

RESEARCH ARTICLE

Cloning of nine glucocorticoid receptor isoforms from the slender African lungfish (*Protopterus dolloi*)

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Abstract

We wanted to clone the glucocorticoid receptor (GR) from slender African lungfish (*Protopterus dolloi*) for comparison to the *P. dolloi* mineralocorticoid receptor (MR), which we had cloned and were characterizing, as well as for comparison to the GRs from humans, elephant shark and zebrafish. However, although sequencing of the genome of the Australian lungfish (*Neoceratodus forsteri*), as well as, that of the West African lungfish (*Protopterus annectens*) were reported in the first three months of 2021, we could not retrieve a GR sequence with a BLAST search of GenBank, when we submitted our research for publication in July 2021. Moreover, we were unsuccessful in cloning the GR from slender African lungfish using a cDNA from the ovary of *P. dolloi* and PCR primers that had successfully cloned a GR from elephant shark, *Xenopus* and gar GRs. On October 21, 2021 the nucleotide sequence of West African lungfish (*P. annectens*) GR was deposited in GenBank. We used this GR sequence to construct PCR primers that successfully cloned the GR from the slender spotted lungfish. Here, we report the sequences of nine *P. dolloi* GR isoforms and explain the basis for the previous failure to clone a GR from slender African lungfish using PCR primers that cloned the GR from elephant shark, *Xenopus* and gar. Studies are underway to determine corticosteroid activation of these slender African lungfish GRs.

Introduction

The glucocorticoid receptor (GR) belongs to the nuclear receptor family, a diverse group of transcription factors that arose in multicellular animals [1–4]. The GR has many key roles in the physiology of humans and other terrestrial vertebrates and fish [5–8]. Important for

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understanding the function of the GR is that it is closely related to the mineralocorticoid receptor (MR) [9–11]. These two steroid receptors evolved from a duplication of an ancestral corticoid receptor (CR) in a jawless fish (cyclostome), which has descendants in modern lampreys and hagfish [11–13]. A distinct GR and MR first appear in cartilaginous fishes (Chondrichthyes) [1, 9, 11, 14, 15], which diverged from bony vertebrates about 450 million years ago [16, 17].

Lungfishes are important in the transition of vertebrates from water to land [18–22], and aldosterone activation of the MR is important in this process [11, 22–25]. Aldosterone, the main physiological mineralocorticoid in humans and other terrestrial vertebrates [26–29], first appears in lungfish [21–23]. To investigate the origins of aldosterone signaling, we cloned the MR from slender spotted African lungfish (*P. dolloi*) and studied its activation by aldosterone, other corticosteroids and progesterone [30]. To continue our investigation of early events in the evolution of the GR and MR, we sought to clone the *P. dolloi* GR for comparison with *P. dolloi* MR, as well as with the GR in coelacanths, zebrafish and humans. However, a BLAST search with the sequence of the GR from coelacanth and zebrafish did not retrieve the sequence of *P. dolloi* GR or any other lungfish GR from GenBank. Nor could we clone the *P. dolloi* GR using a cDNA from *P. dolloi* ovary using PCR primers that had successfully cloned a GR from elephant shark GR [15] and chicken, alligator and frog GRs [31]. Fortunately, on October 21, 2021 the nucleotide sequence of African lungfish (*P. annectens*) GR was deposited in GenBank, which gave us sufficient information for PCR primers to clone nine isoforms of *P. dolloi* GR. Here we report the sequences of these nine *P. dolloi* GR isoforms and explain the basis for the previous failure to clone a GR from slender African lungfish using PCR primers that previously cloned the GR from elephant shark, *Xenopus* and gar [15, 31, 32]. Our analysis of these nine GR sequences indicates that they evolved by alternative splicing and gene duplication [33, 34].

Results and discussion

Multiple sequence alignment of nine *P. dolloi* GR isoforms

Fig 1 shows a multiple sequence alignment of the nine isoforms of *P. dolloi* GR. The nine *P. dolloi* GRs cluster into three groups: group I (GR-A1, GR-A2), group II (GR-B1, GR-B2, GR-B3) and group III (GR-C1, GR-C2, GR-C3, GR-C4). GR-A2 begins at “MMDP”, a sequence motif that is conserved in all nine GRs.

The multiple alignment reveals that these nine slender African lungfish GRs evolved through alternative splicing and gene duplications (Fig 1). GR-A2 appears to be a product of alternative splicing of GR-A1. GR-C4 appears to be a product of alternative splicing of one or more GR-C isoforms, which supports a GR gene duplication in *P. dolloi* genome. There also is evidence for gene duplications among the *P. dolloi* GRs. MLSE at the beginning of GR-A1 is conserved in GR-B2 and GR-C2. A closely following YAPAD sequence is conserved in all *P. dolloi* GR isoforms. Fifteen of the first sixteen amino acids at the amino terminus of GR-A-1 are conserved in GR-B2 and GR-C2 (Fig 1A). This amino acid sequence is highly conserved in the other seven GRs. The rest of GR-A2 beginning at MMDPAGALNSLNGTQSLNKY is identical in GR-A1, and this amino acid sequence is highly conserved in the other seven GRs. MPFESLKYYAPAD is conserved at the beginning of GR-B3 and GR-C3. Beginning at the conserved MMDP sequence in the N-terminal domain, the two GR-A isoforms differ at 55 positions from the three GR-B and the four GR-C isoforms.

