

# Evaluation of nitrous oxide reduction in solid carbon source-driven counter-diffusional biofilm denitrification system

Yingrui Liu<sup>a,1</sup>, Feng Chen<sup>a,1</sup>, Yanying He<sup>a</sup>, Yufen Wang<sup>a</sup>, Tingting Zhu<sup>a,\*</sup>, Yindong Tong<sup>a</sup>, Yingxin Zhao<sup>a</sup>, Bing-Jie Ni<sup>b</sup>, Yiwen Liu<sup>a,\*</sup>

<sup>a</sup> School of Environmental Science and Engineering, Tianjin University, Tianjin 300072, PR China

<sup>b</sup> School of Civil and Environmental Engineering, University of New South Wales, Sydney, New South Wales, 2052, Australia

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## ABSTRACT

Solid carbon-driven biofilm system can provide sufficient carbon source for denitrification, while its counter-diffusional structure could inevitably induce the delayed carbon-nitrogen contact and electron transport, further affecting carbon footprints mainly contributed by nitrous oxide (N<sub>2</sub>O) at wastewater treatment plants (WWTPs). However, the detailed understanding of N<sub>2</sub>O dynamics during solid-phase denitrification (SPD) has not been disclosed. In this work, a fixed bed bioreactor driven by polycaprolactone (PCL) was constructed and operated over 180 days, achieving 97 %-99 % of total nitrogen (TN) removal efficiency. Biochemical results indicated that under the condition that each nitrogen oxide (NO<sub>x</sub>) concentration was maintained at 30 mg-N/L, the electron competition between upstream and downstream electron pools was still observed during PCL-driven denitrification even providing sufficient carbon source. For example, under the coexistent nitrate (NO<sub>3</sub><sup>-</sup>) + nitrite (NO<sub>2</sub><sup>-</sup>) + N<sub>2</sub>O condition, few electrons (i.e., 12.6 %) distributed to N<sub>2</sub>O reductase (Nos), significantly decreasing the N<sub>2</sub>O reduction rate (i.e., 1.42 mg/g VSS/h). Under the condition that TN concentration was maintained at 30 mg-N/L, the TN removal rate in the scheme containing NO<sub>3</sub><sup>-</sup> + NO<sub>2</sub><sup>-</sup> + N<sub>2</sub>O was observed to be 1.75–2.3 times higher than that of the scheme with sole NO<sub>x</sub> of 30 mg-N/L. This suggested that when treating wastewater containing multiple NO<sub>x</sub>, the PCL-driven biofilm denitrification system can not only relatively improve the total nitrogen removal efficiency, but also relatively alleviate N<sub>2</sub>O emissions. The higher abundance of *Bacteroidota* and *Comamonadaceae* ensured the stable carbon source release and nitrogen conversion states.

## 1. Introduction

Wastewater treatment plants (WWTPs) typically remove nitrate or nitrite through denitrification processes to avoid the negative effects of excess nitrogen on aquatic ecosystems (Duan et al., 2021; Hatzikioseyian et al., 2023). Among physical, chemical, and biological methods, the biological approach is deemed the most effective and promising (Zhang et al., 2025). Biological denitrification can be categorized as two metabolic pathways, namely heterotrophic and autotrophic processes, with the former being the more common and practical pathway, offering the advantages of high efficiency and low treatment cost (Fu et al., 2022; Hu et al., 2023). In general, heterotrophic denitrification occurs under anaerobic or anoxic conditions through biological heterotrophic reduction, during which denitrifying bacteria complete the denitrification process by consuming organic substrates to obtain sufficient

electrons (Fang et al., 2020). Complete heterotrophic denitrification is the successive reduction of nitrate (NO<sub>3</sub><sup>-</sup>) → nitrite (NO<sub>2</sub><sup>-</sup>) → nitric oxide (NO) → nitrous oxide (N<sub>2</sub>O) → nitrogen gas (N<sub>2</sub>) with the participation of various reductases (Zheng et al., 2024). Imbalances in the reduction rates of adjacent denitrification steps, i.e., changes in external factors such as the limited available carbon sources, would result in the accumulation of specific nitrogen oxide intermediates.

Liquid carbon sources such as sodium acetate, glucose, and methanol mainly collected through sludge treatment are the most commonly used electron donors for heterotrophic denitrification (Liu et al., 2022; Wang et al., 2024a; Wang et al., 2022b). However, water-soluble organic matter could carry the risk of insufficient or excessive dosage in operation and toxicity or flammability during storage and transportation, as well as require complex monitoring and control process. In contrast to conventional liquid carbon sources, solid carbon sources exhibit the

\* Corresponding authors.

E-mail addresses: [ttzhu@tju.edu.cn](mailto:ttzhu@tju.edu.cn) (T. Zhu), [yiwen.liu@tju.edu.cn](mailto:yiwen.liu@tju.edu.cn) (Y. Liu).

<sup>1</sup> Yingrui Liu and Feng Chen contributed equally to the work.

advantages of easy management, infrequent addition, low sludge yield, controllable carbon release, and appreciable denitrification rate (Zhang et al., 2022a). In this regard, solid-phase denitrification (SPD) has been considered as a promising alternative technology. Solid carbon sources comprise synthetic biodegradable polymers and natural materials, with the former allowing for higher denitrification efficiency and more stable effluent quality (Chu and Wang, 2013). Solid carbon sources can serve as both the electron donor and biofilm carrier, thus leading to the unique counter-diffusional mass transfer behavior. The dense biofilm that grows and attaches to the polymer surface provides a protective zone for the internal denitrifiers, thereby enhancing the resilience of functional bacteria to shock loadings (Fu et al., 2022). Among the various biodegradable polymers, polycaprolactone (PCL) is the most widely studied and applied with respect to better bioavailability and more attractive economics (Zhang et al., 2022a).

