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Asymmetric Vinylogous Aldol-type Reactions of Aldehydes with Allyl Phosphonate and Sulfone

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SUMMARY

Two catalytic asymmetric vinylogous aldol-type reactions of aldehydes with allyl phosphonate and allyl sulfone have been uncovered in good to high yields for the first time. The bulky ligand—(R)-DTBM-SEGPHOS—was found to be the key to perfectly control both regio- and enantioselectivities. Transformations of the vinylogous products (including Horner-Wadsworth-Emmons and Julia olefinations) were successfully realized by virtue of the phosphonate and sulfone moieties. Moreover, the present methodology was successfully applied in the asymmetric synthesis of natural products.

INTRODUCTION

Asymmetric vinylogous aldol reaction (VAR) has been one of the most important reactions in the synthesis of complex natural products, especially in the synthesis of polyketides. In the past decades, catalytic asymmetric VAR of aldehydes or ketones has evolved from classical Mukaiyama vinylogous aldol reaction (Denmark et al., 2005; Casiraghi et al., 2011; Pansare and Paul, 2011; Bisai, 2012; Kalesse et al., 2014; Hosokawa, 2018a, 2018b) to direct vinylogous aldol reaction (DVAR) (Li and Yin, 2018; Otsuka et al., 2013; Zhu et al., 2013; Li et al., 2014; Han and Chang, 2016; Jing et al., 2016; Ray and Mukherjee, 2018), which enjoys the advantages of easy reaction protocol and high atom economy (Trost, 1991; Anastas and Crabtree, 2009; Newhouse et al., 2009). However, the nucleophiles in DVAR were mainly limited to unsaturated carbonyl compounds and their close derivatives (Bai et al., 2017). Unsaturated phosphonates or phosphine oxides, as well as unsaturated sulfones, have never been investigated as prenucleophiles in vinylogous aldol-type reactions.

In such reactions, synthetically versatile chiral vinylogous products containing α , β -unsaturated phosphonate or phosphine oxide moiety or α , β -unsaturated sulfone motif (Nishida et al., 2008; Xue et al., 2011; Konno et al., 2012; Lefevre et al., 2013; Hornillos et al., 2015; Lim and Hayashi, 2015, 2017; Wang and Hayashi, 2018; El-Awa et al., 2009; Alba et al., 2010; Nielsen et al., 2010; Zhu and Lu, 2010; Moteki et al., 2010; Quintard et al., 2011; Moure et al., 2011; Nishimura et al., 2012; Halskov et al., 2012; Zhou et al., 2012; Hernández-Toribio et al., 2012), would be produced. Furthermore, phosphonates and phosphine oxides had great applications in medicinal and agricultural chemistry (Mucha et al., 2011; Ordóñez et al., 2012; Corbridge, 2013; Horsman and Zechel, 2017). Sulfones, especially α , β -unsaturated sulfones, were widely distributed in biologically active compounds, even in the commercial pharmaceuticals (Meadows and Gervay-Hague, 2006; Dunny et al., 2013; Woo et al., 2014; Fang et al., 2016). Therefore it is highly desirable to achieve a vinylogous aldol-type reaction of unsaturated phosphonates or phosphine oxides, as well as unsaturated sulfones.

One inherent difficulty faced in the vinylogous aldol-type reaction of allyl phosphonate or allyl sulfone is the control of regioselectivity. The addition of allyl phosphonate to aldehyde was investigated by Yuan and co-workers in detail (Scheme 1A) (Yuan et al., 1990, 1991). Treating allyl phosphonate with ⁿBuLi in tetrahydrofuran (THF) at -70° C afforded the corresponding delocalized allylic carbanion, which reacted with benzaldehyde to give a mixture of α - and γ -adducts. α -Addition was the natural tendency and thus dominated the addition pathways, which led to the vinylogous product (γ -adduct) in significantly low yield. Increasing the steric hindrance of the alkyl group in phosphonate only led to a slight improvement of the γ -selectivity.

Furthermore, the addition of allyl sulfone to benzaldehyde catalyzed by a phosphine-based strong base was studied by Verkade and co-workers, which delivered the α -adduct in 63% yield at -78°C (Scheme 1B) (Kisanga and Verkade, 2002). The same reaction promoted by stoichiometric ⁿBuLi also afforded the

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https://doi.org/10.1016/j.isci. 2019.03.010 A Reported Addition of Allyl Phosphonate to Benzaldehyde Dominated by α-Selectivity

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B Reported Addition of Allyl Sulfone to Benzaldehyde Dominated by α-Selectivity



c This Work: Catalytic Asymmetric Vinylogous Aldol-Type Reactions of Allyl Phosphonate and Allyl Sulfone (γ-Selectivity)



D Selected Natural Products Synthesized with Horner-Wadsworth-Emmons (HWE) Olefination or Julia Olefination



Scheme 1. Aldol-type Reactions of Aldehydes with Allyl Phosphonate and Sulfone and Selected Natural Products Synthesized with Horner-Wadsworth-Emmons and Julia Olefinations

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 α -adduct exclusively, which was employed to prepare acyclic 2-phenylsulfonyl 1,3-diene in 89% yield (Cuvigny et al., 1983, 1986; Chinkov et al., 2003). Obviously, it is challenging to overcome the inherent α -selectivity in the aldol-type reactions of allyl phosphonate and allyl sulfone. The other difficulty in the DVAR is the remote asymmetric induction as the functional group (aldehyde, ketone, ester, and amide generally, and phosphonate or sulfone here) is far from the reactive γ -position in form, which was viewed as a challenge in asymmetric catalysis (Shirokawa et al., 2004). To the best of our knowledge, there is no reported enantioselective method to carry out a vinylogous aldol-type reaction of allyl phosphonate or allyl sulfone (Scheme 1C).

The α -carbanions of phosphonate and sulfone have been employed as nucleophiles in Horner-Wadsworth-Emmons (HWE) and Julia olefinations, which were identified as two prominent synthetic tools to assemble the carbon-carbon double bond in organic synthesis, especially in the total synthesis of complex natural products (Kobayashi et al., 2018; Ma et al., 2010) (such as callipeltoside A, dictyostatin, indolizomycin and ambruticin) (Scheme 1D) (Trost et al., 2002; Ho et al., 2013; Kim et al., 1993; Liu and Jacobson, 2001). The products from the saturation of vinylogous aldol-type products (3/5) would be suitable substrates for HWE and Julia olefinations for further structure elaboration. Herein, we report two asymmetric vinylogous aldol-type reactions of aldehydes with allyl phosphonate and allyl sulfone catalyzed by a bulky chiral copper(I) complex and an organic base. The deprotonation of allyl phosphonate or allyl sulfone would generate a nucleophilic allylcopper(I) species, which afforded the vinylogous product through an asymmetric allylation via a six-member ring transition state (Scheme 1C).

RESULTS AND DISCUSSION

The reaction of allyl sulfone 4'/4 and benzaldehyde (2a) was investigated as a model reaction in the presence of $Cu(CH_3CN)_4PF_6$, phosphine ligand, and Barton's base (Table 1). In all cases with 4', the vinylogous product 5a' was obtained with unsatisfactory regio- and enantioselectivities (entries 1–9). (*R*)-DTBM-SEGPHOS, an effective bulky ligand, led to good control of the enantioselectivity in our previously reported catalytic asymmetric aldol reaction of unsaturated esters and aldehydes (Zhang and Yin, 2018) and gave excellent control of the regioselectivity. The enantioselectivity was improved from 43% to 76% by switching phenyl to 2-pyridinyl (entry 10). Lowering the temperature to $-40^{\circ}C$ resulted in 97% enantiomeric excess (ee) for 5a (entry 11). The moderate yield was enhanced to 98% by changing the ratio of 2a/4, increasing the amount of Barton's base to 30 mol %, and prolonging the reaction time to 36 h (entries 12–14). Finally, the catalyst loading was successfully decreased to 3 mol % without changing both regio- and enantioselectivities (entry 15).

By modifying the optimized reaction conditions for **4** (3 equiv. **1**, 20 mol % Barton's base, and -10° C), the substrate scope of aldehydes **2** in the reaction with allyl phosphonate **1** was studied (Table 2). Aromatic aldehydes with electron-withdrawing groups or electron-donating groups were competent substrates to generate the corresponding vinylogous products uniformly in good yields with both excellent regioselectivity (>20/1) and excellent enantioselectivity (>95% ee) (3a-3o). Moreover, the reaction was not sensitive to the position of a substituent on the phenyl ring of the aromatic aldehydes. Even the sterically congested *ortho*-CF₃-benzaldehyde afforded the product **31** in 91% yield with 95% ee. 1-Naphthaldehyde was also an excellent substrate (**3p**). Moreover, the present reaction conditions were applicable to various heteroaromatic aldehydes (**3q-3x**). Although the yields were moderate in some cases, both regio- and enantioselectivities were excellent. Particularly noteworthy are the aldehydes containing a pyridine motif (**3t**) and a carbazole motif (**3x**) as these functional groups potentially can coordinate to the metal center and thus deactivate the catalyst.

 α , β -Unsaturated aldehydes also served as suitable substrates (**3y-3ai**). Aryls, heteroaryls, alkyls, and vinyls with substituent were well tolerated at the β -position of the α , β -unsaturated aldehydes. Moreover, functional groups, such as alkyl chloride (**3af**), *tert*-butyldimethylsilyl (TBS)-ether (**3ag**), alkynyl (**3ah**), and prenyl (**3ai**), remained intact in the present reaction conditions. These functional groups offer the opportunity for further structure elaboration. It is noteworthy that acrolein (**2ab**), susceptible to conjugate addition, served as a suitable substrate to give the vinylogous product **3ab** in moderate yield with excellent enantioselectivity. The chiral aldehydes, including α , β -unsaturated aldehyde **2aj** derived from (–)-citronellal, (–)-perillaldehyde (**2ak**), and (–)-myrtenal (**2al**), were also investigated with both (*R*)-DTBM-SEGPHOS and (*S*)-DTBM-SEGPHOS. In both cases, the products (**3aj**, **3aj**', **3ak**, **3ak**', **3al**, and **3al**') were obtained in good yields with excellent diastereoselectivity, which indicated that the asymmetric



Table 1. Optimization of the Reaction Conditions^a



 $\begin{array}{ll} \mbox{Ar}=\mbox{Ph}, \mbox{ (R)-} & \mbox{R}=\mbox{H}, \mbox{Ar}=\mbox{Ph}, \mbox{(R)-QUINAP}\\ \mbox{BINAP}; & \mbox{R}=\mbox{H}, \mbox{Ar}=\mbox{3,5-}^{r}\mbox{Bu}_{2}\mbox{-4-OMe-Ph}, \\ \mbox{Ar}=\mbox{Tol}, \mbox{(R)-} & \mbox{(R)-DTBM-SEGPHOS}\\ \mbox{Tol}-\mbox{BINAP} & \mbox{(R)-}\mbox{DTBM-SEGPHOS}\\ \end{array}$



(R,R)-Ph-BPE

HPLC, high-performance liquid chromatography; NMR, nuclear magnetic resonance

^a4/4', 0.1 mmol; 2a, 0.2 mmol.

^bDetermined by ¹H NMR analysis of reaction crude mixture using mesitylene as an internal standard.

 $^{\rm c}\mbox{Determined}$ by chiral-stationary-phase HPLC analysis.

^d**4**. 0.2 mmol; **2a**. 0.1 mmol.

^e36 h.

 ${}^{f}Cu(CH_{3}CN)_{4}PF_{6}$ and ligand, 3 mol %, Barton's base = $2 - {}^{t}Bu - 1, 1, 3, 3$ -tetramethylguanidine.

introduction was dominated by the copper(I)-catalyst. The absolute configuration of **3a** was determined to be *R* by transforming it to a reported compound (for details, see Supplemental Information). The absolute configurations of other products were tentatively assigned by analogy.





Table 2. Substrate Scope of Aldehydes in the Reaction with 1^a

(Continued on next page)



Table 2. Continued

HPLC, high-performance liquid chromatography; NMR, nuclear magnetic resonance

^a2, 0.3 mmol; 1, 0.9 mmol. Isolated yield was reported. Regio- and diastereoselectivity were determined by ¹H NMR analysis of reaction crude mixture. Enantioselectivity was determined by chiral-stationary-phase HPLC analysis.

^bGram-scale synthesis.

^c(S)-DTBM-SEGPHOS was used.

Moreover, the substrate scope of aldehydes in the reaction with allyl sulfone 4 was evaluated (Table 3). Various aromatic aldehydes with a substituent at ortho-, meta-, or para-position were suitable substrates. Both 1-naphthyl and 2-naphthyl aldehydes were well applicable. The vinylogous products (5a-5h, 5k, 5l, 5n, 50, 5p, and 5am-5aq) were isolated in moderate to excellent yields with excellent regio- and enantioselectivities. Heteroaromatic aldehydes also served as competent substrates without compromising enantioselectivity (5q, 5r, 5u, 5x, 5ar and 5as). As for α , β -unsaturated aldehydes, aryls, heteroaryls, vinyls with substituent, and alkyls were accepted at β -position (5y, 5z, 5aa, 5ad, 5ae, 5at, 5au, and 5ah). Moreover, the reaction conditions were successfully applied to chiral natural products bearing α , β -unsaturated aldehyde moiety, such as (-)-perillaldehyde (2ak) and (-)-myrtenal (2al). The corresponding vinylogous products (5ak, 5ak', 5al, and 5al') were obtained in moderate yields with high diastereoselectivity. It is evident that in the case of (-)-myrtenal with (S)-DTBM-SEGPHOS, mismatch phenomenon was observed. The absolute configuration of 5a was assigned to be R by its transformation to a known compound (for details, see Supplemental Information). Analogically, the stereochemistry of other products was dictated tentatively. It should be pointed out that the gram-scale syntheses of both 3a and 5a were successfully carried out with constant results. Moreover, it should be mentioned that aliphatic aldehydes afforded α -adducts mainly in low yields at the present reaction conditions, which is a limitation of the present reactions. However, the vinylogous products of aliphatic aldehydes could be potentially accessed by the transformations of vinylogous products of various α , β -unsaturated aldehydes by means of the carbon-carbon double bond.

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Table 3. Substrate Scope of Aldehydes in the Reaction with 4^a

(Continued on next page)



Table 3. Continued

HPLC, high-performance liquid chromatography; NMR, nuclear magnetic resonance

^a2, 0.3 mmol; 4, 0.6 mmol. Isolated yield was reported. Regio- and diastereoselectivity were determined by ¹H NMR analysis of reaction crude mixture. Enantioselectivity was determined by chiral-stationary-phase HPLC analysis.

^bGram-scale synthesis.

^c(S)-DTBM-SEGPHOS was used.

To get insights into the mechanism, $rac \cdot \alpha \cdot 3a$ and $rac \cdot \alpha \cdot 5a$ (racemic α -adducts) prepared according to a reported and a modified reaction procedure (for details, see Supplemental Information) were subjected to the standard reaction conditions, respectively (Scheme 2A). It was found that $rac \cdot \alpha \cdot 3a$ was completely consumed and 3a was observed in 40% yield with 99% ee, together with benzaldehyde (2a), allyl phosphonate 1, and α,β -unsaturated phosphonate 6. These results clearly indicated that *retro*-aldol reaction of $rac \cdot \alpha \cdot 3a$ proceeded to afford benzaldehyde (2a) and allyl phosphonate 1. One portion of allyl phosphonate 1 reacted with benzaldehyde (2a) to give the vinylogous product 3a in 40% yield with 99% ee in the presence of 5 mol % copper(I) catalyst and 20 mol % Barton's base. One portion of allyl phosphonate 1 isomerized to α,β -unsaturated phosphonate 6, whereas the other portion of allyl phosphonate 1 remained. The same tendency was also observed in the *retro*-aldol reaction of $rac \cdot \alpha \cdot 5a$. This phenomenon indicated that significantly reversible α -addition led to the transformation of α -adducts to γ -adducts, which finally led to excellent control of the regioselectivity.

Rac-3a and *rac-5a* prepared by $Cu(CH_3CN)_4PF_6$ -*rac-DTBM-SEGPHOS-catalyzed* reactions were also submitted to the standard reaction conditions, respectively (Scheme 2B). Thin-layer chromatography, ¹H nuclear magnetic resonance, and chiral high-performance liquid chromatographic analyses of the reaction crude mixtures indicated that slow and inefficient *retro*-vinylogous additions occurred, as **3a** was obtained in 83% yield with -18% ee, whereas **5a** was generated in 70% yield with -8% ee. It was obvious that the *retro*-vinylogous aldol reactions of both (*R*)-**3a** and (*R*)-**5a** proceeded selectively, which resulted in the slight enrichment of (S)-**3a** and (S)-**5a** in the reaction mixtures. However, these *retro*-vinylogous aldol reactions were very slow and inefficient, which would not have detrimental effect on the enantioselectivity in the catalytic asymmetric vinylogous aldol-type reactions of allyl phosphonate **1** and allyl sulfone **4**. Based on these important experimental observations and literatures (Bazán-Tejeda et al., 2006; Yamaguchi et al., 2007; Bouaouli et al., 2018), a possible reaction pathway was proposed in Supplemental Information.

The transformations of the vinylogous aldol products (**3a** and **5a**) were carried out as shown in Scheme 3. The cleavage of unsaturated double bond in **3a** was easily achieved through ozonolysis to deliver diol **8** in 67% yield after the reduction of generated aldehyde moiety with NaBH₄. After being protected as TBS-ether, **3a** was reduced to phosphonate **10** with H₂ in the presence of Pd/C. **10** Was easily transformed to α , β -unsaturated compounds **11** and **12** via α -functionalization and subsequent HWE olefination. The sulfone moiety in **5a** was successfully removed without touching the double bond to afford ester **13** in 52% yield after the protection of the alcohol motif. Sulfone **15** was easily accessed through the reduction of the unsaturated double bond and the protection of the hydroxyl group in **5a**, which was transformed to olefin **16** in 67% yield in two steps by α -functionalization and the following removal of the sulfone group. Moreover, chiral diol **18** was synthesized from **5a** in 74% yield with >20/1 diastereoisomeric ratio (dr) in two steps through intramolecular oxo-Michael addition and the



A Experimental Insights into the Retro-Aldol Reaction of α-Adducts





Scheme 2. Trials for retro-Aldol Reactions of α-Adducts and γ-Adducts

cleavage of the generated acetal motif. Furthermore, the synthetic utilities of the present methodology were showcased by its applications in the asymmetric synthesis of yashabushidiol B and the formal asymmetric synthesis of (+)-cryptocaryalactone (for the details, see Supplemental Information). Moreover, our synthetic route provided a straightforward method for the asymmetric synthesis of various chiral diols **19**. Some of the diols **19** (both natural and man-made) exhibited significant anti-proliferative activity on some human cancer cell lines (Narasim-hulu et al., 2009; Yokosuka et al., 2002).

Limitations of Study

Aliphatic aldehydes were not applicable in the present reactions as α -adducts were obtained in low yields and no vinylogous products were generated. Fortunately, α , β -unsaturated aldehydes served as competent substrates and their vinylogous products could be potentially converted to the vinylogous products of aliphatic aldehydes through the transformations of carbon-carbon double bond.

Conclusion

In summary, two copper(I)-(*R*)-DBTM-SEGPHOS complex-catalyzed asymmetric vinylogous aldol-type reactions of aldehydes with allyl phosphonate and allyl sulfone were disclosed. These two reactions enjoyed advantages of 100% atomic economy, mild reaction conditions, easy reaction protocol, broad substrate scope, excellent regioselectivity, and excellent enantioselectivity. The mechanistic studies revealed a

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significantly reversible α -addition process and a slightly reversible γ -addition process, which accounted for the perfect control of the regioselectivity in these two vinylogous aldol-type reactions. Finally, various transformations of the vinylogous products (including HWE and Julia olefinations) were successfully carried out by means of phosphonate and sulfone. Application of the present methodology in the asymmetric synthesis of complex natural products is currently on the way in our laboratory.

METHODS

All methods can be found in the accompanying Transparent Methods supplemental file.

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SUPPLEMENTAL INFORMATION

Supplemental Information can be found online at https://doi.org/10.1016/j.isci.2019.03.010.

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AUTHOR CONTRIBUTIONS

L.Y. conceived the project and designed the experiments. W.-J.Y. and C.-Y.Z. performed and analyzed the experiments. L.Y. wrote the manuscript. W.-J.Y. wrote the Supplemental Information and contributed other related materials. All the authors discussed the results and commented on the manuscript.

DECLARATION OF INTERESTS

The authors declare no competing interests.

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REFERENCES

Alba, A.-N.R., Companyó, X., and Rios, R. (2010). Sulfones: new reagents in organocatalysis. Chem. Soc. Rev. *39*, 2018–2033.

Anastas, P.T., and Crabtree, R.H. (2009). Handbook of Green Chemistry-Green Catalysis (Wiley-VCH: Weinheim).

Bai, X., Zeng, G., Shao, T., and Jiang, Z. (2017). Catalytic enantioselective γ-selective additions of 2-allylazaarenes to activated ketones. Angew. Chem. Int. Ed. 56, 3684–3688.

Bazán-Tejeda, B., Bluet, G., Broustal, G., and Campagne, J.-M. (2006). α,β-Unsaturated δ-lactones from copper-catalyzed asymmetric vinylogous Mukaiyama reactions of aldehydes: scope and mechanistic insights. Chem. Eur. J. 12, 8358–8366.

Bisai, V. (2012). Organocatalytic asymmetric vinylogous aldol reactions. Synthesis 44, 1453–1463.

Bouaouli, S., Spielmann, K., Vrancken, E., Campagne, J.-M., and Gérard, H. (2018). Mechanism of enolate transfer between Si and Cu. Chem. Eur. J. 24, 6617–6624.

Casiraghi, G., Battistini, L., Curti, C., Rassu, G., and Zanardi, F. (2011). The vinylogous aldol and related reactions: ten years of progress. Chem. Rev. 111, 3076–3154.

Chinkov, N., Majumdar, S., and Marek, I. (2003). Stereoselective preparation of dienyl zirconocene complexes via a tandem allylic C–H bond activation-elimination sequence. J. Am. Chem. Soc. 125, 13258–13264.

Corbridge, D.E.C. (2013). Phosphorus: Chemistry, Biochemistry and Technology, Sixth Edition (CRC Press). Cuvigny, T., Hervé du Penhoat, C., and Julia, M. (1983). Synthesis with sulfones (XXX): stereoselective synthesis of arenesulfonyl-1,3dienes. Tetrahedron Lett. 24, 4315–4318.

Cuvigny, T., Hervé du Penhoat, C., and Julia, M. (1986). Syntheses with sulfones XLVI: stereoselective preparation of 2-benzenesulfonyl-1,3-dienes and 2-benzenesulfonyl-1,4-dienes. Tetrahedron 42, 5329–5336.

Denmark, S.E., Heemstra, J.R., Jr., and Beutner, G.L. (2005). Catalytic, enantioselective, vinylogous aldol reactions. Angew. Chem. Int. Ed. 44, 4682–4698.

Dunny, E., Doherty, W., Evans, P., Malthouse, J.P.G., Nolan, D., and Knox, A.J.S. (2013). Vinyl sulfone-based peptidomimetics as antitrypanosomal agents: design, synthesis, biological and computational evaluation. J. Med. Chem. 56, 6638–6650.

El-Awa, A., Noshi, M.N., du Jourdin, X.M., and Fuchs, P.L. (2009). Evolving organic synthesis fostered by the pluripotent phenylsulfone moiety. Chem. Rev. 109, 2315–2349.

Fang, Y., Luo, Z., and Xu, X. (2016). Recent advances in the synthesis of vinyl sulfones. RSC Adv. *6*, 59661–59676.

Halskov, K.S., Johansen, T.K., Davis, R.L., Steurer, M., Jensen, F., and Jørgensen, K.A. (2012). Cross-trienamines in asymmetric organocatalysis. J. Am. Chem. Soc. 134, 12943– 12946.

