

***Aldh2* gene reduces oxidative stress in the bladder by regulating the NF- κ B pathway in a mouse model of ketamine-induced cystitis**

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Abstract. Aldehyde dehydrogenase 2 (*aldh2*) serves an important role in the development of organ injury. Therefore, the present study investigated the effects of *aldh2* on the oxidative stress response in a mouse model of ketamine-induced cystitis (KIC). A total of 60 8-week-old male Institute of Cancer Research wild-type (WT) mice and 45 *aldh2* knock-out (KO) mice were randomized to receive low-dose ketamine (30 mg/kg), high-dose ketamine (60 mg/kg) or normal saline (controls). At 4, 8 and 12 weeks post-injection, bladder tissues were harvested and used to investigate the protective mechanisms of *aldh2* on bladder function. The results demonstrated that *aldh2* KO mice exhibited significant weight loss following chronic ketamine injection compared with that in WT mice. Furthermore, ketamine treatment increased the urination rate ($P < 0.05$), pathological score ($P < 0.05$), levels of the oxidative stress product malondialdehyde ($P < 0.05$) in addition to reducing the expression of the anti-oxidative stress enzyme superoxide dismutase ($P < 0.05$) and glutathione-SH ($P < 0.05$). Oxidative stress in *aldh2* KO mice was also found to significantly enhance the expression of proteins associated with the NF- κ B signaling pathway, which promoted the expression of inducible nitric oxide synthase ($P < 0.05$) and cyclooxygenase-2 ($P < 0.05$) further. Finally, *aldh2* KO mice demonstrated higher severity of fibrosis in the submucosal and muscular layers of the bladder. In conclusion, the present study suggests that *aldh2* serves a protective role in preventing inflammation and fibrosis in KIC.

Introduction

Ketamine, also known as K powder, is a substance that has been widely abused in recent years (1). Chronic ketamine abusers frequently suffer from severe lower urinary tract syndrome (LUTS), which is characterized by the excessive frequency of urination, nocturia, suprapubic discomfort and occasional hematuria (2). Pathologically, it is similar to interstitial cystitis, also known as ketamine-induced cystitis (KIC) (3). However, the pathophysiology of bladder dysfunction in patients with KIC remains to be elucidated.

Accumulating evidence has indicated that oxidative stress-mediated injury serves an important role in KIC (4,5). During oxidative stress, ATP production from the mitochondrial respiratory chain triggers bladder hyperactivity. Ketamine injections increase the production of intracellular reactive oxygen species (ROS), which stimulates electron leakage from the mitochondrial respiratory chain complexes (4). ROS can be removed from the body by enzymes (for example, SOD and GSH-px) and non-enzymatic (for example GSH, Vit C and Vit E) antioxidants, which serves as the main defense system (6). During oxidative stress, high ROS levels induce lipid peroxidation and the formation of malondialdehyde (MDA), which is an end-product of oxidation that influences the mitochondrial respiratory chain complex and the activities of key enzymes (For example, SOD and GSH) (7). Superoxide dismutase (SOD) and glutathione-sulfhydryl (GSH) are the main antioxidant enzymes that degrade and scavenge free radicals *in vivo* (8). A previous study revealed that chronic ketamine treatment increased MDA levels in the rat bladder and reduced the expression of antioxidant enzymes SOD and GSH (Mi *et al*, 2016, unpublished data). This indicated that ketamine can target the bladder urothelium to release ROS and induce inflammation in the bladder. The release of ROS in bladder urothelium is mediated by the cytochrome C oxidase pathway, which induces detrusor hyperactivity. Furthermore, oxidative stress triggers the production of large quantities of proinflammatory mediators nitric oxide (NO) and prostaglandin E2 from macrophages. These products are generated by inducible nitric oxide synthase (iNOS) and cyclooxygenase-2 (COX-2), respectively (9). These inflammatory mediators increase vascular permeability in the urothelium, resulting in ulceration, erythrocyte accumulation (hemorrhage), monocyte infiltration and increased interstitial fibrosis between the detrusor smooth muscle tracts in the rat bladder injured by ketamine treatment (10).

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Abbreviations: WT, wild-type; KO, knock-out; HK, high dose ketamine group; KNS, knock-out normal saline control group; KHK, knock-out high dose ketamine group; WNS, wild-type normal saline control group; WHK, wild-type high dose ketamine group

Key words: ketamine-induced cystitis, aldehyde dehydrogenase 2, oxidative stress, cyclooxygenase-2, inducible nitric oxide synthase

Aldehyde dehydrogenase 2 (*aldh2*) is a mitochondrial enzyme that regulates aldehyde metabolism by eliminating cytotoxic aldehydes, thereby reducing oxidative stress and inhibiting the production of ROS-related toxic products (11). In particular, ~40% of the population in Asia has a defective *aldh2* gene compared with that in Europe and Africa, where the prevalence is <5% (12). Individuals with the *aldh2* allele deficiency are highly susceptible to adverse reactions to ethanol and other stimulatory factors (For example, 4-hydroxy-2-nonenal and hypoxia) due to the accumulation of aldehydes caused by the lack of *aldh2* enzymes. *Aldh2* has been previously identified as a potential prognostic marker for bladder urothelial carcinoma (13). In a follow-up study performed in the USA, *aldh2* variants were found to be associated with a shorter time to first recurrence of bladder cancer (14). This finding was supported by another previous study by Ferreira-Teixeira *et al* (15), who revealed that *aldh2* has the potential to predict the progression and metastasis of invasive bladder cancer. Although a number of studies have demonstrated the anti-oxidant effect of *aldh2* in bladder tumors, its role in cystitis remains poorly understood.

It has been previously reported that ketamine treatment induces the translocation of the NF- κ B subunit p65 into the nucleus, activates COX-2 expression and production of prostaglandin E2 in bladder tissues (16). As an important anti-oxidative stress protein in the body, *aldh2* has been found to suppress the production of ROS and the related toxic aldehyde product MDA, in turn inhibiting apoptosis by suppressing the NF- κ B signaling pathway in human pulmonary artery smooth muscle cells (5). Based on these aforementioned findings, the present study hypothesized that *aldh2* may serve a role in the development of KIC.

Materials and methods

Animal models. A total of 45 *aldh2* knock-out (*aldh2*KO) and 60 wild-type (WT) male Institute of Cancer Research (ICR) mice (weight, 25±5 g; age, 8 weeks) were used for the present study. The *aldh2*KO mice were obtained from the Genomics Center Laboratory of Guangxi Medical University (Nanning, China; Fig. S1). The genotype of the mice was verified by PCR using tail tissues cut from the *aldh2* KO mice (17). The *aldh2* specific primer (DNA) was designed on the BLAST website (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>): Forward, 5'-GCTGGGCTGACAAAGTACCAT-3' and reverse, 5'-TTGATCAAGTTGGCCACGTA-3' (Takara Bio Technology Co., Ltd.). The *aldh2* genotype was verified by Takara Taq™ Version 2.0 (Code No. R004Q; Takara Biotechnology Co., Ltd.) according to the manufacturer's instructions. The thermocycling conditions were as follows: Initial denaturation at 98°C for 10 sec, followed by 30 cycles of 55°C for 30 sec and 72°C for 1 min. Agarose gel (1.5%) and ethidium bromide (0.5 µg/ml) were used in the experiment. WT mice were purchased from Hunan Slack Jingda Experimental Animal Co., Ltd. All mice were kept at 23±1°C and 50±5% humidity, with 12 h light/dark cycles and had free access to food and water. The present study was approved by the Ethics Committee of Guangxi Medical University (approval no. 20180129).

