [https://doi.org/10.1016/S0031-9384\(04\)00348-8](https://doi.org/10.1016/S0031-9384(04)00348-8)
60. Yip CC (1992) The insulin-binding domain of insulin receptor is encoded by exon 2 and exon 3. *J Cell Biochem* 48(1):19–25. <https://doi.org/10.1002/jcb.240480105>
61. Gabbouj S et al (2019) Altered insulin signaling in Alzheimer's disease brain – special emphasis on PI3K-Akt pathway. *Front Neurosci* 13:629. <https://doi.org/10.3389/fnins.2019.00629>
62. Kim B, Feldman EL (2012) Insulin resistance in the nervous system. *Trends Endocrinol Metab* 23(3):133–141. <https://doi.org/10.1016/j.tem.2011.12.004>
63. Risso G, Blaustein M, Pozzi B, Mammi P, Srebrow A (2015) Akt/PKB: one kinase, many modifications. *Biochem J* 468(2):203–214. <https://doi.org/10.1042/BJ20150041>
64. Evans RM, Hui S, Perkins A, Lahiri DK, Poirier J, Farlow MR (2004) Cholesterol and APOE genotype interact to influence Alzheimer disease progression. *Neurol* 62(10):1869–1871. <https://doi.org/10.1212/01.WNL.0000125323.15458.3F>
65. Yao W-D et al (2004) Identification of PSD-95 as a regulator of dopamine-mediated synaptic and behavioral plasticity. *Neuron* 41(4):625–638. [https://doi.org/10.1016/S0896-6273\(04\)00048-0](https://doi.org/10.1016/S0896-6273(04)00048-0)
66. Jeanneteau F, Deinhardt K (2011) Fine-tuning MAPK signaling in the brain: the role of MKP-1. *Commun Integr Biol* 4(3):281–283. <https://doi.org/10.4161/cib.4.3.14766>
67. Ahmed N, Thornalley PJ (2007) Advanced glycation end-products: what is their relevance to diabetic complications? *Diabetes Obes Metab* 9(3):233–245. <https://doi.org/10.1111/j.1463-1326.2006.00595.x>
68. Jaganjac M, Tirosh O, Cohen G, Sasson S, Zarkovic N (2013) Reactive aldehydes – second messengers of free radicals in

- diabetes mellitus. *Free Radic Res* 47(sup1):39–48. <https://doi.org/10.3109/10715762.2013.789136>
69. Lu J, Randell E, Han Y, Adeli K, Krahn J, Meng QH (2011) Increased plasma methylglyoxal level, inflammation, and vascular endothelial dysfunction in diabetic nephropathy. *Clin Biochem* 44(4):307–311. <https://doi.org/10.1016/j.clinbiochem.2010.11.004>
 70. McLellan AC, Thornalley PJ, Benn J, Sonksen PH (1994) Glyoxalase system in clinical diabetes mellitus and correlation with diabetic complications. *Clin Sci* 87(1):21–29. <https://doi.org/10.1042/cs0870021>
 71. Rabbani N (2022) Methylglyoxal and glyoxalase 1—a metabolic stress pathway-linking hyperglycemia to the unfolded protein response and vascular complications of diabetes. *Clin Sci* 136(11):819–824. <https://doi.org/10.1042/CS20220099>
 72. Sousa Silva M, Gomes RA, Ferreira AEN, Ponces Freire A, Cordeiro C (2013) The glyoxalase pathway: the first hundred years... and beyond. *Biochem J* 453(1):1–15. <https://doi.org/10.1042/BJ20121743>
 73. Thornberry NA, Gallwitz B (2009) Mechanism of action of inhibitors of dipeptidyl-peptidase-4 (DPP-4). *Best Pract Res Clin Endocrinol Metab* 23(4):479–486. <https://doi.org/10.1016/j.beem.2009.03.004>
 74. Kazachkov M, Yu P (2005) A novel HPLC procedure for detection and quantification of aminoacetone, a precursor of methylglyoxal, in biological samples. *J Chromatogr B* 824(1–2):116–122. <https://doi.org/10.1016/j.jchromb.2005.07.006>
 75. Reichard GA, Skutches CL, Hoeldtke RD, Owen OE (1986) Acetone metabolism in humans during diabetic ketoacidosis. *Diabetes* 35(6):668–674. <https://doi.org/10.2337/diab.35.6.668>
 76. Nigro C et al (2019) Dicarbonyl stress at the crossroads of healthy and unhealthy aging. *Cells* 8(7):749. <https://doi.org/10.3390/cells8070749>
 77. Rabbani N, Thornalley PJ (2018) Advanced glycation end products in the pathogenesis of chronic kidney disease. *Kidney Int* 93(4):803–813. <https://doi.org/10.1016/j.kint.2017.11.034>
 78. Thornalley PJ (2005) Dicarbonyl intermediates in the Maillard reaction. *Ann N Y Acad Sci* 1043(1):111–117. <https://doi.org/10.1196/annals.1333.014>
 79. I. Allaman, M. BÄölander, and P. J. Magistretti (2015) “Methylglyoxal, the dark side of glycolysis,” *Front. Neurosci.*, 9, <https://doi.org/10.3389/fnins.2015.00023>.