Han, J.-L., and Chang, C.-H. (2016). An asymmetric assembly of spirooxindole dihydropyranones through a direct enantioselective organocatalytic vinylogous aldol-cyclization cascade reaction of 3-alkylidene oxindoles with isatins. Chem. Commun. (Camb.) 52, 2322–2325.

Hernández-Toribio, J., Padilla, S., Adrio, J., and Carretero, J.C. (2012). Catalytic asymmetric synthesis of α -quaternary proline derivatives by 1,3-dipolar cycloaddition of α -silylimines. Angew. Chem. Int. Ed. 51, 8854–8858.

Ho, S., Bucher, C., and Leighton, J.L. (2013). A highly step-economical synthesis of dictyostatin. Angew. Chem. Int. Ed. *52*, 6757–6761.

Hornillos, V., Vila, C., Otten, E., and Feringa, B.L. (2015). Catalytic asymmetric synthesis of phosphine boronates. Angew. Chem. Int. Ed. 54, 7867–7871.

Horsman, G.P., and Zechel, D.L. (2017). Phosphonate biochemistry. Chem. Rev. 117, 5704–5783.

Hosokawa, S. (2018a). Remote asymmetric induction reactions using a *E*,*E*-vinylketene silyl *N*,*O*-acetal and the wide range stereocontrol strategy for the synthesis of polypropionates. Acc. Chem. Res. *51*, 1301–1314.

Hosokawa, S. (2018b). Recent development of vinylogous Mukaiyama aldol reactions. Tetrahedron Lett. *59*, 77–88.

Jing, Z., Bai, X., Chen, W., Zhang, G., Zhu, B., and Jiang, Z. (2016). Organocatalytic enantioselective vinylogous aldol reaction of allyl aryl ketones to activated acyclic ketones. Org. Lett. 18, 260–263.

Kalesse, M., Cordes, M., Symkenberg, G., and Lu, H.-H. (2014). The vinylogous Mukaiyama aldol reaction (VMAR) in natural product synthesis. Nat. Prod. Rep. 31, 563–594.



Kim, G., Chu-Moyer, M.Y., Danishefsky, S.J., and Schulte, G.K. (1993). The total synthesis of indolizomycin. J. Am. Chem. Soc. *115*, 30–39.

Kisanga, P.B., and Verkade, J.G. (2002). $P(RNCH_2CH_{2)3}N$ -Catalyzed 1,2-addition reactions of activated allylic synthons. J. Org. Chem. 67, 426–430.

Kobayashi, K., Tanaka, K., III, and Kogen, H. (2018). Recent topics of the natural product synthesis by Horner-Wadsworth-Emmons reaction. Tetrahedron Lett. *59*, 568–582.

Konno, T., Shimizu, K., Ogata, K., and Fukuzawa, S. (2012). Rhodium-catalyzed enantioselective hydrogenation of unsaturated phosphonates by Click-Ferrophos ligands. J. Org. Chem. 77, 3318– 3324.

Lefevre, N., Brayer, J.-L., Folléas, B., and Darses, S. (2013). Chiral α -amino phosphonates via rhodium-catalyzed asymmetric 1,4-addition reactions. Org. Lett. 15, 4274–4276.

Li, H., and Yin, L. (2018). Recent progress on direct catalytic asymmetric vinylogous reactions. Tetrahedron Lett. *59*, 4121–4135.

Li, T.-Z., Jiang, Y., Guan, Y.-Q., Sha, F., and Wu, X.-Y. (2014). Direct enantioselective vinylogous aldol-cyclization cascade reaction of allyl pyrazoleamides with isatins: asymmetric construction of spirocyclic oxindoledihydropyranones. Chem. Commun. (Camb.) *50*, 10790–10792.

Lim, K.M.-H., and Hayashi, T. (2015). Rhodiumcatalyzed asymmetric arylation of allyl sulfones under the conditions of isomerization into alkenyl sulfones. J. Am. Chem. Soc. 137, 3201–3204.

Lim, K.M.-H., and Hayashi, T. (2017). Dynamic kinetic resolution in rhodium-catalyzed asymmetric arylation of phospholene oxides. J. Am. Chem. Soc. 139, 8122–8125.

Liu, P., and Jacobson, E.N. (2001). Total synthesis of (+)-Ambruticin. J. Am. Chem. Soc. *123*, 10772–10773.

Ma, J.-H., Wang, F., Wang, J.-X., and You, Q.-D. (2010). Progress of Julia olefination in total synthesis of natural products. Chin. J. Org. Chem. 30, 1615–1623.

Meadows, D.C., and Gervay-Hague, J. (2006). Vinyl sulfones: synthetic preparations and medicinal chemistry applications. Med. Res. Rev. 26, 793–814.

Moteki, S.A., Xu, S., Arimitsu, S., and Maruoka, K. (2010). Design of structurally rigid *trans*-diaminebased Tf-amide organocatalysis with a dihydroanthracene framework for asymmetric conjugate additions of heterosubstituted aldehydes to vinyl sulfones. J. Am. Chem. Soc. 132, 17074–17076.

Moure, A.L., Arrayás, R.G., and Carretero, J.C. (2011). Catalytic asymmetric conjugate boration of α , β -unsaturated sulfones. Chem. Commun. (Camb.) 47, 6701–6703.

Mucha, A., Kafarski, P., and Berlicki, Ł. (2011). Remarkable potential of the α-aminophosphonate/phosphinate structure motif in medicinal chemistry. J. Med. Chem. 54, 5955–5980.

Narasimhulu, M., Reddy, T.S., Mahesh, K.C., Krishna, A.S., Rao, J.V., and Venkateswarlu, Y. (2009). Synthesis of yashahushidiol and its analogues and their cytotoxic activity against cancer cell lines. Bioorg. Med. Chem. Lett. 19, 3125–3127.

Newhouse, T., Baran, P.S., and Hoffmann, R.W. (2009). The economies of synthesis. Chem. Soc. Rev. 38, 3010–3021.

Nielsen, M., Jacobsen, C.B., Holub, N., Paixão, M.W., and Jørgensen, K.A. (2010). Asymmetric organocatalysis with sulfones. Angew. Chem. Int. Ed. 49, 2668–2679.

Nishida, G., Noguchi, M., Hirano, M., and Tanaka, K. (2008). Enantioselective synthesis of P-stereogenic alkynylphosphine oxides by Rhcatalyzed [2+2+2] cycloaddition. Angew. Chem. Int. Ed. 47, 3410–3413.

Nishimura, T., Takiguchi, Y., and Hayashi, T. (2012). Effect of chiral diene ligands in rhodiumcatalyzed asymmetric addition of arylboronic acids to α , β -unsaturated sulfonyl compounds. J. Am. Chem. Soc. 134, 9086–9089.

Ordóñez, M., Sayago, F.J., and Cativiela, C. (2012). Synthesis of quaternary α-aminophosphonic acids. Tetrahedron 68, 6369–6412.

Otsuka, Y., Takada, H., Yasuda, S., Kumagai, N., and Shibasaki, M. (2013). Direct catalytic asymmetric addition of allylic cyanides to aldehydes for expeditious access to enantioenriched unsaturated δ -valerolactones. Chem. Asian J. 8, 354–358.

Pansare, S.V., and Paul, E.K. (2011). The organocatalytic vinylogous aldol reaction: recent advances. Chem. Eur. J. 17, 8770–8779.

Quintard, A., Alexakis, A., and Mazet, C. (2011). Access to high levels of molecular complexity by one-pot iridium/enamine asymmetric catalysis. Angew. Chem. Int. Ed. *50*, 2354–2358.

Ray, B., and Mukherjee, S. (2018). Direct catalytic enantioselective vinylogous aldol reaction of allyl ketones to pyrazole-4,5-diones. J. Org. Chem. *83*, 10871–10880.

Shirokawa, S., Kamiyama, M., Nakamura, T., Okada, M., Nakazaki, A., Hosokawa, S., and Kobayashi, S. (2004). Remote asymmetric induction with vinylketene silyl N,O-acetal. J. Am. Chem. Soc. 126, 13604–13605.

Trost, B.M. (1991). The atom economy—a search for synthetic efficiency. Science 254, 1471–1477.

Trost, B.M., Dirat, O., and Gunzner, J.L. (2002). Callipeltoside A: assignment of absolute and relative configuration by total synthesis. Angew. Chem. Int. Ed. 41, 841–843. Wang, Z., and Hayashi, T. (2018). Rhodiumcatalyzed enantioposition-selective hydroarylation of divinylphosphine oxides with aryl boroxines. Angew. Chem. Int. Ed. 57, 1702– 1706.

Woo, S.Y., Kim, J.H., Moon, M.K., Han, S.-H., Yeon, S.K., Choi, J.W., Jang, B.K., Song, H.J., Kang, Y.G., Kim, J.W., et al. (2014). Discovery of vinyl sulfones as a novel class of neuroprotective agents toward Parkinson's disease therapy. J. Med. Chem. *57*, 1473–1487.

Xue, Z.-Y., Li, Q.-H., Tao, H.-Y., and Wang, C.-J. (2011). A facile Cu(I)/TF-BiphamPhos-catalyzed asymmetric approach to unnatural *a*-amino acid derivatives containing *gem*-bisphosphonates. J. Am. Chem. Soc. 133, 11757–11765.

Yamaguchi, A., Aoyama, N., Matsunaga, S., and Shibasaki, M. (2007). Ba-catalyzed direct Mannich-type reactions of a β , γ -unsaturated ester providing β -methyl aza-Morita–Baylis– Hillman-type products. Org. Lett. *9*, 3387–3390.

Yokosuka, A., Mimaki, Y., Sakagami, H., and Sashida, Y. (2002). New diarylheptanoids and diarylheptanoid glucosides from the rhizomes of *tacca chantrieri* and their cytotoxic activity. J. Nat. Prod. 65, 283–289.

Yuan, C., Yao, J., and Li, S. (1990). Studies on organophosphorus compounds XLIV. Structural effect of electrophiles on the regioselectivity of carbanion derived from dialkyl allylphosphonates. Phosphorus Sulfur Silicon Relat. Elem. 53, 21–27.

Yuan, C., Yao, J., and Li, S. (1991). Studies on organophosphorus compounds XLV. Structural effects of ester alkyl group on dialkyl allylphosphonate carbanion on the regioselectivity of electrophilic addition. Phosphorus Sulfur Silicon Relat. Elem. 55, 125–131.

Zhang, H.-J., and Yin, L. (2018). Asymmetric synthesis of α , β -unsaturated δ -lactones through copper(I)-catalyzed direct vinylogous aldol reaction. J. Am. Chem. Soc. 140, 12270–12279.

Zhou, T., Peters, B., Maldonado, M.F., and Govender, T. (2012). Enantioselective synthesis of chiral sulfones by Ir-catalyzed asymmetric hydrogenation: a facile approach to the preparation of chiral allylic and homoallylic compounds. J. Am. Chem. Soc. 134, 13592– 13595.

Zhu, Q., and Lu, Y. (2010). Stereocontrolled creation of all-carbon quaternary stereocenters by organocatalytic conjugate addition of oxindoles to vinyl sulfone. Angew. Chem. Int. Ed. 49, 7753–7756.

Zhu, B., Zhang, W., Lee, R., Han, Z., Wang, W., Tan, D., Huang, K.-W., and Jiang, Z. (2013). Direct asymmetric vinylogous aldol reaction of allyl ketones with isatins: divergent synthesis of 3-hydroxy-2-oxindole derivatives. Angew. Chem. Int. Ed. *52*, 6666–6670. ISCI, Volume 14

Supplemental Information

Asymmetric Vinylogous Aldol-type

Reactions of Aldehydes

with Allyl Phosphonate and Sulfone

Wen-Jun Yue, Cheng-Yuan Zhang, and Liang Yin

Supplement Information

Asymmetric Vinylogous Aldol-Type Reactions of Aldehydes with Allyl Phosphonate and Sulfone

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Copies of product NMR spectra







Figure S4. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3a, related to Table 2



Figure S3. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3a, related to Table 2



Figure S5. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3a**, related to **Table 2**



Figure S7. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3b, related to Table 2





Figure S8. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3b, related to Table 2

Figure S9. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound 3b, related to Table 2





Figure S11. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3c, related to Table 2





Figure S12. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3c, related to Table 2



Figure S14. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3d, related to Table 2



Figure S13. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3d, related to Table 2



Figure S15. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3d, related to Table 2



Figure S16. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3e, related to Table 2







Figure S18. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3e, related to Table 2



Figure S19. ¹H NMR (400 MHz, $CDCl_3$) spectrum of compound 3f, related to Table 2

Figure S20. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3f, related to Table 2





Figure S21. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3f, related to Table 2



Figure S22. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3g, related to Table 2

Figure S23. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3g, related to Table 2





Figure S24. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3g, related to Table 2



Figure S25. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3h, related to Table 2







Figure S27. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3h**, related to **Table 2**



Figure S29. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3i, related to Table 2





Figure S29. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3i, related to Table 2



Figure S30. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3j, related to Table 2







Figure S32. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3j, related to Table 2

Figure S33. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound **3***j*, related to **Table 2**










Figure S36. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3k, related to Table 2

Figure S37. 19 F NMR (376 MHz, CDCl₃) spectrum of compound 3k, related to Table 2





Figure S39. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3l, related to Table 2



Figure S38. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3l, related to Table 2



Figure S40. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3l, related to Table 2

Figure S41. 19 F NMR (376 MHz, CDCl₃) spectrum of compound 3l, related to Table 2





Figure S42. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3m, related to Table 2







Figure S44. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3m**, related to Table 2



Figure S46. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3n, related to Table 2





Figure S47. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3n**, related to Table 2



Figure S48. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 30, related to Table 2







Figure S50. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **30**, related to Table 2



Figure S51. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3p**, related to **Table 2**

Figure S52. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3p**, related to Table 2





Figure S53. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3p**, related to Table 2



Figure S54. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3q, related to Table 2

Figure S55. 13 C NMR (100 MHz, CDCl₃) spectrum of compound 3q, related to Table 2





Figure S56. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3q**, related to Table 2



Figure S58. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3r**, related to Table 2



36



Figure S59. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3r**, related to Table 2



Figure S61. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3s, related to Table 2



Figure S60. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3s, related to Table 2



Figure S62. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3s, related to Table 2



Figure S64. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3t, related to Table 2



40



Figure S65. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3t, related to Table 2



Figure S66. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3u**, related to Table 2







Figure S68. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3u**, related to Table 2



Figure S69. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3v, related to Table 2







Figure S71. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3v**, related to Table 2



Figure S72. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3w, related to Table 2







Figure S74. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3w**, related to Table 2



Figure S75. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3x, related to Table 2

Figure S76. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3x, related to Table 2





Figure S77. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3x, related to Table 2



Figure S78. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3y, related to Table 2







Figure S80. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3y**, related to Table 2



Figure S81. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3z, related to Table 2

Figure S82. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3z, related to Table 2





Figure S83. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3z**, related to Table 2



Figure S85. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3aa, related to Table 2



Figure S84. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3aa, related to Table 2



Figure S86. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3aa, related to Table 2







Figure S89. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3ab, related to Table 2


Figure S90. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3ac, related to Table 2

Figure S91. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3ac, related to Table 2





Figure S92. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3ac**, related to **Table 2**



Figure S93. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3ad, related to Table 2





Figure S95. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3ad, related to Table 2



Figure S97. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3ae, related to Table 2





Figure S98. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3ae, related to Table 2



Figure S99. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3af, related to Table 2

Figure S100. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3af, related to Table 2





Figure S101. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3af**, related to **Table 2**



Figure S102. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3ag, related to Table 2

Figure S103. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3ag, related to Table 2







Figure S106. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3ah, related to Table 2



Figure S105. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3ah, related to Table 2



Figure S107. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3ah, related to Table 2



Figure S108. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3ai, related to Table 2

Figure S109. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3ai, related to Table 2





Figure S110. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3ai**, related to **Table 2**



Figure S111. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3aj, related to Table 2

Figure S112. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 3aj, related to Table 2





Figure S113. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3aj**, related to **Table 2**



Figure S114. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3aj', related to Table 2



Figure S116. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3aj'**, related to **Table 2**



Figure S117. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **3ak**, related to **Table 2**

190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 fl (ppm)

10 0

-10

230

210



Figure S119. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3ak**, related to **Table 2**



Figure S120. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3ak', related to Table 2

Figure S121. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound **3ak'**, related to **Table 2**





Figure S122. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3ak**', related to **Table 2**





Figure S123. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 3al, related to Table 2



Figure S125. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **3al**, related to **Table 2**



Figure S126. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 3al', related to Table 2







Figure S128. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 3al', related to Table 2



Figure S129. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 5a, related to Table 3

Figure S130. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5a, related to Table 3





Figure S131. 1 H NMR (400 MHz, CDCl₃) spectrum of compound **5b**, related to Table 3







Figure S133. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound **5b**, related to **Table 3**



Figure S134. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5c, related to Table 3

Figure S135. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5c, related to Table 3





Figure S136. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 5d, related to Table 3

Figure S137. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5d, related to Table 3











Figure S140. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5f, related to Table 3

fl (ppm)

60 50

190 180 170 160 150 140 130 120 110 100 90 80 70



Figure S142. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5g, related to Table 3

Figure S143. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5g, related to Table 3





Figure S144. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **5h**, related to Table 3

Figure S145. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5h, related to Table 3





Figure S147. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5k, related to Table 3




Figure S148. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound 5k, related to Table 3



Figure S149. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 5l, related to Table 3

Figure S150. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5l, related to Table 3





Figure S151. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound 5l, related to Table 3



Figure S152. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **5n**, related to Table 3

Figure S153. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5n, related to Table 3





Figure S154. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 50, related to Table 3

fl (ppm)

110 100 90

80 70 60

50

40 30 20 10 0

220 210 200 190

180 170 160

150 140 130 120



Figure S156. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **5p**, related to Table 3

Figure S157. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5p, related to Table 3





Figure S158. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 5q, related to Table 3

Figure S159. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5q, related to Table 3





Figure S160. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 5r, related to Table 3

Figure S161. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5r, related to Table 3





Figure S162. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **5u**, related to Table 3







Figure S164. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5x, related to Table 3



Figure S166. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5y, related to Table 3

Figure S167. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5y, related to Table 3





Figure S168. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5z, related to Table 3

190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 fl (ppm)

10 0 -10

230

210



Figure S170. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 5aa, related to Table 3







230

210

190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fl (ppm)



Figure S174. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5ae, related to Table 3







Figure S176. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5ah, related to Table 3

Figure S177. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5ah, related to Table 3





Figure S179. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5ak, related to Table 3





Figure S180. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5ak', related to Table 3

Figure S181. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5ak', related to Table 3





Figure S182. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5al, related to Table 3

Figure S183. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5al, related to Table 3





Figure S184. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5al', related to Table 3

Figure S185. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5al', related to Table 3





190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 fl (ppm)

0 -10

230

210



Figure S188. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5an, related to Table 3

Figure S189. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5an, related to Table 3





Figure S190. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5ao, related to Table 3

Figure S191. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5ao, related to Table 3











Figure S194. ¹⁹F NMR (376 MHz, CDCl₃) spectrum of compound **5ap**, related to Table 3



Figure S195. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5aq, related to Table 3

Figure S196. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5aq, related to Table 3





Figure S197. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5ar, related to Table 3

Figure S198. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5ar, related to Table 3





Figure S199. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5as, related to Table 3

Figure S200. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5as, related to Table 3





Figure S201. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5at, related to Table 3

Figure S202. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 5at, related to Table 3





Figure S203. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 5au, related to Table 3







Figure S205. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 9, related to Scheme 3

Figure S206. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 9, related to Scheme 3





Figure S207. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 9, related to Scheme 3



Figure S208. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 10, related to Scheme 3

Figure S209. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 10, related to Scheme 3





Figure S210. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 10, related to Scheme 3



Figure S211. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 11, related to Scheme 3

Figure S212. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 11, related to Scheme 3





Figure S213. 1 H NMR (400 MHz, CDCl₃) spectrum of compound 12, related to Scheme 3




Figure S216. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 14, related to Scheme 3



130



Figure S217. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 15, related to Scheme 3

Figure S218. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 15, related to Scheme 3





Figure S220. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 16, related to Scheme 3





Figure S221. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 17, related to Scheme 3



Figure S224. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 18, related to Scheme 3





Figure S225. ¹H NMR (400 MHz, CDCl₃) spectrum of compound **19**, related to Scheme 3

Figure S226. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 19, related to Scheme 3





Figure S227. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound **19**, related to Scheme 3







Figure S230. ³¹P NMR (162 MHz, CDCl₃) spectrum of compound 20, related to Scheme 3



Figure S231. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 21, related to Scheme 3

Figure S232. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 21, related to Scheme 3





Figure S233. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 22, related to Scheme 3

Figure S234. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 22, related to Scheme 3





Figure S235. ¹H NMR (400 MHz, CDCl₃) spectrum of compound 23, related to Scheme 3

Figure S236. ¹³C NMR (100 MHz, CDCl₃) spectrum of compound 23, related to Scheme 3





Figure S238. ¹³C NMR (100 MHz, CDCl₃) spectrum of yashabushidiol B, related to Scheme 3



Transparent Methods

All reagents were obtained commercially unless otherwise noted. Nuclear Magnetic Resonance (NMR) spectra were acquired on an Agilent 400 or Bruker 400 spectrometer. For ¹H NMR, chemical shifts were reported in δ ppm referenced to an internal SiMe₄ standard. For ¹⁹F NMR, CFCl₃ was used as the reference with chemical shift at 0 ppm. For ¹³C NMR, chemical shifts were reported in the scale relative to NMR solvent (CDCl₃: 77.0 ppm) as an internal reference. ³¹P NMR spectra were referenced externally to phosphoric acid. Multiplicities are reported using the following abbreviations: br = broad, s = singlet, d = doublet, t = triplet, q = broadquartet, p = pentet, m = multiplet. Mass spectra (EI) were measured on Agilent Technologies 5973N GC-MS. High-resolution mass spectra (EI) were measured on Waters Micromass GCT Premier spectrometer. Mass spectra (ESI) were measured on Agilent Technologies 1100 Series LC-MS. High-resolution mass spectra (ESI) were measured on Thermo Scientific LTQ FT Ultra FT-MS. Mass spectra (DART) and high-resolution mass spectra (DART) were measured on Thermo Fisher Scienticfic LTQ FTICR-MS. Infrared (IR) spectra were recorded on Thermo Scientific Nicolet iS5 FT-IR. Optical rotation was measured using a 1 mL cell with 1.0 dm path length on a JASCO P-1030 polarimeter. HPLC analysis was conducted on a Shimadzu HPLC system equipped with Daicel chiral-stationary-phase columns (4.6 mm×250 mm).

The procedure for preparation of 2ah: A solution of (triphenylphosphoranylidene)-acetaldehyde (3.04 g, 10 mmol, 1.0 equiv) and dec-5-ynal (1.52 g, 10 mmol, 1.0 equiv) in absolute chloroform (concentration of the aldehyde: 0.3 M) was refluxed until no further reaction progress was monitored by GC/MS. Then the reaction mixture was adsorbed on a small amount of silica gel and was purified by column chromatography (petroleum ether/ethyl acetate = 100/1 to 80/1) to afford the aldehyde **2ah** (0.54 g, 3 mmol, 30% yield) as a pale green oil.

General procedure for catalytic asymmetric direct vinylogous aldol-type reaction of aldehydes and allyl phosphonate:

Procedure A:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (5.6 mg, 0.15 mmol, 0.05 equiv) and (*R*)-DTBM-SEGPHOS (17.7 mg, 0.15 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (2.0 mL, 0.15 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl phosphonate **1** (160.4 mg, 0.9 mmol, 3.0 equiv) and aldehyde **2** (0.3 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -10 °C, Barton's Base (12 µL, 0.06 mmol, 0.20 equiv) was added. The resulting reaction mixture was stirred at -10 °C for 48 hours. Then, the reaction mixture was quenched by acetic acid (300 µL (0.4 M in THF), 0.12 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -10 °C. After solvent was removed under reduced pressure, the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate/methanol) to give the desired product.

