



Review Glyoxalases in Urological Malignancies

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Abstract: Urological cancers include a spectrum of malignancies affecting organs of the reproductive and/or urinary systems, such as prostate, kidney, bladder, and testis. Despite improved primary prevention, detection and treatment, urological cancers are still characterized by an increasing incidence and mortality worldwide. While advances have been made towards understanding the molecular bases of these diseases, a complete understanding of the pathological mechanisms remains an unmet research goal that is essential for defining safer pharmacological therapies and prognostic factors, especially for the metastatic stage of these malignancies for which no effective therapies are currently being used. Glyoxalases, consisting of glyoxalase 1 (Glo1) and glyoxalase 2 (Glo2), are enzymes that catalyze the glutathione-dependent metabolism of cytotoxic methylglyoxal (MG), thus protecting against cellular damage and apoptosis. They are generally overexpressed in numerous cancers as a survival strategy by providing a safeguard through enhancement of MG detoxification. Increasing evidence suggests that glyoxalases, especially Glo1, play an important role in the initiation and progression of urological malignancies. In this review, we highlight the critical role of glyoxalases as regulators of tumorigenesis in the prostate through modulation of various critical signaling pathways, and provide an overview of the current knowledge on glyoxalases in bladder, kidney and testis cancers. We also discuss the promise and challenges for Glo1 inhibitors as future anti-prostate cancer (PCa) therapeutics and the potential of glyoxalases as biomarkers for PCa diagnosis.

Keywords: Glyoxalases; urological malignancies; prostate cancer; kidney cancer; bladder cancer testicular cancer

1. Introduction

Urological cancers consist of prostate, kidney, bladder and testis cancers. Excluding testicular cancer, they are among the ten most frequent malignancies in man [1]. According to the most recent cancer statistics, renal, bladder and prostate cancers constituted more than 33% of all cancers in the United States in 2016 [2]. Despite improved primary prevention, early detection and treatment, urological cancers are still characterized by increasing incidence and mortality worldwide [1], and despite the significant progress in understanding the pathology of these malignancies, the study of the mechanisms involved remains an unmet, but essential, research goal if we are to define safer pharmacological therapies and prognostic factors, especially for the metastatic stage of these neoplastic diseases whose prognosis, unfortunately even today, remains poor. Prostate cancer (PCa) is a major health concern in the older male population all over the world and is the sixth most common cause of cancer-related deaths in the world [3]. Androgen receptor plays a crucial role in the development of PCa and androgen deprivation therapy is the first-line therapy for newly diagnosed PCa patients [4]. Nevertheless, most patients progress to castration-resistant PCa, very often associated with metastases [5]. Bladder cancer (BCa) incidence is three times higher among males than females [6]. Nearly 386,000 newly diagnosed cases and about 150,000 deaths are reported annually worldwide [4]. Renal cell carcinoma (RCC) is the eighth most common cancer in the United States [7] and its incidence is steadily rising in most areas of the world [8]. Total or partial nephrectomy is the optimal primary

treatment. Nevertheless, RCC recurs in 20–40% of patients after resection, which is associated with a worse tumor stage and grade [9]. To further exacerbate the situation, at least some malignancies of the urinary tract are age-related, therefore, it is highly likely that their incidence may continue to rise as a result of global population aging.

Multiple factors are involved in the genesis and progression of urological malignancies, including hypoxia, hormone influence, genetic predisposition, oxidative and glycative stresses and inflammation [10–18]. With regards to oxidative stress, glycative stress and inflammation, a complex interplay is known to exist between these biological phenomena, so that each of them fuels the other in a fine-tuned regulatory circuitry.

A crucial common factor in this interplay is represented by methylglyoxal (MG), an endogenous dicarbonyl metabolite [19], formed during cell metabolism, especially glycolysis (Figure 1), so that MG is particularly abundant in actively proliferating cancerous cells, that primarily rely on glucose metabolism to support their elevated growth demands, according to the so-called "Warburg effect" [20]. MG is a potent glycating agent [19], able to rapidly and spontaneously react with free amino groups in proteins, with DNA and lipids, generating a large family of adducts, called advanced glycation end products (AGEs) [21] (Figure 1).



Figure 1. Formation and modification of macromolecules of/by methylglyoxal (MG). In cells, the majority of MG is formed non-enzymatically as a by-product of glycolysis, in particular, from the degradation of dihydroxyacetone phosphate (DHAP) and glyceraldehyde-3-phosphate (G3P) [22,23]. Minor sources of MG are derived from the hydrolysis and dephosphorylation of DHAP and G3P by triosephosphate isomerase [24], from aminoacetone during catabolism of threonine and from hydroxyacetone in the metabolism of acetone [25,26]. MG can also be formed by the decomposition of lipid peroxidation products [27,28], autoxidation of glucose and degradation of glycated proteins [29–31]. MG can modify arginine (Arg) residues of proteins to generate MG-derived hydroimidazolones (MG-H1, MG-H2, MG-H3), argpyrimidine and tetrahydropyrimidine (THP), as well as lysine (Lys) residues to form N ϵ -(1-carboxyethyl)lysine (CEL) and 1,3-di(N ϵ -lysino)-4-methyl-imidazolium (MOLD). MG also induces stable modifications of DNA bases (2'-deoxyguanosine, dG) and lipids.

In particular, by reacting mainly with arginine residues of proteins, MG forms, primarily, hydroimidazolone adducts (MG-H1, MG-H2 and MG-H3) [32–34], and, secondly, argpyrimidine [35] and tetrahydropyrimidine (THP) [36]. MG can also react with lysine residues of proteins forming N ϵ -(1-carboxyethyl)lysine (CEL) [37] and 1,3-di(N ϵ -lysino)-4-methyl-imidazolium (MOLD) [38]. The interaction of MG with DNA leads preferentially to the formation of imidazopurinone MGdG [39–42], while, by reacting with basic phospholipids, MG forms lipid adducts [42,43]. MG is a crucial factor

in the induction of oxidative stress and in the generation of reactive oxygen species (ROS) [40,44–48]. The major route by which MG generates ROS is by directly modifying proteins, e.g., mitochondrial membrane proteins [49,50], antioxidant enzymes [50,51], endothelial nitric oxide synthase [52] and NADPH oxidase [53]. Moreover, MG, mainly through MG-H1, which serves as a ligand for the receptor of AGEs (RAGE), is an important pro-inflammatory agent [40,54–56].

Increased levels of MG and MG-modified macromolecules, particularly, MG-modified proteins ("the dicarbonyl proteome", [41]) can drive several biological responses, including cell death [57–62], cell transformation [63–66] and senescence [67–69], contributing to cell and tissue dysfunction in aging and disease [41,70] (Figure 2).



Figure 2. Involvement of methylglyoxal (MG)-derived advanced glycation end products (AGEs) in aging and disease. MG-derived AGEs are an important piece of the puzzle representing the pathogenesis of several health problems. NAFLD: non-alcoholic fatty liver disease.

Under physiological conditions, intracellular concentrations of MG and, consequently, of MG-derived AGEs, are strictly regulated. Methylglyoxal is metabolized mainly by glyoxalases [71–77], with minor metabolism by aldoketo reductases (AKRs) and aldehyde dehydrogenases (ADHs) [19,40,78–81] that convert it to hydroxyacetone and pyruvate, respectively [82]. Glyoxalases are cellular enzymes whose increased expression and activity in tumor tissues and/or cell lines [77,83–85], including those from the urogenital tract [86–90], have been widely documented [77,83–90]. In the present review, we provide an overview of the role of glyoxalases in the onset and progression of human urogenital malignancies and glyoxalases' potential as targets for anticancer drug development and biomarkers for diagnosis or prognosis of urological cancers.

2. Glyoxalases

Glyoxalases comprise two enzymes, named glyoxalase 1 (Glo1) and glyoxalase 2 (Glo2). The two enzymes have been historically described to work in tandem as part of a system where Glo1 catalyzes the isomerization of the hemithioacetal formed non-enzymatically by the reaction of MG with reduced glutathione (GSH) into S-D-lactoylglutathione, and Glo2 the hydrolysis of S-D-lactoylglutathione to D-lactate, thereby reforming GSH [21] (Figure 3).



Figure 3. The glyoxalase system. The glyoxalase system consists of glyoxalase 1 (Glo1), glyoxalase 2 (Glo2), and a catalytic amount of glutathione (GSH). Acting as the rate-limiting enzyme, Glo1 transforms the hemithioacetal spontaneously formed between MG and GSH to the thioester S-D-lactoylglutathione. In turn, Glo2 catalyzes the hydrolysis of S-D-lactoylglutathione to D-lactate regenerating GSH.

Glo1 activity can be regulated through several mechanisms including transcriptional regulation [21,59,84,91] and by post-translational modifications [21]. In particular, the promoter region of *Glo1* contains various regulatory elements, including binding sites for activator protein-2 α (AP-2 α), early gene 2 factor isoform 4 (E2F4), nuclear transcription factor- κ B (NF- κ B), and activator protein-1 (AP-1), as well as antioxidant response (ARE), metal-response (MRE), and insulin-response (IRE) elements [91,92] (Figure 4). It has been shown that AP-2 α , E2F4, nuclear factor erythroid 2-related factor 2 (Nrf2) and NF- κ B enhance the activity of Glo1 promoter, and up-regulate Glo1 expression

[93,94]. Phosphorylation, NO-mediated modification and glutathionylation have been described as post-translational modifications of Glo1 [21,95–98] (Figure 4).



