

# Method for Evaluation of the Requirements of B-group Vitamins Using Tryptophan Metabolites in Human Urine

Katsumi Shibata, Junko Hirose and Tsutomu Fukuwatari

Department of Nutrition, School of Human Cultures, the University of Shiga Prefecture, Hikone, Shiga, Japan.

**ABSTRACT:** Tryptophan metabolism is directly involved with B-group vitamins such as vitamin B<sub>2</sub>, niacin, and vitamin B<sub>6</sub>, and indirectly with vitamin B<sub>1</sub> and pantothenic acid. We evaluated the validity of requirements of B-group vitamins set by the *Dietary Reference Intakes for the Japanese (DRI-J)*. We investigated the fate of dietary tryptophan in 10 Japanese adult men who ate the same diet based on *DRI-J* during a 4-week study. Vitamin mixtures were administered based on the amounts in the basal diet during weeks 2, 3, and 4. Daily urine samples were collected eight times (days 1 and 5 in each week). Administration of vitamin mixtures had no effect on tryptophan metabolites such as anthranilic acid, kynurenic acid, xanthurenic acid, 3-hydroxyanthranilic acid, and quinolinic acid within individuals. Surplus administration of B-group vitamins against *DRI-J* requirements did not elicit beneficial effects on tryptophan metabolism. Our findings supported the requirements of B-group vitamins set by the *DRI-J*.

**KEYWORDS:** tryptophan, quinolinic acid, human, urine, vitamin

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**CORRESPONDENCE:** kshibata@shc.usp.ac.jp

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## Introduction

The vitamin niacin can be synthesized from the essential amino acid tryptophan (Trp).<sup>1</sup> B-group vitamins are involved in the metabolism of Trp: the pyridoxal 5'-phosphate (PLP)-dependent enzyme kynureninase,<sup>2,3</sup> the flavin adenine dinucleotide-dependent enzyme kynurenine 3-monooxygenase (which needs the reduced form of nicotinamide adenine dinucleotide phosphate as a coenzyme),<sup>4,5</sup> and the PLP-dependent enzyme kynurenine aminotransferase<sup>6</sup> (Fig. 1).

After ingestion of Trp, riboflavin-deficient rats were shown to excrete abnormally high amounts of anthranilic acid (AnA) and kynurenic acid (KA).<sup>7,8</sup> The reduced flux from the conversion of kynurenine to 3-hydroxykynurenine gives rise to the formation of KA and AnA. Rats deficient in vitamin B<sub>6</sub> excrete abnormally large amounts of xanthurenic acid (XA).<sup>9</sup>

Based on the information detailed above, we developed a method to evaluate the requirements of B-group vitamins set by the *Dietary Reference Intakes for the Japanese (DRIs-J)* in 2010.<sup>10</sup> If the intake of B-group vitamins is lower than required, urinary excretion of Trp metabolites would be affected. We investigated the effect of a gradual increase in the intake of vitamins B<sub>2</sub>, B<sub>6</sub>, and other B-group vitamins on the metabolism of Trp to quinolinic acid (QA).

The method proposed in this manuscript is for an evaluation of the necessity of B-group vitamins based on the individual Trp metabolism ability, and therefore, is applied to evaluate those in DRIs of many countries.

## Methods

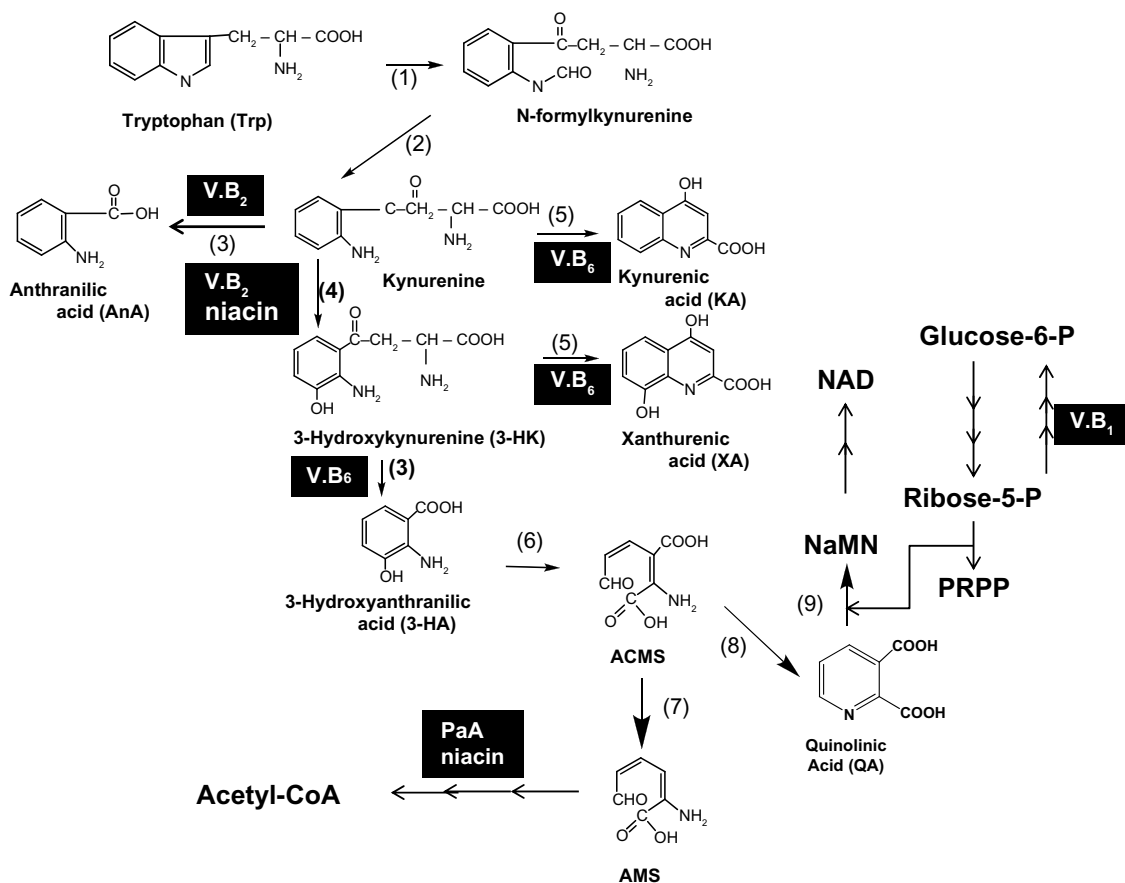
The study protocol was approved by the Ethics Committee of the University of Shiga Prefecture (Shiga, Japan) and was conducted according to the guidelines laid down in the Declaration of Helsinki. All subjects provided written informed consent to participate in the study after being informed of its protocol and purpose.

**Chemicals.** AnA and QA were purchased from Wako Pure Chemical Industries (Osaka, Japan). XA, KA, and 3-hydroxyanthranilic acid (3-HA) were purchased from Tokyo Chemical Industry (Tokyo, Japan).

