



# Commentary: Dietary Glutamic Acid, Obesity, and Depressive Symptoms in Patients With Schizophrenia

Tauseef A. Khan<sup>1,2,3†</sup>, John L. Sievenpiper<sup>1,2,3,4,5†</sup> and John D. Fernstrom<sup>6,7\*</sup>

<sup>1</sup> Department of Nutritional Sciences, Temerty Faculty of Medicine, University of Toronto, Toronto, ON, Canada, <sup>2</sup> Toronto 3D Knowledge Synthesis and Clinical Trials Unit, St. Michael's Hospital, Toronto, ON, Canada, <sup>3</sup> Clinical Nutrition and Risk Factor Modification Centre, St. Michael's Hospital, Toronto, ON, Canada, <sup>4</sup> Division of Endocrinology & Metabolism, Department of Medicine, St. Michael's Hospital, Toronto, ON, Canada, <sup>5</sup> Scientist, Li Ka Shing Knowledge Institute, St. Michael's Hospital, Toronto, ON, Canada, <sup>6</sup> Department of Psychiatry, University of Pittsburgh School of Medicine, Pittsburgh, PA, United States, <sup>7</sup> Department of Pharmacology & Chemical Biology, University of Pittsburgh School of Medicine, Pittsburgh, PA, United States

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## A commentary on

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### \*Correspondence:

John D. Fernstrom  
fernstromjd@upmc.edu

†These authors have contributed  
equally to this work

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## INTRODUCTION

Kumar et al. (1) collected self-reported 24-h recalls of foods and beverages consumed by patients with schizophrenia. From these data, they estimated 24-h intakes of dietary protein and the glutamic acid present in dietary protein. These glutamate intake estimates were evaluated to assess the relationship between 24-hr glutamate intake and depression [Beck Depression Inventory scale (BDI)]. They concluded that no significant relationship was found between dietary glutamate intake and mood. However, among those who were not obese, higher intakes of glutamate (i.e., protein) were associated with greater depression.

In the introduction, the investigators note that it is important to understand the importance of environmental factors in the development of psychiatric disorders. Their contribution concerns the dietary intake of glutamate, an amino acid which is also a brain neurotransmitter. Their interpretation of the findings in non-obese individuals seems to be that ingesting dietary protein, which contains glutamate, raises blood glutamate concentrations sufficiently to cause an increase in glutamate penetration into brain, where it acts on neuronal glutamate receptors to cause depression.

## STATISTICAL ISSUES

The authors conclusions are based on a linear regression analysis of estimated dietary glutamic acid intake with BDI score. We are concerned about these results based upon non-normality of the data. In Table 1 of their paper, the BDI data appear to be non-normally distributed based upon mean ( $14.5 \pm SD 10.2$ ) and range (2–50). For any normally distributed variable, about two-thirds of the cases are within 1-SD of the mean (in this case, 4.3 and 24.7) and 95% will be within two SDs of the mean (in this case, –5.9 and 34.9) (2). In addition, the mean is smaller than twice the SD, which also indicates that the data are not normal (3). The data are clearly positively skewed. This indicates that most data points cluster at low ends of the BDI scale with outliers at the high end. This issue is exacerbated by the stratification by obesity, as it limits the number of observations in each group and the obese have the tendency for higher intake of glutamate ( $p = 0.086$ ).

The linear regression analysis employed in this study assumes that the data have normal distribution (though strictly it should be the residuals which are assumed to be normal); without transformation of the BDI (e.g., log transformation to normalize the distribution) this approach becomes inappropriate (4, 5). This is made clear when we attempt to estimate BDI using their regression equation for the obesity group with reasonable assumptions. The resulting estimates are uninterpretable at best and most likely biased. The slope of BDI risk with glutamic acid intake (2.39 BDI per glutamic acid grams/day) appears exaggerated as the estimates include both negative and very high BDI values. Furthermore, it is possible that a linear or straight line itself might not be the best fit. A non-linear or threshold analysis may show interesting or even opposite results.

We believe that the conclusions based upon a linearity assumption in this paper are spurious. If the authors were to run the analysis with appropriate transformation or explore non-linear or threshold analysis using appropriate methods (6), then we surmise that the correction of the large skew would effectively flatten the slope and would make the relationship non-significant or clinically trivial.

