

Following in Emil Fischer's Footsteps: A Site-Selective Probe of Glucose Acid–Base Chemistry

Published as part of *The Journal of Physical Chemistry virtual special issue "Daniel Neumark Festschrift"*.

Sebastian Malerz,[◆] Karen Mudryk,[◆] Lukáš Tomaník, Dominik Stemer, Uwe Hergenahn, Tillmann Buttersack, Florian Trinter, Robert Seidel, Wilson Quevedo, Claudia Goy, Iain Wilkinson, Stephan Thürmer, Petr Slaviček,* and Bernd Winter*



Cite This: *J. Phys. Chem. A* 2021, 125, 6881–6892



Read Online

ACCESS |



Metrics & More

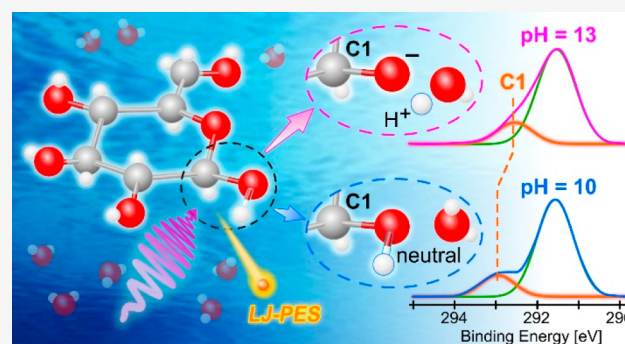


Article Recommendations



Supporting Information

ABSTRACT: Liquid-jet photoelectron spectroscopy was applied to determine the first acid dissociation constant (pK_a) of aqueous-phase glucose while simultaneously identifying the spectroscopic signature of the respective deprotonation site. Valence spectra from solutions at pH values below and above the first pK_a reveal a change in glucose's lowest ionization energy upon the deprotonation of neutral glucose and the subsequent emergence of its anionic counterpart. Site-specific insights into the solution-pH-dependent molecular structure changes are also shown to be accessible via C 1s photoelectron spectroscopy. The spectra reveal a considerably lower C 1s binding energy of the carbon site associated with the deprotonated hydroxyl group. The occurrence of photoelectron spectral fingerprints of cyclic and linear glucose prior to and upon deprotonation are also discussed. The experimental data are interpreted with the aid of electronic structure calculations. Our findings highlight the potential of liquid-jet photoelectron spectroscopy to act as a site-selective probe of the molecular structures that underpin the acid–base chemistry of polyprotic systems with relevance to environmental chemistry and biochemistry.



INTRODUCTION

Glucose is a ubiquitous monosaccharide of major significance in living organisms.^{1,2} It is the precursor of many oligo- and polysaccharides that mediate cell–cell communication,³ build up the scaffold of cells,^{4–6} or serve as energy storage units.^{7–10} It is a natural energy source synthesized via the conversion of solar energy into chemical energy by plants during photosynthesis.^{11,12} Consequently, it plays a central role in the metabolic pathways that govern the flow of energy and matter that sustain life.¹³ As a result, it has also become relevant in the investigation of renewable energy technologies that seek to mimic nature, in particular, with the demonstration of alkaline glucose fuel cells.^{14,15}

The chemistry associated with the use of glucose as a fuel source, in both living organisms and technological devices, is inherently related to the nature of its structure–function relationship and acid–base chemistry in aqueous solution. Despite this, there remains much to be learned about the acid–base properties of this fundamental molecule. This is perhaps surprising because the structure of glucose has been intensively studied since the turn of the 19th century, when Emil Fischer (Nobel Prize in Chemistry 1902) reported the chemical synthesis of D-(+)-glucose and demonstrated its

stereoisomeric forms.^{16,17} But it is only with advancing experimental and theoretical methods,^{18–21} in particular, photoelectron spectroscopy (PES) from an aqueous solution,^{22–25} that previously inaccessible molecular structural details can now be resolved.

With an elemental composition of $C_6H_{12}O_6$, glucose is an aldohexose with an aldehyde group at the C1 position in the Fischer projection. In aqueous solution, glucose predominantly adopts the six-membered closed-ring pyranose structure (>99%), in which the C1 atom forms a hemiacetal linkage to the C5 atom. In smaller quantities, the five-membered, closed-ring furanose (<0.5%) and linear (<0.05%) forms are also present.^{26,27} Traditionally, the stereochemistry in glucose molecules is denoted by the relative orientation of the hydroxyl group at the C5 site, which points to the left in the L form and to the right in the D form, as viewed in the Fischer

Received: May 28, 2021

Revised: July 14, 2021

Published: July 30, 2021

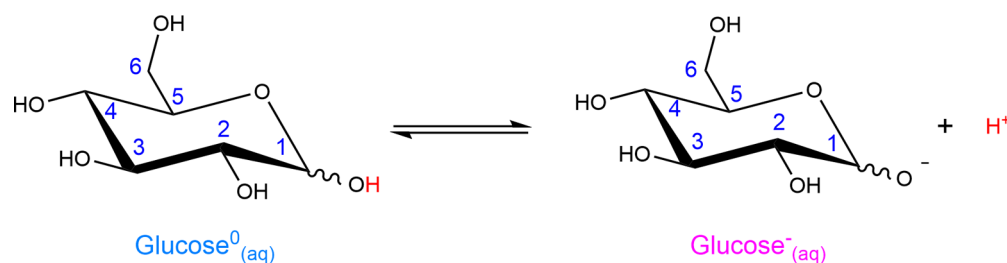


Figure 1. Schematics of the predominantly adopted six-membered closed-ring protonated ($\text{glucose}^0_{(\text{aq})}$) and deprotonated ($\text{glucose}^-_{(\text{aq})}$) structures of glucose in aqueous solution.^{40,42} C–OH sites are labeled 1–6, and the nomenclature is used throughout the text. The zigzag line is used to indicate the two possible orientations of the hydroxyl group at the C1 site, downward or upward, in the α and β anomeric forms, respectively.

projection. As with many biomolecules, such as amino acids, proteins, and DNA, glucose exhibits a remarkable degree of homochirality. In nature, the right-handed D form dominates.^{28,29} Upon cyclization into the pyranose form, a new stereocenter emerges at the C1 site. At this anomeric center, the stereochemistry is denoted depending on the relative orientation of the hemiacetal hydroxyl group as α or β . Because there are multiple stereocenters within the molecule, α - and β -glucose are diastereomers, which exhibit different physicochemical properties. In solution, the α and β forms of glucose are in equilibrium with the open-ring form and therefore constantly interconvert through a process known as mutarotation,³⁰ with β -glucose being favored ($\sim 38\%$ α -glucose vs $\sim 62\%$ β -glucose) due to the anomeric effect;^{27,31} however, this equilibrium can be shifted by tuning the solution pH³² or through the addition of inorganic salts.³³ Glucose is also known to readily isomerize under alkaline conditions to fructose and mannose via Lobry de Bruyn–Alberda van Ekenstein transformations, a process of significance in the preparation of liquid hydrocarbon fuels.^{34,35}

Glucose is a weak acid with at least two reported acid-ionization (*i.e.*, deprotonation) equilibria and acidity constants ($\text{p}K_{\text{a}}$) of 12.1 ($\text{p}K_{\text{a}1}$) and 13.9 ($\text{p}K_{\text{a}2}$),^{36,37} which highlight its enhanced reactivity in alkaline media.^{38,39} Previous determinations of glucose $\text{p}K_{\text{a}}$ values involved the use of site-insensitive high-performance liquid chromatography³⁹ and titration-based methods.^{36,40,41} Consequently, ambiguity remains regarding the extent to which the transition defined at $\text{p}K_{\text{a}1}$ involves charge sharing with C–OH groups beyond the C1 site.⁴² Schematic representations of both the neutral ($\text{glucose}^0_{(\text{aq})}$) and deprotonated ($\text{glucose}^-_{(\text{aq})}$) forms^{40,42} of aqueous glucose are shown in Figure 1.

Whereas site-sensitive methods such as X-ray PES have been applied to study glucose in the solid phase,^{43–45} this approach is unable to access the behavior of aqueous-phase glucose, which is expected to be significantly affected by intra- and intermolecular hydrogen bonding.^{32,40,46} In this context, a local, site-selective, solution-phase characterization of the molecular structure of glucose is needed to better understand its acid–base behavior. To meet this challenge, we leverage the unique experimental capabilities of liquid-jet PES (LJ-PES),^{22,47} which enable us to probe the overall and local electronic structure of protonated and deprotonated solvated glucose via valence and site-specific measurements, respectively. This approach has been previously applied to investigate pH-dependent electronic-structure changes and protonation sites in amino acids.^{48,49}

Herein, we present a combined experimental and theoretical study of the molecular structure of glucose in the pH range from 2 to 13. Utilizing soft X-ray LJ-PES, we monitor (mostly

alkaline) pH-dependent changes in the solution-phase valence and C 1s PES spectra of glucose. Specifically, we probe aqueous-phase binding energy (BE) shifts associated with the most acidic C–OH group(s) and determine the $\text{p}K_{\text{a}1}$ value. To gain insight into the existence of single or multiple deprotonation equilibria related to $\text{p}K_{\text{a}1}$, we investigate the associated chemical structures and confirm principal spectral assignments with the aid of electronic-structure calculations. Our work highlights LJ-PES as a methodology to site-selectively probe ionization equilibria in solvated species, in contrast with previous site-unspecific measurements.