**Subjects.** Male students and faculty members were recruited from the University of Shiga Prefecture. Participants diagnosed with a cold or influenza, and those who had taken multivitamin supplements at least once during the previous month, were excluded. All subjects were non-smokers and passed a standard medical examination at the university. Of the 12 apparently healthy male Japanese subjects who participated in this study, 10 subjects (age, 19–55 (mean ± standard deviation (SD), 26.8 ± 11.0) years) completed the study.

**Study design.** All subjects ( $n = 10$ ) were housed in the same facility and given the same diet. The height, body weight, and body mass index of the 10 subjects was 174.3 ± 4.2 cm, 66.5 ± 8.3 kg, and 21.9 ± 2.3 kg/m<sup>2</sup>, respectively. The experimental period was 4 weeks.

Breakfast consisted of bread (126 g), butter (7 g), ham (38 g), yoghurt (90 g), tomatoes (40 g), lettuce (40 g), and



**Figure 1.** Trp to QA and related pathways for B-group vitamins. ACMS,  $\alpha$ -amino- $\beta$ -carboxymuconate- $\epsilon$ -semialdehyde; AMS,  $\alpha$ -aminomuconate- $\epsilon$ -semialdehyde; NaMN, nicotinic acid mononucleotide; PRPP, 5-phosphoribosyl-1-pyrophosphate. (1) tryptophan 2,3-dioxygenase (TDO), (2) formylase, (3) kynureninase, (4) kynurenine 3-monoxygenase, (5) kynurenine aminotransferase, (6) 3-hydroxyanthranilic acid 3,4-dioxygenase (3-HADO), (7)  $\alpha$ -amino- $\beta$ -carboxymuconate- $\epsilon$ -semialdehyde decarboxylase, (8) non-enzymatic reaction (occurs spontaneously), (9) quinolinic acid phosphoribosyltransferase.

milk (200 mL). Lunch comprised rice (300 g), toasted and seasoned laver (1.4 g), luncheon meat (95 g), boiled egg (55 g), raw cabbage (50 g), mayonnaise (10 g), miso soup (miso, 10 g), and Japanese tea (200 mL). Dinner consisted of rice (300 g), soy sauce-flavored Pacific saury (70 g), tofu (soybean curd) (150 g), *katsuo-bushi* (1.5 g), spinach (leaves, boiled) (50 g), sesame seeds (1 g), soy sauce (6 g), kiwi fruit (50 g), and Japanese tea (200 mL). Cheese (25 g) and jelly fruit mix (200 g) was given as a midnight snack. Nutrient elements are shown in Table 1.

Subjects were allowed to drink natural mineral water freely (*Asahi Oisii Mizu Rokko*, Asahi Soft Drinks, Tokyo, Japan). Nutrients were calculated using the *Standard Tables of Food Composition in Japan (2010)*.<sup>11</sup> Only water-soluble vitamins in the food were assessed by us.

Subjects consumed the diet from day-1 to day-5 each week. However, the dietary regimen was relaxed on day-6 and day-7 of each week. This period of 5 days was considered to permit attainment of a steady nutritional status for these vitamins.<sup>12,13</sup> Approximately one-fold, three-fold, and seven-fold amounts of synthesized water-soluble vitamin mixtures

as vitamin mixtures “ $\alpha$ ”, “ $\beta$ ”, and “ $\gamma$ ” as shown in the *DRI-J (2010)*<sup>10</sup> were made (Table 2). Participants were given only the diet for the first week, the diet with vitamin mixture- $\alpha$  for the second week, the diet with vitamin mixture- $\beta$  for the third week, and the diet with vitamin mixture- $\gamma$  for the fourth week. One-third of the dose was put into a small gelatinous capsule, and the capsule administered after breakfast, lunch, and dinner (ie, three times daily).

Twenty four-hour urine samples were collected on day-1 and day-5 of each week. That is, urine samples were collected on a total of eight occasions (days 1, 5, 8, 12, 15, 19, 22, and 26).

**Analyses.** Trp metabolites KA,<sup>14</sup> AnA,<sup>15</sup> XA,<sup>16</sup> 3-HA,<sup>16</sup> and QA<sup>17</sup> were measured as described previously. In briefly, for the measurements of these compounds in urine, the acidified urine sample was passed through a 0.45- $\mu$ m microfilter. The filtrate (20  $\mu$ L) was injected directly into the respective HPLC systems.

**Statistical analyses.** Nonparametric Friedman test for repeated measures following Dunn’s post test was used to analyze statistical differences. Intra- and inter-individual variations were calculated using analysis of variance. Pearson’s

**Table 1.** Composition of the diet fed to male Japanese subjects during the study.

	BREAKFAST	LUNCH	DINNER	SNACK	TOTAL
Energy, <i>kcal</i>	694	977	830	225	2,726
Energy, <i>MJ</i>	2.90	4.08	3.47	1.07	11.39
Protein, <i>g</i>	28.6	30.7	31.7	10.3	101.2
Fat, <i>g</i>	26.9	38.2	19.5	6.5	91.1
Carbohydrate, <i>g</i>	84.7	123.3	127.8	30.9	366.7
<b>Vitamins</b>					
Vitamin A, $\mu\text{gRAE}^1$	152	166	271	65	653
Vitamin D, $\mu\text{g}$	0.9	1.1	9.2	0.0	11.1
Vitamin E, <i>mg</i>	1.66	2.26	4.50	0.28	8.69
Vitamin K, $\mu\text{g}$	21	81	158	1	260
Vitamin B <sub>1</sub> , <i>mg</i>	0.47 (0.37)	0.35 (0.34)	0.26 (0.50)	0.05 (0.05)	1.12 (1.26)
Vitamin B <sub>2</sub> , <i>mg</i>	0.56 (0.45)	0.73 (0.50)	0.33 (0.29)	0.10 (0.05)	1.72 (1.29)
Vitamin B <sub>6</sub> , <i>mg</i>	0.27 (0.23)	0.33 (0.23)	0.50 (0.36)	0.02 (0.02)	1.12 (0.84)
Vitamin B <sub>12</sub> , $\mu\text{g}$	1.0 (1.01)	2.7 (3.11)	8.8 (5.60)	0.8 (0)	13.3 (9.72)
Niacin, $\text{mgNE}^2$	9.8 (10.3)	9.2 (10.0)	10.5 (14.7)	2.0 (1.3)	31.5 (36.3)
Pantothenic acid, <i>mg</i>	2.4 (2.8)	2.6 (2.4)	1.5 (1.4)	0.1 (0.2)	6.6 (6.8)
Folate, $\mu\text{g}$	95 (98)	126 (81)	132 (97)	14 (10)	367 (286)
Biotin, $\mu\text{g}$	10.8 (15.6)	18.4 (14.5)	4.8 (9.0)	0.0 (1.7)	34.0 (42.4)
Vitamin C, <i>mg</i>	36 (22)	37 (34)	45 (45)	7 (43)	125 (144)
<b>Minerals</b>					
Na, <i>mg</i>	1,190	1,777	741	289	3,998
K, <i>mg</i>	872	556	856	123	2,406
Ca, <i>mg</i>	382	148	335	164	1,028
Mg, <i>mg</i>	72	85	148	11	315
P, <i>mg</i>	531	470	533	195	1,728
Fe, <i>mg</i>	1.2	3.9	3.7	0.3	9.2
Zn, <i>mg</i>	2.8	4.4	3.9	0.8	11.9
Cu, <i>mg</i>	0.23	0.54	0.77	0.02	1.56
Mn, $\mu\text{g}$	0.39	1.53	1.82	0.02	3.76
I, $\mu\text{g}$	51	94	1	0	146
Se, $\mu\text{g}$	39	24	10	0	74
Cr, $\mu\text{g}$	1	1	1	0	3
Mo, $\mu\text{g}$	36	104	95	0	235

