

## 

**Citation:** Xia H, Huang X, Deng S, Xu H, Yang Y, Liu X, et al. (2021) *DNAH11* compound heterozygous variants cause heterotaxy and congenital heart disease. PLoS ONE 16(6): e0252786. https://doi.org/10.1371/journal. pone.0252786

Editor: Tao Cai, NIDCR/NIH, UNITED STATES

Received: November 20, 2020

Accepted: May 21, 2021

Published: June 16, 2021

**Copyright:** © 2021 Xia et al. This is an open access article distributed under the terms of the <u>Creative</u> Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: The data involved in the study cannot be shared publicly because of ethical and legal concerns, regarding sensitive information. Reasonable requests to access the datasets by the qualified researchers can be sent to the Institutional Review Board of the Third Xiangya Hospital, Central South University (xy3irb@163. com).

**Funding:** The study was supported by grants from National Natural Science Foundation of China (81670216, 81873686 and 81800219), Natural Science Foundation of Hunan Province, China **RESEARCH ARTICLE** 

# *DNAH11* compound heterozygous variants cause heterotaxy and congenital heart disease

# Hong Xia<sup>1,2</sup>, Xiangjun Huang<sup>3</sup>, Sheng Deng<sup>4</sup>, Hongbo Xu<sup>1</sup>, Yan Yang<sup>5</sup>, Xin Liu<sup>1</sup>, Lamei Yuan<sup>1</sup>\*, Hao Deng<sup>0</sup><sup>1</sup>\*

 Center for Experimental Medicine, the Third Xiangya Hospital, Central South University, Changsha, Hunan, China, 2 Department of Emergency, the Third Xiangya Hospital, Central South University, Changsha, Hunan, China, 3 Department of General Surgery, The First Affiliated Hospital of Hunan University of Chinese Medicine, Changsha, Hunan, China, 4 Department of Pharmacy, Xiangya Hospital, Central South University, Changsha, Hunan, China, 5 Department of Neurology, the Third Xiangya Hospital, Central South University, Changsha, Hunan, China

\* hdeng008@yahoo.com (HD); yuanlamei229@163.com (LY)

### Abstract

Heterotaxy (HTX), a condition characterized by internal organs not being arranged as expected relative to each other and to the left-right axis, is often accompanied with congenital heart disease (CHD). The purpose was to detect the pathogenic variants in a Chinese family with HTX and CHD. A non-consanguineous Han Chinese family with HTX and CHD, and 200 unrelated healthy subjects were enlisted. Exome sequencing and Sanger sequencing were applied to identify the genetic basis of the HTX family. Compound heterozygous variants, c.3426-1G>A and c.4306C>T (p.(Arg1436Trp)), in the dynein axonemal heavy chain 11 gene (DNAH11) were identified in the proband via exome sequencing and further confirmed by Sanger sequencing. Neither c.3426-1G>A nor c.4306C>T variant in the DNAH11 gene was detected in 200 healthy controls. The DNAH11 c.3426-1G>A variant was predicted as altering the acceptor splice site and most likely affecting splicing. The DNAH11 c.4306C>T variant was predicted to be damaging, which may reduce the phenotype severity. The compound heterozygous variants, c.3426-1G>A and c.4306C>T, in the DNAH11 gene might be the pathogenic alterations resulting in HTX and CHD in this family. These findings broaden the variant spectrum of the DNAH11 gene and increase knowledge used in genetic counseling for the HTX family.

#### Introduction

Human and other vertebrate visceral organs are asymmetric to the left-right (LR) axis. Normal organ placement relative to the LR axis is referred to as situs solitus (SS). Abnormal positioning is heterotaxy (HTX) or situs inversus (SI) [1]. A randomized placement of visceral organs relative to each other and LR axis is HTX. Visceral organ placement in a perfect mirror-image position along the LR axis is SI [2].

HTX is an extremely clinical heterogeneous disorder. HTX prevalence is about 1 in 10,000 newborns [3]. HTX, SI, and SS may present in different families or accidentally in the same

(2019JJ50927, 2020JJ3057 and 2020JJ4830), Scientific Research Project of Health Commission of Hunan Province, China (B2019174), Special Emergency Project from Science and Technology Department of Hunan Province (2020SK3032) and the Hunan Provincial Innovation Foundation for Postgraduate, China (CX20190254). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing interests:** The authors have declared that no competing interests exist.

family [1]. HTX is often connected with congenital heart disease (CHD) and is classified as either isolated or syndromic [4, 5]. At least 59 syndromes have been reported to associate either HTX or SI [5].

HTX is a genetic heterogeneous disorder with incomplete penetrance. It may be inherited in an autosomal recessive, autosomal dominant or X-linked fashion. It associates with genetic factors, environmental modifiers, and developmental randomness [4, 5]. As of the date of this writing (November 2020), nine loci (*HTX1-9*) and eight causative genes, including the Zic family member 3 gene (*ZIC3*), the cripto, FRL-1, cryptic family 1 gene (*CFC1*), the activin A receptor type 2B gene (*ACVR2B*), the nodal growth differentiation factor gene (*NODAL*), the cilia and flagella associated protein 53 gene (*CFAP53*), the matrix metallopeptidase 21 gene (*MMP21*), the polycystin 1 like 1, transient receptor potential channel interacting gene (*PKD1L1*) and the meiosis specific nuclear structural 1 gene (*MNS1*) have been reported as associating with isolated HTX [6–14].

The great clinical and genetic heterogeneity presents challenges to correctly identifying causative variants in individuals with HTX phenotype using Sanger sequencing. Exome sequencing, a cost-saving measure, has been used in the genetic investigation of HTX [12–14]. Our previous study has successfully identified pathogenic variants in two Chinese families with SI phenotype via exome sequencing [15, 16]. In this study, exome sequencing and Sanger sequencing detected the compound heterozygous variants, c.3426-1G>A and c.4306C>T (p. (Arg1436Trp)), in the dynein axonemal heavy chain 11 gene (*DNAH11*, NM\_001277115.1) in a Han Chinese family with HTX and CHD.

#### Materials and methods

#### Subjects

A two-generation, non-consanguineous Han family, from Yiyang, Hunan, China, with HTX and CHD was enrolled from May to July 2019. Four family members including the proband (II:2), his unaffected mother (I:2), and two unaffected siblings (II:1 and II:3) participated (Fig 1A). The proband (II:2) was diagnosed, at age five, as having cyanotic complex CHD due to dyspnea, cyanosis, poor activity endurance and cardiac murmurs. His first operation occurred at sixteen after he was admitted to hospital due to severe shortness of breath. He presented with cyanotic lips. The apical impulse was detected at the right fifth intercostal space. A systolic ejection murmur was heard in the right second intercostal space. Chest X-ray revealed dextrocardia (Fig 1B). Echocardiography showed complex CHD, including single ventricle, incomplete transposition of the great arteries, and pulmonary valve stenosis. No abdominal organ abnormalities were detected in the proband. Dyspnea, cyanosis, dextrocardia, cardiac murmurs, or cardiovascular malformations were not detected in his parents or other siblings. Two hundred unrelated subjects (100 males and 100 females, aged 16.5±5.5 years) who did not have dyspnea, cyanosis, dextrocardia, or cardiovascular anomalies were recruited as healthy controls from May to October 2019. The study was approved by the Institutional Review Board of the Third Xiangya Hospital, Central South University (Changsha, China) in accordance with the Declaration of Helsinki. The study was conducted from May to December 2019. Written informed consent forms were obtained, and venous blood samples were taken from all participants. The authors had no access to information that could identify individual participants during or after data collection.