We also note that the antipsychotics were not identified, but described as chlorpromazine therapeutic equivalents. Each antipsychotic (and antidepressant) should have been identified, and included in the analysis, as several have notable effects on body weight (7, 8). Moreover, smoking should have been included as a factor in this study. Many psychotic and depressed patients smoke (9), and smoking is associated with a lower body weight, while smoking cessation can increase body weight (10, 11).

## DIETARY GLUTAMATE AND BRAIN GLUTAMATE NEUROTRANSMISSION: METABOLIC CONSIDERATIONS

The authors infer in their introduction that ingesting glutamate can lead directly to increases in brain glutamate levels and adversely modify brain functions via its neurotransmitter role. However, in this case, they misunderstand key metabolic features of dietary glutamate handling by the body. Almost all glutamate ingested each day derives from dietary protein [glutamate is abundant in dietary proteins (12)]. Free glutamate [naturally present in foods and added as monosodium glutamate (MSG)] makes a modest contribution by comparison (12). During digestion and absorption, almost all glutamate present in food (95+%) is catabolized by enterocytes (13). Consequently, only a very small fraction of ingested glutamate enters the circulation. This explains why ingesting a protein-containing (and MSG-containing) meal raises plasma glutamate levels by no more than 2-fold (14, 15). Such increases in plasma glutamate are

insufficient to push glutamate into brain, owing to the metabolic properties of capillary endothelial cells in brain. These cells are joined by tight junctions, and form the “blood-brain barrier” (BBB) (16). For glutamate, they employ several energy-dependent transporters that prevent glutamate passage from blood into brain (16). The result is that consuming large amounts of glutamate in food, even when it raises plasma glutamate, does not elevate brain glutamate levels. The articles Kumar et al. (1) cite to support glutamate administration affecting brain function involve giving direct systemic injections to animals, thus bypassing gastrointestinal metabolism (17–22). Such injections produce abnormally large increments in plasma glutamate, sufficient to overwhelm the BBB barrier to glutamate [17-fold increments or more are required (23)]. In humans, plasma glutamate concentrations can be raised as much as 10-fold under artificial conditions (a single, enormous, oral dose of MSG ingested in a liquid in the fasting state) (24, 25). However, even with such non-physiologic increases [humans do not willingly consume such large amounts of pure MSG, because it tastes unpleasant (26)], glutamate does not penetrate into brain or alter brain function (24).

A more likely path through which dietary glutamate could influence the brain is via its interaction in the alimentary canal with glutamate receptors that occur in the mouth, stomach and intestines. This route is physiologic. For example, glutamate in food binds to the amino acid taste receptor in the mouth [which in humans is tuned specifically to sodium glutamate (27)], leading ultimately to the perception of umami taste by the brain via the activation of sensory nerves in the face (28). The taste of sodium glutamate is sensed as pleasant, and is hypothesized to direct amino acid and protein-seeking behavior (28). Glutamate also binds to glutamate receptors on gastrointestinal vagal afferents that provide sensory information to brainstem and higher brain centers involved in the reflex control of GI and other functions (29). Behavioral effects, where observed in animals after consuming orally-administered glutamate [e.g., (30)], may thus be mediated via this indirect neural pathway, rather than by a direct chemical action of dietary glutamate in the brain.

Finally, Kumar et al. (1) have examined protein ingestion, not MSG ingestion. From a nutritional perspective, it would be unfortunate if those reading their article concluded that individuals with schizophrenia should reduce their daily intake of protein, particularly when many already appear to be consuming too little (Table 1). Current dietary guidelines set an adequate protein intake level at 0.8 g/kg/day (31).

## AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

## REFERENCES

- Kumar P, Kraal AZ, Prawdzik AM, Ringold AE, Ellingrod V. Dietary glutamic acid, obesity, and depressive symptoms in patients with schizophrenia. *Front Psychiatry*. (2020) 11:620097. doi: 10.3389/fpsy.2020.620097
- Acock AC. *A Gentle Introduction to Stata*. 6 ed. College Station TX: Stata Press (2018). 570p.
- Altman DG, Bland JM. Detecting skewness from summary information. *BMJ*. (1996) 313:1200. doi: 10.1136/bmj.313.7066.1200
- Bland JM, Altman DG. Transformations, means, and confidence intervals. *BMJ*. (1996) 312:1079. doi: 10.1136/bmj.312.7038.1079
- Bland JM, Altman DG. Transforming data. *BMJ*. (1996) 312:770. doi: 10.1136/bmj.312.7033.770
- Pek J, Wong O, Wong ACM. How to address non-normality: a taxonomy of approaches, reviewed, and illustrated. *Front Psychol*. (2018) 9:2104. doi: 10.3389/fpsy.2018.02104
- Allison DB, Mentore JL, Heo M, Chandler LP, Cappelleri JC, Infante MC, et al. Antipsychotic-induced weight gain: a comprehensive research synthesis. *Am J Psychiatr*. (1999) 156:1686–96.
- Serretti A, Mandelli L. Antidepressants and body weight: a comprehensive review and meta-analysis. *J Clin Psychiatry*. (2010) 71:1259–72. doi: 10.4088/JCP.09r05346blu
- Lasser K, Boyd JW, Woolhandler S, Himmelstein DU, McCormick D, Bor DH. Smoking and mental illness: a population-based prevalence study. *JAMA*. (2000) 284:2606–10. doi: 10.1001/jama.284.20.2606
- Perkins KA. Metabolic effects of cigarette smoking. *J Appl Physiol*. (1992) 72:401–9. doi: 10.1152/jappl.1992.72.2.401
- Hofstetter A, Schutz Y, Jequier E, Wahren J. Increased 24-hour energy expenditure in cigarette smokers. *N Engl J Med*. (1986) 314:79–82. doi: 10.1056/NEJM198601093140204
- Brosnan JT, Drewnowski A, Friedman MI. Is there a relationship between dietary MSG and obesity in animals or humans? *Amino Acids*. (2014) 46:2075–87. doi: 10.1007/s00726-014-1771-6
- Reeds PJ, Burrin DG, Jahoor F, Wykes L, Henry J, Frazer, et al. Enteral glutamate is almost completely metabolized in first pass by the gastrointestinal tract of infant pigs. *Am J Physiol*. (1996) 270:E413–8. doi: 10.1152/ajpendo.1996.270.3.E413
- Stegink LD, Filer LJ, Baker GL. Plasma and erythrocyte amino acid levels in normal adult subjects fed a high protein meal with and without added monosodium glutamate. *J Nutr*. (1982) 112:1953–60. doi: 10.1093/jn/112.10.1953
- Tsai PJ, Huang PC. Circadian variations in plasma and erythrocyte concentrations of glutamate, glutamine, and alanine in men on a diet without and with added monosodium glutamate. *Metabolism*. (1999) 48:1455–60. doi: 10.1016/S0026-0495(99)90159-2
- Hawkins RA, O’Kane RL, Simpson IA, Vina JR. Structure of the blood-brain barrier and its role in the transport of amino acids. *J Nutr*. (2006) 136 (1 Suppl):218S–26S. doi: 10.1093/jn/136.1.218S
- Quines CB, Rosa SG, Velasquez D, Da Rocha JT, Neto JS, Nogueira CW. Diphenyl diselenide elicits antidepressant-like activity in rats exposed to monosodium glutamate: a contribution of serotonin uptake and Na(+), K(+)-ATPase activity. *Behav Brain Res*. (2016) 301:161–7. doi: 10.1016/j.bbr.2015.12.038
- Quines CB, Rosa SG, Da Rocha JT, Gai BM, Bortolatto CF, Duarte MM, et al. Monosodium glutamate, a food additive, induces depressive-like and anxiogenic-like behaviors in young rats. *Life Sci*. (2014) 107:27–31. doi: 10.1016/j.lfs.2014.04.032
- Lima CB, Soares Gde S, Vitor SM, Castellano B, Andrade da Costa BL, Guedes RC. Neonatal treatment with monosodium glutamate lastingly facilitates spreading depression in the rat cortex. *Life Sci*. (2013) 93:388–92. doi: 10.1016/j.lfs.2013.07.009
