ELSEVIER

Contents lists available at ScienceDirect

# Current Research in Insect Science

journal homepage: www.elsevier.com/locate/cris



# Physiological status is a stronger predictor of nutrient selection than ambient plant nutrient content for a wild herbivore



Marion Le Gall<sup>a,\*</sup>, Mira L. Word<sup>a</sup>, Alioune Beye<sup>b</sup>, Arianne J. Cease<sup>a,c</sup>

<sup>a</sup> School of Sustainability, Arizona State University, Tempe, AZ, United States <sup>b</sup> Direction de la Protection des Végétaux, Nganda, Senegal

<sup>c</sup> School of Life Sciences, Arizona State University, Tempe, AZ, United States

#### ARTICLE INFO

Keywords: Carbohydrates Locust Protein Sex Temporal variation

### ABSTRACT

There is generally a close relationship between a consumer's food and its optimal nutrients. When there is a mismatch, it is hypothesized that mobile herbivores switch between food items to balance nutrients, however, there are limited data for field populations. In this study, we measured ambient plant nutrient content at two time points and contrasted our results with the nutrient ratio selected by wild female and male grasshoppers (*Oedaleus senegalensis*). Few plants were near *O. senegalensis'* optimal protein:carbohydrate ratio (P:C), nor were plants complementary. Grasshoppers collected earlier all regulated for a carbohydrate-biased ratio but females ate slightly more protein. We hypothesized that the long migration undertaken by this species may explain its carbohydrate needs. In contrast to most laboratory studies, grasshoppers collected later did not tightly regulate their P:C. These results suggest that field populations are not shifting their P:C to match seasonal plant nutrient shifts and that mobile herbivores rely on post-ingestive mechanisms in the face of environmental variation. Because this is among the first studies to examine the relationship between ambient nutrient landscape and physiological state our data are a key step in bridging knowledge acquired from lab studies to hypotheses regarding the role ecological factors play in foraging strategies.

# 1. Introduction

There is generally a close relationship between a consumer's food and its optimal nutrient intake (Lee et al., 2003). For instance, the optimal diet for an aphid that feeds on sugary plant sap low in nitrogen is not the same as the optimal diet for a carnivore that feeds on animals rich in lipid and protein but low in carbohydrates (Simpson and Raubenheimer, 2012). This pattern suggests that consumers have adapted to their host foods through post-ingestive regulation, or by adjusting physiological demands through behaviors such as decreased activity, or both. Consumers can also shift their nutrient intake by eating different foods based on their physiological state or environmental factors. For instance, adult honeybees select an increasingly carbohydrate-biased diet as they age (Paoli et al., 2014), and females and males of a given species may have different nutrient requirements (Jensen et al., 2015; Reddiex et al., 2013; Solon-Biet et al., 2015; South et al., 2011). Because there are limited data for free-living populations, the relative importance of nutrient availability versus physiological state on nutrient selection is largely unknown. This is particularly true for herbivores living in highly stochastic

environments where the relationships between nutrient supplies and demands will change rapidly.

There is substantial evidence that the relative amounts of dietary macronutrients (proteins, carbohydrates, lipids) consumers eat is particularly important for fitness correlates in a wide range of organisms (Harrison et al., 2014; Le Couteur et al., 2016; Simpson et al., 2015; Simpson and Raubenheimer, 2012). When given the opportunity, consumers select a ratio of macronutrients based on various external and internal factors such as toxins and pathogens (Deans et al., 2016; Le Gall, 2014; Lee et al., 2006; Povey et al., 2009; Tessnow et al., 2018), predators (Schmitz et al., 2016, 2010), activity level (Clark et al., 2013; Guglielmo, 2010), previous dietary experience (Behmer et al., 2001; Tews et al., 1992), temperature (Clissold et al., 2013; Miller et al., 2009; Musten et al., 1974), sex (Jensen et al., 2015; Maklakov et al., 2008; Reddiex et al., 2013; Solon-Biet et al., 2015; South et al., 2011), and age (Paoli et al., 2014; Roeder and Behmer, 2014). However, much of this work on consumer behavioral nutrient regulation is done in laboratories with colony-reared animals. In contrast, field-based studies tracking foraging behavior usually focus on total energy consumption

https://doi.org/10.1016/j.cris.2020.100004

Received 9 July 2020; Received in revised form 23 October 2020; Accepted 23 October 2020

2666-5158/© 2020 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

<sup>\*</sup> Corresponding author. *E-mail address:* marionlegall314@gmail.com (M. Le Gall).

(Benoit-Bird, 2004; Davies, 1977; Lobel and Ogden, 1981; Nagy et al., 1984; Werner and Mittelbach, 1981) based on the assumption that all consumers are aiming to maximize energy gain (MacArthur and Pianka, 1966; Perry and Pianka, 1997), which is often not the case (Deans et al., 2015; Harrison et al., 2014; Le Couteur et al., 2016; Le Gall and Behmer, 2014; Reddiex et al., 2013; Solon-Biet et al., 2014).

It has often been hypothesized that consumers that do not have the possibility to switch between food items are more reliant on postingestive regulation, as shown by aphids excreting excess sugars as honeydew. However, some mobile herbivores such has grasshoppers exhibit substantial post-ingestive regulatory capacities (Behmer, 2009; Clissold et al., 2010; Le Gall and Behmer, 2014), despite their wellknown capacity to adjust nutrient intake by switching between food items (Behmer, 2009). For example, grasshoppers can differentially secrete enzymes to compensate for protein and carbohydrate deficiencies (Clissold et al., 2010), and post-ingestively regulate phosphorus (Cease et al., 2016; Zhang et al., 2014). This physiological flexibility is thought to be an evolved response to the high level of nutritional heterogeneity in the environment, although functional data describing the multidimensional nutrient landscape of plant protein and carbohydrate content through space and time are scarce (Le Gall et al., 2020b; Lenhart et al., 2015). Thus, we still know comparatively little about how the relative macronutrient availability and demands of field populations change (for exceptions see (Casas et al., 2005, 2003; Lenhart et al., 2015)).

To test the potential for behavioral vs. post-ingestive regulation, we first measured plant nutrient content at two time points and contrasted our results with the nutrient ratio that field-caught grasshoppers selected when they were offered artificial food. Grasshoppers are a good model because they are usually very good at nutrient regulation (Behmer, 2009). This allowed us to create a framework to evaluate which strategies free-ranging animals are adopting in the field. For behavioral regulation to supersede post-ingestive regulation, the animal should be able to acquire plants that are 1) close to its optimal nutrient intake, or 2) unbalanced but complementary to facilitate diet mixing. If these conditions are not met in the environment, then the animal must rely on post-ingestive regulation and/or face decreased performance outcomes.

Second, we measured the plasticity of nutrient regulation for fieldcaught animals collected at different times and of different sexes to determine which factors influenced nutrient selection. We hypothesize that if a specific blend of nutrients is optimal for most physiological needs, then the insects would select a consistent nutrient ratio. If instead there is a trade-off between nutrient availability and physiological demands, then the ratio of nutrients selected by the herbivores would be plastic to match the shift in plant nutrient content. Finally, we hypothesize that if the nutrient ratio selected is influenced more by physiological needs than nutrient availability, the ratio of nutrients ingested should correlate with physiological indicators well, but poorly with changes in the nutritional landscape.

We conducted our study in the West African Sahel. The Sahel is a zone of ecoclimatic and biogeographic transition between the Sahara to the north and the Sudanian Savanna to the south. It undergoes a fast and dramatic shift in plant growth during the rainy season, so we expect plant nutrient content to change rapidly, as we have already observed in cultivated fields of millet (Le Gall et al., 2020a), which should allow us to test our hypotheses. Among herbivores living in the Sahel, the Senegalese grasshopper (*Oedaleus senegalensis*) is described as the main pest of cereal crops in the region and is a grass specialist. To our knowledge, this is the first study to examine the relationship between nutrient availability in the field, physiological state, and nutrient regulation. This a critical step in bridging advancements in nutritional regulation research from lab studies and hypotheses regarding the role ecological factors play in shaping ingestive trade-offs.

