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LETTER TO THE EDITOR

## Longevity and correlation with disease severity of the humoral and cellular response to SARS-CoV-2 infection in haemodialysis patients

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Following severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) infection, the large majority of haemodialysis (HD) patients rapidly seroconvert [1-3], with robust and durable serological responses up to 10 months [4]. These observations appear non-sequitur in view of the well-known immune dysfunction of HD patients, resulting in slower viral clearance and higher case fatality rates of SARS-CoV-2 infection, as well as impaired responses to SARS-CoV-2 vaccination [5]. A potential explanation may be that dialysis patients develop more severe disease with prolonged viral shedding and higher levels of pro-inflammatory cytokines, resulting in more intense immune stimulation. In a study of 2215 HD patients, older and diabetic patients had slightly higher antibody levels, but data on disease severity were not available [6].

In the general population, robust T-cell responses have been observed in asymptomatic patients even in the absence of detectable antibodies, suggesting that T-cell immunity develops independently of disease severity [7]. A cohort of 14 HD patients, sampled a mean of 40 days after coronavirus disease 2019 diagnosis, displayed T-cell responses of similar intensity to those of patients with normal renal function [8]. Currently there are no

data on the durability and correlation with disease severity of T-cell immunity in HD patients.

Of the 569 patients on HD in March 2021, 41 had a history of polymerase chain reaction (PCR)-confirmed SARS-CoV-2 infection, of which 24 had asymptomatic or mild disease and 17 had severe disease requiring hospitalization (Supplementary data, Table S1). Patients were stratified by days between the PCR test and blood sampling into Period 1 [mean 348 days (range 327-378)] and Period 2 [mean 125 days (range 48-200)].

Serological responses persisted up to >1 year, with a faster decay of anti-nucleocapsid protein (anti-N) immunoglobulin G (IgG) than anti-spike protein (anti-S) IgG (Figure 1A and B). Patients with severe disease had higher antibody levels (Table 1), but the decay trajectory was similar. Interferon- $\gamma$  production by peripheral blood CD4+ and CD8+ lymphocytes was unaffected by disease severity (Table 1) and could be detected up to >1 year after infection but did not display a clear pattern of evolution over time (Figure 1C).

In conclusion, the humoral response to SARS-CoV-2 infection in HD patients is a marker of disease severity. Prolonged viral antigen exposure and production of inflammatory cytokines

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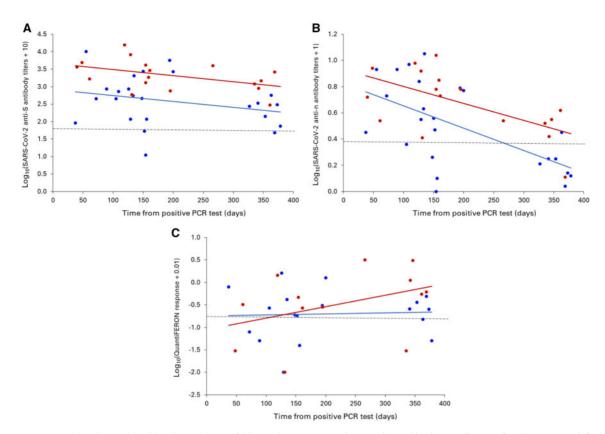