NTD			DBD		
GR-A1	MLSEAR	43	GR-A1	CLVCSDEASGCHYGVLT	422
GR-A2	MMDPAG	27	GR-A2	CLVCSDEASGCHYGVLT	423
GR-B1	MMSVHESRHDTDWC	60	GR-B1	CLVCSDEASGCHYGVLT	455
GR-B2	MLSEVRI	40	GR-B2	CLVCSDEASGCHYGVLT	438
GR-B3	MPFESLKYAPAD	40	GR-B3	CLVCSDEASGCHYGVLT	435
GR-C1	MMSVHESRHDTDWC	60	GR-C1	CLVCSDEASGCHYGVLT	455
GR-C2	MLSEVRI	43	GR-C2	CLVCSDEASGCHYGVLT	438
GR-C3	MPFESLKYAPAD	40	GR-C3	CLVCSDEASGCHYGVLT	435
GR-C4	MMDPAG	27	GR-C4	CLVCSDEASGCHYGVLT	422

GR-A1	SGYPFP	103	GR-A1	CLQAGM	499
GR-A2	SGYPFP	87	GR-A2	CLQAGM	483
GR-B1	SGYPFP	120	GR-B1	CLQAGM	515
GR-B2	SGYPFP	103	GR-B2	CLQAGM	498
GR-B3	SGYPFP	100	GR-B3	CLQAGM	495
GR-C1	SGYPFP	120	GR-C1	CLQAGM	515
GR-C2	SGYPFP	103	GR-C2	CLQAGM	498
GR-C3	SGYPFP	100	GR-C3	CLQAGM	495
GR-C4	SGYPFP	87	GR-C4	CLQAGM	482

GR-A1	MGFYMEVNSKAAG	163	GR-A1	EPDVIYAGYDSTSP	543
GR-A2	MGFYMEVNSKAAG	180	GR-A2	EPDVIYAGYDSTSP	513
GR-B1	MGFYMEVNSKAAG	147	GR-B1	EPDVIYAGYDSTSP	575
GR-B2	MGFYMEVNSKAAG	163	GR-B2	EPDVIYAGYDSTSP	558
GR-B3	MGFYMEVNSKAAG	160	GR-B3	EPDVIYAGYDSTSP	555
GR-C1	MGFYMEVNSKAAG	180	GR-C1	EPDVIYAGYDSTSP	575
GR-C2	MGFYMEVNSKAAG	163	GR-C2	EPDVIYAGYDSTSP	558
GR-C3	MGFYMEVNSKAAG	160	GR-C3	EPDVIYAGYDSTSP	555
GR-C4	MGFYMEVNSKAAG	147	GR-C4	EPDVIYAGYDSTSP	542

GR-A1	ADSLARGQ	223	GR-A1	WMFLM	619
GR-A2	ADSLARGQ	207	GR-A2	WMFLM	603
GR-B1	ADSLARGQ	239	GR-B1	WMFLM	635
GR-B2	ADSLARGQ	223	GR-B2	WMFLM	618
GR-B3	ADSLARGQ	219	GR-B3	WMFLM	615
GR-C1	ADSLARGQ	239	GR-C1	WMFLM	635
GR-C2	ADSLARGQ	222	GR-C2	WMFLM	618
GR-C3	ADSLARGQ	219	GR-C3	WMFLM	615
GR-C4	ADSLARGQ	206	GR-C4	WMFLM	602

GR-A1	GRITDGS	283	GR-A1	FEYILCM	679
GR-B1	GRITDGS	267	GR-A2	FEYILCM	663
GR-B2	GRITDGS	283	GR-B1	FEYILCM	695
GR-B3	GRITDGS	283	GR-B2	FEYILCM	678
GR-C1	GRITDGS	279	GR-B3	FEYILCM	675
GR-C2	GRITDGS	282	GR-C2	FEYILCM	678
GR-C3	GRITDGS	279	GR-C3	FEYILCM	675
GR-C4	GRITDGS	266	GR-C4	FEYILCM	662

GR-A1	HDSQMP	343	GR-A1	KLLDSMH	796
GR-A2	HDSQMP	327	GR-A2	KLLDSMH	780
GR-B1	HDSQMP	359	GR-B2	KLLDSMH	795
GR-B2	HDSQMP	342	GR-B3	KLLDSMH	792
GR-B3	HDSQMP	359	GR-C1	KLLDSMH	812
GR-C1	HDSQMP	342	GR-C2	KLLDSMH	795
GR-C2	HDSQMP	342	GR-C3	KLLDSMH	792
GR-C3	HDSQMP	339	GR-C4	KLLDSMH	799
GR-C4	HDSQMP	326	GR-C4	KLLDSMH	779

GR-A1	TSGGQSY	403	GR-A1	TSGGQSY	403
GR-B1	TSGGQSY	419	GR-B1	TSGGQSY	419
GR-B2	TSGGQSY	403	GR-B2	TSGGQSY	403
GR-B3	TSGGQSY	399	GR-B3	TSGGQSY	399
GR-C1	TSGGQSY	419	GR-C1	TSGGQSY	419
GR-C2	TSGGQSY	402	GR-C2	TSGGQSY	402
GR-C3	TSGGQSY	399	GR-C3	TSGGQSY	399
GR-C4	TSGGQSY	386	GR-C4	TSGGQSY	386