- Hernandez-Bautista RJ, Alarcon-Aguilar FJ, Del CE-VM, Almanza-Perez JC, Merino-Aguilar H, Fainstein MK, et al. Biochemical alterations during the obese-aging process in female and male monosodium glutamate (MSG)-treated mice. *Int J Mol Sci*. (2014) 15:11473–94. doi: 10.3390/ijms150711473
- Bahadoran Z, Mirmiran Z, Ghasemi A. Monosodium glutamate (MSG)-Induced animal model of type 2 diabetes. *Methods Mol Biol*. (2019) 1916:49–65. doi: 10.1007/978-1-4939-8994-2\_3
- Zhu W, Yang F, Cai X, Zhang W, Zhang J, Cai M, et al. Role of glucocorticoid receptor phosphorylation-mediated synaptic plasticity in anxiogenic and depressive behaviors induced by monosodium glutamate. *Naunyn Schmiedebergs Arch Pharmacol*. (2021) 394:151–64. doi: 10.1007/s00210-020-01845-x
- Hu L, Fernstrom JD, Goldsmith PC. Exogenous glutamate enhances glutamate receptor subunit expression during selective neuronal injury in the ventral arcuate nucleus of postnatal mice. *Neuroendocrinology*. (1998) 68:77–88. doi: 10.1159/000054353
- Fernstrom JD, Cameron JL, Fernstrom MH, McConaha C, Weltzin TE, Kaye, et al. Short-term neuroendocrine effects of a large oral dose of monosodium glutamate in fasting male subjects. *J Clin Endocrinol Metab*. (1996) 81:184–91. doi: 10.1210/jcem.81.1.8550750
- Mourtzakis M, Graham TE. Glutamate ingestion and its effects at rest and during exercise in humans. *J Appl Physiol*. (2002) 93:1251–9. doi: 10.1152/jappphysiol.00111.2002
- Yamaguchi S, Takahashi C. Hedonic functions of monosodium glutamate and four basic taste substances used at various concentration levels in single and complex systems. *Agric Biol Chem*. (1984) 48:1077–81. doi: 10.1080/00021369.1984.10866271
- Li X. The T1R receptors mediate mammalian sweet and umami taste. *Am J Clin Nutr*. (2009) 90:733S–7S. doi: 10.3945/ajcn.2009.27462G
- Yarmolinsky DA, Zuker CS, Ryba NJ. Common sense about taste: from mammals to insects. *Cell*. (2009) 139:234–44. doi: 10.1016/j.cell.2009.10.001
- Kitamura A, Sato W, Uneyama H, Torii K, Nijijima A. Effects of intragastric infusion of inosine monophosphate and L- glutamate on vagal gastric afferent activity and subsequent autonomic reflexes. *J Physiol Sci*. (2011) 61:65–71. doi: 10.1007/s12576-010-0121-z
- Onaolapo AY, Onaolapo OJ. Dietary glutamate and the brain: in the footprints of a jekyll and hyde molecule. *Neurotoxicology*. (2020) 80:93–104. doi: 10.1016/j.neuro.2020.07.001
- Institute of Medicine. *Dietary Reference Intakes for Energy, Carbohydrate, Fiber, Fat, Fatty Acids, Cholesterol, Protein, and Amino Acids*. Washington, DC: The National Academies Press (2005). 1358p.

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European Association for the study of Diabetes (EASD), Canadian Cardiovascular Society (CCS), and Obesity Canada/Canadian Association of Bariatric Physicians and Surgeons. He serves or has served as an unpaid scientific advisor for the Food, Nutrition, and Safety Program (FNSP) and the Technical Committee on Carbohydrates of ILSI North America. He is a member of the International Carbohydrate Quality Consortium (ICQC), Executive Board Member of the Diabetes and Nutrition Study Group (DNSG) of the EASD, and Director of the Toronto 3D Knowledge Synthesis and Clinical Trials foundation. His wife is an employee of AB InBev. JF is a scientific adviser to the International Glutamate Technical Committee, Brussels, Belgium.

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