Firstly, 15 WT mice were randomly divided into the ketamine group, which received 30 mg/kg for 4 and 8 weeks to produce the ICR mouse model or the saline group, which

received normal saline (saline group) for 4 and 8 weeks, according to the ICR mouse model previously established by Yeung *et al* (18). Four groups were used with 3-4 WT mice within each group. Ketamine hydrochloride injections (100 mg/2 ml) were purchased from China Fujian Gutian Pharmaceutical Co., Ltd. Injections were administered by an intraperitoneal injection every day at 9 am, where an injection volume of 1 ml liquid volume (saline or ketamine solution) was used to mimic chronic ketamine abuse. All mice were weighed weekly to adjust the doses of ketamine given (30 mg/kg). Mice were sacrificed by cervical dislocation following the 4- and 8-week treatments after an anesthetic (40 mg/kg intravenous pentobarbital; Sigma-Aldrich; Merck KGaA) was administered. Bladder tissues were subsequently obtained and the expression levels of *aldh2* was quantified using quantitative PCR (qPCR), as aforementioned.

In addition, 45 WT and 45 KO mice were randomly divided into the following groups: i) WT normal saline control (WNS); ii) KO normal saline control (KNS); iii) WT low-dose ketamine (WLK; 30 mg/kg); iv) KO low-dose ketamine (KLK; 30 mg/kg); v) WT high-dose ketamine (WHK; 6 mg/kg); and vi) KO high-dose ketamine (KHK; 60 mg/kg). Each group was then further divided into 3 subgroups (4, 8 and 12 weeks, 5 mice in each). Following ketamine administration, mice bladder tissues were obtained from each mouse following euthanasia at 4, 8 and 12 weeks as aforementioned.

Micturition behavior. Micturition frequency was monitored as previously described by Gu *et al* (19). Briefly, the short-term micturition frequency of freely moving mice was observed at the end of weeks 4, 8 and 12 following ketamine treatment. At these time-points, mice were placed in a metabolic cage containing a mesh filter pad, where the short-term urination frequency was recorded. A filter paper was soaked with a saturated copper sulfate solution ($\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$) which was dehydrated at 200°C for 1 h prior to use. When the urine come to contact with the filter paper, the anhydrous CuSO_4 was rehydrated and turned blue. After allowing the mice to move freely for 2 h, the filter paper was removed from the cage and the number of urination events was determined by counting the number of blue dots on the filter paper. Overlapping urination points with distinctly different edges were considered separate urination events and urine points >0.2 cm in diameter were counted (Fig. S2).

Determination of ketamine metabolites in serum and urine. A total of 1 ml of blood was obtained from the mouse's tail. Then the blood was separated by centrifugation at 2,000 x g for 10 min at 4°C. The concentration of ketamine and norketamine in the serum and urine (1 ml) was determined using high-performance liquid chromatography. Briefly, samples were collected on the day before the mice were euthanized. The final mobile phase was prepared via mixing ammonium bicarbonate solution (5 mM) adjusted with concentrated ammonia to a pH of 11.3 and acetonitrile in a ratio of 70:30. A Phenomenex LUX® AMP (Phenomenex) 3 µm, 150x4.6 mm column served as the stationary phase. All chemicals were of analytical grade. Chiral separation experiments were carried out with an Agilent 1260 Series Liquid Chromatograph (Agilent Technologies Inc.), equipped with an autosampler

and a diode array detector. Each analysis was performed at ambient column temperature or a column temperature of 40°C. The measurements were performed under isocratic conditions with a flow rate of 0.5 ml/min and an injection volume of 1 µl. Ultraviolet detection was performed at 200 nm. Data evaluation was performed via a ChemStation for LC 3D Systems Rev. C. 01.07SR2 software (Agilent Technologies GmbH).

Measurement of oxidative stress parameters. Bladder tissues were quickly excised from mice and washed thoroughly with ice-cold normal PBS (pH 7.2). Tissues were sectioned into small pieces by the shear with liquid nitrogen and then homogenized using a glass homogenizer in ice-cold PBS. Next, the solution was centrifuged at 20,000 rpm (41,800 x g) for 10 min at 4°C. The supernatant was used to estimate the levels of oxidative stress indicators SOD, GSH and MDA using their respective ELISA kits (SOD, cat. no. 706002; GSH, cat. no. 703002; MDA, grant no. 700870; Cayman Chemical Company), according to the manufacturer's protocols.

Histopathology and immunohistochemical examinations. Bladder tissues were fixed in 4% phosphate-buffered paraformaldehyde for one day at room temperature, dehydrated in an ascending ethanol gradient, cleared in xylene and embedded in paraffin. The paraffinized tissues were then cut into 5-µm sections and stained with hematoxylin and eosin (H&E) and Masson's trichrome for 5 min at room temperature. Sections were then examined under a light microscope (magnification, x100, x200, x400; Olympus Corporation).

Immunohistochemical examination was performed using the Zhongshan Jinqiao Detection kit (OriGene Technologies, Inc.). Briefly, the tissue sections were deparaffinized and immersed in 3% H₂O₂ for 30 min to quench endogenous peroxidase activity. After being blocked with 2% BSA at room temperature for 30 min, the sections were incubated with iNOS antibody (1:400; cat. no. AF0199; Affinity Biosciences), COX-2 antibody (1:500; cat. no. 12282; Cell Signaling Technology, Inc.) overnight at 4°C. Following incubation with the primary antibodies, the tissue sections were incubated with the appropriate biotinylated secondary antibody (1:5,000; cat. no. TA130016; OriGene Technologies, Inc.) for 30 min at room temperature; after which, they were incubated with 3,3'-diaminobenzidine for 2 min and lightly counterstained with hematoxylin for 1 min at room temperature. Lastly, sections were then examined under a light microscope (magnification, x200; Olympus Corporation).

Western blotting. Harvested bladder tissues (10 mg) were homogenized in liquid nitrogen and re-suspended. The lysates were centrifuged at 14,500 x g for 15 min at 4°C. Protein concentrations were quantified using a bicinchoninic acid Protein Assay kit (Pierce; Thermo Fisher Scientific, Inc.) and 5X protein loading buffer (Beyotime Institute of Biotechnology) was added to the supernatant (specifically 1:4). The samples (25 µg total protein/lane) were separated using 10% SDS-PAGE gel electrophoresis and then transferred onto PVDF membranes. After that, membranes were blocked with 5% BSA at room temperature for 30 min. The membranes were incubated with primary antibodies at 4°C overnight and then washed with TBS-supplemented with

0.1% Tween-20. The primary antibodies used were as follows: iNOS (1:1,000; cat. no. 131205; Cell Signaling technology, Inc.), Aldh2 (1:1,000; cat. no. 108306; Abcam), COX-2 (1:1,000; cat. no. 12282; Cell Signaling technology, Inc.), NF-κB (1:1,500; cat. no. 8242; Cell Signaling technology, Inc.), α-smooth muscle actin (α-SMA; 1:1,000; cat. no. 68463; Cell Signaling technology, Inc.), transforming growth factor-β (TGF-β; 1:1,000; cat. no. 3711; Cell Signaling technology, Inc.), fibronectin (1:1,000; cat. no. AF5335; Affinity Biosciences) and β-actin (1:5,000; cat. no. 12262; Cell Signaling technology, Inc.). This was followed by incubation with the horseradish peroxidase (HRP)-conjugated secondary antibodies (1:1,000; cat. no. A0208; Beyotime Institute of Biotechnology) at 37°C for 1 h. The membranes were next incubated with Novex ECL HRP chemiluminescent substrate reagent kits (Invitrogen; Thermo Fisher Scientific, Inc.) for 30 sec at room temperature and exposed to X-ray film. Protein signals were quantified by scanning densitometry using a FluorChem Q system (Alpha Innotech Corporation). The results of western blotting were quantified using Quantity One Version 4.4.0 software (Bio-Rad Laboratories, Inc.).