METHODS

Experiments. Glucose aqueous solutions of 1 M concentration were prepared at room temperature by dissolving α -D-(+)-glucose crystals (Acros Organics, >99% purity, anhydrous) in Millipore water (55 nS/cm). Measurements were performed from both freshly prepared and several-hour-old solutions, yielding identical spectra and implying that α and β anomers cannot be distinguished in the present experiments. The pH was varied by the dropwise addition of either HCl (10% and 37% w/w) or NaOH (1 M and 10 M) aqueous solutions under constant magnetic stirring. The solution pH was monitored with a pH meter (VWR, pHenomenal 1100L) to produce 1 M samples at pH values of 2.0, 7.4, 10.0, 10.5, 11.0, 11.5, 12.0, 12.5, and 13.0. Glucose aqueous solutions of 1 M concentration without prior pH adjustment were found to exhibit a pH of 4.8 ± 0.2 . Because of the protonated, nonionic nature of pure glucose, NaCl (Aldrich, $\geq 99\%$) in 25–50 mM concentration was added to those samples to ensure sufficient solution conductivity and minimize sample charging due to streaming potentials.⁵⁰ LJ-PES experiments were performed at the U49/2 PGM-1 beamline⁵¹ at BESSY II using the SOL³PES setup⁵² and the P04 beamline⁵³ at PETRA III using the EASI setup.⁵⁴ Although the majority of the measurements were conducted at the P04 beamline, spectra from glucose solutions at two complementary pH values (11 and 12) were subsequently recorded at BESSY II. The latter yielded somewhat lower electron count rates due to the lower photon flux. At BESSY II, experiments were performed with horizontally polarized light, whereas P04 provided circularly polarized light. Both setups were equipped with differentially pumped hemispherical electron analyzers that detected photoelectrons emitted from the sample either at 0° with respect to the light polarization (SOL³PES) or at 50° with respect to the light propagation (EASI). The samples were introduced into the experimental chamber in the form of liquid microjets⁵⁵ using glass capillary nozzles of 25–35 μm inner diameter and 0.6–0.8 mL/min sample flow rates. In the SOL³PES setup, the (vertical) liquid-

jet flow direction was orthogonal with respect to the light polarization and the electron analyzer detection axes. In the EASI setup, the (horizontal) liquid-jet flow was at 90° with respect to both the light propagation (floor plane) and the electron detection (at an angle of 50° to the floor plane) directions. The liquid jet was electrically connected and grounded to the experimental setup by means of a gold wire immersed in the electrically conductive solution (in the case of SOL³PES) or by a small metallic tube inserted into the main polyether ether ketone (PEEK) liquid delivery line (in the case of EASI). Valence and C 1s spectra were recorded as a function of pH using photon energies of 600 eV and 850 eV, respectively. Such photon energies enabled us to produce photoelectrons with approximately 590 eV (valence) and 560 eV (C 1s) kinetic energies, thus ensuring sufficiently large probing depths into the liquid jet⁵⁶ (*i.e.*, probing of fully hydrated glucose molecules). At PETRA III, the overall experimental energy resolution was 230 meV (for 600 eV photon energy) and 280 meV (for 850 eV photon energy), respectively. The complementary measurements at BESSY II had a somewhat lower overall resolution of 380 meV (850 eV only).

Computations. Valence vertical ionization energies (VIEs), that is, BEs, were calculated for glucose structures optimized at the density functional theory (DFT) level using the CAM-B3LYP⁵⁷ functional and the 6-31+G* basis set. All of the optimized structures were confirmed as energy minima via frequency analysis. The polarizable continuum model (PCM)^{58,59} was used to mimic the presence of the water solvent. In particular, the nonequilibrium PCM⁶⁰ was used to describe the fast photoelectron ionization process. To account for specific intermolecular interactions, several nearest water molecules were considered explicitly. Calculations were performed in Gaussian 09 (revision D.01)⁶¹ using the default parameters for the PCM. VIEs were calculated with the DFT-based delta self-consistent field (Δ SCF) approach, followed by a time-dependent DFT (TDDFT) evaluation of the BEs for deeper-lying electrons. The VIEs were then calculated as⁶²

$$\text{VIE}_i = \text{VIE}_{\text{HOMO}} + E_{\text{exc}}(i \rightarrow \text{SOMO})$$

where VIE_i is the VIE of the i th electron and E_{exc} is the excitation energy, restricted to excitations into the singly occupied molecular orbital (SOMO).

Core-level VIEs were calculated using the maximum overlap method (MOM)⁶³ in the Q-Chem 4.3 software.⁶⁴ This approach is centered on ground-state electronic structure techniques while avoiding the variational collapse of the wave function and providing reliable core-electron BEs for solvated systems.⁶⁵ Calculations were performed using the CAM-B3LYP functional and the cc-pVTZ split basis set for all of the H atoms in glucose. For the C and O atoms, the aug-cc-pCVTZ basis set was used. The parameters in the nonequilibrium PCM were set to match those in Gaussian 09, revision D.01⁶¹ (atomic radii from Universal Force Field⁶⁶ and scaling factor $\alpha = 1.1$). C 1s PES spectra were modeled from the calculated VIEs at the optimized geometries via the empirical broadening scheme. In that way, each modeled spectrum was calculated considering six energies originating from six C atoms in the molecule, and the corresponding sum of six Gaussian components was centered at the respective VIEs. The width of each Gaussian is characterized by a standard deviation (σ) of 0.45 eV, which is reasonable for PES spectra of solvated systems.⁶⁷ This value was found to fit the

experimental PES data of fully protonated glucose (*i.e.*, data recorded at pH 10; see Figure 3) and was thus subsequently used as a constant to simulate similar PES spectra for the different deprotonated forms (*i.e.*, considering the deprotonation of different hydroxyl groups).

Calculations of pK_a values were performed using Gaussian 09 (revision D.01)⁶¹ and implementing two different approaches. The first approach was based on a thermodynamic cycle including the deprotonation of glucose in the gas phase. Within that methodology, aqueous-phase deprotonation energetics (*i.e.*, changes in Gibbs free energies, $\Delta G_{(\text{aq})}$) were calculated by evaluating their gas-phase counterpart ($\Delta G_{(\text{g})}$) and adding solvation energies ($\Delta G_{(\text{solv})}$). Thus the energies associated with the solvation of gas-phase reactants ($\Delta G_{(\text{solv},\text{react})}$) and products ($\Delta G_{(\text{solv},\text{prod})}$) were used to calculate $\Delta G_{(\text{aq})}$ as

$$\Delta G_{(\text{aq})} = \Delta G_{(\text{g})} + \Delta G_{(\text{solv},\text{prod})} - \Delta G_{(\text{solv},\text{react})}$$

The gas-phase energetics were evaluated using CAM-B3LYP/aug-cc-pVTZ at a temperature of 298.15 K using frequency calculations to obtain the gas-phase free energies. Solvation energies of protonated and deprotonated glucose were calculated using the PCM. The solvation energy of the proton was taken from the literature to be -265.9 kcal/mol.⁶⁸ The second, more straightforward approach consisted of the implementation of the methodology proposed by Thapa and Schlegel.⁶⁹ This method uses a polarizable solvation model to directly evaluate aqueous-phase free-energy changes. According to the authors, reasonable results can be obtained when using a combination of the ω B97XD⁷⁰ method with a 6-31+G* basis set and the solvation model based on solute electron density (SMD).⁷¹

RESULTS AND DISCUSSION

Valence PES Spectra from Protonated and Deprotonated Aqueous-Phase Glucose. Valence PES spectra from 1 M glucose aqueous solutions at pH 10 (below pK_{a1} , bottom) and 13 (above pK_{a1} , top) are shown in Figure 2. The spectra are representative of protonated ($\text{glucose}^0_{(\text{aq})}$) and deprotonated ($\text{glucose}^-_{(\text{aq})}$) molecules. The data are presented on a BE scale calibrated according to the 1b₁ BE of neat liquid water.⁴ The spectra are almost identical, dominated by the contributions from liquid water corresponding to the ionization of water's 1b₁, (split) 3a₁, and 1b₂ leading orbitals (as labeled in Figure 2; see ref 72 for details). Small contributions from gaseous water from the vapor layer surrounding the liquid jet can also be observed (mainly from 1b_{1(g)}} photoelectrons; see a somewhat sharper peak at ~ 12.5 eV).

We expected that a change in the charge state of glucose upon deprotonation, from $\text{glucose}^0_{(\text{aq})}$ at pH 10 to $\text{glucose}^-_{(\text{aq})}$ at pH 13, would lead to a change in the lowest VIE of the molecule (as demonstrated in previous experiments on the pH-dependent changes in the lowest VIEs of aqueous imidazole,^{73,74} phosphate,⁷⁵ and phenol/phenolate⁷⁶). A small but decisive difference between the pH 10 and pH 13 data is the occurrence of a photoelectron signal near 8.5 eV BE in the pH 13 spectrum. Enlarged views of this spectral region are presented in the figure inset. Gaussian peaks from a cumulative fit analysis highlight the spectral contributions from $\text{glucose}^-_{(\text{aq})}$ (at 8.5 ± 0.3 eV, magenta fill), added hydroxide ($\text{OH}^-_{(\text{aq})}$, striped fill, fixed at 9.2 eV BE, as reported in ref 77),

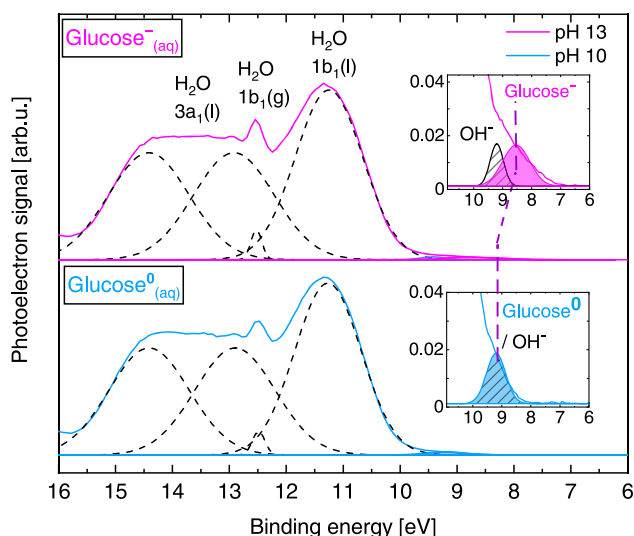


Figure 2. Valence PES spectra of 1 M glucose aqueous solutions at pH 10 (bottom, light blue curve) and 13 (top, magenta curve) measured at a 600 eV photon energy. The dashed lines are Gaussian curves representing signal contributions from water's leading orbitals, $1b_1$ and (split) $3a_1$; liquid (l) and gas-phase (g) signals are assigned.⁷² Spectra intensities are displayed to yield the same height as the $1b_{1(l)}$ peak. The figure insets show enlarged views of the photoelectron features associated with the lowest vertical ionizing transitions in glucose⁰(aq) (light blue fill) and glucose⁻(aq) (magenta fill) as well as from OH⁻(aq) (peak with striped fill). The BE of the latter was fixed during the fitting procedure according to the value reported in ref 77.