 80. Gilbert RP, Brandt RB (1975) Spectrophotometric determination of methyl glyoxal with 2,4-dinitrophenylhydrazine. *Anal Chem* 47(14):2418–2422. <https://doi.org/10.1021/ac60364a003>
 81. Aviles-Olmos I, Limousin P, Lees A, Foltynie T (2013) Parkinson’s disease, insulin resistance and novel agents of neuroprotection. *Brain* 136(2):374–384. <https://doi.org/10.1093/brain/aww009>
 82. Thornalley PJ, Langborg A, Minhas HS (1999) Formation of glyoxal, methylglyoxal and 3-deoxyglucosone in the glycation of proteins by glucose. *Biochem J* 344(1):109–116. <https://doi.org/10.1042/bj3440109>
 83. Wan OW, Chung KKK (2012) The role of alpha-synuclein oligomerization and aggregation in cellular and animal models of Parkinson’s disease. *PLoS ONE* 7(6):e38545. <https://doi.org/10.1371/journal.pone.0038545>
 84. Kurz A et al (2011) Alpha-synuclein deficiency leads to increased glyoxalase I expression and glycation stress. *Cell Mol Life Sci* 68(4):721–733. <https://doi.org/10.1007/s00018-010-0483-7>
 85. Zhang C et al (2011) Parkin, a p53 target gene, mediates the role of p53 in glucose metabolism and the Warburg effect. *Proc Natl Acad Sci USA* 108(39):16259–16264. <https://doi.org/10.1073/pnas.1113884108>
 86. Ferrer I, Martinez A, Blanco R, Dalfó E, Carmona M (2011) Neuropathology of sporadic Parkinson disease before the appearance of parkinsonism: preclinical Parkinson disease. *J Neural Transm* 118(5):821–839. <https://doi.org/10.1007/s00702-010-0482-8>
 87. Gómez A, Ferrer I (2009) Increased oxidation of certain glycolysis and energy metabolism enzymes in the frontal cortex in Lewy body diseases. *J Neurosci Res* 87(4):1002–1013. <https://doi.org/10.1002/jnr.21904>
 88. Sandyk R (1993) The relationship between diabetes mellitus and Parkinson’s disease. *Int J Neurosci* 69(1–4):125–130. <https://doi.org/10.3109/00207459309003322>
 89. Marques A et al (2018) Glucose dysregulation in Parkinson’s disease: too much glucose or not enough insulin? *Parkinsonism Relat Disord* 55:122–127. <https://doi.org/10.1016/j.parkreldis.2018.05.026>
 90. Athauda D, Foltynie T (2016) Insulin resistance and Parkinson’s disease: a new target for disease modification? *Prog Neurobiol* 145–146:98–120. <https://doi.org/10.1016/j.pneurobio.2016.10.001>
 91. Unger J, Livingston J, Moss A (1991) Insulin receptors in the central nervous system: localization, signalling mechanisms and functional aspects. *Prog Neurobiol* 36(5):343–362. [https://doi.org/10.1016/0301-0082\(91\)90015-S](https://doi.org/10.1016/0301-0082(91)90015-S)
 92. Baskin DG, Porte D, Guest K, Dorsa DM (1983) Regional concentrations of insulin in the rat brain*. *Endocrinol* 112(3):898–903. <https://doi.org/10.1210/endo-112-3-898>
 93. Kuwabara T et al (2011) Insulin biosynthesis in neuronal progenitors derived from adult hippocampus and the olfactory bulb. *EMBO Mol Med* 3(12):742–754. <https://doi.org/10.1002/emmm.201100177>
 94. Duarte AI, Moreira PI, Oliveira CR (2012) Insulin in central nervous system: more than just a peripheral hormone. *J Aging Res* 2012:1–21. <https://doi.org/10.1155/2012/384017>
 95. Morris JK et al (2014) Insulin resistance and gray matter volume in neurodegenerative disease. *Neurosci* 270:139–147. <https://doi.org/10.1016/j.neuroscience.2014.04.006>
 96. Tong M, Dong M, de la Monte SM (2009) Brain insulin-like growth factor and neurotrophin resistance in Parkinson’s disease and dementia with Lewy bodies: potential role of manganese neurotoxicity. *JAD* 16(3):585–599. <https://doi.org/10.3233/JAD-2009-0995>
 97. Kawahito S, Kitahata H, Oshita S (2009) Problems associated with glucose toxicity: role of hyperglycemia-induced oxidative stress. *WJG* 15(33):4137. <https://doi.org/10.3748/wjg.15.4137>
 98. Eriksson JW (2007) Metabolic stress in insulin’s target cells leads to ROS accumulation - a hypothetical common pathway causing insulin resistance. *FEBS Lett* 581(19):3734–3742. <https://doi.org/10.1016/j.febslet.2007.06.044>
 99. Brownlee M (2005) The pathobiology of diabetic complications. *Diabetes* 54(6):1615–1625. <https://doi.org/10.2337/diabetes.54.6.1615>
 100. Jenner P, Olanow CW (2006) The pathogenesis of cell death in Parkinson’s disease. *Neurol* 66(10):24–36. https://doi.org/10.1212/WNL.66.10_suppl_4.S24
 101. Liu X, Yamada N, Maruyama W, Osawa T (2008) Formation of dopamine adducts derived from brain polyunsaturated fatty acids. *J Biol Chem* 283(50):34887–34895. <https://doi.org/10.1074/jbc.M805682200>
 102. Fan H et al (2015) Methionine sulfoxide reductase A negatively controls microglia-mediated neuroinflammation via inhibiting ROS/MAPKs/NF-κB signaling pathways through a catalytic antioxidant function. *Antioxid Redox Signal* 22(10):832–847. <https://doi.org/10.1089/ars.2014.6022>
 103. Zhang J, Zheng Y, Luo Y, Du Y, Zhang X, Fu J (2019) Curcumin inhibits LPS-induced neuroinflammation by promoting microglial M2 polarization via TREM2/TLR4/NF-κB pathways in BV2 cells. *Mol Immunol* 116:29–37. <https://doi.org/10.1016/j.molimm.2019.09.020>

104. Jayaraj RL, Azimullah S, Beiram R, Jalal FY, Rosenberg GA (2019) Neuroinflammation: friend and foe for ischemic stroke. *J Neuroinflammation* 16(1):142. <https://doi.org/10.1186/s12974-019-1516-2>
105. Bhuvanendran S, Bakar SNS, Kumari Y, Othman I, Mohd FS, Hassan Z (2019) Embelin improves the spatial memory and hippocampal long-term potentiation in a rat model of chronic cerebral hypoperfusion. *Sci Rep* 9(1):14507. <https://doi.org/10.1038/s41598-019-50954-y>
106. Rajchgot T et al (2019) Neurons and microglia; a sickly-sweet duo in diabetic pain neuropathy. *Front Neurosci* 13:25. <https://doi.org/10.3389/fnins.2019.00025>
107. Liu M, Gao L, Zhang N (2019) Berberine reduces neuroglia activation and inflammation in streptozotocin-induced diabetic mice. *Int J Immunopathol Pharmacol* 33:205873841986637. <https://doi.org/10.1177/2058738419866379>
