

CKJ REVIEW

Sodium–glucose cotransporter inhibition in polycystic kidney disease: fact or fiction

Baris Afsar¹, Rengin Elsurer Afsar¹, Atalay Demiray², Sevval Altay², Hakan Korkmaz³, Abdulmecit Yildiz⁴, Adrian Covic⁵, Alberto Ortiz⁶ and Mehmet Kanbay⁷

¹Department of Medicine, Division of Nephrology, Suleyman Demirel University School of Medicine, Isparta, Turkey, ²Department of Medicine, Koc University School of Medicine, Istanbul, Turkey, ³Department of Medicine, Division of Endocrinology, Suleyman Demirel University School of Medicine, Isparta, Turkey, ⁴Department of Medicine, Division of Nephrology, Uludag University School of Medicine, Bursa, Turkey, ⁵Nephrology Clinic, Dialysis and Renal Transplant Center, ‘C.I. PARHON’ University Hospital, and ‘Grigore T. Popa’ University of Medicine, Iasi, Romania, ⁶Department of Medicine, Universidad Autonoma de Madrid and IIS-Fundacion Jimenez Diaz, Madrid, Spain and ⁷Department of Medicine, Division of Nephrology, Koc University School of Medicine, Istanbul, Turkey

Correspondence to: Mehmet Kanbay; E-mail: mkanbay@ku.edu.tr

ABSTRACT

Autosomal dominant polycystic kidney disease (ADPKD) is the most prevalent hereditary kidney disease. Recent evidence suggests that the pathogenesis of ADPKD is a complex web of abnormal cellular processes including altered cell signaling, disordered cell metabolism, impaired autophagy, increased apoptosis, mitochondrial dysfunction and chronic inflammation. Sodium–glucose cotransporter (SGLT) inhibitors (SGLTi) reduce body weight, blood pressure and blood glucose levels, have kidney and cardiovascular protective activity, and have been reported to decrease inflammation, increase autophagy and improve mitochondrial dysfunction. We now review results from preclinical studies on SGLTi for ADPKD identified through a systematic search of the MEDLINE, Cochrane Library, Embase and PubMed databases. Potential underlying mechanisms for the conflicting results reported as well as implications for clinical translation are discussed, as ADPKD patients were excluded from clinical trials exploring kidney protection by SGLT2 inhibitors (SGLT2i). However, they were not excluded from cardiovascular safety trials or trials for cardiovascular conditions. A post-hoc analysis of the kidney function trajectories and safety of SGLT2i in ADPKD patients enrolled in such trials may provide additional information. In conclusion, SGLT2i are cardio- and nephroprotective in diverse clinical situations. Currently, it is unclear whether ADPKD patients may benefit from SGLT2i in terms of kidney function preservation, and their safety in this population remains unexplored. We propose a roadmap to address this unmet clinical need.

Keywords: apoptosis, autophagy, canagliflozin, dapagliflozin, polycystic kidney disease, SGLT inhibitors

Received: 12.10.2021; Editorial decision: 26.1.2022

© The Author(s) 2022. Published by Oxford University Press on behalf of the ERA. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial License (<https://creativecommons.org/licenses/by-nc/4.0/>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

INTRODUCTION

Autosomal dominant polycystic kidney disease (ADPKD) is the most common inherited kidney disease, which is characterized by the formation of numerous fluid-filled cysts primarily within the kidneys, often progressing to end-stage renal disease (ESRD) as renal cysts increase in number and size. Most cases are caused by mutations of either the polycystin-1 (PKD1) (75–85%) or polycystin-2 (PKD2) (15–25%) genes, the former having generally a worse prognosis [1, 2]. Sodium–glucose cotransporter-2 (SGLT2) inhibitors (SGLT2i) were introduced as antidiabetic drugs; however, more recently clinical trials have shown that they have heart and kidney protective effects in person with or without diabetes [3]. A systematic review of the literature was conducted to identify publications that detail the effect of sodium–glucose cotransporter (SGLT) inhibition in the treatment of ADPKD. The search was conducted in the following electronic databases: MEDLINE, Cochrane Library, Embase and PubMed. These electronic databases were searched on (until October 2021) using a structured search string, including the terms ADPKD and SGLT, ADPKD and SGLT_i, PKD and SGLT, and PKD and SGLT_i.

PATHOGENESIS OF ADPKD

The pathogenesis of ADPKD is a complex web of defective cellular processes including altered cellular signaling, increased apoptosis, impaired autophagy, mitochondrial dysfunction, increased aerobic glycolysis associated with hyperglycemia (Warburg Effect) and chronic inflammation [4]. Better understanding of these abnormal cellular processes has highlighted several treatment possibilities; however, few have fulfilled their initial promises and met the theoretical and pathophysiological expectations. The poor translation of preclinical animal studies to humans has several contributors, from animal models that do not reproduce the same molecular defect (PKD1 or PKD2 deficiency) to use of doses that cannot be achieved in human studies due to side effects. Thus, the treatment of ADPKD still remains a challenge revealing the unmet need for new therapeutic options, despite the availability of tolvaptan, a selective, competitive vasopressin receptor 2 (V2R) antagonist [5].

A diverse set of intracellular signaling pathways are dysregulated and contribute to the pathogenesis of ADPKD including the mammalian target of rapamycin (mTOR), AMP-activated protein kinase (AMPK), vasopressin-mediated cyclic adenosine monophosphate (cAMP) and extracellular signal-regulated kinases (ERK) pathways [6–8]. Inflammation, mitochondrial dysfunction and metabolic reprogramming have also been held responsible for the formation and expansion of cysts. Finally, as for any chronic, slowly progressive form of CKD, the reduction in functional kidney mass may result in glomerular hyperfiltration and development of focal segmental glomerular lesions. The pathogenesis of ADPKD was examined and thoroughly explained in reviews and beyond the scope of the article [4].

THERAPEUTIC APPROACHES

Targeting cAMP signaling

Vasopressin binding to V2R induces cyst formation and transepithelial fluid secretion by increasing cAMP, especially in distal tubules [9]. Thus, suppressing vasopressin is a logical approach for reducing cysts within the kidneys. This may be achieved by daily water intake of 3–4 L over 24 h, although the feasibility and efficacy on ADPKD of long-term vasopressin suppres-

sion through increased water intake has not been demonstrated [10]. In addition, tolvaptan slows cyst and total kidney volume (TKV) growth and estimated glomerular filtration rate (eGFR) loss in persons with ADPKD [11–13]. However, tolvaptan is still an expensive medication, not available widely, and has significant side-effects (hepatotoxicity, polyuria) leading to intolerance and drug withdrawal in some patients. Additionally, residual risk for CKD progression remains unacceptably high.