During biological nitrogen removal in wastewater treatment,  $N_2O$ , the intermediate product of denitrification with global warming potential of approximately 300 times greater than that of carbon dioxide ( $CO_2$ ), is inevitably produced (Liu et al., 2025).  $N_2O$  has caused profound negative impacts on the ozone layer for its high global warming potential and long lifetime (Dai et al., 2021). It has been indicated that in recent decades, atmospheric  $N_2O$  concentration has increased at an average rate of 0.26 % per year (Huang et al., 2023). Reportedly,  $N_2O$  accounted for ca. 23 % of the direct emissions from total wastewater treatment plants worldwide (He et al., 2023a). Denitrification could act as both a source and sink of  $N_2O$  (Tong et al., 2024). Therefore, understanding the pathway of  $N_2O$  production and controlling the denitrification process as an effective  $N_2O$  sink rather than a net source is considered crucial opportunities to reduce the greenhouse effect and carbon footprint (He et al., 2023b; He et al., 2024b). Regarding  $N_2O$  production during the SPD process, Liu et al. (2019) observed  $N_2O$  accumulation in an aerobic granular sludge (AGS) pre-reactor and a poly (butylene succinate)-supported SPD post-reactor. However, little is known about the dynamics of  $N_2O$  production during SPD.

The aim of this study was to investigate the  $N_2O$  turnovers (i.e., production and consumption) under different combinations of electron acceptors and different nitrogen loading conditions in a counter-diffusion biofilm system represented by PCL. An upflow fixed-bed reactor was constructed and operated for >180 days to form a stable biofilm on the surface of the solid carbon source, followed by multiple sets of batch experiments and microbial characterization. In addition, this study provided an inner-cycle mode method that can be used to determine the cyclic variation of  $N_2O$  in granular counter-diffusion biofilms.

## 2. Results and discussion

### 2.1. SPD reactor performance

Since the amount of organic carbon released from solid carbon sources is regulated by bacteria that respond to bulk nitrate concentration, SPD systems can be considered as non-carbon-limiting conditions. An up-flow fixed bed reactor was operated for a period of 180 days. The denitrification performance of the reactor and the characteristic of the synthetic wastewater were presented in Fig. 1 and Table S1, respectively. Stable  $NO_3^-$  removal rates were obtained within the first 15 days, after which a long operation period was taken for biofilm formation, development, and stabilization on the PCL pellets surface. The total nitrogen removal efficiency was above 97 % at 154–180 days, indicating a higher denitrification performance.

### 2.2. Electron competition

Fig. 2A–G showed the variation of nitrogen oxide ( $NO_x$ ) profiles, which was based on the data of the first 1 h in schemes A–G (Table 1). Schemes A–C represented the sole  $NO_3^-$ ,  $NO_2^-$  and  $N_2O$  addition,

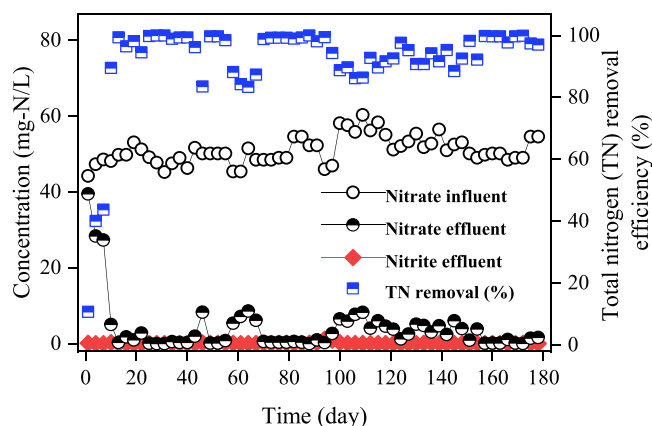


Fig. 1. The performance of the up-flow solid-phase denitrification reactor during 180 days of operation.

respectively. Schemes d–F represented  $NO_3^-+NO_2^-$ ,  $NO_3^-+N_2O$ ,  $NO_2^-+N_2O$  and  $NO_3^-+NO_2^-+N_2O$ , respectively. Each  $NO_x$  concentration in each scheme was 30 mg-N/L. The profiles of  $NO_3^-$ ,  $NO_2^-$  and  $N_2O$  in schemes A–C and E were displayed a linearly downward trend, which was consistent with previous study (Liu et al., 2022). Notably, the rate of decreasing  $NO_3^-$  levels in schemes D and G (i.e., both  $NO_3^-$  and  $NO_2^-$  were present simultaneously) was higher than the rate observed in scheme A. Additionally, the  $NO_2^-$  level remained almost constant at ca. 25 mg-N/L, indicating that the reduction of  $NO_3^-$  to  $NO_2^-$  occurred at a rate approximately equal to the  $NO_2^-$  consumption rate. In scheme E (i.e.,  $NO_3^-+N_2O$ ),  $NO_3^-$  and  $N_2O$  concentrations decreased by 8.5 and 3.5 mg-N/L, respectively. While, the  $N_2O$  level increased by ca. 2.8 mg-N/L in scheme F (i.e.,  $NO_2^-+N_2O$ ).

In Fig. 3A, the net  $NO_3^-$ ,  $NO_2^-$ , and  $N_2O$  reduction rates were 0.84, 2.25, and 1.66 mg N/g VSS/h, respectively. In schemes D and G (i.e.,  $NO_3^-$  and  $NO_2^-$  simultaneously existed), the  $NO_3^-$  reduction rate was obviously the highest (i.e., 2.58 and 2.62 mg/g VSS/h, respectively). In schemes F and G (i.e.,  $NO_2^-$  and  $N_2O$  simultaneously existed), the  $NO_2^-$  reduction rate (3.76 and 2.6 mg/g VSS/h, respectively) was apparently higher than the  $N_2O$  reduction rate (2.81 and 1.35 mg/g VSS/h, respectively).