#### Exome sequencing

Genomic DNA was obtained from blood samples via a saturated phenol-chloroform extraction method [17]. A microgram of genomic DNA from the proband (II:2) was exome sequenced at



**Fig 1. Pedigree of a Chinese heterotaxy family, chest X-ray of the heterotaxy patient and the** *DNAH11* **Sanger sequencing electropherograms.** (A) Pedigree of a Chinese heterotaxy family. N, normal; V1, the *DNAH11* c.3426-1G>A variant; V2, the *DNAH11* c.4306C>T variant. The slash indicates deceased individual, the fully shaded symbol indicates the affected individual, and the arrow indicates the proband. (B) Chest X-ray of the family member (II:2) revealed dextrocardia. (C, D) The compound heterozygosity for the *DNAH11* variants, c.3426-1G>A and c.4306C>T, in the individual (II:2) with heterotaxy.

https://doi.org/10.1371/journal.pone.0252786.g001

BGI-Shenzhen, China. The library with targeted exome was captured by the Agilent SureSelect Human All Exon V6 and sequenced using a BGISEQ-500 platform. Average sequencing depth was 267.63×. Debased reads were filtered and the clean reads were mapped to the human reference genome (UCSC database version hg19, http://genome.ucsc.edu/) using a Burrows-Wheeler Aligner tool (BWA, version 0.7.15). Duplicate reads were marked by Picard-tools (version 2.5.0). Single nucleotide polymorphisms (SNPs) and insertions/deletions (Indels) were detected by a HaplotypeCaller of Genome Analysis Toolkit (GATK, version 3.3.0). Variant annotation and prediction were performed by SnpEff tool (https://pcingola.github.io/ SnpEff/). Variants with minor allele frequency  $\geq$ 1% in the following databases were eliminated: the SNP database version 141 (dbSNP141), the 1000 Genomes Project (1000 genomes release phase 3), and the NHLBI Exome Sequencing Project (ESP) 6500 [18]. The functional impacts of nonsynonymous SNPs or Indels were predicted by the Sorting Intolerant from Tolerant (SIFT) and the Protein Variation Effect Analyzer (PROVEAN, version 1.1.3, http:// provean.jcvi.org/index.php). The effects of splice site variants on splicing were predicted by the Human Splicing Finder (HSF, version 3.1, http://www.umd.be/HSF3/).

#### Sanger sequencing, variant analysis and protein structure modeling

Candidate variants were validated by Sanger sequencing. A co-segregation analysis between possible causative variants and this family's HTX phenotype was performed [19]. Paired primer sequences for the candidate variants (c.3426-1G>A and c.4306C>T) in the *DNAH11* gene were synthesized respectively as follows: 5'-TGTTGCCAGTTTCATGATAGAGA-3' and 5'-TACAGCCAGAAGATGCACCA-3'; 5'-TTCACCAGCCTTTAGGCAAA-3' and 5'-TCTCAG TCCCCAGCTCTTTC-3'. The recorded frequencies of the candidate variants in the Genome Aggregation Database (gnomAD, version 3.0, http://gnomad.broadinstitute.org) were further checked, and pathogenic variant databases, including the Leiden Open Variation Database (LOVD, v.3.0, https://www.lovd.nl/3.0/home), the Human Gene Mutation Database (HGMD, http://www.hgmd.cf.ac.uk/ac/index.php), and ClinVar (https://www.ncbi.nlm.nih.gov/clinvar/) were referred to interpreting variants. The American College of Medical Genetics and Genomics (ACMG) sequence variant interpretation guidelines, and recommendations for interpreting the loss of function pathogenic criterion (PVS1) created by ClinGen Sequence Variant Interpretation Working Group, were used to categorize the identified variants [20, 21].

The protein structures of wild-type and variant-type were predicted via the online SWISS-MODEL tool (http://www.swissmodel.expasy.org), and the visualized structures were constructed using the PyMOL software (version 1.7, Schrödinger, LLC, Portland, U.S.A.).

#### Results

Exome sequencing generated 240,827,260 clean reads. Approximately 99.94% of the clean reads were mapped to the human reference genome. A total of 105,788 SNPs and 18,688 Indels were detected. After a series of optimization screening measures, two variants, c.3426-1G>A and c.4306C>T in the *DNAH11* gene, were discovered. Other possible disease-causing alterations in the pathogenic genes known for HTX, SI or CHD were eliminated.

The compound heterozygous variants, a splicing variant, c.3426-1G>A, and a missense variant, c.4306C>T (p.(Arg1436Trp)), in the *DNAH11* gene, were further confirmed in the HTX patient (II:2) via Sanger sequencing (Fig 1C and 1D). The heterozygous c.3426-1G>A variant in the *DNAH11* gene was detected in his mother (I:2). Neither c.3426-1G>A nor c.4306C>T variant in the *DNAH11* gene was discovered in his elder sister (II:1). The compound heterozygous variants, c.3426-1G>A and c.4306C>T, in the *DNAH11* gene were also identified in his brother (II:3) who had no HTX or SI phenotype. This was in accord with prior reports that SI and HTX may occur in about half of PCD patients, approximately 59.1% of individuals or 42.9% of affected siblings having the *DNAH11* biallelic variants [22, 23]. His father's genotyping was indefinable as he (I:1) passed away. Due to the presence of the *DNAH11* c.4306C>T variant in two brothers (II:2 and II:3) and the absence in the mother (I:2), the variant might be not *de novo*, but inherited from his father (I:1), further supporting that the disorder was inherited in an autosomal recessive manner in this family. Neither *DNAH11* c.3426-1G>A nor c.4306C>T variant was detected in the 200 healthy controls.