108. Sivitz WI, Yorek MA (2010) Mitochondrial dysfunction in diabetes: from molecular mechanisms to functional significance and therapeutic opportunities. *Antioxid Redox Signal* 12(4):537–577. <https://doi.org/10.1089/ars.2009.2531>
109. Ji R-R, Suter MR (2007) p38 MAPK, microglial signaling, and neuropathic pain. *Mol Pain* 3:1744–8069–3–33. <https://doi.org/10.1186/1744-8069-3-33>
110. Dukic-Stefanovic S, Gasic-Milenkovic J, Deuther-Conrad W, Münch G (2003) Signal transduction pathways in mouse microglia N-11 cells activated by advanced glycation end-products (AGEs): AGE signalling pathways in N-11 cells. *J Neurochem* 87(1):44–55. <https://doi.org/10.1046/j.1471-4159.2003.01988.x>
111. Imamura K, Hishikawa N, Sawada M, Nagatsu T, Yoshida M, Hashizume Y (2003) Distribution of major histocompatibility complex class II-positive microglia and cytokine profile of Parkinson's disease brains. *Acta Neuropathol* 106(6):518–526. <https://doi.org/10.1007/s00401-003-0766-2>
112. McGeer PL, Itagaki S, Boyes BE, McGeer EG (1988) Reactive microglia are positive for HLA-DR in the substantia nigra of Parkinson's and Alzheimer's disease brains. *Neurology* 38(8):1285–1285. <https://doi.org/10.1212/WNL.38.8.1285>
113. Waterman C et al (2020) Moringa isothiocyanate-rich seed extract delays the onset of diabetes in UC Davis type-2 diabetes mellitus rats. *Sci Rep* 10(1):8861. <https://doi.org/10.1038/s41598-020-65722-6>
114. Cane HPCA et al (2020) Macrophylloflavone: a new biflavonoid from *Garcinia macrophylla* mart. (Clusiaceae) for antibacterial, antioxidant, and anti-type 2 diabetes mellitus activities. *Sci World J* 2020:1–14. <https://doi.org/10.1155/2020/2983129>
115. DiK B, Coşkun D, BahçiVan E, Üney K (2021) Potential antidiabetic activity of benzimidazole derivative albendazole and lansoprazole drugs in different doses in experimental type 2 diabetic rats. *Turk J Med Sci* 51(3):1578–1585. <https://doi.org/10.3906/sag-2004-38>
116. Dharmalingam M et al (2020) Efficacy and safety of remogliflozin etabonate, a new sodium glucose co-transporter-2 inhibitor, in patients with type 2 diabetes mellitus: a 24-week, randomized, double-blind, active-controlled trial. *Drugs* 80(6):587–600. <https://doi.org/10.1007/s40265-020-01285-0>
117. Guo L et al (2021) Comparison of clinical efficacy and safety of metformin sustained-release tablet (II) (Dulening) and metformin tablet (Glucophage) in treatment of type 2 diabetes mellitus. *Front Endocrinol* 12:712200. <https://doi.org/10.3389/fendo.2021.712200>
118. Shankar A (2021) An efficacy and safety study of remogliflozin in obese Indian type 2 diabetes mellitus patients who were inadequately controlled on insulin glargine plus other oral hypoglycemic agents. *CDR* 17(7):e122120189341. <https://doi.org/10.2174/1573399817666201222102520>
119. An W et al (2021) Mechanisms of Rhizoma Coptidis against type 2 diabetes mellitus explored by network pharmacology combined with molecular docking and experimental validation. *Sci Rep* 11(1):20849. <https://doi.org/10.1038/s41598-021-00293-8>
120. Sun Y et al (2020) Active ingredients and mechanism of action of Rhizoma Coptidis against type 2 diabetes based on network-pharmacology and bioinformatics. *Curr Med Sci* 40(2):257–264. <https://doi.org/10.1007/s11596-020-2182-4>
121. Morales Ramos JG, Esteves Pairazamán AT, Mocarro Willis MES, Collantes Santisteban S, Caldas Herrera E (2021) Medicinal properties of *Morus alba* for the control of type 2 diabetes mellitus: a systematic review. *f1000Res* 10:1022. <https://doi.org/10.12688/f1000research.55573.1>
122. Nganou-Gnindjio CN, Ngati Nyonga D, Wafeu GS, Nga EN, Sobngwi E (2022) Cardiometabolic effects of ZingiberOfficinale Roscoe extracts in type 2 diabetic Cameroonian patients after six weeks of add-on therapy : a single clinical-arm trial. *Annales de Cardiologie et d'Angéiologie* 71(3):160–165. <https://doi.org/10.1016/j.ancard.2021.09.010>
123. Terauchi Y, Nakama T, Spranger R, Amano A, Inoue T, Niemoeller E (2020) Efficacy and safety of insulin glargine/lixisenatide fixed-ratio combination (iGLARLIXI 1:1) in Japanese patients with type 2 diabetes mellitus inadequately controlled on oral antidiabetic drugs: a randomized, 26-week, open-label, multicentre study: the LIXILAN JP-O2 randomized clinical trial. *Diabetes Obes Metab* 22(S4):14–23. <https://doi.org/10.1111/dom.14036>
124. Xu X et al (2020) Antidiabetic effects of Gegen Qinlian decoction via the gut microbiota are attributable to its key ingredient berberine. *Genomics Proteomics Bioinformatics* 18(6):721–736. <https://doi.org/10.1016/j.gpb.2019.09.007>
125. Ren L, Cheng Y, Qin F (2020) Herbal formula Gegen-Qinlian decoction for type 2 diabetes mellitus: a meta-analysis of randomized controlled trials. *Evid-Based Complement Altern Med* 2020:1–11. <https://doi.org/10.1155/2020/3907920>
126. Yin B, Bi Y-M, Fan G-J, Xia Y-Q (2020) Molecular mechanism of the effect of Huanglian Jiedu decoction on type 2 diabetes mellitus based on network pharmacology and molecular docking. *J Diabetes Res* 2020:1–24. <https://doi.org/10.1155/2020/5273914>
127. Pan L, Li Z, Wang Y, Zhang B, Liu G, Liu J (2020) Network pharmacology and metabolomics study on the intervention of traditional Chinese medicine Huanglian decoction in rats with type 2 diabetes mellitus. *J Ethnopharmacol* 258:112842. <https://doi.org/10.1016/j.jep.2020.112842>
128. Hamdan A, Haji Idrus R, Mokhtar MH (2019) Effects of Nigella sativa on type-2 diabetes mellitus: a systematic review. *IJERPH* 16(24):4911. <https://doi.org/10.3390/ijerph16244911>
129. Tong X et al (2018) Structural alteration of gut microbiota during the amelioration of human type 2 diabetes with hyperlipidemia by metformin and a traditional Chinese herbal formula: a multicenter, randomized, open label clinical trial. *mBio* 9(3):02392–17. <https://doi.org/10.1128/mBio.02392-17>