In Fig. 3B, the electron consumption rates of each reductase corresponding to  $NO_x$  reduction rates were calculated (Pan et al., 2019). During scheme A,  $NO_2^-$ , NO, and  $N_2O$  were formed as denitrifying intermediates. The highest electron consumption rates were observed for *Nar* (0.11 mmol e/g VSS/h) in scheme A, *Nir* (0.16 mmol e/g VSS/h) in scheme B, and *Nos* (0.1 mmol e/g VSS/h) in scheme C. This result indicated that electron competition inevitably occurred in all tests when the potential electron consumption rate (PEC) exceeded the maximum electron supply rate (MES) according to previous study. The total electron consumption rate in the multiple  $NO_x$  schemes was significantly higher than that in the sole  $NO_x$  schemes. It was in agreement with the results observed in previous studies of both forward-diffusion and counter-diffusion biofilm systems (Pan et al., 2019; Roy et al., 2021), suggesting that the PCL-driven SPD system consistently and steadily provided sufficient carbon sources for  $NO_x$  reduction. However, the  $N_2O$  reduction rate and the electron consumption rate of *Nos* were consistently the lowest. This was likely due to electron competition between the upstream electron pool (i.e., the quinol pool, where electrons were directly utilized by *Nar*) and the downstream electron pools (i.e., periplasmic small c-type cytochromes, which accepted electrons from the quinol pool and then transfer them to *Nir* and *Nos* via the bc1 complex) (Liu et al., 2025), electrons diffusing from inner to outer layer allocated to *Nos* was the least.

Fig. 3C illustrated the electron distribution patterns among *Nar*, *Nir*, and *Nos* in schemes D, E, F and G in the PCL-driven SPD system. Obviously, electrons consistently tended to flow toward *Nar* (35.1–48.5 %),

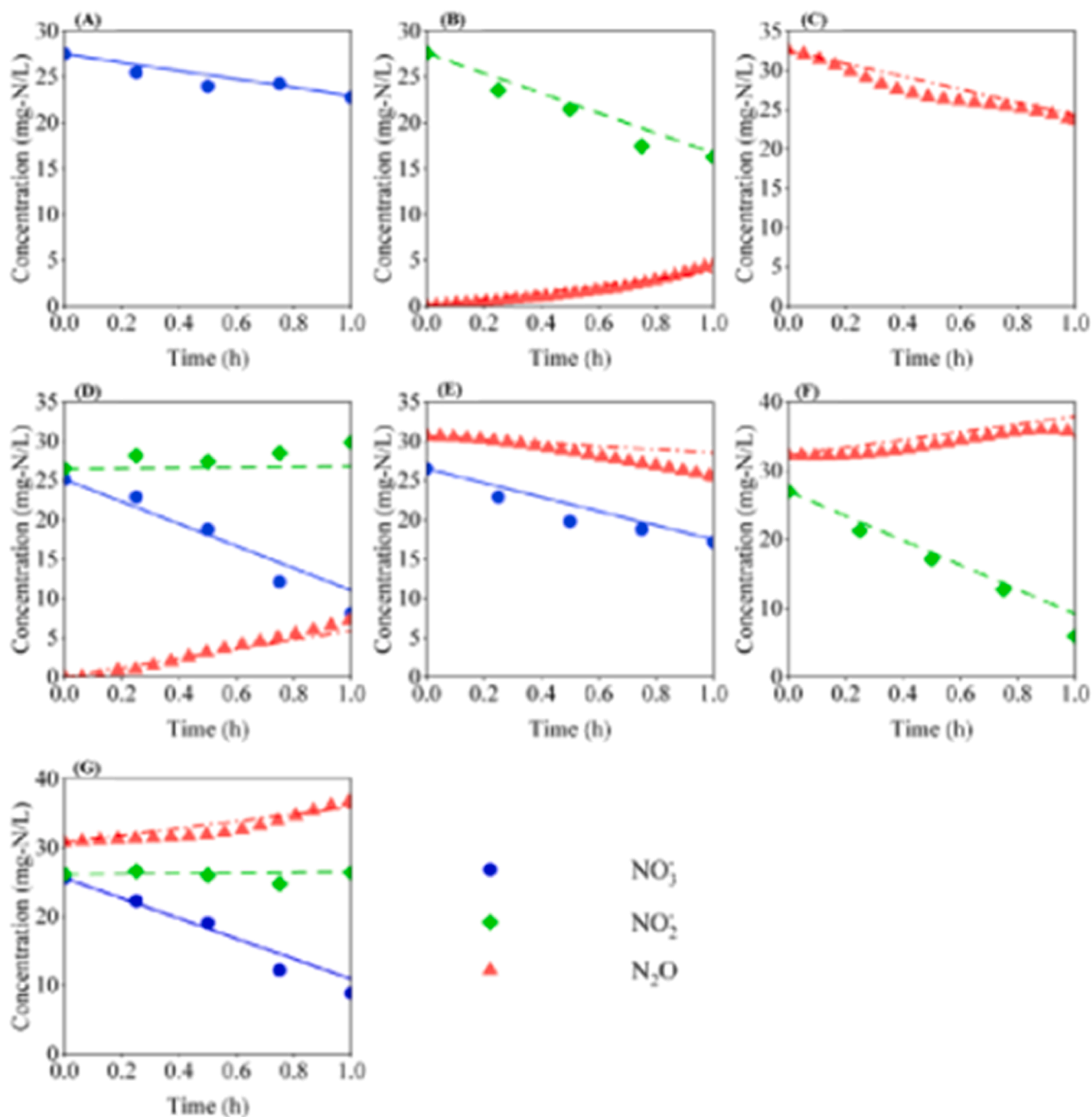


Fig. 2.  $\text{NO}_3^-$ ,  $\text{NO}_2^-$  and  $\text{N}_2\text{O}$  profiles during electron competition experiment from the PCL-supported biofilm denitrification system.

confirming the above results (Fig. 3A and 3B) that electrons preferentially flowed toward the upstream electron pool. Compared to schemes E and F, more electrons flowed toward *Nar* in scheme G, indicating electron competition between the upstream and downstream electron pools. Overall, electron competition between the two electron pools was the primary reason for the lower  $\text{N}_2\text{O}$  reduction rate in the PCL-driven SPD system.