GR-A1	GVSGF	438	GR-A1	GVSGF	438
GR-A2	GVSGF	422	GR-A2	GVSGF	422
GR-B1	GVSGF	454	GR-B1	GVSGF	454
GR-B2	GVSGF	437	GR-B2	GVSGF	437
GR-B3	GVSGF	434	GR-B3	GVSGF	434
GR-C1	GVSGF	454	GR-C1	GVSGF	454
GR-C2	GVSGF	437	GR-C2	GVSGF	437
GR-C3	GVSGF	434	GR-C3	GVSGF	434
GR-C4	GVSGF	421	GR-C4	GVSGF	421

Fig 1. Multiple alignment of the amino acid sequences slender African lungfish glucocorticoid receptors. Total RNA was isolated from *P. dolloi* ovary and translated into cDNA. PCR was performed using four primer sets based on the sequence of *P. annectens* GR, as described in the Methods section. The amplified DNA fragments were sub-cloned into a vector for sequence analysis. Similar to other steroid receptors, slender African lungfish GR can be divided into four functional domains [6, 8], consisting of a ligand-binding domain (LBD) at the C-terminus, a DNA-binding domain (DBD) in the center that is joined to the LBD by a short hinge domain (hinge), and a domain at the amino-terminus (NTD). GenBank accession no. BDF84376 for GR-A1, BDF84377 for GR-A2, BDF84378 for GR-B1, BDF84379 for GR-B2, BDF84380 for GR-B3, BDF84381 for GR-C1, BDF84382 for GR-C2, BDF84383 for GR-C3, and BDF84384 for GR-C4. Sequences were aligned with Clustal W [35], as described in the Methods section.

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Comparison of slender African lungfish GRs and West African lungfish GRs

To better understand sequence conservation and divergence among lungfish GRs, we compared GR-A1, GR-B1 and GR-C1, which are the three longest slender African lungfish GRs, with the four West African lungfish glucocorticoid receptor sequences in GenBank (Fig 2).

The multiple sequence alignment, shown in Fig 2, reveals strong sequence conservation in the DBD, with a difference at only one position containing a semi-conserved phenylalanine-tyrosine. The sequences in the LBD and hinge domains of slender African lungfish GR and