130. Ida S et al (2021) Effects of oral antidiabetic drugs and glucagon-like peptide-1 receptor agonists on left ventricular diastolic function in patients with type 2 diabetes mellitus: a systematic review and network meta-analysis. *Heart Fail Rev* 26(5):1151–1158. <https://doi.org/10.1007/s10741-020-09936-w>
131. Doumas M, Imprialos K, Stavropoulos K, Reklou A, Sachinidis A, Athyros VG (2018) Combination of SGLT-2 inhibitors and GLP-1 receptor agonists: potential benefits in surrogate and hard endpoints. *CPD* 24(17):1879–1886. <https://doi.org/10.2174/1381612824666180604113653>
132. Luk AOY et al (2021) Glucose-lowering drugs and outcome from COVID-19 among patients with type 2 diabetes mellitus: a population-wide analysis in Hong Kong. *BMJ Open* 11(10):052310. <https://doi.org/10.1136/bmjopen-2021-052310>

133. Machado-Duque ME, Garcia DA, Emura-Vélez MH, Gaviria-Mendoza A, Machado-Alba JE (2021) Prevalence of the use of aspirin and statins for preventing cardiovascular events in the Colombian population with type 2 diabetes mellitus: comparison of 2008 and 2018. *J Prim Care Community Health* 12:215013272110070. <https://doi.org/10.1177/21501327211007015>
134. Mann JFE, Muskiet MHA (2021) Incretin-based drugs and the kidney in type 2 diabetes: choosing between DPP-4 inhibitors and GLP-1 receptor agonists. *Kidney Int* 99(2):314–318. <https://doi.org/10.1016/j.kint.2020.08.036>
135. Xiang AS et al (2021) Trends in glycaemic control and drug use in males and females with type 2 diabetes: Results of the AUSTRALIAN NATIONAL DIABETES AUDIT from 2013 to 2019. *Diabetes Obes Metab* 23(12):2603–2613. <https://doi.org/10.1111/dom.14506>
136. Piera-Mardemootoo C, Lambert P, Faillie J-L (2021) Efficacy of metformin on glycemic control and weight in drug-naïve type 2 diabetes mellitus patients: a systematic review and meta-analysis of placebo-controlled randomized trials. *Therapies* 76(6):647–656. <https://doi.org/10.1016/j.therap.2018.01.006>
137. T. Iijima et al (2021) “Acute effect of add-on therapy with tofogliflozin, a sodium glucose co-transporter 2 inhibitor, on 24-hours glucose profile and glycaemic variability evaluated by continuous glucose monitoring in patients with type 2 diabetes receiving dipeptidyl peptidase-4 inhibitors,” *Int J Clin Pract*, 75(11), <https://doi.org/10.1111/ijcp.14732>.
138. Habicht S, Ludwig C, Yang R, Krawinkel M (2014) Momordica charantia and type 2 diabetes: from in vitro to human studies. *CDR* 10(1):48–60. <https://doi.org/10.2174/1573399809666131126152044>
139. Rauniyar GP, Sinha R, Chapagain K, Maskey R, Pandey DR (2021) Effects of Momordica charantia (karela/bitterguard) in type 2 diabetic patients taking allopathic drugs: a pilot study. *Kathmandu Univ Med J (KUMJ)* 19(74):143–147
140. García-Calzón S et al (2020) Epigenetic markers associated with metformin response and intolerance in drug-naïve patients with type 2 diabetes. *Sci. Transl. Med.* 12(561):eaaz1803. <https://doi.org/10.1126/scitranslmed.aaz1803>
141. Guo J, Tao H, Cao Y, Ho C-T, Jin S, Huang Q (2016) Prevention of obesity and type 2 diabetes with aged citrus peel (*Chenpi*) extract. *J Agric Food Chem* 64(10):2053–2061. <https://doi.org/10.1021/acs.jafc.5b06157>
142. Suksomboon N, Poolsup N, Punthanitisarn S (2016) Effect of *aloe vera* on glycaemic control in prediabetes and type 2 diabetes: a systematic review and meta-analysis. *J Clin Pharm Ther* 41(2):180–188. <https://doi.org/10.1111/jcpt.12382>
143. Dong Y et al (2016) Metabolomics study of type 2 diabetes mellitus and the antidiabetic effect of berberine in Zucker diabetic fatty rats using Uplc-ESI-Hdms: metabolomics of T2DM and anti-diabetic effect of berberine in ZDF rats. *Phytother Res* 30(5):823–828. <https://doi.org/10.1002/ptr.5587>
144. Yang L et al (2019) *Pueraria lobata* for diabetes mellitus: past, present and future. *Am J Chin Med* 47(07):1419–1444. <https://doi.org/10.1142/S0192415X19500733>
145. Komuro I, Kadowaki T, Bodegård J, Thuresson M, Okami S, Yajima T (2021) Lower heart failure and chronic kidney disease risks associated with sodium-glucose cotransporter-2 inhibitor use in Japanese type 2 diabetes patients without established cardiovascular and renal diseases. *Diabetes Obes Metab* 23(S2):19–27. <https://doi.org/10.1111/dom.14119>
146. Kawai Y et al (2022) Comparison of effects of SGLT-2 inhibitors and GLP-1 receptor agonists on cardiovascular and renal outcomes in type 2 diabetes mellitus patients with/without albuminuria: a systematic review and network meta-analysis. *Diabetes Res Clin Pract* 183:109146. <https://doi.org/10.1016/j.diabres.2021.109146>
147. Wei X-B, Wei W, Ding L-L, Liu S-Y (2021) Comparison of the effects of 10 GLP-1 RA and SGLT2 inhibitor interventions on cardiovascular, mortality, and kidney outcomes in type 2 diabetes: a network meta-analysis of large randomized trials. *Prim Care Diabetes* 15(2):208–211. <https://doi.org/10.1016/j.pcd.2020.08.017>
148. Herrera Comoglio R, Vidal Guitart X (2021) Cardiovascular events and mortality among type 2 diabetes mellitus patients newly prescribed first-line blood glucose-lowering drugs monotherapies: a population-based cohort study in the Catalan electronic medical record database, SIDIAP, 2010–2015. *Primary Care Diabetes* 15(2):323–331. <https://doi.org/10.1016/j.pcd.2020.11.002>
149. Bao P et al (2021) Efficacy and safety of acupuncture combined with Chinese herbal medicine in the treatment of type 2 diabetes mellitus: a protocol for a systematic review and meta-analysis. *Medicine* 100(43):e27658. <https://doi.org/10.1097/MD.00000000000027658>