### 2.3. $\text{NO}_x$ reduction and $\text{N}_2\text{O}$ accumulation in the PCL-driven spd system

Figs. 4–6 demonstrated the variations of  $\text{NO}_x$  levels during the 2–4 h batch tests in 11 schemes, which contained sole and multiple  $\text{NO}_x$  addition schemes (i.e., A–G) with each  $\text{NO}_x$  concentration of 30 mg-N/L,

and multiple  $\text{NO}_x$  addition schemes (i.e., H–K) with TN concentration of 30 mg-N/L (Table 1). In Fig. 4, the concentrations of  $\text{NO}_3^-$ ,  $\text{NO}_2^-$ , or  $\text{N}_2\text{O}$  decreased by 21.2, 25.3 and 29.9 mg-N/L within 4.0 h, 2.75 h and 3.5 h, respectively. The reduction rates were consistent with the results from the first hour (Fig. 2 and 3A). In Fig. 4B, the  $\text{N}_2\text{O}$  concentration initially showed an increasing trend, followed by a decreasing trend at 2.75 h, and the maximum accumulation  $\text{N}_2\text{O}$  concentration exceeding 10 mg-N/L. This indicated that electrons were utilized by *Nos* for the  $\text{N}_2\text{O}$  reduction as  $\text{NO}_2^-$  was consumed, further highlighting the electron competition between downstream electron pools (Fig. 3C). Similar  $\text{N}_2\text{O}$  accumulation has been observed in a previous powdered degradable microplastic-activated sludge system (He et al., 2024a). The possible reason was that PCL may significantly reduce the relative abundance of

**Table 1**

Electron acceptors addition scheme.

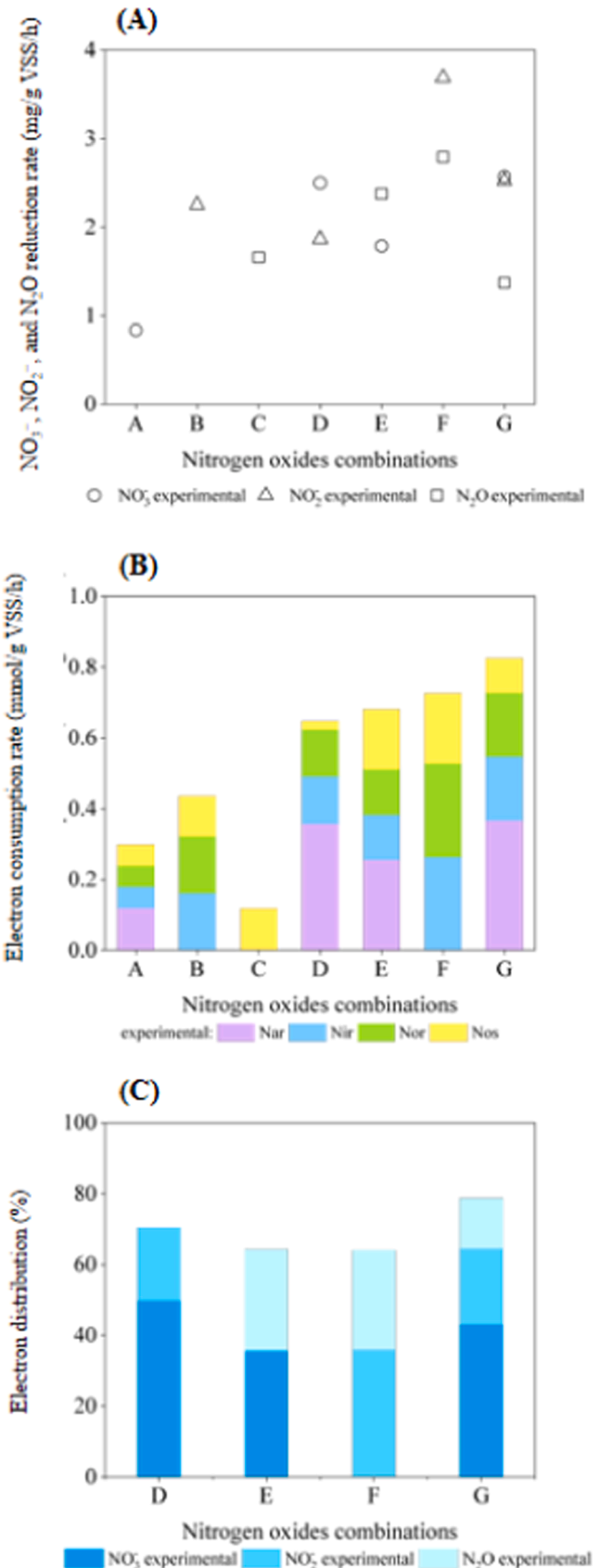
Data set 1	A	B	C	
Electron acceptor added	NO <sub>3</sub>	NO <sub>2</sub>	N <sub>2</sub> O	
Added concentration	30	30	30	
Total NOx concentration	30	30	30	
Data set 2	D	E	F	G
Electron acceptor added	NO <sub>3</sub> +NO <sub>2</sub>	NO <sub>3</sub> +N <sub>2</sub> O	NO <sub>2</sub> +N <sub>2</sub> O	NO <sub>3</sub> +NO <sub>2</sub> +N <sub>2</sub> O
Added concentration	30+30	30+30	30+30	30+30+30
Total NOx concentration	60	60	60	90
Data set 3	H	I	J	K
Electron acceptor added	NO <sub>3</sub> +NO <sub>2</sub>	NO <sub>3</sub> +N <sub>2</sub> O	NO <sub>2</sub> +N <sub>2</sub> O	NO <sub>3</sub> +NO <sub>2</sub> +N <sub>2</sub> O
Added concentration	15+15	15+15	15+15	10+10+10
Total NOx concentration	30	30	30	30

Concentration unit: mg-N/L.

denitrifying bacteria and key genes involved in the electron transfer system, which could be related to significant variations in extracellular polymeric substance composition.