**Notes:** Values were calculated from the *Standard Tables of Food Composition in Japan (2010)*. Parenthesis numbers in water-soluble vitamins were measured by us. <sup>1</sup>RAE = retinol activity equivalent. <sup>2</sup>NE (niacin equivalent was calculated based on the assumption that 1 mg of nicotinamide can be synthesized from 60 mg of Trp, and that 100 g of protein contains 1.0 g of Trp).

correlation coefficients were calculated to determine the association between experimental days and the concentration of each compound detected in urine.  $P < 0.05$  was considered significant. Prism 5.0 (GraphPad, San Diego, CA, USA) was used for statistical analyses.

## Results

### Extent of urinary excretion of Trp metabolites.

Figure 2 shows the mean changes in urinary excretion of AnA, KA, XA, 3-HA, and QA. Administration of a vitamin mixture did not affect urinary excretion of Trp metabolites.

Urinary excretion amounts (mean  $\pm$  SD,  $n = 80$ ) of AnA, KA, XA, 3-HA, and QA were  $1.7 \pm 0.8$ ,  $11 \pm 2.4$ ,  $3.7 \pm 0.8$ ,  $6.6 \pm 1.9$ , and  $17 \pm 4.6$   $\mu\text{mol/day}$ , respectively. Figure 3 shows the individual changes in urinary excretion of Trp metabolites. Table 3 shows the individual mean daily values of L-Trp metabolites collected eight times during the experimental period. These urine excretion amounts were observed differences among subjects. Intra-individual coefficient of variation (CV) of AnA, KA, 3-HA, and QA was 7, 18, 15, 17, and 49%, respectively. Inter-individual CV of AnA, KA, XA, 3-HA, and QA was 55, 80, 35, 61, and

**Table 2.** Composition of the vitamin mixture administered during weeks 2, 3, and 4.

	WEEK 2 VITAMIN MIXTURE “α”	WEEK 3 VITAMIN MIXTURE “β”	WEEK 4 VITAMIN MIXTURE “γ”
Thiamin <sup>1</sup> , mg mg as dietary vitamin B <sub>1</sub> *	1.4 2.4	4.2 7.0	8.4 13.9
Riboflavin, mg mg as dietary vitamin B <sub>2</sub> *	1.6 2.4	4.8 7.2	9.6 14.4
Pyridoxine <sup>2</sup> , mg mg as dietary vitamin B <sub>6</sub> *	1.4 2.0	4.2 5.9	8.4 11.8
Cyanocobalamin, μg μg as dietary vitamin B <sub>12</sub> *	2.4 4.8	7.2 14.4	14.4 28.8
Nicotinamide, mg mg as dietary niacin*	15 25.5	45 76.5	90 153
Pantothenic acid <sup>3</sup> , mg mg as dietary pantothenic acid*	6 8.4	18 25.2	36 50.4
Pteroylmonoglutamic acid, mg mg as dietary folate*	0.24 0.48	0.72 1.44	1.44 2.88
Biotin, μg μg as dietary biotin*	50 65	150 195	300 390
Ascorbic acid, mg mg as dietary vitamin C*	100 100	300 300	600 600

**Notes:** <sup>1</sup>Thiamin hydrochloride was used and the value expressed as thiamin itself. <sup>2</sup>Pyridoxine hydrochloride was used and the value expressed as pyridoxine itself. <sup>3</sup>Ca pantothenate was used and the value expressed as pantothenic acid itself. \*Relative biological values of synthesized vitamins against dietary vitamins are used: vitamin B<sub>1</sub>, 1.7; vitamin B<sub>2</sub>, 1.5; vitamin B<sub>6</sub>, 1.4; niacin, 1.7; pantothenic acid, 1.4; vitamin B<sub>12</sub>, 2.0; folate, 2.0; biotin, 1.3; vitamin C, 1. These values were taken from [39] and [40].

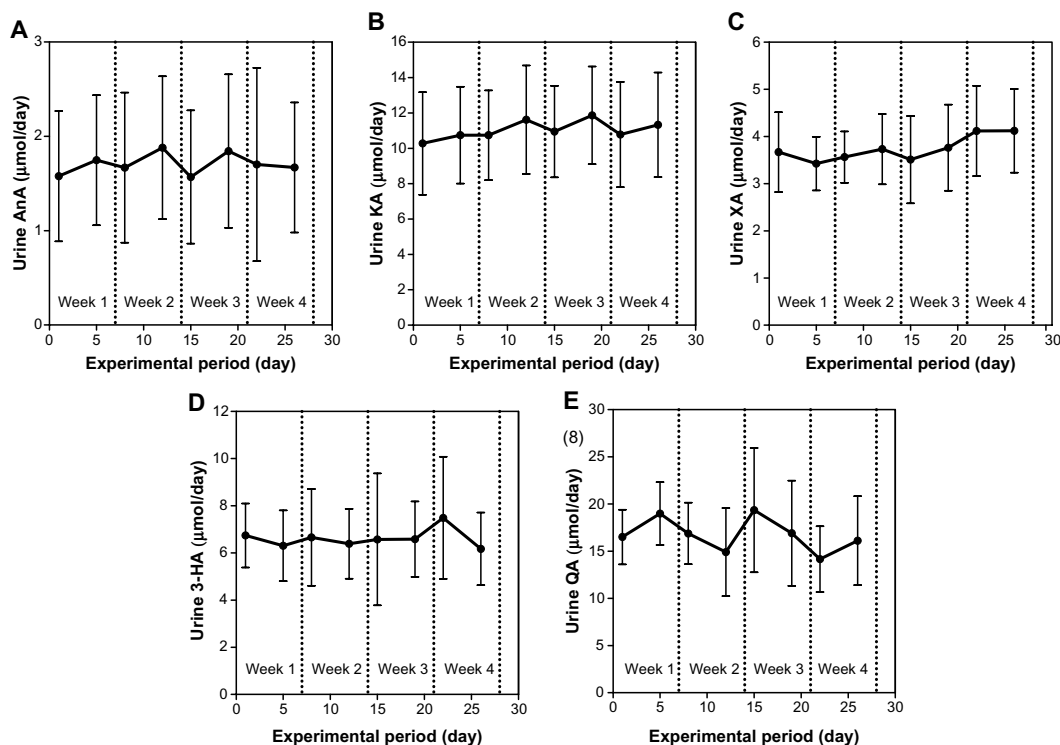
71%, respectively. Intra-individual CV of these metabolites was lower than the inter-individual CV. As information, we showed “Supplemental Table 1”, which was shown the values in terms of creatinine.