Figure 4. Mechanisms of glyoxalase 1 (Glo1) regulation. Glo1 can be regulated at transcriptional and post-translational levels. Transcriptional regulators are activator protein- 2α (AP- 2α), early gene 2 factor isoform 4 (E2F4), nuclear factor erythroid 2-related factor 2 (Nrf2), nuclear transcription factor–kB (NF- κ B), antioxidant response (ARE), metal-response (MRE) and insulin-response (IRE) elements, and activator protein-1 (AP-1). It has been shown that AP- 2α , E2F4, Nrf2 and NF- κ B enhance (switch on symbol) the activity of Glo1 promoter, and up-regulate Glo1 expression. As to *Glo1* regulation by ARE, MRE, IRE and AP-1, a clear demonstration has not been provided yet (question mark symbol). Post-translational modification can occur via glutathionylation, phosphorylation and nitrosylation. GSH: reduced glutathione, NO: nitric oxide; R: the "variable" or "R" group of proteins.

Glo2 expression can be up-regulated by the transcription factors p63 and p73 [99], steroid hormones [88], androgen receptor [90] and phosphatase and tensin homologue (PTEN)/ phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT)/mammalian target of rapamycin (mTOR) signaling [89] (Figure 5). Extensive information about physical and chemical properties of glyoxalases has been largely described in excellent previous reviews [100–104].



Figure 5. Transcriptional regulation of glyoxalase 2 (*Glo2*). Transcriptional positive regulators are (a) the transcription factors p63 and p73; (b) steroid hormones; (c) androgen receptor (AR) and (d) phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT)/mammalian target of rapamycin (mTOR) signaling, one of the most important pathways involved in prostate cancer (PCa) progression, negatively regulated by the tumor suppressor phosphatase and tensin homologue (PTEN).

3. Prostate Cancer (PCa)

3.1. Glyoxalase 1 (Glo1)

The first interest in the role of Glo1 in PCa dates back to the 1990s, when Ayoub et al. [105] and Di llio et al. [106] measured Glo1 specific activity in human tumor cell lines in vitro or cancerous and non-cancerous tissues. Although PCa tissues were poorly represented and PCa cell lines not clearly identified, these results suggested a role for Glo1 in PCa biology, its enzymatic activity being higher in cancerous cells than non-malignant ones. The first seminal observations of the potential role of Glo1 in PCa onset were made later by Davidson et al. [107] who performed a quantitative PAGE-based Glo1 assay on 22 PCa and ten normal prostate tissues to assess its activity, and thereby establish its clinical importance in this neoplasm. These results demonstrated that Glo1 activity differed substantially and significantly between non-cancerous and cancerous specimens, with a densitometric analysis showing differences in the range from five- to eight-fold in PCa compared with non-cancerous samples. Notably, a distinctly high activity of Glo1 was observed in all 22 PCa specimens and with little to no activity detected in ten out of ten non-malignant specimens. Although preliminary, these results provided new impetus to the investigation of Glo1 expression, either at mRNA, protein or functional level, in PCa tissues and normal counterparts or in differentially aggressive and invasive PCa [89,90,108–110]. Overall, these studies strongly suggested Glo1 as an important new target in PCa genesis and progression. Overexpression of Glo1 in PCa cells would maintain low intracellular levels of the glycolysis-derived cytotoxic metabolite MG, as a survival defense strategy. Indeed, in vitro studies performed with PCa cell lines, clearly demonstrated just such a role [57,58,89], in addition to pointing out the pro-survival function of this protein by eluding apoptosis in a mechanism involving

the desensitization of NF- κ B pathway [58]. Only one study, based on the proteomic analysis of normal and malignant prostate tissues, identified Glo1 among the proteins lost in PCa [111]. While this result was not further validated, it is worth raising a note of caution here. The role of Glo1 in PCa carcinogenesis has supported the view of Glo1 both as a marker for diagnosis of PCa and a potential target for the development of novel anti-cancer or chemotherapeutic agents for PCa, which were indeed confirmed by some studies. In particular, Chavan et al. [112] suggested serum Glo1 levels as a supplemental biomarker to differentiate between malignant and non-malignant diseases of the prostate in patients with PSA in the range of 4–20 ng/mL. Similarly, we first explored the association of Glo1 Ala111Glu (A111E) (419A/C) functional polymorphism, associated with a loss-of-function mutation. Our findings supported the idea of a pathogenic role of Glo1 in PCa as well as identified an additional candidate for risk assessment in PCa patients and an independent prognostic factor for survival [113]. Although further investigations with a larger number of participants are absolutely required to confirm these findings, these explorative studies have stimulated further investigations important for PCa diagnosis and encourage further progress in this field. So far, the gold standard in diagnosis for urological neoplasms involves tissue biopsy, and early screening methods are rare. Some existing biomarkers such as prostate-specific antigen (PSA) may be useful in PCa screening, but its utility is hampered by its poor specificity, leading to over-diagnosis and over-treatment, which limits its application [114–116]. With regards to Glo1 as a drug target, Sharkey et al. [117] provided the first demonstration that a competitive inhibitor of Glo1 effectively inhibited the growth of PCa tumors in mice, when delivered as the diethyl ester prodrug. Notably, the antitumor effects of this Glo1 inhibitor were comparable to those observed with the standard antitumor agents. Carmustine (BCNU) is an anticancer agent and another putative inhibitor of Glo1, while β -glucan is a unique, nontoxic polysaccharide extracted from mushrooms. In 2002, treatment of PC3 PCa cells with a combination of BCNU with β -glucan demonstrated a sensitized cytotoxic effect which was associated with a striking (approximately 80%) inactivation of Glo1 [118]. More recently, it has been also reported that curcumin inhibits Glo1 with the consequent intracellular accumulation of toxic levels of MG and GSH, that, by modulating various metabolic cellular pathways, inhibit growth of malignant cells [119]. Finally, Baunacke et al. [109] identified ethyl pyruvate as an agent targeting Glo1 that reduced some malignancy-associated properties of PCa cells while Valenti et al. [120] found that 3-bromopyruvate induced rapid human PCa cell death by affecting, among other targets, Glo1. Although these results convincingly suggested a crucial role for Glo1 in PCa, we believe that further clinical and/or in animal studies are necessary in the future before drawing definitive conclusions.

3.2. Glyoxalase 2 (Glo2)

In marked contrast to Glo1, the role of Glo2 in PCa has been relatively overlooked [89,90,105,106]. Ayoub et al. [105] and Di Ilio et al. [106] first evaluated Glo2 enzymatic activity in human PCa cell lines and tissues. They found that Glo2 enzyme activity in cancerous cells was not so different from that of non-cancerous cells. However, the small number of PCa samples considered in those studies significantly limited the strength of the conclusions drawn on the biological significance of Glo2 in this neoplasia. Only recently we demonstrated a causative role of Glo2 in PCa [89,90] providing in vivo and in vitro evidence for a role of this enzyme in prostate carcinogenesis. In particular, we showed that Glo2 was selectively expressed in PCa but not in the luminal compartment of the adjacent benign epithelium, consistently in all the examined cases (n = 20). Moreover, we demonstrated that Glo2 expression in malignant prostate cells was dependent on androgen receptor, in line with another previous exploratory study by our group [88], and was linked to enhanced cell proliferation and resistance to apoptosis through a mechanism involving the p53-p21 axis. Hence, our results demonstrated, for the first time, a role of Glo2 in prostate tumorigenesis as well as suggesting a possible mechanism. Both genetic and environmental factors participate in PCa pathogenesis [121,122]. Nevertheless, the molecular biology and mechanisms of prostate carcinogenesis remain to be further elucidated in order to identify additional diagnostic factors. As mentioned above, while the possibility of biomarkers for PCa has