Targeting mTOR signaling

Sirolimus and everolimus inhibit mTOR complex 1 (mTORC1) but not mTOR complex 2 (mTORC2). Although sirolimus and everolimus showed promising results in animal models, these findings unfortunately were not replicated in clinical trials [6, 14]. A new class of drugs, mTOR kinase inhibitors, directly binds to mTOR kinase and inhibits both mTORC1 and mTORC2. It was suggested that combined mTOR1 and mTOR 2 inhibitions may be more effective than solely inhibiting mTOR1 in PKD. PP242, an mTOR kinase inhibitor, reduced renal enlargement and cyst numbers in the Han:SPRD rat model of PKD. Compared with sirolimus, PP242 has a higher anti-proliferative effect and is a more effective mTORC1 inhibitor and less toxic on bone marrow, T and B cells [15, 16].

Targeting AMPK signaling

Metformin activation of AMPK and resulting inhibition of mTORC1 signaling lead to attenuation of PKD in the *Pkd1* mutant mice [17]. However, a phase II clinical trial of metformin for ADPKD identified several limitations such as gastrointestinal side effects and reduced bioavailability following a hepatic first-pass effect [18]. In this regard, the clinically maximum tolerable oral dose of metformin (i.e. 2.0 g/day) may still not be sufficient for AMPK activation in the kidney [19, 20]. Despite this, two ongoing phase 3 clinical trials are comparing metformin with placebo [NCT04939935: Implementation of Metformin therapy to Ease Decline of Kidney Function in Polycystic Kidney Disease (IMPEDE-PKD), expected to be completed by 2026] or tolvaptan [NCT0376460: Metformin vs Tolvaptan for Treatment of Autosomal Dominant Polycystic Kidney Disease (METROPOLIS) expected to be completed by 2022].

Targeting fat metabolism

Hepatocyte nuclear factor 4a (Hnf4a) and the peroxisome proliferator-activated receptor- α (PPAR α) are two major proteins responsible for decreased Fatty Acid Oxidation (FAO) in PKD [21]. Loss of Hnf4a function in PKD results in more severe cystic disease [21], while fenofibrate, a PPAR α agonist, enhanced FAO and reduced cysts in an orthologous ADPKD model [22]. However, the clinical evaluation of this drug is limited by its potential to increase serum creatinine [23]. The mechanism is unclear and interference with creatinine secretion has been suggested. In any case, the increase in serum creatinine is rapidly reversible upon stopping fenofibrate and in fact, in a post-hoc analysis of a clinical trial, fenofibrate reduced albuminuria and slowed eGFR loss over 5 years in persons with diabetes [24].

Targeting energy metabolism

In ADPKD, impaired FAO is related to decreased mitochondrial Tricarboxylic Acid (TCA) cycle efficiency [25], rendering PKD cells mostly dependent on glucose and aerobic glycolysis instead of mitochondrial oxidative phosphorylation, known as the ‘Warburg effect’ [26, 27]. This shift generates 4 ATP and 2 lactate

molecules instead of 36 ATP molecules, and lactate is used to build macromolecules in highly proliferative cells such as cells lining cyst tubules [28]. Also, local hypoxia with mitochondrial dysfunction including mitochondrial fragmentation, swelling and reduction in mitochondrial DNA copy number were frequently observed in PKD [29–32]. Transcription and activity of glycolytic enzymes were increased and those of gluconeogenic enzymes decreased in kidneys of Han:SPRD Cy/+ rats. Administration of 2-deoxyglucose (2DG), a glycolytic pathway inhibitor, decreased intrarenal lactate levels, ERK1/2 phosphorylation and cell proliferation, retarded cyst progression and attenuated renal functional decline in cystic rats [33, 34]. Thus, mitochondrial dysfunction with altered metabolic arrangements such as increased glycolysis and decreased oxidative phosphorylation appeared as the main characteristic features of PKD. However, therapeutic options are highly limited for adjustment of this altered metabolic state and mitochondrial dysfunction. SGLT2i seem like a promising agent for targeting energy metabolism and mitochondrial dysfunction.

Besides pharmacological therapies, some dietary strategies were also considered to slow down the progression of PKD such as daily energy restriction, time restricted feeding, intermittent fasting and ketogenic diets [35, 36]. Time-restricted feeding, compared with isocaloric ad libitum feeding, reduced blood glucose and increased ketogenesis, which inhibited renal cyst proliferation and growth. These findings are not merely due to time-restricted feeding since ad libitum administration of a ketogenic diet similarly inhibited renal cyst growth. In addition, acute fasting in rat, mouse and feline models of PKD induced significant apoptosis in cyst-lining epithelial cells but not in normal tissue, resulting in decreased renal cystic burden. These beneficial findings were all replicated with a ketogenic diet based on β -hydroxybutyrate; therefore, the metabolic state of ketosis seemed to be the crucial mediator of beneficial effects by inhibiting cyst growth and fibrosis via altering PKD-related signaling pathways like mTOR and STAT3 [27]. A recent retrospective case series also showed the feasibility of ketogenic diet for patients with ADPKD, concluding significant weight loss, improved blood pressure management and slight improvements in eGFR, with some safety concerns such as hyperlipidemia [37]. Overall, studies suggested that dietary restriction was beneficial for PKD progression by inducing ketosis, as renal cyst cells in PKD seem to be metabolically inflexible and thus unable to adapt to alternative fuel sources apart from glucose [27]. Moreover, several studies showed that ketogenic status is associated not only with better energy metabolism, but also with improved mitochondrial function and autophagy, which eventually contributed to controlling of inflammation and fibrosis [38].

These findings are supported by several studies that showed obesity and hyperglycemia were also correlated with faster disease progression in ADPKD [39–41], as well as functional and structural kidney damage in PKD [42]. Interestingly, one of the putative mechanisms of action of SGLT2i to promote heart and kidney protection is by increasing ketones such as β -hydroxybutyrate, which may also regulate histone post-translational modifications to regulate gene expression [43]. Thus, beneficial effects of SGLT2i seem closely linked to its efficacy on metabolism similar to the metabolic state of ketosis and β -hydroxybutyrate.