# 2. Material and methods

2.1. O. senegalensis nutritional preferences and physiological variables over time

# 2.1.1. The Senegalese grasshopper

*Oedaleus senegalensis* is a grass-feeder and a major pest of millet and other cereal crops of subsistence agriculture in the Sahel zone of West Africa. The field station where we ran the experiment belongs to the Direction de la Protection des Vegetaux (DPV), 13°49′23.8″N 15°24′19.4″W. It is located in the village of Nganda, in the West Central Agricultural Region of Senegal where most millet and groundnut production takes place.

We estimated grasshopper abundance in various agricultural fields (millet, groundnut, and fallow fields) with sweep net surveys within 20 m of each plot where vegetation samples were taken. The same researcher conducted all surveys by evenly sweeping 20 times, each a 180° arc approximately one m apart along a straight line.

# 2.1.2. Nutrient selection, mass gain, and body lipid content

We collected 5th instar grasshoppers at two time points in August 2017 around the DPV field station. We initiated the experiments on 04-Aug-2017 (early) and 17-Aug-2017, 2017 (late). We weighed and put grasshoppers in individual aerated plastic containers  $(15 \times 10 \text{ cm})$ . Each cage contained a water tube, a perch for roosting, and two dishes containing artificial diet. We prepared and dried the food in our laboratory at Arizona State University (United States) following the method developed by Simpson and Abisgold (1985). In total, we made three diets varying in protein to carbohydrate ratios but otherwise isocaloric: p35:c7, p28:c14, and p7:c35. These diets were chosen because they span extreme protein:carbohydrate ratios, therefore ensuring that the IT is not outside of the range offered to the grasshoppers. For each diet, "p" stands for percent of protein in the diet, and "c" stands for percent of carbohydrates in the diet. The protein component of all foods was a 3:1:1 mix of casein, peptone, and albumen, while the digestible carbohydrate (henceforth carbohydrates) component was a 1:1 mix of sucrose and dextrin. All foods contained similar amounts of Wesson's salt (2.4%), cholesterol (0.5%), linoleic acid (0.5%), ascorbic acid (0.3%) and vitamin mix (0.2%) (Dadd, 1961). The remainder of the diet was cellulose, a non-nutritive bulking agent.

We gave the grasshoppers one of two treatments of pre-weighed diet pairings: p7:c35 & p35:c7 or p7:c35 & p28:c14 and used 20–25 grasshoppers per treatment (approximately half males and half females). After three days, the diets were removed and dried for 24–36 h at 60 °C and then re-weighed to record consumption at the nearest 0.1 mg. Grasshoppers were weighed at the beginning and the end of the experiment to calculate mass gain. We extracted lipids from each individual grasshopper by using a series of three 24 h chloroform washes on the dried carcasses. Lipid content was calculated by subtracting post-chloroform wash dry mass from pre-chloroform wash dry mass.

# 2.2. Plant survey and chemical analysis

#### 2.2.1. Plant survey

Plant surveys and collection took place on 5-Aug-2017 (early) and on 17-Aug-2017 (late). We randomly selected 5 quadrats of 10 m by 10 m within a fallow and millet field. We selected fallow and millet fields because these are where grasshoppers have been shown to be the most abundant (Le Gall et al., 2020b; Toure et al., 2013; Word et al., 2019). For each quadrat, we recorded the three most abundant plant species and presented the data for grasses (*O. senegalensis* is a grassfeeder). For each plant species, we collected enough plant material to obtain 20 mg of dry sample (different specimens of the same species were pooled together) (Fig. ESM2). We chose the most abundant species because grass diversity and overall plant biomass were low (Fig ESM2) but grasshopper density high (Fig. ESM1), therefore the most abundant grass were likely to be representative of the plants consumed by the grasshoppers.

# 2.2.2. Chemical analysis

Upon collection, we dried the plants in an oven at 60 °C for 48 h and brought back the samples to our laboratory at Arizona State University (United States). We ground plant samples for 30 s at 200 rpm using a Retsch MM 400 ball mill. We measured plant protein content with a Bradford assay and the non-structural carbohydrate content using the phenol-sulfuric acid method (Deans et al., 2018).

## 2.3. Contrasting nutrient regulation with nutrient availability

For animals to meet their optimal nutrient intake without relying on post-ingestive mechanisms there are two possibilities: 1) plant protein:carbohydrate ratio closely resembles the optimal protein:carbohydrate ratio (aka Intake Target, thereafter IT) measured in the choice experiment or 2) unbalanced but complementary foods are available so that by mixing among plants to achieve an optimal ratio. To test the first hypothesis, we plotted the self-selected protein:carbohydrate ratios from the choice experiment as nutritional rails ( $\mathbf{y} = \text{mx}, m$ =Intake Target) and calculated the Euclidian distance separating plant nutritional content from that optimal rail in the nutrient space for the two time points. The longer the Euclidian Distance, the further an animal is from the optimal P:C ratio.

Euclidian distance =  $|mx_{plantprotein} - y_{plantcarbohydrates}|/\sqrt{(1 + m^2)}$ where  $(x_{plantprotein}, y_{carbohydrates})$  are the coordinates of each plant in the nutrient landscape.

To test the second hypothesis, we compared the median protein:carbohydrate ratio of plants (50% of plants fall on either side of this number) with the self-selected protein:carbohydrate ratio measured in the choice experiment. If the median plant protein:carbohydrate ratio and the self-selected number of plants are different, it means there are more plants on one side of the nutrient space relative to the IT, making diet mixing for an optimal balance challenging.

Methodological consideration: all the plants presented are acceptable host plants for *O. senegalensis*. We acknowledge that we did not account for non-nutritional factors (e.g., plant structure or plant defenses) that could skew host plant selection. However, measuring and ranking these non-nutritional factors was not feasible in a single study, so for the purpose of this study, we only compare variation in total plant nutrients.

# 2.4. Statistical analysis

#### 2.4.1. Grasshopper density in the field

Grasshopper density in fallow, millet, and groundnut fields at the two time points was compared using ANOVA.

#### 2.4.2. Choice experiment

The amount of protein and carbohydrate eaten for each treatment was compared using MANCOVA techniques with start mass as a covariate to correct for size differences among individuals. We used a Pillai's test statistic. In all grasshopper-related analyses, we included sex as an independent variable. Protein, carbohydrate, and total food intakes were compared using ANCOVA techniques. Grasshopper wet start masses were compared using ANCOVA techniques. Mass gain and lipid content were analyzed with ANCOVA techniques. We reported the effects of protein and carbohydrate intake on mass gain and lipid content. To account for size differences protein and carbohydrate intakes were plotted as mass corrected ratios. Temperatures were analyzed using an ANOVA.

#### 2.4.3. Plant nutrient analyses and relationship to the ITs

Plant P:C ratio was analyzed with a MANOVA and plant protein and carbohydrate content with ANOVAs. Finally, we compared the Euclidian distances of plant p:c to the ITs using ANOVA techniques; median plant p:c ratio and median ITs were compared using a median test score.

## Table 1

Results fr	om the	ANCOVA	for p	protein	intake,	carbohydrate	intake,	and total
food intal	ke at tw	o time poi	nts.					