and glucose⁰(aq) (light blue fill, overlapping with the OH⁻(aq) feature). Note that lifetime broadening effects are negligible in the case of valence ionization, and a cumulative Gaussian fit approach is thus appropriate. The overall error is a combination of the fitting error reported from the least-squares fitting procedure, the experimental resolution (from both the light source and the electron analyzer) of ~ 0.23 eV, and the error associated with the calibration of the BE scale. As previously stated, the BE scale is referenced to the $1b_1$ BE of neat liquid water, which, however, may deviate slightly from the $1b_1$ BE value of the solution. Solute-induced changes in the electronic structure of water and the work function of the solution may occur, which shift the BE of the photoelectron features. A slight shift of the liquid $1b_1$ peak for very high (up to 8 M) concentrations of NaI has been discussed lately by some of the coauthors⁷⁸ and is being further investigated by our group. Notably, we do not expect any detectable reference-level changes from the 1 M glucose solute, similarly from the small amounts of HCl_(aq) and NaOH_(aq) added to the solutions. On this basis, we estimate cumulative valence peak BE errors of 0.3 eV. Our interpretation of the spectral changes observed upon changing the solution pH from 10 to 13 was corroborated by our calculations, which revealed glucose⁻(aq) to be the prevalent species at pH 13, as opposed to glucose⁰(aq) at pH 10, as shown in the following.

We have evaluated the respective valence VIEs for the minimum-energy structures of protonated and deprotonated glucose in both the α and β anomeric forms. The calculated structures are consistent with the energetic minima previously published,^{79,80} showing only minor energy differences between the α and β forms for both gas- and aqueous-phase glucose. (See Table S1 in the Supporting Information (SI) for details.) Because of the very high charge density at the O atom, the

addition of explicit solvent molecules is required to reach quantitatively correct values in calculations of the electron BEs for glucose⁻(aq). (The calculated results are not sensitive to the explicit solvent addition for glucose⁰(aq).)⁸¹ The highest occupied molecular orbital (HOMO), associated with the lowest VIE, is of π character and is delocalized over the whole molecule for neutral glucose⁰(aq). However, the HOMO is more localized on the single C and single O of the deprotonated hydroxyl group in glucose⁻(aq).

The first VIE of fully protonated glucose (glucose⁰(aq)), with the explicit inclusion of close-range water interactions represented by two water molecules, was calculated to be 9.09 eV (Table 1), in good agreement with the onset of the

Table 1. Calculated First Three Vertical Ionization Energies (VIEs) of Fully Protonated (glucose⁰(aq)) and Singly Deprotonated (glucose⁻(aq)) Aqueous-Phase Glucose in Electronvolts^a

	glucose ⁰ (aq)	glucose ⁻ (aq)
1. VIE (HOMO)	9.09	7.95
2. VIE (HOMO-1)	9.40	8.87
3. VIE (HOMO-2)	9.66	9.44

^aCalculations were performed by applying the hybrid model with two or six explicitly solvating water molecules for glucose⁰(aq) and glucose⁻(aq), respectively.

spectrum at pH 10. The solvent shift (*i.e.*, the difference between the gas- and solution-phase VIEs) is ~ 1 eV. For glucose⁻(aq), the calculated first VIE is at 7.95 eV (using six explicit water molecules; the larger number of explicit solvating molecules is needed due to a slower BE convergence compared with the neutral species; see Table S1 in the SI), >1 eV lower than that of glucose⁰(aq), in reasonable agreement with the 8.5 eV feature observed from the experimental data recorded at pH 13. (A solvent shift of almost 2 eV is observed in this case.) Note that the calculated VIEs without the inclusion of specific water interaction are much lower (6.75 eV). The VIEs for the various deprotonation sites and for the different anomers are rather close in energy, as presented in Table S1 in the SI for both α and β anomers.

Overall, we conclude that the valence PES spectrum recorded at pH 13 (*i.e.*, above pK_{a1}) reflects the acid ionization of glucose's hydroxyl groups. However, site-specific insights into the molecular structural changes that take place upon deprotonation, that is, which C–OH groups are involved in the deprotonation equilibria, cannot be inferred from the valence spectra. The reason is that the valence energies of the different hydroxyl groups are too similar to be resolved, and the signal contributions from the solvent as well as the OH⁻(aq) solution component overlap with the primary feature of interest. Site-specific and more differential information is revealed in the C 1s core-level PES spectra, which thus serve as a probe of more specific acid–base properties of glucose, as will be presented in the following.

C 1s Core-Level PES Spectra from Aqueous-Phase Glucose: pH-Dependent Changes. Figure 3 shows C 1s PES spectra from 1 M glucose aqueous solutions in the 2–13 pH range. As in Figure 2, the data are presented on a BE scale, but this time they are calibrated based on the liquid water O 1s BE⁸² measured from each solution.^a In this way, (pH-dependent) sample surface-charging effects are canceled out, and the observed small energy shifts (up to ~ 300 meV; see the

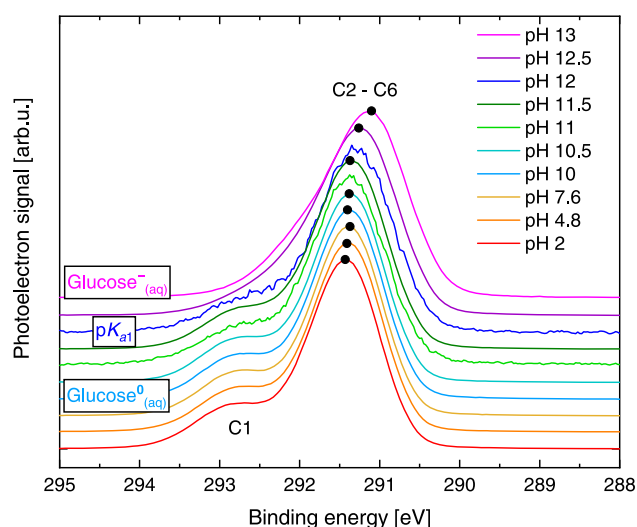


Figure 3. C 1s PES spectra from 1 M glucose aqueous solutions, with pH ranging from 2 to 13, measured at an 850 eV photon energy. The spectra are scaled to yield the same height of the 291.5 eV BE peak and are additionally presented with a vertical offset to highlight the pH-dependent spectral changes. The black dots at the peak of each spectrum serve as a guide for the energy change of the main (C2–C6) peak as a function of pH. Spectra at pH 11 and 12 were measured at the lower-photon-flux beamline and correspondingly have lower signal-to-noise levels. (See the Methods section.) These spectra are manually shifted such that the respective C2–C6 peak centers match the trend of peak maxima (highlighted using black dots).

black dots at the peak of each spectrum to guide the eye) of the overall C 1s spectra are argued to reflect true pH-dependent BE changes, as similarly observed and confirmed by our calculations, see the next section. We further stress that the amounts of $\text{HCl}_{(\text{aq})}$ and $\text{NaOH}_{(\text{aq})}$ solutes added for pH adjustment are too small to lead to a detectable change in the electronic structure of water.⁷⁸

With reference to available PES studies from solid-phase (crystalline) glucose⁴³ and carbon spheres grown from glucose solutions,⁴⁴ the glucose C 1s aqueous-phase spectral features can be crudely assigned to ionization from the C1 and C2–C6 atoms, as labeled in Figure 3. In the 2–10 pH range, we observe a main C 1s peak near 291.5 eV BE accompanied by a higher energy shoulder near 293.0 eV, with no noticeable pH-dependent spectral changes. The latter implies the (near-sole) existence of a single-charge-state species, that is, neutral glucose ($\text{glucose}^0_{(\text{aq})}$) up to pH 10. At pH values higher than 10, the higher energy shoulder becomes less clear, as it appears to move toward lower BEs, consistent with observations from the valence data presented in the previous section. This is indicative of an increase in the fraction of deprotonated glucose molecules up to pH 13 (above $\text{pK}_{\text{a}1}$), when $\text{glucose}^-_{(\text{aq})}$ becomes the prevalent species. (Considering a $\text{pK}_{\text{a}2}$ of 13.9,^{36,37} a fraction of doubly ionized molecules should also be present.)

Qualitatively, the larger C 1s BE of the higher-energy shoulder in $\text{glucose}^0_{(\text{aq})}$ with respect to $\text{glucose}^-_{(\text{aq})}$ reflects the additional positive charge at the specific C site in the former. The observations described here highlight pH-dependent, C-site-specific spectral changes in the C 1s PES data, that is, a site-selective probe of molecular structure changes in glucose upon deprotonation. Details regarding the identity of the C–OH groups involved in the acid dissociation process

highlighted in the data are provided by our calculations, as presented in the following section.

Assignment of the C 1s PES Spectrum of Glucose: Electronic Structure Theory Calculations. The C 1s BEs of each individual C atom (C1–C6) were evaluated using the MOM method combined with the CAM-B3LYP/aug-cc-pCVTZ approach. The BEs were calculated in the gas and aqueous phases for both the α - and β -glucose forms, focusing on the C 1s BE changes taking place upon deprotonation. The calculations considered microsolvation of glucose by the addition of a single explicit water molecule; the addition of further water molecules did not lead to any significant change in the BE. The calculated BEs are summarized in Table S2 in the SI. As follows, we will exclusively discuss the results for the β anomer; the results for the α anomer are very similar.