Targeting autophagy

Autophagy and autophagosome fusion with lysosomes (i.e. autophagic flux) are impaired in PKD [26, 44–46]. In a Pkd1 mutant zebrafish model, knockdown of autophagy protein Atg5 pro-

moted cystogenesis, whereas treatment with the autophagy inducer Beclin-1 reduced cyst formation and growth [45]. However, a natural autophagy enhancer trehalose was ineffective in a Pkd1 mutant model [44]. Of note, mTORC-1 inhibits autophagy, and various rapalogs enhance autophagy and reduce cystogenesis in PKD [26, 45]. Although the drug arsenal for targeting autophagy was still under investigation, dietary interventions including a ketogenic diet were shown to improve parameters related to autophagy and improved inflammatory state with less fibrosis [36].

Targeting hypoxia-inducible factor

In ADPKD, cyst growth may compromise tissue perfusion and activate hypoxia-inducible factor (HIF), as HIF-1 α and HIF-2 α levels are associated with cyst burden [47]. Although, HIF-1 α is thought to play an active role in the process of cyst expansion and pericyclic angiogenesis, the HIF-1 inhibitor 2-methoxyestradiol (2ME2) had no significant effect on kidney volume or cyst volume density [46] and the impact of HIF stabilizers in clinical use or clinical trials for uremic anemia on ADPKD progression has not been adequately explored [48]. Even though they seemed to provide crucial benefits for PKD and anemia management of CKD patients, HIF-prolyl hydroxylase inhibitors are not currently favored due to a lack of data on their potential effects on cyst development or growth [47].

In addition to the aforementioned metabolic effects of SGLT2i, they have been proposed to play a beta-blocker-like effect in the kidneys, decreasing energy demand by proximal tubule cells, and may be expected to protect proximal tubule cells from a hypoxic environment [3]. Therefore, SGLT2i decrease renal hypoxia, enhance nutrient deprivation signaling, suppress HIF-1 α and activate HIF-2 α , which promotes erythrocytosis [49]. In a recent review of Patel and Dahl [47], effects of SGLT2i and HIF-prolyl hydroxylase inhibitors on ADPKD were compared in detail, suggesting that SGLT2i might provide a better alternative to tolvaptan rather than HIF-prolyl hydroxylase inhibitors for patients with ADPKD.

Targeting glomerular hyperfiltration

In ADPKD, higher baseline albuminuria was associated with faster eGFR loss [50]. Indeed, tolvaptan decreased albuminuria compared with placebo, independent of blood pressure. Treatment efficacy of tolvaptan on changes in TKV and eGFR was more readily detected in patients with higher albuminuria. Together with the observation that tolvaptan causes an early, reversible dip in eGFR, followed by a slower eGFR slope [51], similar to renin-angiotensin system blockers and SGLT2i [52], this observation supports a role of glomerular hyperfiltration of remaining nephrons in the progression of ADPKD. Indeed, in long-term studies, tolvaptan slowed eGFR decline, regardless of its impact on TKV, which was initially considered to be its mechanism of action [53]. This knowledge further supports the potential benefit of SGLT2i in ADPKD.

SGLT INHIBITION

SGLTs are a family of proteins mediating the transepithelial transport of glucose in the proximal tubule of nephrons and intestinal mucosa of small intestines. SGLT2 is a high-capacity, low-affinity transporter, which mediates the remaining 90% of glucose reabsorption in the proximal renal tubule [54, 55]. SGLT inhibition decreases Na⁺-glucose reabsorption in the proximal tubule, thus decreasing energy needs, induces osmotic diuresis and increases distal tubule fluid rate. This leads to an increase

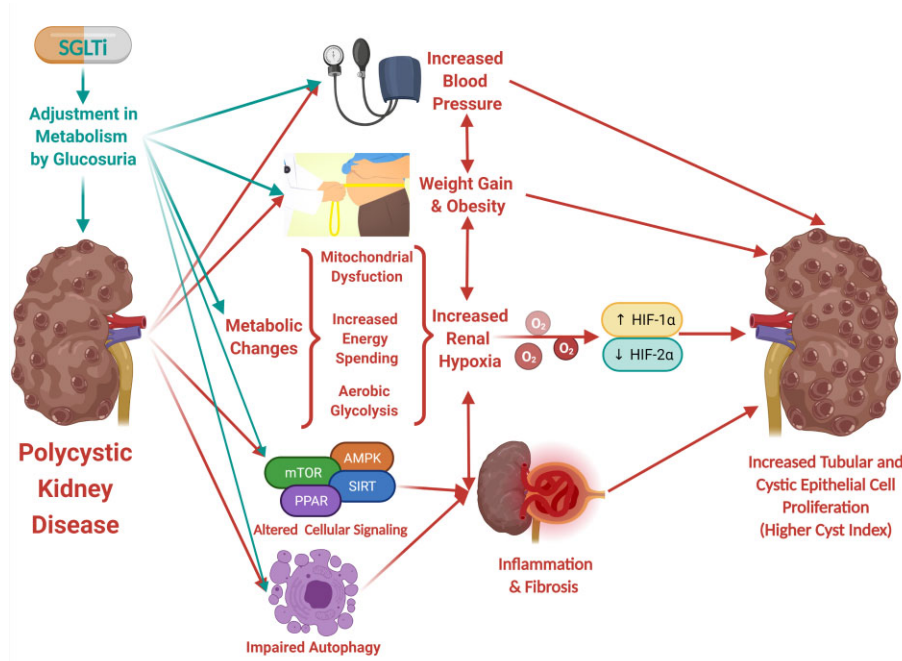


FIGURE 1: The suggested mechanisms of polycystic kidney disease (PKD) leading to higher tubular and cystic epithelial cell proliferation with higher cyst index are shown with red arrows. The possible beneficial effects of SGLT2 on polycystic kidney disease (PKD) are shown with green arrows as they can interfere in multiple pathological processes of PKD by adjustments in metabolism via their glucosuric effects. SGLT2, sodium-glucose co-transporter inhibitor; HIF, hypoxia-inducible factor.

in afferent arteriolar tone because of tubuloglomerular feedback and thus corrects intra-glomerular hypertension and decreases hyperfiltration and its consequences such as albuminuria and metabolic overload of proximal tubules. Furthermore, SGLT2i have many other benefits such as reduction in HbA1c, blood pressure and weight reduction thanks to their glucosuric effect, which helps to get rid of excess glucose from body. Therefore, SGLT2 inhibition could be regarded as an antagonist of excessive carbohydrate consumption for the metabolism. Also, several studies have shown that SGLT2 inhibition has cardioprotective and nephroprotective effects in both diabetic and nondiabetic patients. Mechanisms contributing these benefits are still under investigation; however, switching metabolism to a ketogenic state (e.g. increased β -hydroxybutyrate) by getting rid of excess glucose appears as the main mechanism. Importantly, an anti-inflammatory impact has been described for SGLT2i, as well as increased autophagy and improvement of mitochondrial dysfunction [38, 56]. Since PKD is associated with altered glucose metabolism, impaired autophagy, increased apoptosis and mitochondrial dysfunction, SGLT inhibition appears to be a very promising approach for the treatment (Figure 1). As the main beneficial mechanism of SGLT inhibition has similarity to a reduced carbohydrate diet, whether diet with reduced carbohydrate intake can achieve the same benefits also needs to be elucidated. While the impact of SGLT inhibition on PKD pathogenesis has been investigated over the last few years, studies are limited to preclinical models and results are conflicting. Below, we critically discuss the available and still needed studies.