been investigated for some molecules, their prospects in clinical application still need to be further evaluated [114–116,123]. There is an urgent need for additional biomarkers that supplement PSA. Since we demonstrated that the luminal Glo2 is able to drive prostate tumorigenesis and that it is not expressed in benign luminal cells but only in PCa, we proposed that it might represent a novel marker in the pathological diagnosis of early PCa, by distinguishing between benign and malignant lesions. In fact, detection of Glo2 staining in luminal cells of the benign gland during diagnostic investigations might be indicative of an initial/ongoing neoplastic transformation process. In addition, we found in tissue sections, that Glo2 was also intensely expressed in the basal cells of benign glands, even though this basal cell-localized Glo2 was not involved in PCa genesis. The histological diagnosis of PCa, at least in difficult cases, is based upon prostate gland architectural and cytological features, with basal cell loss as a hallmark of malignancy. When the growth pattern is obscured, as can be the case in core needle biopsies with few suspicious glands, assessing these basal cells immunohistochemically is the usual auxiliary approach in confirming or excluding malignancy [124]. A variety of basal cell markers have been suggested. The standard markers used routinely in surgical pathology are high molecular weight cytokeratins (e.g., $34\beta E12$, CK5/6) or p63, both of which stain basal cells with high sensitivity and specificity [124]. However, in some cases, false-negative staining may occur because of patchy cytoplasmic staining, making a definitive diagnosis difficult. We suggested that Glo2 immunostaining could represent a novel additional marker for basal cells in the benign prostate and its absence in malignant cells, consequent to the loss of basal cells, might have been useful as a complement to existing methods to facilitate the pathological diagnosis of PCa. In particular, since the loss of basal cells is an early event during tumorigenesis, we believe that Glo2 might be especially important for the early diagnosis of PCa. The concept of a metabolic pathway comprising Glo2 and Glo1, is a widely accepted idea. However, this view was first challenged in 1970 with a report of Glo2 deficiency in erythrocytes [125]. The authors studied this deficiency in family members for three generations demonstrating that this condition is transmitted as an autosomal recessive trait. Moreover, they observed that this deficiency was absent in leucocytes, while Glo1 activity was entirely normal in both leukocytes and erythrocytes, and was three times more active than Glo2 [126]. Additional studies in other eukaryotic organisms (reviewed in [101,126]) suggested further exceptions to the concept of a pathway composed of Glo1 and Glo2, acting sequentially. In our study, the results from IHC, confirmed in PCa cell models by multidisciplinary approaches, showed that the localization of Glo1 was opposite to that of Glo2. In all 20 cases of non-malignant glands, Glo1 staining was consistently observed only in the luminal but not in the basal cells, even with the use of different Glo1 antibody solutions, suggesting an independent role of the two proteins. Moreover, when attempting to demonstrate a possible role of Glo1 in association with Glo2 in PCa genesis via cell growth control we did not find any robust dependency. We suggest that Glo2 involvement in prostate tumorigenesis is independent of Glo1, at least as far as cell growth control is concerned, and/or that Glo1 may be involved in PCa genesis via the control of other tumorigenesis-related biological processes, possibly induced by factors in the cancer microenvironment. Further research is needed to clarify this intriguing finding. On the other hand, as mentioned above, we recently found that Glo2, this time in cooperation with Glo1, is also involved in human PCa progression as part of a mechanism driven by PTEN/PI3K/AKT/mTOR signaling with involvement of PKM2 and estrogen receptor α [89]. In particular, we found that both enzymes were up-regulated in both aggressive PCa tissues and cell lines and this up-regulation mediated maintenance of low intracellular levels of the glycolysis-derived cytotoxic metabolites MG and LSG, very likely as a survival defense strategy. Importantly, Glo2 silencing did not alter the behavior of benign cells, suggesting that targeting this protein might represent a strategy to selectively inhibit advanced PCa. Additional in vivo studies are absolutely required to further prove this function for both enzymes. Hence, despite significant research efforts and important new insights, the role of glyoxalases in the pathology and progression of PCa remains opaque; much more work must still be done.

4. Renal Cancer (RCa)

4.1. Glyoxalase 1 (Glo1)

Back in 1995, Di Ilio and colleagues evaluated Glo1 enzyme activity in the cytosolic fractions prepared from 15 samples of neoplastic and healthy kidney tissues from patients with primary renal cell carcinoma [106]. However, no significant differences were observed. Ten years later, we investigated Glo1 expression, at both mRNA and protein level, in a human renal carcinoma (clear cell adenocarcinoma, CCA), known to be strongly resistant to chemotherapeutic drugs, and in pair-matched normal tissues [87]. This study provided clear evidence for significantly higher (about nine-fold) levels of Glo1 mRNA and activity (2.5-fold) and markedly reduced intracellular levels of pro-apoptotic MG in CCA than in control tissues, suggesting a mechanism explaining, at least in part, the refractoriness of CCA to a number of apoptosis-inducing chemotherapy treatments. In fact, despite the discovery of a number of new cytotoxic agents in the recent years, kidney cancer treatment is still unsatisfactory, mainly due to a marked resistance of this tumor towards these drugs. Hence, we suggested that the synthesis of new molecules selectively down-regulating Glo1 expression in cancer cells might be an alternative method of therapeutic treatment for this neoplasia, in particular CCA. Our study was later extended by Tanaka and colleagues who studied Glo1 expression in human renal tumor cell lines with diverse metastatic potential [127]. By using two-dimensional electrophoresis followed by liquid chromatography-tandem mass spectrometry, they found that protein Glo1 appeared to be directly proportional to the metastatic potential of the studied clones, and was also the only protein which consistently varied with the metastatic potentials of CCR clones: Glo1 was increased in highly metastatic cell lines compared with those with lower metastatic potential and this was confirmed by Western blot analysis. These results, in agreement with ours [87], suggested that the increased Glo1 expression in more aggressive CCR cells might provide advantages to the metastatic potential in aggressive clones, and that Glo1 inhibition might be a therapeutic approach for the treatment of metastatic CCR. A novel mechanism underlying increased Glo1 expression in tumors is *Glo1* gene amplification [72]. A quite recent study demonstrated a direct link between *Glo1* gene amplification, overexpression and increased sensitivity to Glo1 inhibition in 6.5% of RCa, and suggested that cell lines with *Glo1* amplification were significantly more sensitive to *Glo1* inhibition than those without. Moreover, it was suggested that Glo1 might represent a useful target for therapy in cancers with Glo1 amplification, such as RCa. As for studies on the association of the *Glo1* A111E (419A/C) polymorphism with the risk of RCa, they have been limited to only one study in a cohort of 214 patients with CCA [128]. This study demonstrated the link between the A111E Glo1 single nucleotide polymorphism (SNP) and the presence of CCR. In particular, a higher frequency of the C allele and genotype CC in patients with CCR was observed in comparison with the control group. This result would be in accordance with the fact that the C allele is linked to a higher enzymatic activity of Glo1, which was previously observed in another human urological malignancy [113]. However, although this study clearly demonstrated Glo1 involvement in kidney cancer, it identified some potential limitations, the most important of which was the lack of a biological link, since Glo1 activity was not measured either in tissue or blood samples.

4.2. Glyoxalase 2 (Glo2)

A clear connection between Glo2 and RCa was first established in 1995 when, once again, Di Ilio and colleagues [106] evaluated Glo2 enzyme activity in the cytosolic fractions from 15 samples of neoplastic or healthy kidney tissues of patients with primary RCC. In this study, Di Ilio et al. [106] revealed a significantly lower (approximately four-fold) Glo2 activity in malignant than in non-malignant kidney tissues. Only one study by our group followed this exploratory observation [87]. In agreement with Di Ilio et al. [106], we demonstrated a significant decrease of Glo2 enzyme activity (approximately two-fold) in the tumor compared with non-tumor tissues.

5. Bladder Cancer (BCa)

Glyoxalase 1 (Glo1) and Gyoxalase 2 (Glo2)

Little information is available on glyoxalases in BCa [86,106]. A study [106] reported glyoxalases' activity in six samples of normal bladder and in the tumor and non-tumor samples of eight bladders. Large inter-individual variations were observed in both Glo1 and Glo2 activities which, according to the authors, made the definition of normal baseline level difficult to set. Although no significant difference in Glo1 means activity was found between tumor and non-tumor samples of BCa, relatively higher tumor Glo1 activity, compared with the corresponding non-tumor tissues, was found in five samples. As to Glo2, no significant differences were observed between the cancerous component and the normal counterpart. Next, we studied *Glo1* and *Glo2* gene expression and enzyme activity in human BCa compared with the corresponding normal mucosa [86]. In particular, samples of these tissues were collected from 26 patients with superficial (SBCa) or invasive bladder cancer (IBCa). Both Glo1 mRNA expression and activity were significantly increased in SBCa samples, while they remained unchanged in IBCa samples, compared with the normal mucosa. Conversely, Glo2 showed a higher activity in the tumor (either SBCa or IBCa samples) versus normal tissues. These results suggested that the differing levels of Glo1 activity level and gene expression between the SBCa and IBCa samples might have helped in their differential diagnosis.

6. Testis Cancer (TCa)

Glo1 and Glo2

As far as we know, only Di Ilio and colleagues reported on the activity of glyoxalases in TCa more than twenty years ago [106]. However, with the exception of one sample, they found Glo1 and Glo2 activity essentially identical between tumor and non-tumor samples. Significantly more research is needed to investigate the role of glyoxalases in this urogenital malignancy.