Procedure B:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (5.6 mg, 0.15 mmol, 0.05 equiv) and (S)-DTBM-SEGPHOS (17.7 mg, 0.15

mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (2.0 mL, 0.15 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl phosphonate **1** (160.4 mg, 0.9 mmol, 3.0 equiv) and aldehyde **2** (0.3 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -10 °C, Barton's Base (12 μ L, 0.06 mmol, 0.20 equiv) was added. The resulting reaction mixture was stirred at -10 °C for 48 hours. Then, the reaction mixture was quenched by acetic acid (300 μ L (0.4 M in THF), 0.12 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -10 °C. After solvent was removed under reduced pressure, the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate/methanol) to give the desired product.

General procedure for catalytic asymmetric direct vinylogous aldol-type reaction of aldehydes and allyl sulfone:

Procedure A:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (5.6 mg, 0.15 mmol, 0.05 equiv) and (*R*)-DTBM-SEGPHOS (17.7 mg, 0.15 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (2.0 mL, 0.15 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl sulfone **4** (109.9 mg, 0.6 mmol, 2.0 equiv) and aldehyde **2** (0.3 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -40 °C, Barton's Base (18 µL, 0.09 mmol, 0.30 equiv) was added. The resulting reaction mixture was stirred at -40 °C for 36 hours. Then, the reaction mixture was quenched by acetic acid (300 µL (0.4 M in THF), 0.12 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -40 °C. After solvent was removed under reduced pressure, the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate) to give the desired product.

Procedure B:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (5.6 mg, 0.15 mmol, 0.05 equiv) and (*S*)-DTBM-SEGPHOS (17.7 mg, 0.15 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (2.0 mL, 0.15 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl sulfone **4** (109.9 mg, 0.6 mmol, 2.0 equiv) and aldehyde **2** (0.3 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -40 °C, Barton's Base (18 µL, 0.09 mmol, 0.30 equiv) was added. The resulting reaction mixture was stirred at -40 °C for 36 hours. Then, the reaction mixture was quenched by acetic acid (300 µL (0.4 M in THF), 0.12 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -40 °C. After solvent was removed under reduced pressure, the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate) to give the desired product.

The procedure for determination of the absolute configuration of 3a

Absolute configuration of 3a was determined by its transformation to (*R*)-1-phenylpropane-1,3-diol as shown below and the comparison of its optical rotation with the one reported in literature (Denmark et. al., 2004).



Figure S239, related to Table 2

Ozone was bubbled into a solution of **3a** (110 mg, 0.39 mmol, 1.0 equiv) in MeOH (5.0 mL) at -78 °C until the appearance of a persistent blue color (about 30 min). The reaction solution was then allowed to warm up to 0 °C and the mixture was subsequently treated with NaBH₄ (73.8 mg, 1.95 mmol, 5 equiv.) at 0°C. The reaction mixture was allowed to warm up to room temperature and was stirred for additional 2 hours. Then, the reaction was quenched by H_2O (5 mL) and extracted with DCM (15 mL×3). The combined organic layers were dried over Na₂SO₄. After removal of solvent under reduced pressure, the crude was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to afford (*R*)-1-phenylpropane-1,3-diol (39 mg, colorless oil, 67% yield).

The procedure for determination of the absolute configuration of 5a

Absolute configuration of 5a was determined by its transformation to (*R*)-1-phenylpropane-1,3-diol as shown below and the comparison of its optical rotation with the one reported in literature (Denmark et. al., 2004).



Figure S240, related to Table 3

Ozone was bubbled into a solution of **5a** (94 mg, 0.33 mmol, 1.0 equiv) in MeOH (5.0 mL) at -78 °C until the appearance of a persistent blue color (about 30 min). The reaction solution was then allowed to warm up to 0 °C and the mixture was subsequently treated with NaBH₄ (62.4 mg, 1.65 mmol, 5 equiv.) at 0°C. The reaction mixture was allowed to warm up to room temperature and was stirred for additional 2 hours. Then, the reaction was quenched by H₂O (5 mL) and extracted with DCM (15 mL×3). The combined organic layers were dried over Na₂SO₄. After removal of solvent under reduced pressure, the crude was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to afford (*R*)-1-phenylpropane-1,3-diol (21 mg, colorless oil, 42% yield).

The procedure for preparation of rac-3a:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (9.3 mg, 0.025 mmol, 0.05 equiv) and *rac*-DTBM-SEGPHOS (29.5 mg, 0.025 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (2.0 mL, 0.25 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl phosphonate **1** (267.3 mg, 1.5 mmol, 3.0 equiv) and

aldehyde **2a** (53.1mg, 0.5 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -10 °C, Barton's Base (17.1mg, 0.10 mmol, 0.20 equiv) was added. The resulting reaction mixture was stirred at -10 °C for 48hours. Then, the reaction mixture was quenched by acetic acid (500 μ L(0.4 M in THF), 0.20 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -10 °C. Then the volatives were under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate/methanol = 30/15/1) to give *rac-3a*(128.0 mg, 90% yield) as a colorless oil.

The procedure for preparation of rac-5a:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (11.2 mg, 0.030 mmol, 0.05 equiv) and *rac*-DTBM-SEGPHOS (35.4 mg, 0.030 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (2.0 mL, 0.30 M) was added via a syringe. The mixture was stirred for 15 minutes to give a colorless catalyst solution. Then allyl sulfone **4** (220.0 mg, 1.2 mmol, 2.0 equiv) and aldehyde **2a** (63.7mg, 0.6 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -40 °C, Barton's Base (30.8mg, 0.18 mmol, 0.30 equiv) was added. The resulting reaction mixture was stirred at -40 °C for 12 hours. Then, the reaction mixture was quenched by acetic acid (600 µL (0.4 M in THF), 0.20 mmol, 0.40 equiv), and was stirred for additional 20 minutes at -40 °C. Then the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/2) to give *rac*-5a (110.0 mg, 63% yield) as pale green powders.

The procedure for preparation of $rac-\alpha-3a$:

*rac-3***a** was prepared according to a reported procedure (Yuan et. al., 1991). A dried 50 mL round bottom flask equipped with a magnetic stirring bar was charged with allyl phosphonate **1** (534.5 mg, 3.0 mmol, 1.0 equiv) under N₂ atmosphere. Anhydrous THF (10 mL) was added via a syringe. The mixture was cooled to -78 °C and was stirred for 10 minutes. Then "BuLi (1.3 mL (2.5 M solution in hexane), 3.15 mmol, 1.05 equiv) was added via a syringe. After 30 minutes, benzaldehyde **2a** (318.4 mg, 3 mmol, 1.0 equiv) was added via a syringe and the mixture was stirred for 30 minutes. The reaction was quenched by saturated aqueous NH₄Cl (5 mL) at -78 °C. The aqueous phase was extracted with ethyl acetate (20 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate/methanol = 14/7/1) to give *rac-α-3a* (724.9 mg, 85% yield, dr = 2.5/1) as a colorless oil.

The procedure for preparation of $rac - \alpha - 5a$:

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with LDA (1.0 mL (2 M solution in hexane/THF), 2 mmol, 1.0 equiv) under N₂ atmosphere. Anhydrous THF (2 mL) was added via a syringe. The mixture was cooled to -78 °C and HMPA (358.4 mg, 2 mmol, 1.0 equiv) was added via a syringe. The resulting mixture was stirred at -78 °C for 30 minutes and then allyl sulfone **5** (439.8 mg, 2.4 mmol, 1.2 equiv) was added. After 30 minutes, benzaldehyde **2a** (318.4 mg, 3 mmol, 1.5 equiv) was added and the resulting mixture was stirred for 20 minutes. The reaction was quenched by saturated aqueous NH₄Cl (5 mL) at -78 °C. The aqueous phase was extracted with ethyl acetate (10 mL × 3). The combined organic extracts were dried over

anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/1) to give *rac*- α -5a (101.0 mg, 15% yield, dr = 1/1) as pale green powders.

Proposed Mechanism for the Copper(I)-Catalyzed Asymmetric Aldol-Type Reaction:



Figure S241, Proposed Mechanism, related to Scheme 2

Based on these experimental observations and literature proposals, a postulated reaction pathway was given as shown above. In the presence of copper(I) complex U and Barton's Base, the deprotonation of substrate 1/4 occurred smoothly to give allylcopper(I) species V, which might form an equilibrium with allylcopper(I) species W. The $\overline{\alpha}$ -addition of V with aldehyde 2produced copper(I) alkoxide complex X, which afforded α -adduct after protonation with substrate 1/4. As demonstrated by the experiments, the α -addition was a significantly reversible process. It waspossible that the $\overline{\gamma}$ -addition of W with aldehyde 2 also furnished copper(I) alkoxide complex X. The γ -addition of allylcopper(I) species V generated copper(I) alkoxide complex Y through a six-memberring transition state, which was identified as a slightly reversible process. The protonation of Y with additional substrate 1/4 led to γ -adduct.

The procedure for gram-scale preparation of vinylogous product 3a:

A dried 100 mL round bottom flaske quipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (74.5 mg, 0.20 mmol, 0.05 equiv) and (*R*)-DTBM-SEGPHOS (235.9 mg, 0.20 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (40 mL, 0.2 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl phosphonate **1** (2.140 g, 12 mmol, 3.0 equiv) and benzaldehyde **2a** (424.5 mg, 4.0 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -10 °C, Barton's Base (137.0 mg, 0.80 mmol, 0.20 equiv) was added. The resulting reaction mixture was stirred at -10 °C for 48 hours. Then, the reaction mixture was quenched by acetic acid (4 mL (0.4 M in THF), 1.6 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -10 °C. After solvent was removed under reduced pressure, the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate/methanol = 30/15/1) to give product **3a** (0.990 g, 85% yield, 99% ee) as a colorless oil.

The procedure for gram-scale preparation of vinylogous product 5a:

A dried 100 mL round bottom flaske quipped with a magnetic stirring bar was charged with $[Cu(CH_3CN)_4]PF_6$ (74.5 mg, 0.20 mmol, 0.05 equiv) and (*R*)-DTBM-SEGPHOS (235.9 mg, 0.20 mmol, 0.05 equiv) in a glove box under Ar atmosphere. Anhydrous THF (40 mL, 0.1 M) was added via a syringe. The mixture was stirred at room temperature for 15 minutes to give a colorless catalyst solution. Then allyl sulfone **4** (1.466 g, 12 mmol, 3.0 equiv) and benzaldehyde **2a** (424.5 mg, 4.0 mmol, 1.0 equiv) were added sequentially. After the mixture was cooled to -40 °C, Barton's Base (205.5 mg, 1.20 mmol, 0.30 equiv) was added. The resulting reaction mixture was stirred at -40 °C for 36 hours. Then, the reaction mixture was quenched by acetic acid (4 mL (0.4 M in THF), 1.6 mmol, 0.40 equiv) and was stirred for additional 20 minutes at -40 °C. After solvent was removed under reduced pressure, the residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/2) to give product **5a** (1.100 g, 95% yield, 97% ee) as pale green powders.





A dried 50 mL round bottom flask equipped with a magnetic stirring bar was charged with **3a** (250 mg, 0.88 mmol, 1.0 equiv) and 2,6-lutidine (189 mg, 1.76 mmol, 2.0 equiv) under N₂ atmosphere. After the mixture was cooled to -10 °C, TBSOTf (465 mg, 1.76 mmol, 2.0 equiv) was added via a syringe. The resulting mixture was stirred at -10 °C for 7 hours. After removing the volatiles under reduced pressure, the crude was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to give product **9** (312 mg, 90% yield) as a colorless oil.





A dried 25 mL round bottom flask equipped with a magnetic stirring bar was charged with **9** (79.7 mg, 0.20 mmol, 1.0 equiv), Pd/C (16 mg, 5% w/w) and EtOH (4 mL). The resulting mixture was stirred at room temperature for 3 hours with a ballon filled with H₂. The black solids were filtered off and washed thoroughly with EtOH. The filtrate was concentrated under reduced pressure to give the crude, which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to give product **10** (78.5 mg, 98% yield) as a colorless oil.





(0.1 mL (2 M solution in hexane/THF), 2 mmol, 1.0 equiv) under N₂ atmosphere. Anhydrous THF (0.2 mL) was added via a syringe. The mixture was cooled to -78 °C and **10** (38.1 mg, 0.095 mmol, 1.0 equiv) in THF (0.5 mL) was added dropwise via a syringe. The resulting mixture was stirred at -78 °C for 5 minutes and then EtOCOOEt (11.8 mg, 0.10 mmol, 1.05 equiv) in THF (0.5 mL) was added dropwise via a syringe. The resulting mixture was stirred at -78 °C for 30 minutes and then was warmed to 0 °C. Benzaldehyde **2a** (11.1 mg, 0.105 mmol, 1.1 equiv) in THF (0.5 mL) was added dropwise via a syringe. The resulting mixture was stirred at room temperature overnight and then was quenched by saturated aqueous NH₄Cl (2 mL). The aqueous phase was extracted with diethyl ether (10 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 100/1) to give **11** (32.5 mg, 81% yield, E/Z > 20/1) as a colorless oil.



Figure S245, Transformations, related to Scheme 3

A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with LDA (0.105 mL (2 M solution in hexane/THF), 2 mmol, 1.0 equiv) under N₂ atmosphere. Anhydrous THF (0.2 mL) was added via a syringe. The mixture was cooled to -78 °C and **10** (40.1 mg, 0.10 mmol, 1.0 equiv) in THF (0.5 mL) was added dropwise via a syringe. The resulting mixture was stirred at -78 °C for 5 minutes and then EtOCOOEt (13.0 mg, 0.105 mmol, 1.05 equiv) in THF (0.5 mL) was added dropwise via a syringe. The resulting mixture was stirred at -78 °C for 30 minutes and then was warmed to 0 °C. Cinnamaldehyde **2y** (14.5 mg, 0.11 mmol, 1.1 equiv) in THF (0.5 mL) was added dropwise via a syringe. The resulting mixture was stirred at room temperature overnight and then was quenched by saturated aqueous NH₄Cl (2 mL). The aqueous phase was extracted with diethyl ether (10 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 100/1) to give **11** (32 mg, 71% yield, E/Z = 5/1) as a colorless oil.

Transformations of vinylogous product 5a:





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with **5a** (57.9 mg, 0.2 mmol, 1.0 equiv) under N₂ atmosphere. SmI₂ (10 mL (0.1 M solution in THF), 1 mmol, 5.0 equiv) was added via a syringe. The mixture was cooled to -20 $^{\circ}$ C and HMPA (0.8 mL) was added dropwise via a syringe. The resulting mixture was stirred at -20 $^{\circ}$ C for 2 hours, Then the reaction mixture was concentrated under reduced pressure to give the crude which was used in next step without further purification.

To the solution of above crude (0.2 mmol, 1.0 equiv) in toluene (2 mL) were added DMAP (4.8 mg, 0.04 mmol, 0.10 equiv) and benzoic anhydride (136 mg, 0.6 mmol, 1.5 equiv). The resulting mixture was stirred at room temperature for 10 hours. Then the reaction mixture was concentrated under reduced pressure to give the crude, which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 50/1) to give **13** (26 mg, 52% yield) as a pale yellow oil.





A dried 100 mL round bottom flask equipped with a magnetic stirring bar was charged with **5a** (1.00 g, 3.46 mmol, 1.0 equiv) under N₂ atmosphere. THF (40 mL) was added via a syringe. The mixture was cooled to 0 °C and LiBH(Et)₃ (4.5 mL (1 M solution in THF), 4.50 mmol, 1.3 equiv) was added dropwise via a syringe. The resulting mixture was stirred at room temperature for 4 hours. Then the reaction was quenched by saturated aqueous NH₄Cl (20 mL). The aqueous phase was extracted with ethyl acetate (50 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to give **14** (932 mg, 92% yield) as white powders.





A dried 50 mL round bottom flask equipped with a magnetic stirring bar was charged with **14** (697 mg, 2.40 mmol, 1.0 equiv) and 2,6-lutidine (514 mg, 4.80 mmol, 2.0 equiv) under N₂ atmosphere. After cooling to -10 °C, TBSOTf (1.27 g, 4.80 mmol, 2.0 equiv) was added via a syringe. The resulting mixture was stirred at -10 °C for 12 hours. After removing the volatiles under reduced pressure, the crude was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/1) to give product **15** (908 mg, 93% yield) as a colorless oil.





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with **15** (81.2 mg, 0.20 mmol, 1.0 equiv) under N_2 atmosphere. Anhydrous DME (5 mL) was added via a syringe. The mixture was cooled to -78 °C and KHMDS (0.40 mL (1 M solution in THF), 0.40 mmol, 2.0 equiv) was added via a syringe. After 3 minutes, benzaldehyde **2a** (31.8 mg, 0.30 mmol, 1.5 equiv) was added via a syringe and the resulting mixture was stirred for 2 hours. Then the reaction mixture was warm to room temperature and stirred for 12 hours. The reaction was

quenched by saturated aqueous NH₄Cl (5 mL). The aqueous phase was extracted with ethyl acetate (20 mL \times 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 100/1) to give **16** (47 mg, 67% yield) as a colorless oil.





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with **15** (40.6 mg, 0.10 mmol, 1.0 equiv) under N₂ atmosphere. Anhydrous THF (3 mL) was added via a syringe. The mixture was cooled to -78 °C and was stirred for 10 minutes. Then ^{*n*}BuLi (0.08 mL (1 M solution in THF), 0.20 mmol, 2.0 equiv) was added via a syringe. After 30 minutes, PhCOCl (21.1 mg, 0.15 mmol, 1.5 equiv) was added and the resulting mixture was stirred for 2 hours. Then the reaction was quenched by saturated aqueous NH₄Cl (5 mL). The aqueous phase was extracted with ethyl acetate (10 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure to give the crude which was used in next step without further purification.

The solution of above crude (0.1 mmol, 1.0 equiv) in THF (2 mL) was added to a mixture of activated Zn powder (180 mg) ,THF (4 mL) and $H_2O(4 mL)$. The resulting mixture was stirred at room temperature for 4 hours. The solids were filtered off and washed thoroughly with DCM. The filtrate was dried over anhydrous Na_2SO_4 and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 20/1) to give **17** (25 mg, 68% yield) as a colorless oil.





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with **5a** (63.1 mg, 0.20 mmol, 1.0 equiv) under N₂ atmosphere. Anhydrous THF (2 mL) was added via a syringe. The mixture was cooled to 0 °C and was stirred for 10 minutes. Then benzaldehyde **2a** (23.4 mg, 0.22 mmol, 1.1 equiv) and LiHMDS (0.2 mL (1 M solution in THF), 0.20 mmol, 1.0 equiv) were added via a syringe. After 15 minutes, benzaldehyde **2a** (23.4 mg, 0.22 mmol, 1.1 equiv) and LiHMDS (0.2 mL (1 M solution in THF), 0.20 mmol, 1.1 equiv) and LiHMDS (0.2 mL (1 M solution in THF), 0.20 mmol, 1.1 equiv) and LiHMDS (0.2 mL (1 M solution in THF), 0.20 mmol, 1.0 equiv) was added again. This procedure was repeated twice. Then the resulting reaction mixture was quenched by saturated aqueous NH₄Cl (5 mL). The aqueous phase was extracted with ethyl acetate (10 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure to give the crude which was used in next step without further purification.

The above crude (0.2 mmol, 1.0 equiv) was added to HOAc (4 mL, 80% in water). The resulting reaction mixture was heating to 80° C and stirred at this temperature overnight. Then the

resulting reaction mixture was quenched by saturated aqueous NaHCO₃ (20 mL). The aqueous phase was extracted with diethyl ether (20 mL \times 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure. The residue was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 1/2) to give **18** (45.5 mg, 74% yield) as white powders.

Synthetic Application of the Methodology:





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with *ent-3y* (93.1 mg, 0.30 mmol, 1.0 equiv) and 2,6-lutidine (64.3 mg, 0.60 mmol, 2.0 equiv) under N₂ atmosphere. After the mixture was cooled to -10 °C, TBSOTf (158.6 mg, 0.60 mmol, 2.0 equiv) was added. The resulting mixture was stirred at -10 °C for 4 hours. After removing the volatiles under reduced pressure, the crude was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to give product **19** (105.7 mg, 83% yield) as a colorless oil.





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with CuCl (3.0 mg, 0.03 mmol, 0.10 equiv), *rac*-BINAP (22.5 mg, 0.036 mmol, 0.12 equiv) and NaO'Bu (4.3 mg, 0.045 mmol, 0.15 eqiv) in a glove box under Ar atmosphere. **19** (127.5 mg, 0.30 mmol, 1.0 equiv) and B₂(Pin)₂ (152.4 mg, 0.6 mmol, 2.0 equiv) were added under N₂ atmosphere. Anhydrous THF (3.0 mL) was added via a syringe. The mixture was stirred at room temperature for 15 minutes. Then MeOH (19.2 mg, 0.6 mmol, 2.0 equiv) was added. The resulting reaction mixture was stirred at room temperature for 24 hours. Then, water (3 ml) and NaBO₃ H₂O (138.6 mg, 0.90 mmol, 3.0 equiv) were added sequentially. The mixture was stirred at room temperature for additional 3 hours. Then the resulting reaction mixture was quenched by saturated aqueous NH₄Cl (5 mL). The aqueous phase was extracted with diethyl ether (10 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure to give the crude which was used in next step without further purification.

To the solution of above crude (0.30 mmol, 1.0 equiv) in DCM (18 mL) was added 4Å molecular sieves (350 mg) and PCC (516 mg, 2.40 mmol, 8.0 equiv). The resulting mixture was stirred at room temperature for 12 hours. The solids were filtered off and washed thoroughly with ethyl acetate. The filtrate was concentrated under reduced pressure to give the crude which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 3/1) to give product **20** (97.8 mg, 74% yield) as a colorless oil.





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with **20** (61 mg, 0.14 mmol, 1.0 equiv) under N₂ atmosphere. THF (2.0 mL) was added via a syringe. Then Ba(OH)₂ (29.5 mg, 0.17 mmol, 1.25 equiv) was added. The resulting mixture was stirred for 30 minutes at room temperature and then benzaldehyde **2a** (15.3 mg, 0.15 mmol, 1.05 equiv) in THF/H₂O (2 mL, 40/1) was added dropwise via a syringe. The resulting mixture was stirred at room temperature for 2 hours. Then the reaction was quenched by saturated aqueous NH₄Cl (3 mL). The aqueous phase was extracted with diethyl ether (10 mL \times 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure to give the crude, which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 100/1) to give product **21** (46.1 mg, 85% yield) as a colorless oil.





A 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with **21** (42 mg, 0.107 mmol, 1.0 equiv) and THF (2.0 mL). Then HCl (0.21 mL (3 M solution in water), 0.63 mmol, 6.0 equiv) was added. The resulting mixture was stirred at room temperature for 4 hours. Then the reaction was quenched by saturated aqueous NaHCO₃ (3 mL). The aqueous phase was extracted with diethyl ether (10 mL \times 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure to give the crude, which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 6/1) to give product **22** (21.1 mg, 71% yield) as white powders.





A dried 25 mL Schlenk tube equipped with a magnetic stirring bar was charged with MeBH(OAc)₃ (157.9 mg, 0.60 mmol, 6.0 equiv) under N₂ atmosphere. Anhydrous CH₃CN (0.5 mL) and HOAc (0.5 mL) were added via syringes. The resulting mixture was stirred at room temperature for 30 minutes. Then the resulting mixture was cooled to -20 °C. **22** (27.8 mg, 0.10 mmol, 1.0 equiv) in anhydrous CH₃CN (1 mL) was added dropwise via a syringe. The resulting mixture was stirred at -20 °C for 4 hours. Then the reaction was quenched by saturated aqueous sodium potassium tartarate and saturated aqueous NaHCO₃. The aqueous phase was extracted with diethyl ether (10 mL × 3). The combined organic extracts were dried over anhydrous Na₂SO₄ and the volatives were removed under reduced pressure to give the crude, which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to give product **23** (25.2 mg, 90% yield, dr = 8/1) as white powders (Diastereoselectivity was determined by ¹H NMR

analysis of reaction crude mixture).