**Percentage urinary excretion of Trp metabolites.** Trp intake was calculated from the protein intake based on the assumption that Trp content in protein is 1%. In the present study, 100 g of protein was administered every day, so Trp intake was 1,000 mg/day or 5 mmol/day. Percentage urinary excretion (mean ± SD, n = 80) of AnA, KA, XA, 3-HA, and QA was 0.034 ± 0.015, 0.22 ± 0.055, 0.075 ± 0.016, 0.13 ± 0.038, and 0.38 ± 0.093% against the Trp intake, respectively. Table 4 shows the individual mean percentage urinary excretion. These were observed differences among subjects.

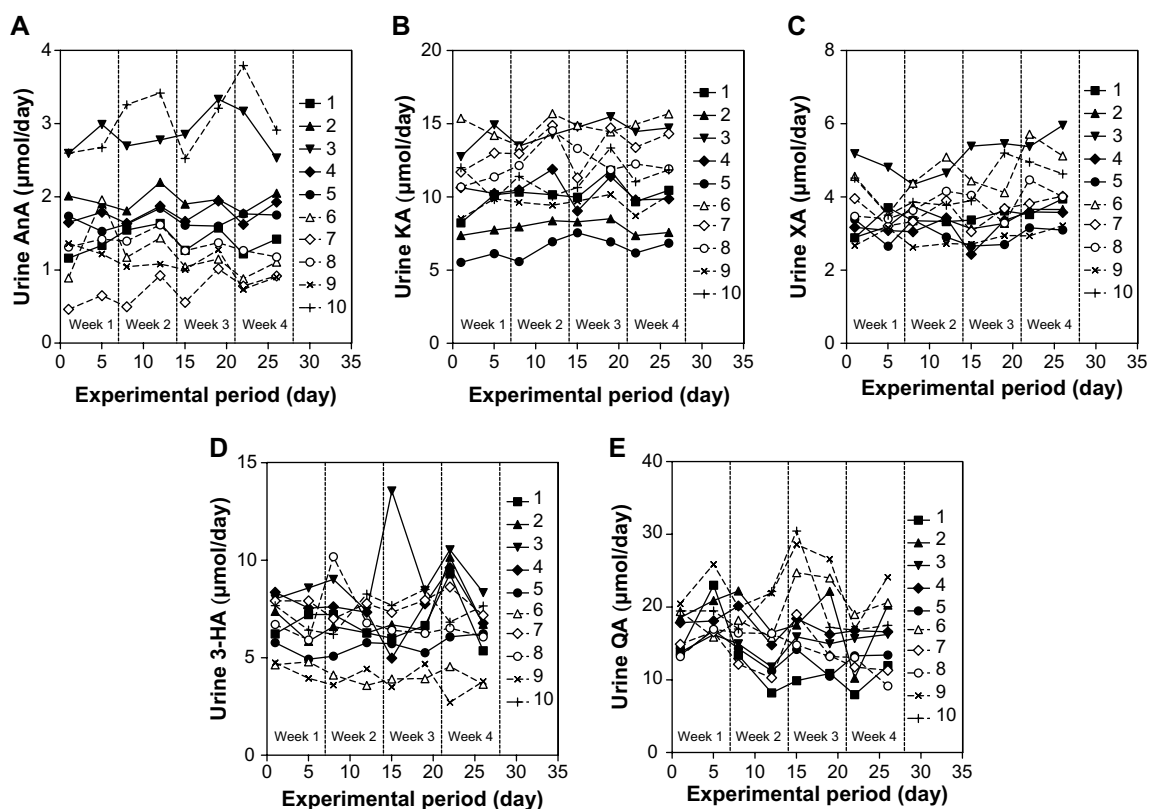
**Association between each metabolite of Trp.** Relationships between each metabolite of Trp were analyzed (Fig. 4). Direct proportional relationships were observed between AnA and 3-HA (Fig. 4C), KA and XA (Fig. 4E), and XA and 3-HA (Fig. 4H). An inverse proportional relationship was observed between 3-HA and QA (Fig. 4J).

## Discussion

The metabolism of the Trp–QA pathway is affected by female hormones,<sup>18–20</sup> so the present study was conducted in male Japanese subjects. It is known that rats deficient in vitamin B<sub>6</sub> excrete abnormally large amounts of XA.<sup>2</sup> Kynureninase is



**Figure 2.** Daily mean changes in urinary excretion of AnA (A), KA (B), XA (C), 3-HA (D), and QA (E) during the study. Each symbol represents the mean ± SD (n = 10). Week 1, diet only; week 2, diet with vitamin mixture “α” (two-fold); week 3, diet with vitamin mixture “β” (four-fold); week 4, diet with vitamin mixture “γ” (seven-fold).



**Figure 3.** Individual daily changes in urinary excretion of AnA (A), KA (B), XA (C), 3-HA (D), and QA (E) during the study. Each symbol represents the value for each subject. The numbers in figure indicate the subject number. Intra-individual coefficient of variation (CV) of AnA, KA, 3-HA, and QA was 7.4, 17.7, 15.1, 17.4, and 48.9%, respectively. Inter-individual CV of AnA, KA, XA, 3-HA, and QA was 43.2, 79.8, 35.0, 61.1, and 71.3%, respectively.

**Table 3.** Individual mean values of daily L-Trp metabolites collected eight times in male Japanese subjects.

SUBJECT NUMBER	AnA $\mu\text{mol/day}$	KA $\mu\text{mol/day}$	XA $\mu\text{mol/day}$	3-HA $\mu\text{mol/day}$	QA $\mu\text{mol/day}$
1	$1.40 \pm 0.16^{\text{bcd}}$	$10.10 \pm 0.98^{\text{bd}}$	$3.46 \pm 0.31^{\text{ad}}$	$6.78 \pm 1.19^{\text{ac}}$	$12.42 \pm 4.81^{\text{c}}$
2	$1.95 \pm 0.14^{\text{ab}}$	$7.90 \pm 0.46^{\text{bcd}}$	$3.37 \pm 0.30^{\text{bcd}}$	$7.01 \pm 1.34^{\text{ac}}$	$18.55 \pm 3.95^{\text{abc}}$
3	$2.87 \pm 0.28^{\text{a}}$	$14.35 \pm 0.86^{\text{a}}$	$5.14 \pm 0.52^{\text{a}}$	$9.23 \pm 1.96^{\text{a}}$	$14.95 \pm 1.52^{\text{bc}}$
4	$1.76 \pm 0.14^{\text{ac}}$	$10.42 \pm 0.91^{\text{ad}}$	$3.21 \pm 0.38^{\text{bcd}}$	$7.50 \pm 1.32^{\text{ab}}$	$17.37 \pm 1.62^{\text{abc}}$
5	$1.68 \pm 0.11^{\text{ad}}$	$6.46 \pm 0.72^{\text{d}}$	$2.89 \pm 0.31^{\text{d}}$	$5.60 \pm 0.47^{\text{bc}}$	$13.39 \pm 1.94^{\text{c}}$
6	$1.21 \pm 0.35^{\text{bcd}}$	$14.83 \pm 0.77^{\text{a}}$	$4.62 \pm 0.68^{\text{a}}$	$4.14 \pm 0.46^{\text{cd}}$	$19.70 \pm 3.32^{\text{abc}}$
7	$0.73 \pm 0.21^{\text{d}}$	$13.28 \pm 1.33^{\text{ab}}$	$3.62 \pm 0.37^{\text{ad}}$	$7.71 \pm 0.52^{\text{ab}}$	$13.65 \pm 2.96^{\text{bc}}$
8	$1.35 \pm 0.13^{\text{bcd}}$	$12.26 \pm 1.19^{\text{ac}}$	$3.81 \pm 0.42^{\text{ac}}$	$6.85 \pm 1.38^{\text{ac}}$	$14.14 \pm 2.57^{\text{bc}}$
9	$1.08 \pm 0.20^{\text{cd}}$	$9.51 \pm 0.61^{\text{bcd}}$	$2.87 \pm 0.23^{\text{cd}}$	$3.92 \pm 0.68^{\text{cd}}$	$23.07 \pm 3.91^{\text{a}}$
10	$3.05 \pm 0.45^{\text{a}}$	$11.27 \pm 1.15^{\text{ad}}$	$4.29 \pm 0.62^{\text{ab}}$	$7.40 \pm 0.84^{\text{ab}}$	$20.00 \pm 4.60^{\text{ab}}$
All subjects	$1.71 \pm 0.75$	$11.04 \pm 2.74$	$3.74 \pm 0.81$	$6.61 \pm 1.89$	$16.72 \pm 4.60$