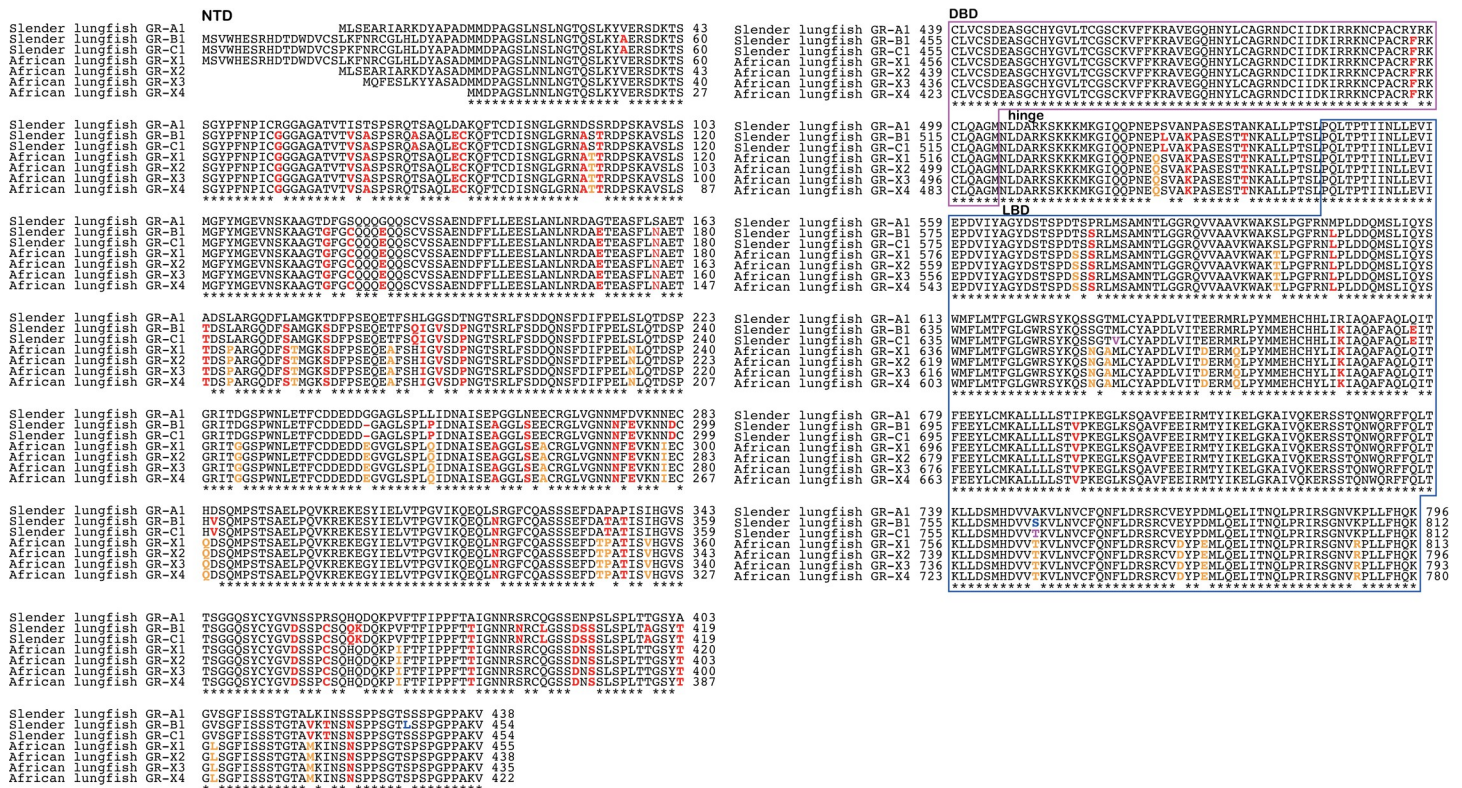


Fig 2. Multiple alignment of the amino acid sequences of three African lungfish GRs and four West African lungfish GRs. West African lungfish glucocorticoid receptor sequences were downloaded from GenBank (Accessions XP_043925084 for X1, XP_043925085 for X2, XP_043925087 for X3, XP_043925088 for X4). Sequences were aligned with Clustal W [35], as described in the Methods section.

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West African lungfish GR also are highly conserved. There are small segments of sequence divergence in the NTD, but most of the NTD is conserved. Overall slender African lungfish GRs and African lungfish GRs are very similar to each other.

Comparison of the amino acid sequences of slender African lungfish GR, West African lungfish GR, coelacanth GR, zebrafish GR and human GR

To begin to understand the relationship of lungfish GRs to other selected GRs, we constructed a multiple sequence alignment of slender African lungfish GR with West African lungfish GR, coelacanth GR, zebrafish GR and human GR (Fig 3). The DBD and hinge domains are highly conserved in all GRs. There is good sequence conservation of the LBD in all six GRs. However, there is an interesting pattern of sequence conservation in the NTD. There is excellent sequence conservation in the NTD among slender African lungfish GR, West African lungfish GR, coelacanth GR and human GR. The stronger conservation of the NTD in lungfish GRs with human GR than with zebrafish GR, indicates that the NTD in zebrafish GR has diverged from the other GRs.

Comparison of functional domains in slender African lungfish GR with domains in West African lungfish GR, coelacanth GR, zebrafish GR and human GR

Fig 4 shows the percent identity in the comparison of the different functional domains on slender African lungfish GR with the GR and MR from other vertebrates.