Figure S257, Synthetic application, related to Scheme 3

A dried 25 mL round bottom flask equipped with a magnetic stirring bar was charged with **23** (25 mg, 0.09 mmol, 1.0 equiv), Pd/C (27.4 mg, 5% w/w) and EtOH (2 mL). The resulting mixture was stirred for 2 hours at room temperature with a ballon filled with H₂. The black solids were filtered off and washed thoroughly with EtOH. The filtrate was concentrated under reduced pressure to give the crude, which was purified by silica gel column chromatography (petroleum ether/ethyl acetate = 2/1) to give product **yashabushidiol B** (23 mg, 88% yield) as white powders.

Characterization of all compounds:



¹**H NMR (400 MHz, CDCl₃)** δ 9.44 (d, *J* = 7.9 Hz, 1H), 6.80 (dt, *J* = 15.6, 6.8 Hz, 1H), 6.07 (dd, *J* = 15.6, 7.9 Hz, 1H), 2.44–2.34 (m, 2H), 2.19–2.12 (m, 2H), 2.10–2.06 (m, 2H), 1.72–1.55 (m, 2H), 1.47–1.24 (m, 4H), 0.84 (t, *J* = 7.1 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 193.88, 157.83, 133.26, 81.35, 78.64, 31.59, 31.08, 27.11, 21.87, 18.32, 18.21, 13.54 ppm.

MS(EI) m/z [M-H]⁺:177.00.

HRMS(EI) m/z [M]⁺: calcd. 178.1358, found 178.1359.

IR (film):2933, 2320, 1698, 1652, 1286 cm⁻¹.



3a: Procedure A, 78 mg, colorless liquid, 91% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.42–7.12 (m, 5H), 6.76 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.66 (dd, *J* = 21.2, 17.1 Hz, 1H), 4.81 (dd, *J* = 7.4, 5.5 Hz, 1H), 4.07–3.88 (m, 4H), 3.34 (s, 1H), 2.80–2.50 (m, 2H), 1.26 (td, *J* = 7.0, 5.4 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.89 (d, *J* = 4.9 Hz), 143.80, 128.36, 127.50, 125.78, 119.28 (d, *J* = 186.6 Hz), 72.50, 61.68 (d, *J* = 5.5 Hz), 43.94 (d, *J* = 22.0 Hz), 16.24 (d, *J* = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.05 ppm.

MS(ESI) m/z [M+H]⁺: 285.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 285.1250, found 285.1250.

IR (film): 3361, 2984, 1632, 1259, 1020, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = +19.72$ (*c* = 1.780, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 13/3, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 11.8 min, t_R(minor) = 13.4 min, ee = 99%.



Figure S258, the HPLC spectrum of compound 3a, related to Table 2



3b: Procedure A, 73 mg, colorless liquid, 81% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.32 (dd, *J* = 8.6, 5.5 Hz, 2H), 7.01 (t, *J* = 8.7 Hz, 2H), 6.75 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.66 (dd, *J* = 21.1, 17.1 Hz, 1H), 4.81 (t, *J* = 7.9 Hz, 1H), 4.23–3.82 (m, 4H), 3.62 (d, *J* = 3.5 Hz, 1H), 2.77–2.52 (m, 2H), 1.26 (td, *J* = 7.1, 3.7 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 162.09 (d, J = 245.5 Hz), 149.55 (d, J = 5.0 Hz), 139.59 (d, J = 3.0 Hz), 127.42 (d, J = 8.1 Hz), 119.58 (d, J = 186.6 Hz), 115.17 (d, J = 21.3 Hz), 71.88 (d, J = 0.7 Hz), 61.70 (d, J = 5.4 Hz), 44.01 (d, J = 22.1 Hz), 16.24 (d, J = 6.5 Hz) ppm.

¹⁹F NMR (376 MHz, CDCl₃) δ -115.10~-115.18 (m) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.88 ppm.

MS(ESI) m/z [M+H]⁺: 303.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 303.1154, found 303.1155.

IR (film): 3354, 2984, 1633, 1510, 1260, 1026 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +20.98 (*c* = 2.070, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 18.5 min, t_R(minor) = 19.9 min, ee = 99%.





Figure S259, the HPLC spectrum of compound 3b, related to Table 2



3c: Procedure A, 71 mg, colorless liquid, 74% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.31–7.19 (m, 4H), 6.75 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.65 (dd, *J* = 21.1, 17.1 Hz, 1H), 4.80 (t, *J* = 7.4 Hz, 1H), 4.06–3.89 (m, 4H), 3.78 (d, *J* = 3.2 Hz, 1H), 2.73–2.51 (m, 2H), 1.26 (td, *J* = 7.1, 2.7 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.49 (d, J = 5.0 Hz), 142.37, 133.11, 128.48, 127.19, 119.63 (d, J = 186.6 Hz), 71.81 (d, J = 1.3 Hz), 61.73 (d, J = 5.5 Hz), 43.93 (d, J = 22.1 Hz), 16.24 (d, J = 6.5 Hz) ppm.
³¹P NMR (162 MHz, CDCl₃) δ 17.83 ppm.

MS(ESI) m/z [M+H]⁺: 319.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 319.0860, found 319.0863.

IR (film): 3354, 2988, 1632, 1260, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28} = +18.26$ (*c* = 2.470, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 18.5 min, t_R(minor) = 19.9 min, ee = 99%.



Figure S260, the HPLC spectrum of compound 3c, related to Table 2



3d: Procedure A, 84 mg, colorless liquid, 77% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.46 (d, *J* = 8.4 Hz, 2H), 7.23 (d, *J* = 8.4 Hz, 2H), 6.76 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.66 (dd, *J* = 21.0, 17.1 Hz, 1H), 4.80 (t, *J* = 6.3 Hz, 1H), 4.03–3.84 (m, 4H), 3.43 (s, 1H), 2.72–2.54 (m, 2H), 1.27 (td, *J* = 7.1, 2.9 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.28 (d, J = 5.2 Hz), 142.76, 131.48 , 127.52, 121.32, 119.84 (d, J = 186.5 Hz), 71.93 (d, J = 1.2 Hz), 61.74 (d, J = 5.5 Hz), 43.87 (d, J = 22.0 Hz), 16.27 (d, J = 6.5 Hz) ppm. ³¹P NMR (162 MHz, CDCl₃) δ 17.75 ppm.

MS(ESI) m/z [M+H]⁺: 363.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 363.0353, found 363.0353.

IR (film): 3352, 2988, 1630, 1260, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28} = +16.74$ (*c* = 1.450, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 19.4 min, t_R(minor) = 20.8 min, ee = 98%.



Figure S261, the HPLC spectrum of compound 3d, related to Table 2



3e: Procedure A, 113 mg, colorless liquid, 92% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.66 (d, *J* = 8.3 Hz, 2H), 7.10 (d, *J* = 8.2 Hz, 2H), 6.75 (ddt, *J* = 22.1, 17.1, 7.0 Hz, 1H), 5.66 (dd, *J* = 20.9, 17.1 Hz, 1H), 4.79 (t, *J* = 6.2 Hz, 1H), 4.05–3.86 (m, 4H), 3.33 (s, 1H), 2.79–2.21 (m, 2H), 1.27 (td, *J* = 7.1, 2.8 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.27 (d, *J* = 5.0 Hz), 143.42, 137.46, 127.78, 119.86 (d, *J* = 186.4 Hz), 92.91, 72.02 (d, *J* = 1.3 Hz), 61.75 (d, *J* = 5.4 Hz), 43.85 (d, *J* = 22.0 Hz), 16.30 (d, *J* = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.77 ppm.

MS(ESI) m/z [M+H]⁺: 411.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 411.0217, found 411.0215.

IR (film): 3354, 2986, 1634, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{28}$ = +16.00 (*c* = 2.60, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 39/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 64.7 min, t_R(minor) = 70.9 min, ee = 99%.



Figure S262, the HPLC spectrum of compound 3e, related to Table 2



3f: Procedure A, 72 mg, colorless liquid, 80% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.22 (d, *J* = 8.0 Hz, 2H), 7.14 (d, *J* = 8.0 Hz, 2H), 6.74 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.66 (dd, *J* = 21.2, 17.1 Hz, 1H), 4.77 (t, *J* = 7.9 Hz, 1H), 4.05–3.86 (m, 4H), 3.05 (d, *J* = 3.4 Hz, 1H), 2.80–2.51 (m, 2H), 2.33 (s, 3H), 1.26 (q, *J* = 6.9 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.79 (d, J = 4.9 Hz), 140.69, 137.28, 129.09, 125.72, 119.41 (d, J = 186.3 Hz), 72.51 (d, J = 1.2 Hz), 61.64 (d, J = 5.4 Hz), 43.87 (d, J = 21.9 Hz), 21.06, 16.26 (d, J = 6.6 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.01 ppm.

MS(ESI) m/z [M+H]⁺: 341.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 341.1876, found 341.1879.

IR (film): 3371, 2985, 1635, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +15.25 (*c* = 1.510, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 19.5 min, t_R(minor) = 23.9 min, ee = 98%.





Figure S263, the HPLC spectrum of compound 3f, related to Table 2



3g: Procedure A, 92 mg, colorless liquid, 90% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.35 (d, *J* = 8.3 Hz, 2H), 7.27 (d, *J* = 8.3 Hz, 2H), 6.77 (ddt, *J* = 22.1, 17.1, 6.9 Hz, 1H), 5.69 (dd, *J* = 21.3, 17.1 Hz, 1H), 4.79 (dd, *J* = 7.3, 5.6 Hz, 1H), 4.08–3.89 (m, 4H), 3.10 (s, 1H), 2.83–2.53 (m, 2H), 1.40–1.15 (m, 15H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 150.52, 149.98 (d, J = 5.1 Hz), 140.66, 125.52, 125.30, 119.26 (d, J = 186.3 Hz), 72.39, 61.65 (d, J = 5.5Hz), 43.77 (d, J = 22.0 Hz), 34.46, 31.31, 16.27 (d, J = 6.5 Hz)ppm.
³¹P NMR (162 MHz, CDCl₃) δ 18.09 ppm.

MS(ESI) m/z [M+H]⁺: 299.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 299.1407, found 299.1405.

IR (film): 3366, 2963, 1635, 1230, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{29} = +16.03$ (*c* = 3.325, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 37/3, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 23.6 min, t_R(minor) = 25.7 min, ee = > 99%.



Figure S264, the HPLC spectrum of compound 3g, related to Table 2



3h: Procedure A, 84 mg, colorless liquid, 85% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.27 (d, *J* = 7.6 Hz, 2H), 7.22 (d, *J* = 8.4 Hz, 2H), 6.75 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.67 (dd, *J* = 21.1, 17.1 Hz, 1H), 4.79 (t, *J* = 7.4 Hz, 1H), 4.20–3.66 (m, 4H), 3.09 (d, *J* = 3.1 Hz, 1H), 2.74–2.54 (m, 2H), 2.47 (s, 3H), 1.27 (td, *J* = 7.0, 5.0 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.52 (d, J = 5.2 Hz), 140.54, 137.75, 126.59, 126.33, 119.65 (d, J = 186.3 Hz), 72.26 (d, J = 1.4 Hz), 61.70 (d, J = 5.5 Hz), 43.82 (d, J = 22.0 Hz), 16.28 (d, J = 6.0 Hz), 15.83 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.88 ppm.

MS(ESI) m/z [M+H]⁺: 331.10.

0-

10.0

HRMS(ESI) m/z [M+H]⁺: calcd. 331.1127, found 331.1126.

IR (film): 3366, 2988, 1635, 1260, 1025, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29}$ = +15.85 (*c* = 1.485, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 27.6 min, t_R(minor) = 30.4 min, ee = > 99%.



25.0

30.0

35.0

40.0

min

Figure S265, the HPLC spectrum of compound 3h, related to Table 2

20.0

15.0



3i: Procedure A, 76 mg, colorless liquid, 81% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, *J* = 8.7 Hz, 2H), 6.87 (d, *J* = 8.6 Hz, 2H), 6.74 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.68 (dd, *J* = 21.1, 17.1 Hz, 1H), 4.77 (t, *J* = 7.9 Hz, 1H), 4.17–3.86 (m, 4H), 3.79 (s, 3H), 2.79 (d, *J* = 3.3 Hz, 1H), 2.75–2.53 (m, 2H), 1.27 (q, *J* = 7.0 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 159.11, 149.66 (d, J = 5.1 Hz), 135.73, 127.02, 119.53 (d, J = 186.5 Hz), 113.82, 72.34 (d, J = 1.2 Hz), 61.66 (d, J = 5.4 Hz), 55.25, 43.82 (d, J = 21.9 Hz), 16.27 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.95 ppm.

MS(ESI) m/z [M+H]⁺: 315.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 315.1356, found 315.1355.

IR (film): 3368, 2988, 1612, 1260, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28} = +16.00$ (*c* = 1.600, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 29.2 min, t_R(minor) = 33.3 min, ee = 97%.



Figure S266, the HPLC spectrum of compound 3i, related to Table 2



3j: Procedure A, 88 mg, colorless liquid, 83% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.39 (d, *J* = 8.6 Hz, 2H), 7.19 (d, *J* = 8.2 Hz, 2H), 6.79 (ddt, *J* = 22.0, 17.2, 7.0 Hz, 1H), 5.70 (dd, *J* = 20.9, 17.2 Hz, 1H), 4.87 (t, *J* = 7.6 Hz, 1H), 4.11–3.84 (m, 4H), 3.25 (d, *J* = 3.4 Hz, 1H), 2.74–2.56 (m, 2H), 1.27 (td, *J* = 7.1, 3.3 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.13 (d, *J* = 5.0 Hz), 148.49 (d, *J* = 1.8 Hz), 142.38, 127.18, 120.92, 120.40 (d, *J* = 257.0 Hz), 119.07, 71.85 (d, *J* = 1.3 Hz), 61.73 (d, *J* = 5.5 Hz), 43.93 (d, *J* = 22.1 Hz), 16.23 (d, *J* = 6.5 Hz) ppm.

¹⁹F NMR (376 MHz, CDCl₃) δ -57.96 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.71 ppm.

MS(ESI) m/z [M+H]⁺: 369.10.

10.0

12.5

15.0

HRMS(ESI) m/z [M+H]⁺: calcd. 369.1073, found 369.1071.

IR (film): 3361, 2989, 1636, 1260, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +14.54 (*c* = 1.100, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 19/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 20.9 min, t_R(minor) = 22.8 min, ee = 99%.



Figure S267, the HPLC spectrum of compound 3j, related to Table 2

17.5

20.0

22.5

25.0

27.5

30.0

32.5

min


3k: Procedure A, 82 mg, colorless liquid, 90% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.52 (t, *J* = 6.9 Hz, 1H), 7.27–7.18 (m, 1H), 7.14 (t, *J* = 7.4 Hz, 1H), 7.05–6.95 (m, 1H), 6.81 (ddt, *J* = 24.0, 17.1, 6.9 Hz, 1H), 5.68 (dd, *J* = 21.2, 17.1 Hz, 1H), 5.16 (dd, *J* = 10.7, 5.6 Hz, 1H), 4.05–3.85 (m, 4H), 3.78 (d, *J* = 4.3 Hz, 1H), 2.78–2.58 (m, 2H), 1.25 (q, *J* = 7.0 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 159.39 (d, J = 245.2 Hz), 149.63 (d, J = 5.0 Hz), 130.88 (d, J = 13.4 Hz), 128.79 (d, J = 8.2 Hz), 127.29 (d, J = 4.4 Hz), 124.23 (d, J = 3.4 Hz), 119.43 (d, J = 186.4 Hz), 115.06 (d, J = 21.7 Hz), 66.33, 61.69 (d, J = 5.5Hz), 42.72 (d, J = 22.1 Hz), 16.23 (d, J = 6.5 Hz) ppm.

¹⁹F NMR (376 MHz, CDCl₃) δ -119.42~-119.56 (m) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.97 ppm.

MS(ESI) m/z [M+H]⁺: 303.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 303.1156, found 303.1155.

IR (film): 3353, 2986, 1634, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +23.86 (*c* = 2.480, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 12.7 min, t_R(minor) = 16.7 min, ee = 98%.



Figure S268, the HPLC spectrum of compound 3k, related to Table 2



3I: Procedure A, 96 mg, colorless liquid, 91% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.84 (d, *J* = 7.8 Hz, 1H), 7.59 (dd, *J* = 14.9, 7.7 Hz, 2H), 7.36 (t, *J* = 7.6 Hz, 1H), 6.88 (ddt, *J* = 22.0, 17.1, 6.9 Hz, 1H), 5.71 (dd, *J* = 21.1, 17.1 Hz, 1H), 5.34–5.14 (m, 1H), 4.05–3.94 (m, 4H), 3.91 (d, *J* = 3.3 Hz, 1H), 2.69–2.49 (m, 2H), 1.28 (td, *J* = 7.1, 2.3 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.81 (d, J = 5.0 Hz), 143.29, 132.23, 127.63, 127.39, 126.42 (q, J = 30.6 Hz), 125.32 (q, J = 5.9 Hz), 124.30 (q, J = 273.9 Hz), 119.39 (d, J = 186.8 Hz), 67.86, 61.70 (d, J = 5.5 Hz), 44.13 (d, J = 22.4 Hz), 16.22 (d, J = 6.5 Hz) ppm.

¹⁹F NMR (376 MHz, CDCl₃) δ -58.23 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 19.97 ppm.

MS(ESI) m/z [M+Na]⁺: 375.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 353.1124, found 353.1122.

IR (film): 3342, 2985, 1632, 1259, 1056, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +28.63 (*c* = 2.655, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 9.5 min, t_R(minor) = 13.9 min, ee = 95%.



Figure S269, the HPLC spectrum of compound 31, related to Table 2



3m: Procedure A, 86 mg, colorless liquid, 91% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.34 (dd, *J* = 7.5, 1.4 Hz, 1H), 7.29–7.21 (m, 1H), 6.95 (t, *J* = 7.4 Hz, 1H), 6.87 (d, *J* = 8.2 Hz, 1H), 6.85–6.71 (m, 1H), 5.69 (dd, *J* = 21.4, 17.2 Hz, 1H), 5.06 (dd, *J* = 12.4, 6.0 Hz, 1H), 4.10–3.90 (m, 4H), 3.84 (s, 3H), 3.05 (d, *J* = 5.8 Hz, 1H), 2.72–2.67 (m, 2H), 1.28 (td, *J* = 7.1, 4.2 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 156.17, 150.25 (d, J = 4.8 Hz), 131.30, 128.49, 126.68, 120.72, 118.96 (d, J = 186.4 Hz), 110.35, 68.94 (d, J = 1.1 Hz), 61.59 (d, J = 5.4 Hz), 55.21, 42.08 (d, J = 21.9 Hz), 16.29 (d, J = 6.6 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.20 ppm.

MS(ESI) m/z [M+Na]⁺: 337.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 315.1356, found 315.1354.

IR (film): 3365, 2982, 1632, 1239, 1026, 756 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +22.46 (*c* = 2.095, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 27.3 min, t_R(minor) = 34.0 min, ee = 98%.





Figure S270, the HPLC spectrum of compound 3m, related to Table 2



3n: Procedure A, 77 mg, colorless liquid, 81% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.36 (s, 1H), 7.29–7.18 (m, 3H), 6.76 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.66 (dd, *J* = 21.1, 17.1 Hz, 1H), 4.87–4.69 (m, 1H), 4.07–3.90 (m, 5H), 2.73–2.53 (m, 2H), 1.27 (td, *J* = 7.1, 3.0 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.54 (d, J = 4.9 Hz), 146.12, 134.21, 129.65, 127.49, 125.97, 123.95, 119.53 (d, J = 186.6 Hz), 71.78 (d, J = 1.0 Hz), 61.78 (d, J = 5.4 Hz), 43.88 (d, J = 22.1 Hz), 16.23 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.89 ppm.

MS(ESI) m/z [M+H]⁺: 319.05.

0

HRMS(ESI) m/z [M+H]⁺: calcd. 319.0860, found 319.0863.

IR (film): 3346, 2984, 1635, 1259, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29}$ = +18.19 (*c* = 2.420, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 14.8 min, t_R(minor) = 18.4 min, ee = 98%.





Figure S271, the HPLC spectrum of compound 3n, related to Table 2



3o: Procedure A, 96 mg, colorless liquid, 88% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.52 (s, 1H), 7.38 (d, *J* = 7.8 Hz, 1H), 7.27 (d, *J* = 7.1 Hz, 1H), 7.20 (t, *J* = 7.7 Hz, 1H), 6.77 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.67 (dd, *J* = 21.1, 17.1 Hz, 1H), 4.80 (t, *J* = 7.9 Hz, 1H), 4.07–3.90 (m, 4H), 3.82 (d, *J* = 3.7 Hz, 1H), 2.74–2.49 (m, 2H), 1.27 (td, *J* = 7.1, 3.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.37 (d, J = 5.1 Hz), 146.30, 130.51, 129.99, 128.89, 124.42, 122.52, 119.71 (d, J = 186.5 Hz), 71.81 (d, J = 1.3 Hz), 61.78 (d, J = 5.5 Hz), 43.90 (d, J = 22.1 Hz), 16.28 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.83 ppm.

MS(ESI) m/z [M+H]⁺: 363.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 363.0355, found 363.0353.

IR (film): 3352, 2986, 1630, 1260, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +14.69 (*c* = 2.265, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 15.8 min, t_R(minor) = 20.3 min, ee = 97%.



Figure S272, the HPLC spectrum of compound 30, related to Table 2



3p: Procedure A, 94 mg, pale yellow liquid, 94% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.03 (d, *J* = 8.1 Hz, 1H), 7.89–7.79 (m, 1H), 7.76 (d, *J* = 8.2 Hz, 1H), 7.68 (d, *J* = 7.1 Hz, 1H), 7.56–7.42 (m, 3H), 6.89 (ddt, *J* = 22.0, 17.1, 6.9 Hz, 1H), 5.69 (dd, *J* = 21.1, 17.1 Hz, 1H), 5.62–5.56 (m, 1H), 4.05–3.77 (m, 4H), 3.21 (d, *J* = 3.4 Hz, 1H), 2.91–2.58 (m, 2H), 1.24 (dt, *J* = 12.9, 7.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.95 (d, J = 5.0 Hz), 139.28, 133.72, 130.04, 128.95, 128.05, 126.13, 125.55, 125.42, 123.00, 122.80, 119.34 (d, J = 186.5 Hz), 69.39, 61.68 (d, J = 7.1 Hz), 43.01 (d, J = 22.0 Hz), 16.26 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.02 ppm.

MS(ESI) m/z [M+H]⁺: 335.15.

0

15.0

HRMS(ESI) m/z [M+H]⁺: calcd. 335.1407, found 335.1406.

IR (film): 3356, 2983, 1635, 1259, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29}$ = +38.34 (*c* = 1.620, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 22.2 min, t_R(minor) = 24.6 min, ee = 99%.



22.5

25.0

27.5

30.0

min

Figure S273, the HPLC spectrum of compound 3p, related to Table 2

20.0

17.5



3q: Procedure A, 70 mg, colorless liquid, 85% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.35 (d, *J* = 1.0 Hz, 1H), 6.76 (ddt, *J* = 22.1, 17.2, 6.9 Hz, 1H), 6.31 (dd, *J* = 3.1, 1.8 Hz, 1H), 6.25 (d, *J* = 3.2 Hz, 1H), 5.73 (dd, *J* = 21.0, 17.2 Hz, 1H), 4.83 (t, *J* = 6.6 Hz, 1H), 4.14–3.88 (m, 4H), 3.73 (s, 1H), 2.77 (t, *J* = 6.7 Hz, 2H), 1.28 (td, *J* = 7.1, 1.4 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 155.87, 149.22 (d, *J* = 5.1 Hz), 141.84, 119.45 (d, *J* = 186.7 Hz), 110.09, 106.16, 66.01 (d, *J* = 1.2 Hz), 61.75 (d, *J* = 5.6 Hz), 40.39 (d, *J* = 22.3 Hz), 16.23 (d, *J* = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.96 ppm.