FIGURE 1: SARS-CoV-2 (A) anti-S IgG titres, (B) anti-N IgG titres and (C) QuantiFERON response in HD patients with a history of PCR-confirmed SARS-CoV-2 infection that had asymptomatic or mild disease (blue circles) or severe disease (red circles) as a function of the time interval between the lowest cycle threshold value and serum sampling. Anti-S IgG was measured by a chemiluminescent microparticle immunoassay (CMIA) on the ARCHITECT i System (SARS-CoV-2 IgG II Quant assay, Abbott Laboratories, Abbott Park, IL, USA). The cut-off for positivity (dashed line) was 50 AU/mL and the conversion factor to the World Health Organization binding antibody units  $(BAU/mL) \ is \ BAU/mL = 0.142 \times AU/mL. \ Anti-N \ IgG \ was \ measured \ by \ a \ CMIA \ on \ the \ same \ ARCHITECT \ i \ analyser \ (SARS-CoV-2 \ IgG \ assay, \ Abbott). \ A \ signal: cut-off \ ratio \ \ge 1.4 \times 10^{-1} \ same \ ARCHITECT \ i \ analyser \ (SARS-CoV-2 \ IgG \ assay, \ Abbott).$ (dashed line) was interpreted as reactive. The QuantiFERON SARS-CoV-2 test (Qiagen, Venlo, The Netherlands) measures the secretion of interferon-y by peripheral blood  $CD4^{+} \ and \ CD8^{+} \ lymphocytes \ upon \ SARS-CoV-2 \ glycoprotein \ stimulation. \ The \ threshold for \ positivity \ (dashed \ line) \ is \ 0.15 \ IU/mL.$ 

Table 1. Humoral and cellular response to SARS-CoV-2 infection stratified by disease severity and interval between infection and sampling

	Mild disease (95% CI)	Severe disease (95% CI)	P-value <sup>a</sup>
Period 1 (16 March 2020–30 August 2020)	n = 7	n = 6	
Anti-S IgG (GMT)	187 (96–362)	1306 (633–2695)	0.003
Anti-N IgG (GMT)	1.62 (1.29–2.03)	2.87 (2.05–4.03)	0.016
QuantiFERON (GMC)	0.21 (0.11–0.40)	0.69 (0.18–2.74)	0.154
Period 2 (31 August 2020–2 March 2021)	n = 17	n=11	
Anti-S IgG (GMT)	525 (227–1215)	2658 (1483–4763)	0.010
Anti-N IgG (GMT)	4.10 (2.93–5.73)	6.18 (4.77–8.00)	0.097
QuantiFERON (GMC)	0.20 (0.08–0.46)	0.17 (0.05–0.62)	0.880

<sup>&</sup>lt;sup>a</sup>According to linear models for log<sub>10</sub>-transformed outcomes. GMT, geometric mean titre; GMC, geometric mean concentration.

due to a more severe disease course may explain the intense and long-lasting antibody production in HD patients. Conversely, the cellular immune response is unaffected by disease severity and appears to be governed by as yet undetermined factors.

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## **CONFLICT OF INTEREST STATEMENT**

None declared.

## REFERENCES

- 1. De Vriese AS, Reynders M. IgG antibodyresponse to SARS-CoV-2 infection and viral RNA persistence in patients on maintenance hemodialysis. Am J Kidney Dis 2020; 76: 440-441
- 2. Sakhi H, Dahmane D, Attias P et al. Kinetics of anti-SARS-CoV-2 IgG antibodies in hemodialysis patients six months after infection. J Am Soc Nephrol 2021; 32: 1033-1036

- Clarke CL, Prendecki M, Dhutia A et al. Longevity of SARS-CoV-2 immune responses in hemodialysis patients and protection against reinfection. Kidney Int 2021; 99: 1470–1477
- Dudreuilh C, Roper T, Breen C et al. IgG SARS-CoV-2 antibodies persist at least for 10 months in patients on hemodialysis. Kidney Int Rep 2021; 6: 1961–1964
- Ikizler TA, Coates PT, Rovin BH et al. Immune response to SARS-CoV-2 infection and vaccination in patients receiving kidney replacement therapy. Kidney Int 2021; 99: 1275–1279
- Anand S, Montez-Rath ME, Han J et al. Serial SARS-CoV-2 receptor-binding domain antibody responses in patients receiving dialysis. Ann Intern Med 2021; 174: 1073–1080
- Sekine T, Perez-Potti A, Rivera-Ballesteros O et al. Robust T cell immunity in convalescent individuals with asymptomatic or mild COVID-19. Cell 2020; 183: 158–168.e14
- 8. Anft M, Blazquez-Navarro A, Paniskaki K et al. SARS-CoV-2-reactive cellular and humoral immunity in hemodialysis population. Kidney Int 2021; 99: 1489–1490