$\begin{array}{cccccccccccccccccccccccccccccccccccc$	Variable	Source	df	F-ratio	P-value
Wet start mass 1,65 0.0140 0.9060   Sex 1,65 29.3025 <0.0001*	Early				
Sex 1,65 29.3025 <0.0001*   Treatment 1,65 0.0395 0.8430   Carbohydrate intake (g) Wet start mass 1,65 0.0095 0.9228   Sex 1,65 0.0095 0.9228   Sex 1,65 0.0085 0.9268   Food intake (g) Wet start mass 1,65 0.0141 0.9060   Sex 1,65 32.7863 <0.0001*	Protein intake (g)	Treatment	1,65	0.1928	0.6621
Treatment 1,65 0.0395 0.8430   Carbohydrate intake (g) Wet start mass 1,65 0.0095 0.9228   Sex 1,65 0.0085 0.9268   Food intake (g) Wet start mass 1,65 0.0141 0.9060   Food intake (g) Wet start mass 1,65 0.0141 0.9060   Carbohydrate intake (g) Wet start mass 1,47 0.9858 0.3262   Protein intake (g) Wet start mass 1,47 0.0731 0.7882   Carbohydrate intake (g) Wet start mass 1,47 0.0731 0.7882   Carbohydrate intake (g) Wet start mass 1,47 0.3938 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 0.0363 0.8492   Protein intake (g) Treatment 1,47 0.2284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.9669 0.1636<		Wet start mass	1,65	0.0140	0.9060
Carbohydrate intake (g) Wet start mass Sex 1,65 0.0095 0.9228   Sex 1,65 24.4067 <0.0001°		Sex	1,65	29.3025	< 0.0001*
Sex 1,65 24.4067 <0.0001*   Treatment 1,65 0.0085 0.9268   Food intake (g) Wet start mass 1,65 0.0141 0.9060   Sex 1,65 32.7863 <0.0001*		Treatment	1,65	0.0395	0.8430
Treatment 1,65 0.0085 0.9268   Food intake (g) Wet start mass 1,65 0.0141 0.9060   Sex 1,65 32.7863 <0.0001*	Carbohydrate intake (g)	Wet start mass	1,65	0.0095	0.9228
$\begin{array}{llllllllllllllllllllllllllllllllllll$		Sex	1,65	24.4067	< 0.0001*
Sex 1,65 32.7863 <0.0001*   Late Treatment 1,47 0.9858 0.3262   Protein intake (g) Wet start mass 1,47 0.0731 0.7882   Sex 1,47 0.0731 0.7882   Treatment 1,47 2.2871 0.1376   Carbohydrate intake (g) Wet start mass 1,47 0.3938 0.5335   Gardo intake (g) Wet start mass 1,47 0.0312 0.0324   Food intake (g) Wet start mass 1,47 0.0312 0.0374*   Sex 1,47 0.0363 0.8492   Food intake (g) Treatment 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5599 0.2149   Protein intake (g) Treatment 1113 1.9669 0.1636   Sex 1113 1.9640 0.3001 Wet start mass 1113 0.2703 0.6042		Treatment	1,65	0.0085	0.9268
Late Treatment 1,47 0.9858 0.3262   Protein intake (g) Wet start mass 1,47 3.6039 0.0642   Sex 1,47 0.0731 0.7882   Treatment 1,47 2.2871 0.1376   Carbohydrate intake (g) Wet start mass 1,47 0.3938 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 0.0338 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 0.0284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.9669 0.1636   Sex 1113 1.9669 0.1636 Sex 1113 1.9669 0.1636   <	Food intake (g)	Wet start mass	1,65	0.0141	0.9060
Treatment 1,47 0.9858 0.3262   Protein intake (g) Wet start mass 1,47 3.6039 0.0642   Sex 1,47 0.0731 0.7882   Treatment 1,47 2.2871 0.1376   Carbohydrate intake (g) Wet start mass 1,47 4.3067 0.0438*   Sex 1,47 0.3938 0.5335 Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 4.6047 0.0374*   Sex 1,47 0.2284 0.6351   Whole model Treatment 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.9669 0.1636   Sex 1113 1.9669 0.1636 Sex 1113 1.34224 0.0004*   Time point 1113 5.8485 0.0172* Carbohydrate intake (g) Treatment 1113 0.2703 0.6042   Sex 1113 1.0840 0.3001 Wet start mass 1113		Sex	1,65	32.7863	< 0.0001*
Protein intake (g) Wet start mass Sex 1,47 3,6039 0.0642   Sex 1,47 0.0731 0.7882   Treatment 1,47 2.2871 0.1376   Carbohydrate intake (g) Wet start mass 1,47 4.3067 0.0438*   Sex 1,47 0.0393 0.5335 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 0.0284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.9669 0.1636   Sex 1113 1.9669 0.1636 Sex 1113 1.9669 0.1636   Sex 1113 1.9669 0.1636 Sex 0.0014* 1113 0.2703 0.6042   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001 Wet start mass 1113 <t< td=""><td>Late</td><td></td><td></td><td></td><td></td></t<>	Late				
Sex 1,47 0.0731 0.7882   Treatment 1,47 0.0731 0.7882   Treatment 1,47 2.2871 0.1376   Carbohydrate intake (g) Wet start mass 1,47 4.3067 0.0438*   Sex 1,47 0.3938 0.5335 Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 4.6047 0.0374* Sex 1,47 0.2284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 1.9424 0.0004* Time point 1113 1.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042 Sex 1113 1.6032 0.2082   Food intake (g) <t< td=""><td></td><td>Treatment</td><td>1,47</td><td>0.9858</td><td>0.3262</td></t<>		Treatment	1,47	0.9858	0.3262
Treatment 1,47 2.2871 0.1376   Carbohydrate intake (g) Wet start mass 1,47 4.3067 0.0438*   Sex 1,47 0.3938 0.5335 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 0.2034 0.6351   Whole model  Sex 1,47 0.2284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Treatment 1113 1.9669 0.1636   Sex 1113 1.9640 0.0301 Wet start mass 1113 1.0840 0.3001   Wet start mass 1113 1.0840 0.3001 Wet start mass 1113 0.2703 0.6042   Sex 1113 1.6322 0.2082 Sex 1113 1.632 0.2082   Carbohydrate intake (g) Treatment 1113 1.0840 </td <td>Protein intake (g)</td> <td>Wet start mass</td> <td>1,47</td> <td>3.6039</td> <td>0.0642</td>	Protein intake (g)	Wet start mass	1,47	3.6039	0.0642
Carbohydrate intake (g) Wet start mass Sex 1,47 4,3067 0.0438*   Sex 1,47 0.3938 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 4.6047 0.0374*   Sex 1,47 0.2284 0.6551   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 1.9669 0.1636 Sex 1113 1.4224 0.0004*   Time point 1113 5.8485 0.0172* Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042 Sex 1113 0.2703 0.6042   Sex 1113 1.52377 0.0002* Time point 1113 1.6332 0.2822   Food intake (g) <		Sex	1,47	0.0731	0.7882
Sex 1,47 0.3938 0.5335   Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 4.6047 0.0374*   Sex 1,47 0.2284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 1.34224 0.0004* Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Treatment	1,47	2.2871	0.1376
Treatment 1,47 0.0512 0.8221   Food intake (g) Wet start mass 1,47 4.6047 0.0374*   Sex 1,47 0.2284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 1.34224 0.0004* 1113 1.34224 0.0004*   Time point 1113 5.8485 0.0172* 0.6042 1113 1.0840 0.3001   Wet start mass 1113 1.6322 0.2082 1113 1.6322 0.2082   Food intake (g) Treatment 1113 1.0056 0.9406   Wet start mass 1113 1.0814 0.3007	Carbohydrate intake (g)	Wet start mass	1,47	4.3067	0.0438*
Food intake (g) Wet start mass Sex 1,47 4.6047 0.0374*   Whole model 1,47 0.2284 0.6351   Whole model 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 1.34224 0.0004*   Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 1.6322 0.2082   Food intake (g) Treatment 1113 1.6032 0.2082   Food intake (g) Treatment 1113 1.0056 0.9406		Sex	1,47	0.3938	0.5335
Sex 1,47 0.2284 0.6351   Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.34224 0.0004*   Sex 1113 1.34224 0.0004* 1113 1.8400 0.3001   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 1.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Treatment	1,47	0.0512	0.8221
Whole model Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 13.4224 0.0004*   Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 0.2703 0.6042   Sex 1113 0.2377 0.0002*   Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 1.0814 0.3007	Food intake (g)	Wet start mass	1,47	4.6047	0.0374*
Time point 1113 0.0363 0.8492   Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 13.4224 0.0004*   Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.2882   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Sex	1,47	0.2284	0.6351
Protein intake (g) Treatment 1113 1.5559 0.2149   Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 13.4224 0.0004*   Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 15.032 0.282   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007	Whole model				
Protein intake (g) Wet start mass 1113 1.9669 0.1636   Sex 1113 13.4224 0.0004*   Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.282   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Time point	1113	0.0363	0.8492
Sex 1113 13.4224 0.0004*   Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007	Protein intake (g)	Treatment	1113	1.5559	0.2149
Time point 1113 5.8485 0.0172*   Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.282   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007	Protein intake (g)	Wet start mass	1113	1.9669	0.1636
Carbohydrate intake (g) Treatment 1113 1.0840 0.3001   Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.282   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Sex	1113	13.4224	0.0004*
Wet start mass 1113 0.2703 0.6042   Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Time point	1113	5.8485	0.0172*
Sex 1113 15.2377 0.0002*   Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007	Carbohydrate intake (g)	Treatment	1113	1.0840	0.3001
Time point 1113 1.6032 0.2082   Food intake (g) Treatment 1113 0.0056 0.9406   Wet start mass 1113 1.0814 0.3007		Wet start mass	1113	0.2703	0.6042
Food intake (g) Treatment 1113 0.0056 0.9406 Wet start mass 1113 1.0814 0.3007		Sex	1113	15.2377	0.0002*
Wet start mass 1113 1.0814 0.3007		Time point	1113	1.6032	0.2082
	Food intake (g)	Treatment	1113	0.0056	0.9406
Sex 1113 17.2722 <0.0001*		Wet start mass	1113	1.0814	0.3007
		Sex	1113	17.2722	< 0.0001*