Reverse transcription (RT)-qPCR (RT-qPCR)

Total RNA extraction and RT. Minced bladder tissues were treated with TRIzol[®] reagent (Thermo Fisher Scientific, Inc.) according to the manufacturer's protocol, which were fully lysed and mixed. A total of 50% by volume of chloroform was added and the upper aqueous phase was collected following centrifugation (4°C; 15,000 x g). Equal volumes of isopropanol were added and RNA was precipitated by centrifugation (4°C; 15,000 x g). RNA was washed with 70% alcohol, dried and dissolved in RNA-free enzyme water. Subsequently, the purity and concentration of the isolated RNA were detected using an ultra-micronucleic acid detector AIGS (SuiZhen Biotechnology Co., Ltd.). RNA was reverse-transcribed to cDNA using a reverse transcription kit MonAmp[™] Mix (cat. no. MR05201; Monad Biotech Co., Ltd.), according to the manufacturer's protocol. Obtained cDNA was stored at -20°C until use.

qPCR and data analysis. The qPCR reaction system consisted of 5 µl SYBR Green Premix Taq (cat. no. RN04006M; Monad Biotech Co., Ltd.), 1 µl cDNA, 0.3 µl forward primer (10 µM), 0.3 µl reverse primer (10 µM) and 3.4 µl H₂O. The thermocycling conditions were as follows: Initial denaturation at 95°C for 30 sec, followed by 40 cycles of 95°C for 5 sec, 60°C for 30 sec and 72°C for 15 sec. Dissolution curve was produced using the standard dissolution curve program (Agilent Aria Software; version 1.5; Agilent Technologies, Inc.). The PCR reaction was carried out on an ABI 7500 quantitative PCR instrument (Thermo Fisher Scientific, Inc.). The relative mRNA expression data were calculated using the 2^{-ΔΔC_q} method (20). The relative expression value of a gene was normalized against the expression of Actb from control group (WNS) mRNA. The Data was analyzed using the SPSS 22.0 software (IBM Corp). Primer sequences are presented in Table I.

Statistical analysis. In the aldh2 gene expression experiment, 4 mice were used. While in the rest of the experiments, 5 mice were analysed. All data are presented as mean ± standard error

Table I. Oligonucleotide sequences of the primer pairs used for reverse transcription-quantitative PCR.

Primer	Forward sequence (5'-3')	Reverse sequence (5'-3')
β -actin	CAGCCTTCCTTCTTGGGTAT	TGGCATAGAGGTCTTTACGG
COX-2	CAGATGACTGCCCAACTCCC	TGAACCCAGGTCTCTCGCTTA
iNOS	TGGAGCGAGTTGTGGATTGT	GTGAGGGCTTGGCTGAGTGA
NF- κ B	ACACGAGGCTACAACCTCTGC	GGTACCCCCAGAGACCTCAT
α -SMA	GTACCCAGGCATTGCTGACA	GCTGGAAGGTAGACAGCGAA
TGF- β	AGGGCTACCATGCCAACTTC	CCACGTAGTAGACGATGGGC
Fibronectin	ATGAGAAGCCTGGATCCCCT	GGAAGGGTAACCAGTTGGGG
<i>Aldh2</i>	TTCGGGGACGTAAAAGACGG	GGTGTCTTCTCCGGCATAG

COX-2, cyclooxygenase 2; iNOS, inducible nitric oxide synthase; α -SMA, α -smooth muscle actin; TGF- β , transforming growth factor β ; *aldh2*, aldehyde dehydrogenase 2.

of the mean. Statistical analyses were performed using the Graph Pad Prism software (version 7.0; GraphPad Software, Inc.). Mean differences were compared with one-way ANOVA followed by multiple comparison by Tukey's test. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

*Ketamine increases *aldh2* mRNA expression in WT mice.* The mRNA expression levels of *aldh2* in bladder tissues from WT mice were measured after ketamine treatment. At 4 weeks, the relative *aldh2* mRNA levels were found to be 1.06 ± 0.37 in the saline group and 1.38 ± 0.15 in the ketamine group (Fig. 1A). There was no significant difference between these two groups ($P = 0.078$). At 8 weeks, the relative mRNA levels of *aldh2* in the WT mice in the ketamine treatment group was 1.84 ± 0.24 , which was significantly higher compared with 1.02 ± 0.15 in the saline group ($P < 0.01$; Fig. 1A). These results suggest that long-term treatment with ketamine increased the expression of *aldh2* mRNA in the bladder tissues of WT mice.

*Ketamine treatment increases the frequency of urination and suppresses weight gain in *aldh2* KO mice.* Results of *Aldh2* KO and WT mice were compared, which revealed that prolonged ketamine treatment caused urinary dysfunction (Fig. 1B) and suppressed weight gain (Fig. 1C) in KO mice. At week 8, the frequency of urination in the KHK group was 10.33 ± 0.47 , which was significantly higher compared with 8.00 ± 0.82 in the WHK group ($P < 0.05$; Fig. 1B). Similar trends were recorded at week 12 (KHK vs. WHK; 13.33 ± 0.47 vs. 10.67 ± 0.47 ; $P < 0.01$; Fig. 1B). In terms of body weight, the weight of mice in the HK group at week 12 was 28.76 ± 0.93 g in WT mice and 27.67 ± 1.12 g in KO mice ($P = 0.032$), where the average weight gain from the beginning of the experiment to week 12 was 9.12 ± 1.03 g in WT mice and 7.68 ± 0.69 g in KO mice ($P = 0.042$) in the HK group (Fig. 1C). Similar results were observed in the LK group (Fig. 1C). There were no significant differences in urination frequency or body weight changes between WT and KO mice in the NS group. These results suggest that ketamine treatment significantly hindered weight gain whilst increasing urination frequency in KO mice compared with that in WT mice.

*Ketamine metabolism is unaffected by *aldh2* KO.* The potential effects of *aldh2* on ketamine metabolism in WT and *aldh2* KO mice was next investigated. According to the serum and urine measurements of ketamine and norketamine, the levels in the *Aldh2* KO group did not differ significantly compared with those in the WT group (Table II). These observations suggest that ketamine and its metabolites induced similar toxic effects in bladder tissues in WT and KO mice.

*Ketamine increases oxidative stress parameters in *aldh2* KO mice.* ELISA were performed to quantify the levels of oxidative stress factors in bladder tissues. Results in Fig. 2A demonstrated the effects of different treatment periods and doses of ketamine on oxidative stress in the mouse bladder tissues at 12 weeks in all groups and at 4, 8 and 12 weeks in the HK group. These assays reported that levels of anti-oxidation factors SOD ($P < 0.01$) and GSH ($P < 0.01$) were significantly decreased, whilst levels of the lipid peroxidation indicator MDA ($P < 0.01$) was significantly increased in *aldh2* KO mice compared with those in the WT mice at week 12 in the HK group. At week 12, the KHK group exhibited a significant decline in the level of SOD (5.05 ± 0.56 U/ml) and GSH (49.75 ± 13.45 μ M) and a significant increase in MDA (70.79 ± 8.28 nmol/ml) in bladder tissues compared with WHK (SOD, 10.35 ± 1.04 U/ml; $P < 0.01$; GSH, 124.49 ± 26.02 μ M; $P < 0.05$; MDA, 41.97 ± 5.01 nmol/ml; $P < 0.01$).