MS(ESI) m/z [M+H]⁺: 275.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 275.1043, found 275.1045.

IR (film): 3361, 2985, 1635, 1226, 1020, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{29} = +9.75$ (*c* = 3.005, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 37/3, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 34.0 min, t_R(minor) = 39.4 min, ee = > 99%.



Figure S274, the HPLC spectrum of compound 3q, related to Table 2



3r: Procedure A, 79 mg, colorless liquid, 91% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.22 (dd, *J* = 4.8, 1.2 Hz, 1H), 6.94 (dd, *J* = 7.9, 3.0 Hz, 2H), 6.76 (ddt, *J* = 22.1, 17.1, 6.9 Hz, 1H), 5.71 (dd, *J* = 21.1, 17.1 Hz, 1H), 5.06 (t, *J* = 6.4 Hz, 1H), 4.22–3.70 (m, 5H), 2.91–2.59 (m, 2H), 1.27 (td, *J* = 7.1, 3.9 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.34 (d, J = 5.3 Hz), 147.97, 126.54, 124.42, 123.59, 119.53 (d, J = 186.4 Hz), 68.40, 61.75 (d, J = 5.4 Hz), 44.02 (d, J = 22.1 Hz), 16.24 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.94 ppm.

MS(ESI) m/z [M+H]⁺: 291.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 291.0814, found 291.0813.

IR (film): 3342, 2984, 1634, 1259, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{29} = +9.24$ (*c* = 2.640, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 18.3 min, t_R(minor) = 20.3 min, ee = > 99%.



Figure S275, the HPLC spectrum of compound 3r, related to Table 2



3s: Procedure A, 63 mg, colorless liquid, 76% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.43–7.34 (m, 2H), 6.77 (ddt, *J* = 22.0, 17.2, 6.9 Hz, 1H), 6.40 (s, 1H), 5.81–5.68 (m, 1H), 4.82 (t, *J* = 5.2 Hz, 1H), 4.17–3.89 (m, 4H), 2.88 (d, *J* = 2.5 Hz, 1H), 2.76–2.57 (m, 2H), 1.30 (td, *J* = 7.1, 1.6 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.22 (d, J = 5.2 Hz), 143.39, 139.07, 128.25, 119.88 (d, J = 186.6 Hz), 108.37, 65.37 (d, J = 1.4 Hz), 61.71 (d, J = 5.4 Hz), 42.62 (d, J = 22.1 Hz), 16.28 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.97 ppm.

MS(ESI) m/z [M+H]⁺: 275.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 275.1043, found 275.1041.

IR (film): 3368, 2989, 1631, 1260, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{27}$ = +10.84 (*c* = 1.070, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 21.1 min, t_R(minor) = 25.1 min, ee = 98%.



Figure S276, the HPLC spectrum of compound 3s, related to Table 2



3t: Procedure A, 47 mg, colorless liquid, 55% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.57 (s, 1H), 8.47 (d, *J* = 3.4 Hz, 1H), 7.74 (d, *J* = 7.8 Hz, 1H), 7.29 (t, *J* = 6.2 Hz, 1H), 6.81 (ddt, *J* = 24.1, 17.1, 6.9 Hz, 1H), 5.71 (dd, *J* = 20.9, 17.1 Hz, 1H), 4.90 (dd, *J* = 7.4, 5.3 Hz, 1H), 4.13–3.81 (m, 4H), 2.75–2.53 (m, 2H), 1.27 (t, *J* = 7.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.17 (d, J = 4.3 Hz), 148.59, 147.55, 139.53, 133.73, 123.46, 120.00 (d, J = 186.8 Hz), 70.13, 61.77 (d, J = 5.7 Hz), 43.81 (d, J = 22.1 Hz), 16.25 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.62 ppm.

MS(ESI) m/z [M+Na]⁺: 308.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 286.1203, found 286.1203.

IR (film): 3355, 2983, 1634, 1229, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{25} = +34.45$ (*c* = 4.000, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 49.9 min, t_R(minor) = 56.1 min, ee = > 99%.



Figure S277, the HPLC spectrum of compound 3t, related to Table 2



3u: Procedure A, 87 mg, pale yellow liquid, 85% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.87–7.78 (m, 2H), 7.48–7.28 (m, 3H), 6.85 (ddt, *J* = 22.0, 17.1, 6.9 Hz, 1H), 5.69 (dd, *J* = 21.0, 17.1 Hz, 1H), 5.32–5.07 (m, 1H), 4.03–3.85 (m, 4H), 3.49 (d, *J* = 3.9 Hz, 1H), 2.93–2.66 (m, 2H), 1.30–1.18 (m, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.57 (d, J = 4.9 Hz), 140.92, 138.75, 136.93, 124.43, 124.07, 122.94, 122.51, 122.06, 119.56 (d, J = 186.5 Hz), 68.17 (d, J = 1.3 Hz), 61.72 (d, J = 6.1 Hz), 42.05 (d, J = 22.1 Hz), 16.27 (d, J = 6.6 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.89 ppm.

MS(ESI) m/z [M+H]⁺: 341.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 341.0971, found 341.0972.

IR (film): 3351, 2988, 1630, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +32.45 (*c* = 1.465, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 21.2 min, t_R(minor) = 23.3 min, ee = > 99%.



Figure S278, the HPLC spectrum of compound 3u, related to Table 2



3v: Procedure A, 79 mg, colorless liquid, 81% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.49 (d, *J* = 7.4 Hz, 1H), 7.42 (d, *J* = 8.0 Hz, 1H), 7.31–7.09 (m, 2H), 6.82 (ddt, *J* = 24.0, 17.1, 6.9 Hz, 1H), 6.63 (s, 1H), 5.73 (dd, *J* = 20.9, 17.1 Hz, 1H), 4.96 (t, *J* = 6.2 Hz, 1H), 4.39 (s, 1H), 4.05–3.79 (m, 4H), 3.00–2.64 (m, 2H), 1.19 (td, *J* = 7.0, 1.2 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.66 (d, *J* = 5.4 Hz), 154.70, 149.02, 128.03, 124.08, 122.75, 120.99, 119.75 (d, *J* = 181.9 Hz), 111.11, 102.88, 66.58, 61.78 (d, *J* = 5.4 Hz), 40.45 (d, *J* = 22.4 Hz), 16.17 (d, *J* = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.85 ppm.

MS(ESI) m/z [M+H]⁺: 325.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 325.1199, found 325.1200.

IR (film): 3341, 2988, 1632, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{29} = +16.59$ (*c* = 2.590, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 19.1 min, t_R(minor) = 20.3 min, ee = 97%.



Figure S279, the HPLC spectrum of compound 3v, related to Table 2



3w: Procedure A, 92 mg, colorless liquid, 90% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.78 (d, *J* = 7.6 Hz, 1H), 7.67 (d, *J* = 7.2 Hz, 1H), 7.36–7.23 (m, 2H), 7.15 (s, 1H), 6.79 (ddt, *J* = 24.0, 17.1, 6.9 Hz, 1H), 5.70 (dd, *J* = 21.0, 17.1 Hz, 1H), 5.12 (t, *J* = 6.3 Hz, 1H), 4.21 (s, 1H), 4.01–3.75 (m, 4H), 2.87–2.67 (m, 2H), 1.23–1.11 (m, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 148.87 (d, *J* = 4.9 Hz), 148.44, 139.39, 139.26, 124.27, 124.16, 123.43, 122.39, 120.18, 119.98 (d, *J* = 186.0 Hz), 69.05, 61.77 (d, *J* = 5.2 Hz), 43.67 (d, *J* = 22.3 Hz), 16.17 (dd, *J* = 6.5, 3.6 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.78 ppm.

MS(ESI) m/z [M+H]⁺: 341.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 341.0971, found 341.0972.

IR (film): 3336, 2988, 1635, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29}$ = +11.64 (*c* = 1.760, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 60.1 min, t_R(minor) = 64.5 min, ee = > 99%.



Peak#	Ret. Time	Area%	
1	60.119	99.708	Ā
2	64.493	0.292	
	1	I	* * * *

Figure S280, the HPLC spectrum of compound 3w, related to Table 2



3x: Procedure A, 70 mg, colorless liquid, 58% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.08 (d, *J* = 7.5 Hz, 2H), 7.53–7.33 (m, 4H), 7.23 (t, *J* = 7.4 Hz, 1H), 6.96–6.65 (m, 1H), 5.70 (dd, *J* = 20.9, 17.3 Hz, 1H), 5.06–4.96 (m, 1H), 4.36 (q, *J* = 7.2 Hz, 2H), 4.03–3.80 (m, 4H), 2.91–2.69 (m, 2H), 1.90–1.70 (br, 1H), 1.42 (t, *J* = 7.2 Hz, 3H), 1.30–1.10 (m, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.76 (d, J = 4.9 Hz), 140.24, 139.60, 133.97, 125.80, 123.58, 122.86, 122.69, 120.39, 119.58 (d, J = 186.1 Hz), 118.89, 117.82, 108.51, 108.49, 73.50 (d, J = 1.1 Hz), 61.65 (d, J = 5.3 Hz), 44.27 (d, J = 21.8 Hz), 37.57, 16.77 (d, J = 6.5 Hz), 13.78 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.97 ppm.

MS(ESI) m/z [M+Na]⁺: 424.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 402.1829, found 402.1829.

IR (film): 3361, 2985, 1635, 1260, 1025, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = +14.31$ (*c* = 0.460, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK IF-3, hexane/*i*-PrOH = 8/1, flow rate: 0.9 mL/min, λ = 207 nm, t_R(major) = 47.2 min, t_R(minor) = 53.7 min, ee = > 99%.



Figure S281, the HPLC spectrum of compound 3x, related to Table 2



3y: Procedure A, 71 mg, colorless liquid, 76% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.42–7.17 (m, 5H), 6.82 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 6.60 (d, *J* = 15.9 Hz, 1H), 6.21 (dd, *J* = 15.9, 6.6 Hz, 1H), 5.77 (dd, *J* = 21.0, 17.1 Hz, 1H), 4.55–4.35 (m, 1H), 4.11–3.92 (m, 4H), 2.69 (s, 1H), 2.66–2.46 (m, 2H), 1.27 (q, *J* = 7.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.33 (d, J = 5.1 Hz), 136.33, 131.12, 130.78, 128.55, 127.78, 126.46, 119.82 (d, J = 186.4 Hz), 71.16 (d, J = 1.2 Hz), 61.74 (d, J = 5.3 Hz), 42.18 (d, J = 22.0 Hz), 16.27 (d, J = 6.6 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.89 ppm.

MS(ESI) m/z [M+Na]⁺: 333.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 311.1407, found 311.1405.

IR (film): 3361, 2983, 1631, 1228, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +1.10 (*c* = 1.150, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 23.4 min, t_R(minor) = 25.6 min, ee = 97%.



Figure S282, the HPLC spectrum of compound 3y, related to Table 2



3z: Procedure A, 76 mg, colorless liquid, 78% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 7.32 (t, *J* = 7.5 Hz, 2H), 7.28–7.18 (m, 3H), 6.80 (ddt, *J* = 21.9, 17.1, 7.0 Hz, 1H), 6.52 (s, 1H), 5.77 (dd, *J* = 21.0, 17.1 Hz, 1H), 4.43–4.23 (m, 1H), 4.12–3.93 (m, 4H), 2.68–2.49 (m, 2H), 2.38 (d, *J* = 3.2 Hz, 1H), 1.88 (s, 3H), 1.28 (q, *J* = 7.2 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.58 (d, *J* = 4.9 Hz), 139.07, 137.15, 128.92, 128.10, 126.57, 126.19, 119.43 (d, *J* = 186.7 Hz), 76.11 (d, *J* = 1.2 Hz), 61.71 (d, *J* = 5.4 Hz), 40.13 (d, *J* = 22.0 Hz), 16.29 (d, *J* = 6.5 Hz), 13.49 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.90 ppm.

MS(ESI) m/z [M+Na]⁺: 347.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 325.1562, found 325.1562.

IR (film): 3366, 2985, 1634, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = -11.16 (*c* = 1.260, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK IA, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 19.6 min, t_R(minor) = 22.4 min, ee = 98%.



Figure S283, the HPLC spectrum of compound 3z, related to Table 2



3aa: Procedure A, 81 mg, colorless liquid, 85% yield.

¹H NMR (400 MHz, CDCl₃) δ 7.19–7.13 (m, 1H), 6.98–6.93 (m, 2H), 6.89–6.68 (m, 2H), 6.05 (dd, *J* = 15.7, 6.4 Hz, 1H), 5.77 (dd, *J* = 21.0, 17.2 Hz, 1H), 4.51–4.31 (m, 1H), 4.10–3.98 (m, 4H), 2.65–2.45 (m, 2H), 2.34–2.10 (br, 1H), 1.29 (m, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.28 (d, J = 4.8 Hz), 141.44, 130.61, 127.36, 126.09, 124.48, 123.97, 119.85 (d, J = 186.4 Hz), 70.81 (d, J = 1.2 Hz), 61.80 (d, J = 5.4 Hz), 42.10 (d, J = 22.0 Hz), 16.27 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.89 ppm.

MS(ESI) m/z [M+H]⁺: 317.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 317.0971, found 317.0970.

IR (film): 3358, 2986, 1631, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = +3.05$ (*c* = 0.680, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 22.6 min, t_R(minor) = 27.4 min, ee = 98%.



Figure S284, the HPLC spectrum of compound 3aa, related to Table 2



3ab: Procedure A, 48 mg, colorless liquid, 68% yield.

¹H NMR (400 MHz, CDCl₃) δ 6.78 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.93–5.82 (m, 1H), 5.81–5.67 (m, 1H), 5.27 (d, *J* = 17.2 Hz, 1H), 5.15 (d, *J* = 10.4 Hz, 1H), 4.27 (q, *J* = 6.1 Hz, 1H), 4.17–3.97 (m, 4H), 2.64 (s, 1H), 2.58–2.38 (m, 2H), 1.32 (t, *J* = 7.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.33 (d, J = 5.0 Hz), 139.90, 119.66 (d, J = 186.9 Hz), 115.28, 71.22 (d, J = 1.2 Hz), 61.72 (d, J = 5.3 Hz), 41.78 (d, J = 22.0 Hz), 16.30 (d, J = 6.4 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.96 ppm.

MS(ESI) m/z [M+Na]⁺: 257.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 235.1094, found 235.1094.

IR (film): 3379, 2985, 1633, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{28}$ = +2.52 (*c* = 1.060, CHCl₃, 93% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 13.7 min, t_R(minor) = 14.9 min, ee = 93%.





Figure S285, the HPLC spectrum of compound 3ab, related to Table 2



3ac

3ac: Procedure A, 48 mg, colorless liquid, 58% yield, E/Z = 6/1 (**2ac** was used as a mixture (E/Z = 6/1)).

¹**H NMR (400 MHz, CDCl₃)** δ 6.76 (ddt, *J* = 21.9, 17.1, 7.0 Hz, 1H), 6.19 (dd, *J* = 15.2, 10.4 Hz, 1H), 6.07–5.90 (m, 1H), 5.82–5.62 (m, 2H), 5.55 (dd, *J* = 15.2, 6.8 Hz, 1H), 4.28 (q, *J* = 6.4 Hz, 1H), 4.11–3.98 (m, 4H), 2.56 (s, 1H), 2.51–2.43 (m, 2H), 1.75 (d, *J* = 7.0 Hz, 3H), 1.42–1.20 (m, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.56 (d, J = 5.0 Hz), 131.88, 131.26, 130.53, 130.34, 119.46 (d, J = 186.6 Hz), 70.87 (d, J = 1.3 Hz), 61.71 (d, J = 5.5 Hz), 42.15 (d, J = 21.9 Hz), 18.05, 16.28 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.06 ppm.

MS(ESI) m/z [M+Na]⁺: 297.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 275.1407, found 275.1407.

IR (film): 3363, 2962, 1634, 1260, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{25} = +8.32$ (*c* = 0.510, CHCl₃, 97% ee, *E*/*Z* = 6/1).

HPLC: DAICEL CHIRALPAK IC, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 16.8 min, t_R(minor) = 19.6 min, ee = 97%.



Figure S286, the HPLC spectrum of compound 3ac, related to Table 2



3ad: Procedure A, 51 mg, colorless liquid, 68% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 6.76 (ddt, *J* = 24.1, 17.1, 7.0 Hz, 1H), 5.82–5.62 (m, 2H), 5.50 (dd, *J* = 15.3, 5.9 Hz, 1H), 4.21 (dd, *J* = 12.8, 6.4 Hz, 1H), 4.17–3.95 (m, 4H), 2.55–2.36 (m, 2H), 2.24 (s, 1H), 1.69 (d, *J* = 6.3 Hz, 3H), 1.32 (t, *J* = 7.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.55 (d, J = 4.5 Hz), 132.95, 127.57, 119.50 (d, J = 185.8 Hz), 71.25, 61.69 (d, J = 5.5 Hz), 42.06 (d, J = 21.9 Hz), 17.61, 16.31 (d, J = 6.4 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.02 ppm.

MS(ESI) m/z [M+H]⁺: 249.10.

HRMS(ESI) m/z [M+H]⁺: calcd. 249.1250, found 249.1251.

IR (film): 3386, 2985, 1633, 1259, 1020, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{29} = +5.38$ (*c* = 1.155, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 14.5 min, t_R(minor) = 15.9 min, ee = 95%.



Figure S287, the HPLC spectrum of compound 3ad, related to Table 2



3ae

3ae: Procedure A, 59 mg, colorless liquid, 71% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 6.76 (ddt, *J* = 24.1, 17.1, 7.0 Hz, 1H), 5.83–5.61 (m, 2H), 5.48 (dd, *J* = 15.4, 6.8 Hz, 1H), 4.22 (q, *J* = 6.4 Hz, 1H), 4.17–3.96 (m, 4H), 2.52–2.41 (m, 2H), 2.32 (s, 1H), 2.10–1.89 (m, 2H), 1.45–1.33 (m, 2H), 1.32 (t, *J* = 7.1 Hz, 6H), 0.90 (t, *J* = 7.4 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.59 (d, J = 5.0 Hz), 132.59, 131.81, 119.42 (d, J = 187.0 Hz), 71.24, 61.67 (d, J = 5.2 Hz), 42.17 (d, J = 21.9 Hz), 34.16, 22.17, 16.29 (d, J = 6.5 Hz), 13.61 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.03 ppm.

MS(ESI) m/z [M+H]⁺: 277.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 277.1563, found 277.1563.

IR (film): 3384, 2960, 1635, 1230, 1098, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{29} = +5.67$ (*c* = 1.510, CHCl₃, 93% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 12.8 min, t_R(minor) = 13.8 min, ee = 93%.



Figure S288, the HPLC spectrum of compound 3ae, related to Table 2



3af: Procedure A, 69 mg, colorless liquid, 71% yield.

¹H NMR (400 MHz, CDCl₃) δ 6.76 (ddt, J = 24.1, 17.1, 7.0 Hz, 1H), 5.81–5.61 (m, 2H), 5.51 (dd, J = 15.4, 6.6 Hz, 1H), 4.22 (q, J = 6.3 Hz, 1H), 4.17–3.09 (m, 4H), 3.54 (t, J = 6.6 Hz, 2H), 2.46 (t, J = 6.5 Hz, 2H), 2.25 (s, 1H), 2.17–1.96 (m, 2H), 1.85–1.72 (m, 2H), 1.57–1.45 (m, 2H), 1.32 (t, J = 7.1 Hz, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ 149.41 (d, J = 5.0 Hz), 132.29, 131.79, 119.62 (d, J = 187.1 Hz), 71.10 (d, J = 1.1 Hz), 61.73 (d, J = 5.4 Hz), 44.83, 42.14 (d, J = 22.0 Hz), 31.94, 31.28, 26.20, 16.32 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.97 ppm.

MS(ESI) m/z [M+Na]⁺: 347.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 325.1330, found 325.1332.

IR (film): 3379, 2987, 1635, 1260, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +5.66 (*c* = 0.940, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 18.1 min, t_R(minor) = 20.0 min, ee = 97%.





Figure S289, the HPLC spectrum of compound 3af, related to Table 2



3ag: Procedure A, 59 mg, colorless liquid, 47% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 6.76 (ddt, *J* = 24.1, 17.2, 7.0 Hz, 1H), 5.81–5.61 (m, 2H), 5.48 (dd, *J* = 15.4, 6.8 Hz, 1H), 4.21 (q, *J* = 6.4 Hz, 1H), 4.17–3.96 (m, 4H), 3.60 (t, *J* = 6.3 Hz, 2H), 2.45 (t, *J* = 6.4 Hz, 2H), 2.22 (s, 1H), 2.14–1.96 (m, 2H), 1.57–1.47 (m, 2H), 1.46–1.36 (m, 2H), 1.32 (t, *J* = 7.1 Hz, 6H), 0.89 (s, 9H), 0.05 (s, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.55 (d, J = 5.1 Hz), 132.62, 131.80, 119.51 (d, J = 186.9 Hz), 71.23 (d, J = 1.3 Hz), 62.94, 61.70 (d, J = 5.5 Hz), 42.15 (d, J = 21.9 Hz), 32.26, 31.86, 29.66, 25.93, 25.29, 18.33, 16.31 (d, J = 6.4 Hz), -5.31 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.03 ppm.

MS(ESI) m/z [M+Na]⁺: 443.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 421.2534, found 421.2534.

IR (film): 3381, 2930, 1633, 1255, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +3.45 (*c* = 1.100, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 7/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 19.1 min, t_R(minor) = 20.9 min, ee = 98%.



Figure S290, the HPLC spectrum of compound 3ag, related to Table 2



3ah: Procedure A, 91 mg, colorless liquid, 85% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 6.76 (ddt, *J* = 22.1, 17.1, 7.0 Hz, 1H), 5.81–5.61 (m, 2H), 5.51 (dd, *J* = 15.4, 6.7 Hz, 1H), 4.22 (q, *J* = 6.4 Hz, 1H), 4.17–3.98 (m, 4H), 2.56–2.44 (m, 2H), 2.23–2.01 (m, 7H), 1.62–1.49 (m, 2H), 1.54–1.33 (m, 4H), 1.32 (t, *J* = 7.1 Hz, 6H), 0.91 (t, *J* = 7.1 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.43, 132.29, 131.85 (d, *J* = 2.6 Hz), 119.58 (d, *J* = 187.0 Hz), 80.66, 79.47, 71.19, 61.71 (d, *J* = 5.6 Hz), 42.11 (d, *J* = 21.9 Hz), 31.18, 31.13, 28.42, 21.89, 18.38, 18.18, 16.32 (d, *J* = 6.4 Hz), 13.59 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.97 ppm.

MS(ESI) m/z [M+H]⁺: 357.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 357.2189, found 357.2191.

IR (film): 3379, 2932, 1634, 1275, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29}$ = +3.32 (*c* = 1.110, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 15/1, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 28.5 min, t_R(minor) = 30.6 min, ee = 98%.





Figure S291, the HPLC spectrum of compound 3ah, related to Table 2



3ai: Procedure A, 80 mg, colorless liquid, 68% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 6.76 (ddt, *J* = 22.0, 17.1, 7.0 Hz, 1H), 5.74 (dd, *J* = 21.3, 17.1 Hz, 1H), 5.21 (dd, *J* = 8.6, 0.7 Hz, 1H), 5.19–5.00 (m, 2H), 4.51 (dd, *J* = 14.4, 6.7 Hz, 1H), 4.17–4.00 (m, 4H), 2.55–2.33 (m, 2H), 2.28 (s, 1H), 2.13–1.93 (m, 8H), 1.68 (s, 6H), 1.60 (s, 6H), 1.32 (t, *J* = 7.1 Hz, 6H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.74 (d, J = 4.7 Hz), 139.24, 135.37, 131.29, 126.56, 124.21, 123.58, 119.31 (d, J = 187.0 Hz), 67.04 (d, J = 1.0 Hz), 61.66 (d, J = 5.6 Hz), 42.42 (d, J = 21.7 Hz), 39.62, 39.45, 26.66, 26.29, 25.64, 17.63, 16.68, 16.29 (d, J = 6.5 Hz), 15.96 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.11 ppm.