*Notes*: Time point refers to the time of the testing (early or late August), Treatment refers to the food pairing (p7:c35 & p35:c7 and p7:c35 & p28:c14); wet start mass was used as a covariate to adjust for size differences among insects, and sex was included as a cofactor. For each treatment we used 20–35 grasshoppers. \* P < 0.05.

# 3. Results

# 3.1. O. senegalensis density in the field

We found few *O. senegalensis* nymphs during our surveys as many had already molted, thus we presented the data for adults only. Grasshoppers were a lot more abundant in fallow fields at the beginning of the rainy season than in other field types at either time points ( $\approx$ 39 grasshoppers per transect vs. 0–4 grasshoppers per transect, Electronic Supplementary Material (Fig. S1)).

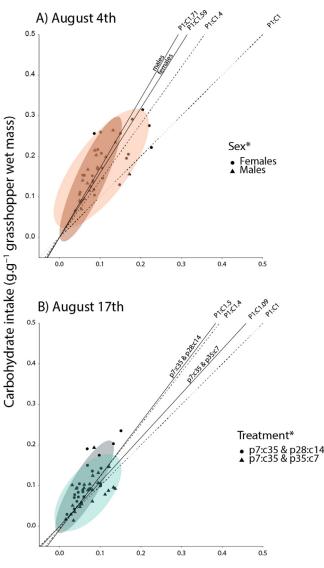
# 3.2. O. senegalensis nutritional preferences over time

# 3.2.1. Early: IT, protein intake, carbohydrate intake, and food intake

At the beginning of the rainy season (August 5th, early), grasshoppers (5th instar nymphs) actively regulated for a specific protein:carbohydrate ratio (Table S2). There were significant differences between males and females, with males regulating for a more carbohydrate-biased ratio than females (P1:C1.71 *vs.* P1:c1.59) (Fig. 1, Table S2). As a result, females ingested more protein, about twice more than males, and males more carbohydrates, about 25% more than females (Fig. 1, Table 1). Females also consumed proportionally 7% more food than males (Fig. 1, Table 1).

# 3.2.2. Late: IT, protein intake, carbohydrate intake, and food intake

Later in the rainy season (August 17th, late), insects (5th instar nymphs) did not regulate their nutrient intake as there was no convergence to a statistically indistinguishable point in nutrient space and



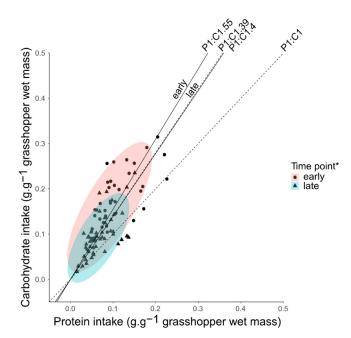


**Fig. 1.** Panel A: mass corrected protein:carbohydrate intakes by grasshoppers early (warm color) and panel B late (cold color). Straight lines represent the nutritional rails defined by the average protein:carbohydrate ratio intake. Dotted lines represent the average protein:carbohydrates ratio defined by the food pairings if consumption was random. Ellipses represent 95% confidence intervals for males and females (panel A) and food pairing treatments (panel B). For each treatment we used 20–25 grasshoppers. \*<0.05.

there were no differences between sexes (Fig. 1, Table S2). Insect start mass was significant for carbohydrate and total food intakes later in the season (Fig. 1, Table 1). In summary, later in the season, larger insects consumed more food which resulted in greater nutrient intakes.

# 3.2.3. Whole model: ITs, protein intake, carbohydrate intake, and food intake

When we compared the two time points, we found that grasshoppers selected different optimal protein:carbohydrate ratios. The ratio selected became less carbohydrate-biased later in the rainy season (P1: C1.55 vs. P1:C1.39) (Fig. 2. Table S2). The effect of time superseded both the effect of sex observed early in the season and of treatment later in the season (Table S2). For the whole model, females ate more protein, more carbohydrates, and overall more food than males (Table 1). Insects caught earlier ate more carbohydrates than insects caught later (Fig. 2, Table 1).



**Fig. 2.** Mass corrected protein:carbohydrate intakes by grasshoppers on August 4th (early = warm color) and August 17th (late = cold color). Straight lines represent the nutritional rails defined by the average protein:carbohydrate ratio calculated from the MANCOVA analysis (see Table 1). Dotted lines represent the average protein:carbohydrates ratio defined by the food pairings if consumption is random. Ellipses represent 95% confidence intervals for each time point. For each treatment we used 20–25 grasshoppers. \*<0.05.

#### Table 2

Results of ANOVA for wet start mass, and ANCOVAs for mass gain and lipid content.

Variable	Source	df	F-ratio	P-value
Wet start mass (g)	Time point	1118	77.7183	<0.0001*
	Sex	1118	128.1881	< 0.0001*
	Time point * Sex	1118	74.9377	< 0.0001*
	Time point	1112	13.0561	0.0005*
	Wet start mass	1112	83.7876	< 0.0001*
Mass gain (g)	Sex	1112	0.9801	0.3244
	Protein intake	1112	14.7505	0.0002*
	Carbohydrate intake	1112	3.7335	0.0560
	Time point	1111	1.4740	0.2274
Lipids (mg)	Wet start mass	1111	17.6075	< 0.0001*
	Sex	1111	0.0751	0.7845
	Protein intake	1111	1.1578	0.2844
	Carbohydrate intake	1111	1.6167	0.2063

*Notes:* Time point refers to the time of the testing (early or late August); wet start mass was used as a covariate to adjust for size differences among insects, and sex was included as a cofactor. Interactive terms are presented only when significant. For each treatment we used 20–35 grasshoppers. \* P < 0.05.

## 3.3. Physiological variables over time

## 3.3.1. Start mass

Initially (August 4th) there were no differences between male's and female's weights. However, the female grasshoppers we collected later were significantly heavier, they were about twice the mass of early females, and twice the mass of males at either time point, roughly 0.4 g vs. 0.2 g (Fig. 3, panel A, Table 2).