Ketamine induces more severe pathological damage in KO mice. At week 12, modest inflammatory cell infiltration and massive intravascular congestion were observed via H&E staining in the submucosal layer of the bladder tissues of mice in the WHK group (Fig. 2B). In tissues from mice in the KHK group, this appeared to be more severe, where the mucosal barrier had disintegrated with numerous inflammatory cells accumulated in the submucosa and extensive edema in the bladder mucosa (Fig. 2B). No edema could be observed in the bladder walls of tissues from the WNS and KNS control groups, where the bladder mucosal barrier was intact (Fig. 2B). All these qualitative descriptions were pointed out by using arrows. Immunohistochemical analysis revealed that COX-2 and iNOS protein expression in KO mice was enhanced beneath the bladder mucosa compared with

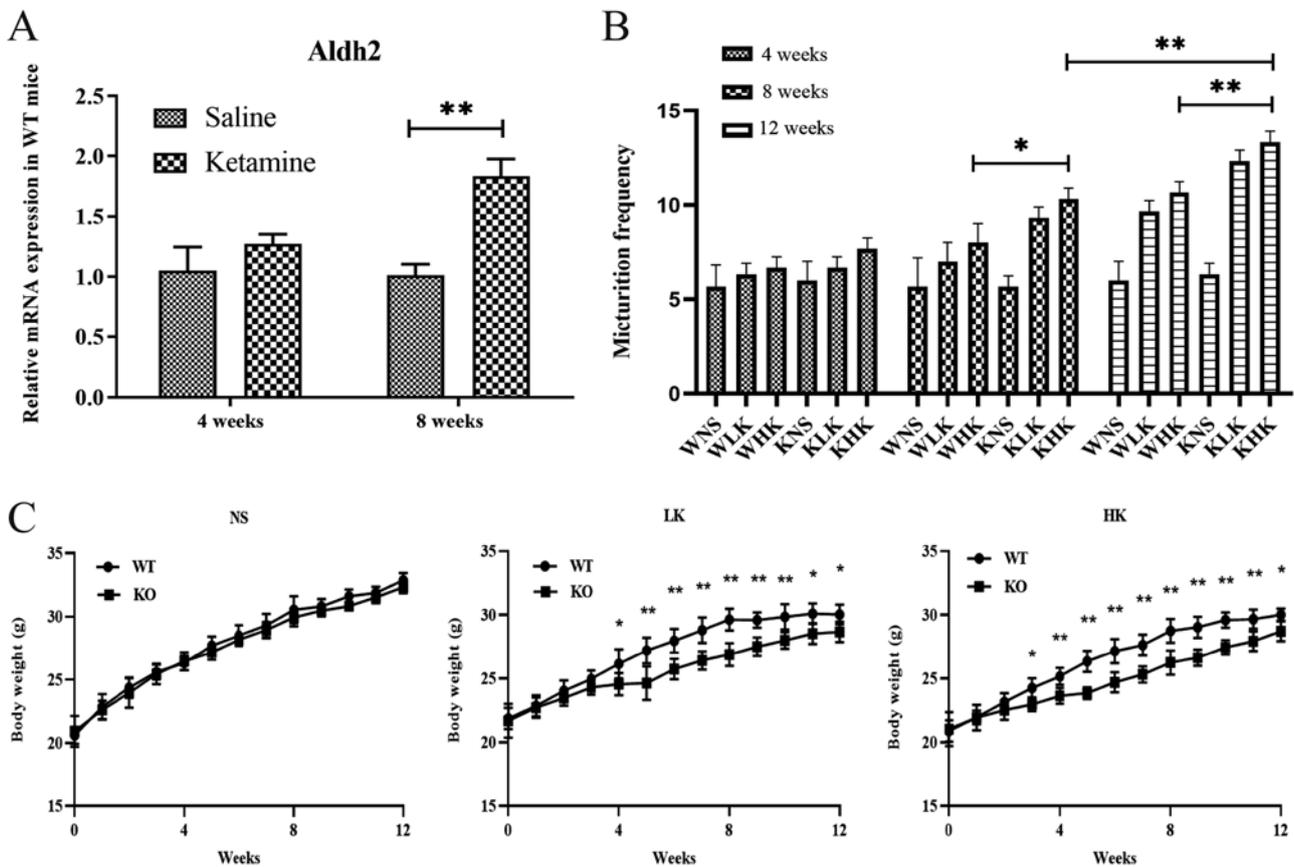


Figure 1. Ketamine treatment increases *aldh2* mRNA expression in WT mice and increases urination frequency and reduces body weight in KO mice. (A) Reverse transcription-quantitative PCR was performed to measure the mRNA expression of *aldh2* in WT mice at 4 and 8 weeks following saline and ketamine treatment. (B) Changes in urination frequency of KO and WT mice during 4-, 8- and 12-week ketamine treatments. (C) Weekly weight changes of KO and WT mice (each week, group WT vs. group KO). Data were presented as the mean \pm standard error of the mean from ≥ 3 experimental repeats. * $P < 0.05$ and ** $P < 0.01$. Aldh2, aldehyde dehydrogenase 2; WT, wild-type; KO, knock-out; WNS, wild-type normal saline control group; WLK, wild-type low-dose ketamine group; WHK, wild-type high-dose ketamine group; KNS, knock-out normal saline control group; KLK, knock-out low-dose ketamine group; KHK, knock-out high-dose ketamine group; NS, normal saline; LK, low-dose ketamine; HK, high-dose ketamine; w, week.

WT mice (Fig. 2C). In week 4, COX-2 protein expression in KHK group increased more than WHK and iNOS protein expression in KHK and WHK group showed little difference. When in week 12, the staining densities of COX-2 and iNOS in the KHK group were markedly higher compared with those in the WHK group (Fig. 2C). Masson trichrome staining of the bladder tissues revealed that the bladder submucosa and muscular layers were not notably affected in the WNS and KNS groups at week 12 (Fig. 2D). By contrast, the lamina propria and submucosa structure had disintegrated in the WHK and KHK groups, where the tissues exhibited diffuse interstitial fibrosis. Furthermore, the area infiltrated by collagen fibers was larger and the thickening of the knot tissues was clearer in the KHK group compared with that in the WHK group. All these qualitative descriptions are presented by arrows.

Inflammation of ketamine-induced cystitis is more severe in KO mice. Furthermore, the expression of NF- κ B, iNOS and COX-2, indicators associated with inflammation, was verified by measuring their protein and mRNA levels. The results demonstrated that in the KHK group, the protein expression levels of COX-2, iNOS and NF- κ B were increased by 1.63 ($P < 0.01$), 1.41 ($P < 0.01$) and 1.51 ($P < 0.01$) fold, respectively,

compared with those in the WHK group in week 12, in a concentration-dependent manner (Fig. 3A). In addition, when comparing the LK groups (week 12) or week 8 (HK groups), the results also demonstrated that NF- κ B ($P < 0.05$), iNOS ($P < 0.05$) and COX-2 ($P < 0.01$) protein expression was enhanced in KO mice compared with those in WT mice. At 12 weeks in the HK group, the mRNA levels of NF- κ B, COX-2 and iNOS were significantly higher (all $P < 0.01$) in the KO group (2.620 ± 0.350 , 2.903 ± 0.188 and 3.307 ± 0.242 , respectively) compared with those in the WT group (1.884 ± 0.311 , 1.787 ± 0.116 and 2.083 ± 0.286 , respectively; Fig. 3B). Furthermore, when comparing LK groups (week 12) or week 8 (HK groups), the results also demonstrated that NF- κ B ($P < 0.05$), iNOS ($P < 0.05$) and COX-2 ($P < 0.05$) protein expressions were enhanced in KO mice compared with those in WT mice.