The HSF tool predicted that the *DNAH11* c.3426-1G>A variant alters acceptor splice site and most probably affects splicing. The *DNAH11* c.4306C>T variant was predicted to result in substituting tryptophan for arginine at codon 1436 (p.(Arg1436Trp)) and it would be damaging with a prediction score of 0.001 by SIFT, and be deleterious with a score of -3.10 by PRO-VEAN. The allele frequencies in the gnomAD were  $1.31 \times 10^{-5}$  for c.3426-1G>A and  $3.09 \times 10^{-4}$ for c.4306C>T. The two variants were not recorded in LOVD, but deposited in HGMD, responsible for ciliary dyskinesia and bronchiectasis, respectively. The *DNAH11* c.3426-1G>A variant was interpreted as pathogenic in the ClinVar database, while the c.4306C>T variant



**Fig 2. Structural modeling displayed conformational changes of the DNAH11 p.(Arg1436Trp) variant.** (A) The cartoon representation of the DNAH11 protein. (B) The stick model of the arginine at residue 1436 (p.Arg1436). (C) The stick model of the mutant tryptophan at residue 1436 (p. Trp1436).

https://doi.org/10.1371/journal.pone.0252786.g002

had conflicting interpretations of pathogenicity (uncertain significance or benign). According to guidelines for the sequence variant interpretation, the variants, c.3426-1G>A and c.4306C>T, were classified as "pathogenic" (PVS1+PM2+PP5) and "uncertain significance" (PM2), respectively. Conformational changes after the arginine at residue 1436 (p.Arg1436) altered to tryptophan (p.Trp1436) were displayed by structural modeling (Fig 2).

#### Discussion

HTX occurs in approximately 6.3%-12.1% of patients having primary ciliary dyskinesia (PCD) [24, 25]. In 2002, a homozygous c.8533C>T (p.(Arg2845\*)) variant and a heterozygous c.8990G>A (p.(Arg2997Gln)) variant in the *DNAH11* gene were firstly detected in two

patients with SI and most likely PCD, respectively [26]. Subsequently compound heterozygous variants, c.12363C>G (p.(Tyr4121\*)) and c.13531\_\*36del (p.(Ala4511\_Ala4516delinsGln)), in the *DNAH11* gene were identified in a family with PCD, including a patient with SI [27]. Biallelic variants in the *DNAH11* gene may be responsible for about 22% of PCD patients with normal ciliary ultrastructure. It was reported that SI occurred in 54.5% of PCD patients, HTX in about 4.5% of PCD patients, and SS in about 41% of PCD patients with *DNAH11* biallelic variants [22].

The *DNAH11* gene locates in 7p15.3, contains 82 exons, and encodes a ciliary outer dynein arm with 4516 amino acids. The protein contains an N-terminal tail domain, six ATPase associated diverse cellular activities (AAA) domains, four P-loops, a helix-1-microtubule-binding-helix-2 domain, and a C-domain [26].

The embryonic expression of *Dnah11*, a mouse homolog of human *DNAH11*, also called "left-right dynein" (*lrd*), is detected primarily in non-ciliated cells and highly restricted in the node at mouse embryonic day 7.5. It also expresses in many ciliated cells in newborn and adult mice [28]. Dnah11-containing monocilia in the node generate a leftward nodal flow which may determine the normal left-right patterns in mouse embryos [29, 30].

The *iv/iv* mice, resulting from the homozygous missense p.Glu2271Lys variant in the *Dnah11* gene, presented with an abnormal nodal flow, randomized expression of lateralized genes, a high incidence of cardiovascular anomaly, and randomized left-right development. Half of them exhibited either HTX or SI, and half exhibited SS [28, 30-34]. Phenotypes indicative of human PCD, including rhinitis, sinusitis, otitis media, static tracheal cilia, and sperm motility defects, were also discovered in the *iv/iv* mice [34].

Compound heterozygous *DNAH11* variants, responsible for about 14.3% of patients with CHD and HTX, may be a frequent genetic basis of familial CHD and HTX syndrome [35]. The reported compound heterozygous variants in the *DNAH11* gene and their various phenotypes were summarized in Table 1 [22, 23, 27, 34–43].

In this study, the compound heterozygous variants, c.3426-1G>A and c.4306C>T (p. (Arg1436Trp)), in the *DNAH11* gene were identified in the HTX patient and his unaffected brother. Different clinical phenotypes (HTX, SI or SS) of the siblings with the *DNAH11* gene biallelic variants have been reported in some families [22, 40]. HTX and SI occurring in about 59% of individuals with the *DNAH11* biallelic variants may be connected with clinical heterogeneity of LR asymmetry disorders with incomplete penetration, or additional genetic variants in the family member II:3 serving a suppressor in the *DNAH11* pathway. It might also be associated with environmental modifiers and randomized left-right development [1, 22].

The *DNAH11* c.3426-1G>A variant in the homozygous state was previously identified in a fetus with structural abnormalities, including single atrium and ventricle, pulmonary stenosis, and right isomerism [44]. The compound heterozygous *DNAH11* missense variants, c.4306C>T (p.(Arg1436Trp)) and c.6118C>T (p.(Arg2040Cys)), were reported in a patient with bronchiectasis, a clinical manifestation of PCD [39]. Neither variants were detected in the enrolled 200 healthy controls. Allele frequency of either variant in the total population was very low. The *DNAH11* c.3426-1G>A variant was predicted to alter the acceptor splice site, and the c.4306C>T variant was predicted to be damaging. Together with the pathogenicity evidence, the c.3426-1G>A and c.4306C>T variants in the *DNAH11* gene, as a novel combination, could be the pathogenic variants for HTX and CHD in this family, and the missense variant may reduce the phenotype severity.

*DNAH11* biallelic variants were detected in patients with ciliary dysfunction, HTX, SI and CHD phenotypes [23, 40]. The patient refused to permit either the nasal curettage or the bronchial mucosal biopsy to perform an analysis on the ciliary ultrastructure and the waveform. No obvious chronic airway infection symptoms or signs, such as recurrent rhinitis, sinusitis, otitis