Simulated C 1s PES spectra for $\text{glucose}^0_{(\text{aq})}$ and $\text{glucose}^-_{(\text{aq})}$ are presented in Figure 4. The corresponding experimental

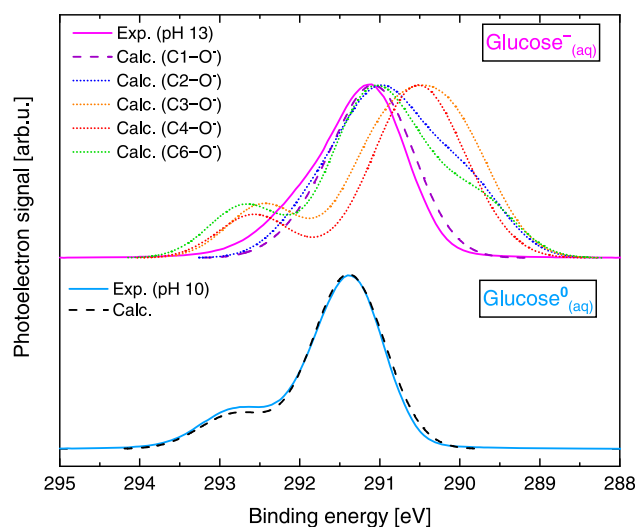


Figure 4. Comparison of experimental (full line) and calculated (dashed and dotted lines) C 1s PES data for $\text{glucose}^0_{(\text{aq})}$ (pH 10, bottom) and $\text{glucose}^-_{(\text{aq})}$ (pH 13, top). All calculated curves were shifted in binding energy by an offset value determined such that the main $\text{glucose}^0_{(\text{aq})}$ peak overlaps with the experimental data at pH 10, as explained in the text. Calculated spectra considering deprotonation at different C–OH sites are shown in the top panel. Note that there is no C5–OH group to deprotonate in the cyclic form.

data (C 1s PES spectra recorded at pH 10 and 13, respectively, as shown in Figure 3) are plotted for comparison. For $\text{glucose}^0_{(\text{aq})}$ (bottom panel), the main C 1s feature in the calculated spectra is overlapped with the experimental data. The same spectral shift was applied to the calculated $\text{glucose}^-_{(\text{aq})}$ curves (top panel), which were produced considering deprotonation at different C–OH sites. The BE shift of the C 1s main feature between the experimental and calculated spectra amounted to 240 meV. Such a magnitude accounts for the (expected) few hundred millielectronvolts shift between theory and experiment due to the known shortcomings of DFT calculations in providing core-level BEs on an absolute scale. (We assume that the absolute value is shifted due to the localized nature of the C 1s electron, yet the differences between various forms are faithfully described.) Nonetheless, consistent with the experimental data, our calculations show that all BEs are shifted toward lower values when going from $\text{glucose}^0_{(\text{aq})}$ to $\text{glucose}^-_{(\text{aq})}$. (See Figure 3.)

For glucose⁰_(aq), the shape of the calculated, fully protonated (cyclic glucose, pyranose form) spectrum (black dashed line) matches that of the experimental data (light-blue full line). For glucose⁻_(aq), a match in spectral shape is observed only with the spectrum calculated considering deprotonation at C1 (purple dashed line). Our calculations show that if we deprotonate any of the other C sites (blue, orange, red, and green dotted lines), then we are left with a separated minor peak at higher energies or a broad shoulder at lower energies. These features are, however, not observed experimentally. Consequently, the experimentally observed higher energy shoulder discussed in the previous section can be safely assigned to the C1 group, whereas the main C 1s feature stems from five close-lying (protonated) components. A more detailed spectral decomposition analysis will be discussed in the next section, with contributions from both theory and experiment.

Our computations were complemented by quantifying the BEs of the energetically unfavorable noncyclic forms of glucose.⁴² The results are presented in Figure 5, with the

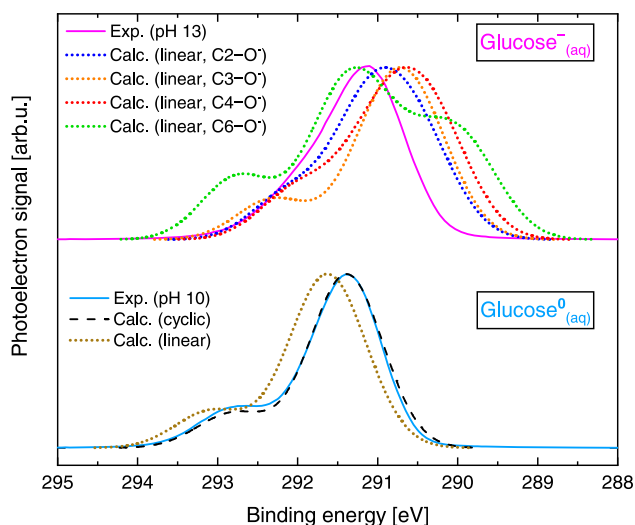


Figure 5. Comparison of experimental (full line) and calculated cyclic and linear (dashed and dotted lines, respectively) C 1s PES spectra of glucose⁰_(aq) (pH 10, bottom) and glucose⁻_(aq) (pH 13, top). The simulated spectral curves were all shifted by the same value, chosen to overlap the pH 10 data, as explained in the text. Calculated spectra of the linear forms of glucose considering deprotonation at different C sites are shown in the top panel.

computed spectra aligned on the BE scale, as discussed for Figure 4. The absolute calculated BE values are shown in Table S2 in the SI but are not considered in the interpretation of the experimental data because an error on the order of few hundreds of millielectronvolts is expected from DFT-based calculations, as performed here. However, more information can be inferred from the structure of the peak. On the basis of a comparison of experimental and calculated spectral shapes for glucose⁰_(aq) (bottom panel), the difference between the cyclic and linear forms is minor. (The linear form has a somewhat larger splitting between the two peaks.) For glucose⁻_(aq) (top panel), spectra from linear structures with a deprotonated hydroxyl group at the C3 and C6 sites exhibit rather pronounced shoulders that are not observed experimentally. Note also that spectra corresponding to acid ionization at C2 and C4 are too wide, and the linear structure

deprotonated at the C5 position does not represent a stable energy minimum. (Calculations for C1 are not shown because there is no C1–OH group in the noncyclic form.) Furthermore, according to our calculations, noncyclic forms are at least 26 kJ/mol (0.27 eV) higher in energy than the cyclic form, so the relative fraction of noncyclic forms should be <0.003%.

Spectral Decomposition of the C 1s PES Spectrum of Glucose. With the aim of identifying the individual spectral contribution from the C1–OH group in the experimental data, we performed a cumulative Gaussian fit analysis of the pH 10 and pH 13 glucose C 1s PES spectra from Figure 3 (glucose⁰_(aq) and glucose⁻_(aq), respectively). The results are presented in Figure 6.

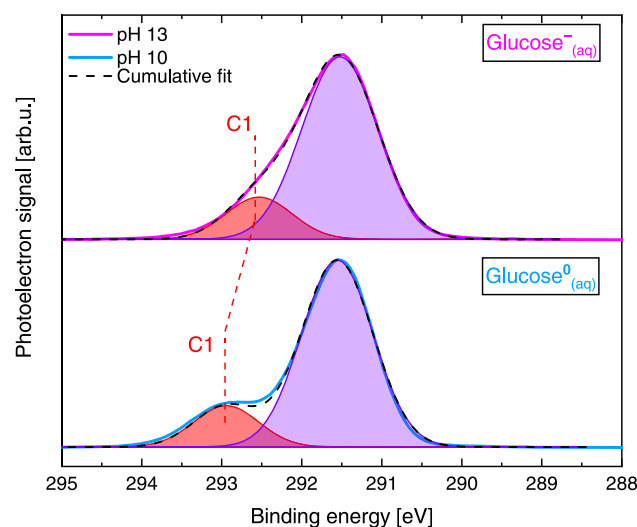


Figure 6. PES spectra of 1 M glucose aqueous solutions at pH 10 and 13, reproduced from Figure 3. The dashed lines indicate cumulative Gaussian fits. Red and purple fills highlight the associated C1 fit component and single-fit component accounting for C2–C6, respectively.

Both the pH 10 (Figure 6, bottom) and pH 13 (Figure 6, top) spectra were fit using two Gaussian curves with the area ratio between the C1 (higher energy shoulder) and the main C2–C6 feature constrained to 1:5. This is indeed a sensible approach because in the pH 10 spectrum, the C1 contribution (highlighted in red) is sufficiently separated from the signal of all of the other C sites (in agreement with our calculations, which indicate a 1.2 eV peak separation; see Table S2 in the SI). The main feature (highlighted in purple) is composed of five C contributions, that is, a convolution of the C2–C6 site signals. The obtained C1 and C2–C6 component BEs are 292.9 ± 0.2 and 291.5 ± 0.2 eV for glucose⁰_(aq) and 292.5 ± 0.2 and 291.5 ± 0.2 eV for glucose⁻_(aq). The 0.4 eV C1 energy shift toward lower BEs in going from glucose⁰_(aq) to glucose⁻_(aq) reflects the larger electron density at the respective carbon site due to the deprotonation of the associated hydroxyl group. As for the valence spectra, the overall error is a combination of the fitting error reported from the least-squares fitting procedure, the experimental resolution, and the error associated with the calibration of the BE scale (calibrated using liquid water's O 1s core level). Because of the somewhat higher photon energy used, the experimental energy resolution is slightly worse, amounting to 0.28 eV (PETRA III) and to 0.38 eV (BESSY II). However, for the C 1s BE values, the