7. Concluding Remarks

7.1. Prostate Cancer (PCa)

Studies so far performed on the role of glyoxalases in urological cancers have mainly focused on PCa. These studies pointed to a role for Glo1 in prostate carcinogenesis, especially during PCa progression, according to which increased expression and activity of Glo1 would be permissive for survival and growth of tumors with relatively high glycolytic rates and related high fluxes of MG formation. Hence, in PCa, the use of specific inhibitors of Glo1 might indeed represent an effective anticancer strategy. Several natural compounds have been proposed as Glo1 inhibitors attenuating the growth of PCa [109,118–120]. In addition, these studies demonstrated that as well as Glo1, Glo2 represents a novel contributor to PCa progression [90] and, more importantly, that Glo1/Glo2 silencing did not alter the behavior of benign cells [90], suggesting that targeting glyoxalases may represent a strategy to selectively inhibit advanced PCa. There is an urgent need for in vivo studies to validate these important conclusions and move them towards potential clinical application. While the traditional cooperative role of Glo1/Glo2 appears valid in PCa progression, in PCa genesis it appears that Glo1 and Glo2 have independent roles, not working sequentially as a metabolic system, helping cast further doubt on whether the glyoxalase pathway is really a pathway [101], at least in some settings. Additional studies are needed to shed new light on the role of these intricate and intriguing proteins in the specific context of PCa and, more widely, in the oncologic ambit. Finally, studies performed on the role of glyoxalases in PCa demonstrated that Glo1 and Glo2 may represent novel additional markers in the pathological diagnosis of advanced and early PCa, respectively. We hope that urologist and oncologist alike, thanks to these new advances, may find renewed interest in these proteins which are frequently overlooked in cancer research.

7.2. Renal and Bladder Cancers (RCa, BCa)

The role of glyoxalases in renal and bladder cancers remains comparatively understudied. The little information that is available needs to be further corroborated but supports the potential use of these proteins as biomarkers of these neoplastic diseases, whose incidence continues to rapidly increase worldwide. In particular, glyoxalases might be useful in the early detection and post-surgical follow up [14,128] since no biomarkers currently exist for either application [129]. In fact, RCa and BCa lack specific predictive biomarkers meaning that diagnosis frequently only follows the presentation of advanced symptoms [129,130].

7.3. Testis Cancer (TCa)

The study of the role of glyoxalases in TCa is today basically still a "virgin field". TCas are a group of heterogeneous, biologically diverse and clinically challenging neoplasms. Despite the relatively low incidence and mortality rates, a subgroup of patients with disseminated disease relapse after conventional therapy and have a dismal prognosis. Moreover, TCs afflict mostly young men and have therapeutic peculiarities, with some patients showing resistance to cisplatin-based treatments and others being troubled by irreversible side effects, such as infertility [131,132]. It is to be hoped that these disease features will provide powerful motivation for further research in this area.

8. Future Directions

Research on glyoxalases has experienced frequent and surprising turns ever since they were discovered. Research on glyoxalases in urological malignancies might reveal, as has already occurred in the past, novel and interesting roles of these proteins and bring them to the limelight once more.

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Abbreviations

Glo1	Glyoxalase 1
Glo2	Glyoxalase 2
MG	Methylglyoxal
AGEs	Advanced glycation end products
AKRs	Aldoketo reductases
ADHs	Aldehyde dehydrogenases
PCa	Prostate cancer
BCa	Bladder cancer
RCC	Renal cell carcinoma
RCa	Renal cancer
ROS	Reactive oxygen species
GSH	Glutathione
PSA	Prostate-specific antigen
BCNU	Carmustine
CCA	Clear cell adenocarcinoma
SNP	Single nucleotide polymorphism
SBCa	Superficial bladder cancer
IBCa	Invasive bladder cancer
TCa	Testis cancer

References

- 1. Torre, L.A.; Bray, F.; Siegel, R.L.; Ferlay, J.; Lortet-Tieulent, J.; Jemal, A. Global cancer statistics, 2012. *CA Cancer J. Clin.* **2015**, 65, 87–108. [CrossRef] [PubMed]
- Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2016. CA Cancer J. Clin. 2016, 66, 7–30. [CrossRef] [PubMed]
- 3. Liu, X.; Wu, Q.; Li, L. Functional and therapeutic significance of EZH2 in urological cancers. *Oncotarget* 2017, *8*, 38044–38055. [CrossRef] [PubMed]
- 4. Ferlay, J.; Shin, H.R.; Bray, F.; Forman, D.; Mathers, C.; Parkin, D.M. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. *Int. J. Cancer* 2010, *127*, 2893–2917. [CrossRef] [PubMed]
- 5. Kahn, B.; Collazo, J.; Kyprianou, N. Androgen receptor as a driver of therapeutic resistance in advanced prostate cancer. *Int. J. Biol. Sci.* **2014**, *10*, 588–595. [CrossRef] [PubMed]
- 6. Ploeg, M.; Aben, K.K.; Kiemeney, L.A. The present and future burden of urinary bladder cancer in the world. *World J. Urol.* **2009**, *27*, 289–293. [CrossRef] [PubMed]
- Siegel, R.; Naishadham, D.; Jemal, A. Cancer statistics, 2013. CA Cancer J. Clin. 2013, 63, 11–30. [CrossRef] [PubMed]
- 8. Sun, M.; Thuret, R.; Abdollah, F.; Lughezzani, G.; Schmitges, J.; Tian, Z.; Shariat, S.F.; Montorsi, F.; Patard, J.J.; Perrotte, P.; et al. Age-adjusted incidence, mortality, and survival rates of stage-specific renal cell carcinoma in North America: A trend analysis. *Eur. Urol.* **2011**, *59*, 135–141. [CrossRef] [PubMed]
- 9. Chin, A.I.; Lam, J.S.; Figlin, R.A.; Belldegrun, A.S. Surveillance strategies for renal cell carcinoma patients following nephrectomy. *Rev. Urol.* **2006**, *8*, 1–7. [PubMed]
- 10. Udensi, U.K.; Tchounwou, P.B. Oxidative stress in prostate hyperplasia and carcinogenesis. *J. Exp. Clin. Cancer Res.* **2016**, *35*, 139. [CrossRef] [PubMed]
- 11. Gakis, G. The role of inflammation in bladder cancer. *Adv. Exp. Med. Biol.* **2014**, *816*, 183–196. [CrossRef] [PubMed]
- 12. Yang, S.; Pinney, S.M.; Mallick, P.; Ho, S.M.; Bracken, B.; Wu, T. Impact of Oxidative Stress Biomarkers and Carboxymethyllysine (an Advanced Glycation End Product) on Prostate Cancer: A Prospective Study. *Clin. Genitourin. Cancer* **2015**, *13*, e347–e351. [CrossRef] [PubMed]
- 13. Elangovan, I.; Thirugnanam, S.; Chen, A.; Zheng, G.; Bosland, M.C.; Kajdacsy-Balla, A.; Gnanasekar, M. Targeting receptor for advanced glycation end products (RAGE) expression induces apoptosis and inhibits prostate tumor growth. *Biochem. Biophys. Res. Commun.* **2012**, *417*, 1133–1138. [CrossRef] [PubMed]
- 14. Guo, Y.; Xia, P.; Zheng, J.J.; Sun, X.B.; Pan, X.D.; Zhang, X.; Wu, C.Z. Receptors for advanced glycation end products (RAGE) is associated with microvessel density and is a prognostic biomarker for clear cell renal cell carcinoma. *Biomed. Pharmacother.* **2015**, *73*, 147–153. [CrossRef] [PubMed]
- Hirakawa, Y.; Inagi, R. Glycative Stress and Its Defense Machinery Glyoxalase 1 in Renal Pathogenesis. *Int. J. Mol. Sci.* 2017, *18*, 174. [CrossRef] [PubMed]
- Mikhaylova, O.; Stratton, Y.; Hall, D.; Kellner, E.; Ehmer, B.; Drew, A.F.; Gallo, C.A.; Plas, D.R.; Biesiada, J.; Meller, J.; et al. VHL-regulated MiR-204 suppresses tumor growth through inhibition of LC3B-mediated autophagy in renal clear cell carcinoma. *Cancer Cell* 2012, *21*, 532–546. [CrossRef] [PubMed]
- Solomon, D.A.; Kim, J.S.; Bondaruk, J.; Shariat, S.F.; Wang, Z.F.; Elkahloun, A.G.; Ozawa, T.; Gerard, J.; Zhuang, D.; Zhang, S.; et al. Frequent truncating mutations of STAG2 in bladder cancer. *Nat. Genet.* 2013, 45, 1428–1430. [CrossRef] [PubMed]
- 18. Attard, G.; Parker, C.; Eeles, R.A.; Schröder, F.; Tomlins, S.A.K.; Tannock, I.; Drake, C.G.; de Bono, J.S. Prostate cancer. *Lancet* 2016, *387*, 70–82. [CrossRef]
- 19. Rabbani, N.; Thornalley, P.J. Dicarbonyl stress in cell and tissue dysfunction contributing to ageing and disease. *Biochem. Biophys. Res. Commun.* 2015, 458, 221–226. [CrossRef] [PubMed]
- 20. Ward, P.S.; Thompson, C.B. Metabolic reprogramming: A cancer hallmark even warburg did not anticipate. *Cancer Cell* **2012**, *21*, 297–308. [CrossRef] [PubMed]
- 21. Bellahcène, A.; Nokin, M.J.; Castronovo, V.; Schalkwijk, C. Methylglyoxal-derived stress: An emerging biological factor involved in the onset and progression of cancer. *Semin. Cancer Biol.* **2017**. [CrossRef] [PubMed]
- 22. Richard, J.P. Mechanism for the formation of methylglyoxal from triosephosphates. *Biochem. Soc. Trans.* **1993**, 21, 549–553. [CrossRef] [PubMed]

