





Draft Genome Sequence of the Syntrophic Lactate-Degrading Bacterium *Tepidanaerobacter syntrophicus* JL^T

Norihisa Matsuura, Akiko Ohashi, Dieter M. Tourlousse, Yuji Sekiguchi

Biomedical Research Institute, National Institute of Advanced Industrial Science and Technology (AIST), Tsukuba, Ibaraki, Japan

We report here a high-quality draft genome sequence of the type strain (JL) of *Tepidanaerobacter syntrophicus*, an obligately anaerobic and moderately thermophilic bacterium, which is able to perform syntrophic lactate degradation with hydrogenotrophic methanogens. The genome comprises 2.43 Mb in 9 scaffolds, with a G+C content of 38.6%.

Received 15 December 2015 Accepted 21 December 2015 Published 11 February 2016

Citation Matsuura N, Ohashi A, Tourlousse DM, Sekiguchi Y. 2016. Draft genome sequence of the syntrophic lactate-degrading bacterium *Tepidanaerobacter syntrophicus* JL^T. Genome Announc 4(1):e01712-15. doi:10.1128/genomeA.01712-15.

Copyright © 2016 Matsuura et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license

Address correspondence to Yuji Sekiguchi, y.sekiguchi@aist.go.jp.

epidanaerobacter syntrophicus is a moderately thermophilic anaerobic bacterium capable of degrading primary aliphatic alcohols and lactate in syntrophic association with hydrogenotrophic methanogens (1). The bacterium was originally isolated from sludge in a full-scale thermophilic (55°C) anaerobic digester decomposing municipal solid wastes, and it was described as a new species of a new genus in the class Clostridia of the phylum Firmicutes. As a closely related species, Tepidanaerobacter acetatoxydans was also isolated and characterized (2), and its genome content was reported (3-5). One of the characteristic differences between the two species lies in the range of substrates utilized, particularly in syntrophic coculture with hydrogenotrophs. While T. acetatoxydans can perform anaerobic syntrophic acetate oxidation, such a capability was not observed for T. syntrophicus (1). Comparative genomics of the two species may allow us to infer potential genetic features underlying these phenotypic differences related to syntrophy. With this aim, the genome of the type strain of *T. syntrophicus* (strain JL^T JCM 12098^T, NBRC 100060^T, DSM 15584^T) was sequenced.

The draft genome of T. syntrophicus JL^T was generated with a paired-end Nextera XT sequencing library (400- to 600-bp inserts) and a Nextera mate-pair library (1- to 14-kb inserts). The libraries were sequenced on an Illumina NextSeq instrument generating 2 \times 150-bp paired-end reads (200 \times and 50 \times coverages for the paired-end and mate-pair libraries, respectively). The raw reads were quality trimmed and filtered using Trimmomatic version 0.32 (6). Trimmed reads from the paired-end library were merged with FLASH version 1.2.11 (7), and those from the matepair library were processed with NextClip version 1.3.1 (8). For the mate-pair library, the resulting reads in categories A, B, and C were used. Assembly was performed using SPAdes version 3.6.0 (9), followed by additional scaffolding and manual refinement of the assembly, as described previously (10). Annotation of the genome was performed within the Integrated Microbial Genomes (IMG) platform (11).

The final high-quality draft assembly of T. syntrophicus JL^T contained 10 contigs in 9 scaffolds, for a total length of 2,427,925 bp. The genome had a G+C content of 38.6% and was

predicted to contain 2,301 protein-coding sequences and 76 RNA genes. Four complete sets of rRNA genes were identified. Although motility was not observed in our previous study (1), genome annotation indicated the presence of a nearly complete set of genes encoding flagellar proteins, as well as genes for methylaccepting chemotaxis proteins, suggesting that strain JL^T may be motile under certain conditions. The average amino acid identity (AAI) between strain JL^T and *T. acetatoxydans* (accession no. NC_019954.2) was estimated to be 77.9% using CompareM (version 0.0.5; GitHub); the two strains shared 1,576 orthologous genes. Further detailed analysis, including genome-scale metabolic modeling, will contribute to our understanding of the physiology of *T. syntrophicus*, including its syntrophic properties.

Nucleotide sequence accession numbers. The whole-genome shotgun project has been deposited at DDBJ/EMBL/GenBank under the accession number BCMU00000000 (BioProject no. PRJDB4393). The version described in this paper is version BCMU01000000.

ACKNOWLEDGMENT

This research was partly supported by a grant-in-aid for JSPS Fellows from the Japan Society for the Promotion of Science (JSPS) to N.M.

REFERENCES

- 1. Sekiguchi Y, Imachi H, Susilorukmi A, Muramatsu M, Ohashi A, Harada H, Hanada S, Kamagata Y. 2006. *Tepidanaerobacter syntrophicus* gen. nov., sp. nov., an anaerobic, moderately thermophilic, syntrophic alcohol- and lactate-degrading bacterium isolated from thermophilic digested sludges. Int J Syst Evol Microbiol 56:1621–1629. http://dx.doi.org/10.1099/ijs.0.64112-0.
- 2. Westerholm M, Roos S, Schnürer A. 2011. *Tepidanaerobacter acetatoxydans* sp. nov., an anaerobic, syntrophic acetate-oxidizing bacterium isolated from two ammonium-enriched mesophilic methanogenic processes. Syst Appl Microbiol 34:260–266. http://dx.doi.org/10.1016/j.syapm.2010.11.018.
- Müller B, Sun L, Schnürer A. 2013. First insights into the syntrophic acetate-oxidizing bacteria—a genetic study. Microbiol Open 2:35–53. http://dx.doi.org/10.1002/mbo3.50.
- Müller B, Manzoor S, Niazi A, Bongcam-Rudloff E, Schnürer A. 2015. Genome-guided analysis of physiological capacities of *Tepidanaerobacter acetatoxydans* provides insights into environmental adaptations and syn-

- trophic acetate oxidation. PLoS One 10:e0121237. http://dx.doi.org/10.1371/journal.pone.0121237.
- 5. Manzoor S, Bongcam-Rudloff E, Schnürer A, Müller B. 2013. First genome sequence of a syntrophic acetate-oxidizing bacterium, *Tepidan-aerobacter acetatoxydans* strain rel. Genome Announc 1(1):. http://dx.doi.org/10.1128/genomeA.00213-12.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics 30:2114–2120. http:// dx.doi.org/10.1093/bioinformatics/btu170.
- Magoč T, Salzberg SL. 2011. FLASH: fast length adjustment of short reads to improve genome assemblies. Bioinformatics 27:2957–2963. http:// dx.doi.org/10.1093/bioinformatics/btr507.
- Leggett RM, Clavijo BJ, Clissold L, Clark MD, Caccamo M. 2014. NextClip: an analysis and read preparation tool for Nextera Long Mate Pair libraries. Bioinformatics 30:566–568.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. http://dx.doi.org/10.1089/cmb.2012.0021.
- Sekiguchi Y, Ohashi A, Parks DH, Yamauchi T, Tyson GW, Hugenholtz P. 2015. First genomic insights into members of a candidate bacterial phylum responsible for wastewater bulking. Peer J 3:e740.
- 11. Markowitz VM, Chen I-MA, Chu K, Szeto E, Palaniappan K, Pillay M, Ratner A, Huang J, Pagani I, Tringe S, Huntemann M, Billis K, Varghese N, Tennessen K, Mavromatis K, Pati A, Ivanova NN, Kyrpides NC. 2014. IMG/M 4 version of the integrated metagenome comparative analysis system. Nucleic Acids Res 42:D568–D573. http://dx.doi.org/10.1093/nar/gkt919.