From Fig. 5, NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup> and N<sub>2</sub>O exhibited a distinct reduction sequence, i.e., electrons were first utilized by NO<sub>3</sub><sup>-</sup> for its reduction and then NO<sub>2</sub><sup>-</sup> levels showed a decreasing trend as NO<sub>3</sub><sup>-</sup> was nearly depleted, followed by N<sub>2</sub>O. On top of this phenomenon, NO<sub>2</sub><sup>-</sup> represented an apparent accumulation during the degradation of NO<sub>3</sub><sup>-</sup>, as was the relationship between NO<sub>2</sub><sup>-</sup> and N<sub>2</sub>O. Especially, the presence of NO<sub>2</sub><sup>-</sup> caused N<sub>2</sub>O to initially accumulate at the start of the test (i.e., 0–1 h and 0–3 h in Fig. 5C and 5D, respectively), followed by its subsequent consumption (i.e., 1–3 h and 3–4 h in Fig. 5C and 5D, respectively). This was inconsistent with the results observed in previous co-diffusion biofilm systems (Pan et al., 2019), which may be attributed to the counter-diffusion structure of the PCL-driven SPD system. In counter-diffusion biofilms (Chen et al., 2019), the flow directions of NO<sub>x</sub> and carbon sources were opposite, causing a delay in the flow of electrons from the upstream electron pool to the downstream electron pools, further hindering the flow of electrons toward *Nir* and *Nos*, subsequently slowing down the NO<sub>2</sub><sup>-</sup> and N<sub>2</sub>O reduction rate. It is worth noting that the addition of NO<sub>2</sub><sup>-</sup> and/or N<sub>2</sub>O at the same concentration accelerated the NO<sub>3</sub><sup>-</sup> reduction rate, which indicated that the addition of various NO<sub>x</sub> promoted the utilization of PCL by microorganisms (Zhang et al., 2022a), thus releasing more electrons and increasing the NO<sub>3</sub><sup>-</sup> reduction rate (Liu et al., 2019). Similarly, the presence of NO<sub>2</sub><sup>-</sup> with N<sub>2</sub>O also increased the NO<sub>2</sub><sup>-</sup> reduction rate. However, the NO<sub>2</sub><sup>-</sup> reduction rate was reduced when NO<sub>3</sub><sup>-</sup> was present simultaneously.

Compared to Fig. 5 (i.e., schemes d–f), in Fig. 6 (i.e., schemes H–K), the reaction time was reduced to within 1.5–2 h after the concentration of each NO<sub>x</sub> profiles in each scheme decreased by half. Similarly, no significant change of NO<sub>x</sub> reduction sequence was observed (Fig. 6), indicating that the decreasing N<sub>2</sub>O reduction rate was mainly attributed to the electron competition between the upstream and downstream electron pools within the counter-diffusion biofilm, rather than the diffusion rate of NO<sub>x</sub> from outer to inner layer in the aqueous phase. In addition, under the condition of the same concentration of total NO<sub>x</sub>-N, the total nitrogen removal duration of single NO<sub>x</sub> schemes was 1.5–2 times as long as that of multiple NO<sub>x</sub> schemes. This may be because even if the total NO<sub>x</sub> demonstrated the same level (30 mg-N/L), multiple NO<sub>x</sub> types can still stimulate the PCL to release more electrons, thereby simultaneously performing the reduction reaction. This result was different from that of biofilm systems that initially feed sufficient carbon sources, because PCL-driven SPD systems can continuously supply sufficient carbon sources from the inner layer of the biofilm (Zhang et al., 2022a). Therefore, PCL-driven SPD system can better solve the dilemma



**Fig. 3.** NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup> and N<sub>2</sub>O net reduction rates (A), electron consumption rates by *Nar*, *Nir*, *Nor*, and *Nos* (B), percentages of electrons distributed to the added electron acceptors under addition schemes D, E, F, G (C).

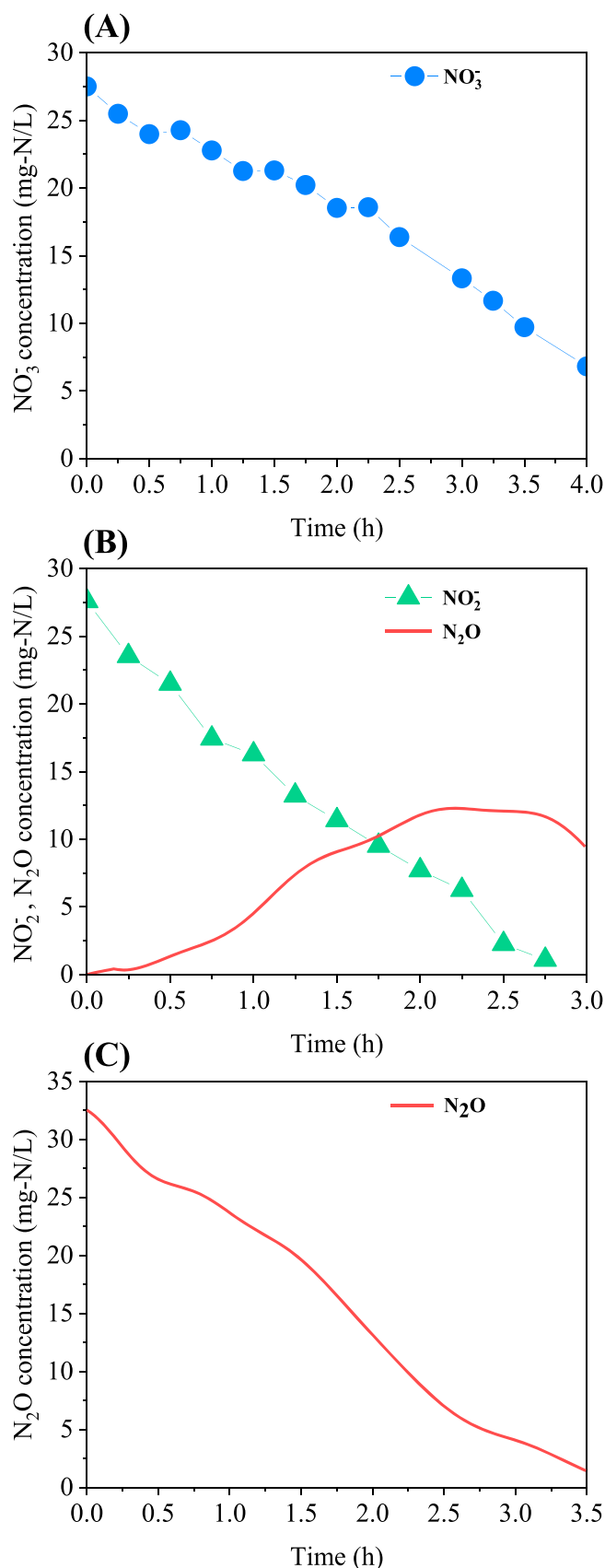


Fig. 4. The variations of NO<sub>3</sub><sup>-</sup>, NO<sub>2</sub><sup>-</sup> and N<sub>2</sub>O concentrations in schemes A (A), B (B) and C (C).

