

REVIEW

Open Access

How tissue injury alarms the immune system and causes a systemic inflammatory response syndrome

Jérôme Pugin*

Abstract

Systemic inflammation is very prevalent among critically ill patients, particularly those with extensive tissue injury. Although downstream mediators (cytokines) and effector cells (phagocytes) have been identified, proximal mediators originating from injured tissues remained elusive. Alarmins (“danger signals”) released by necrotic/injured cells have been identified recently and certainly play a role in triggering local and systemic inflammation in critically ill patients. The most promising alarmin candidates are of mitochondrial origin, i.e. mitochondrial DNA and the chemotactic factor fMet-Leu-Phe (fMLP). ATP also is released from necrotic tissues and stimulates the assembly of the inflammasome, leading to the production of proinflammatory cytokines, such as interleukin (IL)-1 β . The identification of novel alarmins opens new therapeutic avenues for the treatment of severe SIRS, and SIRS-dependent organ dysfunction.

Keywords: SIRS, Inflammation, Alarmins, Danger signals, Mitochondria, Pro-inflammatory cytokines

Review

Introduction

Clinicians have recognized for a long time that critically ill patients often present with a stress response associating tachycardia and tachypnea, together with an “inflammatory” response characterized by fever and leukocytosis. The systemic inflammatory response syndrome (SIRS) concept was coined in 1991 by the late Roger C. Bone [1]. SIRS criteria were rapidly incorporated into the definition of sepsis during the first ACCP/SCCM consensus conference published two decades ago [2]. Sepsis was defined as bacterial infection with a systemic response, associating at least two of four SIRS criteria [3]. These definitions, although not perfect, still hold today, and are used in all sepsis studies, as well as at the bedside. Aside from its use in the sepsis definition, SIRS was rapidly recognized as a very prevalent syndrome in intensive care units (ICU), even in non-infected patients. Up to 93% of surgical ICU patients were reported to meet SIRS criteria, with ICU mortality ranging from 7–9% in the absence of an infection [3]. It also was epidemiologically demonstrated

that SIRS may precede sepsis [4]. Because of its high prevalence in ICUs and the absence of a clear demonstration of a link between clinical SIRS and systemic inflammation, some of our colleagues have proposed to abandon the SIRS concept [5]. Interestingly, new tentative sepsis definitions also included a “response” item, like in the PIRO concept (predisposition, infection, *response*, and organ dysfunction) [6]. “Symptoms” such as heart rate, respiratory frequency, and band forms were retained in a PIRO score utile for stratification of critically ill patients and outcome prediction [7].

It was convincingly shown since the late 1980s that SIRS in critically ill patients was associated with the circulation of pro- and anti-inflammatory cytokines, even in the absence of infection [8]. It also was demonstrated that the balance of pro/anti-inflammatory mediators was pro-inflammatory in the inflamed organ (bioactive TNF and IL-1 β in lungs from ARDS patients, in ascitic fluid during pancreatitis, etc.) but was found to be anti-inflammatory in the circulation [9,10]. It was proposed that the normal response to local infection or tissue injury was to produce a *local* inflammatory reaction, attracting effectors phagocytes at the site of injuries or infection [11]. In contrast, an excess of anti-inflammatory mediators (soluble TNF

Correspondence: jerome.pugin@unige.ch
Intensive Care - SIRS Unit, University Hospitals of Geneva, 1211 Geneva 14, Switzerland

receptors, IL-1RA, IL-10) prevented “systemic inflammation” within the circulatory compartment [11].

Although effector cells and their mediators have been described during SIRS (phagocytes, cytokines, chemokines) [12], what lacked in the paradigm of tissue injury-induced inflammation was the identification of endogenous mediators originating from injured tissues that were capable of generating this inflammatory/immune reaction [13]. It is the purpose of this review to discuss recent findings in the field of the pathogenesis of SIRS and the relevance of novel candidate endogenous danger signals.