Slender lungfish GR	MSVWHESRHDTDWDVCSPKFNRCGLHLDYAPADM	MDPAGSLNSLNGTQSLKYAERSDKTS	60
African lungfish GR	MSVWHESRHDTDWDVCSLKFNRCGLHLDYASADM	MDPAGSLNNLNGTQSLKYVVERSDKTS	60
Coelacanth GR		MMESEKVLNTSGGD-SLKCVD-YSKRE	25
Human GR		MDSKESLTPGREE-NPSSVL-AQERG	24
Zebrafish GR		MDQGGLENGK-----KRDERL	16
Slender lungfish GR	SGYPFNPICGGAGA---TVTVSASPSRQ-ASAQLECKQFTCD	ISNGLGRNASTRDPSKA	116
African lungfish GR	SGYPFNPICGGAGA---TVTVSASPSRQ-TSAQLECKQFTCD	ISNGLGRNASTRDPSKA	116
Coelacanth GR	EYVDFNPFVFRGAPI---PLTPASSLLLV-SEADSPRQLASG	DTNKGVSNGATPQDLSCA	81
Human GR	DVMDFFYKTLRGATV---KVSASSPSLAVASQSDSKORRLLD	DFPKGVSNSNAQPDLSKA	81
Zebrafish GR	NLTLDYNKRATEGLPRRIQSTMSVAPTSMPVQAGPMQPVSG	ITPNGLSNSPTQPEHTSS	76
Slender lungfish GR	VSLSMGFYMGEVNSKAASTGFGCQQEQQSCVSSAENDFFLLE	ESLANLNRDAETEASF-	175
African lungfish GR	VSLSMGFYMGEVNSKAASTGFGCQQEQQSCVSSAENDFFLLE	ESLANLNRDAETEASF-	175
Coelacanth GR	VSESMGLYMGEVNSKAASTGFGCQQEQQSCVSSAENDFFLLE	ESLANLNRDAETEASF-	140
Human GR	VSLSMGLYMGEVNSKAASTGFGCQQEQQSCVSSAENDFFLLE	ESLANLNRDAETEASF-	140
Zebrafish GR	VS---SIFGDDSELKLLGKEQRALQQTLV-PFTLGDLSLGL	ESLANLNRDAETEASF-	132
Slender lungfish GR	LNAETDTSARGQDFSTMGKSDPSEQEFQSHIGVSDPN	GTSLRFLSDDQNSFDIFPELSL	235
African lungfish GR	LNAETDTSARGQDFSTMGKSDPSEQEFQSHIGVSDPN	GTSLRFLSDDQNSFDIFPELSL	235
Coelacanth GR	SLAPGQVSL---DNGSGMAKDLSEQETFAQT-DSDPN	GNLFFPPDQAAFDLQELDL	196
Human GR	SSASTAVSAAPTEKFPKTHSVSSEQHLKQ-TGTNG	GNVKLYTDDQSTFDLQDFL	199
Zebrafish GR	GGVDPNFLPKTEDFSPMIKGDMDLDQDSFGHIGKVDV	GNHFLSDFN--TLDLQDFEL	190
Slender lungfish GR	QTDSPGRITDGSFVNLETFCDDDED-DGAGLSP	LP-IDNAISEAGGLSEECRGLVGNNNFE	293
African lungfish GR	QTDSPGRITDGSFVNLETFCDDDED-DGAGLSP	LP-IDNAISEAGGLSEECRGLVGNNNFE	294
Coelacanth GR	TPCSPGKE---NPWSLDPIYDGGGR--GLLSPLA-ADDP	PFLMAAVANEDCKSLVTNTSQ	250
Human GR	SSGSPGKETNESPWRSDLLIDE--N--CLSLP	LAGEDDSFLLLEGNSNEDCKPLILPDKP	255
Zebrafish GR	DGSPSDFYVAD-----DAFLSTIG-EDAL	LSLEPTN-----LDRDSKAAV--SGSN	233
Slender lungfish GR	VKNNDCHVSQMPSTSAELPQVKREKESYTELVT	PGVVKOEQLNRGFCOASSEF----	349
African lungfish GR	VKNIECQDSQMPSTSAELPQVKREKESYTELVT	PGVVKOEQLNRGFCOASSEF----	350
Coelacanth GR	STNNECNLFIPDLSSQLSQQKSDKEGYTELL	TPGVVKOETLGRSVCOANLTAASAT-TA	309
Human GR	KIKDNGDLVSSPNSVTLQVTEKEDFTELC	TPGVVKOETLGRSVCOASFPGANIGNK	315
Zebrafish GR	TLNGTASSLSLSTANSILPNIKVEKDSIT	QLCTPGVVKOENTGASVCOGGLHS-----	286
Slender lungfish GR	TATISIHGVSTSGGQSYCYGVDS--SPCSQ	QKQDKPVEFTFIPFTTIGNNRNRC	407
African lungfish GR	PATISVHGVSTSGGQSYCYGVDS--SPCSQ	QKQDKPVEFTFIPFTTIGNNRNRC	408
Coelacanth GR	NSSISIHGVSTSGGQMYHYDVAAGAVSSAQ	QDPDKPIFNFIPIVSTAEINWNR	369
Human GR	MSAISVHGVSTSGGQMYHYDMN--TASLS	QKQDKPVEFTFIPPIVSGENWNR	373
Zebrafish GR	-TPTINICGVTTSSGQSLFGNSSTAVVGLQ	DOKPVEFTFIPMYTTLTSSGDGWS	345
Slender lungfish GR	SSLSPLTGGSYTGSGFTSSSTGTAVTKNSNP	-PSGTSSSPGPPAKVCLVCSDEASGCH	466
African lungfish GR	SSLSPLTGGSYTGSGFTSSSTGTAMKINSNP	-PSGTSSSPGPPAKVCLVCSDEASGCH	467
Coelacanth GR	NSTPPLGNVNASGRSGFASSYSPGTRTATPT	--PSSSTSSGPPHKLCLVCSDEASGCH	427
Human GR	NL-TSLGTLNFPGRTVFNGYSSPSMRPDVSS	PPSSSTATGPPPKLCLVCSDEASGCH	432
Zebrafish GR	SGMQQRASLCTFSKN--ES--SSPYRPE	DST--ATSSAGGTGTHKILCLVCSDEASGCH	398
Slender lungfish GR	YGVLTGCSCKVFFKRAVEGQHNYLCAGRND	CIIDKIRRNKCPACRFKRCLOAGMNL	526
African lungfish GR	YGVLTGCSCKVFFKRAVEGQHNYLCAGRND	CIIDKIRRNKCPACRFKRCLOAGMNL	527
Coelacanth GR	YGVLTGCSCKVFFKRAVEGQHNYLCAGRND	CIIDKIRRNKCPACRFKRCLOAGMNL	487
Human GR	YGVLTGCSCKVFFKRAVEGQHNYLCAGRND	CIIDKIRRNKCPACRYRKCLOAGMNL	492
Zebrafish GR	YGVLTGCSCKVFFKRAVEGQHNYLCAGRND	CIIDKIRRNKCPACRFKRCLOAGMNL	458
Slender lungfish GR	SKKMKGIQOPNEP--LVAKPASESTNKALL	PTSLPOLTPTIINLLEVIETDVIYAGYD	584
African lungfish GR	SKKMKGIQOPNEP--LVAKPASESTNKALL	PTSLPOLTPTIINLLEVIETDVIYAGYD	585
Coelacanth GR	SKKLNKMKGN-LS---S-KEQATPPLPERAV	PASVATLPTMTISLEAIEPISILYSYD	542
Human GR	TKKIKIGIQATT---GVSQETSNPNKTI	VATLPTLPTLVLLEVIETDVIYAGYD	549
Zebrafish GR	SKSKARQAGKVIQQQSIPERNLPLPEARAL	VKPMPOLVPTLVLLEVIETDVIYAGYD	518
Slender lungfish GR	STSPTDSSRLMSAMNTLGGQVVA	AAVKWAKLPGFRNLPLDDQMSLIQYSW	644
African lungfish GR	STSPTDSSRLMSAMNTLGGQVVA	AAVKWAKLPGFRNLPLDDQMSLIQYSW	645
Coelacanth GR	STIPDTHCRMLTALNKGGRQVVA	AAVKWAKLPGFRNLHLDQMVLLQYSW	602
Human GR	SSVPDSTWRIMTTLNMLGGQVVA	AAVKWAKLPGFRNLHLDQMTLLQYSW	609
Zebrafish GR	STIPDTSVRLMTLNLGGQVVA	AAVKWAKLPGFRNLHLDQMTLLQYSW	578
Slender lungfish GR	WRSYKQSSGTVLCYAPDLVITTEERMRL	LFYMMEHCHHLIKIAQAFALQITFEEY	704
African lungfish GR	WRSYKQSSGTVLCYAPDLVITTEERMRL	LFYMMEHCHHLIKIAQAFALQITFEEY	705
Coelacanth GR	WRSYQANGSMLCFAPDLIINEQRM	QLPCMYEQCHMLKIASSEFRLQVSYE	662
Human GR	WRSYQSSANLFCFAPDLIINEQRM	QLPCMYEQCHMLYVSELHRLQVSYE	669
Zebrafish GR	WRSYQHCNGNMLCFAPDLIINEERM	LFYMSDQCEQMLKISNEFVRLQVSTEE	638
Slender lungfish GR	LLLSTVPKEGLKSQAVFEEIRMTYIKELG	KAIQKERSSTQNQRFFQLTKLLDSMH	764
African lungfish GR	LLLSTVPKEGLKSQAVFEEIRMTYIKELG	KAIQKERSSTQNQRFFQLTKLLDSMH	765
Coelacanth GR	LLLSTIPQEGGLKSQPVFDEIRMTYIKELG	KAIQKERSSTQNQRFFQLTKLLDSMH	722
Human GR	LLLSSVPKDGGLKSQELFDEIRMTYIKELG	KAIQKERSSTQNQRFFQLTKLLDSMH	729
Zebrafish GR	LLLNTVPRKDGGLKSQSVFDELRMSYIKELG	KAIQKERSSTQNQRFFQLTKLLDSMH	698
Slender lungfish GR	TKVLNVCFQNFDRSRVVEYDMLQELITN	QLPRIKRSNVKPLLFHQK	812
African lungfish GR	TKVLNVCFQNFDRSRVVEYDMLQELITN	QLPRIKRSNVKPLLFHQK	813
Coelacanth GR	KELLKICRHTFVDKLSVEFPEMLAEIT	SNQLPKVTSGSKALLFHON	770
Human GR	ENLLNYCFQTFDKTMSIEFPEMLAEIT	SNQLPKVTSGSKALLFHON	777
Zebrafish GR	GGLNFCYTFVNKLSVEFPEMLAEIT	SNQLPKFKDGSVKPLLFHQK	746