Product Probability of the sector	Patients	Gender	Ethnicity	Sequence variants <sup>#</sup>	PCD phenotypes	Situs	CHD	Ciliary motion	References
Matrix Matrix Signal (C)	-	F	Chinese	c.73G>A, p.(Ala25Thr)	BR, rhinosinusitis	SI	-	KS	[36]
<table-container>      Net     Note     Section     Sect</table-container>				c.5702A>C, p.(Glu1901Ala)					
Image: section of the section of	OP41-II:1	М	Caucasian	c.350A>T, p.(Glu117Val)	NRD, sinusitis, OM	SI	-	KS (H)	[22]
5862 F C C C C C C C C C C C C C   4803 M C <				c.7148T>C, p.(Leu2383Pro)					
Image: biologic state in the state in th	#5062	F	Chinese	c.727A>G, p.(Ile243Val)	na	HTX	+	CD (I+R)	[35]
<table-container>      9501     Number Notation     Second Second</table-container>				c.10829A>T, p.(Asp3610Val)					
<table-container>  P44 N4 V C4060-A, P, TP302' Rinomania (M) HTX A RS(N) [7] [2] [2] [2] [2] [2] [2] [2] [2] [2] [2</table-container>	#5031	М	Chinese	c.846G>C, p.(Met282Ile)	na	HTX	+	CD (I+R +D)	[35]
A F Iula 683 Hold 683 Hold 683 Hold 683 Hold 613 Hold 683 Hold	P44	М		c.2406G>A, p.(Trp802*)	Rhinosinusitis	HTX	+	KS (R)	[37]
8 M (A1305) (ATPR177) B. Recurrent pneumonia, BR SI (a) (b)   1212 na Calso(>, p(Pro1321(a)) B na </td <td>A</td> <td>F</td> <td>Italian</td> <td>c.883-1G&gt;A</td> <td>A: BR, rhinitis, rhinosinusitis, otitis</td> <td>SI</td> <td>-</td> <td>KS (I+H)</td> <td>[38]</td>	A	F	Italian	c.883-1G>A	A: BR, rhinitis, rhinosinusitis, otitis	SI	-	KS (I+H)	[38]
B212 na Chines c.1307>C.p.(Phet34Leu) BR na na<	В	М		c.4130G>A, p.(Trp1377*)	B: Recurrent pneumonia, BR	SI			
$ \begin{array}{ c c c c } \hline \begin{tabular}{ c c } \hline \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	B212	na	Chinese	c.1300T>C, p.(Phe434Leu)	BR	na	na	na	[39]
\$5065     M     Chinese     c.1339G>A,p.(Gly447Arg) c.34707-G, p.(Leu]157Arg)     na     HTX     +     CD (I+R)     [55]       Patien1 M     Polish, C.1648del, p.(Arg550G)/57 h2)     na     HTX     +     na     [40]       Patien1 Z     na     C.1648del, p.(Arg550G)/57 h2)     na     HTX     +     na     [40]       PCD761     F     Caucasia     C2275-G>, p.(Arg3888G)%)     NRD, BR, sinusitis, OM     SI     -     KS (H)     [22]       Family 1-1     F     Finnish     c.23416-S, p.(Gla/31Ly)     Rhinosinusitis, OM     SI     -     KS (Static)     [41]       Pamily 1-2     M     C.7645+5G>A     Rhinosinusitis <om< td="">     SI     -     KS (Static)     [41]       Pamily 1-2     Ma     Chinese     c.2449G&gt;C,p.(Ap807His)     BR     na     na     na     na     [39]       C12256C&gt;A, p.(Ya4848Met)     E     Come     c.2446S&gt;C, p.(Ap807His)     BR     na     na     na     [39]       P28     Ma     Chinese     c.2446S&gt;A, p.(Ya4848Met)     BR<!--</td--><td></td><td></td><td></td><td>c.6983C&gt;T, p.(Pro2328Leu)</td><td></td><td></td><td></td></om<>				c.6983C>T, p.(Pro2328Leu)					
$ \begin{array}{ c c c } \begin{tabular}{ c c } \hline \begin{tabular}{ c c c } \hline \begin{tabular}{ c c c c } \hline \begin{tabular}{ c c c c c c } \hline \hline \begin{tabular}{ c c c c } \hline \hline \begin{tabular}{ c c c c } \hline \hline \begin{tabular}{ c c c c c } \hline \hline \begin{tabular}{ c c c c c c c c c c } \hline \begin{tabular}{ c c c c c c c c c } \hline \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	#5065	М	Chinese	c.1339G>A, p.(Gly447Arg)	na	НТХ	+	CD (I+R)	[35]
Patient 1 M Polish (2772G>A, p.(Merg2alle) (-1166CC>T, p.(Arg3888Cy)) na HTX				c. 3470T>G, p.(Leu1157Arg)					
Patient 2 Patient 2 and PCD7-61Patient 2 rel c.2772G-A, p.(Met92411c) c.11662C>T, p.(Arg4988Cys)NRD, BR, sinusitis, OMPatient 2 rel rel rel rel rel rel rel c.2275-1G>C, p.Tyr759_Glu888Cys)NRD, BR, sinusitis, OMSI rel rel rel rel rel rel rel rel rel rel 	Patient 1	М	Polish	c.1648del, p.(Arg550Glyfs*16)	na	HTX	+	na	[40]
$ \begin{array}{ c c c c c } \hline \begin{tabular}{ c c c c } \hline \begin{tabular}{ c c c c c c c } \hline \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	Patient 2	na		c.2772G>A, p.(Met924Ile)					
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $				c.11662C>T, p.(Arg3888Cvs)					
$ \begin{array}{ c c c c c c } \hline \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	PCD761	F	Caucasian	c.2275-1G>C, p.Tvr759 Glu889del	NRD, BR, sinusitis, OM	SI	-	KS (H)	[22]
Family 1-1FFinnishC2341G>A, p.(Gla781LyS)Rhinosinusitis, OMS1-KS (Static)PCD (D)Family 1-2Mc.7645+5G>ARhinosinusitisSSSS-KS (Static)PCD (D)B082naChinesec.2419G>C, p.(Asp807His)BRnanananana[39]B082naChinesec.2419G>C, p.(Asp807His)BRnanananana[39]B012naChinesec.2419G>C, p.(Asp807His)BRnanananana[39]C12258C>A, p.(Iva086')Fc.2482G>T, p.(Arg829Cys)Rhinosinusitis, OMSS-PCD (R)[37]B036naChinesec.2482G>A, p.(Va1848Met)BRnananana[39]B036naChinesec.2542G>A, p.(Va1848Met)BRnanana[39]B136naChinesec.2542G>A, p.(Va1848Met)BRnanana[39]c11174MItalianc.2753G>T, p.(Ghy4204Phe)Bronchitis, rhinitis, sinusitis, otitisSS-PCD (I)[23]11174MItalianc.2134(C-T, p.(Ser4024Phe)Bronchitis, rhinitis, sinusitis, otitisSS-PCD (I)[23]11174MItalianc.2342G>A, p.(Gla94SSerfs'10)Recurrent chest infection, rhinitisSS-PCD (I)[23]11174FEnglishc.23420G>C, p.(Leu1077)Rhinosinusitis, OM <td></td> <td>c.13213del, p.(Arg4405Alafs*2)</td> <td></td> <td></td> <td></td>				c.13213del, p.(Arg4405Alafs*2)					
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Family 1–1	F	Finnish	c.2341G > A, p.(Glu781Lvs)	Rhinosinusitis, OM	SI	-	KS (Static) PCD (D)	[41]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	Family 1–2	M		c.7645+5G>A	Rhinosinusitis	SS			
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	B082	na	Chinese	c.2419G>C, p.(Asp807His)	BR	na	na	na	[39]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $				c.2542G > A, p.(Val848Met)					
$\begin{array}{ c c c c c } \hline \begin{tabular}{ c c c c } \hline \begin{tabular}{ c c c c c c } \hline \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	B012	na	Chinese	c.2419G>C, p.(Asp807His)	BR	na	na	na	[39]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	0012			$c.12258C>A. p.(Tyr4086^*)$					
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	P28	М	Chinese	c.2485C>T. p.(Arg829Cvs)	Rhinosinusitis, OM	SS	-	PCD (R)	[37]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	120			c.5608C>T, p.(Pro1870Ser)					
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	B036	na	Chinese	c.2542G>A, p.(Val848Met)	BR	na	na	na	[39]
BI70naChinesec.2542G>A, p.(Val848Met) c.11624A>G, p.(Lys875Arg)BRnananananana[39]B156naChinesec.2542G>A, p.(Val848Met) c.12071C>T, p.(Ser4024Phe)BRnanananana[39]11174MItalian c.12776_12801delinsATA, p. Phe4266_Asn4267delinsIleBronchitis, rhinitis, sinusitis, otitisSS-PCD (I)[23]#4FEnglish c.13240dup, p.(Gln945Serfs*10) c.13240dup, p.(Thr4414Asnfs*34)Recurrent chest infection, rhinitis energianSS-PCD (I)[42]B142naChinese c.2912A>G, p.(Asp971Gly) 	0000			c.9260A > G. p.(Lvs3087Arg)					
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	B170	na	Chinese	c.2542G>A, p.(Val848Met)	BR	na	na	na	[39]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $				$c_{11624A>G} p(V_{us}^{(1)})$					
$\begin{array}{c c c c c c c } \hline \begin{tabular}{ c c c c c c c c c c c c c c c c c c c$	B156	na	Chinese	c.2542G>A, p.(Val848Met)	BR	na	na	na	[39]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $				c.12071C>T. p.(Ser4024Phe)		1			
$\begin{array}{cccccccccccccccccccccccccccccccccccc$	11174	М	Italian	c 2753G > T p (Glv918Val)	Bronchitis rhinitis sinusitis otitis	SS	_	PCD (I)	[23]
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $				c.12796_12801delinsATA, p. Phe4266_Asn4267delinsIle					
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	#4	F	English	c.2832dup. p (Gln945Serfs*10)	Recurrent chest infection, rhinitis	SS	-	PCD	[42]
B142naChinese $c.2912A > G, p.(Asp971Gly)$ $c.11396T > C, p.(Ile3799Thr)$ BRnanana[39]P25MChinese $c.3020T > G, p.(Leu1007^*)$ $c.3470T > G, p.(Leu1157Arg)$ Rhinosinusitis, OMSI-KS (N)[37]#6FEnglish $c.3220G > T, p.(Glu1074^*)$ $c.13069C > T, p.(Arg4357^*)$ Recurrent chest infectionSI-KS (Static)[42]II:2MChinese $c.3426-1G > A$ $c.4306C > T, p.(Arg1436Trp)-HTX+naThis study$				c.13240dup, p.(Thr4414Asnfs*34)					
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	B142	na	Chinese	c.2912A>G, p.(Asp971Gly)	BR	na	na	na	[39]
P25MChinese $c.3020T>G, p.(Leu1007^*)$ $c.3470T>G, p.(Leu1157Arg)$ Rhinosinusitis, OMSI-KS (N)[37]#6FEnglish $c.3220G>T, p.(Glu1074^*)$ $c.13069C>T, p.(Arg4357^*)$ Recurrent chest infectionSI-KS (Static)[42]II:2MChinese $c.3426-1G>A$ $c.4306C>T, p.(Arg1436Trp)-HTX+naThis study$				c.11396T>C, p.(Ile3799Thr)					
$ \begin{array}{c c c c c c c c c c c c c c c c c c c $	P25	М	Chinese	c.3020T>G, p.(Leu1007*)	Rhinosinusitis. OM	SI	-	KS (N)	[37]
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$				c.3470T>G, p.(Leu1157Arg)					
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	#6	F	English	c.3220G>T. p.(Glu1074*)	Recurrent chest infection	SI	-	KS (Static)	[42]
II.2 M Chinese $\frac{c.3426-1G>A}{c.4306C>T, p.(Arg1436Trp)}$ - HTX + na This study		-	8-1011	c.13069C>T. p.(Arg4357*)					[]
c.4306C>T, p.(Arg1436Trp)	II:2	м	Chinese	c.3426-1G>A	-	нтх	+	na	This study
				c.4306C>T, p.(Arg1436Trp)					