#### 3.3.2. Mass gain

Over the course of the three-day feeding experiment, we observed that insects captured later gained about three times more mass than the grasshoppers captured at the beginning of the month. We found

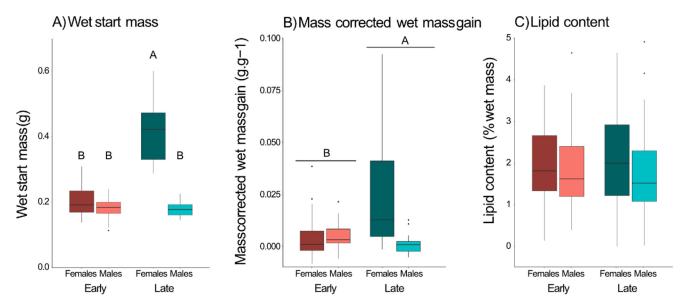


Fig. 3. Panel A: start wet mass ( $\pm$  SE); panel B: mass corrected wet mass gain ( $\pm$  SE); panel C: lipid content ( $\pm$  SE) for female (dark color) and male (light color) grasshoppers early (warm color) and late (cold color). For each treatment we used 20–25 grasshoppers. Different letters indicate statistical differences. \* <0.05.

# Table 3

Results of MANOVA for grass protein and carbohydrate ratios at two time points and ANOVAs for grass protein and carbohydrate content at two time points.

Variable	Source	df	F-ratio	P-value
Grass P:C ratio	Time point	1,32	25.13	<0.0001*
	Land-use	1,32	43.67	<0.0001*
Grass protein (%)	Time point	1,33	10.29	<0.01*
	Land-use	1,33	70.22	<0.0001*
Grass carbohydrates (%)	Time point	1,32	30.57	<0.0001*
	Land-use	1,32	0.61	0.44

*Notes*: Time point refers to the time of the testing (early or late August), and land-use to the type of field where grasses were collected (millet or fallow). \* P < 0.05.

that protein intake had a significant negative effect on mass gain, but carbohydrate intake did not (Fig. 3, panel B, Table 2).

# 3.3.3. Lipid content

Lipid content was proportional to insect mass and was not affected by time point, sex, or nutrient intake over the course of three days (Fig. 3, panel C, Table 2).

# 3.4. Relationship between plant nutrient content and ITs

3.4.1. Plant protein: carbohydrate ratio, protein and carbohydrate content We found that most grasses sampled were carbohydrate biased. Over the course of two weeks, their protein and carbohydrate shifted significantly and became increasingly less carbohydrate biased (from P1:C2.26 to P1:C1.30). We found that protein content increased over time and carbohydrate content decreased (Fig. 4, Table 3).

#### 3.4.2. Euclidian distance between ITs and plant P:C ratio

We found no differences in Euclidian distances to the ITs over time, meaning that at no time points were the insects statistically closer to their respective IT (Fig. 4, Table S3). However, the *p*-value was smaller later compared to earlier in the rainy season, suggesting "early" grasshoppers were best able to find plants that contained their optimal nutrient balance (Table S3).

#### 3.4.3. Plant distribution in regard to ITs: median test

The median protein:carbohydrate ratio selected by the insects was significantly different from the median plant protein:carbohydrate ratio, meaning that at no point was there an equal number of plants on either side of the optimal protein:carbohydrate ratio. That difference was smaller later compared to earlier in the rainy season (fewer plants were skewed toward a specific nutrient), suggesting that even if grasshoppers caught later were unable to find plants containing their optimal nutrient ratio, they could attain nutrient balance by eating from among different plants (Fig. 4, Table S4).

# 4. Discussion

Our data support the hypothesis that mobile herbivores rely on postingestive mechanisms, in addition to behavioral selection, to regulate nutrient acquisition in the face of environmental variation. While diet mixing is possible in this environment, it would require extensive foraging efforts since when insects were regulating and abundant (early season), few plants were more protein-biased than their optimal nutrient intake (IT) (Fig. 4). ITs correlated poorly with grass nutrient content but varied with age and sex, highlighting that behavioral nutrient regulation is dynamic and likely based on physiological state rather than short-term shifts in nutrient availability.

When given the choice, male and female grasshoppers collected early actively regulated for a specific IT. Males selected a slightly more carbohydrate-biased ratio than females: P1:C1.71 vs. P1:C1.59 (Fig. 1, panel A). The difference between males and females may be because acquisition and consumption of protein by females is a critical factor for egg production (Clark et al., 2013; Lee, 2010; Maklakov et al., 2008; Reddiex et al., 2013; Solon-Biet et al., 2015; South et al., 2011). Nevertheless, for both sexes, the ITs were carbohydrate biased. In a separate study on adults of the same species, we found that females preferred leaves of plants that were more carbohydrate-biased but, contrasting with the results obtained here with artificial diets, adult males did not differentiate between leaves of different nutrient content (Le Gall et al., accepted). Later in the season, there were no differences between males and females, and insects ate randomly (Fig. 1, panel B). We found that females collected later were heavier than females collected earlier, as well as males at either time point (Fig. 3), perhaps because of ovariole development (Smith, 1964). To understand these sex-dependent shifts in nutritional regulation, longer-term studies are needed (Roeder and

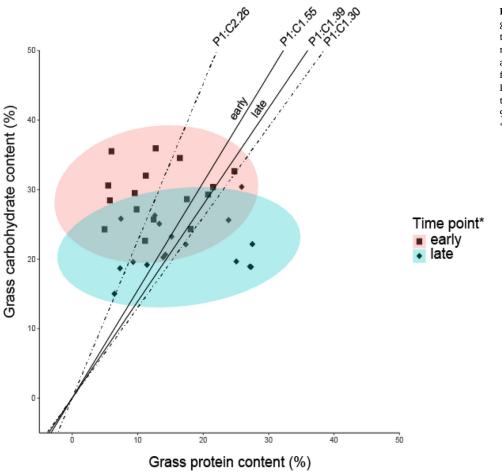


Fig. 4. Protein and carbohydrate content of grasses in fallow and millet fields at two time points (early and late). Straight lines represent the nutritional rails defined by the average protein:carbohydrate ratio calculated from the choice experiment (see Table 1). Dotted lines represent the average grass protein:carbohydrate ratio. Ellipses represent the 95% confidence intervals for each time point. \*<0.05.

Behmer, 2014), particularly since for hemimetabolous insects, nutrient acquisition during both juvenile and adult stages fuels major biological processes like reproduction and migration.

Migration has substantial impacts on nutritional demands for all animals. This is particularly true for insects as they have the highest flight metabolism known (Kammer and Heinrich, 1978). Insect migrations are usually fueled by lipid reserves acquired largely by increased carbohydrate consumption (Blem, 1980; Rankin and Burchsted, 1992; Uvarov, 1928; Weis-Fogh, 1956). Oedaleus senegalensis adults are highly migratory: at the beginning of the rainy season, the first generation usually migrate from South to North following the Intertropical Convergence Zone (Elamin et al., 2013; Holt and Colvin, 1997; Maiga et al., 2008). O. senegalensis can fly up to 350 km in a night (Cheke, 1990; Riley and Reynolds, 1990, 1983). Despite differences in nutrient selection between sexes and collection date, all nutrient ratios selected were carbohydrate biased, highlighting the key role of this nutrient for this species. However, we found no difference in body lipid content for the two food pairings (p7:c35 & p28:c14 and p7:c35 & p35:c7), over time, or between sexes. In other grasshopper species (non-migratory) wide variations in lipid levels (2 to 5.8%) are recorded when fed different foods in no-choice experiments (Le Gall et al., 2020b). We expect this species would have varying body lipid content if constrained to diets that vary in protein:carbohydrate ratios in a no-choice experiment. A potential explanation for the lack of difference in lipid proportion between males and females is that flight energetics could be solely dictated by overall mass, thus maintaining approximately 2% body lipid may be sufficient for migration in this species (Table 2). We also found a negative correlation between mass gain and protein intake, perhaps because of the strong effect that protein has on satiety (Abisgold and Simpson, 1988; Gosby et al., 2014; Simpson and Raubenheimer, 2005). In another study, it appeared that carbohydrates had a positive effect on mass gain *via* fat deposition (Le Gall and Behmer, 2014); we did not observe an effect of carbohydrate consumption on fat deposition, likely due to the shorter experimental time period (3 days vs. whole instar duration – roughly two weeks).