of decreasing NO<sub>x</sub> reduction rate caused by electron competition and insufficient carbon source in multiple NO<sub>x</sub> schemes. However, the N<sub>2</sub>O accumulation cannot be avoided simultaneously. For instance, under the condition of the total NO<sub>x</sub> level at 30 mg-N/L, the maximum N<sub>2</sub>O accumulation in schemes H and K reached 12.3 and 14.2 mg-N/L, respectively, primarily due to the inevitable NO<sub>2</sub><sup>-</sup> accumulation. Fortunately, at the 2 h, the N<sub>2</sub>O concentrations in both schemes H and K decreased to below 2.5 mg-N/L, whereas in scheme C (i.e., sole N<sub>2</sub>O addition), the N<sub>2</sub>O concentration remained at 12.8 mg-N/L. These results clearly demonstrated that the PCL-driven SPD system was effective in alleviating N<sub>2</sub>O emissions for treating wastewater containing multiple NO<sub>x</sub>. At WWTPs, the actual municipal wastewater was often discharged in the form of multiple NO<sub>x</sub> simultaneously (van Dijk et al., 2021). Therefore, PCL-driven SPD systems were advantageous in reducing total nitrogen removal.

#### 2.4. Microbial analysis

The surface sludge of PCL-driven biofilm on day 85 (R1) and day 180 (R2) were obtained and the microbial communities were analyzed. The relative abundance of bacterial populations at the phylum level was characterized in Fig. S1, the main microbial group with N<sub>2</sub>O emission potential was *Proteobacteria* with 58.39 % and 40.86 % in R1 and R2, respectively, followed by *Bacteroidota* with 22.46 % and 33.31 % in R1 and R2, respectively. 33.31 % in R1 and R2, respectively. *Proteobacteria* was dominant in the whole community and its important role in organic matter and denitrification (Shen et al., 2013). *Bacteroidota* can play a role in the decomposition of macromolecules in wastewater treatment (Shen et al., 2020), and their coexistence in the system contributes to the hydrolysis of PCL. According to the release mechanism of solid carbon sources, hydrolytic enzymes were required to first break down macromolecules into small molecules that can be utilized by microorganisms, and the hydrolysis process was also considered as an important rate-limiting step. At the genus level, the dominant taxon was *unclassified\_f Comamonadaceae* with 46.39 % and 31.41 % in R1 and R2, respectively (Fig. S2). *Comamonadaceae* were commonly detected in solid phase denitrification systems responsible for polymer degradation, and were the main PCL degrading denitrifying bacteria with the ability to reduce NO<sub>3</sub><sup>-</sup> to NO<sub>2</sub><sup>-</sup> (Wu et al., 2013). These results indicated that the carbon-nitrogen counter-diffusion biofilm system maintained stable carbon source release and nitrogen conversion states, providing a stable microbial environment that ensured the reliability of biochemical tests results.

### 3. Conclusion

In the carbon and nitrogen counter-diffusion biofilm system, the NO<sub>3</sub><sup>-</sup> reduction rate was the fastest in the multiple NO<sub>x</sub> schemes, where the total electron consumption rate was significantly higher than in the sole NO<sub>x</sub> addition schemes. The fiercer electron competition was obtained between the upstream and downstream electron pool, resulting in inevitable N<sub>2</sub>O accumulations. The NO<sub>x</sub> sequential reduction (NO<sub>3</sub><sup>-</sup> → NO<sub>2</sub><sup>-</sup> → N<sub>2</sub>O) was clearly observed under different nitrogen loading conditions, which further indicated that carbon release from solid carbon sources was a non-limiting factor. The *Comamonadaceae* was the dominant species at the genus level for denitrification.

### 4. Materials and methods

#### 4.1. Reactor startup and continuous operation

PCL stands out as a promising biodegradable polymer due to its considerable carbon source release rate and denitrification efficiency as well as high bioavailability (Zhang et al., 2022a). Therefore, PCL was selected as biofilm carriers and external electron donors in this study. PCL exhibited a spherical pellets shape with a diameter of ca. 3 mm, a