- 23. Phillips, S.A.; Thornalley, P.J. The formation of methylglyoxal from triose phosphates. Investigation using a specific assay for methylglyoxal. *Eur. J. Biochem.* **1993**, *212*, 101–105. [CrossRef] [PubMed]
- 24. Pompliano, D.L.; Peyman, A.; Knowles, J.R. Stabilization of a reaction intermediate as a catalytic device: Definition of the functional role of the flexible loop in triosephosphate isomerase. *Biochemistry* **1990**, *29*, 3186–3194. [CrossRef] [PubMed]
- 25. Ray, M.; Ray, S.J. Aminoacetone oxidase from goat liver. Formation of methylglyoxal from aminoacetone. *Biol. Chem.* **1987**, *262*, 5974–5977.
- Reichard, G.A., Jr.; Skutches, C.L.; Hoeldtke, R.D.; Owen, O.E. Acetone metabolism in humans during diabetic ketoacidosis. *Diabetes* 1986, 35, 668–674. [CrossRef] [PubMed]
- 27. Baynes, J.W.; Thorpe, S.R. Glycoxidation and lipoxidation in atherogenesis. *Free Radic. Biol. Med.* **2000**, *28*, 1708–1716. [CrossRef]
- 28. Esterbauer, H.; Gebicki, J.; Puhl, H.; Jürgens, G. The role of lipid peroxidation and antioxidants in oxidative modification of LDL. *Free Radic. Biol. Med.* **1992**, *13*, 341–390. [CrossRef]
- 29. Chetyrkin, S.; Mathis, M.; Pedchenko, V.; Sanchez, O.A.; McDonald, W.H.; Hachey, D.L.; Madu, H.; Stec, D.; Hudson, B.; Voziyan, P. Glucose autoxidation induces functional damage to proteins via modification of critical arginine residues. *Biochemistry* **2011**, *50*, 6102–6112. [CrossRef] [PubMed]
- 30. Thornalley, P.J.; Langborg, A.; Minhas, H.S. Formation of glyoxal, methylglyoxal and 3-deoxyglucosone in the glycation of proteins by glucose. *Biochem. J.* **1999**, *344 Pt 1*, 109–116. [CrossRef] [PubMed]
- 31. Giacco, F.; Brownlee, M. Oxidative stress and diabetic complications. *Circ. Res.* **2010**, *107*, 1058–1070. [CrossRef] [PubMed]
- Ahmed, N.; Argirov, O.K.; Minhas, H.S.; Cordeiro, C.A.; Thornalley, P.J. Assay of advanced glycation endproducts (AGEs): Surveying AGEs by chromatographic assaywith derivatization by 6-aminoquinolyl-*N*-hydroxysuccinimidyl-carbamate and application to Nepsilon-carboxymethyl-lysineand Nepsilon-(1-carboxyethyl)lysine-modified albumin. *Biochem. J.* 2002, 364 Pt 1, 1–14. [PubMed]
- 33. Ahmed, N.; Thornalley, P.J. Chromatographic assay of glycation adducts in human serum albumin glycated in vitro by derivatization with 6-aminoquinolyl-*N*-hydroxysuccinimidyl-carbamate and intrinsic fluorescence. *Biochem. J.* **2002**, *364 Pt* **1**, 15–24. [CrossRef] [PubMed]
- 34. Westwood, M.E.; Thornalley, P.J. Molecular characteristics of methylglyoxal-modified bovine and human serum albumins. Comparison with glucose-derived advanced glycation endproduct-modified serum albumins. *J. Protein Chem.* **1995**, *14*, 359–372. [CrossRef] [PubMed]
- Shipanova, I.N.; Glomb, M.A.; Nagaraj, R.H. Protein modification by methylglyoxal: Chemical nature and synthetic mechanism of a major fluorescent adduct. *Arch. Biochem. Biophys.* 1997, 344, 29–36. [CrossRef] [PubMed]
- Oya, T.; Hattori, N.; Mizuno, Y.; Miyata, S.; Maeda, S.; Osawa, T.; Uchida, K. Methylglyoxal modification of protein. Chemical and immunochemical characterization of methylglyoxal-arginine adducts. *J. Biol. Chem.* 1999, 274, 18492–18502. [CrossRef] [PubMed]
- Ahmed, M.U.; Brinkmann Frye, E.; Degenhardt, T.P.; Thorpe, S.R.; Baynes, J.W. *N*-epsilon-(carboxyethyl) lysine, a product of the chemical modification of proteins by methylglyoxal, increases with age in human lens proteins. *Biochem. J.* 1997, 324 Pt 2, 565–570. [CrossRef] [PubMed]
- Odani, H.; Shinzato, T.; Usami, J.; Matsumoto, Y.; Brinkmann Frye, E.; Baynes, J.W.; Maeda, K. Imidazolium crosslinks derived from reaction of lysine with glyoxal and methylglyoxal are increased in serum proteins of uremic patients: Evidence for increased oxidative stress in uremia. *FEBS Lett.* 1998, 427, 381–385. [CrossRef]
- Rabbani, N.; Xue, M.; Weickert, M.O.; Thornalley, P.J. Multiple roles of glyoxalase 1-mediated suppression of methylglyoxal glycation in cancer biology—Involvement in tumour suppression, tumour growth, multidrug resistance and target for chemotherapy. *Semin. Cancer Biol.* 2017. [CrossRef] [PubMed]
- 40. Piperi, C.; Adamopoulos, C.; Papavassiliou, A.G. Potential of glycative stress targeting for cancer prevention. *Cancer Lett.* **2017**, *390*, 153–159. [CrossRef] [PubMed]
- 41. Rabbani, N.; Xue, M.; Thornalley, P.J. Methylglyoxal-induced dicarbonyl stress in aging and disease: First steps towards glyoxalase 1-based treatments. *Clin. Sci. (Lond.)* **2016**, *130*, 1677–1696. [CrossRef] [PubMed]
- Thornalley, P.J.; Rabbani, N. Glyoxalase in tumourigenesis and multidrug resistance. *Semin. Cell Dev. Biol.* 2011, 22, 318–325. [CrossRef] [PubMed]