The danger theory

The long time prevalent theory of the immune system was based on tolerance of self-antigens developed mostly during the fetal life by the deletion of auto-reactive lymphocytes in the thymus and bone marrow. In this theory, only remain during *extra utero* life immune cells capable of reacting to non-self antigens preventing autoimmunity, and conferring immunity to microbes and allografts. Sir Frank Macfarlane Burnet and Peter Medawar were awarded the Nobel Prize in 1960 for this innovative concept for self-/non-self-discrimination. Ample experimental evidence supported this paradigm; however, it appeared that it was somewhat reductive and could not completely explain various situations of tolerance to foreign antigens or immune/inflammatory reactions to endogenous antigens. How to explain with the self-/non-self-discrimination theory the absence of sepsis after tooth brushing where bacteremia is frequent? Why pregnant women do not reject their fetus, although half of its antigens coming from the father should be recognized as non-self? Why lactating women do not reject their breasts despite the expression in adulthood of “novel” milk proteins? In addition, because the immune system should be tolerant to self-antigens, how is autoimmunity explained?

Early in the 1990s, Polly Matzinger came up with the concept that the immune system did not care about self- vs. non-self but discriminated antigens based on the fact that were sensed as dangerous or not [14]. This “danger theory” may explain why pregnant women do not reject their nondangerous fetus and lactating women do not recognize newly expressed milk protein as dangerous. This also explains why circulating indolent gingival bacteria do not generate a sepsis syndrome after tooth brushing, because they are not seen as danger by our immune system. In this theory, what is essential for the immune system to react to an antigen is a signal of tissue injury, sensing the danger [15,16]. For the immune system to react, two signals are needed: one from the antigen and another one from tissue suffering. Vaccination is another example validating the danger model. Vaccines are usually efficacious only with a certain degree of tissue injury induced by the irritating effect at the site of

injection of the adjuvant. The sole injection of the microbial antigen is usually well tolerated and does not induce a protective immune response. The danger theory was well captured by Polly Matzinger in an interview given to the New York Times in 1998: *“Imagine a community in which the police accept anyone they met during elementary school and kill any new migrant. That’s the Self/Nonself Model. In the Danger Model, tourists and immigrants are accepted, until they start breaking windows. Only then, do the police move to eliminate them. In fact, it doesn’t matter if the window breaker is a foreigner or a member of the community.”* [17].

This danger theory also makes a lot of sense to the intensivist for several reasons [13]. SIRS could arise from an overwhelmed and systemic inflammatory reaction to massive release of danger signals from injured tissues. Examples of such situations in the intensive care unit are numerous: multiple trauma, pancreatitis, ischemia-reperfusion injury, shock, massive transfusions, and major surgery [13]. The devastating effects of sepsis may be explained by a synergistic activation of the inflammatory/immune system by foreign bacterial antigens and bacteria-induced tissue injury, further increased by shock, leading to organ dysfunction [13]. This synergism also may explain why patients with SIRS have such overwhelmed responses to bacterial infections acquired during their ICU stay.

Endogenous danger signals (alarmins)

What was lacking to support the danger theory was the identification of molecules originating from host cells signaling tissue injury to the immune system (alarmins, danger-associated molecular patterns, DAMPs). The definition of an alarmin could be a molecule released by injured cells/tissues that is capable of producing a local proinflammatory reaction, and attract to the site of injury effector cells such as phagocytes [18]. Alarmins not only may generate tissue inflammation after injury, but they also could act synergistically with microbial non-self-antigens to enhance the inflammatory reaction [19]. In case of massive tissue injury, the local production of alarmins could spillover into the circulation and produce systemic symptoms, such as fever, leukocytosis, as well as a stress humoral and neural response [19]. Leukocytes activated by alarmins may then relay and amplify the inflammatory process through the secretion of cytokines, the release of molecules, such as enzymes, lipid mediators, reactive oxygen, and nitrogen species [20]. The importance of tissue injury and the presence of microbial co-factors may account for the magnitude of the local vs. systemic inflammatory response (SIRS), vasodilation, and end-organ dysfunction.

Alarmins represent nothing less than the missing molecular link between tissue injury and the inflammatory

response to tissue suffering. Conceptually, alarmins should be molecules present inside of cells sheltered from the immune system, and only released during tissue injury, most probably during cells necrosis or plasma membrane rupture [21]. Indeed, it has been shown that cell death by apoptosis does not produce inflammation. In addition, receptors for these danger signals should exist in leukocytes, and possibly in other cell types sensing the presence of endogenous alarmins that have become extracellular.