MS(ESI) m/z [M+Na]⁺: 421.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 399.2659, found 399.2659.

IR (film): 3385, 2927, 1633, 1270, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +2.53 (*c* = 2.675, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 37/3, flow rate: 0.8 mL/min, λ = 207 nm, t_R(major) = 19.3 min, t_R(minor) = 23.1 min, ee = 99%.



Figure S292, the HPLC spectrum of compound 3ai, related to Table 2



3aj: Procedure A, 56 mg, colorless liquid, 52% yield, 15/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹H NMR (400 MHz, CDCl₃) δ 6.77 (ddt, J = 22.1, 17.1, 7.0 Hz, 1H), 5.80–5.60 (m, 2H), 5.48 (dd, J = 15.3, 6.7 Hz, 1H), 5.09 (t, J = 7.1 Hz, 1H), 4.32–4.16 (m, 1H), 4.17–3.95 (m, 4H), 2.56–2.40 (m, 2H), 2.40 (s, 1H), 2.11–1.81 (m, 4H), 1.68 (s, 3H), 1.60 (s, 3H), 1.55–1.45 (m, 1H), 1.40–1.23 (m, 7H), 1.22–1.07 (m, 1H), 0.87 (d, J = 6.6 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.58 (d, J = 4.8 Hz), 132.97, 131.15, 124.64, 119.47 (d, J = 187.1 Hz), 109.99, 71.20 (d, J = 1.2 Hz), 61.69 (d, J = 6.4 Hz), 42.21 (d, J = 21.9 Hz), 39.48, 36.62, 32.43, 25.67, 25.48, 19.32, 17.61, 16.31 (d, J = 6.4 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.04 ppm.

MS(ESI) m/z [M+Na]⁺: 381.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 359.2346, found 359.2346.

IR (film): 3381, 2912, 1633, 1231, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{28}$ = +6.51 (*c* = 2.205, CHCl₃, 15/1 dr).



3aj': Procedure B, 65 mg, colorless liquid, 60% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹H NMR (400 MHz, CDCl₃) δ 6.77 (ddt, J = 22.1, 17.1, 7.0 Hz, 1H), 5.79–5.58 (m, 2H), 5.48 (dd, J = 15.3, 6.8 Hz, 1H), 5.09 (t, J = 7.1 Hz, 1H), 4.29–4.16 (m, 1H), 4.17–4.00 (m, 4H), 2.56–2.36 (m, 2H), 2.38 (s, 1H), 2.12–1.75 (m, 4H), 1.68 (s, 3H), 1.60 (s, 3H), 1.55–1.41 (m, 1H), 1.38–1.24 (m, 7H), 1.22–1.08 (m, 1H), 0.86 (d, J = 6.6 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.56 (d, J = 5.0 Hz), 132.96, 131.30, 131.18, 124.64, 119.46 (d, J = 187.1 Hz), 71.27 (d, J = 1.3 Hz), 61.69 (d, J = 5.2 Hz), 42.22 (d, J = 21.9 Hz), 39.53, 36.67, 32.40, 25.67, 25.48, 19.30, 17.61, 16.31 (d, J = 6.4 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.03 ppm.

MS(ESI) m/z [M+H]⁺: 381.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 359.2346, found 359.2347.

IR (film): 3380, 2964, 1633, 1231, 1028 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28} = -5.73$ (*c* = 1.875, CHCl₃, > 20/1 dr).



3ak: Procedure A, 57 mg, colorless liquid, 58% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹H NMR (400 MHz, CDCl₃) δ 6.73 (ddt, *J* = 24.1, 17.1, 6.9 Hz, 1H), 5.83–5.64 (m, 2H), 4.71 (d, *J* = 9.3 Hz, 2H), 4.15 (t, *J* = 6.6 Hz, 1H), 4.12–4.00 (m, 4H), 2.57–2.35 (m, 2H), 2.30–1.91 (m, 5H), 1.91–1.82 (m, 1H), 1.73 (s, 3H), 1.55–1.37 (m, 1H), 1.38–1.21 (m, 7H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.92 (d, J = 4.5 Hz), 149.54, 138.71, 123.61, 119.09 (d, J = 186.7 Hz), 108.71, 74.40, 61.68 (d, J = 5.4 Hz), 41.19, 39.82 (d, J = 21.9 Hz), 30.39, 27.42, 23.81, 20.69, 16.32 (d, J = 6.4 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.06 ppm.

MS(ESI) m/z [M+H]⁺: 329.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 329.1876, found 329.1877.

IR (film): 3379, 2988, 1636, 1260, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = -25.80$ (*c* = 1.330, CHCl₃, > 20/1 dr).



3ak': Procedure B, 78 mg, colorless liquid, 79% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹H NMR (400 MHz, CDCl₃) δ 6.74 (ddt, *J* = 24.1, 17.2, 7.0 Hz, 1H), 5.82–5.65 (m, 2H), 4.71 (d, *J* = 12.2 Hz, 2H), 4.14 (t, *J* = 6.2 Hz, 1H), 4.16–4.00 (m, 4H), 2.59–2.39 (m, 3H), 2.21–2.05 (m, 3H), 2.02–1.90 (m, 1H), 1.89–1.75 (m, 1H), 1.73 (s, 3H), 1.56–1.49 (m, 1H), 1.36–1.20 (m, 7H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 150.03 (d, J = 4.9 Hz),149.5, 138.49, 122.37, 119.08 (d, J = 187.0 Hz), 108.68, 74.10 (d, J = 1.0 Hz), 61.68 (d, J = 5.4 Hz), 41.05, 40.12 (d, J = 22.0 Hz), 30.27, 27.30, 24.50, 20.73, 16.31 (d, J = 6.3 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.10 ppm.

MS(ESI) m/z [M+H]⁺: 329.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 329.1876, found 329.1876.

IR (film): 3379, 2985, 1642, 1260, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = -38.08$ (*c* = 1.735, CHCl₃, > 20/1 dr).



3al: Procedure A, 87 mg, colorless liquid, 88% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹**H NMR (400 MHz, CDCl₃)** δ 6.78 (ddt, *J* = 23.9, 17.1, 6.8 Hz, 1H), 5.74 (dd, *J* = 21.1, 17.1 Hz, 1H), 5.48 (s, 1H), 4.14 (t, *J* = 6.0 Hz, 1H), 4.16–3.96 (m, 4H), 2.52–2.35 (m, 3H), 2.32–2.19 (m, 3H), 2.16–2.00 (m, 2H), 1.37–1.24 (m, 9H), 1.16 (d, *J* = 8.6 Hz, 1H), 0.82 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.97 (d, J = 4.9 Hz), 149.51, 119.20 (d, J = 187.0 Hz), 118.07, 73.05 (d, J = 1.1 Hz), 61.63 (d, J = 3.8 Hz), 42.05, 40.83, 39.64 (d, J = 22.0 Hz), 37.79, 31.66, 31.01, 26.11, 21.39, 16.31 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.03 ppm.

MS(ESI) m/z [M+H]⁺: 329.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 329.1876, found 329.1876.

IR (film): 3384, 2914, 1635, 1260, 1027, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = -9.99$ (*c* = 1.540, CHCl₃, > 20/1 dr).



3al': Procedure B, 60 mg, colorless liquid, 61% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹**H NMR (400 MHz, CDCl₃)** δ 6.74 (ddt, *J* = 24.0, 17.1, 6.9 Hz, 1H), 5.72 (dd, *J* = 21.2, 17.1 Hz, 1H), 5.48 (s, 1H), 4.14 (t, *J* = 6.5 Hz, 1H), 4.16–4.00 (m, 4H), 2.48–2.34 (m, 3H), 2.31–2.18 (m, 3H), 2.15–2.00 (m, 2H), 1.37–1.24 (m, 9H), 1.12 (d, *J* = 8.6 Hz, 1H), 0.84 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 150.00 (d, J = 4.8 Hz), 149.02, 119.10 (d, J = 187.0 Hz), 118.53, 72.99 (d, J = 0.9 Hz), 61.64 (d, J = 5.4 Hz), 41.82, 40.84, 39.57 (d, J = 22.0 Hz), 37.77, 31.60, 31.03, 26.04, 21.35, 16.30 (d, J = 6.5 Hz) ppm.

³¹P NMR (162 MHz, CDCl₃) δ 18.11 ppm.

MS(ESI) m/z [M+H]⁺: 329.15.

HRMS(ESI) m/z [M+H]⁺:calcd. 329.1876, found 329.1875.

IR (film): 3379, 2988, 1631, 1260, 1028, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{29} = -24.74$ (*c* = 1.870, CHCl₃, > 20/1 dr).



5a: Procedure A, 83 mg, pale green solid, 96% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.66 (s, 1H), 8.06 (d, *J* = 7.5 Hz, 1H), 7.94 (t, *J* = 7.7 Hz, 1H), 7.57–7.47 (m, 1H), 7.36–7.24 (m, 5H), 7.15 (dt, *J* = 15.2, 7.2 Hz, 1H), 6.60 (d, *J* = 15.2 Hz, 1H), 5.24–4.49 (m, 1H), 2.78 (d, *J* = 3.2 Hz, 1H), 2.76–2.66 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.21, 150.15, 146.36, 142.95, 138.25, 129.96, 128.61, 127.96, 127.13, 125.63, 121.91, 72.36, 41.34 ppm.

MS(ESI) m/z [M+H]⁺: 290.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 290.0845, found 290.0846.

IR (film): 3502, 2914, 1630, 1428, 1170, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +32.29 (*c* = 1.050, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 31.9 min, t_R(minor) = 36.1 min, ee = 97%.



Figure S293, the HPLC spectrum of compound 5a, related to Table 3



5b: Procedure A, 81 mg, colorless crystal, 88% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.70 (d, *J* = 4.1 Hz, 1H), 8.07 (d, *J* = 7.9 Hz, 1H), 7.99–7.85 (m, 1H), 7.53 (dd, *J* = 7.1, 5.2 Hz, 1H), 7.30 (dd, *J* = 8.5, 5.4 Hz, 2H), 7.13 (dt, *J* = 15.2, 7.3 Hz, 1H), 7.00 (t, *J* = 8.7 Hz, 2H), 6.60 (d, *J* = 15.2 Hz, 1H), 4.89 (t, *J* = 6.2 Hz, 1H), 2.81–2.62 (m, 2H), 2.49 (s, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 162.31 (d, J = 246.2 Hz), 158.24 , 150.15, 145.86, 138.61 (d, J = 3.2 Hz), 138.25, 130.25, 127.31 (d, J = 8.1 Hz), 127.13, 121.81, 115.48 (d, J = 21.4 Hz), 71.78, 41.43 ppm.

¹⁹F NMR (376 MHz, CDCl₃) δ -112.86 \sim -117.88 (m) ppm.

MS(ESI) m/z [M+Na]⁺: 329.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 308.0751, found 308.0752.

IR (film): 3405, 2921, 1428, 1276 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +21.10 (*c* = 0.250, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 41.4 min, t_R(minor) = 47.7 min, ee = 97%.





Figure S294, the HPLC spectrum of compound 5b, related to Table 3



5c: Procedure A, 90 mg, colorless crystal, 93% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.68 (d, *J* = 4.6 Hz, 1H), 8.06 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.53 (dd, *J* = 7.1, 5.2 Hz, 1H), 7.31–7.21 (m, 4H), 7.12 (dt, *J* = 15.2, 7.3 Hz, 1H), 6.58 (d, *J* = 15.2 Hz, 1H), 4.89 (t, *J* = 6.3 Hz, 1H), 2.78 (s, 1H), 2.69 (t, *J* = 6.6 Hz, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.14, 150.15, 145.78, 141.34, 138.28, 133.59, 130.31, 128.73, 127.18, 127.01, 121.84, 71.69, 41.34 ppm.

MS(ESI) m/z [M+Na]⁺: 345.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 324.0456, found 324.0455.

IR (film): 3494, 2919, 1630, 1453, 1163 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +27.84 (*c* = 0.625, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 27.5 min, t_R(minor) = 29.7 min, ee = 97%.





Figure S295, the HPLC spectrum of compound 5c, related to Table 3



5d: Procedure A, 94 mg, white powder, 85% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.70 (d, *J* = 4.1 Hz, 1H), 8.07 (d, *J* = 7.8 Hz, 1H), 7.96 (td, *J* = 7.8, 1.6 Hz, 1H), 7.54 (dd, *J* = 6.6, 4.8 Hz, 1H), 7.43 (d, *J* = 8.4 Hz, 2H), 7.20 (d, *J* = 8.3 Hz, 2H), 7.12 (dt, *J* = 15.2, 7.3 Hz, 1H), 6.59 (d, *J* = 15.2 Hz, 1H), 4.87 (t, *J* = 6.2 Hz, 1H), 2.69 (t, *J* = 7.0 Hz, 2H), 2.55 (s, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.21, 150.16, 145.61, 141.80, 138.25, 131.69, 130.41, 127.33, 127.15, 121.79, 121.76, 71.78, 41.27 ppm.

MS(ESI) m/z [M+Na]⁺: 389.90.

HRMS(ESI) m/z [M+H]⁺: calcd. 367.9951, found 367.9951.

IR (film): 3490, 2924, 1428, 1262 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +30.84 (*c* = 0.335, CHCl₃, 92% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 29.8 min, t_R(minor) = 31.7 min, ee = 92%.





Figure S296, the HPLC spectrum of compound 5d, related to Table 3



5e: Procedure A, 102 mg, white powder, 82% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.66 (d, *J* = 4.5 Hz, 1H), 8.04 (d, *J* = 7.8 Hz, 1H), 7.95 (td, *J* = 7.7, 1.6 Hz, 1H), 7.61 (d, *J* = 8.3 Hz, 2H), 7.54 (ddd, *J* = 7.5, 4.7, 0.9 Hz, 1H), 7.19–6.99 (m, 3H), 6.56 (d, *J* = 15.2 Hz, 1H), 4.85 (t, *J* = 6.2 Hz, 1H), 3.20 (s, 1H), 2.77–2.56 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 157.99, 150.18, 145.93, 142.67, 138.35, 137.55, 130.21, 127.63, 127.26, 121.92, 93.26, 71.69, 41.25 ppm.

MS(ESI) m/z [M+Na]⁺: 437.70.

HRMS(ESI) m/z [M+H]⁺: calcd. 415.9812, found 415.9812.

IR (film): 3493, 2919, 1630, 1427, 1163 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +28.57 (*c* = 1.835, CHCl₃, 91% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 5/1, flow rate: 0.72 mL/min, λ = 254 nm, t_R(major) = 70.4 min, t_R(minor) = 75.0 min, ee = 91%.





Figure S297, the HPLC spectrum of compound 5e, related to Table 3


5f: Procedure A, 87 mg, white powder, 96% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.67 (d, *J* = 4.2 Hz, 1H), 8.05 (d, *J* = 7.9 Hz, 1H), 7.93 (td, *J* = 7.8, 1.6 Hz, 1H), 7.51 (ddd, *J* = 7.5, 4.7, 0.9 Hz, 1H), 7.25–7.05 (m, 5H), 6.59 (d, *J* = 15.2 Hz, 1H), 4.93–4.75 (m, 1H), 2.84–2.58 (m, 3H), 2.32 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.20, 150.16, 146.53, 140.00, 138.25, 137.64, 129.84, 129.25, 127.11, 125.57, 121.90, 72.21, 41.32, 21.09 ppm.

MS(ESI) m/z [M+Na]⁺: 326.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 304.1002, found 304.1002.

IR (film): 3514, 2921, 1630, 1428, 1270 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +35.05 (*c* = 1.165, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 36.0 min, t_R(minor) = 39.0 min, ee = 97%.



Figure S298, the HPLC spectrum of compound 5f, related to Table 3



5g: Procedure A, 94 mg, colorless liquid, 91% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.67 (d, *J* = 4.6 Hz, 1H), 8.07 (d, *J* = 7.9 Hz, 1H), 7.94 (td, *J* = 7.8, 1.6 Hz, 1H), 7.51 (ddd, *J* = 7.6, 4.8, 0.9 Hz, 1H), 7.35 (d, *J* = 8.4 Hz, 2H), 7.25 (d, *J* = 8.3 Hz, 2H), 7.22–7.06 (m, 1H), 6.62 (d, *J* = 15.2 Hz, 1H), 4.89–4.78 (m, 1H), 2.91–2.59 (m, 3H), 1.30 (s, 9H).ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.19, 150.92, 150.18, 146.67, 139.99, 138.28, 129.78, 127.14, 125.50, 125.40, 121.94, 72.13, 41.24, 34.52, 31.30 ppm.

MS(ESI) m/z [M+Na]⁺: 368.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 346.1471, found 346.1469.

IR (film): 3507, 2989, 1461, 1260, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +28.35 (*c* = 1.790, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK IG-3, hexane/*i*-PrOH = 3/1, flow rate: 0.6 mL/min, λ = 254 nm, t_R(major) = 41.0 min, t_R(minor) = 38.7 min, ee = 95%.





Figure S299, the HPLC spectrum of compound 5g, related to Table 3



5h: Procedure A, 91 mg, pale green liquid, 90% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.70 (d, *J* = 4.6 Hz, 1H), 8.07 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.56–7.49 (m, 1H), 7.31–7.18 (m, 4H), 7.19–7.06 (m, 1H), 6.60 (d, *J* = 15.2 Hz, 1H), 4.86 (t, *J* = 6.2 Hz, 1H), 2.76–2.63 (m, 2H), 2.47 (s, 3H), 2.43 (s, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 157.86, 150.15, 146.45, 139.97, 138.40, 137.82, 129.86, 127.29, 126.45, 126.27, 121.97, 71.73, 41.16, 15.68 ppm.

MS(ESI) m/z [M+Na]⁺: 357.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 336.0723, found 336.0723.

IR (film): 3393, 2921, 1428, 1270 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +23.63 (*c* = 0.420, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 11/5, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 48.7 min, t_R(minor) = 52.8 min, ee = 97%.





Figure S300, the HPLC spectrum of compound 5h, related to Table 3



5am: Procedure A, 79 mg, colorless liquid, 76% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.62 (d, *J* = 4.3 Hz, 1H), 8.02 (d, *J* = 7.8 Hz, 1H), 7.97–7.87 (m, 3H), 7.51 (dd, *J* = 6.9, 4.9 Hz, 1H), 7.38 (d, *J* = 8.2 Hz, 2H), 7.13 (dt, *J* = 15.2, 7.2 Hz, 1H), 6.56 (d, *J* = 15.2 Hz, 1H), 4.96 (t, *J* = 6.1 Hz, 1H), 3.89 (s, 3H), 3.65 (s, 1H), 2.70 (t, *J* = 6.7 Hz, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 166.81, 157.87, 150.13, 148.20, 146.01, 138.39, 130.15, 129.76, 129.35, 127.28, 125.62, 121.96, 71.68, 52.14, 41.21 ppm.

MS(ESI) m/z [M+Na]⁺: 369.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 348.0900, found 348.0900.

IR (film): 3493, 2952, 1717, 1429, 1026, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +28.54 (*c* = 0.289, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 53.8 min, t_R(minor) = 59.7 min, ee = 98%.



Figure S301, the HPLC spectrum of compound 5am, related to Table 3



5k: Procedure A, 88 mg, colorless liquid, 95% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.70 (d, *J* = 4.4 Hz, 1H), 8.08 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.5 Hz, 1H), 7.60–7.48 (m, 1H), 7.46 (td, *J* = 7.5, 1.2 Hz, 1H), 7.29–7.09 (m, 3H), 7.03–6.96 (m, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.24–5.18 (m, 1H), 2.86–2.66 (m, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 160.59, 158.18 (d, J = 8.1 Hz), 150.16, 145.98, 138.26, 130.20, 129.87 (d, J = 13.2 Hz), 129.31 (d, J = 8.3 Hz), 127.13, 127.00 (d, J = 4.2 Hz), 124.43 (d, J = 3.5 Hz), 121.87, 115.34 (d, J = 21.6 Hz), 66.42 (d, J = 2.5 Hz), 40.03 ppm.

¹⁹**F NMR (376 MHz, CDCl₃)** δ -119.29~-119.40 (m) ppm.

MS(ESI) m/z [M+Na]⁺: 329.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 308.0751, found 308.0751.

IR (film): 3490, 2989, 1456, 1275, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +35.66 (*c* = 0.670, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 11/5, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 30.0 min, t_R(minor) = 28.7 min, ee = 98%.



etector A 2	254nm			
_				1
1	Peak#	Ret. Time	Area%	
	1	28.684	1.149	
	2	29.980	98.851	~

Figure S302, the HPLC spectrum of compound 5k, related to Table 3



5an: Procedure A, 94 mg, colorless liquid, 85% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.69 (d, *J* = 4.1 Hz, 1H), 8.08 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.58–7.43 (m, 3H), 7.36–7.16 (m, 2H), 7.13 (td, *J* = 7.8, 1.6 Hz, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.33–5.18 (m, 1H), 2.96 (s, 1H), 2.86–2.77 (m, 1H), 2.66–2.55 (m, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.21, 150.17, 146.21, 141.85, 138.28, 132.68, 130.08, 129.20, 127.84, 127.18, 127.15, 121.91, 121.47, 71.07, 39.61 ppm.

MS(ESI) m/z [M+Na]⁺: 389.85.

HRMS(ESI) m/z [M+H]⁺: calcd. 367.9951, found 367.9951.

IR (film): 3493, 2960, 1632, 1428, 1198 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +69.81 (*c* = 1.070, CHCl₃, 92% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 28.3 min, t_R(minor) = 30.3 min, ee = 92%.



Figure S303, the HPLC spectrum of compound 5an, related to Table 3



5ao: Procedure A, 89 mg, colorless liquid, 98% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.71 (d, *J* = 4.3 Hz, 1H), 8.09 (d, *J* = 7.8 Hz, 1H), 7.95 (td, *J* = 7.8, 1.4 Hz, 1H), 7.53 (dd, *J* = 7.0, 4.8 Hz, 1H), 7.46 (d, *J* = 6.9 Hz, 1H), 7.24–7.09 (m, 4H), 6.64 (d, *J* = 15.2 Hz, 1H), 5.12 (t, *J* = 6.2 Hz, 1H), 2.68 (t, *J* = 6.4 Hz, 2H), 2.36 (s, 1H), 2.31 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.30, 150.18, 146.49, 140.97, 138.23, 134.13, 130.55, 129.94, 127.71, 127.10, 126.48, 124.99, 121.84, 68.79, 40.16, 18.99 ppm.

MS(ESI) m/z [M+Na]⁺: 326.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 304.1002, found 304.1003.

IR (film): 3405, 2918, 1462, 1276, cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +40.80 (*c* = 0.345, CHCl₃, 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 32.2 min, t_R(minor) = 40.8 min, ee = 99%.



Figure S304, the HPLC spectrum of compound 5ao, related to Table 3



5I: Procedure A, 87 mg, colorless liquid, 81% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.71 (d, *J* = 4.2 Hz, 1H), 8.10 (d, *J* = 7.9 Hz, 1H), 7.96 (td, *J* = 7.8, 1.6 Hz, 1H), 7.78 (d, *J* = 7.9 Hz, 1H), 7.67–7.49 (m, 3H), 7.39 (t, *J* = 7.6 Hz, 1H), 7.21 (dt, *J* = 15.2, 7.4 Hz, 1H), 6.65 (d, *J* = 15.2 Hz, 1H), 5.35–5.25 (m, 1H), 2.77–2.56 (m, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.29, 150.16, 145.91, 141.98, 138.25, 132.42, 130.28, 127.93, 127.40, 127.13, 125.61, 125.55, 122.82, 121.83, 67.73, 41.33 ppm.