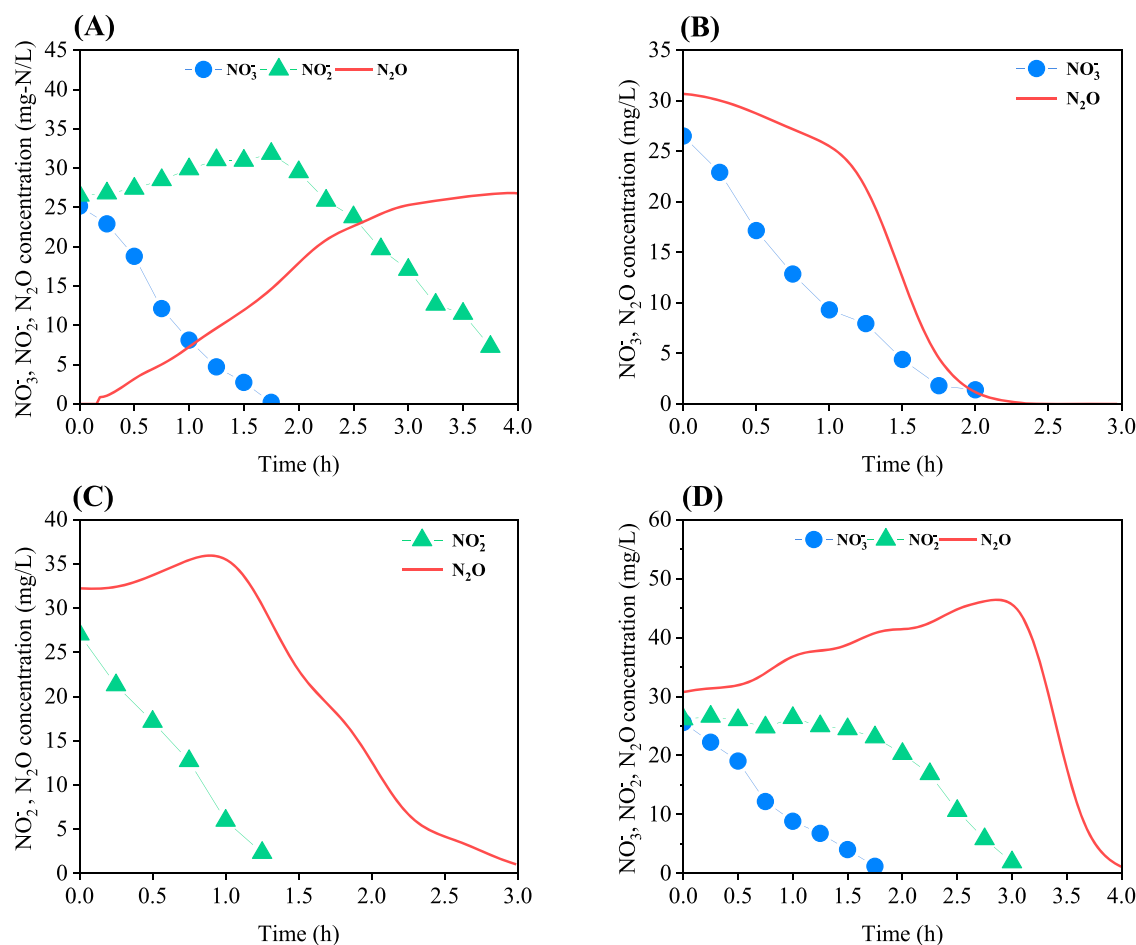


Fig. 5. The variations of  $\text{NO}_3^-$ ,  $\text{NO}_2^-$  and  $\text{N}_2\text{O}$  concentrations in schemes D (A), E (B), F (C) and G (D).

bulk density of 0.78 g/mL, a real density of 1.15 g/mL, and an average molecular weight of 50,000 Da. A laboratory-scale up-flow SPD cylindrical reactor was operated at 23–25 °C room temperature. The reactor was made of polyvinyl chloride, 36 cm in height, 6 cm in inner diameter, and 1.0 L in effective volume.

The reactor was packed to a filling ratio of 75 % with PCL pellets and inoculated with activated sludge from a secondary sedimentation tank. The reactor was wrapped in sunshade cloth to eliminate the possible proliferation of algae (Chu and Wang, 2013). The HRT of reactor was 3 h, and the influent pH was approximately controlled at 7.5–8. The reactor was continuously loaded with synthetic wastewater containing 50 mg-N/L  $\text{NO}_3^-$ . The composition and proportion of synthetic wastewater and trace elements were consistent with previous studies and presented in Table S1 (Zhang et al., 2022b). The reactor was operated for more than six months.

## 4.2. Batch tests

### 4.2.1. Batch reactor setup

After over six months of operation, stable  $\text{NO}_3^-$  removal efficiency and microbial biomass were obtained in the reactor, and a dense biofilm was formed on the surface of PCL pellets. Before batch tests, to eliminate the effects of  $\text{NO}_x$  compounds and organic carbon that may remain in the reactor, it is necessary to stop the continuous feed and drain the liquid from the reactor in advance as well as wash the biofilm and PCL pellets 2–3 times. Subsequently, the reactor was modified to an internal circulation mode. To be specific, a peristaltic pump was used to connect the reactor outlet to the inlet during a batch test, with the nitrogen-containing wastewater in the reactor internally circulated from bottom

to top. Finally, the synthesis wastewater was injected into the reactor until no headspace was left. The entire batch tests lasted for approximately 3–4 h.

### 4.2.2. Batch test operation

In order to provide the reactor with anoxic conditions in all batch tests, the mixed liquor was sprayed with  $\text{N}_2$  beforehand up to 20 min. The reactor was sealed to ensure that  $\text{N}_2\text{O}$  remained in the liquid phase. According to the addition scheme of nitrogen compounds in Table S2,  $\text{NO}_3^-$  and  $\text{NO}_2^-$  stock solutions were added to obtain various target  $\text{NO}_x$  concentrations of the mixture. The  $\text{N}_2\text{O}$  stock solution (i.e., 300 mg N/L) was freshly prepared through sparging Milli-Q water with 45 %  $\text{N}_2\text{O}$  gas for 10 min. Mixed liquor samples were collected from approximately the middle part of the reactor every 15 min by a syringe, and instantly filtered through a 0.22  $\mu\text{m}$  filter and stored at 4 °C for  $\text{NO}_3^-$  and  $\text{NO}_2^-$  analysis (Yang et al., 2022). An online  $\text{N}_2\text{O}$  microsensor ( $\text{N}_2\text{O}$ -R, Unisense A/S, Aarhus, Denmark) was used to monitor liquid-phase  $\text{N}_2\text{O}$  constantly. The microsensor was calibrated before each test. The pH of each batch was controlled within the optimal range for  $\text{NO}_x$  reduction (i.e.,  $8.0 \pm 0.1$ ) and regulated with 1 M HCl or 1 M NaOH solution (Pan et al., 2012). According to standard methods,  $\text{NO}_3^-$  and  $\text{NO}_2^-$  concentrations were examined at the end of experiments (APHA, 1998).