Fig 3. Multiple sequence alignment of slender African lungfish GR, West African lungfish GR, coelacanth GR, zebrafish GR and human GR. Glucocorticoid receptor sequences were downloaded from GenBank (Accession no. NP_000167 for human GR, XP_005996162 for coelacanth GR, and NP_001018547 for zebrafish GR) and aligned with Clustal W [35], as described in the Methods section. The NTD in zebrafish GR has gaps and sequence differences with the other GRs.

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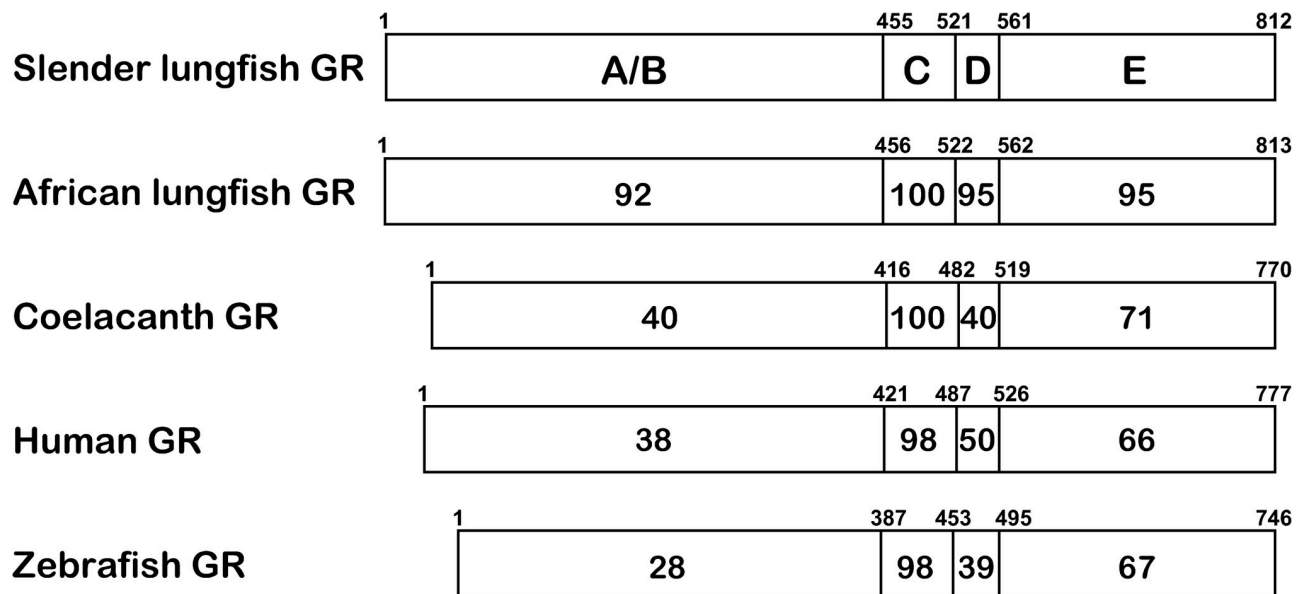