150. Liu D, Chen H, Song F, Ahmed MA, Wu H (2020) Adverse drug events observed with the novel sodium/glucose co-transporter 2 inhibitor ipragliflozin for the treatment of patients with type 2 diabetes mellitus: a systematic review and meta-analysis of randomized studies. *Adv Ther* 37(10):4356–4369. <https://doi.org/10.1007/s12325-020-01471-2>
151. Gui Q, Xu Z, Xu K, Yang Y (2016) The efficacy of ginseng-related therapies in type 2 diabetes mellitus: an updated systematic review and meta-analysis. *Medicine* 95(6):e2584. <https://doi.org/10.1097/MD.0000000000002584>
152. Li T et al (2021) Efficacy and safety of Shenqi Jiangtang granules plus oral hypoglycemic agent in patients with type 2 diabetes mellitus: a systematic review and meta-analysis of 15 RCTs. *Medicine* 100(5):e23578. <https://doi.org/10.1097/MD.00000000000023578>
153. Tian Y et al (2020) Baihu Jia Renshen decoction for type 2 diabetic mellitus: a protocol for systematic review and meta-analysis. *Medicine* 99(19):20210. <https://doi.org/10.1097/MD.00000000000020210>
154. Piao C et al (2020) Treatment of type 2 diabetes with Tianqi Jiangtang capsule: a systematic review and meta-analysis of randomized controlled trials. *Medicine* 99(21):e19702. <https://doi.org/10.1097/MD.00000000000019702>
155. Jingjing L, Hongmei Z, Yanping L, Bin W (2016) Meta-analysis of the efficacy in treatment of primary sjögren’s syndrome: traditional Chinese medicine vs Western medicine. *J Tradit Chin Med* 36(5):596–605. [https://doi.org/10.1016/S0254-6272\(16\)30078-4](https://doi.org/10.1016/S0254-6272(16)30078-4)
156. Ali MM et al (2022) Impact of Cotadutide drug on patients with type 2 diabetes mellitus: a systematic review and meta-analysis. *BMC Endocr Disord* 22(1):113. <https://doi.org/10.1186/s12902-022-01031-5>
157. Li S et al (2019) Exploring the protective effect of ShengMai-Yin and Ganmaidazao decoction combination against type 2 diabetes mellitus with nonalcoholic fatty liver disease by network pharmacology and validation in KKAY mice. *J Ethnopharmacol* 242:112029. <https://doi.org/10.1016/j.jep.2019.112029>
158. Cao Y, Sun Z, Huang H, Lin A, Liu Y (2022) Comparative analysis of absorbed ingredients and metabolites, and pharmacokinetic studies of Zhimu-Huangbai herb pair in the plasma of normal and type 2 diabetes mellitus rats by UHPLC-linear trap quadrupole-orbitrap MS and LC-MS/MS. *J Sep Sci* 45(3):664–676. <https://doi.org/10.1002/jssc.202100582>
159. Yan Z et al (2020) Integrated metabolomics and gut microbiome to the effects and mechanisms of naoxintong capsule on type 2 diabetes in rats. *Sci Rep* 10(1):10829. <https://doi.org/10.1038/s41598-020-67362-2>
160. E. Kutoh, A. N. Kuto, A. Wada, R. Kurihara, and R. Kojima (2021) “Complementary effects on glycaemic and non-glycaemic

- parameters between responders and non-responders treated with pioglitazone and canagliflozin in drug-naïve subjects with type 2 diabetes,” *Int J Clinical Practice*, 75(12), <https://doi.org/10.1111/ijcp.14914>.
161. Zhou S et al (2021) Canagliflozin could improve the levels of renal oxygenation in newly diagnosed type 2 diabetes patients with normal renal function. *Diabetes Metab* 47(5):101274. <https://doi.org/10.1016/j.diabet.2021.101274>
 162. Navval-Esfahlan E, Raftar M, Asghari S, Imani H, Asghari-Jafarabadi M, Karimi-Avval S (2021) Effect of French maritime pine bark extract supplementation on metabolic status and serum vascular cell adhesion molecule-1 levels in patients with type 2 diabetes and microalbuminuria. *Complement Ther Med* 58:102689. <https://doi.org/10.1016/j.ctim.2021.102689>
 163. Pan J et al (2021) The effectiveness of traditional Chinese medicine Jinlida granules on glycemic variability in newly diagnosed type 2 diabetes: a double-blinded, randomized trial. *J Diabetes Res* 2021:1–8. <https://doi.org/10.1155/2021/6303063>
 164. M. Guo et al (2014) “Metformin may produce antidepressant effects through improvement of cognitive function among depressed patients with diabetes mellitus,” *Clin Exp Pharmacol Physiol*, <https://doi.org/10.1111/1440-1681.12265>.
 165. Sgrò P, Emerenziani GP, Antinozzi C, Sacchetti M, Di Luigi L (2021) Exercise as a drug for glucose management and prevention in type 2 diabetes mellitus. *Curr Opin Pharmacol* 59:95–102. <https://doi.org/10.1016/j.coph.2021.05.006>
 166. Li C, He J-Z, Zhou X-D, Xu X (2017) Berberine regulates type 2 diabetes mellitus related with insulin resistance. *Zhongguo Zhong Yao Za Zhi* 42(12):2254–2260. <https://doi.org/10.19540/j.cnki.cjmm.20170307.014>
 167. Silva ML, Bernardo MA, Singh J, de Mesquita MF (2022) Cinnamon as a complementary therapeutic approach for dysglycemia and dyslipidemia control in type 2 diabetes mellitus and its molecular mechanism of action: a review. *Nutrients* 14(13):2773. <https://doi.org/10.3390/nu14132773>
 168. Pivari F, Mingione A, Brasacchio C, Soldati L (2019) Curcumin and type 2 diabetes mellitus: prevention and treatment. *Nutrients* 11(8):1837. <https://doi.org/10.3390/nu11081837>
 169. Wirngo FE, Lambert MN, Jeppesen PB (2016) The physiological effects of dandelion (*Taraxacum Officinale*) in type 2 diabetes. *Rev Diabet Stud* 13(2–3):113–131. <https://doi.org/10.1900/RDS.2016.13.113>
 170. García-Ulloa AC et al (2021) Clinician prescription of lipid-lowering drugs and achievement of treatment goals in patients with newly diagnosed type 2 diabetes mellitus. *BMJ Open Diab Res Care* 9(1):e001891. <https://doi.org/10.1136/bmjdr-2020-001891>
 171. Fu J-Y, Zhang Y (2021) Mechanism of Astragali Radix-Coptis Rhizoma pair in treating type 2 diabetes mellitus based on network pharmacology. *Zhongguo Zhong Yao Za Zhi* 46(18):4808–4815. <https://doi.org/10.19540/j.cnki.cjmm.20210609.705>
 172. K. K. Oh (2022) “A network pharmacology study to investigate bioactive compounds and signaling pathways of garlic (*Allium sativum* L.) husk against type 2 diabetes mellitus,” *Journal of Food Biochemistry*, 46(7), <https://doi.org/10.1111/jfbc.14106>.