¹⁹F NMR (376 MHz, CDCl₃) δ -58.23 ppm.

MS(ESI) m/z [M+Na]⁺: 379.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 358.0719, found 358.0718.

IR (film): 3494, 2925, 1132 cm⁻¹.

Optical rotation: $[\alpha]_D^{25} = +54.75$ (*c* = 2.750, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 19.9 min, t_R(minor) = 21.4 min, ee = 97%.





Figure S305, the HPLC spectrum of compound 5l, related to Table 3



5ap: Procedure A, 74 mg, colorless liquid, 80% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.65 (d, *J* = 4.2 Hz, 1H), 8.05 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.57–7.47 (m, 1H), 7.32–7.22 (m, 1H), 7.19–7.01 (m, 3H), 6.93 (td, *J* = 8.3, 2.2 Hz, 1H), 6.58 (d, *J* = 15.2 Hz, 1H), 4.96–4.86 (m, 1H), 3.20 (d, *J* = 3.7 Hz, 1H), 2.80–2.62 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 162.87 (d, J = 246.5 Hz), 158.09, 150.15, 145.91, 145.66 (d, J = 6.8 Hz), 138.32, 130.24, 130.13 (d, J = 8.2 Hz), 127.19, 121.89, 121.22 (d, J = 2.9 Hz), 114.69 (d, J = 21.1 Hz), 112.60 (d, J = 22.0 Hz), 71.66, 41.27 ppm.

¹⁹**F NMR (376 MHz, CDCl₃)** δ -112.27~-112.39 (m) ppm.

MS(ESI) m/z [M+Na]⁺: 330.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 308.0751, found 308.0751.

IR (film): 3316, 2915, 1428, 1232, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +36.50 (*c* = 1.355, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 40.2 min, t_R(minor) = 43.0 min, ee = 97%.



Figure S306, the HPLC spectrum of compound 5ap, related to Table 3



5n: Procedure A, 79 mg, colorless liquid, 81% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.70 (d, *J* = 4.6 Hz, 1H), 8.08 (d, *J* = 7.9 Hz, 1H), 7.96 (td, *J* = 7.8, 1.5 Hz, 1H), 7.59–7.50 (m, 1H), 7.34 (s, 1H), 7.29–7.08 (m, 4H), 6.62 (d, *J* = 15.2 Hz, 1H), 4.96–4.85 (m, 1H), 2.71 (t, *J* = 6.7 Hz, 2H), 2.63 (d, *J* = 3.5 Hz, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.21, 150.16, 145.67, 144.92, 138.28, 134.56, 130.39, 129.91, 128.09, 127.14, 125.79, 123.77, 121.81, 71.75, 41.28 ppm.

MS(ESI) m/z [M+H]⁺: 323.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 324.0456, found 324.0457.

IR (film): 3396, 2924, 1428, 1270 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +37.10 (*c* = 0.300, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 26.8 min, t_R(minor) = 31.1 min, ee = 98%.



Figure S307, the HPLC spectrum of compound 5n, related to Table 3



50: Procedure A, 84 mg, colorless liquid, 76% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.71 (d, *J* = 4.6 Hz, 1H), 8.09 (d, *J* = 7.9 Hz, 1H), 7.96 (td, *J* = 7.8, 1.6 Hz, 1H), 7.59–7.46 (m, 2H), 7.40 (d, *J* = 7.7 Hz, 1H), 7.27–7.05 (m, 3H), 6.62 (d, *J* = 15.3 Hz, 1H), 4.87 (t, *J* = 6.3 Hz, 1H), 2.71 (t, *J* = 6.8 Hz, 2H), 2.53 (s, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.23, 150.16, 145.63, 145.15, 138.28, 131.05, 130.41, 130.20, 128.71, 127.14, 124.25, 122.76, 121.80, 71.71, 41.30 ppm.

MS(ESI) m/z [M+H]⁺: 367.90.

HRMS(ESI) m/z [M+H]⁺: calcd. 367.9951, found 367.9951.

IR (film): 3485, 2961, 1428, 1261, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +33.07 (*c* = 1.850, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 30.0 min, t_R(minor) = 35.5 min, ee = 98%.





Figure S308, the HPLC spectrum of compound 50, related to Table 3



5aq: Procedure A, 90 mg, colorless liquid, 88% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.61 (d, J = 4.7 Hz, 1H), 8.01 (d, J = 7.9 Hz, 1H), 7.90–7.74 (m, 5H), 7.52–7.38 (m, 4H), 7.19 (dt, J = 15.2, 7.3 Hz, 1H), 6.61 (d, J = 15.2 Hz, 1H), 5.10–4.99 (m, 1H), 2.91–2.73 (m, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.24, 150.08, 146.04, 140.15, 138.10, 133.15, 133.05, 130.18, 128.57, 127.98, 127.67, 127.00, 126.33, 126.10, 124.51, 123.45, 121.70, 72.59, 41.23 ppm.

MS(ESI) m/z [M+Na]⁺: 361.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 340.1002, found 340.1002.

IR (film): 3494, 2925, 1427, 1162 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +30.68 (*c* = 2.200, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 46.5 min, t_R(minor) = 50.4 min, ee = 97%.



Figure S309, the HPLC spectrum of compound 5aq, related to Table 3



5p: Procedure A, 99 mg, colorless liquid, 97% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.61 (d, *J* = 4.6 Hz, 1H), 8.04–7.94 (m, 2H), 7.92–7.80 (m, 2H), 7.73 (d, *J* = 8.2 Hz, 1H), 7.61 (d, *J* = 7.1 Hz, 1H), 7.50–7.43 (m, 3H), 7.40 (t, *J* = 7.7 Hz, 1H), 7.25 (dt, *J* = 15.2, 7.2 Hz, 1H), 6.60 (d, *J* = 15.2 Hz, 1H), 5.69–5.58 (m, 1H), 3.24 (s, 1H), 2.93–2.67 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.06, 150.14, 146.79, 138.58, 138.29, 133.69, 129.83, 129.74, 129.00, 128.27, 127.16, 126.30, 125.66, 125.42, 122.96, 122.62, 121.92, 69.02, 40.37 ppm.

MS(ESI) m/z [M+Na]⁺: 362.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 340.1002, found 340.1003.

IR (film): 3493, 2989, 1427, 1275 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26} = +60.65$ (*c* = 2.035, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK IG-3, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 108.8 min, t_R(minor) = 92.3 min, ee = 95%.





Figure S310, the HPLC spectrum of compound 5p, related to Table 3



5q: Procedure A, 70 mg, colorless liquid, 81% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.70 (d, *J* = 4.2 Hz, 1H), 8.08 (d, *J* = 7.8 Hz, 1H), 7.95 (td, *J* = 7.8, 1.5 Hz, 1H), 7.53 (dd, *J* = 7.2, 5.0 Hz, 1H), 7.35 (s, 1H), 7.21–6.98 (m, 1H), 6.65 (d, *J* = 15.3 Hz, 1H), 6.39–6.00 (m, 2H), 4.90 (t, *J* = 6.0 Hz, 1H), 2.85 (t, *J* = 6.8 Hz, 2H), 2.65 (s, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.18, 154.82, 150.15, 145.47, 142.28, 138.26, 130.22, 127.15, 121.89, 110.27, 106.62, 65.89, 37.80 ppm.

MS(ESI) m/z [M+Na]⁺: 301.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 280.0638, found 280.0639.

IR (film): 3349, 2924, 1631, 1429, 1163 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +21.31 (*c* = 0.535, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 61.1 min, t_R(minor) = 67.4 min, ee = 98%.





Figure S311, the HPLC spectrum of compound 5q, related to Table 3



5r: Procedure A, 74 mg, colorless liquid, 84% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.68 (d, *J* = 4.4 Hz, 1H), 8.07 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.5 Hz, 1H), 7.56–7.45 (m, 1H), 7.25–7.20 (m, 1H), 7.15 (dt, *J* = 15.2, 7.2 Hz, 1H), 7.00–6.88 (m, 2H), 6.64 (d, *J* = 15.2 Hz, 1H), 5.14 (t, *J* = 6.3 Hz, 1H), 2.99 (s, 1H), 2.89–2.71 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.13, 150.16, 146.71, 145.61, 138.29, 130.30, 127.16, 126.79, 124.96, 124.03, 121.91, 68.34, 41.39 ppm.

MS(ESI) m/z [M+Na]⁺: 317.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 296.0410, found 296.0410.

IR (film): 3392, 2922, 1630, 1428, 1238 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +21.45 (*c* = 0.735, CHCl₃, 98% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 60.7 min, t_R(minor) = 70.8 min, ee = 98%.



Figure S312, the HPLC spectrum of compound 5r, related to Table 3



5ar: Procedure A, 67 mg, colorless liquid, 76% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.66 (d, *J* = 4.4 Hz, 1H), 8.06 (d, *J* = 7.8 Hz, 1H), 7.94 (td, *J* = 7.8, 1.5 Hz, 1H), 7.60–7.44 (m, 1H), 7.38–7.18 (m, 1H), 7.23–7.10 (m, 2H), 7.05 (d, *J* = 4.9 Hz, 1H), 6.60 (d, *J* = 15.2 Hz, 1H), 4.98 (t, *J* = 6.0 Hz, 1H), 2.99 (s, 1H), 2.75 (t, *J* = 6.5 Hz, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.14, 150.16, 146.19, 144.41, 138.30, 130.03, 127.18, 126.52, 125.30, 121.91, 121.20, 68.56, 40.64 ppm.

MS(ESI) m/z [M+Na]⁺: 317.95.

HRMS(ESI) m/z [M+H]⁺: calcd. 296.0410, found 296.0410.

IR (film): 3493, 3005, 1427, 1276 cm⁻¹.

Optical rotation: $[\alpha]_D^{26} = +30.34$ (*c* = 1.115, CHCl₃, > 99% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 40.5 min, t_R(minor) = 48.1 min, ee = > 99%.



Figure S313, the HPLC spectrum of compound 5ar, related to Table 3



5u: Procedure A, 88 mg, colorless liquid, 85% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.63 (d, *J* = 4.6 Hz, 1H), 8.02 (d, *J* = 7.9 Hz, 1H), 7.90 (td, *J* = 7.8, 1.5 Hz, 1H), 7.86–7.76 (m, 2H), 7.50–7.45 (m, 1H), 7.39–7.30 (m, 3H), 7.28–7.16 (m, 1H), 6.60 (d, *J* = 15.2 Hz, 1H), 5.31–5.20 (m, 1H), 3.25 (s, 1H), 2.96–2.76 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.04, 150.14, 146.24, 140.89, 138.30, 137.95, 136.68, 130.04, 127.18, 124.56, 124.20, 122.99, 122.85, 121.95, 121.91, 67.87, 39.40 ppm.

MS(ESI) m/z [M+Na]⁺: 367.95.

150

HRMS(ESI) m/z [M+H]⁺: calcd. 346.0566, found 346.0568.

IR (film): 3493, 3054, 1428, 1276, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +44.56 (*c* = 1.435, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK IE, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 76.5 min, t_R(minor) = 72.0 min, ee = 95%.



2

76.507

97.404

95.0 mi

Figure S314, the HPLC spectrum of compound 5u, related to Table 3

77.5



5as: Procedure A, 100 mg, colorless liquid, 78% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.66 (d, *J* = 4.5 Hz, 1H), 8.13 (d, *J* = 7.7 Hz, 1H), 8.03 (d, *J* = 7.8 Hz, 1H), 7.91 (t, *J* = 7.7 Hz, 1H), 7.60 (d, *J* = 7.8 Hz, 1H), 7.54 (s, 1H), 7.49 (dd, *J* = 7.5, 4.7 Hz, 1H), 7.32 (t, *J* = 7.7 Hz, 1H), 7.27–7.14 (m, 2H), 6.66 (d, *J* = 15.2 Hz, 1H), 5.16 (t, *J* = 6.2 Hz, 1H), 2.91 (t, *J* = 6.7 Hz, 2H), 2.77 (s, 1H), 1.66 (s, 9H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.16, 150.15, 149.52, 146.18, 138.23, 135.79, 130.03, 127.98, 127.11, 124.70, 122.70, 122.67, 122.46, 121.80, 119.48, 115.44, 83.96, 66.21, 39.56, 28.16 ppm.

MS(ESI) m/z [M+Na]⁺: 451.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 429.1479, found 429.1479.

IR (film): 3507, 2979, 1731, 1428, 1254 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +23.31 (*c* = 1.225, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK IG-3, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 82.7 min, t_R(minor) = 67.5 min, ee = 97%.



300				
200	Peak#	Ret. Time	Area%	
20	1	67.541	1.729	
50 - 00 -	2	82.683	98.271	
50				
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Figure S315, the HPLC spectrum of compound 5as, related to Table 3



5x: Procedure A, 56 mg, colorless liquid, 46% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.52 (d, *J* = 4.6 Hz, 1H), 8.06–7.97 (m, 2H), 7.94 (d, *J* = 7.9 Hz, 1H), 7.72 (td, *J* = 7.8, 1.6 Hz, 1H), 7.46 (t, *J* = 7.6 Hz, 1H), 7.40–7.35 (m, 2H), 7.35–7.28 (m, 2H), 7.27–7.10 (m, 2H), 6.57 (d, *J* = 15.2 Hz, 1H), 5.11–4.92 (m, 1H), 4.31 (q, *J* = 7.2 Hz, 2H), 2.97 (s, 1H), 2.88–2.68 (m, 2H), 1.39 (t, *J* = 7.2 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.09, 150.01, 146.73, 140.22, 139.56, 138.09, 133.57, 129.76, 126.94, 125.84, 123.49, 122.78, 122.69, 121.76, 120.48, 118.91, 117.67, 109.99, 108.55, 72.93, 41.76, 37.56, 13.82 ppm.

MS(ESI) m/z [M+Na]⁺: 429.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 407.1351, found 407.1352.

IR (film): 3506, 2977, 1427, 1233, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +29.12 (*c* = 2.140, CHCl₃, 93% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 98.2 min, t_R(minor) = 89.8 min, ee = 93%.



Γ	Poak#	Rot Time	Area%	
ļ	rean#	Ret. Time	Alea/0	
	1	89.842	3.697	
	2	98.215	96.303	
				T
				Λ

Figure S316, the HPLC spectrum of compound 5x, related to Table 3



5y: Procedure A, 79 mg, pale green solid, 84% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.63 (d, *J* = 4.1 Hz, 1H), 8.05 (d, *J* = 7.9 Hz, 1H), 7.88 (td, *J* = 7.8, 1.6 Hz, 1H), 7.45 (ddd, *J* = 7.6, 4.7, 0.9 Hz, 1H), 7.35–7.09 (m, 6H), 6.71–6.51 (m, 2H), 6.18 (dd, *J* = 15.9, 6.5 Hz, 1H), 4.55–4.45 (m, 1H), 2.92 (s, 1H), 2.68–2.52 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.09, 150.15, 146.29, 138.28, 136.17, 131.10, 130.50, 129.97, 128.56, 127.87, 127.16, 126.53, 121.90, 70.75, 39.59 ppm.

MS(ESI) m/z [M+Na]⁺: 338.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 316.1002, found 316.1003.

IR (film): 3494, 2922, 1428, 1276 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26}$ = +14.38 (*c* = 1.910, CHCl₃, 94% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 37.6 min, t_R(minor) = 42.1 min, ee = 94%.





Figure S317, the HPLC spectrum of compound 5y, related to Table 3



5z: Procedure A, 50 mg, pale green solid, 51% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.66 (d, *J* = 4.4 Hz, 1H), 8.07 (d, *J* = 7.9 Hz, 1H), 7.89 (td, *J* = 7.8, 1.6 Hz, 1H), 7.47 (dd, *J* = 6.8, 4.8 Hz, 1H), 7.41–7.25 (m, 2H), 7.23–7.13 (m, 4H), 6.68 (d, *J* = 15.2 Hz, 1H), 6.52 (s, 1H), 4.38 (t, *J* = 6.0 Hz, 1H), 2.75–2.55 (m, 2H), 2.10 (s, 1H), 1.85 (d, *J* = 1.1 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.35, 150.15, 146.37, 138.47, 138.15, 136.92, 129.80, 128.92, 128.12, 127.03, 126.69, 126.58, 121.77, 75.69, 37.65, 13.54 ppm.

MS(ESI) m/z [M+Na]⁺: 352.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 330.1158, found 330.1159.

IR (film): 3305, 2921, 1427, 1163 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = -1.98 (*c* = 0.250, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK IG-3, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 48.5 min, t_R(minor) = 39.7 min, ee = 95%.



Figure S318, the HPLC spectrum of compound 5z, related to Table 3



5aa: Procedure A, 77 mg, yellow liquid, 80% yield.

¹H NMR (400 MHz, CDCl₃) δ 8.65 (d, *J* = 4.3 Hz, 1H), 8.06 (d, *J* = 7.8 Hz, 1H), 7.89 (td, *J* = 7.8, 1.5 Hz, 1H), 7.47 (dd, *J* = 7.2, 5.0 Hz, 1H), 7.23–7.07 (m, 2H), 7.02–6.86 (m, 2H), 6.75–6.59 (m, 2H), 6.00 (dd, *J* = 15.7, 6.4 Hz, 1H), 4.55–4.35 (m, 1H), 2.93 (s, 1H), 2.66–2.47 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.07, 150.17, 146.13, 141.22, 138.28, 130.03, 129.93, 127.40, 127.17, 126.32, 124.64, 124.32, 121.91, 70.47, 39.50 ppm.

MS(ESI) m/z [M+Na]⁺: 343.95.

250

HRMS(ESI) m/z [M+H]⁺: calcd. 322.0566, found 322.0566.

IR (film): 3494, 2923, 1428, 1249 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +14.19 (*c* = 1.445, CHCl₃, 92% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 39.6 min, t_R(minor) = 49.5 min, ee = 92%.



Figure S319, the HPLC spectrum of compound 5aa, related to Table 3

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5ad: Procedure A, 38 mg, colorless liquid, 50% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.0 Hz, 1H), 8.10 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.53 (dd, *J* = 6.7, 4.8 Hz, 1H), 7.13 (dt, *J* = 15.2, 7.3 Hz, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.82–5.62 (m, 1H), 5.55–5.46 (m, 1H), 4.35–4.20 (m, 1H), 2.63–2.43 (m, 2H), 2.06 (s, 1H), 1.68 (d, *J* = 5.7 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.33, 150.17, 146.40, 138.24, 132.37, 129.73, 128.19, 127.10, 121.83, 70.91, 39.50, 17.61 ppm.

MS(ESI) m/z [M+Na]⁺: 276.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 254.0845, found 254.0846.

IR (film): 3514, 2918, 1428, 1276, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27}$ = +10.63 (*c* = 0.600, CHCl₃, 93% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/*i*-PrOH = 3/1, flow rate: 0.6 mL/min, λ = 254 nm, t_R(major) = 37.1 min, t_R(minor) = 43.2 min, ee = 93%.





Figure S320, the HPLC spectrum of compound 5ad, related to Table 3



5ae: Procedure A, 59 mg, colorless liquid, 70% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.7 Hz, 1H), 8.10 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.7 Hz, 1H), 7.52 (ddd, *J* = 7.6, 4.7, 1.0 Hz, 1H), 7.13 (dt, *J* = 15.2, 7.3 Hz, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.78–5.62 (m, 1H), 5.47 (dd, *J* = 15.4, 6.9 Hz, 1H), 4.37–4.17 (m, 1H), 2.57–2.49 (m, 2H), 2.06–1.93 (m, 3H), 1.42–1.32 (m, 2H), 0.88 (t, *J* = 7.4 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.38, 150.17, 146.34, 138.21, 133.31, 131.20, 129.73, 127.07, 121.80, 70.97, 39.57, 34.12, 22.11, 13.61 ppm.

MS(ESI) m/z [M+Na]⁺: 304.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 282.1158, found 282.1160.

IR (film): 3405, 2957, 1428, 1170, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{27} = +8.41$ (*c* = 0.560, CHCl₃, 97% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.6 mL/min, λ = 254 nm, t_R(major) = 30.6 min, t_R(minor) = 34.8 min, ee = 97%.





Figure S321, the HPLC spectrum of compound 5ae, related to Table 3



5at: Procedure A, 90 mg, colorless liquid, 88% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.70 (d, J = 4.1 Hz, 1H), 8.09 (d, J = 7.9 Hz, 1H), 7.92 (td, J = 7.8, 1.6 Hz, 1H), 7.51–7.45 (m, 1H), 7.42–7.36 (m, 2H), 7.34–7.30 (m, 2H), 7.28–7.21 (m, 1H), 7.16 (dt, J = 15.2, 7.3 Hz, 1H), 6.77–6.60 (m, 2H), 6.54 (d, J = 15.7 Hz, 1H), 6.41 (dd, J = 15.2, 10.4 Hz, 1H), 5.80 (dd, J = 15.2, 6.6 Hz, 1H), 4.54–4.36 (m, 1H), 2.66–2.50 (m, 2H), 2.21 (s, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.29, 150.18, 145.97, 138.24, 136.85, 134.13, 133.61, 131.70, 130.09, 128.62, 127.79, 127.58, 127.10, 126.41, 121.84, 70.59, 39.53 ppm.

MS(ESI) m/z [M+Na]⁺: 364.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 342.1158, found 342.1158.

IR (film): 3492, 2924, 1630, 1450, 1162 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{28}$ = +13.92 (*c* = 1.930, CHCl₃, 96% ee).

HPLC: DAICEL CHIRALPAK IBN-3, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 44.2 min, t_R(minor) = 80.0 min, ee = 96%.





Figure S322, the HPLC spectrum of compound 5at, related to Table 3



5au: Procedure A, 88 mg, colorless liquid, 80% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.1 Hz, 1H), 8.10 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.53 (ddd, *J* = 7.6, 4.7, 1.0 Hz, 1H), 7.47–7.37 (m, 2H), 7.36–7.29 (m, 3H), 7.14 (dt, *J* = 15.2, 7.3 Hz, 1H), 6.71–6.53 (m, 2H), 6.34 (dd, *J* = 15.2, 10.9 Hz, 1H), 5.81 (dd, *J* = 15.3, 7.8 Hz, 2H), 4.53–4.36 (m, 1H), 2.66–2.20 (m, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.17, 150.20, 145.89, 140.28, 138.33, 136.38, 131.44, 130.40, 130.15, 128.33, 128.27, 127.21, 123.17, 121.90, 112.41, 92.64, 88.51, 70.17, 39.41 ppm.

MS(ESI) m/z [M+Na]⁺: 388.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 366.1158, found 366.1158.

IR (film): 3493, 2989, 1428, 1275, 750 cm⁻¹.

Optical rotation: $[\alpha]_D^{25} = +33.73$ (*c* = 0.900, CHCl₃, 94% ee).

HPLC: DAICEL CHIRALPAK ID, hexane/i-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 35.2 min, t_R(minor) = 38.9 min, ee = 94%.



Figure S323, the HPLC spectrum of compound 5au, related to Table 3



5ah: Procedure A, 76 mg, colorless liquid, 70% yield.

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.5 Hz, 1H), 8.10 (d, *J* = 7.8 Hz, 1H), 7.96 (td, *J* = 7.8, 1.6 Hz, 1H), 7.53 (dd, *J* = 7.0, 4.8 Hz, 1H), 7.13 (dt, *J* = 15.2, 7.2 Hz, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.77–5.62 (m, 1H), 5.50 (dd, *J* = 15.4, 6.7 Hz, 1H), 4.36–4.28 (m, 1H), 2.52 (t, *J* = 6.5 Hz, 2H), 2.38 (s, 1H), 2.20–2.05 (m, 6H), 1.61–1.32 (m, 6H), 0.90 (t, *J* = 7.1 Hz, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.27, 150.17, 146.42, 138.27, 132.30, 131.76, 129.69, 127.13, 121.85, 80.71, 79.46, 70.80, 39.57, 31.17, 31.08, 28.31, 21.89, 18.38, 18.16, 13.60 ppm.