Fig 4. Comparison of functional domains of slender lungfish GR with domains in West African lungfish GR, coelacanth GR, zebrafish GR, human GR. Comparison of domains in slender African lungfish GR with GRs from West African lungfish, coelacanths, humans and zebrafish and MRs from slender African lungfish, West African lungfish, humans and zebrafish. The functional NTD (A/B), DBD (C), hinge (D) and LBD (E) domains are schematically represented with the numbers of amino acid residues and the percentage of amino acid identity depicted.

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As shown in Fig 4, the DBD and LBD are highly conserved in all GRs. For example, slender African lungfish GR and human GR have 98% and 66% identity in DBD and LBD, respectively. There are similar % identities between corresponding DBDs and LBDs in lungfish GR and other GRs. This strong conservation of the DBD and LBD contrasts with the lower sequence identity between the NTD of slender African lungfish GR and human GR (38%) and even lower sequence identity with the NTD in zebrafish GR (28%).

Phylogenetic analysis

To better understand the relationships among the nine *P. dolloi* GRs and four *P. annectens* GRs, we constructed the phylogenetic tree, shown in Fig 5. In this phylogeny, the four African lungfish GRs cluster into one group. Slender African lungfish GR-A1 and GR-A2 are in a separate branch from the other slender African lungfish GRs. GR-A2 appears to be formed by alternative splicing of GR-A1. GR-B1, GR-B2 and GR-B3 cluster. GR-C3 and GR-C4 cluster, and GR-C4 appears to be formed by alternative splicing of GR-C3.

Basis for the failure to clone *P. dolloi* GR

Fig 6 shows the location of the PCR primers that we used to successfully clone GRs from chicken, alligator and frog [31]. Due to the strong conservation of the GR and MR these PCR primers retrieved partial sequences from both the GR and MR in chicken, alligator and frog. The full sequences of these GRs and MRs was achieved in the next step using RACE. Our failure to clone *P. dolloi* GR was due using WQRFYQ instead of WQRFFQ for the 1st/2nd-reverse primer. When we used WQRFFQ we were able to clone *P. dolloi* GR.

Summary

P. dolloi contains nine GR isoforms, in contrast to *P. annectens*, which contains four GR isoforms. We do not know how many GR isoforms are in Australian lungfish (*Neoceratodus*

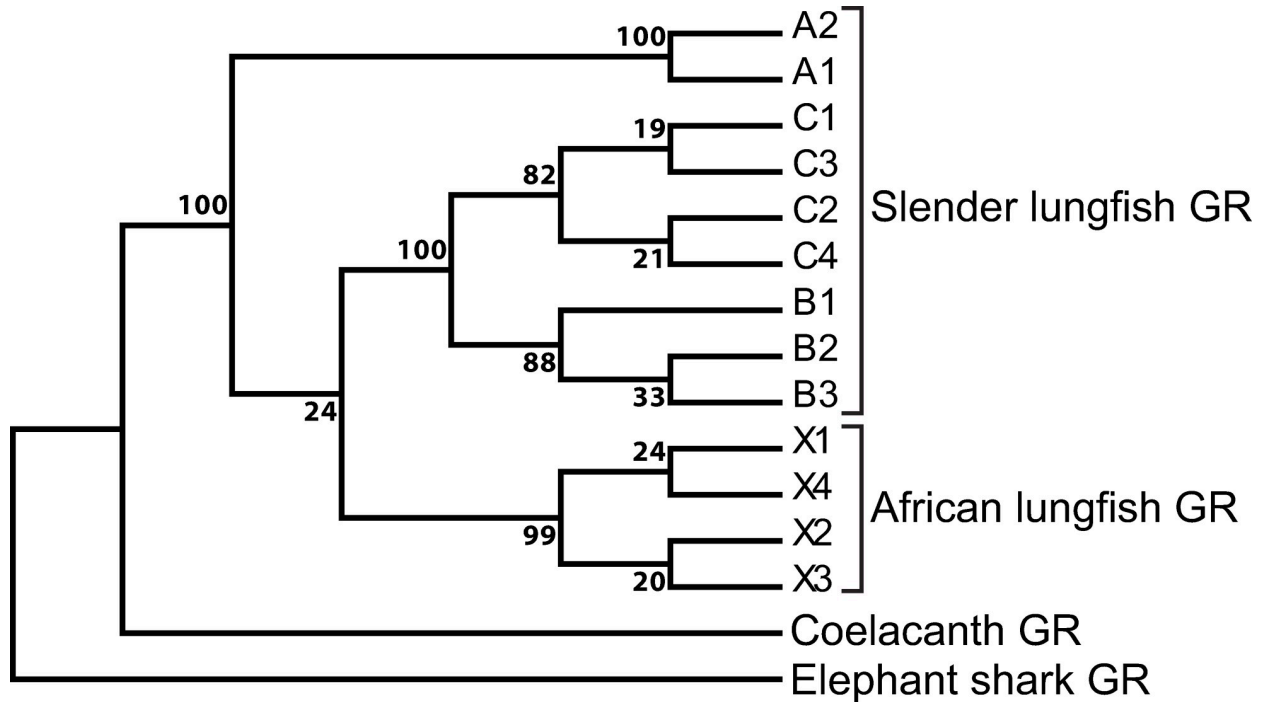


Fig 5. Phylogeny of slender African lungfish glucocorticoid receptors, West African lungfish glucocorticoid receptors, coelacanth GR and elephant shark GR. MEGA5 [36] was used to construct this phylogeny. Statistics are based on 1,000 runs.