Besides migration, another striking feature of locust densitydependent polyphenism is a release on diet breadth constraints. Gregarious locusts are notorious for eating more plant species than solitarious locusts and they will more readily eat plant toxins (COPR, 1982; Despland and Simpson, 2005a). In model locusts, the change in food choices is thought to be, in part, an antipredator strategy where eating plant toxins make the conspicuous swarms unpalatable to predators (Despland, 2005; Despland and Simpson, 2005a, 2005b; Sword, 2002; Sword et al., 2000). However, in lab colonies of the desert locust, this does not translate in changes in nutrient regulation and similar ITs were recorded for gregarious and solitarious nymphs (Simpson et al., 2002). Oedaleus senegalensis is a non-model locust (Song, 2010), and its densitydependent polyphenism is poorly understood but appears to be less pronounced than for model species. For instance, the nymphs do not appear to form large marching bands, however, the adults migrate large distances and populations densities are highly variables with regular outbreaks (Abdelmanan, 2010; Bal et al., 2015; Cheke, 1990; El Naim et al., 2014; Elamin et al., 2013; Riley and Reynolds, 1983). There was a locust upsurge the summer we ran the experiments for the current study. Insects were found at high densities in the study area, as shown by the sweep net survey early in the season (Fig. ESM1). The adult density was lower when we recorded density the second time, likely because most had already migrated north. Thus, grasshoppers collected later in the season were potentially shifting into migratory status, which may explain their increased diet breadth and lack of nutrient regulation.

Unlike more sessile herbivores such as caterpillars, grasshoppers have been shown to be tight nutrient regulators, at least during nymphal development (Behmer, 2009). Our new data suggest that this might not consistently be the case for field populations and that there may be some sex-specific differences, although more studies are needed to demonstrate whether this is a general phenomenon or field or species-specific.

Beyond changes in the nutritional landscape, other environmental factors can influence feeding habits and nutrient demands. Temperature variation can be important in determining consumption in Orthoptera (Clissold et al., 2013; Miller et al., 2009) but there was only  $\sim$ 3 °C variation in temperatures during the period of our study (data not shown). Differences in predator (Hawlena et al., 2012) and pathogen (Graham et al., 2014) exposure can also induce a shift in foraging behavior. Foraging is a dangerous exercise due to the increased risk for predation (Bernays, 1997) and herbivores may rely more heavily on postingestive regulation when predators and pathogens are abundant. In the current study, insects that were found later may have been exposed to more diseases due to the advancement of the rainy season supporting more pathogens and predators. Such increased predator and pathogen exposure may have contributed to the herbivores having relaxed nutrient selection later in the rainy season.

Our data show that the extraordinary capacity for behavioral nutrient regulation that laboratory grasshoppers and locusts exhibit is not the only tool used by field populations to meet their nutritional requirements. Post-ingestive regulation (e.g., limiting nutrients can be preferentially extracted by increasing the production of digestive enzymes (Clissold et al., 2010); excess carbohydrate can be respired (Zanotto et al., 1997) or stored as lipid; excess protein can be eliminated as uric acid and in volatile form (Harrison, 1995; Zanotto et al., 1993) or used to produce energy via gluconeogenesis (Thompson, 2000)) is likely necessary for field-caught *O. senegalensis* grasshoppers to meet their optimal nutrient needs, which are carbohydrate-biased. The capacity to be flexible by incorporating varying degrees of nutrient selection during foraging and post-ingestive regulation may be particularly important for animals inhabiting highly dynamic xeric habitats with short growing seasons.

#### Funding

This work was supported by the National Science Foundation, DEB-1313693 to AJC.

# Author contributions

MLG and AC conceived the ideas and designed methodology; MLG, MW, AB, and AC collected the data in the field; MLG analyzed the data; MLG and AC led the writing of the manuscript. All authors contributed critically to the drafts and gave final approval for publication.

Data will be available from the Dryad Digital Repository.

#### **Declaration of Competing Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

# Acknowledgments

The authors thank La Direction de la Protection des Végétaux (DPV) for their collaboration and assistance on this project, in particular Emile Victor Coly, Ousmane Diene, Elhadji Omar Dieng, Aliou Badji, and Ansou Kamara. The Nganda DPV Phytosanitary base: Mamadou Diallo, Idrissa Biaye, Sidy Kaerou Badiane, Ismaela Thiow, Ibrahim Sadio, Makam Cisse, Manga, and Papa Ndiaye; Mamour Toure for advice working on *O. senegalensis* and field equipment support, Natalia Thompson for running the chemical assays, and Rick Overson for helpful discussions.