**Notes:** Values are the mean  $\pm$  SD for L-Trp metabolites collected eight times for all subjects. The means without a common superscripted letter in a same column differ,  $P < 0.05$ , determined by one-way ANOVA followed by Nonparametric Friedman test for repeated measures following Dunn's post test. Values for all subjects are the mean  $\pm$  SD for 80 samples.

a PLP enzyme present in the soluble fraction.<sup>9</sup> Deficiency of vitamin B<sub>6</sub> reduces the activity of this enzyme. Kynurenine aminotransferase is also a PLP enzyme<sup>6</sup> but its activity is tolerant to deficiency of vitamin B<sub>6</sub> because the enzyme is localized in the inner membrane of mitochondria.<sup>3</sup> Thus, in vitamin B<sub>6</sub>-deficient

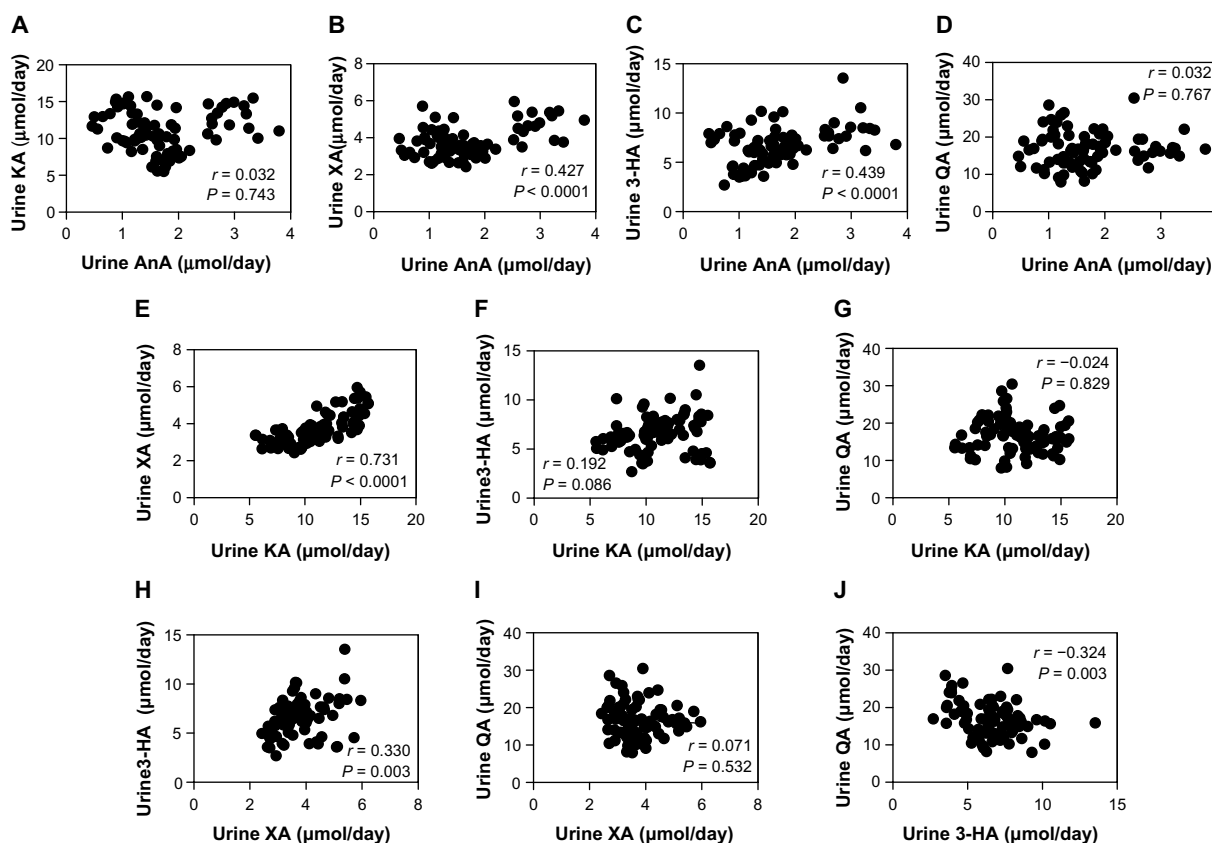
rats, kynurenine is metabolized to 3-hydroxykynurenine, but the conversion of 3-hydroxykynurenine to 3-HA (catalyzed by kynureninase) decreases,<sup>3</sup> with the accumulated 3-hydroxykynurenine being metabolized to XA. It is well known that the urinary excretion of XA abnormally increases