High mobility group box 1 (HMGB1)

Since the appearance of the concept of endogenous danger signals, several candidate alarmins have been proposed [18]. HMGB1 is a DNA chaperone concentrated in the cell nucleus first demonstrated to induce cell migration and differentiation. Its role in the pathogenesis of sepsis was reported in 1999 by the group of Kevin Tracey [22] and by others in the context of severe SIRS [23,24]. These investigators showed that HMGB1 was released by macrophages stimulated by lipopolysaccharide (LPS), tumor necrosis factor- α (TNF- α), and IL-1 β [22] and that high HMGB1 was measured in plasma from patients with septic shock [25]. Administration of antibodies to HMGB1 prevented late death in a murine model of sepsis [22]. In contrast, the injection of recombinant HMGB1 increased septic mouse lethality [22]. Since this pioneer work, many groups have shown a proinflammatory activity of HMGB1 and have identified putative surface leukocyte receptors for HMGB1 (RAGE, TLR2, and TLR4) [26,27]. It has been recently well established that most of—if not all—the inflammatory activity was not contained in the HMGB1 protein, but rather in bacterial contaminants bound to it, explaining TLR-dependent cell activation, HMGB1 only playing a role of chaperone for bacterial products, including LPS [28,29]. HMGB1 also can be released from dying cells and become a potential extracellular danger signal, but its role as an alarmin remains to be clearly demonstrated [30-32].

Heat shock proteins

HSPs are ubiquitous proteins, present in the cytoplasm, mitochondria, the cell nucleus, and act as molecular chaperones for proteins. They can be secreted by stressed cells and are released by necrotic cells [33]. The potential pro-inflammatory and immune-inducing effects of extracellular HSPs were first proposed in 1993, later thought to be mediated by TLR2 and TLR4 receptors. As with HMGB1, highly purified HSPs lacked cytokine function [29]. It is now believed that HSPs, like HMGB1, act more as molecular chaperones for bacterial products (LPS, flagellin, and lipopeptides) and co-factors for cell activation to pathogen-associated molecular patterns. Therefore,

the role of HSPs and HMGB1 in sterile inflammation and SIRS remains elusive [28,29].

Other putative alarmins

S100 proteins (also called calgranulins) are a group of calcium-binding proteins that have been proposed as potential alarmins, particularly those expressed in phagocytes (S100A8, -A9, and -A12) through their interaction with RAGE and TLR4 [34,35]. Again, no evidence exists to show a clear danger signal effect of these proteins in the absence of bacterial products. Crystals of uric acid were shown to induce the assembly of the NALP3 inflammasome, leading to the cleavage of pro-IL-1 β into mature IL-1 β by caspase-1 [36]. Whereas uric acid is now recognized as a local danger signal in the pathogenesis of gout arthritis, none reported a role of this alarmin in systemic inflammation. Recently, another alarmin released by necrotic cells has been proposed: IL-33 [37]. This mediator acts through the receptor ST2 at the surface of CD8⁺ T cells, enhancing T cell clonal expansion and was shown to be necessary for the control of RNA and DNA viral infections in mice [37].

Mitochondrial alarmins

Mitochondria are ancient bacteria that have become endosymbionts and then organelles specialized with the evolution into oxygen-dependent energy producing factories (ATP production, cellular respiration) [38]. Two important features of mitochondria are that: 1) they have retained some molecules of bacterial origin throughout the evolution, and 2) they escape the immune system due to their obligatory intracellular location but can potentially be recognized as non-self when they are found extracellularly. Evidence has recently accumulated that mitochondrial alarmins mediate inflammation [39]. It was recently demonstrated that mitochondrial DNA (resembling to bacterial DNA) was found extracellularly after tissue injury, in the plasma from multiple trauma patients [40]. MtDNA, like bacterial DNA, is proinflammatory via the activation of Toll-like receptor 9 present in many cell types. The formulated peptide fMet-Leu-Phe (or fMLP) is another molecule present in the wall of both bacteria and mitochondria, but absent in any other human structure. It was shown that the release of mitochondria during tissue injury attracted phagocytes via the chemoattractant fMLP, through its ligation to the fMLP receptor (FPR1) present at the surface of neutrophils [40]. By far, mitochondrial DAMPs are to date the best candidate endogenous danger signals; they respond to all alarmin criteria [18,41]. Injected to animals, mitochondrial preparations induce lung injury resembling to that seen during ARDS [40]. A schematic view of the action of mitochondrial danger signals acting as alarmins is proposed in Figure 1.