Table 1. Clinical phenotypes of individuals with DNAH11 compound heterozygous variants.

(Continued)

Patients	Gender	Ethnicity	Sequence variants <sup>#</sup>	PCD phenotypes	Situs	CHD	Ciliary motion	References
P45	F	Chinese	c.3470T>G, p.(Leu1157Arg)	Rhinosinusitis, OM	SI	-	KS (R)	[37]
			c.6727C>G, p.(Arg2243Gly)					
#7	М	English	c.3544C>T, p.(Arg1182*)	Recurrent chest infection, rhinitis	SS	-	PCD	[42]
			c.8798-5G>A					
#1	М	English	c.3727G>T, p.(Glu1243*)	Recurrent chest infection, rhinitis	SS	-	PCD (Static)	[42]
			c.13531_13532insTTCAGGCTGAAGA, p. (Ala4511Valfs*13)					
PCD1077	F	Caucasian	c.3901G>T, p.(Glu1301*)	NRD, sinusitis, OM	SI	-	KS	[22]
			c.11804C>T, p.(Pro3935Leu)					
OP406-II:1	М	Caucasian	c.4254+5G>T	na	SI		KS (H)	[22]
OP406-II:2	F		c.4726-1G>A	NRD, sinusitis	SS		PCD (H)	
B099	na	Chinese	c.4306C>T, p.(Arg1436Trp)	BR	na	na	na	[39]
			c.6118C>T, p.(Arg2040Cys)					
#2	F	English	c.4395_4398del, p.(Ser1465Argfs*6)	Recurrent chest infection, rhinitis	SS	-	PCD (Static)	[42]
			c.7642C>T, p.(Gln2548*)					
P32	М	Chinese	c.4457T>A, p.(Leu1486Gln)	NRD, rhinosinusitis, OM	SS	-	PCD (N)	[37]
			c.10006G>T, p.(Ala3336Ser)					
9003	F	American	c.4505A>C, p.(Gln1502Pro)	NRD, recurrent pneumonia, BR,	HTX	+	CD (H)	[43]
			c.9376G>A, p.(Glu3126Lys)	sinusitis, OM				
PCD106	М	Caucasian	c.4516_4517del, p.(Leu1506Serfs*11)	Sinusitis, OM	SS	-	PCD	[22]
PCD108	М		c.7266+1G>A	NRD, sinusitis, OM	SI		KS (H)	
B185	na	Chinese	c.4898C>A, p.(Ser1633Tyr)	BR	na	na	na	[39]
			c.9260A>G, p.(Lys3087Arg)					
11228	М	Italian	c.4922C>G, p.(Ser1641*)	Bronchitis, rhinitis	SI	-	KS	[23]
			c.9304G>A, p.(Gly3102Ser)					
#5707	М	Chinese	c.5473dup, p.(Gln1825Profs*23)	na	HTX	+	CD (I+R)	[35]
			c.8275T>C, p.(Phe2759Leu)					
			c.13183C>T, p.(Arg4395*)					
#3	F	English	c.5506C>T, p.(Arg1836*)	Recurrent chest infection	SI	-	KS (Static)	[42]
		0	c.5636T>A, p.(Leu1879*)	-				
PCD565	М	Caucasian	c.5778+1G>A	NRD, BR, sinusitis, OM	SI	-	KS (H)	[22]
			c.13061T>A, p.(Leu4354His)					
PCD812	м	Caucasian	c.5815G>A, p.(Glv1939Arg)	NRD, sinusitis, OM	SI	-	KS	[22]
			c.13373C>T, p.(Pro4458Leu)					
PCD157	F	Caucasian	c.6244C>T, p.(Arg2082*)	NRD, BR, sinusitis, OM	SI	-	KS (H)	[22]
			c.11929G>T, p.(Glu3977*)					
#5130	F	Chinese	c.6785T>C, p.(Ile2262Thr)	na	HTX	+	CD (R+D)	[35]
			c.11398G>C, p.(Asp3800His)					
B178	na	Chinese	c.6905A>C, p.(His2302Pro)	BR	na	na	na	[39]
			c.13112C>T, p.(Pro4371Leu)					
B073	na	Chinese	c.7292G>T. p.(Ser2431[le)	BR na	na	na	na	[39]
			c.9017C>T.p.(Thr3006Met)					
P17	м	Chinese	c.7292G>T. p. (Ser2431Ile)	NRD, rhinosinusitis OM	SI	-	KS (R)	[37]
P30	М		c.7364A>C. p.(Asp2455Ala)	NRD	SS	+	PCD (R)	
150			c 9017C>T. p (Thr3006Met)					
			c.13373C>T. p.(Pro4458Len)	-				
	1	1				1	1	