**Table 4.** Mean percentage urinary excretion of Trp metabolites against Trp intake in each subject.

SUBJECT NUMBER	ANA%	KA%	XA%	3-HA%	QA%
1	0.024 ± 0.003 <sup>bcd</sup>	0.176 ± 0.017 <sup>bd</sup>	0.060 ± 0.005 <sup>ab</sup>	0.118 ± 0.021 <sup>ac</sup>	0.216 ± 0.084 <sup>c</sup>
2	0.034 ± 0.002 <sup>ab</sup>	0.138 ± 0.008 <sup>cd</sup>	0.059 ± 0.005 <sup>bcd</sup>	0.122 ± 0.023 <sup>ac</sup>	0.323 ± 0.069 <sup>abc</sup>
3	0.050 ± 0.005 <sup>a</sup>	0.250 ± 0.015 <sup>a</sup>	0.090 ± 0.009 <sup>a</sup>	0.161 ± 0.034 <sup>e</sup>	0.261 ± 0.027 <sup>bc</sup>
4	0.031 ± 0.002 <sup>ac</sup>	0.182 ± 0.016 <sup>ad</sup>	0.056 ± 0.007 <sup>bcd</sup>	0.131 ± 0.023 <sup>ab</sup>	0.303 ± 0.028 <sup>abc</sup>
5	0.029 ± 0.002 <sup>ad</sup>	0.113 ± 0.013 <sup>d</sup>	0.052 ± 0.005 <sup>d</sup>	0.098 ± 0.008 <sup>bc</sup>	0.234 ± 0.034 <sup>c</sup>
6	0.021 ± 0.006 <sup>bcd</sup>	0.259 ± 0.013 <sup>a</sup>	0.081 ± 0.012 <sup>a</sup>	0.072 ± 0.008 <sup>cd</sup>	0.343 ± 0.058 <sup>abc</sup>
7	0.013 ± 0.004 <sup>d</sup>	0.231 ± 0.023 <sup>ab</sup>	0.063 ± 0.006 <sup>ad</sup>	0.134 ± 0.009 <sup>ab</sup>	0.238 ± 0.052 <sup>bc</sup>
8	0.024 ± 0.002 <sup>bcd</sup>	0.214 ± 0.021 <sup>ac</sup>	0.066 ± 0.007 <sup>ac</sup>	0.119 ± 0.024 <sup>ac</sup>	0.247 ± 0.045 <sup>bc</sup>
9	0.019 ± 0.004 <sup>cd</sup>	0.166 ± 0.011 <sup>bcd</sup>	0.050 ± 0.004 <sup>cd</sup>	0.068 ± 0.012 <sup>cd</sup>	0.402 ± 0.068 <sup>a</sup>
10	0.053 ± 0.008 <sup>a</sup>	0.196 ± 0.020 <sup>ad</sup>	0.075 ± 0.011 <sup>ab</sup>	0.129 ± 0.015 <sup>ab</sup>	0.349 ± 0.080 <sup>ab</sup>
All subjects	0.034 ± 0.015	0.222 ± 0.055	0.075 ± 0.016	0.133 ± 0.038	0.377 ± 0.093

**Notes:** Values are the mean ± SD for L-Trp metabolites collected eight times for all subjects. The means without a common superscripted letter in a same column differ,  $P < 0.05$ , determined by one-way ANOVA followed by Nonparametric Friedman test for repeated measures following Dunn's post test. Values for all subjects are the mean ± SD for 80 samples. Trp intake was calculated from the protein intake based on the assumption that protein contains 1% Trp. In the present experiment, 100 g of protein was administered each day, so Trp intake was 1,000 mg/day or 5 mmol/day.



**Figure 4.** Association between each metabolite of Trp. (A), AnA and KA; (B) AnA and XA; (C) AnA and 3-HA; (D) AnA and QA; (E), KA and XA; (F) KA and 3-HA; (G) KA and QA; (H), XA and 3-HA; (I) XA and QA; (J) 3-HA and QA. Pearson's coefficient and P-values are shown in each figure.

when a large amount of Trp is administered to rats and humans with a deficiency of vitamin B<sub>6</sub>.<sup>9</sup> But, we did not perform such a Trp administration experiment.

The concentrations of pyridine nucleotide coenzymes in liver, kidney, and other tissues are regulated by the point of

nicotinamide phosphoribosyltransferase reaction which gets a feed-back regulation by NAD<sup>+</sup>; nicotinamide phosphoribosyltransferase is inhibited by a physiological concentration of NAD<sup>+</sup> in all tissues.<sup>21</sup> Thus, the tissue concentrations of pyridine nucleotide coenzymes are kept constant. Even

if nicotinamide itself is administered, the concentrations of tissue pyridine nucleotide coenzymes do not increase. Therefore, administration of vitamin mixture in the present experiment might not affect the activities of kynurenine 3-monooxygenase which needs NADPH as a coenzyme, and of Trp 2,3-dioxygenase which is inhibited by high concentration of NADPH.<sup>22</sup>

We found that administration of B-group vitamins did not affect any of the amounts of Trp metabolites, regardless of the vitamin amounts. These results suggest that a deficiency of vitamins affects Trp metabolism, but that surplus administration of vitamins does not have any effect on Trp metabolism. These observations show that the intake levels of B-group vitamins set by *DRI-J* are suitable for the metabolism of Trp to QA.

Previously, we reported that the young Japanese women consumed  $\approx 0.7$  g/day (3.5 mmol/day) of Trp.<sup>23</sup> Comparisons between Japanese men and women are not precise. However, differences in the results of dietary Trp were not observed between men and women.<sup>23</sup>

The most striking proportional relationship was obtained between urinary levels of KA and XA. The reactions kynurenine  $\rightarrow$  KA and 3-hydroxykynurenine  $\rightarrow$  XA are catalyzed by the same enzyme: kynurenine aminotransferase. This might be one of the reasons why a close relationship was observed. Differences were observed in the urinary excretion amounts of KA and XA among subjects, so genetic differences in the expression of kynurenine aminotransferase might exist. A moderate proportional relationship was observed between urinary levels of AnA and 3-HA, which may be because the reactions kynurenine  $\rightarrow$  AnA and 3-hydroxykynurenine  $\rightarrow$  3-HA are catalyzed by the same enzyme: kynureninase. A proportional relationship was also observed between urinary levels of XA and 3-HA, both of which arise from 3-hydroxykynurenine. Formation of XA and 3-HA might be dependent upon the formation of 3-hydroxykynurenine.

Conversely, a weak inverse proportional relationship was observed between urinary levels of 3-HA and QA. This phenomenon is interesting and may be related to the organ-to-organ relationship in Trp metabolism. Terakata et al.<sup>24</sup> showed that the urinary excretion ratio of QA/3-HA (99.1/21.3 nmol/day) was 4.7 in wild-type mice and was 0.35 (18.5/53.3 nmol/day) in tryptophan 2,3-dioxygenase-knockout (TDO-KO) mice. The lower ratio denotes lower TDO activity in the liver. In TDO-KO mice, the liver cannot synthesize 3-HA from Trp because TDO (the initial enzyme of the Trp-kynurenine pathway in the liver) is absent. Thus, hepatic 3-HA in TDO-KO mice originates from extra-hepatic tissues. 3-Hydroxyanthranilic acid 3,4-dioxygenase (3-HADO) activity is  $>1000$ -fold higher than that of the other enzymes associated with Trp metabolism in the liver.<sup>25</sup> Therefore, even if 3-HA is formed in the liver, all of it is metabolized to  $\alpha$ -amino- $\beta$ -carboxymuconate- $\epsilon$ -semialdehyde (ACMS). As a result, if the Trp-kynurenine pathway in the liver is operating well, accumulation of 3-HA

does not occur and urinary excretion of 3-HA is negligible. Furthermore, QA is synthesized from 3-HA by the catalysis of 3-HADO in the liver, but the direct product is not QA, it is ACMS. One part of ACMS that could escape from the attack of  $\alpha$ -amino- $\beta$ -carboxymuconate- $\epsilon$ -semialdehyde decarboxylase undergoes cyclization to form QA (see Fig. 1). This spontaneous and non-enzymatic reaction occurs only in the liver because 3-HADO is present in large quantities only in the liver.<sup>24</sup> Hence, the origin of urinary 3-HA is extra-hepatic tissues and that of QA is the liver. The liver can incorporate exogenous 3-HA but not kynurenine and 3-hydroxykynurenine released from extra-hepatic tissues.<sup>26</sup> The inverse relationship that we observed suggests that QA formation is controlled by 3-HA incorporation into the liver from extra-hepatic tissues (Fig. 5).