- 43. Shoji, N.; Nakagawa, K.; Asai, A.; Fujita, I.; Hashiura, A.; Nakajima, Y.; Oikawa, S.; Miyazawa, T. LC-MS/MS analysis of carboxymethylated and carboxyethylated phosphatidylethanolamines in human erythrocytes and blood plasma. *J. Lipid Res.* **2010**, *51*, 2445–2453. [CrossRef] [PubMed]
- 44. Chang, T.; Wang, R.; Wu, L. Methylglyoxal-induced nitric oxide and peroxynitrite production in vascular smooth muscle cells. *Free Radic. Biol. Med.* **2005**, *38*, 286–293. [CrossRef] [PubMed]
- 45. Wang, H.; Meng, Q.H.; Chang, T.; Wu, L. Fructose-induced peroxynitrite production is mediated by methylglyoxal in vascular smooth muscle cells. *Life Sci.* **2006**, *79*, 2448–2454. [CrossRef] [PubMed]
- 46. Chang, T.; Untereiner, A.; Liu, J.; Wu, L. Interaction of methylglyoxal and hydrogen sulfide in rat vascular smooth muscle cells. *Antioxid. Redox Signal.* **2010**, *12*, 1093–10100. [CrossRef] [PubMed]
- 47. Nagaraj, R.H.; Oya-Ito, T.; Bhat, M.; Liu, B. Dicarbonyl stress and apoptosis of vascular cells: Prevention by alphaB-crystallin. *Ann. N. Y. Acad. Sci.* **2005**, *1043*, 158–165. [CrossRef] [PubMed]
- 48. Chan, W.H.; Wu, H.J. Methylglyoxal and high glucose co-treatment induces apoptosis or necrosis in human umbilical vein endothelial cells. *J. Cell. Biochem.* **2008**, *103*, 1144–1157. [CrossRef] [PubMed]
- Rosca, M.G.; Monnier, V.M.; Szweda, L.I.; Weiss, M.F. Alterations in renal mitochondrial respiration in response to the reactive oxoaldehyde methylglyoxal. *Am. J. Physiol. Ren. Physiol.* 2002, 283, F52–F59. [CrossRef] [PubMed]
- 50. Roy, A.; Ahir, M.; Bhattacharya, S.; Parida, P.K.; Adhikary, A.; Jana, K.; Ray, M. Induction of mitochondrial apoptotic pathway in triple negative breast carcinoma cells by methylglyoxal via generation of reactive oxygen species. *Mol. Carcinog.* **2017**. [CrossRef] [PubMed]
- 51. Wu, L.; Juurlink, B.H. Increased methylglyoxal and oxidative stress in hypertensive rat vascular smooth muscle cells. *Hypertension* **2002**, *39*, 809–814. [CrossRef] [PubMed]
- 52. Su, Y.; Qadri, S.M.; Wu, L.; Liu, L. Methylglyoxal modulates endothelial nitric oxide synthase-associated functions in EA.hy926 endothelial cells. *Cardiovasc. Diabetol.* **2013**, *12*, 134. [CrossRef] [PubMed]
- Phalitakul, S.; Okada, M.; Hara, Y.; Yamawaki, H. Vaspin prevents methylglyoxal-induced apoptosis in human vascular endothelial cells by inhibiting reactive oxygen species generation. *Acta Physiol. (Oxf.)* 2013, 209, 212–219. [CrossRef] [PubMed]
- Vulesevic, B.; McNeill, B.; Giacco, F.; Maeda, K.; Blackburn, N.J.; Brownlee, M.; Milne, R.W.; Suuronen, E.J. Methylglyoxal-Induced Endothelial Cell Loss and Inflammation Contribute to the Development of Diabetic Cardiomyopathy. *Diabetes* 2016, 65, 1699–1713. [CrossRef] [PubMed]
- 55. Ishibashi, Y.; Matsui, T.; Nakamura, N.; Sotokawauchi, A.; Higashimoto, Y.; Yamagishi, S.I. Methylglyoxal-derived hydroimidazolone-1 evokes inflammatory reactions in endothelial cells via an interaction with receptor for advanced glycation end products. *Diabetes Vasc. Dis. Res.* 2017, 14, 450–453. [CrossRef] [PubMed]
- Yang, C.T.; Meng, F.H.; Chen, L.; Li, X.; Cen, L.J.; Wen, Y.H.; Li, C.C.; Zhang, H. Inhibition of Methylglyoxal-Induced AGEs/RAGE Expression Contributes to Dermal Protection by N-Acetyl-L-Cysteine. *Cell. Physiol. Biochem.* 2017, 41, 742–754. [CrossRef] [PubMed]
- 57. Antognelli, C.; Mezzasoma, L.; Fettucciari, K.; Talesa, V.N. A novel mechanism of methylglyoxal cytotoxicity in prostate cancer cells. *Int. J. Biochem. Cell Biol.* **2013**, *45*, 836–844. [CrossRef] [PubMed]
- Antognelli, C.; Mezzasoma, L.; Fettucciari, K.; Mearini, E.; Talesa, V.N. Role of glyoxalase I in the proliferation and apoptosis control of human LNCaP and PC3 prostate cancer cells. *Prostate* 2013, 73, 121–132. [CrossRef] [PubMed]
- Antognelli, C.; Palumbo, I.; Aristei, C.; Talesa, V.N. Glyoxalase I inhibition induces apoptosis in irradiated MCF-7 cells via a novel mechanism involving Hsp27, p53 and NF-κB. *Br. J. Cancer* 2014, 111, 395–406. [CrossRef] [PubMed]
- 60. Chan, W.H.; Wu, H.J.; Shiao, N.H. Apoptotic signaling in methylglyoxal-treated human osteoblasts involves oxidative stress, c-Jun N-terminal kinase, caspase-3, and p21-activated kinase 2. *J. Cell. Biochem.* **2007**, *100*, 1056–1069. [CrossRef] [PubMed]
- Angeloni, C.; Malaguti, M.; Rizzo, B.; Barbalace, M.C.; Fabbri, D.; Hrelia, S. Neuroprotective effect of sulforaphane against methylglyoxal cytotoxicity. *Chem. Res. Toxicol.* 2015, 28, 1234–1245. [CrossRef] [PubMed]
- Wang, Y.H.; Han, Y.P.; Yu, H.T.; Pu, X.P.; Du, G.H. Protocatechualdehyde prevents methylglyoxal-induced mitochondrial dysfunction and AGEs-RAGE axis activation in human lens epithelial cells. *Eur. J. Pharmacol.* 2014, 738, 374–383. [CrossRef] [PubMed]

- 63. Antognelli, C.; Gambelunghe, A.; Muzi, G.; Talesa, V.N. Glyoxalase I drives epithelial-to-mesenchymal transition via argpyrimidine-modified Hsp70, miR-21 and SMAD signalling in human bronchial cells BEAS-2B chronically exposed to crystalline silica Min-U-Sil 5: Transformation into a neoplastic-like phenotype. *Free Radic. Biol. Med.* **2016**, *92*, 110–125. [CrossRef] [PubMed]
- 64. Dobler, D.; Ahmed, N.; Song, L.; Eboigbodin, K.E.; Thornalley, P.J. Increased dicarbonyl metabolism in endothelial cells in hyperglycemia induces anoikis and impairs angiogenesis by RGD and GFOGER motif modification. *Diabetes* **2006**, *55*, 1961–1969. [CrossRef] [PubMed]
- 65. Liu, H.; Zhang, N.; Tian, D. MiR-30b is involved in methylglyoxal-induced epithelial-mesenchymal transition of peritoneal mesothelial cells in rats. *Cell. Mol. Biol. Lett.* **2014**, *19*, 315–329. [CrossRef] [PubMed]
- 66. Hirahara, I.; Ishibashi, Y.; Kaname, S.; Kusano, E.; Fujita, T. Methylglyoxal induces peritoneal thickening by mesenchymal-like mesothelial cells in rats. *Nephrol. Dial. Transplant.* **2009**, *24*, 437–447. [CrossRef] [PubMed]
- 67. Navarrete Santos, A.; Jacobs, K.; Simm, A.; Glaubitz, N.; Horstkorte, R.; Hofmann, B. Dicarbonyls induce senescence of human vascular endothelial cells. *Mech. Ageing Dev.* **2017**, *166*, 24–32. [CrossRef] [PubMed]
- Kani, S.; Nakayama, E.; Yoda, A.; Onishi, N.; Sougawa, N.; Hazaka, Y.; Umeda, T.; Takeda, K.; Ichijo, H.; Hamada, Y.; et al. Chk2 kinase is required for methylglyoxal-induced G2/M cell-cycle checkpoint arrest: Implication of cell-cycle checkpoint regulation in diabetic oxidative stress signaling. *Genes Cells* 2007, 12, 919–928. [CrossRef] [PubMed]
- 69. Kang, Y.; Edwards, L.G.; Thornalley, P.J. Effect of methylglyoxal on human leukaemia 60 cell growth: Modification of DNA G1 growth arrest and induction of apoptosis. *Leuk. Res.* **1996**, *20*, 397–405. [CrossRef]
- 70. Rabbani, N.; Xue, M.; Thornalley, P.J. Dicarbonyls and glyoxalase in disease mechanisms and clinical therapeutics. *Glycoconj. J.* 2016, *33*, 513–525. [CrossRef] [PubMed]
- Zhang, S.; Liang, X.; Zheng, X.; Huang, H.; Chen, X.; Wu, K.; Wang, B.; Ma, S. Glo1 genetic amplification as a potential therapeutic target in hepatocellular carcinoma. *Int. J. Clin. Exp. Pathol.* 2014, *7*, 2079–2090. [PubMed]
- 72. Rabbani, N.; Xue, M.; Thornalley, P.J. Activity, regulation, copy number and function in the glyoxalase system. *Biochem. Soc. Trans.* **2014**, *42*, 419–424. [CrossRef] [PubMed]
- 73. Hu, X.; Yang, X.; He, Q.; Chen, Q.; Yu, L. Glyoxalase 1 is up-regulated in hepatocellular carcinoma and is essential for HCC cell proliferation. *Biotechnol. Lett.* **2014**, *36*, 257–263. [CrossRef] [PubMed]
- 74. Rabbani, N.; Thornalley, P.J. Glyoxalase 1 modulation in obesity and diabetes. *Antioxid. Redox Signal.* **2017**. [CrossRef] [PubMed]
- Geoffrion, M.; Du, X.; Irshad, Z.; Vanderhyden, B.C.; Courville, K.; Sui, G.; D'Agati, V.D.; Ott-Braschi, S.; Rabbani, N.; Thornalley, P.J.; et al. Differential effects of glyoxalase 1 overexpression on diabetic atherosclerosis and renal dysfunction in streptozotocin-treated, apolipoprotein E-deficient mice. *Physiol. Rep.* 2014, 2, e12043. [CrossRef] [PubMed]
- 76. Ahmed, U.; Dobler, D.; Larkin, S.J.; Rabbani, N.; Thornalley, P.J. Reversal of hyperglycemia-induced angiogenesis deficit of human endothelial cells by overexpression of glyoxalase 1 in vitro. *Ann. N. Y. Acad. Sci.* **2008**, *1126*, 262–264. [CrossRef] [PubMed]
- 77. Chiavarina, B.; Nokin, M.J.; Durieux, F.; Bianchi, E.; Turtoi, A.; Peulen, O.; Peixoto, P.; Irigaray, P.; Uchida, K.; Belpomme, D.; et al. Triple negative tumors accumulate significantly less methylglyoxal specific adducts than other human breast cancer subtypes. *Oncotarget* **2014**, *5*, 5472–5482. [CrossRef] [PubMed]
- Thornalley, P.J. Protecting the genome: Defence against nucleotide glycation and emerging role of glyoxalase I overexpression in multidrug resistance in cancer chemotherapy. *Biochem. Soc. Trans.* 2003, *31 Pt 6*, 1372–1377. [CrossRef] [PubMed]
- 79. Suzuki, K.; Koh, Y.H.; Mizuno, H.; Hamaoka, R.; Taniguchi, N. Overexpression of aldehyde reductase protects PC12 cells from the cytotoxicity of methylglyoxal or 3-deoxyglucosone. *J. Biochem.* **1998**, *123*, 353–357. [CrossRef] [PubMed]
- Feather, M.S.; Flynn, T.G.; Munro, K.A.; Kubiseski, T.J.; Walton, D.J. Catalysis of reduction of carbohydrate 2-oxoaldehydes (osones) by mammalian aldose reductase and aldehyde reductase. *Biochim. Biophys. Acta* 1995, 1244, 10–16. [CrossRef]
- Lee, H.M.; Seo, J.H.; Kwak, M.K.; Kang, S.O. Methylglyoxal upregulates *Dictyostelium discoideum* slug migration by triggering glutathione reductase and methylglyoxal reductase activity. *Int. J. Biochem. Cell Biol.* 2017, 90, 81–92. [CrossRef] [PubMed]