Table 1. (Continued)

(Continued)

Patients	Gender	Ethnicity	Sequence variants <sup>#</sup>	PCD phenotypes	Situs	CHD	Ciliary motion	References
#730	na	Caucasian	c.7772C>T, p.(Pro2591Leu)	NRD, rhinitis, sinusitis, OM	SS	-	PCD	[34]
			c.8698C>T, p.(Arg2900*)				(erratic)	
OP98-II:1	М	Caucasian	c.7914G>C, p.Trp2604*	BR, sinusitis, OM	SI	-	KS (H)	[22]
OP98-II:2	М	]	c.13330_13333dup, p.(Ile4445Asnfs*4)		SS		PCD (H)	
С	М	Italian	c.8114A>G, p.(His2705Arg)	NRD, recurrent pneumonia, sinusitis, otitis	SI	+	KS (I+H)	[38]
			c.10264G>A, p.(Gly3422Arg)					
B167	na	Chinese	c.9260A>G, p.(Lys3087Arg)	BR	na	na	na	[ <u>39</u> ]
			c.11647C>T, p.(Leu3883Phe)					
B163	na	Chinese	c.9260A>G, p.(Lys3087Arg)	BR	na	na	na	[39]
			c.13175C>T, p.(Thr4392Met)					
P23	F	Chinese	c.9539T>A, p.(Leu3180*)	Rhinosinusitis, OM	SI	-	KS (R)	[37]
			c.9706C>T, p.(Arg3236*)					
#5045	М	Chinese	c.10379C>A, p.(Thr3460Lys)	na	HTX	+	CD (R+D)	[35]
			c.13273G>A, p.(Gly4425Ser)					
PCD1126	F	Asian	c.12064G>C, p.(Ala4022Pro)	BR, sinusitis	SS	-	PCD (H)	[22]
			c.13500_13504dup, p.(Thr4502Argfs*15)					
II-2	М	German	erman c.12363C>G, p.Tyr4121* c.13531_*36del, p.Ala4511_Ala4516delinsGln	Recurrent pneumonia, bronchitis, sinusitis, BR (II-4, 6, 11), OM (II-3, 4, 9)	SI	-	KS	[27]
							PCD	
II-3, 4	F				SS		PCD	
II-6, 9, 11	М				SS			
OP235-II:1	F	Caucasian	c.12697C>T, p.(Gln4233*)	BR, sinusitis, OM	SS	-	PCD (H)	[22]
OP235-II:2	F		c.12980T>C, p.(Leu4327Ser)	NRD, BR, sinusitis, OM	SI		KS (H)	]
PCD918	F	Asian	c.13065_13067del, p.(Leu4356del)	BR, NRD, sinusitis, OM	SS	-	PCD	[22]
PCD919	М		c.13075C>T, p.(Arg4359*)		HTX		KS (H)	

#### Table 1. (Continued)

<sup>#</sup> Description of the sequence variants is recalibrated in accordance with the Human Genome Variation Society nomenclature for variants (<u>http://varnomen.hgvs.org/</u>) using the reference sequence (NM\_001277115.1).

BR, bronchiectasis; CD, ciliary dysfunction; CHD, congenital heart disease; D, discordance; F, female; H, hyperkinetic; HTX, heterotaxy; I, immotile; KS, Kartagener syndrome; M, male; N, nearly normal; na, not available; NRD, neonatal respiratory distress; OM, otitis media; PCD, primary ciliary dyskinesia; R, restricted; SI, situs inversus; SS, situs solitus; +, present; -, not present.

https://doi.org/10.1371/journal.pone.0252786.t001

media, bronchitis, frequent pneumonias, or bronchiectasis were observed in the two family members having the *DNAH11* compound heterozygous variants. However, the very minor or preclinical phenotypes of PCD, such as rhinitis and bronchitis, might be overlooked and a diagnosis of mild PCD may be not excluded. In this study, there is a limitation that the father's genotyping is unavailable due to his death. He died from an accidental injury, without complaining of PCD-related rhinitis, sinusitis, otitis media, bronchitis, bronchiectasis, or cardiovascular anomaly.

In conclusion, the compound heterozygous variants, c.3426-1G>A and c.4306C>T, in the *DNAH11* gene might be the pathogenic alterations in this family with HTX and CHD. Our findings broaden the variant spectrum of the *DNAH11* gene and provide additional information usable in genetic counseling for this family.

#### Acknowledgments

The authors are grateful to the participating individuals and investigators for their co-operation and efforts in collecting clinical and genetic information and DNA specimens.

#### **Author Contributions**

**Conceptualization:** Hong Xia, Hao Deng.

Data curation: Hong Xia, Xiangjun Huang, Sheng Deng.

Formal analysis: Hong Xia, Sheng Deng, Hongbo Xu, Hao Deng.

Funding acquisition: Hong Xia, Lamei Yuan, Hao Deng.

Investigation: Hong Xia, Xiangjun Huang, Yan Yang, Xin Liu.

Methodology: Hong Xia, Lamei Yuan, Hao Deng.

Project administration: Hong Xia, Hao Deng.

Resources: Hong Xia.

Software: Lamei Yuan.

Supervision: Lamei Yuan, Hao Deng.

Validation: Lamei Yuan, Hao Deng.

Visualization: Hong Xia, Hao Deng.

Writing - original draft: Hong Xia, Hao Deng.

Writing – review & editing: Hong Xia, Lamei Yuan, Hao Deng.