Trp metabolism is known to be affected by disorders such as schizophrenia,<sup>27</sup> multiple trauma,<sup>28</sup> celiac disease,<sup>29</sup> alcoholism,<sup>30</sup> HIV infection,<sup>31</sup> malignant tumors,<sup>32</sup> cardiovascular disease,<sup>33</sup> and hormones (eg, steroid hormones).<sup>34</sup> Rayne et al.<sup>30</sup> reported that an increase in dietary levels of niacin are correlated significantly with increases in levels of KA and XA in humans. However, our research team could not reproduce such results in humans<sup>35</sup> or rats.<sup>36</sup> That is, addition of niacin did not elicit increases in urinary excretion of KA and XA.

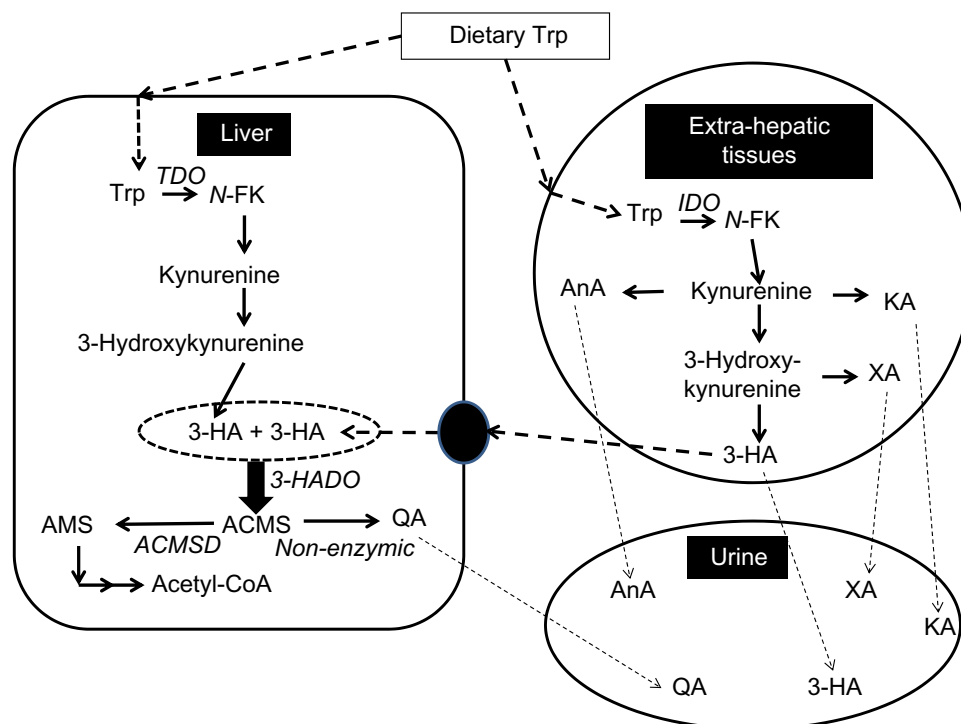
Surplus administration of B-group vitamins above the required amount did not elicit beneficial effects on the metabolism of Trp to QA. That is, we clarified that if the intake of B-group vitamins is of the required amount, the Trp-QA pathway operates within homeostasis. The values presented here, and those in a previous report,<sup>23</sup> provide a reference value of each Trp metabolite. In addition, we reported that supplementation of healthy women with  $\leq 5.0$  g/day of L-Trp confers no adverse effects.<sup>13,37</sup> In those studies,<sup>13,37</sup> urinary excretion amounts of vitamin B<sub>2</sub> and 4-pyridoxic acid (catabolite of vitamin B<sub>6</sub>) did not change even when subjects (who we administered intake levels of B-group vitamins set by the *DRI-J*) were administered 5.0 g/day of L-Trp.

## Conclusion

Administration of vitamin mixtures did not have an effect on the Trp metabolites AnA, KA, XA, 3-HA, and QA between male Japanese subjects. These results show that intake levels of B-group vitamins set by the *DRI-J* are suitable for Trp-QA metabolism. The method described here is useful for evaluation of the nutritional status of individual B-group vitamins. Evaluation of 24-h urine collections is important, but not practical. We have reported a circadian pattern of nicotinamide catabolism.<sup>38</sup> In the future, we will investigate the circadian patterns of Trp metabolites to enable evaluation of single urine collections.

## Author Contributions

KS designed the study and drafted the manuscript. KS, JH, TF carried out the experiments. JH, TF reviewed the manuscript



**Figure 5.** Proposed metabolism of Trp-QA in humans. Solid lines denote the metabolic reaction. Dotted lines indicate the flow of transport. TDO, tryptophan 2,3-dioxygenase; 3-HADO, 3-hydroxyanthranilic acid 3,4-dioxygenase; ACMSD,  $\alpha$ -amino- $\beta$ -carboxymuconate- $\epsilon$ -semialdehyde decarboxylase; IDO, indoleamine 2,3-dioxygenase. Closed circle is a 3-HA transporter into the liver from extra-hepatic tissues.

and helped in the study design. All authors approved the final manuscript.

### Supplementary Data

**Supplementary Table 1.** Individual mean values of daily L-Trp metabolites collected eight times in male Japanese subjects in terms of  $\mu\text{mol/g}$  creatinine.