MS(ESI) m/z [M+Na]⁺: 384.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 362.1784, found 362.1784.

IR (film): 3514, 2931, 1428, 1276 cm⁻¹.

Optical rotation: $[\alpha]_D^{26} = +7.74$ (*c* = 1.625, CHCl₃, 95% ee).

HPLC: DAICEL CHIRALPAK IG-3, hexane/*i*-PrOH = 3/1, flow rate: 0.8 mL/min, λ = 254 nm, t_R(major) = 22.8 min, t_R(minor) = 21.8 min, ee = 95%.





Figure S324, the HPLC spectrum of compound 5ah, related to Table 3



5ak: Procedure A, 56 mg, colorless liquid, 56% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.1 Hz, 1H), 8.09 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.52 (ddd, *J* = 7.6, 4.7, 0.9 Hz, 1H), 7.10 (dt, *J* = 15.2, 7.2 Hz, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.72 (d, *J* = 4.0 Hz, 1H), 4.70 (d, *J* = 15.9 Hz, 2H), 4.20 (t, *J* = 6.4 Hz, 1H), 2.65–2.44 (m, 2H), 2.21–1.80 (m, 7H), 1.72 (s, 3H), 1.41 (m, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.37, 150.17, 149.42, 146.76, 138.28, 138.22, 129.40, 127.09, 123.97, 121.81, 108.78, 73.91, 41.00, 37.40, 30.28, 27.32, 23.87, 20.73 ppm.

MS(ESI) m/z [M+Na]⁺: 356.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 334.1471, found 334.1471.

IR (film): 3513, 2920, 1642, 1453, 1163 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{26} = -25.34$ (*c* = 1.025, CHCl₃, > 20/1 dr).



5ak': Procedure B, 60 mg, colorless liquid, 60% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.6 Hz, 1H), 8.10 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.56–7.44 (m, 1H), 7.10 (dt, *J* = 14.9, 7.3 Hz, 1H), 6.63 (d, *J* = 15.2 Hz, 1H), 5.72 (s, 1H), 4.80–4.61 (m, 2H), 4.20 (t, *J* = 5.9 Hz, 1H), 2.59–2.49 (m, 2H), 2.21–1.79 (m, 7H), 1.72 (s, 3H), 1.50–1.35 (m, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.43, 150.17, 149.31, 146.67, 138.20, 137.98, 129.55, 127.06, 123.02, 121.78, 108.80, 73.84, 40.89, 37.64, 30.17, 27.17, 24.50, 20.79 ppm.

MS(ESI) m/z [M+Na]⁺: 356.00.

HRMS(ESI) m/z [M+H]⁺: calcd. 334.1471, found 334.1472.

IR (film): 3514, 2921, 1642, 1428, 1270 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = -33.51$ (*c* = 0.480, CHCl₃, > 20/1 dr).



5al: Procedure A, 68 mg, colorless liquid, 68% yield, > 20/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹**H NMR (400 MHz, CDCl₃)** δ 8.71 (d, *J* = 4.4 Hz, 1H), 8.08 (d, *J* = 7.9 Hz, 1H), 7.95 (td, *J* = 7.8, 1.6 Hz, 1H), 7.55–7.50 (m, 1H), 7.18–7.09 (m, 1H), 6.63 (d, *J* = 15.3 Hz, 1H), 5.49 (d, *J* = 1.0 Hz, 1H), 4.20 (t, *J* = 6.0 Hz, 1H), 2.53–2.44 (m, 2H), 2.43–2.33 (m, 1H), 2.29–2.14 (m, 4H), 2.08 (s, 1H), 1.27 (s, 3H), 1.10 (d, *J* = 8.7 Hz, 1H), 0.76 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.34, 150.16, 148.99, 146.89, 138.23, 129.46, 127.08, 121.81, 118.62, 72.66, 41.96, 40.77, 37.78, 37.18, 31.61, 30.98, 26.07, 21.34 ppm.

MS(ESI) m/z [M+H]⁺: 334.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 334.1473, found 334.1473.

IR (film): 3508, 2984, 1630, 1428, 1204 cm⁻¹.

Optical rotation: $[\alpha]_D^{27} = -9.72$ (*c* = 1.995, CHCl₃, > 20/1 dr).



5al': Procedure B, 53 mg, colorless liquid, 53% yield, 10/1 dr (Diastereoselectivity was determined by ¹H NMR analysis of reaction crude mixture).

¹**H NMR (400 MHz, CDCl₃)** δ 8.72 (d, *J* = 4.5 Hz, 1H), 8.08 (d, *J* = 7.8 Hz, 1H), 7.95 (td, *J* = 7.8, 1.5 Hz, 1H), 7.52 (dd, *J* = 7.0, 4.9 Hz, 1H), 7.10 (dt, *J* = 14.7, 7.2 Hz, 1H), 6.62 (d, *J* = 15.3 Hz, 1H), 5.49 (s, 1H), 4.20 (t, *J* = 6.3 Hz, 1H), 2.58–2.42 (m, 2H), 2.39–2.30 (m, 1H), 2.27–2.21 (m, 2H), 2.18 (t, *J* = 5.1 Hz, 1H), 2.08 (s, 1H), 2.02–1.92 (m, 1H), 1.28 (s, 3H), 1.05 (d, *J* = 8.7 Hz, 1H), 0.80 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 158.32, 150.18, 148.56, 146.72, 138.22, 129.47, 127.08, 121.83, 119.00,
72.56, 41.84, 40.77, 37.77, 37.08, 31.56, 31.00, 26.01, 21.35 ppm.

MS(ESI) m/z [M+Na]⁺: 356.05.

HRMS(ESI) m/z [M+H]⁺: calcd. 334.1471, found 334.1472.

IR (film): 3515, 2986, 1428, 1276 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{27} = -31.50$ (*c* = 1.470, CHCl₃, 10/1 dr).



¹H NMR (400 MHz, CDCl₃) δ 7.43–7.28 (m, 5H), 4.97 (dd, *J* = 8.8, 3.6 Hz, 1H), 3.87 (t, *J* = 5.6 Hz, 2H), 2.77 (s, 1H), 2.32 (s, 1H), 2.11–1.87 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 144.4, 128.6, 127.7, 125.7, 74.6, 61.6, 40.5 ppm. Optical rotation: $[α]_D^{25} = +64.7$ (c = 1.000, CHCl₃, 99% ee).



¹H NMR (400 MHz, CDCl₃) δ 7.35–7.19 (m, 5H), 6.71 (ddt, *J* = 21.9, 17.1, 7.1 Hz, 1H), 5.61 (dd, *J* = 21.5, 17.1 Hz, 1H), 4.80 (dd, *J* = 6.5, 5.4 Hz, 1H), 4.07–3.91 (m, 4H), 2.71–2.45 (m, 2H), 1.28 (td, *J* = 7.1, 3.6 Hz, 6H), 0.88 (s, 9H), 0.04 (s, 3H), -0.13 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.54 (d, J = 4.7 Hz), 144.08, 128.14, 127.23, 125.69, 119.41 (d, J = 186.8 Hz), 73.72 (d, J = 1.4 Hz), 61.53 (d, J = 5.4 Hz), 45.62 (d, J = 21.9 Hz), 25.75, 18.13, 16.29 (d, J = 6.8 Hz), -4.74, -5.04 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.79 ppm.

MS(ESI) m/z [M+H]⁺: 399.20.

HRMS(ESI) m/z [M+H]⁺: calcd. 399.2115, found 399.2116.

IR (film): 2982, 1634, 1259, 1029, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +28.54 (*c* = 1.580, CHCl₃).



¹H NMR (400 MHz, CDCl₃) δ 7.32–7.27 (m, 4H), 7.25–7.17 (m, 1H), 4.64 (dd, *J* = 7.3, 3.6 Hz, 1H), 4.15–3.94 (m, 4H), 1.84–1.62 (m, 6H), 1.29 (td, *J* = 7.0, 4.4 Hz, 6H), 0.88 (s, 9H), 0.02 (s, 3H), -0.16 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 145.17, 128.02, 126.92, 125.75, 74.48 (d, J = 1.6 Hz), 61.34 (d, J = 6.5 Hz), 41.63 (d, J = 16.1 Hz), 25.80, 25.61 (d, J = 140.6 Hz), 18.65 (d, J = 5.0 Hz), 18.15, 16.41 (d, J = 6.1 Hz), -4.65, -5.04 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 32.07 ppm.

MS(ESI) m/z [M+H]⁺: 401.20.

HRMS(ESI) m/z [M+H]⁺: calcd. 401.2271, found 401.2271.

IR (film): 2929, 1454, 1270, 1030, 836 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = +38.26$ (*c* = 0.855, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 7.60 (s, 1H), 7.34–7.27 (m, 8H), 7.26–7.17 (m, 2H), 4.76 (t, *J* = 5.8 Hz, 1H), 4.23 (q, *J* = 7.1 Hz, 2H), 2.78–2.62 (m, 1H), 2.58–2.39 (m, 1H), 2.01–1.86 (m, 2H), 1.31 (t, *J* = 7.1 Hz, 3H), 0.89 (s, 9H), 0.03 (s, 3H), -0.12 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 168.38, 145.06, 138.65, 135.47, 132.85, 129.45, 128.42, 128.26, 128.00, 126.93, 125.89, 74.96, 60.70, 39.68, 25.86, 23.83, 18.20, 14.29, -4.70, -4.96 ppm.

MS(ESI) m/z [M+Na]⁺: 447.20.

HRMS(ESI) m/z [M+Na]⁺: calcd. 447.2332, found 447.2332.

IR (film): 2956, 1709, 1630 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +106.96 (*c* = 0.915, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 7.45–7.17 (m, 11H), 6.90–6.75 (m, 2H), 4.77 (t, *J* = 5.7 Hz, 1H), 4.33–4.09 (m, 2H), 2.63–2.38 (m, 2H), 1.98–1.71 (m, 2H), 1.34–1.25 (m, 3H), 0.95 (s, 9H), 0.09 (s, 3H), -0.10 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 167.97, 145.09, 139.06, 138.18, 136.51, 132.12, 128.65, 128.63, 128.05, 127.08, 126.87, 125.92, 123.61, 74.67, 60.50, 40.65, 25.89, 23.19, 18.24, 14.31, -4.65, -4.94 ppm.
MS(ESI) m/z [M+Na]⁺: 473.25.

HRMS(ESI) m/z [M+Na]⁺: calcd. 473.2488, found 473.2488.

IR (film): 2927, 1704, 1621, 1452, 1228 cm⁻¹.

Optical rotation: $[\alpha]_D^{25} = +102.52$ (*c* = 0.890, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 8.08 (d, *J* = 7.2 Hz, 2H), 7.56 (t, *J* = 7.4 Hz, 1H), 7.48–7.39 (m, 4H), 7.37–7.32 (m, 2H), 7.30–7.24 (m, 1H), 6.05 (dd, *J* = 7.5, 6.0 Hz, 1H), 5.79 (ddt, *J* = 17.1, 10.2, 7.0 Hz, 1H), 5.12 (dd, *J* = 17.1, 1.4 Hz, 1H), 5.07 (d, *J* = 10.7 Hz, 1H), 2.89–2.76 (m, 1H), 2.74–2.60 (m, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 181.42, 165.68, 133.21, 132.92, 129.61, 128.43, 128.32, 127.93, 126.43,

118.18 ppm. **MS(ESI) m/z [M+NH₄]⁺:** 270.10. **HRMS(ESI) m/z [M+NH₄]⁺:** calcd. 270.1489, found 270.1488. **IR (film):** 2960, 1723, 1494, 1270, 1026 cm⁻¹. **Optical rotation:** $[\alpha]_D^{25} = -2.676$ (*c* = 0.275, CHCl₃).



¹H NMR (400 MHz, CDCl₃) δ 8.72 (d, *J* = 4.2 Hz, 1H), 8.05 (d, *J* = 7.8 Hz, 1H), 7.95 (td, *J* = 7.7, 1.6 Hz, 1H), 7.58–7.48 (m, 1H), 7.35–7.22 (m, 5H), 4.77–4.58 (m, 1H), 3.55–3.31 (m, 2H), 2.20 (d, *J* = 3.4 Hz, 1H), 1.95–1.77 (m, 4H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 157.05, 150.21, 144.01, 138.17, 128.53, 127.72, 127.35, 125.67, 122.19, 73.69, 51.65, 37.25, 18.84 ppm.

MS(ESI) m/z [M+Na]⁺: 314.00.

HRMS(ESI) m/z [M+Na]⁺: calcd. 314.0821, found 314.0821.

IR (film): 3506, 2997, 1579, 1428, 1270, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +27.68 (*c* = 1.210, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 8.73 (d, *J* = 4.6 Hz, 1H), 8.05 (d, *J* = 7.8 Hz, 1H), 7.94 (td, *J* = 7.7, 1.2 Hz, 1H), 7.54 (dd, *J* = 7.6, 4.7 Hz, 1H), 7.35–7.03 (m, 5H), 4.64 (t, *J* = 5.0 Hz, 1H), 3.47–3.28 (m, 2H), 1.85–1.65 (m, 4H), 0.82 (s, 9H), -0.04 (s, 3H), -0.20 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 157.04, 150.23, 144.52, 138.04, 128.10, 127.24, 127.09, 125.65, 122.25, 74.17, 51.94, 39.11, 25.76, 18.58, 18.09, -4.70, -5.11 ppm.

MS(ESI) m/z [M+Na]⁺: 428.15.

HRMS(ESI) m/z [M+Na]⁺: calcd. 428.1686, found 428.1686.

IR (film): 2955, 1578, 1257, 1164, 777 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = +52.64$ (*c* = 1.520, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 7.53–6.89 (m, 10H), 6.37 (d, *J* = 15.8 Hz, 1H), 6.20 (dt, *J* = 15.8, 6.7 Hz, 1H), 4.70 (dd, *J* = 7.4, 4.8 Hz, 1H), 2.41–2.07 (m, 2H), 1.98–1.65 (m, 2H), 0.90 (s, 9H), 0.04 (s, 3H), -0.15

(s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 145.46, 137.81, 130.58, 129.91, 128.43, 128.01, 126.89, 126.77, 125.88, 125.86, 74.45, 40.41, 29.03, 25.86, 18.22, -4.59, -4.92 ppm.

MS(DART) m/z [M+NH₄]⁺: 370.20.

HRMS(DART) m/z [M+NH₄]⁺: calcd. 370.2561, found 370.2557.

IR (film): 2928, 1600, 1257, 1092, 777 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +27.29 (*c* = 0.920, CHCl₃).



¹**H** NMR (400 MHz, CDCl₃) δ 7.97–7.86 (m, 2H), 7.54 (t, *J* = 7.4 Hz, 1H), 7.49–7.35 (m, 2H), 7.32–7.23(m, 4H), 7.22–7.18 (m, 1H), 4.75–4.63 (m, 1H), 2.94 (t, *J* = 6.5 Hz, 2H), 1.88–1.65 (m, 4H), 0.88 (s, 9H), 0.03 (s, 3H), -0.15 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 200.17, 145.38, 136.97, 132.83, 128.49, 128.00, 127.97, 126.86, 125.82, 74.87, 40.37, 38.48, 25.83, 20.44, 18.19, -4.65, -4.97 ppm.

MS(ESI) m/z [M+Na]⁺: 391.15.

HRMS(ESI) m/z [M+Na]⁺: calcd. 391.2070, found 391.2072.

IR (film): 2927, 1690, 1450, 1270, 1097, 837 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +36.41 (*c* = 0.425, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 8.73 (d, *J* = 4.4 Hz, 1H), 8.12 (d, *J* = 7.9 Hz, 1H), 8.01 (td, *J* = 7.8, 1.6 Hz, 1H), 7.60 (ddd, *J* = 7.6, 4.7, 0.9 Hz, 1H), 7.39–7.22 (m, 5H), 4.98 (dd, *J* = 9.5, 3.0 Hz, 1H), 4.68 (s, 1H), 4.61 (t, *J* = 9.3 Hz, 1H), 3.60 (dd, *J* = 14.9, 9.0 Hz, 1H), 3.53 (s, 1H), 3.49 (dd, *J* = 14.9, 2.3 Hz, 1H), 2.08–1.95 (m, 1H), 1.86 (dt, *J* = 14.3, 3.0 Hz, 1H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 157.43, 149.89, 143.71, 138.70, 128.50, 127.74, 127.69, 125.64, 122.04, 73.74, 66.54, 59.66, 44.67 ppm.

MS(ESI) m/z [M+H]⁺: 308.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 308.0951, found 308.0953.

IR (film): 3444, 2924, 1580, 1428, 1308, 793 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25}$ = +27.58 (*c* = 0.710, CHCl₃).



¹H NMR (400 MHz, CDCl₃) δ 7.38–7.18 (m, 5H), 6.77 (ddt, *J* = 21.8, 17.1, 7.1 Hz, 1H), 6.52 (d, *J* = 15.8

Hz, 1H), 6.15 (dd, J = 15.9, 6.4 Hz, 1H), 5.71 (dd, J = 21.3, 17.1 Hz, 1H), 4.43 (q, J = 5.9 Hz, 1H), 4.11–3.87 (m, 4H), 2.62–2.42 (m, 2H), 1.27 (q, J = 7.2 Hz, 6H), 0.91 (s, 9H), 0.09 (s, 3H), 0.06 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 149.41 (d, *J* = 4.9 Hz), 136.57, 131.83, 129.88, 128.53, 127.57, 126.38, 119.54 (d, *J* = 186.7 Hz), 72.22 (d, *J* = 1.3 Hz), 61.59 (d, *J* = 5.6 Hz), 43.39 (d, *J* = 21.8 Hz), 25.82, 18.18, 16.28 (d, *J* = 6.5 Hz), -4.33, -4.84 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 17.83 ppm.

MS(ESI) m/z [M+Na]⁺: 447.15.

HRMS(ESI) m/z [M+H]⁺: calcd. 425.2271, found 425.2273.

IR (film): 2958, 1635, 1252, 1029 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = -19.61$ (*c* = 1.145, CHCl₃).



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¹**H NMR (400 MHz, CDCl₃)** δ 7.45–7.18 (m, 5H), 6.56 (d, *J* = 15.9 Hz, 1H), 6.18 (dd, *J* = 15.9, 6.5 Hz, 1H), 4.79 (dd, *J* = 12.2, 6.4 Hz, 1H), 4.20–4.03 (m, 4H), 3.13 (dq, *J* = 22.6, 13.5 Hz, 2H), 2.99 (dd, *J* = 15.6, 7.5 Hz, 1H), 2.79 (dd, *J* = 15.6, 5.1 Hz, 1H), 1.41–1.23 (m, 6H), 0.89 (s, 9H), 0.08 (s, 3H), 0.06 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 200.29, 136.55, 131.53, 129.86, 128.53, 127.60, 126.42, 70.16, 62.50 (d, J = 6.4 Hz), 52.13, 43.81 (d, J = 126.7 Hz), 25.80, 18.09, 16.27 (d, J = 6.2 Hz), -4.29, -4.99 ppm.

³¹P NMR (162 MHz, CDCl₃) δ 19.63 ppm.

MS(ESI) m/z [M+Na]⁺: 463.20.

HRMS(ESI) m/z [M+H]⁺: calcd. 441.2221, found 441.2223.

IR (film): 2929, 1716, 1472, 1249, 1026, 780 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = -46.06$ (*c* = 1.000, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 7.65–7.45 (m, 3H), 7.40–7.28 (m, 7H), 7.26–7.20 (m, 1H), 6.78 (d, *J* = 16.2 Hz, 1H), 6.61 (d, *J* = 15.9 Hz, 1H), 6.26 (dd, *J* = 15.9, 6.2 Hz, 1H), 4.90 (dt, *J* = 6.5, 5.5 Hz, 1H), 3.09 (dd, *J* = 14.7, 7.8 Hz, 1H), 2.76 (dd, *J* = 14.7, 4.9 Hz, 1H), 0.87 (s, 9H), 0.06 (s, 3H), 0.06 (s, 3H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 198.50, 143.22, 136.74, 134.52, 132.19, 130.46, 129.50, 128.91, 128.54, 128.31, 127.53, 127.30, 126.44, 70.75, 49.25, 25.83, 18.15, -4.34, -4.94 ppm.

MS(ESI) m/z [M+Na]⁺: 415.15.

HRMS(ESI) m/z [M+Na]⁺: calcd. 415.2064, found 415.2061.

IR (film): 2955, 1689, 1609, 1471, 1249, 836, 779 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = -104.50$ (*c* = 1.000, CHCl₃).


¹H NMR (400 MHz, CDCl₃) δ 7.61–7.41 (m, 3H), 7.38–7.04 (m, 8H), 6.70–6.50 (m, 2H), 6.21 (dd, *J* = 15.9, 5.9 Hz, 1H), 4.90–4.73 (m, 1H), 3.33 (s, 1H), 3.07–2.77 (m, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 200.00, 143.92, 136.56, 134.12, 130.85, 130.40, 130.27, 129.01, 128.55, 128.43, 127.69, 126.49, 126.23, 68.74, 46.88 ppm.

MS(ESI) m/z [M+Li]⁺: 285.10.

HRMS(ESI) m/z [M+Li]⁺: calcd. 285.1463, found 285.1462.

IR (film): 3385, 2924, 1458, 1275, 1260, 749 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = -10.78$ (*c* = 0.615, CHCl₃).



¹**H NMR (400 MHz, CDCl₃)** δ 7.48–7.20 (m, 10H), 6.65 (d, *J* = 15.8 Hz, 2H), 6.31 (dd, *J* = 15.8, 5.9 Hz, 2H), 4.79–4.58 (m, 2H), 2.60 (s, 2H), 1.98 (t, *J* = 5.3 Hz, 2H) ppm.

¹³C NMR (100 MHz, CDCl₃) δ 136.55, 131.62, 130.21, 128.59, 127.70, 126.47, 70.47, 42.64 ppm.

MS(ESI) m/z [M+Na]⁺: 303.10.

HRMS(ESI) m/z [M+Na]⁺: calcd. 303.1356, found 303.1356.

IR (film): 3358, 2954, 2924, 1260, 963, 750 cm⁻¹.

Optical rotation: $[\alpha]_{D}^{25} = +46.20 (c = 0.260, CHCl_{3}).$



yashabushidiol B

¹H NMR (400 MHz, CDCl₃) δ 7.32–7.26 (m, 4H), 7.23–7.13 (m, 6H), 4.05–3.92 (m, 2H), 2.83–2.73 (m, 2H), 2.71–2.58 (m, 2H), 2.30 (d, J = 4.2 Hz, 2H), 1.92–1.71 (m, 4H), 1.67 (t, J = 5.6 Hz, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃) δ 141.82, 128.43, 128.35, 125.89, 68.93, 42.52, 39.08, 32.16 ppm. MS(ESI) m/z [M+Na]⁺: 307.10.

HRMS(ESI) m/z [M+Na]⁺: calcd. 307.1674, found 307.1672.

IR (film): 3285, 2923, 1453, 1061, 919, 727 cm⁻¹.

Optical rotation: $[\alpha]_D^{25} = +5.66$ (*c* = 0.250, CHCl₃). {literature (Hashimoto et. al., 1986), $[\alpha]_D = +7.2$ (CHCl₃)}.

Supplemental References

Denmark, S. E., and Heemstra, J. R. Jr. (2004). Lewis base activation of lewis acids: vinylogous aldol additions of silyl dienol ethers to aldehydes. Synlett 2411–2416.

Yuan, C., Yao, J., and Li, S. (1991). Studies on organophosphorus compounds XLV structure effects of ester alkyl group of dialkyl allylphosphonate carbanion on the regioselectivity of electrophilic addition. Phosphorus, Sulfur Silicon Relat. Elem. *55*, 125–131.

Hashimoto, T., Tori, M., and Asakawa, Y. (1986). Five new diarylheptanoids from the male flowers of alnus sieboldiana. Chem. Pharm. Bull. *34*, 1846–1849.