# References

- Abdelmanan, E.H.A., 2010. Bio-Ecology of the Senegalese Grasshopper Oedaleus Senegalensis (Krauss, 1877) (Orthoptera: Acrididae) in Kordofan Region, Sudan PhD Thesis. University of Kordofan.
- Abisgold, J.D., Simpson, S.J., 1988. The effect of dietary protein levels and haemolymph composition on the sensitivity of the maxillary palp chemoreceptors of locusts. J. Exp. Biol. 135, 215–229.
- Bal, A.B., Ouambama, Z., Moumouni, A., Dieng, I., Maiga, I.H., Gagare, S., Axelsen, J.A., 2015. A simple tentative model of the losses caused by the Senegalese grasshopper, *Oedaleus senegalensis* (Krauss 1877) to millet in the Sahel. Int. J. Pest Manag. 61, 198– 203. doi:10.1080/09670874.2015.1031201.
- Behmer, S.T., 2009. Insect herbivore nutrient regulation. Annu. Rev. Entomol. 54, 165–187.
- Behmer, S.T., Raubenheimer, D., Simpson, S.J., 2001. Frequency-dependent food selection in locusts: a geometric analysis of the role of nutrient balancing. Anim. Behav. 61, 995–1005.
- Benoit-Bird, K.J., 2004. Prey caloric value and predator energy needs: foraging predictions for wild spinner dolphins. Mar. Biol. 145, 435–444.
- Bernays, E.A., 1997. Feeding by lepidopteran larvae is dangerous. Ecol. Entomol. 22, 121–123.
- Blem, C.R., 1980. The energetics of migration, Animal Migration, Orientation, and Navigation. In: Academic Press, Inc, pp. 175–224.
- Casas, J., Driessen, G., Mandon, N., Wielaard, S., Desouhant, E., Van Alphen, J., Lapchin, L., Rivero, A., Christides, J.P., Bernstein, C., 2003. Energy dynamics in a parasitoid foraging in the wild. J. Anim. Ecol. 72, 691–697.
- Casas, J., Pincebourde, S., Mandon, N., Vannier, F., Poujol, R., Giron, D., 2005. Lifetime nutrient dynamics reveal simultaneous capital and income breeding in a parasitoid. Ecology 86, 545–554.
- Cease, A.J., Fay, M., Elser, J.J., Harrison, J.F., 2016. Dietary phosphate affects food selection, post-ingestive phosphorus fate, and performance of a polyphagous herbivore. J. Exp. Biol. 219, 64–72.
- Cheke, R.A., 1990. A migrant pest in the Sahel: the Senegalese grasshopper Oedaleus senegalensis. Phil. Trans. R. Soc. Lond. B 328, 539–553. doi:10.1098/rstb.1990.0126.
- Clark, R.M., McConnell, A., Zera, A.J., Behmer, S.T., 2013. Nutrient regulation strategies differ between cricket morphs that trade-off dispersal and reproduction. Funct. Ecol. 27, 1126–1133. doi:10.1111/1365-2435.12103.
- Clissold, F., Coggan, N., Simpson, S., 2013. Insect herbivores can choose microclimates to achieve nutritional homeostasis. J. Exp. Biol. 216, 2089–2096.
- Clissold, F.J., Tedder, B.J., Conigrave, A.D., Simpson, S.J., 2010. The gastrointestinal tract as a nutrient-balancing organ. Proc. R. Soc. B 277, 1751–1759.
- COPR, 1982. The Locust and Grasshopper Agricultural Manual. Overseas Pest Research, London, p. 690.
- Dadd, R.H., 1961. The nutritional requirements of locusts—IV. Requirements for vitamins of the B complex. Journal of Insect Physiology 6.1 1–12.
- Davies, N.B., 1977. Prey selection and the search strategy of the spotted flycatcher (*Muscicapa striata*): a field study on optimal foraging. Anim. Behav. 25, 1016–1033. doi:10.1016/0003-3472(77)90053-7.
- Deans, C.A., Sword, G.A., Behmer, S.T., 2016. Nutrition as a neglected factor in insect herbivore susceptibility to Bt toxins. Curr. Opin. Insect Sci. 15, 97–103.
- Deans, C.A., Sword, G.A., Behmer, S.T., 2015. Revisiting macronutrient regulation in the polyphagous herbivore *Helicoverpa zea* (Lepidoptera: Noctuidae): new insights via nutritional geometry. J. Insect Physiol. 81, 21–27.
- Deans, C.A., Sword, G.A., Lenhart, P.A., Burkness, E., Hutchison, W.D., Behmer, S.T., 2018. Quantifying plant soluble protein and digestible carbohydrate content, using corn (*Zea mays*) as an exemplar. J. Vis. Exp. doi:10.3791/58164.
- Despland, E., 2005. Diet breadth and anti-predator strategies in desert locusts and other Orthopterans. J. Orthoptera Res. 14, 227–233 doi:10.1665/1082-6467(2005)14[227:DBAASI]2.0.CO;2.
- Despland, E., Simpson, S.J., 2005a. Food choices of solitarious and gregarious locusts reflect cryptic and aposematic antipredator strategies. Anim. Behav. 69, 471–479. doi:10.1016/j.anbehav.2004.04.018.
- Despland, E., Simpson, S.J., 2005b. Surviving the change to warning colouration: density-dependent polyphenism suggests a route for the evolution of aposematism. Chemoecology 15, 69–75.
- El Naim A, Elamin A, Abdalla M: The biology of Senegalese grasshopper Oedaleus senegalensis, Krauss, 1977 (Orthoptera: Acrididae). Int J Adv Life Sci Technol 2014, 1:6–15.
- Elamin, A.E., Abdalla, A.M., El Naim, A.M., 2013. Studies on population dynamics of Senegalese grasshopper (*Oedaleus senegalensis*) in Kordofan of Sudan. World J. Agric. Res. 1, 85–89.
- Gosby, A.K., Conigrave, A.D., Raubenheimer, D., Simpson, S.J., 2014. Protein leverage and energy intake. Obes. Rev. 15, 183–191.
- Graham, R.I., Deacutis, J.M., Pulpitel, T., Ponton, F., Simpson, S.J., Wilson, K., 2014. Locusts increase carbohydrate consumption to protect against a fungal biopesticide. J. Insect Physiol. 69, 27–34.
- Guglielmo, C.G., 2010. Move That Fatty Acid: Fuel Selection and Transport in Migratory Birds and Bats. Oxford University Press.
- Harrison, J.F., 1995. Nitrogen Metabolism and Excretion in Locusts. Nitrogen Metabolism and Excretion. CRC Press, Inc, Boca Raton, pp. 119–131.
- Harrison, S.J., Raubenheimer, D., Simpson, S.J., Godin, J.-G.J., Bertram, S.M., 2014. Towards a synthesis of frameworks in nutritional ecology: interacting effects of protein, carbohydrate and phosphorus on field cricket fitness. Proc. R. Soc. B 281, 20140539. doi:10.1098/rspb.2014.0539.
- Hawlena, D., Strickland, M.S., Bradford, M.A., Schmitz, O.J., 2012. Fear of predation slows plant-litter decomposition. Science 336, 1434–1438. doi:10.1126/science.1220097.

Holt, J., Colvin, J., 1997. A differential equation model of the interaction between the migration of the Senegalese grasshopper, *Oedaleus senegalensis*, its predators, and a seasonal habitat. Ecol. Model. 101, 185–193.

- Jensen, K., McClure, C., Priest, N.K., Hunt, J., 2015. Sex-specific effects of protein and carbohydrate intake on reproduction but not lifespan in *Drosophila melanogaster*. Aging Cell 14, 605–615. doi:10.1111/acel.12333.
- Kammer, A.E., Heinrich, B., 1978. Insect flight metabolism. In: Advances in Insect Physiology. Elsevier, pp. 133–228.
- Le Couteur, D.G., Solon-Biet, S., Cogger, V.C., Mitchell, S.J., Senior, A., de Cabo, R., Raubenheimer, D., Simpson, S.J., 2016. The impact of low-protein highcarbohydrate diets on aging and lifespan. Cell. Mol. Life Sci. 73, 1237–1252. doi:10.1007/s00018-015-2120-y.
- Le Gall, M., 2014. Diet-Mixing in a Generalist Herbivore: Trade-Offs Between Nutrient and Allelochemical Regulation. Texas A&M University.
- Le Gall, M., Behmer, S.T., 2014. Effects of protein and carbohydrate on an insect herbivore: the vista from a fitness landscape. Integr. Comp. Biol. 54, 942–954.
- Le Gall, M., Word, M.L., Thompson, N., Beye, A., Cease, A.J., 2020a. Nitrogen fertilizer decreases survival and reproduction of female locusts by increasing plant protein to carbohydrate ratio. J. Anim. Ecol. 89, 2214–2221. doi:10.1111/1365-2656.13288.
- Le Gall, M., Word, M.L., Thompson, N., Manneh, B., Beye, A., Cease, A.J., 2020b. Linking land use and the nutritional ecology of herbivores: a case study with the Senegalese locust. Funct. Ecol. 34, 167–181.
- Lee, K.P., 2010. Sex-specific differences in nutrient regulation in a capital breeding caterpillar, *Spodoptera litura* (Fabricius). J. Insect Physiol. 56, 1685–1695. doi:10.1016/j.jinsphys.2010.06.014.
- Lee, K.P., Cory, J.S., Wilson, K., Raubenheimer, D., Simpson, S.J., 2006. Flexible diet choice offsets protein costs of pathogen resistance in a caterpillar. Proc. R. Soc. B 273, 823–829.
- Lee, K.P., Raubenheimer, D., Behmer, S.T., Simpson, S.J., 2003. A correlation between macronutrient balancing and insect host-plant range: evidence from the specialist caterpillar *Spodoptera exempta* (Walker). J. Insect Physiol. 49, 1161–1171. doi:10.1016/j.jinsphys.2003.08.013.
- Lenhart, P.A., Eubanks, M.D., Behmer, S.T., 2015. Water stress in grasslands: dynamic responses of plants and insect herbivores. Oikos 124, 381–390. doi:10.1111/oik.01370.
- Lobel, P.S., Ogden, J.C., 1981. Foraging by the herbivorous parrotfish Sparisoma radians. Mar. Biol. 64, 173–183. doi:10.1007/BF00397106.
- MacArthur, R.H., Pianka, E.R., 1966. On optimal use of a patchy environment. Am. Nat. 100, 603–609.
- Maiga, I.H., Lecoq, M., Kooyman, C., 2008. Ecology and management of the Senegalese grasshopper *Oedaleus senegalensis* (Krauss 1877) (Orthoptera: Acrididae). In: West Africa: Review and Prospects. Taylor & Francis, pp. 271–288.
- Maklakov, A.A., Simpson, S.J., Zajitschek, F., Hall, M.D., Dessmann, J., Clissold, F., Raubenheimer, D., Bonduriansky, R., Brooks, R.C., 2008. Sex-specific fitness effects of nutrient intake on reproduction and lifespan. Curr. Biol. 18, 1062–1066. doi:10.1016/j.cub.2008.06.059.
- Miller, G.A., Clissold, F.J., Mayntz, D., Simpson, S.J., 2009. Speed over efficiency: locusts select body temperatures that favour growth rate over efficient nutrient utilization. Proc. R. Soc. B 276, 3581–3589. doi:10.1098/rspb.2009.1030.
- Musten, B., Peace, D., Anderson, G.H., 1974. Food intake regulation in the weanling rat: self-selection of protein and energy. J. Nutr. 104, 563–572.
- Nagy, K.A., Huey, R.B., Bennett, A.F., 1984. Field energetics and foraging mode of kalahari lacertid lizards. Ecology 65, 588–596. doi:10.2307/1941421.
- Paoli, P.P., Donley, D., Stabler, D., Saseendranath, A., Nicolson, S.W., Simpson, S.J., Wright, G.A., 2014. Nutritional balance of essential amino acids and carbohydrates of the adult worker honeybee depends on age. Amino Acids 46, 1449–1458.
- Perry, G., Pianka, E.R., 1997. Animal foraging: past, present and future. Trends Ecol. Evol. Amst. 12, 360–364.
- Povey, S., Cotter, S.C., Simpson, S.J., Lee, K.P., Wilson, K., 2009. Can the protein costs of bacterial resistance be offset by altered feeding behaviour? J. Anim. Ecol. 78, 437–446.
- Rankin, M.A., Burchsted, J.C.A., 1992. The cost of migration in insects. Annu. Rev. Entomol. 37, 533–559.
- Reddiex, A.J., Gosden, T.P., Bonduriansky, R., Chenoweth, S.F., 2013. Sex-specific fitness consequences of nutrient intake and the evolvability of diet preferences. Am. Nat. 182, 91–102. doi:10.1086/670649.
- Riley, J.R., Reynolds, D.R., 1990. Nocturnal grasshopper migration in West Africa: transport and concentration by the wind, and the implications for air-to-air control. Philos. Trans. R. Soc. Lond. B 328, 655–672.
- Riley, J.R., Reynolds, D.R., 1983. A long-range migration of grasshoppers observed in the Sahelian zone of Mali by two radars. J. Anim. Ecol. 52, 167–183. doi:10.2307/4594.