SGLT INHIBITION IN PRECLINICAL PKD

SGLT inhibition has been tested in rat and murine models of PKD, but only the murine model was characterized by *Pkd1* deficiency

as human ADPKD. Thus, the rat models may be suboptimal to extrapolate results to human ADPKD.

Rat PKD

Wang *et al.* studied the effect of phlorizin, an inhibitor of both SGLT1 and SGLT2, in PCKD Han:SPRD Cy rats over a 5-week period. PCKD Han:SPRD cy rats have a mutation in *Anks6* (also called *Pkdr1*) which encodes SamCystin, a protein expressed in proximal tubules and glomeruli [57-58]. The mutation results in increased levels and mislocalization of SamCystin. These rats develop cysts exclusively in proximal tubules that become disconnected from these tubules. Phlorizin induced glycosuria and osmotic diuresis, increased creatinine clearance and decreased albuminuria. The kidney weight and cyst index were also lower in phlorizin-treated rats compared with placebo-treated rats. Moreover, phlorizin reduces cystic epithelial cell proliferation, assessed by Ki67 staining. In addition, phlorizin decreased ERK phosphorylation, which is increased in PKD, in a dose-dependent manner [58]. In another study with the same rat model, dapagliflozin for 5 weeks caused glycosuria and polyuria, increased creatinine and blood urea nitrogen (BUN) clearances, and decreased albuminuria despite failing to slow cyst growth and increasing kidney weight. Tubular epithelial cell proliferation, macrophage infiltration and interstitial fibrosis were similar in dapagliflozin- and vehicle-treated groups [54]. Kidney enlargement was attributed to widening of tubular lumen due to increased diuresis [54] and to tubular hypertrophy, thought to result from increased SGLT1-mediated glucose reabsorption in the S2 and S3 segments of proximal tubules [59]. Thus, in PCKD Han:SPRD rats with proximal tubular cysts, a dual SGLT1/SGLT2 inhibitor increased creatinine clearance, decreased kidney weight and cyst index while SGLT2i also preserved kidney function

deposit lack of an effect on cysts. Overall, the results support a protective effect of SGLT2 inhibition through mechanisms independent of cyst growth, likely dependent on interference with nonspecific mechanism of CKD progression. However, PCKD Han:SPRD Cy rats are not an optimal model of ADPKD as they involved a different protein target.

In PCK rats, dapagliflozin 10 mg/kg/day or vehicle was administered by gavage to 6-week-old male rats ($n = 9$ per group). PCK rats have a *Pkhd1* genetic defect and are thus a model of autosomal recessive PKD [60]. They develop cysts in distal nephrons that remain connected to the tubule [61, 62]. Dapagliflozin increased glucosuria, urine output and creatinine clearance after 3 weeks; however, albuminuria was also increased 4-fold, suggesting that dapagliflozin induced hyperfiltration. Histological analysis and ultrasound showed a higher cyst volume and a 23% higher total kidney weight in rats treated with dapagliflozin. Renal cAMP content and Ki67 staining were similar between dapagliflozin- and vehicle-treated PCK rats [63]. Contrary to PCK rats, normal rats treated with dapagliflozin do not develop evidence of hyperfiltration and albuminuria [64, 65], so the findings in PCK rats were surprising [63]. Since PCK rats are a model of autosomal recessive PKD, the results are not directly relevant for ADPKD.

Murine PKD

Leonhard et al. [1] investigated the effect of salsalate (a non-steroidal anti-inflammatory drug), metformin or canagliflozin on kidney cyst growth in an adult-onset mouse model of PKD caused by *Pkd1* deficiency. Salsalate or metformin plus salsalate increased kidney survival and reduced cystic kidney disease severity compared with untreated mutant mice. However, metformin did not add further protection to that afforded by salsalate alone and neither metformin nor canagliflozin alone was effective [1]. However, canagliflozin-treated mice were not studied in detail. Just kidney survival data were provided. Although at the end of the study kidney survival was similar to control mice, there was a lag time of 14 days (18%) between the development of kidney failure for the first control mouse and for the first canagliflozin mouse. More detailed studies are warranted. In this regard, this was an accelerated model of ADPKD in which both *Pkd1* alleles were inactivated, postnatally resulting in kidney failure at around Day 100 of disease, i.e. in very young mice, unlike human ADPKD for which only one *PKD1* allele is mutated leading to kidney failure at around age 65 years.

Understanding the results obtained in preclinical PKD

None of the preclinical studies was performed in an optimal model of human ADPKD. Overall, SGLT inhibition was protective in a rat model of proximal tubular cystogenesis but not in a rat model of human autosomal recessive PKD or in a murine model of *Pkd1* deficiency with a time course closer to the natural history of human autosomal recessive PKD than to the natural history of ADPKD. In any case, the authors of these manuscripts speculated for several potential explanations for the lack of efficacy. These include that osmotic diuresis induced by glycosuria may promote the dilation of distal tubules and linked cysts [63, 66] (Figure 1). However, human cysts are independent from tubules, and tolvaptan induced large diuresis volumes while being protective. Thus, this putative mechanism is not clinically relevant.