#### References

- Deng H, Xia H, Deng S. Genetic basis of human left-right asymmetry disorders. Expert Rev Mol Med. 2015; 16:e19. https://doi.org/10.1017/erm.2014.22 PMID: 26258520.
- Jacobs JP, Anderson RH, Weinberg PM, Walters HL 3rd, Tchervenkov CI, Del Duca D, et al. The nomenclature, definition and classification of cardiac structures in the setting of heterotaxy. Cardiol Young. 2007; 17 Suppl 2:1–28. https://doi.org/10.1017/S1047951107001138 PMID: 18039396.
- Lin AE, Ticho BS, Houde K, Westgate MN, Holmes LB. Heterotaxy: associated conditions and hospitalbased prevalence in newborns. Genet Med. 2000; 2(3):157–72. https://doi.org/10.1097/00125817-200005000-00002 PMID: 11256661.
- Sutherland MJ, Ware SM. Disorders of left-right asymmetry: heterotaxy and situs inversus. Am J Med Genet C Semin Med Genet. 2009; 151C(4):307–17. <u>https://doi.org/10.1002/ajmg.c.30228</u> PMID: 19876930.
- Peeters H, Devriendt K. Human laterality disorders. Eur J Med Genet. 2006; 49(5):349–62. <u>https://doi.org/10.1016/j.ejmg.2005.12.003</u> PMID: 16461029.
- Gebbia M, Ferrero GB, Pilia G, Bassi MT, Aylsworth A, Penman-Splitt M, et al. X-linked situs abnormalities result from mutations in ZIC3. Nat Genet. 1997; 17(3):305–8. https://doi.org/10.1038/ng1197-305 PMID: 9354794.
- Bamford RN, Roessler E, Burdine RD, Saplakoğlu U, dela Cruz J, Splitt M, et al. Loss-of-function mutations in the EGF-CFC gene CFC1 are associated with human left-right laterality defects. Nat Genet. 2000; 26(3):365–9. https://doi.org/10.1038/81695 PMID: 11062482.
- Kato R, Yamada Y, Niikawa N. De novo balanced translocation (6;18)(q21;q21.3) in a patient with heterotaxia. Am J Med Genet. 1996; 66(2):184–6. https://doi.org/10.1002/(SICI)1096-8628(19961211) 66:2<184::AID-AJMG11>3.0.CO;2-P PMID: 8958327.
- Kosaki R, Gebbia M, Kosaki K, Lewin M, Bowers P, Towbin JA, et al. Left-right axis malformations associated with mutations in ACVR2B, the gene for human activin receptor type IIB. Am J Med Genet. 1999; 82(1):70–6. https://doi.org/10.1002/(sici)1096-8628(19990101)82:1<70::aid-ajmg14>3.0.co;2-y PMID: 9916847.
- Mohapatra B, Casey B, Li H, Ho-Dawson T, Smith L, Fernbach SD, et al. Identification and functional characterization of NODAL rare variants in heterotaxy and isolated cardiovascular malformations. Hum Mol Genet. 2009; 18(5):861–71. https://doi.org/10.1093/hmg/ddn411 PMID: 19064609.

- Perles Z, Cinnamon Y, Ta-Shma A, Shaag A, Einbinder T, Rein AJ, et al. A human laterality disorder associated with recessive CCDC11 mutation. J Med Genet. 2012; 49(6):386–90. https://doi.org/10. 1136/jmedgenet-2011-100457 PMID: 22577226.
- Perles Z, Moon S, Ta-Shma A, Yaacov B, Francescatto L, Edvardson S, et al. A human laterality disorder caused by a homozygous deleterious mutation in MMP21. J Med Genet. 2015; 52(12):840–7. https://doi.org/10.1136/jmedgenet-2015-103336 PMID: 26429889.
- Vetrini F D'Alessandro LC, Akdemir ZC, Braxton A, Azamian MS, Eldomery MK, et al. Bi-allelic mutations in PKD1L1 are associated with laterality defects in humans. Am J Hum Genet. 2016; 99(4):886– 93. https://doi.org/10.1016/j.ajhg.2016.07.011 PMID: 27616478.
- Ta-Shma A, Hjeij R, Perles Z, Dougherty GW, Abu Zahira I, Letteboer SJF, et al. Homozygous loss-offunction mutations in MNS1 cause laterality defects and likely male infertility. PLoS Genet. 2018; 14(8): e1007602. https://doi.org/10.1371/journal.pgen.1007602 PMID: 30148830.
- Deng S, Wu S, Xia H, Xiong W, Deng X, Liao J, et al. Identification of a frame shift mutation in the CCDC151 gene in a Han-Chinese family with Kartagener syndrome. Biosci Rep. 2020; 40(6): BSR20192510. https://doi.org/10.1042/BSR20192510 PMID: 32490514.
- Chen X, Deng S, Xia H, Yuan L, Xu H, Tang S, et al. Identification of a CCDC114 variant in a Han-Chinese patient with situs inversus. Exp Ther Med. 2020; 20(4):3336–42. <u>https://doi.org/10.3892/etm.</u> 2020.9059 PMID: 32855706.
- Xia H, Hu P, Yuan L, Xiong W, Xu H, Yi J, et al. A homozygous MYO7A mutation associated to Usher syndrome and unilateral auditory neuropathy spectrum disorder. Mol Med Rep. 2017; 16(4):4241–6. https://doi.org/10.3892/mmr.2017.7053 PMID: 28731162.
- Hu P, Wu S, Yuan L, Lin Q, Zheng W, Xia H, et al. Compound heterozygous POMT1 mutations in a Chinese family with autosomal recessive muscular dystrophy-dystroglycanopathy C1. J Cell Mol Med. 2017; 21(7):1388–93. https://doi.org/10.1111/jcmm.13068 PMID: 28157257.
- Xia H, Huang X, Xu H, Zhou YA, Gong L, Yang Z, et al. GJB2 c.235delC variant associated with autosomal recessive nonsyndromic hearing loss and auditory neuropathy spectrum disorder. Genet Mol Biol. 2019; 42(1):48–51. https://doi.org/10.1590/1678-4685-gmb-2017-0318 PMID: 30816908.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. Genet Med. 2015; 17(5):405– 24. https://doi.org/10.1038/gim.2015.30 PMID: 25741868.
- Abou Tayoun AN, Pesaran T, DiStefano MT, Oza A, Rehm HL, Biesecker LG, et al. Recommendations for interpreting the loss of function PVS1 ACMG/AMP variant criterion. Hum Mutat. 2018; 39(11):1517– 24. https://doi.org/10.1002/humu.23626 PMID: 30192042.
- 22. Knowles MR, Leigh MW, Carson JL, Davis SD, Dell SD, Ferkol TW, et al. Mutations of DNAH11 in patients with primary ciliary dyskinesia with normal ciliary ultrastructure. Thorax. 2012; 67(5):433–41. https://doi.org/10.1136/thoraxjnl-2011-200301 PMID: 22184204.
- Boaretto F, Snijders D, Salvoro C, Spalletta A, Mostacciuolo ML, Collura M, et al. Diagnosis of primary ciliary dyskinesia by a targeted next-generation sequencing panel: molecular and clinical findings in Italian patients. J Mol Diagn. 2016; 18(6):912–922. https://doi.org/10.1016/j.jmoldx.2016.07.002 PMID: 27637300.
- Kennedy MP, Omran H, Leigh MW, Dell S, Morgan L, Molina PL, et al. Congenital heart disease and other heterotaxic defects in a large cohort of patients with primary ciliary dyskinesia. Circulation. 2007; 115(22):2814–21. https://doi.org/10.1161/CIRCULATIONAHA.106.649038 PMID: 17515466.
- Shapiro AJ, Davis SD, Ferkol T, Dell SD, Rosenfeld M, Olivier KN, et al. Laterality defects other than situs inversus totalis in primary ciliary dyskinesia: insights into situs ambiguus and heterotaxy. Chest. 2014; 146(5):1176–86. https://doi.org/10.1378/chest.13-1704 PMID: 24577564.
- Bartoloni L, Blouin JL, Pan Y, Gehrig C, Maiti AK, Scamuffa N, et al. Mutations in the DNAH11 (axonemal heavy chain dynein type 11) gene cause one form of situs inversus totalis and most likely primary ciliary dyskinesia. Proc Natl Acad Sci U S A. 2002; 99(16):10282–6. https://doi.org/10.1073/pnas. 152337699 PMID: 12142464.
- Schwabe GC, Hoffmann K, Loges NT, Birker D, Rossier C, de Santi MM, et al. Primary ciliary dyskinesia associated with normal axoneme ultrastructure is caused by DNAH11 mutations. Hum Mutat. 2008; 29 (2):289–98. https://doi.org/10.1002/humu.20656 PMID: 18022865.
- Supp DM, Witte DP, Potter SS, Brueckner M. Mutation of an axonemal dynein affects left-right asymmetry in inversus viscerum mice. Nature. 1997; 389(6654):963–6. https://doi.org/10.1038/40140 PMID: 9353118.
- Nonaka S, Shiratori H, Saijoh Y, Hamada H. Determination of left-right patterning of the mouse embryo by artificial nodal flow. Nature. 2002; 418(6893):96–9. <u>https://doi.org/10.1038/nature00849</u> PMID: 12097914.