### REFERENCES

1. Fukuwatari T, Murakami M, Ohta M, et al. Changes in the urinary excretion of the metabolites of the tryptophan-niacin pathway during pregnancy in Japanese women and rats. *J Nutr Sci Vitaminol (Tokyo)*. 2004;50:392–8.
2. Takeuchi F, Otsuka H, Shibata Y. Purification and properties of kynureninase from rat liver. *J Biochem*. 1980;88:987–94.
3. Inada J, Okuno E, Kimura M, Kido R. Intracellular localization and characterization of 3-hydroxykynureninase in human liver. *Int J Biochem*. 1984;16:623–8.
4. Okamoto H, Yamamoto S, Nozaki M, Hayaishi O. On the submitochondrial localization of L-kynurenine-3-hydroxylase. *Biochem Biophys Res Commun*. 1967;26:309–14.
5. Uemura T, Hirai K. L-kynurenine 3-monoxygenase from mitochondrial outer membrane of pig liver: purification, some properties and monoclonal antibodies directed to the enzyme. *J Biochem*. 1998;23:253–62.
6. Okamoto H, Hayaishi O. Intra mitochondrial localization of kynurenine amino-transferase. *J Biol Chem*. 1970;245:3603–5.
7. Porter CC, Clark I, Silber RH. The effect of vitamin deficiencies on tryptophan metabolism in the rat. *Arch Biochem*. 1948;18:339–43.
8. Mason M. The metabolism of tryptophan in riboflavin-deficient rats. *J Biol Chem*. 1953;201:513–8.
9. Yeh JK, Rowen RR. Effects of vitamin B-6 deficiency and tryptophan loading on urinary excretion of tryptophan metabolites in mammals. *J Nutr*. 1977;107:261–71.
10. Shibata K, Fukuwatari T, Imai E, et al. Dietary reference intakes for Japanese 2010: water-soluble vitamins. *J Nutr Sci Vitaminol (Tokyo)*. 2012;58:S67–82.
11. Standard Tables of Food Composition in Japan. Report of the Subdivision on Resources Japan. 2010. The Council for Science and Technology. Ministry of Education, Cultures, Sports, Science and Technology. Publisher: Official Gazette Co-operation of Japan, Tokyo, Japan.
12. Fukuwatari T, Ohta M, Kimura N, Sasaki R, Shibata K. Conversion ratio of tryptophan to niacin in Japanese women fed a purified diet conforming to the Japanese dietary reference intakes. *J Nutr Sci Vitaminol (Tokyo)*. 2004;50:385–91.
13. Hiratsuka C, Sano M, Fukuwatari T, Shibata K. Time-dependent effects of L-tryptophan administration on urinary excretion of L-tryptophan metabolites. *J Nutr Sci Vitaminol (Tokyo)*. 2014;60:255–60.
14. Shibata K. Fluorimetric micro-determination of kynurenic acid, as endogenous blocker of neurotoxicity, by high-performance liquid chromatography. *J Chromatogr*. 1988;430:376–80.
15. Shibata K, Onodera M. Measurement of 3-anthranilic acid and anthranilic acid in urine by high-performance liquid chromatography. *Agric Biol Chem*. 1991;55:143–8.
16. Shibata K, Onodera M. Simultaneous high-performance liquid chromatographic measurement of xanthurenic acid and 3-hydroxyanthranilic acid in urine. *Biosci Biotechnol Biochem*. 1992;56:974.
17. Mawatari K, Oshida K, Inuma F, Watanabe W. Determination of quinolinic acid in human urine by liquid chromatography with fluorimetric detection. *Anal Clin Acta*. 1995;302:179–83.
18. Shibata K, Kondo T, Onodera M. Comparison of tryptophan-nicotinamide metabolism between male and female rats. *Biosci Biotechnol Biochem*. 1993;57:858–9.
19. Shibata K, Kondo T. Effect of progesterone and estrone on the conversion of tryptophan to nicotinamide in rats. *Biochem Biotechnol Biochem*. 1993;57:1890–3.
20. Shibata K, Toda S. Effects of sex hormones on the metabolism of tryptophan to niacin and to serotonin in male rats. *Biosci Biotechnol Biochem*. 1997;61:1200–2.
21. Shibata K, Taguchi H, Nishitani H, et al. End product inhibition of the activity of nicotinamide phosphoribosyltransferase from various tissues of rats by NAD. *Agric Biol Chem*. 1989;53:2283–4.
22. Badawy AA, Evans M. The regulation of rat liver tryptophan pyrrolase activity by reduced nicotinamide-adenine dinucleotide (phosphate). Experiments with glucose and nicotinamide. *Biochem J*. 1976;156:381–90.
23. Hiratsuka C, Fukuwatari T, Shibata K. Fate of dietary tryptophan in young Japanese women. *Int J Trp Res*. 2012;5:33–47.
24. Terakata M, Fukuwatari T, Kadota E, et al. The niacin required for optimum growth can be synthesized from L-tryptophan in growing mice lacking tryptophan-2,3-dioxygenase. *J Nutr*. 2013;143:1046–51.
25. Shibata K, Morita N, Shibata Y, Fukuwatari T. Enzymes that control the conversion of L-tryptophan-nicotinamide and the urinary excretion ratio of ( $N^1$ -methyl-2-pyridone-5-carboxamide +  $N^1$ -methyl-4-pyridone-4-carboxamide)/ $N^1$ -methylnicotinamide in mice. *Biosci Biotechnol Biochem*. 2013;77:2105–11.





26. Shibata K. Efficiency of dietary tryptophan, anthranilic acid, and 3-hydroxyanthranilic acid as niacin in rats. *Vitamins*. 1994;68:579–89.
27. Rayne IR, Walsh EM, Whittenburg JR. Relationship of dietary tryptophan and niacin to tryptophan metabolism in schizophrenics and nonschizophrenics. *Am J Clin Nutr*. 1974;27:565–71.
28. Ploder M, Spittler A, Schroecksadel K, et al. Tryptophan degradation in multiple trauma patients: survivors compared with non-survivors. *Clin Sci*. 2009;116:593–8.
29. Torres MI, Lopez-Casado MA, Lorite P, Rios A. Tryptophan metabolism and indoleamine 2,3-dioxygenase expression in celiac disease. *Clin Exp Immun*. 2007;148:419–24.
30. Rayne IR, Lu GH, Meyer K. Relationship of dietary tryptophan and niacin to tryptophan metabolism in alcoholics and nonalcoholics. *Am J Clin Nutr*. 1974;27:572–9.
31. Manches O, Fernandez MV, Plumas J, Chaperot L, Bhardwai N. Activation of the noncanonical NF- $\kappa$ B pathway by HIV controls a dendritic cell immunoregulatory phenotype. *Proc Natl Acad Sci*. 2012;109:14122–27.
32. Tawara I, Shlomchik WD, Jones A, et al. A crucial role for host APCs in the induction of donor CD4<sup>+</sup>CD25<sup>+</sup> regulatory T cell-mediated suppression of experimental graft-versus-host disease. *J Immunol*. 2010;185:3866–72.
33. Cuffy MC, Silverio AM, Qin L, et al. Induction of indoleamine 2,3-dioxygenase in vascular smooth muscle cells by interferon-gamma contributes to medial immunoprivilege. *J Immunol*. 2007;179:5246–54.
34. Rose DP, Braidman IP. Excretion of tryptophan metabolites as affected by pregnancy, contraceptive steroids, and steroid hormones. *Am J Clin Nutr*. 1971;24:673–83.
35. Fukuwatari T, Shibata K. Effect of nicotinamide administration on the tryptophan-nicotinamide pathway in humans. *Int J Vitam Nutr Res*. 2007;77:255–62.
36. Fukuwatari T, Kurata K, Shibata K. Effects of excess nicotinic acid on growth and the urinary excretion of B-group vitamins and the metabolism of tryptophan in weaning rats. *Shokubin Eiseigaku Zasshi*. 2009;50:80–4.
37. Hiratsuka C, Fukuwatari T, Sano M, Saito K, Sasaki S, Shibata K. Supplementing healthy women with up to 5.0 g/d of L-tryptophan has no adverse effects. *J Nutr*. 2013;143:859–66.
38. Okamoto H, Ishikawa A, Yoshitake H, et al. Diurnal variations in human urinary excretion of nicotinamide catabolites. Effects of stress on the diurnal variations. *Am J Clin Nutr*. 2003;77:406–10.
39. Fukuwatari T, Shibata K. Relative availability of B-group vitamin in a test diet to free vitamin. *J Home Eco Jpn*. 2008;59:403–10.
40. Fukuwatari T, Shibata K. Relative availability of water-soluble vitamins in a white bread diet to free vitamin. *J Home Eco Jpn*. 2009;60:57–63.