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forsteri) because their GR sequences have not been deposited in GenBank. The availability of sequences of *P. dolloi* GRs and *P. annectens* GRs should permit using PCR to clone *N. forsteri* GRs, which would elucidate the number GR isoforms in this lungfish and the relationship of their GRs to the GRs of *P. dolloi* and *P. annectens*.

The response to corticosteroids of any lungfish GR is not known, nor are the functions of the multiple GR isoforms in *P. dolloi* GRs and *P. annectens* GRs. We have initiated studies to

1st-forward primer site

human GR	430	GCHYGV	435
chicken GR	425	GCHYGV	430
Xenopus GR	429	GCHYGV	434
zebrafish GR	396	GCHYGV	401
coelacanth GR	425	GCHYGV	430
P.a lungfish GR	465	GCHYGV	470
P.d lungfish GR	464	GCHYGV	469

2nd-forward primer site

human GR	441	CKVFFK	446
chicken GR	436	CKVFFK	441
Xenopus GR	440	CKVFFK	445
zebrafish GR	407	CKVFFK	412
coelacanth GR	436	CKVFFK	441
P.a lungfish GR	476	CKVFFK	481
P.d lungfish GR	475	CKVFFK	480

1st/2nd-reverse primer site

human GR	712	WQRFYQ	717
chicken GR	707	WQRFYQ	712
Xenopus GR	711	WQRFYQ	716
zebrafish GR	681	WQRFYQ	686
coelacanth GR	705	WQRFYQ	710
P.a lungfish GR	748	WQRFYQ	753
P.d lungfish GR	747	WQRFYQ	752

Fig 6. Location of PCR primers used for cloning of slender African lungfish GR, coelacanth GR, elephant shark GR, zebrafish GR and human GR. The correct 1st/2nd-reverse primer for PCR cloning of *P. dolloi* GR is WQRFFQ instead of WQRFYQ.

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determine corticosteroid activation of *P. dolloi* GRs to begin to elucidate the functions of slender African lungfish GRs. It is interesting that there are multiple isoforms of human GR, due to alternative splicing of human GR, and these isoforms are important in achieving functional diversity of human GR [6, 8, 34, 37]. A similar scenario is likely for *P. dolloi* GRs and *P. annectens* GRs.

Materials and methods

Animals

African lungfish (*Protopterus dolloi*) were purchased from a local commercial supplier. Lungfish were anesthetized in freshwater containing 0.02% ethyl 3-aminobenzoate methanesulfonate (Sigma-Aldrich Corp., St. Louis, MO), and tissue samples were quickly dissected and frozen in liquid nitrogen. We used two individuals of lungfish. All experiments in this study were carried out under the guidelines specified by the Institutional Animal Care and Use Committee at the Hokkaido University (Chairman: Prof. Masahiko Watanabe, permission No. 12-0015). The Institutional Animal Care and Use Committee at the Hokkaido University prospectively approved this research.

Molecular cloning of lungfish *P. dolloi* glucocorticoid receptor

For *P. dolloi* GR cloning, we designed 4 types of forward N-terminal primers:

F-X1: 5' -GTCATTTTCCCCGTGCTTAACGAA-3' ,

F-X2: 5' -GTCTGCAGCTTGAAACTTTGTAAAC-3' ,

F-X3: 5' -GACGAACATGCTGACCCGGATCATAA-3' , and

F-X4: 5' -CATACTGCATTTACCAGAATAGAC-3'

and one C-terminal Reverse primer: R: 5' -GTTAAGGCAAATTTCTGATATTAAGGCAG-3' based on the sequences of *P. annectens* GR (X1: XM_044069149, X2: XM_044069150, X3: XM_044069152, X4: XM_044069153). PCR was performed using four primer sets (F-X1xR, F-X2xR, F-X3xR, and F-X4xR) with ovary cDNA of *P. dolloi*, and the amplified DNA fragments with KOD-plus- DNA polymerase were subcloned into a cloning vector, pCR-BluntII-TOPO, and sequence analysis was performed for 10 or more clones for each primer sets.

Database and sequence analysis

GRs for phylogenetic analysis were collected with Blast searches of Genbank. A phylogenetic tree for GRs was constructed by Maximum Likelihood analysis based on the JTT + G model after sequences were aligned by Clustal W [35]. Statistical confidence for each branch in the tree was evaluated by the bootstrap methods [38] with 1000 replications. Evolutionary analyses were conducted in MEGA5 program [36].

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Methodology: Shin Oana, Xiaozhi Lin.

Resources: Susumu Hyodo.

Supervision: Yoshinao Katsu, Michael E. Baker.

Writing – original draft: Yoshinao Katsu, Laurent Bianchetti, Michael E. Baker.

Writing – review & editing: Yoshinao Katsu, Laurent Bianchetti, Michael E. Baker.

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