- McGrath J, Somlo S, Makova S, Tian X, Brueckner M. Two populations of node monocilia initiate leftright asymmetry in the mouse. Cell. 2003; 114(1):61–73. <u>https://doi.org/10.1016/s0092-8674(03)</u> 00511-7 PMID: 12859898.
- Brueckner M D'Eustachio P, Horwich AL. Linkage mapping of a mouse gene, iv, that controls left-right asymmetry of the heart and viscera. Proc Natl Acad Sci U S A. 1989; 86(13):5035–8. <u>https://doi.org/10.1073/pnas.86.13.5035 PMID: 2740340</u>.
- Hanzlik AJ, Binder M, Layton WM, Rowe L, Layton M, Taylor BA, et al. The murine situs inversus viscerum (iv) gene responsible for visceral asymmetry is linked tightly to the lgh-C cluster on chromosome 12. Genomics. 1990; 7(3):389–93. https://doi.org/10.1016/0888-7543(90)90173-r PMID: 2365357.
- Okada Y, Nonaka S, Tanaka Y, Saijoh Y, Hamada H, Hirokawa N. Abnormal nodal flow precedes situs inversus in iv and inv mice. Mol Cell. 1999; 4(4):459–68. <u>https://doi.org/10.1016/s1097-2765(00)</u> 80197-5 PMID: 10549278.
- Lucas JS, Adam EC, Goggin PM, Jackson CL, Powles-Glover N, Patel SH, et al. Static respiratory cilia associated with mutations in Dnahc11/DNAH11: a mouse model of PCD. Hum Mutat. 2012; 33(3):495– 503. https://doi.org/10.1002/humu.22001 PMID: 22102620.
- Liu S, Chen W, Zhan Y, Li S, Ma X, Ma D, et al. DNAH11 variants and its association with congenital heart disease and heterotaxy syndrome. Sci Rep. 2019; 9(1):6683. <u>https://doi.org/10.1038/s41598-019-43109-6 PMID: 31040315.</u>
- Zhang L, Feng X, Zhang J, Hao Y, Wang Y. Co-occurrence of Moyamoya syndrome and Kartagener syndrome caused by the mutation of DNAH5 and DNAH11: a case report. BMC Neurol. 2020; 20 (1):314. https://doi.org/10.1186/s12883-020-01895-x PMID: 32847546.
- Guo Z, Chen W, Wang L, Qian L. Clinical and genetic spectrum of children with primary ciliary dyskinesia in China. J Pediatr. 2020; 225:157–65.e5. https://doi.org/10.1016/j.jpeds.2020.05.052 PMID: 32502479.
- Pifferi M, Michelucci A, Conidi ME, Cangiotti AM, Simi P, Macchia P, et al. New DNAH11 mutations in primary ciliary dyskinesia with normal axonemal ultrastructure. Eur Respir J. 2010; 35(6):1413–6. https://doi.org/10.1183/09031936.00186209 PMID: 20513915.
- Guan WJ, Li JC, Liu F, Zhou J, Liu YP, Ling C, et al. Next-generation sequencing for identifying genetic mutations in adults with bronchiectasis. J Thorac Dis. 2018; 10(5):2618–30. <u>https://doi.org/10.21037/</u> jtd.2018.04.134 PMID: 29997923.
- 40. Namavarian A, Eid A, Goh ES-Y, Thakur V. A novel DNAH11 variant segregating in a sibship with heterotaxy and implications for genetic counseling. Mol Genet Genomic Med. 2020; 8:e1358. <u>https://doi.org/10.1002/mgg3.1358</u> PMID: 32633470.
- Schultz R, Elenius V, Lukkarinen H, Saarela T. Two novel mutations in the DNAH11 gene in primary ciliary dyskinesia (CILD7) with considerable variety in the clinical and beating cilia phenotype. BMC Med Genet. 2020; 21(1):237. https://doi.org/10.1186/s12881-020-01171-2 PMID: 33243178.
- Shoemark A, Burgoyne T, Kwan R, Dixon M, Patel MP, Rogers AV, et al. Primary ciliary dyskinesia with normal ultrastructure: three-dimensional tomography detects absence of DNAH11. Eur Respir J. 2018; 51(2):1701809. https://doi.org/10.1183/13993003.01809-2017 PMID: 29467202.
- **43.** Nakhleh N, Francis R, Giese RA, Tian X, Li Y, Zariwala MA, et al. High prevalence of respiratory ciliary dysfunction in congenital heart disease patients with heterotaxy. Circulation. 2012; 125(18):2232–42. https://doi.org/10.1161/CIRCULATIONAHA.111.079780 PMID: 22499950.
- 44. Fu F, Li R, Li Y, Nie ZQ, Lei T, Wang D, et al. Whole exome sequencing as a diagnostic adjunct to clinical testing in fetuses with structural abnormalities. Ultrasound Obstet Gynecol. 2018; 51(4):493–502. https://doi.org/10.1002/uog.18915 PMID: 28976722.