- Roeder, K.A., Behmer, S.T., 2014. Lifetime consequences of food proteincarbohydrate content for an insect herbivore. Funct. Ecol. 28, 1135–1143. doi:10.1111/1365-2435.12262.
- Schmitz, O.J., Hawlena, D., Trussell, G.C., 2010. Predator control of ecosystem nutrient dynamics. Ecol. Lett. 13, 1199–1209.
- Schmitz, O.J., Rosenblatt, A.E., Smylie, M., 2016. Temperature dependence of predation stress and the nutritional ecology of a generalist herbivore. Ecology 97, 3119–3130. doi:10.1002/ecy.1524.
- Simpson, S., Clissold, F., Lihoreau, M., Ponton, F., Wilder, S., Raubenheimer, D., 2015. Recent advances in the integrative nutrition of arthropods. Annu. Rev. Entomol. 60, 293–311.
- Simpson, S., Raubenheimer, D., 2012. The Nature of Nutrition: A Unifying Framework From Animal Adaptation to Human Obesity. Princeton University Press.
- Simpson, S.J., Abisgold, J.D., 1985. Compensation by locusts for changes in dietary nutrients: behavioural mechanisms. Physiol. Entomol. 10, 443–452.
- Simpson, S.J., Raubenheimer, D., 2005. Obesity: the protein leverage hypothesis. Obes. Rev. 6, 133–142.
- Simpson, S.J., Raubenheimer, D., Behmer, S.T., Whitworth, A., Wright, G.A., 2002. A comparison of nutritional regulation in solitarious- and gregarious-phase nymphs of the desert locust *Schistocerca gregaria*. J. Exp. Biol. 205, 121–129.
- Smith, D.S., 1964. Ovarioles and developing eggs in grasshoppers. Can. Entomol. 96, 1255–1258. doi:10.4039/Ent961255-9.
- Solon-Biet, S.M., McMahon, A.C., Ballard, J.W.O., Ruohonen, K., Wu, L.E., Cogger, V.C., Warren, A., Huang, X., Pichaud, N., Melvin, R.G., 2014. The ratio of macronutrients, not caloric intake, dictates cardiometabolic health, aging, and longevity in ad libitum-fed mice. Cell Metab. 19, 418–430.
- Solon-Biet, S.M., Walters, K.A., Simanainen, U.K., McMahon, A.C., Ruohonen, K., Ballard, J.W.O., Raubenheimer, D., Handelsman, D.J., Couteur, D.G.L., Simpson, S.J., 2015. Macronutrient balance, reproductive function, and lifespan in aging mice. PNAS 112, 3481–3486. doi:10.1073/pnas.1422041112.
- Song, H., 2010. Density-dependent phase polyphenism in nonmodel locusts: a minireview. Psyche doi:10.1155/2011/741769.
- South, S.H., House, C.M., Moore, A.J., Simpson, S.J., Hunt, J., 2011. Male cockroaches prefer a high carbohydrate diet that makes them more attractive to females: implications for the study of condition dependence. Evol. N.Y. 65, 1594–1606. doi:10.1111/j.1558-5646.2011.01233.x.
- Sword, G.A., 2002. A role for phenotypic plasticity in the evolution of aposematism. Proc. R. Soc. B 269, 1639–1644.
- Sword, G.A., Simpson, S.J., El Hadi, O.T.M., Wilps, H., 2000. Density–dependent aposematism in the desert locust. Proc. R. Soc. B 267, 63–68.
- Tessnow, A.E., Behmer, S.T., Walsh, T.K., Sword, G.A., 2018. Protein-carbohydrate regulation in *Helicoverpa amigera* and *H. punctigera* and how diet protein-carbohydrate content affects insect susceptibility to Bt toxins. J. Insect Physiol. 106, 88–95.
- Tews, J.K., Repa, J.J., Harper, A.E., 1992. Protein selection by rats adapted to high or moderately low levels of dietary protein. Physiol. Behav. 51, 699–712.
- Thompson, S.N., 2000. Pyruvate cycling and implications for regulation of gluconeogenesis in the insect, *Manduca sexta* L. Biochem. Biophys. Res. Commun. 274, 787–793.
- Toure, M., Ndiaye, M., Diongue, A., 2013. Effect of cultural techniques: rotation and fallow on the distribution of *Oedaleus senegalensis* (Krauss, 1877) (Orthoptera: Acrididae) in Senegal. Afr. J. Agric. Res. 8, 5634–5638.
- Uvarov, B.P., 1928. Locusts and Grasshoppers. A Handbook for Their Study and Control. Imperial Bureau of Entomology, London.
- Weis-Fogh, T., 1956. Biology and physics of locust flight II. Flight performance of the desert locust (*Schistocerca gregaria*). Phil. Trans. R. Soc. Lond. B 239, 459–510.
- Werner, E.E., Mittelbach, G.G., 1981. Optimal foraging: field tests of diet choice and habitat switching. Integr. Comp. Biol. 21, 813–829. doi:10.1093/icb/21.4.813.
- Word, M.L., Hall, S.J., Robinson, B., Manneh, B., Beye, A., Cease, A.J., 2019. Soil-targeted interventions could alleviate locust and grasshopper pest pressure in West Africa. Sci. Total Environ (663) 632–643. doi:10.1016/j.scitotenv.2019.01.313.
- Zanotto, F., Gouveia, S.M., Simpson, S., Calder, D., 1997. Nutritional homeostasis in locusts: is there a mechanism for increased energy expenditure during carbohydrate overfeeding? J. Exp. Biol. 200, 2437–2448.
- Zanotto, F.P., Simpson, S.J., Raubenheimer, D., 1993. The regulation of growth by locusts through post-ingestive compensation for variation in the levels of dietary protein and carbohydrate. Physiol. Entomol. 18, 425–434. doi:10.1111/j.1365-3032.1993.tb00617.x.
- Zhang, Z., Elser, J.J., Cease, A.J., Zhang, X., Yu, Q., Han, X., Zhang, G., 2014. Grasshoppers regulate N: P stoichiometric homeostasis by changing phosphorus contents in their frass. PLoS ONE 9. doi:10.1242/jeb.078782.