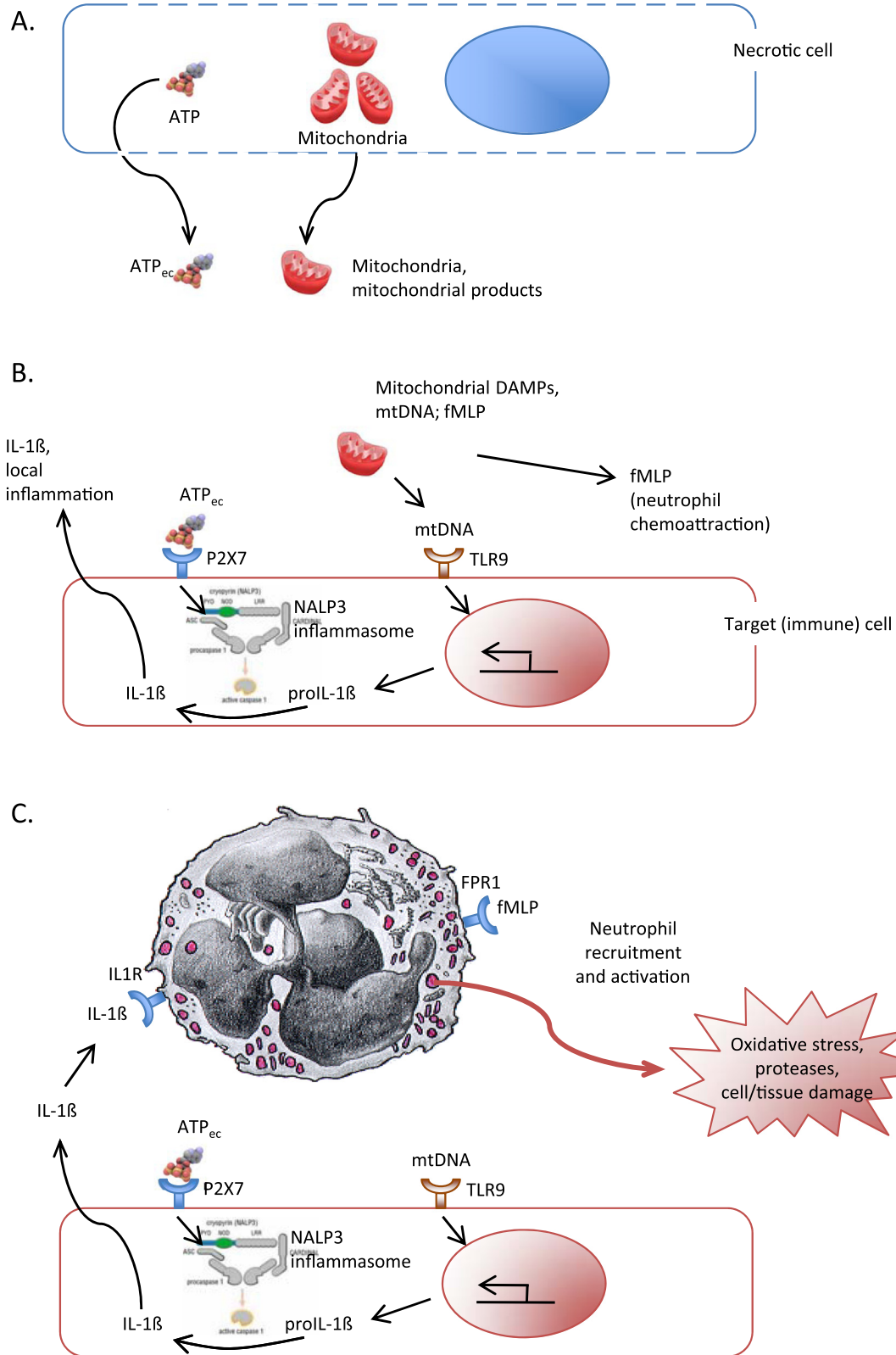


Figure 1 (See legend on next page.)

(See figure on previous page.)