 173. Wang H et al (2021) Potential role of natural plant medicine *Cyclocarya paliurus* in the treatment of type 2 diabetes mellitus. *J Diabetes Res* 2021:1–12. <https://doi.org/10.1155/2021/1655336>
 174. Bailey CJ (2017) Metformin: historical overview. *Diabetologia* 60(9):1566–1576. <https://doi.org/10.1007/s00125-017-4318-z>
 175. Hawley SA et al (2010) Use of cells expressing γ subunit variants to identify diverse mechanisms of AMPK activation. *Cell Metab* 11(6):554–565. <https://doi.org/10.1016/j.cmet.2010.04.001>
 176. Hardie DG (2003) Minireview: The AMP-activated protein kinase cascade: the key sensor of cellular energy status. *Endocrinology* 144(12):5179–5183. <https://doi.org/10.1210/en.2003-0982>
 177. Fullerton MD et al (2013) Single phosphorylation sites in Acc1 and Acc2 regulate lipid homeostasis and the insulin-sensitizing effects of metformin. *Nat Med* 19(12):1649–1654. <https://doi.org/10.1038/nm.3372>
 178. Foretz M et al (2010) Metformin inhibits hepatic gluconeogenesis in mice independently of the LKB1/AMPK pathway via a decrease in hepatic energy state. *J Clin Invest* 120(7):2355–2369. <https://doi.org/10.1172/JCI40671>
 179. He L et al (2009) Metformin and insulin suppress hepatic gluconeogenesis through phosphorylation of CREB binding protein. *Cell* 137(4):635–646. <https://doi.org/10.1016/j.cell.2009.03.016>
 180. Vincent MF, Marangos PJ, Gruber HE, van den Berghe G (1991) Inhibition by AICA riboside of gluconeogenesis in isolated rat hepatocytes. *Diabetes* 40(10):1259–1266. <https://doi.org/10.2337/diab.40.10.1259>
 181. Curry DW, Stutz B, Andrews ZB, Elsworth JD (2018) Targeting AMPK signaling as a neuroprotective strategy in Parkinson’s disease. *JPD* 8(2):161–181. <https://doi.org/10.3233/JPD-171296>
 182. Hang L et al (2021) Conditional disruption of AMP kinase in dopaminergic neurons promotes Parkinson’s disease-associated phenotypes in vivo. *Neurobiol Dis* 161:105560. <https://doi.org/10.1016/j.nbd.2021.105560>
 183. Batandier C et al (2006) The ROS production induced by a reverse-electron flux at respiratory-chain complex 1 is hampered by metformin. *J Bioenerg Biomembr* 38(1):33–42. <https://doi.org/10.1007/s10863-006-9003-8>
 184. Patil SP, Jain PD, Ghumatkar PJ, Tambe R, Sathaye S (2014) Neuroprotective effect of metformin in MPTP-induced Parkinson’s disease in mice. *Neuroscience* 277:747–754. <https://doi.org/10.1016/j.neuroscience.2014.07.046>
 185. Rotermund C, Machetanz G, Fitzgerald JC (2018) The therapeutic potential of metformin in neurodegenerative diseases. *Front Endocrinol* 9:400. <https://doi.org/10.3389/fendo.2018.00400>
 186. Dias V, Junn E, Mouradian MM (2013) The role of oxidative stress in Parkinson’s disease. *J Parkinson’s Dis* 3(4):461–491. <https://doi.org/10.3233/JPD-130230>
 187. Diehn M et al (2009) Association of reactive oxygen species levels and radioresistance in cancer stem cells. *Nature* 458(7239):780–783. <https://doi.org/10.1038/nature07733>
 188. Zhang Q-Q, Li W-S, Liu Z, Zhang H-L, Ba Y-G, Zhang R-X (2020) Metformin therapy and cognitive dysfunction in patients with type 2 diabetes: a meta-analysis and systematic review. *Medicine* 99(10):e19378. <https://doi.org/10.1097/MD.00000000000019378>
 189. Oda SS (2017) Metformin protects against experimental acrylamide neuropathy in rats: metformin and acrylamide neuropathy. *Drug Dev Res* 78(7):349–359. <https://doi.org/10.1002/ddr.21400>
 190. Pérez-Revuelta BI et al (2014) Metformin lowers Ser-129 phosphorylated α -synuclein levels via mTOR-dependent protein phosphatase 2A activation. *Cell Death Dis* 5(5):e1209–e1209. <https://doi.org/10.1038/cddis.2014.175>
 191. Nauck MA (2004) Glucagon-like peptide 1 (GLP-1) in the treatment of diabetes. *Horm Metab Res* 36(11/12):852–858. <https://doi.org/10.1055/s-2004-826175>
 192. Holst JJ (2007) The physiology of glucagon-like peptide 1. *Physiol Rev* 87(4):1409–1439. <https://doi.org/10.1152/physrev.00034.2006>
 193. Orskov C, Holst JJ, Nielsen OV (1988) Effect of truncated glucagon-like peptide-1 [proglucagon-(78–107) amide] on endocrine secretion from pig pancreas, antrum, and nonantral stomach. *Endocrinology* 123(4):2009–2013. <https://doi.org/10.1210/endo-123-4-2009>

194. Rask E et al (2004) Insulin secretion and incretin hormones after oral glucose in non-obese subjects with impaired glucose tolerance. *Metabolism* 53(5):624–631. <https://doi.org/10.1016/j.metabol.2003.11.011>
195. Toft-Nielsen MB et al (2001) Determinants of the impaired secretion of glucagon-like peptide-1 in type 2 diabetic patients. *J Clin Endocrinol Metab* 86(8):3717–3723. <https://doi.org/10.1210/jcem.86.8.7750>
196. Cork SC, Richards JE, Holt MK, Gribble FM, Reimann F, Trapp S (2015) Distribution and characterisation of glucagon-like peptide-1 receptor expressing cells in the mouse brain. *Mol Metab* 4(10):718–731. <https://doi.org/10.1016/j.molmet.2015.07.008>
197. Drucker DJ, Sherman SI, Gorelick FS, Bergenstal RM, Sherwin RS, Buse JB (2010) Incretin-based therapies for the treatment of type 2 diabetes: evaluation of the risks and benefits. *Diabetes Care* 33(2):428–433. <https://doi.org/10.2337/dc09-1499>
198. Liu X-Y, Wang L-X, Chen Z, Liu L-B (2016) Liraglutide prevents beta-amyloid-induced neurotoxicity in SH-SY5Y cells via a PI3K-dependent signaling pathway. *Neurol Res* 38(4):313–319. <https://doi.org/10.1080/01616412.2016.1145914>