Fibrosis levels in ketamine-induced cystitis is more severe in KO mice. Additionally, the protein expression levels of fibrotic markers α -SMA ($P < 0.01$), TGF- β ($P < 0.01$) and fibronectin ($P < 0.05$) were found to be significantly higher in mice in the KHK group compared with those in mice in the WHK group (Fig. 3C), consistent with changes in the expression of inflammatory proteins in KO and WT mice. Subsequently, RT-qPCR analysis revealed that the mRNA expression levels of

Table II. Biochemical indicators of ketamine in blood and urine of mice in the different experimental groups.

A, 4 weeks						
Indicator	C (NS)		LK (30 mg/kg)		HK (60 mg/kg)	
	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)
Serum Ket (ng/ml)	ND	ND	ND	ND	ND	ND
Serum Nket (ng/ml)	ND	ND	ND	ND	ND	ND
Urine Ket (ng/ml)	ND	ND	868±95.9	896±94.3	1068±98.6	1,123±95.3
Urine Nket (ng/ml)	ND	ND	2,2681±734.2	2,0248±870.4	23,425±726.9	24,786±102.3
B, 8 weeks						
Indicator	C (NS)		LK (30 mg/kg)		HK (60 mg/kg)	
	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)
Serum Nket (ng/ml)	ND	ND	ND	ND	ND	ND
Urine Ket (ng/ml)	ND	ND	1,076±103.2	998±87.3	1,324±112.3	1,238±109.3
Urine Nket (ng/ml)	ND	ND	24,564±924.3	23,652±898.3	27,347±923.4	25,863±954.2
C, 12 weeks						
Indicator	C (NS)		LK (30 mg/kg)		HK (60 mg/kg)	
	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)	Aldh2 ⁻ (n=15)	Aldh2 ⁺ (n=15)
Serum Nket (ng/ml)	ND	ND	1.2±0.32	2.3±0.26	4.6±0.67	3.2±0.34
Urine Ket (ng/ml)	ND	ND	1,545±162.7	1,432±93.3	1,636±156.8	1,523±196.2
Urine Nket (ng/ml)	ND	ND	23,691±1,423.3	26,538±1,397.2	28,877±1,452.3	29,376±1,254.8

Data are presented as the mean ± standard error of the mean. Ket, ketamine; nket, norketamine; KO, knock-out; WT, wild-type; -, KO; +, WT; C, control; NS, normal saline; LK, low-dose ketamine; HK, high-dose ketamine; Aldh2, aldehyde dehydrogenase 2; Ket, ketamine; Nket, norketamine; ND, not detected.

α -SMA, TGF- β and fibronectin in KHK vs. WHK mice were 2.814±0.342 vs. 1.967±0.536, 2.237±0.118 vs. 1.524±0.102 and 2.791±0.395 vs. 1.866±0.551, respectively (all P<0.01) in week 12.

Discussion

To the best of our knowledge, the present study is the first to investigate the impact of the *aldh2* gene on ketamine-induced cystitis. Aldh2 is a metabolic enzyme of certain aldehydes, including 4-hydroxynonenal and MDA, which has a number of enzymatic functions, including dehydrogenase and esterase. In addition, *aldh2* oxidizes 4-hydroxynonenal, MDA and other oxidation products and converts them into non-toxic acids, thereby reducing the degree of inflammation and inhibiting apoptosis. Using this function, *aldh2* has been previously reported to protect the kidneys, liver, heart and other organs from damage (21). Hu *et al* (22) demonstrated that inhibiting *aldh2* expression aggravated the inflammatory reaction in rats with sepsis, increasing kidney damage. In another study, Wimborne *et al* (23) previously revealed that activation of

aldh2 reduced the hepatotoxic effects of ethanol and acetaminophen. Additionally, Xu *et al* (24) reported that *aldh2* prevented myocardial damage associated with pulmonary hypertension. These previous studies demonstrated that *aldh2* served important functions in regulating inflammation leading up to organ damage. The low expression of *aldh2* was found to associate closely with renal fibrosis, myocardial fibrosis and urinary tract fibrosis caused by urothelial tumors (25-27). Tang *et al* (25) revealed that *aldh2* can be used as a common potential genetic target for prognosis of various renal fibrosis diseases (renal fibrosis caused by unilateral ureteral obstruction, ischemia-reperfusion injury or cisplatin-induced). Mali *et al* (26) previously discovered that heart damage and myocardial fibrosis as a result of chronic hyperglycemia are associated with reduced *aldh2* activity. A whole genome analysis conducted by Wu *et al* (27) demonstrated *aldh2* is a potential prognostic marker for urothelial carcinoma, where *aldh2* expression associated with that of urothelial fibrosis genes. However, the mechanism of *aldh2* in the development of KIC remains unclear. Due to its significant anti-inflammatory and antifibrotic effects, the present study