Figure 1 Inflammation induced by alarmins released from injured/necrotic cells. **A.** Injured/necrotic cells with membrane breaks release ATP and mitochondrial alarmins (mitochondrial products). **B.** Extracellular ATP stimulates in target (immune) cells the assembly of the NALP3 inflammasome via its interaction with the P_2X_7 receptor, activating caspase-1. Mitochondrial (mt)DNA stimulates the transcription of the pro-IL-1 β gene via its interaction with TLR9 receptor, and the production of pro-IL-1 β that will be cleaved by activated caspase-1 into bioactive IL-1 β , which in turn can be released into the alveolar space and the interstitium and generate local inflammation. fMLP will create chemotactic gradients to attract neutrophils to the injured tissues. **C.** Neutrophils attracted to the injured tissues will be activated further by fMLP and IL-1 β via their respective receptors (FPR1, and IL-1R), generate oxidative burst and release proteases, which could be further deleterious to tissues (neutrophil-dependent tissue injury).

Extracellular ATP

Tissue injury/necrosis also releases cytoplasmic adenosine triphosphate (ATP), which becomes “extracellular” ATP (ATP_{ec}), and activates neighboring cells via the leukocyte P_2X_7 receptor (Figure 1) [42]. Like uric acid, ATP_{ec} activation of cells leads to caspase-1 activation and maturation of IL-1 β via the assembly of the NALP3 inflammasome and participate in local inflammation [42]. Assembling the NALP3 inflammasome probably does not suffice to create inflammation, because the caspase 1 substrates pro-IL-1 β and pro-IL-18 need to be present to generate inflammation [43]. ATP_{ec} might therefore be seen more as a co-alarmin, enhancing the inflammation induced by mitochondrial alarmins or bacterial products such as lipopolysaccharide responsible for the transcriptional activation of the pro-IL-1 β and pro-IL-18 genes [43].

Conclusions

The discovery of potent endogenous danger signals released from injured/necrotic cells clearly adds to our understanding of the pathogenesis of “sterile inflammation,” as well as SIRS when tissue injury is sufficient to produce systemic inflammation. Alarmins represent nothing less than the proximal endogenous mediators linking cell necrosis and the inflammatory response generated by tissue injury. The identification of alarmins from mitochondria that are ancient bacteria sheltered inside of cells makes a lot of sense. Mitochondria have retained throughout the evolution some molecules of their bacterial ascent that, when found extracellularly, can be recognized as “non-self” by cells from the immune system. Other alarmins, such as ATP, also are strictly intracellular molecules that when found extracellularly will be sensed as a “danger molecule,” meaning for the immune system: “rupture of plasma cell membrane and tissue injury.” Although extracellular ATP does not seem to be capable of mediating a great deal of inflammation, it is an important cofactor, an enhancer of the inflammatory reaction in response to mitochondrial danger molecules and to microbial-associated molecular patterns.

Given their proximal location in the inflammatory cascade during tissue inflammation, alarmins and alarmin-dependent pathways represent attractive targets to develop drugs aimed at dampening deleterious inflammatory

reactions following (massive) tissue injury. Examples of such situations in critical care are: multiple trauma, ARDS, ventilator-induced lung injury, ischemia/reperfusion injury, and severe pancreatitis. Blocking alarmin-dependent pathways will certainly dampen neutrophil-dependent tissue injury. It also will decrease innate immune responses and possibly favor bacterial and yeast superinfections, because it will interfere with the recruitment and the activation of neutrophils. Phagocyte recruitment at the site of tissue injury also plays an important role for the removal of cell debris, the restitution of tissue integrity and healing. As with many modulators of the inflammation, finding the right dose of “anti-alarmins,” the proper route of administration, and the adequate time window will be key to develop successful novel drugs designed to combat SIRS and SIRS-induced organ dysfunction.

Abbreviations

ATP_{ec}: Extracellular adenosine triphosphate; fMLP: Formyl-Met-Leu-Phe; PIRO: Predisposition, infection, response, and organ dysfunction; IL-1RA: IL-1 receptor antagonist; DAMPs: Danger-associated molecular patterns; HMGB1: High mobility group box 1; RAGE: Receptor for advanced glycation end products; TLR: Toll-like receptor; HSPs: Heat shock proteins; LPS: Lipopolysaccharide; MtDNA: Mitochondrial DNA; NALP3: NACHT, LRR and PYD domains-containing protein 3.

Competing interests

The author declares that he has no competing interests.

Acknowledgement

This work was supported by the Swiss National Fund for Scientific Research, grant #320080-122034 and 141143.