It has also been suggested that SGLT2 inhibition may increase vasopressin levels. In type 2 diabetic Goto-Kakizaki (GK) rats, after 8 weeks, ipragliflozin decreased body weight, serum glucose

and systolic blood pressure, and increased fluid and food intake, urinary glucose and Na^+ excretion, urine volume and renal osmolar clearance, and most importantly urine vasopressin levels and solute-free water reabsorption (TcH₂O). Urine vasopressin in ipragliflozin-treated rats was negatively and positively associated with fluid balance and TcH₂O, respectively. Ipragliflozin increased expression of SGLT2, aquaporin 2 phosphorylated at Ser269, which is an important clue for strong vasopressin activity, and vasopressin V2 receptor. Thus, the osmotic diuresis induced by SGLT2 inhibition stimulated compensatory fluid intake and renal water reabsorption by increasing vasopressin levels to maintain body fluid volume [67]. Ho et al. [68] suggested that phlorizin could lead to increased endogenous vasopressin levels promoting distal tubular cyst growth, an effect that might not be seen in Han:SPRD rats as the cysts are almost exclusively of proximal tubular origin and unresponsive to vasopressin. Indeed, in type 1 diabetics, levels of copeptin, a more stable marker of vasopressin production, increased in response to empagliflozin under euglycemia and more so under hyperglycemia [69]. Under euglycemia, which would be the clinical conditions for ADPKD patients, the increase was modest (around 24%, to levels of 5.1 ± 2.8 pmol/L) and the clinical implications of this modest increase are unclear. However, in a further diabetic rat model, empagliflozin increased the expression in V2R but decreased protein and mRNA levels of AQP2, and this was associated with increased phosphorylation of AQP2 at S261, a marker of intracellular location, suggesting some blockade of V2R signaling [70]. Overall, the impact of SGLT2 inhibition on vasopressin responses and signaling under nondiabetic chronic conditions is not well understood and merits further clinical studies.

Finally, the impact of SGLT inhibition on PKD may depend on the specificity of SGLT inhibition. Cyst growth, renal function and albuminuria improved in Han:SPRD rats treated with phlorizin, which inhibits both SGLT1 and SGLT2 [58]. Thus, the result may be different with combined SGLT1 and SGLT2 inhibition as compared with SGLT2 inhibition alone, since combined complete SGLT inhibition is more powerful in terms of glycosuria [59]. In this regard, phlorizin is not in clinical use. However, diverse SGLT2i have different relative selectivity for SGLT2 and SGLT1, ranging from 20-fold (sotagliflozin) to 250-fold (canagliflozin) to over 1000-fold (in increasing selectivity order: dapagliflozin, ertugliflozin and empagliflozin, the latest with a selectivity of 2500-fold) [71].

SGLT2 INHIBITION IN CLINICAL ADPKD

Published clinical experience with SGLT2i and ADPKD remains limited or non-existent. Indeed, no publications were found in a PubMed search that also included the main SGLT2i (canagliflozin, dapagliflozin, empagliflozin, ertugliflozin and sotagliflozin) on 9 October 2021. Unfortunately, patients with ADPKD were excluded from clinical trials assessing kidney protection as primary outcome in patients with or without diabetic CKD, such as A Study to Evaluate the Effect of Dapagliflozin on Renal Outcomes and Cardiovascular Mortality in Patients With Chronic Kidney Disease (Dapa-CKD, NCT03036150), which recently demonstrated kidney protection conferred by SGLT2i in nondiabetic patients with CKD [72]. Furthermore, they were also excluded from EMPA-KIDNEY (The Study of Heart and Kidney Protection with Empagliflozin, NCT03594110) a further trial assessing kidney protection by SGLT2 inhibition in nondiabetic patients with CKD expected to be completed by 21 December 2022 [73]. Indeed, none of the ongoing 45 clinical

trials on PKD listed in ClinicalTrials.gov as of 4 October 2021 involves SGLT2 inhibition [74]. Thus, the clinical trial experience is and will remain limited for the foreseeable future. However, clinical trial NCT04680780 is exploring the feasibility of ketogenic dietary interventions and has a secondary endpoint of TKV. Promotion of ketogenesis is one of the putative kidney and cardioprotective mechanisms of action of SGLT2 inhibition [75]. However, ADPKD patients were not excluded from clinical trial assessing cardiovascular safety in persons with type 2 diabetes for assessing efficacy in cardiovascular conditions [76, 77]. Thus, a careful, targeted post-hoc analysis of several of these trials combined in search of persons with ADPKD that may have been enrolled because of coexisting diabetes or cardiovascular disease may provide further clues as to the safety and potential efficacy for kidney protection of SGLT2i in ADPKD.

SAFETY

Regarding safety, a main concern would be the risk of genitourinary infection, as an ascending infection affecting kidney cysts may have potentially severe consequences [78]. As ADPKD patients are more prone to urinary tract and cyst infections than the general population, the detrimental effects of possible infections including faster disease progression in ADPKD patients should be evaluated carefully in future studies. In any case, this information would only be hypothesis generating and would be useful to design further clinical studies as its post-hoc nature, presence of comorbidities and low patient numbers will prevent reaching definitive conclusions. In this regard, another

Table 1. Proposed roadmap to establish the safety and efficacy of SGLT2i for kidney and cardiovascular disease in persons with ADPKD

(A) Preclinical
1. Explore the efficacy and safety of SGLT2i in preclinical models of ADPKD that more closely resemble the kidney conditions: i.e. defective <i>Pkd1</i> gene and long natural history/and long-term SGLT2i treatment, leading to severe kidney disease well into adulthood.
2. Detailed characterization of the impact of SGLT2i on vasopressin and vasopressin signaling under nondiabetic conditions.
(B) Clinical. These sequential steps are proposed:
1. Create an SGLT2i-ADPKD task force.
2. Contact the leadership of all phase 3, large scale trials of cardiovascular safety or cardiovascular outcomes to extract, pool and analyze the safety and outcomes of persons with ADPKD that may have been enrolled in these trials.
3. Create a registry of ADPKD patients treated with SGLT2i for diabetes or cardiovascular conditions, assessing safety (urinary tract infections, copeptin, eGFR trajectories, TKV and albuminuria) as well as kidney efficacy (eGFR trajectories, TKV and albuminuria) outcomes.
4. Design short-term exploratory trials addressing the acute and short-term impact of SGLT2i on vasopressin, vasopressin signaling, eGFR and TKV, as well as safety.
5. Based on the result for steps 2 through 4, decide on the need, feasibility and design of larger, long-term phase 2 and phase 3 trials with endpoint efficacy on kidney protection or add warnings to drug labels regarding potential risks in ADPKD patients.

drug with antidiabetic properties, the thiazolidinedione pioglitazone, was recently found to be safe in a phase 1b clinical trial for ADPKD, following reports of kidney protection effects in rodent PKD [79].

Another safety issue for using SGLT2i in ADPKD is their potential to exacerbation of hypovolemia, hypernatremia and acute kidney injury when combining with tolvaptan, which could prevent vasopressin-mediated water reabsorption [47]. Volume status of ADPKD patients under SGLT2i and tolvaptan treatment should be assessed carefully and their effects should be further explored in future studies.