199. Hunter K, Hölscher C (2012) Drugs developed to treat diabetes, liraglutide and lixisenatide, cross the blood brain barrier and enhance neurogenesis. *BMC Neurosci* 13(1):33. <https://doi.org/10.1186/1471-2202-13-33>
200. Bertilsson G et al (2008) Peptide hormone exendin-4 stimulates subventricular zone neurogenesis in the adult rodent brain and induces recovery in an animal model of Parkinson's disease. *J Neurosci Res* 86(2):326–338. <https://doi.org/10.1002/jnr.21483>
201. Harkavyi A, Abuirmeileh A, Lever R, Kingsbury AE, Biggs CS, Whitton PS (2008) Glucagon-like peptide 1 receptor stimulation reverses key deficits in distinct rodent models of Parkinson's disease. *J Neuroinflammation* 5(1):19. <https://doi.org/10.1186/1742-2094-5-19>
202. Li Y et al (2009) GLP-1 receptor stimulation preserves primary cortical and dopaminergic neurons in cellular and rodent models of stroke and Parkinsonism. *Proc Natl Acad Sci USA* 106(4):1285–1290. <https://doi.org/10.1073/pnas.0806720106>
203. Kosaraju J et al (2013) Saxagliptin: A dipeptidyl peptidase-4 inhibitor ameliorates streptozotocin induced Alzheimer's disease. *Neuropharmacology* 72:291–300. <https://doi.org/10.1016/j.neuropharm.2013.04.008>
204. Rizzo MR, Barbieri M, Boccardi V, Angellotti E, Marfella R, Paolisso G (2014) Dipeptidyl peptidase-4 inhibitors have protective effect on cognitive impairment in aged diabetic patients with mild cognitive impairment. *J Gerontol A Biol Sci Med Sci* 69(9):1122–1131. <https://doi.org/10.1093/gerona/glu032>
205. Bayram E, Litvan I (2020) Lowering the risk of Parkinson's disease with GLP-1 agonists and DPP4 inhibitors in type 2 diabetes. *Brain* 143(10):2868–2871. <https://doi.org/10.1093/brain/awaa287>
206. Nassar NN, Al-Shorbagy MY, Arab HH, Abdallah DM (2015) Saxagliptin: a novel antiparkinsonian approach. *Neuropharmacology* 89:308–317. <https://doi.org/10.1016/j.neuropharm.2014.10.007>
207. C. Ribeiro, A. Silva, S. Viana, and F. Pereira (2012) *Sitagliptin does not protect against MPTP-induced dopaminergic striatal toxicity*.
208. Kern W, Peters A, Fruehwald-Schultes B, Deininger E, Born J, Fehm HL (2001) Improving influence of insulin on cognitive functions in humans. *Neuroendocrinology* 74(4):270–280. <https://doi.org/10.1159/000054694>
209. Freiherr J et al (2013) Intranasal insulin as a treatment for Alzheimer's disease: a review of basic research and clinical evidence. *CNS Drugs* 27(7):505–514. <https://doi.org/10.1007/s40263-013-0076-8>
210. Plum L, Schubert M, Brüning JC (2005) The role of insulin receptor signaling in the brain. *Trends Endocrinol Metab* 16(2):59–65. <https://doi.org/10.1016/j.tem.2005.01.008>
211. Craft S et al (2017) Effects of regular and long-acting insulin on cognition and Alzheimer's disease biomarkers: a pilot clinical trial. *JAD* 57(4):1325–1334. <https://doi.org/10.3233/JAD-161256>
212. Ramalingam M, Kim S-J (2017) Protective effects of activated signaling pathways by insulin on C6 glial cell model of MPP⁺-induced Parkinson's disease. *J Recept Signal Transduction* 37(1):100–107. <https://doi.org/10.3109/10799893.2016.1171342>
213. Heras-Sandoval D, Pérez-Rojas JM, Hernández-Damián J, Pedraza-Chaverri J (2014) The role of PI3K/AKT/mTOR pathway in the modulation of autophagy and the clearance of protein aggregates in neurodegeneration. *Cell Signal* 26(12):2694–2701. <https://doi.org/10.1016/j.cellsig.2014.08.019>
214. Novak P, Pimentel Maldonado DA, Novak V (2019) Safety and preliminary efficacy of intranasal insulin for cognitive impairment in Parkinson disease and multiple system atrophy: a double-blinded placebo-controlled pilot study. *PLoS ONE* 14(4):0214364. <https://doi.org/10.1371/journal.pone.0214364>
215. Kwon KJ, Kim H-J, Shin CY, Han S-H (2010) Melatonin potentiates the neuroprotective properties of resveratrol against beta-amyloid-induced neurodegeneration by modulating AMP-activated protein kinase pathways. *J Clin Neurol* 6(3):127. <https://doi.org/10.3988/jcn.2010.6.3.127>
216. Hundal RS et al (2000) Mechanism by which metformin reduces glucose production in type 2 diabetes. *Diabetes* 49(12):2063–2069. <https://doi.org/10.2337/diabetes.49.12.2063>
217. Chiang M-C, Cheng Y-C, Chen S-J, Yen C-H, Huang R-N (2016) Metformin activation of AMPK-dependent pathways is neuroprotective in human neural stem cells against amyloid-beta-induced mitochondrial dysfunction. *Exp Cell Res* 347(2):322–331. <https://doi.org/10.1016/j.yexcr.2016.08.013>
218. Ng ACT, Delgado V, Borlaug BA, Bax JJ (2021) Diabetes: the combined burden of obesity and diabetes on heart disease and the role of imaging. *Nat Rev Cardiol* 18(4):291–304. <https://doi.org/10.1038/s41569-020-00465-5>
219. Agostini F, Masato A, Bubacco L, Bisaglia M (2021) Metformin repurposing for Parkinson disease therapy: opportunities and challenges. *IJMS* 23(1):398. <https://doi.org/10.3390/ijms23010398>
220. Osborne C, West E, Nolan W, McHale-Owen H, Williams A, Bate C (2016) Glimepiride protects neurons against amyloid- β -induced synapse damage. *Neuropharmacology* 101:225–236. <https://doi.org/10.1016/j.neuropharm.2015.09.030>