### 4.2.3. Batch test design

11 combinations of electron compounds addition schemes (i.e., schemes A–K) were carried out in Table S2. To better understand the impact of the electron competition on  $\text{N}_2\text{O}$  reduction and the electron preference for different nitrogen oxide reductases in the PCL-driven SPD system, each  $\text{NO}_x$  concentration was set to 30 mg-N/L in the sole or

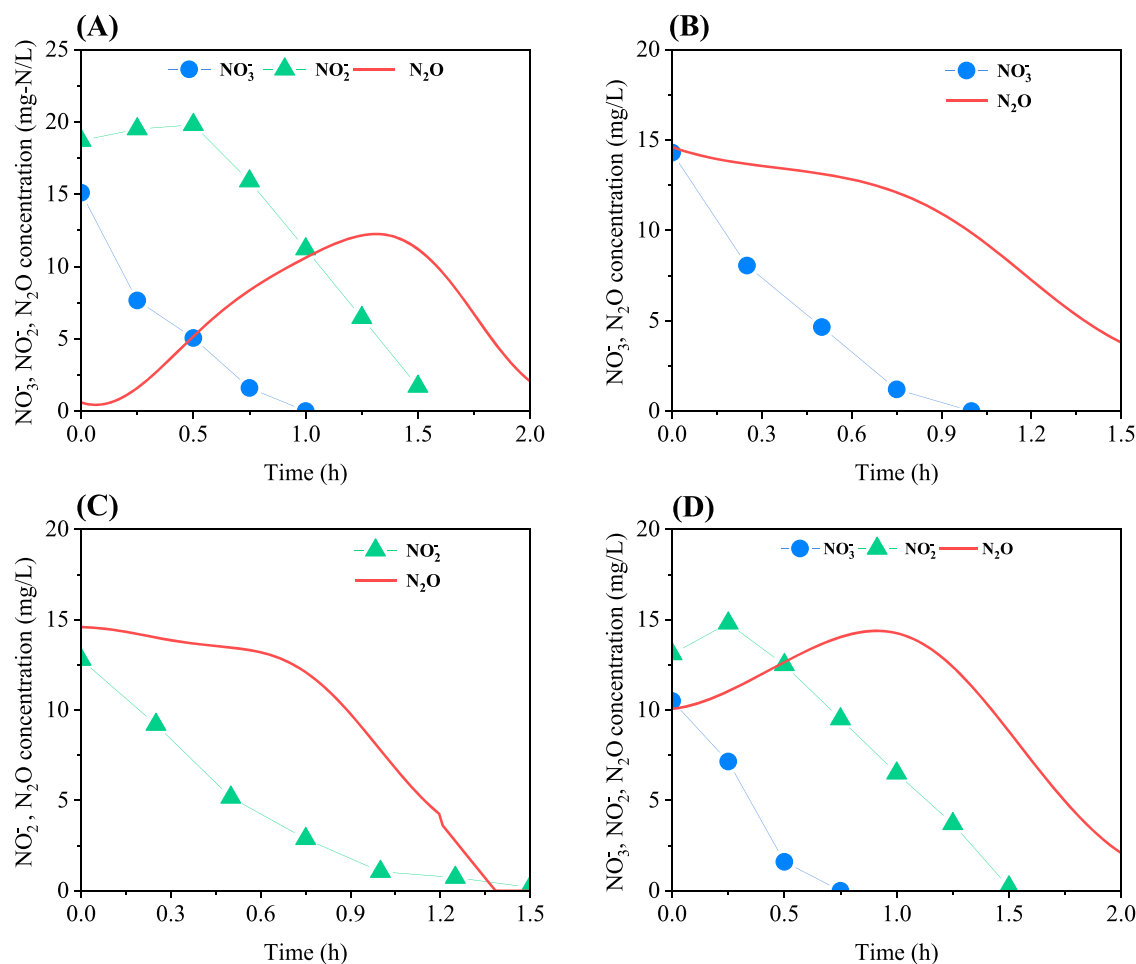


Fig. 6. The variations of  $\text{NO}_3^-$ ,  $\text{NO}_2^-$  and  $\text{N}_2\text{O}$  concentrations in schemes H (A), I (B), J (C) and K (D).

multiple  $\text{NO}_x$  schemes (i.e., A-G). The variation of  $\text{NO}_x$  concentration based on electron competition was obtained from the initial 1 h of batch tests in schemes A-G (Pan et al., 2013). In addition, to further investigated the  $\text{NO}_x$  reduction and  $\text{N}_2\text{O}$  accumulation in the PCL-driven SPD system, the total  $\text{NO}_x$  concentration in the multiple nitrogen combination schemes (i.e., H-K) was adjusted to 30 mg-N/L, which was similar with the  $\text{NO}_x$  concentration of schemes A-C. The variation of  $\text{NO}_x$  concentration was obtained from the 3–4 h batch tests.

#### 4.3. Microbial community analysis

After the biofilm on the surface of PCL particles was stabilized, a certain amount of film hanging particles was taken, and the biofilm was shed through ultrasonic treatment. The shed biofilm was collected in a centrifuge tube, concentrated and frozen for storage at  $-20^\circ\text{C}$ , and the microbial diversity operation and analysis were performed by 16S rRNA sequencing (Wang et al., 2024b; Wang et al., 2024c; Wang et al., 2022a). The main steps are as follows: firstly, DNA sequence extracted by Qubit ssDNA Assay using agarose gel (1 %) electrophoresis; secondly, ABI GeneAmp® 9700 PCR was used to perform PCR amplification of the V4 region of 16S rRNA gene, and the amplification primers were 515F (5' -GTGCCAGCMGCCGCGG-3') and 806R (5' -GGACTACHVGG GTWTCTAAT-3'); then TruSeq™ DNA Sample Prep toolbox was used to construct Miseq library for the sequences that met the requirements (Dai et al., 2024; Guo et al., 2023); finally, paired end sequencing analysis was performed on Illumina Miseq platform according to different sample sequencing requirements.

#### CRediT authorship contribution statement

**Yingrui Liu:** Writing – original draft, Resources, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Feng Chen:** Conceptualization. **Yanying He:** Data curation. **Yufen Wang:** Formal analysis. **Tingting Zhu:** Funding acquisition. **Yindong Tong:** Investigation. **Yingxin Zhao:** Methodology. **Bing-Jie Ni:** Software. **Yiwen Liu:** Writing – review & editing, Funding acquisition.

#### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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#### Supplementary materials

Supplementary material associated with this article can be found, in the online version, at [doi:10.1016/j.wroa.2025.100306](https://doi.org/10.1016/j.wroa.2025.100306).

## Data availability

Data will be made available on request

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