Better understanding of the nature of ADPKD with its pathological processes is crucial for therapeutic selections in the future as some non-pharmacological dietary interventions like ketogenic diet, time-restricted diet, etc. could also provide beneficial effects to patients with ADPKD targeting altered metabolic state. Whether the usage of SGLT2i added more benefit to patients with ADPKD than other dietary interventions such as ketogenic diet or their beneficial effects are solely dependent on their glucosuria, and metabolic effects need to be elucidated in the future. Future studies and randomized controlled trials can be conducted to compare the feasibility, efficacy and the safety profile of SGLT2i and dietary intervention with a ketogenic diet based on β -hydroxybutyrate.

CONCLUSIONS

In conclusion, SGLT2i are currently established heart and kidney protective agents under diabetic and nondiabetic conditions. Thus, ADPKD patients might theoretically benefit from them in terms of kidney and/or heart protection and they may even be prescribed to the patients for cardiovascular conditions. However, clinical experience with SGLT2i is virtually non-existent, as ADPKD patients were excluded from trials with kidney outcomes and whether they were enrolled in trials of cardiovascular outcomes is unclear. If ADPKD patients were enrolled in cardiovascular trials of SGLT2i, their outcomes have not been analyzed specifically. Preclinical studies had conflicting results because they were marred by the use of suboptimal animal models for a complicated human disease. In any case, there is basis for therapeutic concerns regarding the risk of infection and of increased vasopressin levels. Thus, a roadmap should be established aimed at establishing the safety of SGLT2i in persons with ADPKD, as they may need SGLT2i prescription for cardiovascular conditions, and to explore the eventual kidney benefit of this intervention (Table 1).

ACKNOWLEDGEMENTS

M.K. gratefully acknowledges the use of the services and facilities of the Koc University Research Center for Translational Medicine (KUTTAM), funded by the Presidency of Turkey, Presidency of Strategy and Budget. The content is solely the responsibility of the authors and does not necessarily represent the official views of the Presidency of Strategy and Budget.

FUNDING

This study was not funded by any grant.

AUTHORS' CONTRIBUTIONS

Contributed substantially to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work: B.A., R.E.A., S.A., A.D., H.K., A.Y. Drafted the work or revised it critically for important intellectual content: B.A., R.E.A., A.O., A.C. and M.K.

CONFLICT OF INTEREST STATEMENT

A.O. has received consultancy or speaker fees or travel support from Astellas, AstraZeneca, Amicus, Amgen, Fresenius Medical Care, Bayer, Sanofi-Genzyme, Menarini, Kyowa Kirin, Alexion, Idorsia, Chiesi, Otsuka and Vifor Fresenius Medical Care Renal Pharma, and is Director of the Catedra Mundipharma-UAM of diabetic kidney disease and the Catedra Astrazeneca-UAM of chronic kidney disease and electrolytes. A.O. is the Editor-in-Chief of CKJ and M.K. is member of the CKJ editorial board. The other authors declare that they have no conflict of interest.

ETHICAL APPROVAL

This article does not contain any studies with human participants or animals performed by any of the authors.

REFERENCES

- Leonhard WN, Song X, Kanhai AA et al. Salsalate, but not metformin or canagliflozin, slows kidney cyst growth in an adult-onset mouse model of polycystic kidney disease. *EBioMedicine* 2019; 47: 436–445
- Grantham JJ, Mulamalla S, Swenson-Fields KI. Why kidneys fail in autosomal dominant polycystic kidney disease. *Nat Rev Nephrol* 2011; 7: 556–566
- Fernandez-Fernandez B, Sarafidis P, Kanbay M et al. SGLT2 inhibitors for non-diabetic kidney disease: drugs to treat CKD that also improve glycaemia. *Clin Kidney J* 2020; 13: 728–733
- Bergmann C, Guay-Woodford LM, Harris PC et al. Polycystic kidney disease. *Nat Rev Dis Primers* 2018; 4: 50
- Gansevoort RT, Arici M, Benzings T et al. Recommendations for the use of tolvaptan in autosomal dominant polycystic kidney disease: a position statement on behalf of the ERA-EDTA Working Groups on Inherited Kidney Disorders and European Renal Best Practice. *Nephrol Dial Transplant* 2016; 31: 337–348
- Serra AL, Poster D, Kistler AD et al. Sirolimus and kidney growth in autosomal dominant polycystic kidney disease. *N Engl J Med* 2010; 363: 820–829
- Devuyst O, Torres VE. Osmoregulation, vasopressin, and cAMP signaling in autosomal dominant polycystic kidney disease. *Curr Opin Nephrol Hypertens* 2013; 22: 459–470
- Torres VE, Harris PC. Strategies targeting cAMP signaling in the treatment of polycystic kidney disease. *J Am Soc Nephrol* 2014; 25: 18–32
- Belibi FA, Reif G, Wallace DP et al. Cyclic AMP promotes growth and secretion in human polycystic kidney epithelial cells. *Kidney Int* 2004; 66: 964–973
- Wang CJ, Grantham JJ, Wetmore JB. The medicinal use of water in renal disease. *Kidney Int* 2013; 84: 45–53
- Gattone VH II, Wang X, Harris PC et al. Inhibition of renal cystic disease development and progression by a vasopressin V2 receptor antagonist. *Nat Med* 2003; 9: 1323–1326
- Torres VE, Wang X, Qian Q et al. Effective treatment of an orthologous model of autosomal dominant polycystic kidney disease. *Nat Med* 2004; 10: 363–364
- Torres VE, Chapman AB, Devuyst O et al. Tolvaptan in later-stage autosomal dominant polycystic kidney disease. *N Engl J Med* 2017; 377: 1930–1942
- Walz G, Budde K, Mannaa M et al. Everolimus in patients with autosomal dominant polycystic kidney disease. *N Engl J Med* 2010; 363: 830–840
- Ravichandran K, Zafar I, Ozkok A et al. An mTOR kinase inhibitor slows disease progression in a rat model of polycystic kidney disease. *Nephrol Dial Transplant* 2015; 30: 45–53
- Belibi F, Ravichandran K, Zafar I et al. mTORC1/2 and rapamycin in female Han:SPRD rats with polycystic kidney disease. *Am J Physiol Renal Physiol* 2011; 300: F236–F244
- Takiar V, Nishio S, Seo-Mayer P et al. Activating AMP-activated protein kinase (AMPK) slows renal cystogenesis. *Proc Natl Acad Sci USA* 2011; 108: 2462–2467
- Seliger SL, Abebe KZ, Hallows KR et al. A randomized clinical trial of metformin to treat autosomal dominant polycystic kidney disease. *Am J Nephrol* 2018; 47: 352–360
- He L, Wondisford FE. Metformin action: concentrations matter. *Cell Metab* 2015; 21: 159–162
- Chandel NS, Avizonis D, Reczek CR et al. Are metformin doses used in murine cancer models clinically relevant? *Cell Metab* 2016; 23: 569–570
- Menezes LF, Zhou F, Patterson AD et al. Network analysis of a Pkd1-mouse model of autosomal dominant polycystic kidney disease identifies HNF4 α as a disease modifier. *PLoS Genet* 2012; 8: e1003053
- Lakhia R, Yheskel M, Flaten A et al. PPAR α agonist fenofibrate enhances fatty acid β -oxidation and attenuates polycystic kidney and liver disease in mice. *Am J Physiol Renal Physiol* 2018; 314: F122–F131
- Attridge RL, Frei CR, Ryan L et al. Fenofibrate-associated nephrotoxicity: a review of current evidence. *Am J Health Syst Pharm* 2013; 70: 1219–1225
- Davis TM, Ting R, Best JD et al. Effects of fenofibrate on renal function in patients with type 2 diabetes mellitus: the Fenofibrate Intervention and Event Lowering in Diabetes (FIELD) Study. *Diabetologia* 2011; 54: 280–290
- Ishimoto Y, Inagi R, Yoshihara D et al. Mitochondrial abnormality facilitates cyst formation in autosomal dominant polycystic kidney disease. *Mol Cell Biol* 2017; 37: e00337–17
- Rowe I, Chiaravalli M, Mannella V et al. Defective glucose metabolism in polycystic kidney disease identifies a new therapeutic strategy. *Nat Med* 2013; 19: 488–493
- Torres JA, Kruger SL, Broderick C et al. Ketosis ameliorates renal cyst growth in polycystic kidney disease. *Cell Metab* 2019; 30: 1007–1023
- Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science* 2009; 324: 1029–1033
- Che R, Yuan Y, Huang S et al. Mitochondrial dysfunction in the pathophysiology of renal diseases. *Am J Physiol Renal Physiol* 2014; 306: F367–F378
- Li QW, Lu XY, You Y et al. Comparative proteomic analysis suggests that mitochondria are involved in autosomal recessive polycystic kidney disease. *Proteomics* 2012; 12: 2556–2570
- Bernhardt WM, Wiesener MS, Weidemann A et al. Involvement of hypoxia-inducible transcription factors in polycystic kidney disease. *Am J Pathol* 2007; 170: 830–842