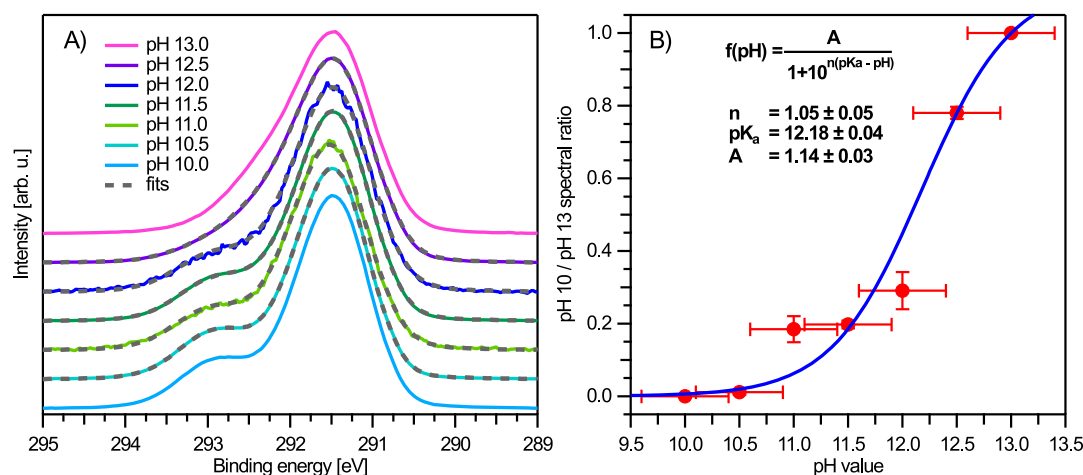


Figure 7. (A) Fits to C 1s PES spectra for intermediate pH values (10.5–12.5) with a model combining the pH 10 (light blue) and pH 13 (magenta) spectra; see the text for details. All spectra were area-normalized and shifted in energy to match the pH 10 spectrum prior to fitting. The vertical offset is added for clarity. (B) Resulting ratio (pH 10/pH 13 spectral contribution) from the fits in panel A as a function of pH value. A fit of a rearranged Henderson–Hasselbalch equation⁸⁴ yields a $\text{pK}_{\text{a}1}$ value of 12.18 ± 0.04 , but considering uncertainties in the pH values when preparing the solutions, 12.2 ± 0.2 is deemed a reasonable result.

calibration errors for the BEs are less of a concern because one can reasonably expect that the O 1s core level is hardly affected by the solute (at least when non-surface-active solutes like glucose are studied). Furthermore, the relative shift of the C1-associated peak can be determined independent of any energy-scale calibration.

The choice of Gaussian fit functions is justified because the aqueous-phase signals are predominantly subject to inhomogeneous (environmental) broadening, yielding principal Gaussian energy broadening terms. We also performed cumulative Voigt-profile fits to capture possible lifetime broadening in core-level photoionization,⁸³ but the results did not converge due to the greater number of degrees of freedom in those fits. However, with an expected 80–100 meV lifetime broadening component of the C 1s peak, which is one order of magnitude smaller than the Gaussian broadening contribution, we do not expect a Voigt fit (either unconstrained or constrained to known lifetime-broadened widths) to yield any appreciable difference in our fit results. In a related context, we have also performed fitting analyses where each C site was represented by a Gaussian with essentially the same peak area and width. Although such a procedure can accurately reproduce the experimental PES spectra, the fits are not unique, and they yield very large fitting errors due to the similar energies of several components. Accordingly, an experimentally meaningful distinction between the individual C2 to C6 BEs is not possible. This explains the use of a single C2–C6 Gaussian in Figure 6, which yields meaningful fitting errors below 100 meV.

Our analysis implies that the C 1s photoionization cross-sections are taken to be identical for the six C sites, which can be considered a reasonable assumption over such a narrow BE range, well above the photoionization threshold. Note that at the near-magic-angle electron collection geometry adopted in the majority of our experiments, the differential photoionization cross-section becomes essentially independent of the β parameter, which could vary with the molecular shape and character at the different C sites.⁵⁶

Determination of $\text{pK}_{\text{a}1}$ and Associated Deprotonated Structures. To determine the $\text{pK}_{\text{a}1}$ value purely from the C 1s PES data, we took the representative experimental spectra of

glucose⁰_(aq) (pH 10) and glucose[−]_(aq) (pH 13) to act as basis curves to fit the spectra associated with the intermediate pH values (10.5–12.5) and determined the ratios of the glucose⁰_(aq) versus glucose[−]_(aq) spectral contributions. All spectra were area-normalized, and the main C 1s feature was overlapped in energy prior to fitting. The basis curves were fit to each spectrum, with the relative basis curve ratio defined as a fitting parameter using the following relation: $\text{Signal}_{\text{pH}10}^{\text{ratio}} + \text{Signal}_{\text{pH}13}^{\text{ratio}}(1 - \text{ratio})$. The results of the fits are shown in Figure 7A, whereas the resulting ratio values are plotted in Figure 7B; the latter can be considered a “photoemission spectroscopy titration”. We also fit the data shown in Figure 7B with a rearranged Henderson–Hasselbalch equation.⁸⁴ The resulting $\text{pK}_{\text{a}1}$ value was found to be 12.18 ± 0.04 . Considering additional uncertainties in the pH values when preparing the solutions, a precision of 0.2, yielding 12.2 ± 0.2 , is reasonable. This result is in excellent agreement with the value reported from titration-based methods ($\text{pK}_{\text{a}1}$ values in the 12.1 to 12.5 range^{36,37,39–41}). The novelty of our LJ-PES approach is the simultaneous determination of the actual deprotonation site, which allows us to associate site-selective spectral changes with a particular acid-ionization constant. Note that our data do not encompass the second acidity constant $\text{pK}_{\text{a}2}$, which is expected at a pH of 13.9.^{36,37} This is also the likely reason that the curve in Figure 7B does not end in a plateau and that rather aqueous-phase glucose is already starting to transition into the double-deprotonated species at higher pH values. Our simple analysis thus does not cover the full picture and is rather meant as a first demonstration of the feasibility of this approach.

To further confirm the assignment of the deprotonation site, we calculated pK_{a} values of glucose upon acid ionization at different C–OH groups. Our results show a robust trend: The pK_{a} value corresponding to acid ionization at C1 is always 1–3 pK_{a} units below the others, irrespective of the method used; the second most easily ionizable C–OH group is located at the C4 site. The calculated values are shown in Table 2, and further analysis is presented in the SI in Table S3. Our results are in agreement with findings by Feng et al.,⁸⁵ who showed that C1 is the best proton donor and is associated with the highest acidity, followed by C4. Notably, though, we observe

Table 2. Glucose Acidity Constants Calculated Using the Thapa and Schlegel⁶⁹ Approach and Assuming a Mixed Explicit/Implicit Model with a Single Solvating Molecule

	C1–OH	C2–OH	C3–OH	C4–OH	C6–OH
calculated pK_{a1}	11.3	15.4	15.5	14.3	17.3

no significant differences in the acidity constants between the two anomeric forms. This latter finding is in contradiction with the report by Feng et al.,⁸⁵ who found pK_{a1} values for C4 and C6 sites to vary between the anomeric forms by more than 2 and 4 pK_a units, respectively. Whereas charge sharing between the two most acidic (C1 and C4) sites has been suggested by Lewis and Schramm³⁹ and cannot be completely excluded based on the C 1s PES experimental data, the present calculations show that C4-deprotonation contributions to the pK_{a1} equilibrium would be negligibly small. The second acid dissociation constant (pK_{a2}) of glucose, assuming deprotonation at C1 followed by deprotonation at C4, was also calculated, yielding a value of 20.8, which is well-separated from the pK_{a1} value.

The values of the acidity constants are controlled by the energetics of the glucose anion produced during deprotonation, which are strongly influenced by the presence of the hydrogen-bonding network, including both glucose–glucose and glucose–water interactions.^{40,46,79,86–88} A summary of the different calculated deprotonated structures of glucose^{−(aq)} together with the respective energies, is shown in Figure 8. Using a relative energy scale, it can be seen that the C1-deprotonated form is energetically preferred.

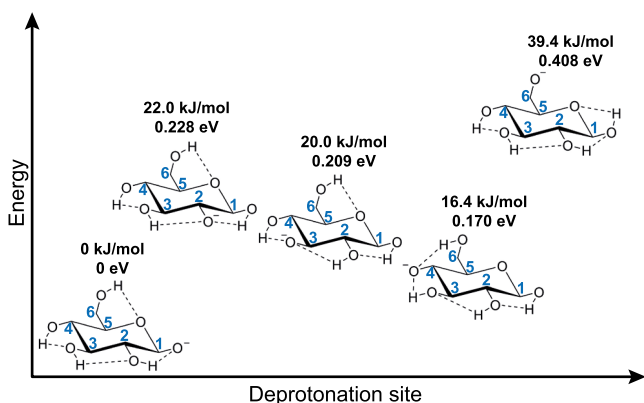


Figure 8. Calculated molecular structures of aqueous-phase β -glucose deprotonated at different C–OH groups. Relative energies (kJ/mol and eV) are also shown and were obtained using free-energy calculations. The presented structures were found as local energetic minima.

CONCLUSIONS

We have investigated the pH-dependent molecular structure changes in glucose by performing LJ-PES experiments over the valence and C 1s spectral regions. With the support of electronic structure calculations, we show how aqueous-phase PES data can be applied to determine the first acid dissociation constant (pK_{a1}) and, more importantly, to unambiguously identify the deprotonation site.

We reported the lowest vertical ionization energies (VIEs), that is, binding energies (BEs) of aqueous-phase glucose from 1 M solutions at pH values below and above pK_{a1} as 9.09 and

7.95 eV, respectively. In addition, our calculations confirmed the signatures of protonated (glucose^{0(aq)}) and deprotonated (glucose^{−(aq)}) species in the experimental data. An experimental VIE of ~ 8.5 eV was determined for glucose^{−(aq)}, almost 1 eV lower than that for its protonated counterpart, glucose^{0(aq)}.