- 82. Baba, S.P.; Barski, O.A.; Ahmed, Y.; O'Toole, T.E.; Conklin, D.J.; Bhatnagar, A.; Srivastava, S. Reductive metabolism of AGE precursors: A metabolic route for preventing AGE accumulation in cardiovascular tissue. *Diabetes* **2009**, *58*, 2486–2497. [CrossRef] [PubMed]
- 83. Rulli, A.; Carli, L.; Romani, R.; Baroni, T.; Giovannini, E.; Rosi, G.; Talesa, V. Expression of glyoxalase I and II in normal and breast cancer tissues. *Breast Cancer Res. Treat.* **2001**, *66*, 67–72. [CrossRef] [PubMed]
- 84. Rulli, A.; Antognelli, C.; Prezzi, E.; Baldracchini, F.; Piva, F.; Giovannini, E.; Talesa, V. A possible regulatory role of 17beta-estradiol and tamoxifen on glyoxalase I and glyoxalase II genes expression in MCF7 and BT20 human breast cancer cells. *Breast Cancer Res. Treat.* **2006**, *96*, 187–196. [CrossRef] [PubMed]
- Cheng, W.; Tsai, M.M.; Tsai, C.Y.; Huang, Y.H.; Chen, C.Y.; Chi, H.C.; Tseng, Y.H.; Chao, I.W.; Lin, W.C.; Wu, S.M.; et al. Glyoxalase-I is a novel prognosis factor associated with gastric cancer progression. *PLoS ONE* 2012, 7, e34352. [CrossRef]
- Mearini, E.; Romani, R.; Mearini, L.; Antognelli, C.; Zucchi, A.; Baroni, T.; Porena, M.; Talesa, V.N. Differing expression of enzymes of the glyoxalase system in superficial and invasive bladder carcinomas. *Eur. J. Cancer* 2002, *38*, 1946–1950. [CrossRef]
- 87. Antognelli, C.; Baldracchini, F.; Talesa, V.N.; Costantini, E.; Zucchi, A.; Mearini, E. Overexpression of glyoxalase system enzymes in human kidney tumor. *Cancer J.* **2006**, *12*, 222–228. [CrossRef] [PubMed]
- Antognelli, C.; Del Buono, C.; Baldracchini, F.; Talesa, V.; Cottini, E.; Brancadoro, C.; Zucchi, A.; Mearini, E. Alteration of glyoxalase genes expression in response to testosterone in LNCaP and PC3 human prostate cancer cells. *Cancer Biol. Ther.* 2007, *6*, 1880–1888. [CrossRef] [PubMed]
- Talesa, V.N.; Ferri, I.; Bellezza, G.; Love, H.D.; Sidoni, A.; Antognelli, C. Glyoxalase 2 Is Involved in Human prostate Cancer Progression as Part of a Mechanism Driven by PTEN/PI3K/AKT/mTOR Signaling with Involvement of PKM2 and ERα. *Prostate* 2017, *77*, 196–210. [CrossRef] [PubMed]
- Antognelli, C.; Ferri, I.; Bellezza, G.; Siccu, P.; Love, H.D.; Talesa, V.N.; Sidoni, A. Glyoxalase 2 drives tumorigenesis in human prostate cells in a mechanism involving androgen receptor and p53-p21 axis. *Mol. Carcinog.* 2017, 56, 2112–2126. [CrossRef] [PubMed]
- 91. Ranganathan, S.; Ciaccio, P.J.; Walsh, E.S.; Tew, K.D. Genomic sequence of human glyoxalase-I: Analysis of promoter activity and its regulation. *Gene* **1999**, *240*, 149–155. [CrossRef]
- 92. Orso, F.; Corà, D.; Ubezio, B.; Provero, P.; Caselle, M.; Taverna, D. Identification of functional TFAP2A and SP1 binding sites in new TFAP2A-modulated genes. *BMC Genom.* **2010**, *11*, 355. [CrossRef] [PubMed]
- 93. Xue, M.; Rabbani, N.; Momiji, H.; Imbasi, P.; Anwar, M.M.; Kitteringham, N.; Park, B.K.; Souma, T.; Moriguchi, T.; Yamamoto, M.; et al. Transcriptional control of glyoxalase 1 by Nrf2 provides a stress-responsive defence against dicarbonyl glycation. *Biochem. J.* 2012, 443, 213–222. [CrossRef] [PubMed]
- 94. Conboy, C.M.; Spyrou, C.; Thorne, N.P.; Wade, E.J.; Barbosa-Morais, N.L.; Wilson, M.D.; Bhattacharjee, A.; Young, R.A.; Tavaré, S.; Lees, J.A.; et al. Cell cycle genes are the evolutionarily conserved targets of the E2F4 transcription factor. *PLoS ONE* **2007**, *2*, e1061. [CrossRef] [PubMed]
- De Hemptinne, V.; Rondas, D.; Toepoel, M.; Vancompernolle, K. Phosphorylation on Thr-106 and NO-modification of glyoxalase I suppress the TNF-induced transcriptional activity of NF-kappaB. *Mol. Cell. Biochem.* 2009, 325, 169–178. [CrossRef] [PubMed]
- 96. Van Herreweghe, F.; Mao, J.; Chaplen, F.W.; Grooten, J.; Gevaert, K.; Vandekerckhove, J.; Vancompernolle, K. Tumor necrosis factor-induced modulation of glyoxalase I activities through phosphorylation by PKA results in cell death and is accompanied by the formation of a specific methylglyoxal-derived AGE. *Proc. Natl. Acad. Sci. USA* 2002, *99*, 949–954. [CrossRef] [PubMed]
- 97. De Hemptinne, V.; Rondas, D.; Vandekerckhove, J.; Vancompernolle, K. Tumour necrosis factor induces phosphorylation primarily of the nitric-oxide-responsive form of glyoxalase I. *Biochem. J.* **2007**, 407, 121–128. [CrossRef] [PubMed]
- Birkenmeier, G.; Stegemann, C.; Hoffmann, R.; Günther, R.; Huse, K.; Birkemeyer, C. Posttranslational modification of human glyoxalase 1 indicates redox-dependent regulation. *PLoS ONE* 2010, 5, e10399. [CrossRef] [PubMed]
- 99. Xu, Y.; Chen, X. Glyoxalase II, a detoxifying enzyme of glycolysis byproduct methylglyoxal and a target of p63 and p73, is a pro-survival factor of the p53 family. *J. Biol. Chem.* **2006**, *281*, 26702–26713. [CrossRef] [PubMed]
- 100. Geng, X.; Ma, J.; Zhang, F.; Xu, C. Glyoxalase I in tumor cell proliferation and survival and as a potential target for anticancer therapy. *Oncol. Res. Treat.* **2014**, *37*, 570–574. [CrossRef] [PubMed]