32. Buchholz B, Schley G, Faria D et al. Hypoxia-inducible factor-1 α causes renal cyst expansion through calcium-activated chloride secretion. *J Am Soc Nephrol* 2014; 25: 465–474
33. Riwanto M, Kapoor S, Rodriguez D et al. Inhibition of aerobic glycolysis attenuates disease progression in polycystic kidney disease. *PLoS One* 2016; 11: e0146654
34. Chiaravalli M, Rowe I, Mannella V et al. 2-deoxy-D-glucose ameliorates PKD progression. *J Am Soc Nephrol* 2016; 27: 1958–1969
35. Carriazo S, Perez-Gomez MV, Cordido A et al. Dietary care for ADPKD patients: current status and future directions. *Nutrients* 2019; 11: 1576
36. Nowak KL, Hopp K. Metabolic reprogramming in autosomal dominant polycystic kidney disease: evidence and therapeutic potential. *Clin J Am Soc Nephrol* 2020; 15: 577–584
37. Sebastian Strubl SO, Jacob AT, Grundmann F et al. Ketogenic dietary interventions in autosomal dominant polycystic kidney disease—a retrospective case series study: first insights into feasibility, safety and effects. *Clin Kidney J* 2021; sfab162
38. Afsar B, Hornum M, Afsar RE et al. Mitochondrion-driven nephroprotective mechanisms of novel glucose lowering medications. *Mitochondrion* 2021; 58: 72–82
39. Reed B, Helal I, McFann K et al. The impact of type II diabetes mellitus in patients with autosomal dominant polycystic kidney disease. *Nephrol Dial Transplant* 2012; 27: 2862–2865
40. Nowak KL, You Z, Gitomer B et al. Overweight and obesity are predictors of progression in early autosomal dominant polycystic kidney disease. *J Am Soc Nephrol* 2018; 29: 571–578
41. Kraus A, Schley G, Kunzelmann K et al. Glucose promotes secretion-dependent renal cyst growth. *J Mol Med (Berl)* 2016; 94: 107–117
42. Sas KM, Yin H, Fitzgibbon WR et al. Hyperglycemia in the absence of cilia accelerates cystogenesis and induces renal damage. *Am J Physiol Renal Physiol* 2015; 309: F79–F87
43. Martinez-Moreno JM, Fontecha-Barriuso M, Martin-Sanchez D et al. Epigenetic modifiers as potential therapeutic targets in diabetic kidney disease. *Int J Mol Sci* 2020; 21: 4113
44. Chou LF, Cheng YL, Hsieh CY et al. Effect of trehalose supplementation on autophagy and cystogenesis in a mouse model of polycystic kidney disease. *Nutrients* 2018; 11: 42
45. Zhu P, Sieben CJ, Xu X et al. Autophagy activators suppress cystogenesis in an autosomal dominant polycystic kidney disease model. *Hum Mol Genet* 2017; 26: 158–172
46. Belibi F, Zafar I, Ravichandran K et al. Hypoxia-inducible factor-1 α (HIF-1 α) and autophagy in polycystic kidney disease (PKD). *Am J Physiol Renal Physiol* 2011; 300: F1235–F1243
47. Patel DM, Dahl NK. Examining the role of novel CKD therapies for the ADPKD patient. *Kidney* 2021; 360: 1036–1041
48. Liu F, Wang J, Ye Q et al. Roxadustat for renal anemia in ESRD from PKD patients: is it safe enough? *J Am Soc Nephrol* 2021; 32: 1005
49. Packer M. Mechanisms leading to differential hypoxia-inducible factor signaling in the diabetic kidney: modulation by SGLT2 inhibitors and hypoxia mimetics. *Am J Kidney Dis* 2021; 77: 280–286
50. Gansevoort RT, Meijer E, Chapman AB et al. Albuminuria and tolvaptan in autosomal-dominant polycystic kidney disease: results of the TEMPO 3:4 Trial. *Nephrol Dial Transplant* 2016; 31: 1887–1894
51. Torres VE, Chapman AB, Devuyst O et al. Multicenter, open-label, extension trial to evaluate the long-term efficacy and safety of early versus delayed treatment with tolvaptan in autosomal dominant polycystic kidney disease: the TEMPO 4:4 Trial. *Nephrol Dial Transplant* 2018; 33: 477–489
52. Sarafidis P, Ferro CJ, Morales E et al. SGLT-2 inhibitors and GLP-1 receptor agonists for nephroprotection and cardioprotection in patients with diabetes mellitus and chronic kidney disease. A consensus statement by the EURECAm and the DIABESITY working groups of the ERA-EDTA. *Nephrol Dial Transplant* 2020; 35: 1825
53. Horie S, Muto S, Kawano H et al. Preservation of kidney function irrelevant of total kidney volume growth rate with tolvaptan treatment in patients with autosomal dominant polycystic kidney disease. *Clin Exp Nephrol* 2021; 25: 467–478
54. Rodriguez D, Kapoor S, Edenhofer I et al. Inhibition of sodium–glucose cotransporter 2 with dapagliflozin in Han:SPRD rats with polycystic kidney disease. *Kidney Blood Press Res* 2015; 40: 638–647
55. Neumiller JJ, White JR Jr, Campbell RK. Sodium–glucose cotransport inhibitors: progress and therapeutic potential in type 2 diabetes mellitus. *Drugs* 2010; 70: 377–385
56. Sen T, Heerspink HJL. A kidney perspective on the mechanism of action of sodium glucose co-transporter 2 inhibitors. *Cell Metab* 2021; 33: 732–739
57. Nagao S, Morita M, Kugita M et al. Polycystic kidney disease in Han:SPRD Cy rats is associated with elevated expression and mislocalization of SamCystin. *Am J Physiol Renal Physiol* 2010; 299: F1078–F1086
58. Wang X, Zhang S, Liu Y et al. Targeting of sodium–glucose cotransporters with phlorizin inhibits polycystic kidney disease progression in Han:SPRD rats. *Kidney Int* 2013; 84: 962–968
59. Abdul-Ghani MA, DeFronzo RA, Norton L. Novel hypothesis to explain why SGLT2 inhibitors inhibit only 30–50% of filtered glucose load in humans. *Diabetes* 2013; 62: 3324–3328
60. Charles River. PCK rat details. Accessed January 20, 2022. <https://www.criver.com/products-services/find-model/pck-rat?region=3611>
61. Brown JH, Bihoreau MT, Hoffmann S et al. Missense mutation in sterile alpha motif of novel protein SamCystin is associated with polycystic kidney disease in (cy/+) rat. *J Am Soc Nephrol* 2005; 16: 3517–3526
62. Hoff S, Halbritter J, Epting D et al. ANKS6 is a central component of a nephronophthisis module linking NEK8 to INVS and NPHP3. *Nat Genet* 2013; 45: 951–956
63. Kapoor S, Rodriguez D, Riwanto M et al. Effect of sodium–glucose cotransport inhibition on polycystic kidney disease progression in PCK rats. *PLoS One* 2015; 10: e0125603
64. Obermeier M, Yao M, Khanna A et al. In vitro characterization and pharmacokinetics of dapagliflozin (BMS-512148), a potent sodium–glucose cotransporter type II inhibitor, in animals and humans. *Drug Metab Dispos* 2010; 38: 405–414
65. Meng W, Ellsworth BA, Nirschl AA et al. Discovery of dapagliflozin: a potent, selective renal sodium-dependent glucose cotransporter 2 (SGLT2) inhibitor for the treatment of type 2 diabetes. *J Med Chem* 2008; 51: 1145–1149
66. Chevalier RL. Pathophysiology of obstructive nephropathy in the newborn. *Semin Nephrol* 1998; 18: 585–593
67. Masuda T, Muto S, Fukuda K et al. Osmotic diuresis by SGLT2 inhibition stimulates vasopressin-induced water reabsorption to maintain body fluid volume. *Physiol Rep* 2020; 8: e14360