We have also reported two C 1s PES fingerprints of aqueous glucose: a main C 1s photoelectron feature with a BE of 291.5 eV and a less intense, higher energy feature at 293 eV. Our calculations show that the latter originates from the photoionization of the C1–OH group in glucose. We found that the C1–OH C 1s BE shifts by 0.4 eV toward lower values upon the deprotonation of glucose, as evident from data recorded at pH values below and above pK_{a1} as well as from our calculations. Our combined experimental and theoretical approach confirms that at pK_{a1} deprotonation occurs almost exclusively at the C1 site (in contrast, deprotonation at the C4 site is negligibly small) and that a cyclic deprotonated structure prevails.

The sensitivity of LJ-PES to local chemical environments enables us to identify spectral fingerprints of pH-dependent, site-specific deprotonation in glucose. In particular, our studies provide a deeper understanding of the correlation between the molecular structure and the biological function in aqueous-phase glucose as well as in other pyranose-based sugars more generally. Thus we demonstrate the use of solution-phase PES as a general methodology to determine pK_a values, expanding on the capabilities of the technique to investigate the acid–base chemistry and structure–function relationship of polyprotic acids.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at <https://pubs.acs.org/doi/10.1021/acs.jpca.1c04695>.

Calculated valence VIEs for aqueous-phase glucose α - and β -anomers in their protonated (glucose^{0(aq)}) and deprotonated (glucose^{−(aq)}) forms. Table S1. Calculated C 1s VIEs for aqueous-phase α - and β -glucose as well as linear (noncyclic, n-) glucose in their protonated (glucose^{0(aq)}) and deprotonated (glucose^{−(aq)}) forms. Table S2. First and second acidity constants (pK_{a1} and pK_{a2} , respectively) calculated for different C–OH groups in α - and β -glucose. Table S3. Sample input for calculations of valence VIEs. Cartesian coordinates of the structures used to calculate valence VIEs. Sample input for calculations of C 1s VIEs. Cartesian coordinates of the structures used to calculate C 1s VIEs. Sample input for calculations of pK_a values. Cartesian coordinates of the structures used to calculate pK_a values. Sample inputs for calculations of pK_a values via the thermodynamic cycle. Cartesian coordinates of structures used to calculate pK_a values via a thermodynamic cycle (PDF)

AUTHOR INFORMATION

Corresponding Authors

Petr Slaviček – Department of Physical Chemistry, University of Chemistry and Technology, Prague 6 16628, Czech Republic; orcid.org/0000-0002-5358-5538; Email: petr.slavicek@vscht.cz

Bernd Winter – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; orcid.org/0000-0002-5597-8888; Email: winter@fhi-berlin.mpg.de

Authors

Sebastian Malerz – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; orcid.org/0000-0001-9570-3494

Karen Mudryk – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; orcid.org/0000-0002-9046-9915

Lukáš Tomaník – Department of Physical Chemistry, University of Chemistry and Technology, Prague 6 16628, Czech Republic; orcid.org/0000-0003-2547-2488

Dominik Stemer – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; orcid.org/0000-0002-5528-1773

Uwe Hergenroth – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; orcid.org/0000-0003-3396-4511

Tillmann Buttersack – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; orcid.org/0000-0002-4547-2656

Florian Trinter – Fritz-Haber-Institut der Max-Planck-Gesellschaft, 14195 Berlin, Germany; Institut für Kernphysik, Goethe-Universität, 60438 Frankfurt am Main, Germany; orcid.org/0000-0002-0891-9180

Robert Seidel – Operando Interfacial Photochemistry, Helmholtz-Zentrum Berlin für Materialien und Energie, 12489 Berlin, Germany; Institut für Chemie, Humboldt-Universität zu Berlin, 12489 Berlin, Germany; orcid.org/0000-0003-2613-4106

Wilson Quevedo – Operando Interfacial Photochemistry, Helmholtz-Zentrum Berlin für Materialien und Energie, 12489 Berlin, Germany; orcid.org/0000-0002-5222-6891

Claudia Goy – Centre for Molecular Water Science (CMWS), Photon Science, Deutsches Elektronen-Synchrotron (DESY), 22607 Hamburg, Germany; orcid.org/0000-0001-5771-8564

Iain Wilkinson – Department of Locally-Sensitive & Time-Resolved Spectroscopy, Helmholtz-Zentrum Berlin für Materialien und Energie, 14109 Berlin, Germany; orcid.org/0000-0001-9561-5056

Stephan Thürmer – Department of Chemistry, Graduate School of Science, Kyoto University, Kyoto 606-8502, Japan; orcid.org/0000-0002-8146-4573

Complete contact information is available at: <https://pubs.acs.org/10.1021/acs.jpca.1c04695>

Author Contributions

◆ S.M. and K.M. contributed equally.

Notes

The authors declare no competing financial interest. Data relevant for this study have been deposited at [10.5281/zenodo.4777079](https://zenodo.org/record/4777079).

ACKNOWLEDGMENTS

We thank Gerard Meijer, Kevin Pagel, and Maïke Lettow for fruitful discussions and critical comments on the manuscript. K.M. acknowledges Nair M. Eguibar for insightful discussions about the biochemistry of glucose. L.T. and P.S. acknowledge

support by the Czech Science Foundation, project no. 21-26601X (EXPRO). L.T. acknowledges Specific University Research grant no. A2_FCHI_2021_028. F.T. and B.W. acknowledge support from the MaxWater initiative of the Max-Planck-Gesellschaft. B.W. acknowledges funding from the European Research Council (ERC) under the European Union's Horizon 2020 research and investigation programme (grant agreement no. 883759). R.S. and W.Q. acknowledge funding from the German Research Foundation through an Emmy-Noether grant (SE 2253/3-1). S.T. acknowledges support from the JSPS KAKENHI grant no. JP20K15229. We thank Helmholtz-Zentrum Berlin for the allocation of synchrotron radiation beamtime at BESSY II. We acknowledge DESY (Hamburg, Germany), a member of the Helmholtz Association HGF, for the provision of experimental facilities. Parts of this research were carried out at PETRA III, and we thank Moritz Hoesch, in particular, as well as the whole beamline staff, the PETRA III chemistry laboratory and crane operators for assistance in using the P04 soft X-ray beamline. Beamtime was allocated for proposal I-20200682.

ADDITIONAL NOTE

“This is the commonly applied procedure in LJ-PES experiments where solute and solvent energies from aqueous solutions are determined from the energy difference between a solute/solvent peak position and the well-known energies of simultaneously measured water gas-phase $1b_1$ or $1a_1$ (O $1s$) peak features. More precisely, the gas-phase referenced ionization energy of liquid water $1b_{1(l)}$ (or O $1s_{(l)}$) has been used as an internal aqueous-solution reference, although the reference was only determined for neat liquid water. As discussed in ref 89, this gas-phase referencing method generally does not produce accurate absolute liquid-phase BEs because of the typically ill-defined charging of the solution surface. Accurate BEs can, however, be obtained when measuring the respective photoelectron peak along with the so-called low-energy spectral cutoff. Note, though, that accurately determined $1b_1$ and O $1s$ BEs from liquid water would still not serve as useful references to infer solute energies, as they neglect any solute-induced effects on the electronic structure of neat liquid water. Despite this, C $1s$ BEs reported in the present work are expected to be accurate within 200–500 meV of the true values.

REFERENCES

- (1) Seeberger, P. H. Monosaccharide Diversity. In *Essentials of Glycobiology*, 3rd ed.; Varki, A., Cummings, R., Esko, J., Eds.; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, NY, 2017.
- (2) Varki, A. Biological roles of oligosaccharides: all of the theories are correct. *Glycobiology* **1993**, *3* (2), 97–130.
- (3) Krauss, G. *Biochemistry of Signal Transduction and Regulation*; 5th ed.; Wiley-VCH: 2014.
- (4) Hon, D. N. S. Cellulose: a random walk along its historical path. *Cellulose* **1994**, *1* (1), 1–25.
- (5) McNeil, M.; Darvill, A. G.; Fry, S. C.; Albersheim, P. Structure and function of the primary cell walls of plants. *Annu. Rev. Biochem.* **1984**, *53* (1), 625–663.
- (6) Marszalek, P. E.; Oberhauser, A. F.; Pang, Y.-P.; Fernandez, J. M. Polysaccharide elasticity governed by chair–boat transitions of the glucopyranose ring. *Nature* **1998**, *396* (6712), 661–664.
- (7) Beck, E.; Ziegler, P. Biosynthesis and Degradation of Starch in Higher Plants. *Annu. Rev. Plant Physiol. Plant Mol. Biol.* **1989**, *40* (1), 95–117.