Received: 28 March 2012 Accepted: 15 April 2012

Published: 12 July 2012

References

1. Bone RC: Toward an epidemiology and natural history of SIRS (systemic inflammatory response syndrome). *JAMA* 1992, **268**:3452–3455.
2. Bone RC, Sibbald WJ, Sprung CL: The ACCP-SCCM consensus conference on sepsis and organ failure. *Chest* 1992, **101**:1481–1483.
3. Pittet D, Rangel-Frausto S, Li N, et al: Systemic inflammatory response syndrome, sepsis, severe sepsis and septic shock: incidence, morbidities and outcomes in surgical ICU patients. *Intensive Care Med* 1995, **21**:302–309.
4. Rangel-Frausto MS, Pittet D, Costigan M, Hwang T, Davis CS, Wenzel RP: The natural history of the systemic inflammatory response syndrome (SIRS) A prospective study. *JAMA* 1995, **273**:117–123.
5. Dear VJL: SIRS I'm sorry to say that I don't like you. *Crit Care Med* 1997, **25**:372–374.
6. Levy MM, Fink MP, Marshall JC, et al: 2001 SCCM/ESICM/ACCP/ATS/SIS International Sepsis Definitions Conference. *Intensive Care Med* 2003, **29**:530–538.

7. Howell MD, Talmor D, Schuetz P, Hunziker S, Jones AE, Shapiro NI: **Proof of principle: the predisposition, infection, response, organ failure sepsis staging system.** *Crit Care Med* 2011, **39**:322–327.
8. Dinarello CA, Gelfand JA, Wolff SM: **Anticytokine strategies in the treatment of the systemic inflammatory response syndrome.** *JAMA* 1993, **269**:1829–1835.
9. Pugin J, Ricou B, Steinberg KP, Suter PM, Martin TR: **Proinflammatory activity in bronchoalveolar lavage fluids from patients with ARDS, a prominent role for interleukin-1.** *Am J Respir Crit Care Med* 1996, **153**:1850–1856.
10. Dugernier TL, Larterre PF, Wittebole X, et al: **Compartmentalization of the inflammatory response during acute pancreatitis: correlation with local and systemic complications.** *Am J Respir Crit Care Med* 2003, **168**:148–157.
11. Munford RS, Pugin J: **Normal responses to injury prevent systemic inflammation and can be immunosuppressive.** *Am J Respir Crit Care Med* 2001, **163**:316–321.
12. de Jong HK, van der Poll T, Wiersinga WJ: **The systemic pro-inflammatory response in sepsis.** *J Innate Immun* 2010, **2**:422–430.
13. Pugin J: **Dear SIRS, the concept of "alarmins" makes a lot of sense!** *Intensive Care Med* 2008, **34**:218–221.
14. Matzinger P: **Tolerance, danger, and the extended family.** *Annu Rev Immunol* 1994, **12**:991–1045.
15. Matzinger P: **An innate sense of danger.** *Ann N Y Acad Sci* 2002, **961**:341–342.
16. Matzinger P: **The danger model: a renewed sense of self.** *Science* 2002, **296**:301–305.
17. Dreifus C: **A Conversation With Polly Matzinger; Blazing an Unconventional Trail to a New Theory of Immunity.** The New York Times; 1998.
18. Bianchi ME: **DAMPs, PAMPs and alarmins: all we need to know about danger.** *J Leukoc Biol* 2007, **81**:1–5.
19. Adib-Conquy M, Cavaillon JM: **Stress molecules in sepsis and systemic inflammatory response syndrome.** *FEBS Lett* 2007, **581**:3723–3733.
20. Oppenheim JJ, Tewary P, de la Rosa G, Yang D: **Alarmins initiate host defense.** *Adv Exp Med Biol* 2007, **601**:185–194.
21. Iyer SS, Pulsikens WP, Sadler JJ, et al: **Necrotic cells trigger a sterile inflammatory response through the Nlrp3 inflammasome.** *Proc Natl Acad Sci USA* 2009, **106**:20388–20393.
22. Wang H, Bloom O, Zhang M, et al: **HMG-1 as a late mediator of endotoxin lethality in mice.** *Science* 1999, **285**:248–251.
23. Yang R, Harada T, Mollen KP, et al: **Anti-HMGB1 neutralizing antibody ameliorates gut barrier dysfunction and improves survival after hemorrhagic shock.** *Mol Med* 2006, **12**:105–114.