68. Ho TA, Godefroid N, Gruzon D et al. Autosomal dominant polycystic kidney disease is associated with central and nephrogenic defects in osmoregulation. *Kidney Int* 2012; 82: 1121–1129
69. Lytvyn Y, Bjornstad P, Katz A et al. SGLT2 inhibition increases serum copeptin in young adults with type 1 diabetes. *Diabetes Metab* 2020; 46: 203–209
70. Chung S, Kim S, Son M et al. Empagliflozin contributes to polyuria via regulation of sodium transporters and water channels in diabetic rat kidneys. *Front Physiol* 2019; 10: 271
71. Cinti F, Moffa S, Impronta F et al. Spotlight on ertugliflozin and its potential in the treatment of type 2 diabetes: evidence to date. *Drug Des Devel Ther* 2017; 11: 2905–2919
72. Heerspink HJL, Stefansson BV, Correa-Rotter R et al. Dapagliflozin in patients with chronic kidney disease. *N Engl J Med* 2020; 383: 1436–1446
73. Herrington WG, Preiss D, Haynes R et al. The potential for improving cardio-renal outcomes by sodium–glucose co-transporter-2 inhibition in people with chronic kidney disease: a rationale for the EMPA-KIDNEY study. *Clin Kidney J* 2018; 11: 749–761
74. ClinicalTrials.gov. https://clinicaltrials.gov/ct2/results?cond=Polycystic+Kidney+Diseases&Search=Apply&recrs=b&recrs=a&recrs=f&recrs=d&age_v=&gndr=&type=&rslt= (4 October, date last accessed)
75. Sarafidis P, Ortiz A, Ferro CJ et al. Sodium–glucose co-transporter-2 inhibitors for patients with diabetic and non-diabetic chronic kidney disease: a new era has already begun. *J Hypertens* 2021; 39: 1090–1097
76. Jensen J, Omar M, Kistorp C et al. Empagliflozin in heart failure patients with reduced ejection fraction: a randomized clinical trial (Empire HF). *Trials* 2019; 20: 374
77. Neal B, Perkovic V, Mahaffey KW et al. Canagliflozin and cardiovascular and renal events in type 2 diabetes. *N Engl J Med* 2017; 377: 644–657
78. Tahrani AA, Barnett AH, Bailey CJ. SGLT inhibitors in management of diabetes. *Lancet Diabetes Endocrinol* 2013; 1: 140–151
79. Blazer-Yost BL, Bacallao RL, Erickson BJ et al. A randomized phase 1b cross-over study of the safety of low-dose pioglitazone for treatment of autosomal dominant polycystic kidney disease. *Clin Kidney J* 2021; 14: 1738–1746