- (8) Smith, A. M.; Zeeman, S. C.; Smith, S. M. Starch degradation. *Annu. Rev. Plant Biol.* **2005**, *56* (1), 73–98.
- (9) Adeva-Andany, M. M.; González-Lucán, M.; Donapetry-García, C.; Fernández-Fernández, C.; Ameneiros-Rodríguez, E. Glycogen metabolism in humans. *BBA clinical* **2016**, *5*, 85–100.
- (10) Adeva-Andany, M. M.; Pérez-Felpete, N.; Fernández-Fernández, C.; Donapetry-García, C.; Pazos-García, C. Liver glucose metabolism in humans. *Biosci. Rep.* **2016**, *36* (6), e00416.
- (11) Leegood, R. C. Photosynthesis. In *Encyclopedia of Biological Chemistry*, 2nd ed.; Lennarz, W. J., Lane, M. D., Eds.; Academic Press: Waltham, MA, 2013; pp 492–496.
- (12) Bassham, J. A. Energy capture and conversion by photosynthesis. *J. Theor. Biol.* **1963**, *4* (1), 52–72.
- (13) Pischetsrieder, M. Chemistry of Glucose and Biochemical Pathways of Biological Interest. *Peritoneal Dial. Int.* **2000**, *20* (2 suppl), 26–30.
- (14) Chen, J.; Zhao, C. X.; Zhi, M. M.; Wang, K.; Deng, L.; Xu, G. Alkaline direct oxidation glucose fuel cell system using silver/nickel foams as electrodes. *Electrochim. Acta* **2012**, *66*, 133–138.
- (15) Zhao, C. X.; Wang, K.; Yan, H.; Xu, G. Output Current Increase in Alkaline Glucose Fuel Cells. *J. Electrochem. Soc.* **2011**, *158* (9), B1055.
- (16) Hudson, C. S. Emil Fischer's discovery of the configuration of glucose. A semicentennial retrospect. *J. Chem. Educ.* **1941**, *18* (8), 353.
- (17) Lichtenthaler, F. W. Emil Fischer's Proof of the Configuration of Sugars: A Centennial Tribute. *Angew. Chem., Int. Ed. Engl.* **1992**, *31* (12), 1541–1556.
- (18) Marianski, M.; Supady, A.; Ingram, T.; Schneider, M.; Baldauf, C. Assessing the Accuracy of Across-the-Scale Methods for Predicting Carbohydrate Conformational Energies for the Examples of Glucose and α -Maltose. *J. Chem. Theory Comput.* **2016**, *12* (12), 6157–6168.
- (19) Delbianco, M.; Kononov, A.; Poveda, A.; Yu, Y.; Diercks, T.; Jiménez-Barbero, J.; Seeberger, P. H. Well-Defined Oligo- and Polysaccharides as Ideal Probes for Structural Studies. *J. Am. Chem. Soc.* **2018**, *140* (16), 5421–5426.
- (20) Hofmann, J.; Hahm, H. S.; Seeberger, P. H.; Pagel, K. Identification of carbohydrate anomers using ion mobility–mass spectrometry. *Nature* **2015**, *526* (7572), 241–244.
- (21) Liu, J.; Cukier, R. I.; Bu, Y.; Shang, Y. Glucose-Promoted Localization Dynamics of Excess Electrons in Aqueous Glucose Solution Revealed by Ab Initio Molecular Dynamics Simulation. *J. Chem. Theory Comput.* **2014**, *10* (10), 4189–4197.
- (22) Seidel, R.; Winter, B.; Bradforth, S. E. Valence Electronic Structure of Aqueous Solutions: Insights from Photoelectron Spectroscopy. *Annu. Rev. Phys. Chem.* **2016**, *67* (1), 283–305.
- (23) Williams, H. L.; Erickson, B. A.; Neumark, D. M. Time-resolved photoelectron spectroscopy of adenosine and adenosine monophosphate photodeactivation dynamics in water microjets. *J. Chem. Phys.* **2018**, *148* (19), 194303.
- (24) Erickson, B. A.; Heim, Z. N.; Pieri, E.; Liu, E.; Martinez, T. J.; Neumark, D. M. Relaxation Dynamics of Hydrated Thymine, Thymidine, and Thymidine Monophosphate Probed by Liquid Jet Time-Resolved Photoelectron Spectroscopy. *J. Phys. Chem. A* **2019**, *123* (50), 10676–10684.
- (25) Ramasesha, K.; Leone, S. R.; Neumark, D. M. Real-Time Probing of Electron Dynamics Using Attosecond Time-Resolved Spectroscopy. *Annu. Rev. Phys. Chem.* **2016**, *67* (1), 41–63.
- (26) Angyal, S. J. The Composition and Conformation of Sugars in Solution. *Angew. Chem., Int. Ed. Engl.* **1969**, *8* (3), 157–166.
- (27) Zhu, Y.; Zajicek, J.; Serianni, A. S. Acyclic Forms of [1-¹³C]Aldohexoses in Aqueous Solution: Quantitation by ¹³C NMR and Deuterium Isotope Effects on Tautomeric Equilibria. *J. Org. Chem.* **2001**, *66* (19), 6244–6251.
- (28) Bonner, W. A. The origin and amplification of biomolecular chirality. *Origins Life Evol. Biospheres* **1991**, *21* (2), 59–111.
- (29) Cintas, P.; Viedma, C. On the physical basis of asymmetry and homochirality. *Chirality* **2012**, *24* (11), 894–908.
- (30) Brønsted, J. N.; Guggenheim, E. A. Contribution to the theory of acid and basic catalysis. The mutarotation of glucose. *J. Am. Chem. Soc.* **1927**, *49* (10), 2554–2584.
- (31) Cui, S. W. *Food Carbohydrates: Chemistry, Physical Properties, and Applications*; Taylor & Francis Group: 2005.
- (32) Sugiyama, H.; Usui, T. The Anomeric Equilibrium of Glucose in Acidic and Basic Media. *Agric. Biol. Chem.* **1980**, *44* (12), 3001–3002.
- (33) Mayes, H. B.; Tian, J.; Nolte, M. W.; Shanks, B. H.; Beckham, G. T.; Gnanakaran, S.; Broadbelt, L. J. Sodium Ion Interactions with Aqueous Glucose: Insights from Quantum Mechanics, Molecular Dynamics, and Experiment. *J. Phys. Chem. B* **2013**, *118* (8), 1990–2000.
- (34) Zhao, S.; Guo, X.; Bai, P.; Lv, L. Chemical Isomerization of Glucose to Fructose Production. *Asian J. Chem.* **2014**, *26*, 4537–4543.
- (35) Climent, M. J.; Corma, A.; Iborra, S. Conversion of biomass platform molecules into fuel additives and liquid hydrocarbon fuels. *Green Chem.* **2014**, *16* (2), 516–547.
- (36) Urban, F.; Shaffer, P. A. The acidic property of sugars. *J. Biol. Chem.* **1932**, *94* (3), 697–715.
- (37) Urban, F.; Williams, R. D. The acidic property of sugars II. *J. Biol. Chem.* **1933**, *100* (1), 237–241.
- (38) Shaffer, P. A.; Friedemann, T. E. Sugar activation by alkali: I. Formation of lactic and saccharinic acids. *J. Biol. Chem.* **1930**, *86* (1), 345–374.
- (39) Lewis, B. E.; Schramm, V. L. Isotope Effect-Mapping of the Ionization of Glucose Demonstrates Unusual Charge Sharing. *J. Am. Chem. Soc.* **2003**, *125* (26), 7872–7877.
- (40) Beenackers, J. A. W. M. B.; Kuster, B. F. M.; van der Baan, H. S. Ionisation and solvation of d-glucose. *Carbohydr. Res.* **1985**, *140* (2), 169–183.
- (41) Degani, C. Ionization constants of sugars: a predominant factor in the cyanogen-induced phosphorylation of sugars. *Carbohydr. Res.* **1971**, *18* (2), 329–332.
- (42) de Wit, G.; Kieboom, A. P. G.; van Bekkum, H. Ionization and mutarotation of hexoses in aqueous alkaline solution as studied by ¹³C-NMR spectroscopy. *Tetrahedron Lett.* **1975**, *16* (45), 3943–3946.
- (43) Stevens, J. S.; Schroeder, S. L. M. Quantitative analysis of saccharides by X-ray photoelectron spectroscopy. *Surf. Interface Anal.* **2009**, *41* (6), 453–462.
- (44) Li, M.; Li, W.; Liu, S. Hydrothermal synthesis, characterization, and KOH activation of carbon spheres from glucose. *Carbohydr. Res.* **2011**, *346* (8), 999–1004.
- (45) Qiao, H.; Xiao, L.; Zheng, Z.; Liu, H.; Jia, F.; Zhang, L. One-pot synthesis of CoO/C hybrid microspheres as anode materials for lithium-ion batteries. *J. Power Sources* **2008**, *185* (1), 486–491.
- (46) El Khadem, H. S.; Ennifar, S.; Isbell, H. S. Evidence of stable hydrogen-bonded ions during isomerization of hexoses in alkali. *Carbohydr. Res.* **1989**, *185* (1), 51–59.
- (47) Winter, B.; Faubel, M. Photoemission from Liquid Aqueous Solutions. *Chem. Rev.* **2006**, *106* (4), 1176–1211.
- (48) Nolting, D.; Aziz, E. F.; Ottosson, N.; Faubel, M.; Hertel, I. V.; Winter, B. pH-Induced Protonation of Lysine in Aqueous Solution Causes Chemical Shifts in X-ray Photoelectron Spectroscopy. *J. Am. Chem. Soc.* **2007**, *129* (45), 14068–14073.
- (49) Ottosson, N.; Børve, K. J.; Spångberg, D.; Bergersen, H.; Sæthre, L. J.; Faubel, M.; Pokapanich, W.; Öhrwall, G.; Björneholm, O.; Winter, B. On the Origins of Core–Electron Chemical Shifts of Small Biomolecules in Aqueous Solution: Insights from Photoemission and ab Initio Calculations of Glycine_{aq}. *J. Am. Chem. Soc.* **2011**, *133* (9), 3120–3130.
- (50) Kurahashi, N.; Karashima, S.; Tang, Y.; Horio, T.; Abulimiti, B.; Suzuki, Y.; Ogi, Y.; Oura, M.; Suzuki, T. Photoelectron spectroscopy of aqueous solutions: streaming potentials of NaX (X = Cl, Br, and I) solutions and electron binding energies of liquid water and X. *J. Chem. Phys.* **2014**, *140* (17), 174506.