24. Levy RM, Mollen KP, Prince JM, et al: **Systemic Inflammation and Remote Organ Injury Following Trauma Require HMGB1.** *Am J Physiol Regul Integr Comp Physiol* 2007, **293**(4):R1538–44.
25. Sunden-Cullberg J, Norrby-Teglund A, Rouhiainen A, et al: **Persistent elevation of high mobility group box-1 protein (HMGB1) in patients with severe sepsis and septic shock.** *Crit Care Med* 2005, **33**:564–573.
26. Kokkola R, Andersson A, Mullins G, et al: **RAGE is the major receptor for the proinflammatory activity of HMGB1 in rodent macrophages.** *Scand J Immunol* 2005, **61**:1–9.
27. Yu M, Wang H, Ding A, et al: **HMGB1 signals through toll-like receptor (TLR) 4 and TLR2.** *Shock* 2006, **26**:174–179.
28. Rouhiainen A, Tumova S, Valmu L, Kalkkinen N, Rauvala H: **Pivotal advance: analysis of proinflammatory activity of highly purified eukaryotic recombinant HMGB1 (amphoterin).** *J Leukoc Biol* 2007, **81**:49–58.
29. Tsan MF: **Heat shock proteins and high mobility group box 1 protein lack cytokine function.** *J Leukoc Biol* 2011, **89**:847–853.
30. Dumitriu IE, Baruah P, Manfredi AA, Bianchi ME, Rovere-Querini P: **HMGB1: guiding immunity from within.** *Trends Immunol* 2005, **26**:381–387.
31. Harris HE, Raucci A: **Alarmin(g) news about danger: workshop on innate danger signals and HMGB1.** *EMBO Rep* 2006, **7**:774–778.
32. Raucci A, Palumbo R, Bianchi ME: **HMGB1: a signal of necrosis.** *Autoimmunity* 2007, **40**:285–289.
33. Calderwood SK, Mambula SS, Gray PJ Jr: **Extracellular heat shock proteins in cell signaling and immunity.** *Ann N Y Acad Sci* 2007, **1113**:28–39.
34. Foell D, Wittkowski H, Roth J: **Mechanisms of disease: a 'DAMP' view of inflammatory arthritis.** *Nature clinical practice* 2007, **3**:382–390.
35. Foell D, Wittkowski H, Vogl T, Roth J: **S100 proteins expressed in phagocytes: a novel group of damage-associated molecular pattern molecules.** *J Leukoc Biol* 2007, **81**:28–37.
36. Martinon F, Petrilli V, Mayor A, Tardivel A, Tschopp J: **Gout-associated uric acid crystals activate the NALP3 inflammasome.** *Nature* 2006, **440**:237–241.
37. Bonilla WV, Frohlich A, Senn K, et al: **The alarmin interleukin-33 drives protective antiviral CD8 T cell responses.** *Science* 2012, **335**:984–989.
38. Dyall SD, Brown MT, Johnson PJ: **Ancient invasions: from endosymbionts to organelles.** *Science* 2004, **304**:253–257.
39. Krysko DV, Agostinis P, Krysko O, et al: **Emerging role of damage-associated molecular patterns derived from mitochondria in inflammation.** *Trends Immunol* 2011, **32**:157–164.
40. Zhang Q, Raoof M, Chen Y, et al: **Circulating mitochondrial DAMPs cause inflammatory responses to injury.** *Nature* 2010, **464**:104–107.
41. Jaecklin T, Otulakowski G, Kavanagh BP: **Do soluble mediators cause ventilator-induced lung injury and multi-organ failure?** *Intensive Care Med* 2010, **36**:750–757.
42. Mariathasan S, Weiss DS, Newton K, et al: **Cryopyrin activates the inflammasome in response to toxins and ATP.** *Nature* 2006, **440**:228–232.
43. McDonald B, Pittman K, Menezes GB, et al: **Intravascular danger signals guide neutrophils to sites of sterile inflammation.** *Science* 2010, **330**:362–366.

doi:10.1186/2110-5820-2-27

Cite this article as: Pugin: How tissue injury alarms the immune system and causes a systemic inflammatory response syndrome. *Annals of Intensive Care* 2012 **2**:27.

Submit your manuscript to a SpringerOpen® journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com