221. Alp H et al (2012) Protective effects of beta glucan and gliclazide on brain tissue and sciatic nerve of diabetic rats induced by streptozosin. *Exp Diabetes Res* 2012:1–7. <https://doi.org/10.1155/2012/230342>
222. Esmaeili MH, Bahari B, Salari A-A (2018) ATP-sensitive potassium-channel inhibitor glibenclamide attenuates HPA axis hyperactivity, depression- and anxiety-related symptoms in a rat model of Alzheimer's disease. *Brain Res Bull* 137:265–276. <https://doi.org/10.1016/j.brainresbull.2018.01.001>
223. Qiu X, Wang Q, Hou L, Zhang C, Wang Q, Zhao X (2021) Inhibition of NLRP3 inflammasome by glibenclamide attenuated dopaminergic neurodegeneration and motor deficits in paraquat and maneb-induced mouse Parkinson's disease model. *Toxicol Lett* 349:1–11. <https://doi.org/10.1016/j.toxlet.2021.05.008>
224. Ju Y-J et al (2020) Glibenclamide modulates microglial function and attenuates A β deposition in 5XFAD mice. *Eur J Pharmacol* 884:173416. <https://doi.org/10.1016/j.ejphar.2020.173416>
225. Landreth G (2007) Therapeutic use of agonists of the nuclear receptor PPAR γ in Alzheimer's disease. *CAR* 4(2):159–164. <https://doi.org/10.2174/156720507780362092>

226. Heneka MT et al (2005) Acute treatment with the PPAR γ agonist pioglitazone and ibuprofen reduces glial inflammation and A β 1–42 levels in APPV7171 transgenic mice. *Brain* 128(6):1442–1453. <https://doi.org/10.1093/brain/awh452>
227. Yu Y et al (2015) Insulin sensitizers improve learning and attenuate tau hyperphosphorylation and neuroinflammation in 3xTg-AD mice. *J Neural Transm* 122(4):593–606. <https://doi.org/10.1007/s00702-014-1294-z>
228. Quinn LP et al (2008) The PPAR γ agonist pioglitazone is effective in the MPTP mouse model of Parkinson's disease through inhibition of monoamine oxidase B: pioglitazone inhibits MAO-B activity in MPTP mice. *Br J Pharmacol* 154(1):226–233. <https://doi.org/10.1038/bjp.2008.78>
229. Hunter RL, Choi D-Y, Ross SA, Bing G (2008) Protective properties afforded by pioglitazone against intrastriatal LPS in Sprague-Dawley rats. *Neurosci Lett* 432(3):198–201. <https://doi.org/10.1016/j.neulet.2007.12.019>
230. Lee EY, Lee JE, Park JH, Shin IC, Koh HC (2012) Rosiglitazone, a PPAR- γ agonist, protects against striatal dopaminergic neurodegeneration induced by 6-OHDA lesions in the substantia nigra of rats. *Toxicol Lett* 213(3):332–344. <https://doi.org/10.1016/j.toxlet.2012.07.016>
231. Corona JC, de Souza SC, Duchon MR (2014) PPAR γ activation rescues mitochondrial function from inhibition of complex I and loss of PINK1. *Exp Neurol* 253:16–27. <https://doi.org/10.1016/j.expneurol.2013.12.012>
232. Corona JC, Duchon MR (2015) PPAR γ and PGC-1 α as therapeutic targets in Parkinson's. *Neurochem Res* 40(2):308–316. <https://doi.org/10.1007/s11064-014-1377-0>
233. Markowicz-Piasecka M, Huttunen KM, Sikora J (2018) Metformin and its sulphonamide derivative simultaneously potentiate anti-cholinesterase activity of donepezil and inhibit beta-amyloid aggregation. *J Enzyme Inhib Med Chem* 33(1):1309–1322. <https://doi.org/10.1080/14756366.2018.1499627>
234. Le Pelletier L et al (2021) Metformin alleviates stress-induced cellular senescence of aging human adipose stromal cells and the ensuing adipocyte dysfunction. *eLife* 10:e62635. <https://doi.org/10.7554/eLife.62635>
235. Pan Y et al (2016) Metformin reduces morphine tolerance by inhibiting microglial-mediated neuroinflammation. *J Neuroinflammation* 13(1):294. <https://doi.org/10.1186/s12974-016-0754-9>
236. X. Chen et al (2018) "A GLP-1 receptor agonist attenuates human islet amyloid polypeptide-induced autophagy and apoptosis in MIN6 cells," *Mol Med Report* <https://doi.org/10.3892/mmr.2018.9741>
237. Jing F, Zou Q, Wang Y, Cai Z, Tang Y (2021) Activation of microglial GLP-1R in the trigeminal nucleus caudalis suppresses central sensitization of chronic migraine after recurrent nitroglycerin stimulation. *J Headache Pain* 22(1):86. <https://doi.org/10.1186/s10194-021-01302-x>
238. Abbas T, Faivre E, Hölscher C (2009) Impairment of synaptic plasticity and memory formation in GLP-1 receptor KO mice: interaction between type 2 diabetes and Alzheimer's disease. *Behav Brain Res* 205(1):265–271. <https://doi.org/10.1016/j.bbr.2009.06.035>
239. Yuan Z et al (2017) A novel GLP-1/GIP dual agonist is more effective than liraglutide in reducing inflammation and enhancing GDNF release in the MPTP mouse model of Parkinson's disease. *Eur J Pharmacol* 812:82–90. <https://doi.org/10.1016/j.ejphar.2017.06.029>
240. Maskery M, Goulding EM, Gengler S, Melchiorson JU, Rosenkilde MM, Hölscher C (2020) The dual GLP-1/GIP receptor agonist DA4-JC shows superior protective properties compared to the GLP-1 analogue liraglutide in the APP/PS1 mouse model of Alzheimer's disease. *Am J Alzheimers Dis Other Demen* 35:153331752095304. <https://doi.org/10.1177/1533317520953041>
241. Roque PS et al (2021) Intranasal insulin rescues repeated anesthesia-induced deficits in synaptic plasticity and memory and prevents apoptosis in neonatal mice via mTORC1. *Sci Rep* 11(1):15490. <https://doi.org/10.1038/s41598-021-94849-3>
242. Brabazon F, Bermudez S, Shaughnessy M, Khayrullina G, Byrnes KR (2018) The effects of insulin on the inflammatory activity of BV2 microglia. *PLoS ONE* 13(8):0201878. <https://doi.org/10.1371/journal.pone.0201878>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.