- (51) Kachel, T. The plane grating monochromator beamline U49/2 PGM1 at BESSY II. *Journal of Large-Scale Research Facilities* **2016**, *2*, A72.
- (52) Seidel, R.; Pohl, M. N.; Ali, H.; Winter, B.; Aziz, E. F. Advances in liquid phase soft-x-ray photoemission spectroscopy: A new experimental setup at BESSY II. *Rev. Sci. Instrum.* **2017**, *88* (7), 073107.
- (53) Viefhaus, J.; Scholz, F.; Deinert, S.; Glaser, L.; Iichen, M.; Seltmann, J.; Walter, P.; Siewert, F. The Variable Polarization XUV Beamline P04 at PETRA III: Optics, mechanics and their performance. *Nucl. Instrum. Methods Phys. Res., Sect. A* **2013**, *710*, 151–154.
- (54) EASI Liquid-Jet PES Instrument Website. Fritz-Haber-Institut, Max-Planck-Gesellschaft, Berlin. <https://www.fhi.mpg.de/236287/experimental-methods> (accessed May 28, 2021).
- (55) Winter, B. Liquid microjet for photoelectron spectroscopy. *Nucl. Instrum. Methods Phys. Res., Sect. A* **2009**, *601* (1–2), 139–150.
- (56) Thürmer, S.; Seidel, R.; Faubel, M.; Eberhardt, W.; Hemminger, J. C.; Bradforth, S. E.; Winter, B. Photoelectron Angular Distributions from Liquid Water: Effects of Electron Scattering. *Phys. Rev. Lett.* **2013**, *111* (17), 173005.
- (57) Yanai, T.; Tew, D. P.; Handy, N. C. A new hybrid exchange–correlation functional using the Coulomb-attenuating method (CAM-B3LYP). *Chem. Phys. Lett.* **2004**, *393* (1), 51–57.
- (58) Mennucci, B.; Tomasi, J. Continuum solvation models: A new approach to the problem of solute's charge distribution and cavity boundaries. *J. Chem. Phys.* **1997**, *106* (12), 5151–5158.
- (59) Cancès, E.; Mennucci, B.; Tomasi, J. A new integral equation formalism for the polarizable continuum model: Theoretical background and applications to isotropic and anisotropic dielectrics. *J. Chem. Phys.* **1997**, *107* (8), 3032–3041.
- (60) Muchová, E.; Slaviček, P. Beyond Koopmans' theorem: electron binding energies in disordered materials. *J. Phys.: Condens. Matter* **2019**, *31* (4), 043001.
- (61) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Petersson, G. A.; Nakatsuji, H.; et al. *Gaussian 09*, revision D.01; Gaussian, Inc.: Wallingford, CT, 2016.
- (62) Pluhařová, E.; Slaviček, P.; Jungwirth, P. Modeling Photoionization of Aqueous DNA and Its Components. *Acc. Chem. Res.* **2015**, *48* (5), 1209–1217.
- (63) Gilbert, A. T. B.; Besley, N. A.; Gill, P. M. W. Self-Consistent Field Calculations of Excited States Using the Maximum Overlap Method (MOM). *J. Phys. Chem. A* **2008**, *112* (50), 13164–13171.
- (64) Shao, Y.; Gan, Z.; Epifanovsky, E.; Gilbert, A. T. B.; Wormit, M.; Kussmann, J.; Lange, A. W.; Behn, A.; Deng, J.; Feng, X.; et al. Advances in molecular quantum chemistry contained in the Q-Chem 4 program package. *Mol. Phys.* **2015**, *113* (2), 184–215.
- (65) Hollas, D.; Pohl, M. N.; Seidel, R.; Aziz, E. F.; Slaviček, P.; Winter, B. Aqueous Solution Chemistry of Ammonium Cation in the Auger Time Window. *Sci. Rep.* **2017**, *7* (1), 756.
- (66) Rappe, A. K.; Casewit, C. J.; Colwell, K. S.; Goddard, W. A.; Skiff, W. M. UFF, a full periodic table force field for molecular mechanics and molecular dynamics simulations. *J. Am. Chem. Soc.* **1992**, *114* (25), 10024–10035.
- (67) Schroeder, C. A.; Pluhařová, E.; Seidel, R.; Schroeder, W. P.; Faubel, M.; Slaviček, P.; Winter, B.; Jungwirth, P.; Bradforth, S. E. Oxidation Half-Reaction of Aqueous Nucleosides and Nucleotides via Photoelectron Spectroscopy Augmented by ab Initio Calculations. *J. Am. Chem. Soc.* **2015**, *137* (1), 201–209.
- (68) Marenich, A. V.; Ho, J.; Coote, M. L.; Cramer, C. J.; Truhlar, D. G. Computational electrochemistry: prediction of liquid-phase reduction potentials. *Phys. Chem. Chem. Phys.* **2014**, *16* (29), 15068–15106.
- (69) Thapa, B.; Schlegel, H. B. Density Functional Theory Calculation of pKa's of Thiols in Aqueous Solution Using Explicit Water Molecules and the Polarizable Continuum Model. *J. Phys. Chem. A* **2016**, *120* (28), 5726–5735.
- (70) Chai, J.-D.; Head-Gordon, M. Long-range corrected hybrid density functionals with damped atom–atom dispersion corrections. *Phys. Chem. Chem. Phys.* **2008**, *10* (44), 6615–6620.
- (71) Marenich, A. V.; Cramer, C. J.; Truhlar, D. G. Universal Solvation Model Based on Solute Electron Density and on a Continuum Model of the Solvent Defined by the Bulk Dielectric Constant and Atomic Surface Tensions. *J. Phys. Chem. B* **2009**, *113* (18), 6378–6396.
- (72) Winter, B.; Weber, R.; Widdra, W.; Dittmar, M.; Faubel, M.; Hertel, I. V. Full Valence Band Photoemission from Liquid Water Using EUV Synchrotron Radiation. *J. Phys. Chem. A* **2004**, *108* (14), 2625–2632.
- (73) Jagoda-Cwiklik, B.; Slaviček, P.; Cwiklik, L.; Nolting, D.; Winter, B.; Jungwirth, P. Ionization of Imidazole in the Gas Phase, Microhydrated Environments, and in Aqueous Solution. *J. Phys. Chem. A* **2008**, *112* (16), 3499–3505.
- (74) Jagoda-Cwiklik, B.; Slaviček, P.; Nolting, D.; Winter, B.; Jungwirth, P. Ionization of Aqueous Cations: Photoelectron Spectroscopy and ab Initio Calculations of Protonated Imidazole. *J. Phys. Chem. B* **2008**, *112* (25), 7355–7358.
- (75) Pluhařová, E.; Ončák, M.; Seidel, R.; Schroeder, C.; Schroeder, W.; Winter, B.; Bradforth, S. E.; Jungwirth, P.; Slaviček, P. Transforming Anion Instability into Stability: Contrasting Photoionization of Three Protonation Forms of the Phosphate Ion upon Moving into Water. *J. Phys. Chem. B* **2012**, *116* (44), 13254–13264.
- (76) Ghosh, D.; Roy, A.; Seidel, R.; Winter, B.; Bradforth, S.; Krylov, A. I. First-Principle Protocol for Calculating Ionization Energies and Redox Potentials of Solvated Molecules and Ions: Theory and Application to Aqueous Phenol and Phenolate. *J. Phys. Chem. B* **2012**, *116* (24), 7269–7280.
- (77) Winter, B.; Faubel, M.; Hertel, I. V.; Pettenkofer, C.; Bradforth, S. E.; Jagoda-Cwiklik, B.; Cwiklik, L.; Jungwirth, P. Electron Binding Energies of Hydrated H₃O⁺ and OH⁻: Photoelectron Spectroscopy of Aqueous Acid and Base Solutions Combined with Electronic Structure Calculations. *J. Am. Chem. Soc.* **2006**, *128* (12), 3864–3865.
- (78) Pohl, M. N.; Muchová, E.; Seidel, R.; Ali, H.; Sršen, Š.; Wilkinson, I.; Winter, B.; Slaviček, P. Do water's electrons care about electrolytes? *Chemical Science* **2019**, *10* (3), 848–865.
- (79) Cramer, C. J.; Truhlar, D. G. Quantum chemical conformational analysis of glucose in aqueous solution. *J. Am. Chem. Soc.* **1993**, *115* (13), 5745–5753.
- (80) Alonso, J. L.; Lozoya, M. A.; Peña, I.; López, J. C.; Cabezas, C.; Mata, S.; Blanco, S. The conformational behaviour of free d-glucose— at last. *Chemical Science* **2014**, *5* (2), 515–522.
- (81) Tomaník, L.; Muchová, E.; Slaviček, P. Solvation energies of ions with ensemble cluster-continuum approach. *Phys. Chem. Chem. Phys.* **2020**, *22* (39), 22357–22368.
- (82) Winter, B.; Aziz, E. F.; Hergenbahn, U.; Faubel, M.; Hertel, I. V. Hydrogen bonds in liquid water studied by photoelectron spectroscopy. *J. Chem. Phys.* **2007**, *126* (12), 124504.
- (83) Campbell, J. L.; Papp, T. Widths of the atomic K–N7 levels. *At. Data Nucl. Data Tables* **2001**, *77* (1), 1–56.
- (84) Hasselbalch, K. A. Die Berechnung der Wasserstoffzahl des Blutes aus der freien und gebundenen Kohlensäure desselben, und die Sauerstoffbindung des Blutes als Funktion der Wasserstoffzahl. *Biochemische Zeitschrift* **1917**, *78*, 112–144.
- (85) Feng, S.; Bagia, C.; Mpourmpakis, G. Determination of Proton Affinities and Acidity Constants of Sugars. *J. Phys. Chem. A* **2013**, *117* (24), 5211–5219.
- (86) Molteni, C.; Parrinello, M. Glucose in Aqueous Solution by First Principles Molecular Dynamics. *J. Am. Chem. Soc.* **1998**, *120* (9), 2168–2171.
- (87) Çarçabal, P.; Jockusch, R. A.; Hüniig, I.; Snoek, L. C.; Kroemer, R. T.; Davis, B. G.; Gamblin, D. P.; Compagnon, I.; Oomens, J.; Simons, J. P. Hydrogen Bonding and Cooperativity in Isolated and Hydrated Sugars: Mannose, Galactose, Glucose, and Lactose. *J. Am. Chem. Soc.* **2005**, *127* (32), 11414–11425.

(88) Suzuki, T. The hydration of glucose: the local configurations in sugar–water hydrogen bonds. *Phys. Chem. Chem. Phys.* **2008**, *10* (1), 96–105.

(89) Thürmer, S.; Malerz, S.; Trinter, F.; Hergenbahn, U.; Lee, C.; Neumark, D. M.; Meijer, G.; Winter, B.; Wilkinson, I. Accurate Vertical Ionization Energy and Work Function Determinations of Liquid Water and Aqueous Solutions. *Chem. Sci.* **2021**, DOI: [10.1039/D1SC01908B](https://doi.org/10.1039/D1SC01908B).