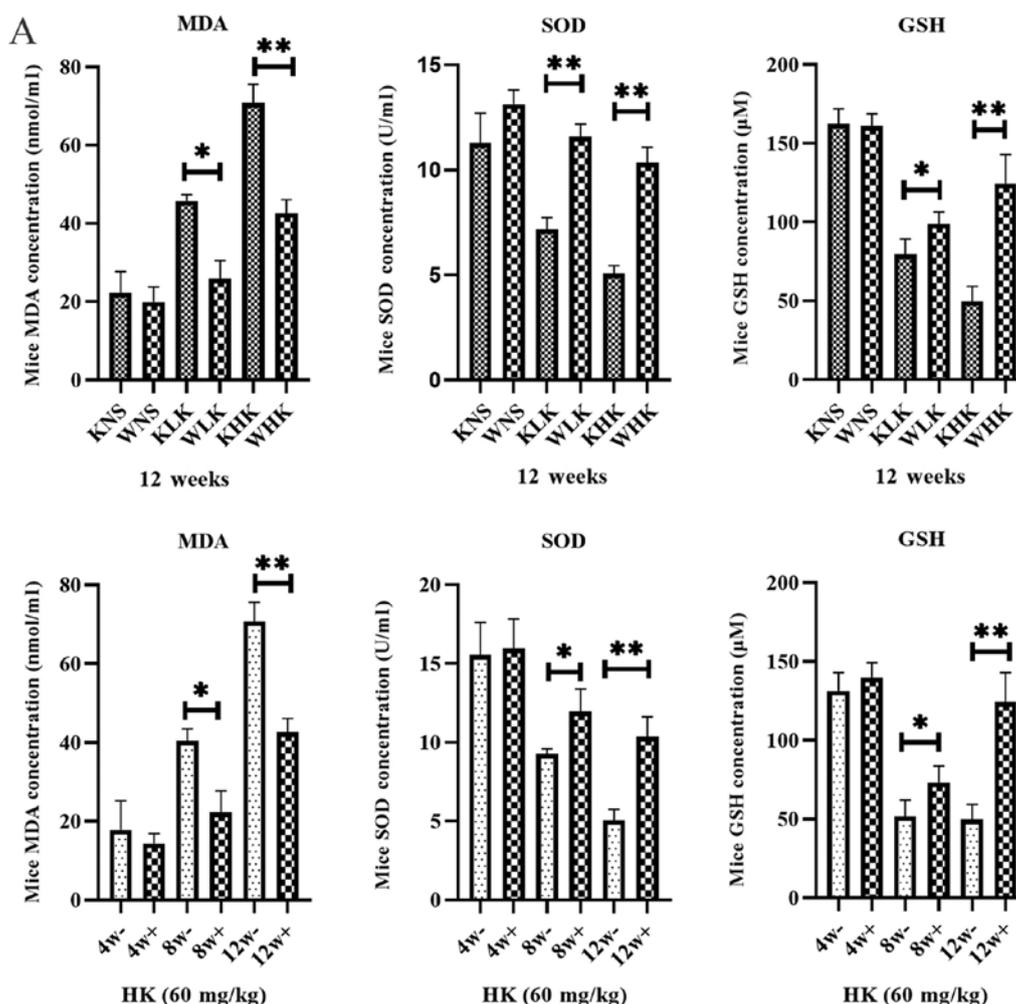


Figure 2. Ketamine increases the level of oxidative stress in *aldh2* KO mice and aggravates pathological damage. (A) Effects of ketamine on parameters of oxidative stress in WT and KO mice at 4, 8 and 12 weeks as detected by ELISA. Negative control mice were treated with NS. * $P < 0.05$ and ** $P < 0.01$. KO, knock-out; NS, normal saline; KHK, knock-out high-dose ketamine group; SOD, superoxide dismutase; GSH, glutathione-sulfhydryl; MDA, malondialdehyde; WHK, wild-type high-dose ketamine group; COX-2, cyclooxygenase 2; iNOS, inducible nitric oxide synthase; WT, wild-type; KNS, knock-out normal saline control group; WNS, wild-type normal saline control group; KLK, knock-out low-dose ketamine group; WLK, wild-type low-dose ketamine group; HK, high-dose ketamine; W, week; -, knock-out; +, wild-type.

hypothesized that *aldh2* has a high probability of being a novel KIC prevention target.

The results of the present study demonstrated that following long-term treatment of ketamine, the expression of *aldh2* in mice was significantly upregulated. Previous studies have reported that the pathogenesis of KIC is associated with inflammation and fibrosis (28,29). Therefore, the present study hypothesized that following long-term treatment of ketamine, the *aldh2* gene may be upregulated to protect against adverse factors, including oxidative stress, inflammation and fibrosis *in vivo*. To investigate this, the long-term effects of ketamine on *aldh2* KO mice were compared with those in WT mice.

KIC is a dynamic and complex process that is closely associated with oxidative stress, which is in turn associated with excessive ROS production (4). The effects of ROS on micturition reflexes have been previously demonstrated in several pathological states of the bladder, including cyclophosphamide-induced hemorrhagic cystitis, ionizing radiation cystitis and partial bladder outlet obstruction (30). High intracellular levels of ROS and the imbalance between the oxidative and antioxidant systems promotes lipid peroxidation and increases

the formation of aldehydes, including that of MDA, which is the end product in kidney tissues. Additionally, GSH and SOD inactivate ROS production in the mitochondria by enhancing the activity of catalase in peroxisomes. Therefore, any aberrant changes in GSH and SOD can cause serious damage to cellular DNA, lipids and proteins (5). The present study demonstrated that under the same doses of ketamine, the levels of oxidative stress in *aldh2* KO mice were significantly higher compared with that in WT mice. In addition, ketamine treatment significantly reduced the expression of antioxidant enzymes whilst enhancing the production of MDA in *aldh2* KO mice.

During KIC, ROS stimulates NF- κ B activation, leading to downstream signaling pathways to induce tissue damage (31,32). Generally, NF- κ B is inactive under physiological conditions. Once activated by phosphorylation, NF- κ B dissociates from its inhibitory unit, nuclear factor of κ light polypeptide gene enhancer in B-cells inhibitor β , freeing the active p65-NF- κ B. p65-NF- κ B then migrates to the nucleus, where it binds to its promoter sequence to activate the expression of pro-inflammatory cytokines (33). In a previous study, hyaluronan instillation treatment significantly inhibited the activation of the NF- κ B