- Sousa Silva, M.; Gomes, R.A.; Ferreira, A.E.; Ponces Freire, A.; Cordeiro, C. The glyoxalase pathway: The first hundred years ... and beyond. *Biochem. J.* 2013, 453, 1–15. [CrossRef] [PubMed]
- Rabbani, N.; Thornalley, P.J. Glyoxalase in diabetes, obesity and related disorders. *Semin. Cell Dev. Biol.* 2011, 22, 309–317. [CrossRef] [PubMed]
- 103. Xue, M.; Rabbani, N.; Thornalley, P.J. Glyoxalase in ageing. Semin. Cell Dev. Biol. 2011, 22, 293–301. [CrossRef] [PubMed]
- 104. Rabbani, N.; Thornalley, P.J. The glyoxalase system—From microbial metabolism, through ageing to human disease and multidrug resistance. *Semin. Cell Dev. Biol.* **2011**, *22*, 261. [CrossRef] [PubMed]
- 105. Ayoub, F.; Zaman, M.; Thornalley, P.; Masters, J. Glyoxalase activities in human tumour cell lines in vitro. *Anticancer Res.* **1993**, *13*, 151–155. [PubMed]
- 106. Di Ilio, C.; Angelucci, S.; Pennelli, A.; Zezza, A.; Tenaglia, R.; Sacchetta, P. Glyoxalase activities in tumor and non-tumor human urogenital tissues. *Cancer Lett.* **1995**, *96*, 189–193. [CrossRef]
- 107. Davidson, S.D.; Cherry, J.P.; Choudhury, M.S.; Tazaki, H.; Mallouh, C.; Konno, S. Glyoxalase I activity in human prostate cancer: A potential marker and importance in chemotherapy. J. Urol. 1999, 161, 690–691. [CrossRef]
- 108. Davidson, S.D.; Milanesa, D.M.; Mallouh, C.; Choudhury, M.S.; Tazaki, H.; Konno, S. A possible regulatory role of glyoxalase I in cell viability of human prostate cancer. *Urol. Res.* 2002, 30, 116–121. [CrossRef] [PubMed]
- 109. Baunacke, M.; Horn, L.C.; Trettner, S.; Engel, K.M.; Hemdan, N.Y.; Wiechmann, V.; Stolzenburg, J.U.; Bigl, M.; Birkenmeier, G. Exploring glyoxalase 1 expression in prostate cancer tissues: Targeting the enzyme by ethyl pyruvate defangs some malignancy-associated properties. *Prostate* 2014, 74, 48–60. [CrossRef] [PubMed]
- Samadi, A.A.; Fullerton, S.A.; Tortorelis, D.G.; Johnson, G.B.; Davidson, S.D.; Choudhury, M.S.; Mallouh, C.; Tazaki, H.; Konno, S. Glyoxalase I phenotype as a potential risk factor for prostate carcinoma. *Urology* 2001, 57, 183–187. [CrossRef]
- 111. Meehan, K.L.; Holland, J.W.; Dawkins, H.J. Proteomic analysis of normal and malignant prostate tissue to identify novel proteins lost in cancer. *Prostate* 2002, *50*, 54–63. [CrossRef] [PubMed]
- 112. Chavan, S.V.; Chavan, N.R.; Balaji, A.; Trivedi, V.D.; Chavan, P.R. A pilot study on the use of serum glyoxalase as a supplemental biomarker to predict malignant cases of the prostate in the PSA range of 4–20 ng/mL. *Indian J. Med. Res.* 2011, 134, 458–462. [PubMed]
- 113. Antognelli, C.; Mezzasoma, L.; Mearini, E.; Talesa, V.N. Glyoxalase 1-419C>A variant is associated with oxidative stress: Implications in prostate cancer progression. *PLoS ONE* **2013**, *8*, e74014. [CrossRef] [PubMed]
- 114. Catalona, W.J.; Southwick, P.C.; Slawin, K.M.; Partin, A.W.; Brawer, M.K.; Flanigan, R.C.; Patel, A.; Richie, J.P.; Walsh, P.C.; Scardino, P.T.; et al. Comparison of percent free PSA, PSA density, and age-specific PSA cutoffs for prostate cancer detection and staging. *Urology* 2000, *56*, 255–260. [CrossRef]
- 115. Talesa, V.N.; Antognelli, C.; Del Buono, C.; Stracci, F.; Serva, M.R.; Cottini, E.; Mearini, E. Diagnostic potential in prostate cancer of a panel of urinary molecular tumor markers. *Cancer Biomark.* 2009, *5*, 241–251. [CrossRef] [PubMed]
- 116. Mearini, E.; Antognelli, C.; Del Buono, C.; Cochetti, G.; Giannantoni, A.; Nardelli, E.; Talesa, V.N. The combination of urine DD3(PCA3) mRNA and PSA mRNA as molecular markers of prostate cancer. *Biomarkers* 2009, 14, 235–243. [CrossRef] [PubMed]
- 117. Sharkey, E.M.; O'Neill, H.B.; Kavarana, M.J.; Wang, H.; Creighton, D.J.; Sentz, D.L.; Eiseman, J.L. Pharmacokinetics and antitumor properties in tumor-bearing mice of an enediol analogue inhibitor of glyoxalase I. *Cancer Chemother. Pharmacol.* 2000, 46, 156–166. [CrossRef] [PubMed]
- 118. Finkelstein, M.P.; Aynehchi, S.; Samadi, A.A.; Drinis, S.; Choudhury, M.S.; Tazaki, H.; Konno, S.J. Chemosensitization of carmustine with maitake beta-glucan on androgen-independent prostatic cancer cells: Involvement of glyoxalase I. J. Altern. Complement. Med. 2002, 8, 573–580. [CrossRef] [PubMed]
- 119. Santel, T.; Pflug, G.; Hemdan, N.Y.; Schäfer, A.; Hollenbach, M.; Buchold, M.; Hintersdorf, A.; Lindner, I.; Otto, A.; Bigl, M.; et al. Curcumin inhibits glyoxalase 1: A possible link to its anti-inflammatory and anti-tumor activity. *PLoS ONE* 2008, *3*, e3508. [CrossRef] [PubMed]
- Valenti, D.; Vacca, R.A.; de Bari, L. 3-Bromopyruvate induces rapid human prostate cancer cell death by affecting cell energy metabolism, GSH pool and the glyoxalase system. *J. Bioenerg. Biomembr.* 2015, 47, 493–506. [CrossRef] [PubMed]

- 121. Balistreri, C.R.; Carruba, G.; Calabrò, M.; Campisi, I.; Di Carlo, D.; Lio, D.; Colonna-Romano, G.; Candore, G.; Caruso, C. CCR5 proinflammatory allele in prostate cancer risk: A pilot study in patients and centenarians from Sicily. *Ann. N. Y. Acad. Sci.* **2009**, 1155, 289–292. [CrossRef] [PubMed]
- Rubin, M.A.; Maher, C.A.; Chinnaiyan, A.M. Common gene rearrangements in prostate cancer. J. Clin. Oncol. 2011, 29, 3659–3668. [CrossRef] [PubMed]
- 123. Fabris, L.; Ceder, Y.; Chinnaiyan, A.M.; Jenster, G.W.; Sorensen, K.D.; Tomlins, S.; Visakorpi, T.; Calin, G.A. The potential of MicroRNAs as prostate cancer biomarkers. *Eur. Urol.* 2016, 70, 312–322. [CrossRef] [PubMed]
- 124. Sailer, V.; Stephan, C.; Wernert, N.; Perner, S.; Jung, K.; Dietel, M.; Kristiansen, G. Comparison of p40 (ΔNp63) and p63 expression in prostate tissues-which one is the superior diagnostic marker for basal cells? *Histopathology* 2013, 63, 50–56. [CrossRef] [PubMed]
- 125. Valentine, W.N.; Paglia, D.E.; Neerhout, R.C.; Konrad, P.N. Erythrocyte glyoxalase II deficiency with coincidental hereditary elliptocytosis. *Blood* **1970**, *36*, 797–808. [PubMed]
- 126. Sousa Silva, M.; Ferreira, A.E.; Gomes, R.; Tomás, A.M.; Ponces Freire, A.; Cordeiro, C. The glyoxalase pathway in protozoan parasites. *Int. J. Med. Microbiol.* **2012**, *302*, 225–229. [CrossRef] [PubMed]
- 127. Tanaka, T.; Kuramitsu, Y.; Wang, Y.; Baron, B.; Kitagawa, T.; Tokuda, K.; Hirakawa, K.; Yashiro, M.; Naito, S.; Nakamura, K. Glyoxalase 1 as a candidate for indicating the metastatic potential of of SN12Chuman renal cell carcinoma cell clones. *Oncol. Rep.* **2013**, *30*, 2365–2370. [CrossRef] [PubMed]
- 128. Chocholatý, M.; Jáchymová, M.; Schmidt, M.; Havlová, K.; Křepelová, A.; Zima, T.; Babjuk, M.; Kalousová, M. Polymorphisms of the receptor for advanced glycation end-products and glyoxalase I in patients with renal cancer. *Tumour Biol.* **2015**, *36*, 2121–2126. [CrossRef] [PubMed]
- 129. Lin, S.Y.; Linehan, J.A.; Wilson, T.G.; Hoon, D.S.B. Emerging Utility of Urinary Cell-free Nucleic Acid Biomarkers for Prostate, Bladder, and Renal Cancers. *Eur. Urol. Focus* **2017**. [CrossRef] [PubMed]
- 130. Truta, A.; Popon, T.A.H.; Saraci, G.; Ghervan, L.; Pop, I.V. Health Related Quality of life in bladder cancer. Current approach and future perspectives. *Clujul Med.* **2017**, *90*, 262–267. [CrossRef] [PubMed]
- 131. Costa, A.L.; Lobo, J.; Jerónimo, C.; Henrique, R. The epigenetics of testicular germ cell tumors: Looking for novel disease biomarkers. *Epigenomics* **2017**, *9*, 155–169. [CrossRef] [PubMed]
- Chieffi, P. An Overview on Predictive Biomarkers of Testicular Germ Cell Tumors. J. Cell. Physiol. 2017, 232, 276–280. [CrossRef] [PubMed]



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