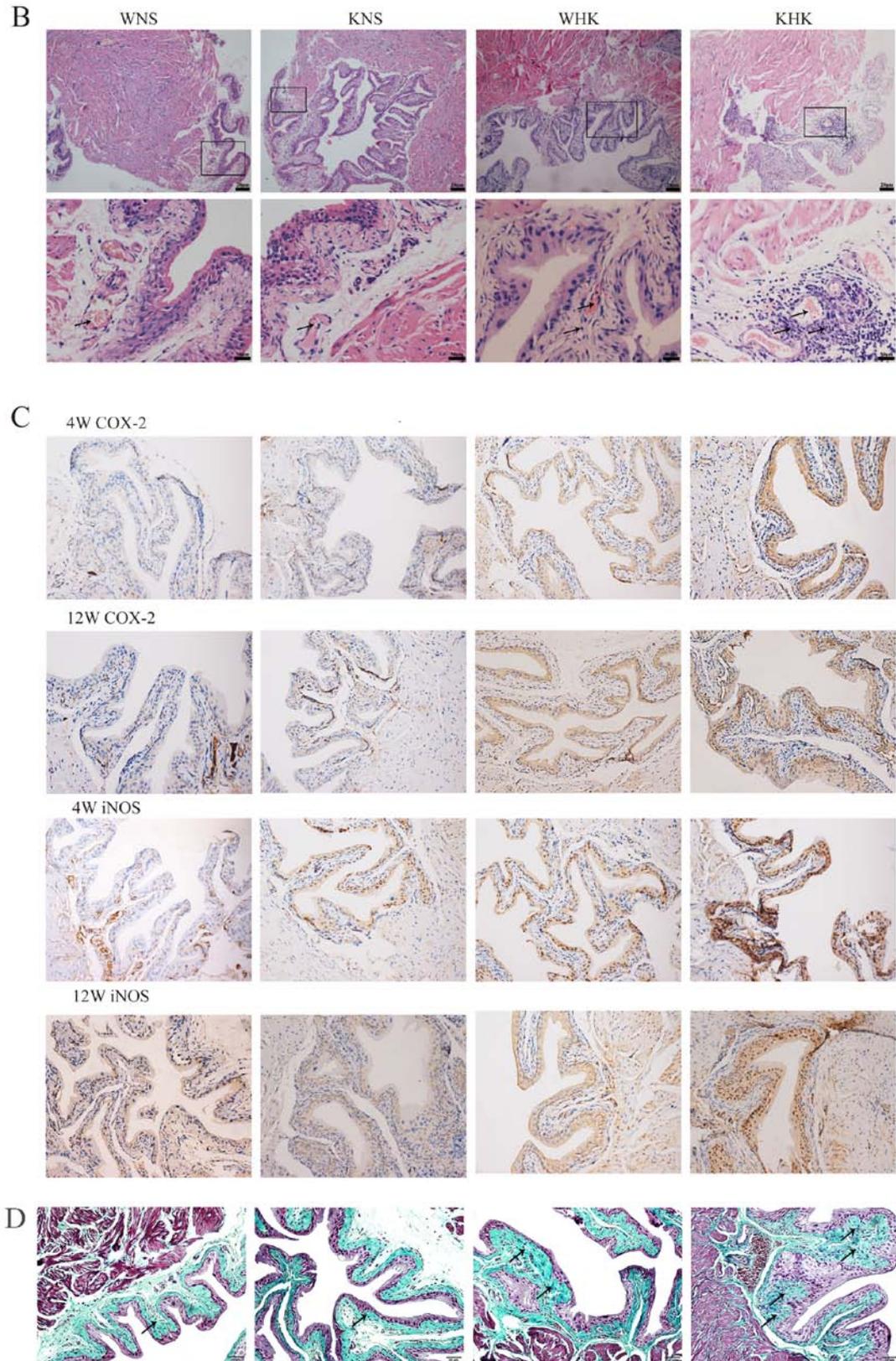


Figure 2. Continued. Ketamine increases the level of oxidative stress in *aldh2* KO mice and aggravates pathological damage. (B) Representative hematoxylin and eosin staining images of bladder tissues from KO and WT mice in week 12. Magnification, x100 for the upper images; x400 for the lower images. (C) Representative immunohistochemical staining images of COX-2 and iNOS proteins in the bladder tissues of KO and WT mice in weeks 4 and 12. The cytoplasm and cell membranes exhibiting brown-yellow colors were considered as positive expression of the target protein, which were mainly confined to the bladder epithelium. Magnification, x200. (D) Representative Masson trichrome staining images of bladder tissues of KO and WT mice in week 12. Collagen fibers stained green, muscle fibers stained red, and nucleus stained blue-brown. Magnification, x200. Data are presented as the mean \pm standard error of the mean from ≥ 3 experimental repeats. * $P < 0.05$ and ** $P < 0.01$. KO, knock-out; NS, normal saline; KHK, knock-out high-dose ketamine group; SOD, superoxide dismutase; GSH, glutathione-sulphydryl; MDA, malondialdehyde; WHK, wild-type high-dose ketamine group; COX-2, cyclooxygenase 2; iNOS, inducible nitric oxide synthase; WT, wild-type; KNS, knock-out normal saline control group; WNS, wild-type normal saline control group; KLK, knock-out low-dose ketamine group; WLK, wild-type low-dose ketamine group; HK, high-dose ketamine; W, week; -, knock-out; +, wild-type.

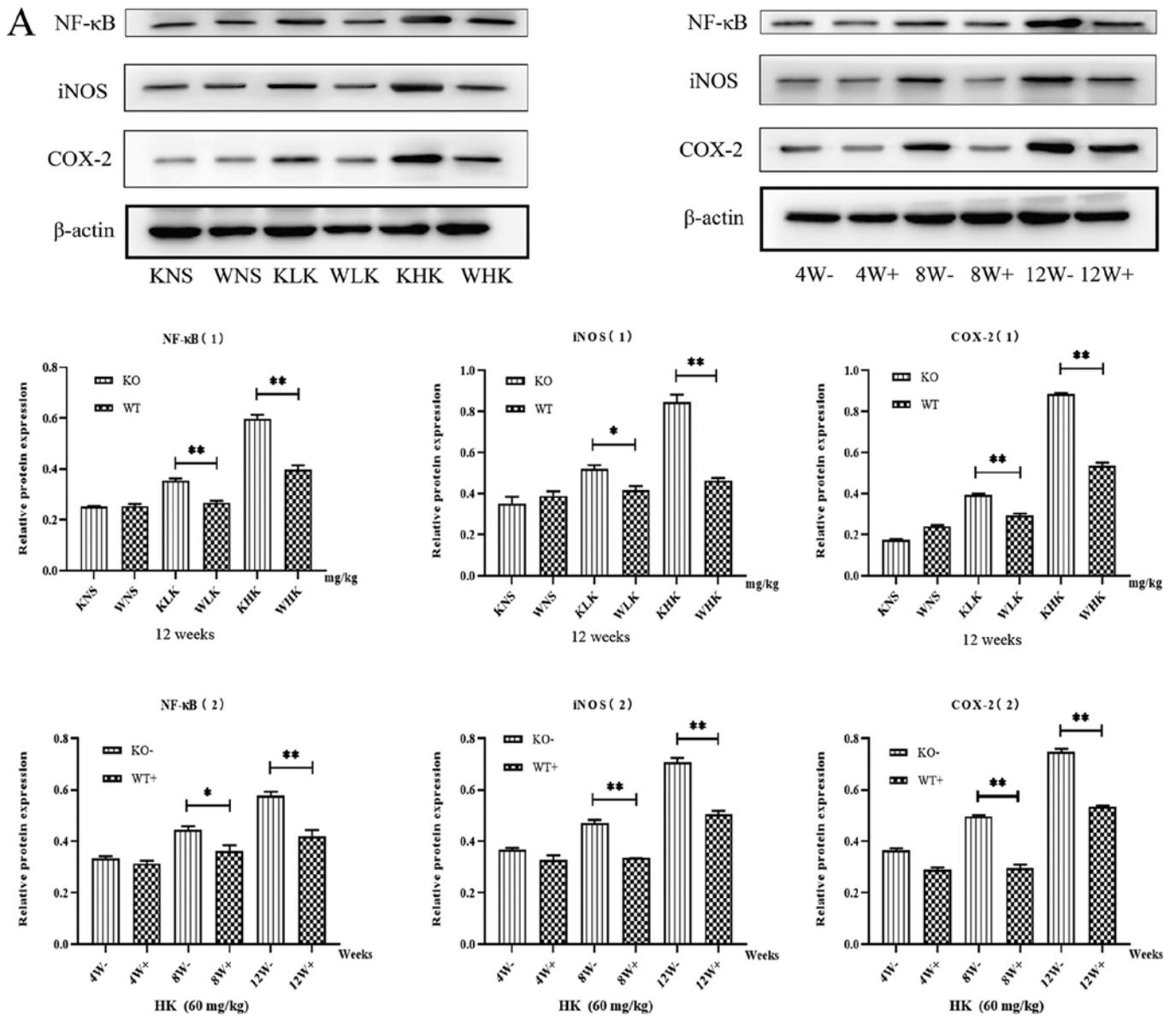


Figure 3. Ketamine treatment induces inflammation and fibrosis in KO mice. (A) Effects of ketamine on inflammation protein COX-2, iNOS and NF-κB levels in mice. *P<0.05 and **P<0.01. Data are presented as the mean ± standard error of the mean from ≥3 experimental repeats. KO, knock-out; KHK, KO high-dose ketamine group; WHK, wild-type high-dose ketamine group; COX-2, cyclooxygenase 2; iNOS, inducible nitric oxide synthase; WT, wild-type; α-SMA, α-smooth muscle actin; TGF-β, transforming growth factor β; KNS, knock-out normal saline control group; WNS, wild-type normal saline control group; KLK, knock-out low-dose ketamine group; WLK, wild-type low-dose ketamine group; W, weeks; -, knock-out; +, wild-type; HK, high-dose ketamine.

signaling pathway by suppressing oxidative stress (34). The results of the present study revealed that the expression of the active NF-κB unit in *aldh2* KO mice was higher compared with that in WT, indicating a more stressful condition. Treating *aldh2* KO mice with ketamine activated the NF-κB pathway, which may be associated with the absence of *aldh2* in its anti-oxidative stress effects. COX-2 and iNOS are sensitive markers of inflammation caused by oxidative stress (35). Previous studies have demonstrated that COX-2 regulated the inflammatory response in rats with KIC via the NF-κB pathway (15,31), such that COX-2 overexpression can serve an important role in altering the urinary pattern during KIC (36). Numerous previous studies on cyclophosphamide-induced cystitis (37,38) revealed the role of COX-2 in bladder overactivity, which is associated with inflammation or hypertrophy. Additionally, treatment

with COX-2 inhibitors has been demonstrated to improve ifosfamide-induced bladder injury and alter urinary overactivity (39). iNOS is usually expressed in macrophages. When activated by pathogens or cytokines, such as ROS, it synthesizes nitric oxide (NO) to promote cell apoptosis (40). A previous study reported that iNOS was upregulated in cyclophosphamide-induced cystitis (38). Other studies have previously revealed that ketamine or its urinary metabolites exerted direct toxic effects on bladder epithelial cells due to the activation of iNOS in the mitochondria and the resulting high levels of NO in urine (19,36). In the present study, COX-2 and iNOS were found to be highly expressed in *aldh2* KO mice.

Numerous animal studies have demonstrated that long-term ketamine treatment can lead to bladder fibrosis, which is an important cause of bladder abnormalities, including decreased

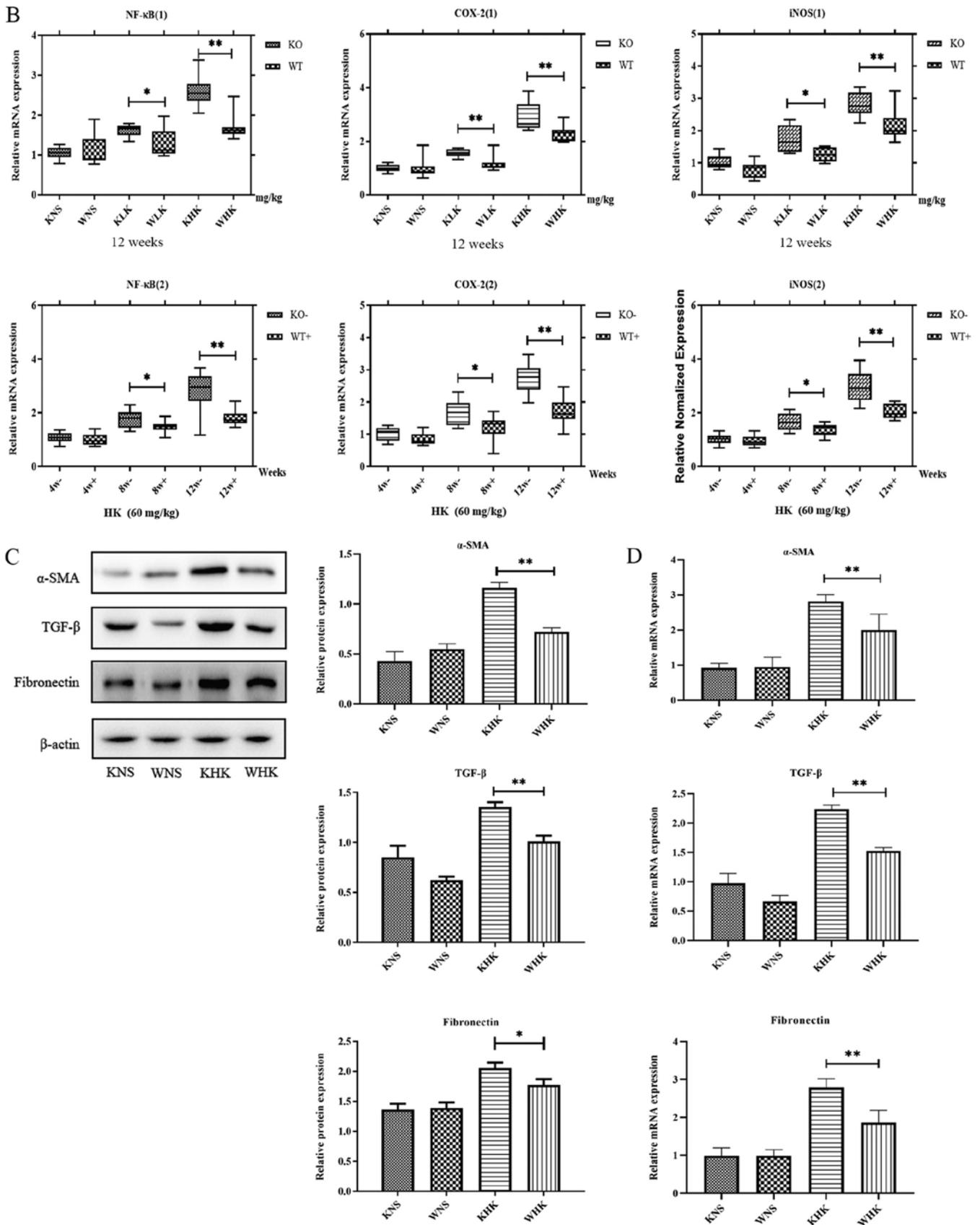


Figure 3. Continued. Ketamine treatment induces inflammation and fibrosis in KO mice. (B) mRNA expression levels of inflammatory proteins COX-2, iNOS and NF-κB in the mouse bladder tissues were detected by reverse transcription-quantitative PCR. (C) Western blotting and (D) reverse transcription-quantitative PCR were performed to measure the expression of α-SMA, TGF-β and fibronectin in mouse bladder tissues at week 12. *P<0.05 and **P<0.01. Data are presented as the mean ± standard error of the mean from ≥3 experimental repeats. KO, knock-out; KHK, KO high-dose ketamine group; WHK, wild-type high-dose ketamine group; COX-2, cyclooxygenase 2; iNOS, inducible nitric oxide synthase; WT, wild-type; α-SMA, α-smooth muscle actin; TGF-β, transforming growth factor β; KNS, knock-out normal saline control group; WNS, wild-type normal saline control group; KLK, knock-out low-dose ketamine group; WLK, wild-type low-dose ketamine group; W, weeks; -, knock-out; +, wild-type; HK, high-dose ketamine.

capacity, lower compliance and impaired detrusor function (41,42). Song *et al* (41) demonstrated that the inflammatory mediators in KIC increased the expression of collagen type-I (COL-I) and α -SMA, which leads to thickening of the bladder basement membrane. Furthermore, Shen *et al* (42) conducted whole genome analysis and reported that the expression of fibrotic genes, including fibronectin, TGF- β 1 and COL-I, were upregulated in bladder tissues with KIC. This enriched expression was considered to be a sensitive marker of active fibrosis development. The present study revealed that compared with WT mice, the severity of ketamine-induced fibrosis in *aldh2* KO mice was considerably higher and the expression of α -SMA, TGF- β 1 and fibronectin were significantly higher. These results indicated that the expression of *aldh2* alleviates bladder fibrosis induced by KIC. Due to the effects of inflammation and fibrosis *in vivo*, *aldh2* KO mice gained weight slower and urinated more frequently compared with that in WT mice. This may be due to the chronic physiological stress caused by inflammation and fibrosis, which were increased in *aldh2* KO mice.

The results of the present study revealed that *aldh2* inhibited the NF- κ B signaling pathway, thereby preventing fibrosis by suppressing lipid peroxidation in KIC and reducing COX-2 and iNOS expression. Data from the present study advises that Asian people should refrain from ingesting ketamine, since they are at a higher risk of developing mutations in the *aldh2* gene, which can result in the development of severe LUTS and bladder contracture. Future studies should include cellular tests and crowd verification experiments to validate the results of the present study. Future studies are also required to clarify whether ketamine-induced cystitis is race-specific.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Author's contributions

XJX and HM designed this study. XJX and SHC performed the experiments. XJX, SHC and HM performed the statistical analysis. XJX and HM drafted and revised the manuscript. HM contributed reagents, materials and experimental platforms. All authors read and approved the final manuscript.

Ethics approval and consent to participate

The present study was approved by the Ethics Committee of Guangxi Medical University (approval no. 20180129).

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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